



This is a repository copy of *Hybrid 'superswarm' leads to rapid divergence and establishment of populations during a biological invasion.*

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/94393/>

Version: Accepted Version

Article:

Roy, D., Lucek, K., Walter, R.P. et al. (1 more author) (2015) Hybrid 'superswarm' leads to rapid divergence and establishment of populations during a biological invasion. *Molecular Ecology*, 24 (21). pp. 5394-5411. ISSN 0962-1083

<https://doi.org/10.1111/mec.13405>

This is the peer reviewed version of the following article: Roy, D., Lucek, K., Walter, R. P. and Seehausen, O. (2015), Hybrid 'superswarm' leads to rapid divergence and establishment of populations during a biological invasion. *Molecular Ecology*, 24: 5394–5411, which has been published in final form at <http://dx.doi.org/10.1111/mec.13405>. This article may be used for non-commercial purposes in accordance with Wiley Terms and Conditions for Self-Archiving (<http://olabout.wiley.com/WileyCDA/Section/id-820227.html>).

Reuse

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

MOLECULAR ECOLOGY

Hybrid 'superswarm' leads to rapid divergence and establishment of populations during a biological invasion

Journal:	<i>Molecular Ecology</i>
Manuscript ID:	MEC-15-0690
Manuscript Type:	Original Article
Date Submitted by the Author:	17-Jun-2015
Complete List of Authors:	Roy, Denis; University of Connecticut, Natural resources and the Environment & Center for Environmental Sciences and Engineering Lucek, Kay; University of Bern, Institute of Ecology and Evolution; EAWAG, Fish Ecology & Evolution Walter, Ryan; California State University Fullerton, Department of Biological Science Seehausen, Ole; University of Bern, Institute of Ecology and Evolution; EAWAG, Fish Ecology & Evolution
Keywords:	Contemporary Evolution, Ecological Genetics, Hybridization, Invasive Species, Population Genetics - Empirical, Speciation

1 **Hybrid ‘superswarm’ leads to rapid divergence and establishment of**
2 **populations during a biological invasion**

3 Denis Roy^{1,§,*}, Kay Lucek^{1,2}, Ryan P. Walter³, Ole Seehausen^{1,2}

4 ¹ Centre for Ecology, Evolution & Biogeochemistry
5 EAWAG Federal Institute of Aquatic Science and Technology
6 Seestrasse 79, 6074 Kastanienbaum
7 Switzerland

8
9 ² Institute for Ecology and Evolution
10 University of Bern
11 Baltzerstrasse 6, 3012 Bern
12 Switzerland

13
14 ³ Department of Biological Science
15 California State University Fullerton
16 Fullerton, CA 92831
17 USA

18
19 [§] Current address: Department of Natural Resources and the Environment and Center for
20 Environmental Sciences and Engineering
21 University of Connecticut
22 3107 Horsebarn Hill Road
23 Storrs, CT 06269-4210
24 USA

25
26
27 **Keywords:** Colonization, Hybridization, Rapid divergence, Stickleback, Switzerland

28 *** Corresponding author**

29

30 **Running title:** Rapidly formed populations by hybridization

31 Abstract

32 Understanding the genetic background of invading species can be crucial information clarifying
33 why they become invasive. Intraspecific genetic admixture among lineages separated in the
34 native ranges may promote the rate and extent of an invasion by substantially increasing standing
35 genetic variation. Here we examine the genetic relationships among threespine stickleback that
36 recently colonized Switzerland. This invasion results from several distinct genetic lineages that
37 colonized multiple locations and have since undergone range expansions, where they coexist and
38 admix in parts of their range. Using 17 microsatellites genotyped for 634 individuals collected
39 from 17 Swiss and two non-Swiss European sites we reconstruct the invasion of stickleback and
40 investigate the potential and extent of admixture and hybridization among the colonizing
41 lineages from a population genetic perspective. Specifically we test for an increase in standing
42 genetic variation in populations where multiple lineages coexist. We find strong evidence of
43 rapid and massive hybridization coupled with the recent development of genetic isolation that
44 has led to the formation of several new genetically distinguishable populations, consistent with a
45 hybrid 'superswarm'. This massive hybridization and population formation event(s) occurred
46 over approximately 140 years and likely fuelled the successful invasion of Swiss waterways. The
47 implications are that multiple colonizations coupled with hybridization can lead to the formation
48 of new stable populations potentially kick-starting speciation and adaptive radiation over very
49 short time.

50 **Introduction**

51 Populations introduced outside their species range may suffer severe genetic bottlenecks and
52 founder effects reducing levels of standing genetic variation available for selection. This can
53 substantially decrease the population's ability to establish and spread into novel environments
54 (Lockwood *et al.* 2007; Dlugosch & Parker 2008; Prentis *et al.* 2008; Simberloff 2009).
55 Consequently, many introduced species persist only locally or briefly after their introduction
56 (Sakai *et al.* 2001; Lockwood *et al.* 2007). Some introduced species, meanwhile, establish viable
57 populations and undergo range expansions despite initial decreases in genetic variation relative
58 to their ancestral population (Lockwood *et al.* 2007; Dlugosch & Parker 2008). Less commonly,
59 introduced species may colonize new geographic regions from multiple, yet genetically distinct
60 sources, which can meet in secondary contact zones after initial range expansions. Within such
61 contact zones distinct lineages can hybridize converting between-lineage genetic variation to
62 within-population genetic variance (Mallet 2007; Prentis *et al.* 2008; Abbott *et al.* 2013;
63 Seehausen *et al.* 2014). This, in turn, increases standing genetic variation and reduces genetic
64 constraints in newly formed hybrid populations, augmenting their genetic potential or
65 adaptability (Mallet 2007; Prentis *et al.* 2008; Nolte & Tautz 2010; Abbott *et al.* 2013;
66 Seehausen *et al.* 2014; Williams *et al.* 2014). Release from former genetic constraints may occur
67 because allelic combinations fixed in parental lineages, expressed through their genetic variance-
68 covariance matrices (VCVs), can be disrupted, the genetic covariance broken and the genetic
69 variance broadened in ensuing hybrids (Buerkle *et al.* 2000; Abbott *et al.* 2013; Seehausen *et al.*
70 2014). Broadened genetic VCVs may better respond to directional selection than narrower ones
71 especially when selection is applied off the main VCV trajectory (assuming loci reflect
72 quantitative traits under selection with some heritability; Schluter 1996; Stepan *et al.* 2002;

73 Schluter & Conte 2009; Seehausen *et al.* 2014). A direct prediction of this is that hybrid lines
74 ought to have greater variance and reduced directionality (i.e., narrowness) in their genetic
75 VCVs than their formative parental lineages (Mallet 2007; Prentis *et al.* 2008; Schluter & Conte
76 2009; Abbott *et al.* 2013; Seehausen *et al.* 2014).

77
78 An increased genetic potential in hybrid populations may facilitate subsequent colonization and
79 establishment, and allow genetically admixed individuals to tap into novel niches within the
80 invaded range not typically occupied by any of its ancestors (Lockwood *et al.* 2007; Yoder *et al.*
81 2010; Williams *et al.* 2014). For hybrids to persist, however, their distribution (in allopatry)
82 and/or the balance between selection and gene flow (in sympatry or parapatry) should help
83 establish reproductive isolation (Grant 1994; Buerkle *et al.* 2000; Mallet 2007; Nolte & Tautz
84 2010; Abbott *et al.* 2013; Seehausen *et al.* 2014). Otherwise, newly formed gene/trait
85 combinations can be quickly eliminated or resorbed into parental lines (Grant 1994; Buerkle *et al.*
86 2000; Mallet 2007; Schluter & Conte 2009). The establishment of such newly adapted,
87 reproductively isolated populations can ultimately lead to the formation of new species (Buerkle
88 *et al.* 2000; Mallet 2007; Nolte & Tautz 2010; Abbott *et al.* 2013; Seehausen *et al.* 2014) and,
89 under the right conditions, facilitate adaptive radiations (Seehausen 2004; Schluter & Conte
90 2009; Nolte & Tautz 2010; Abbott *et al.* 2013).

91
92 Despite an increasing number of both theoretical and empirical studies underscoring the
93 importance of hybridization during biological invasions and species formation (Buerkle *et al.*
94 2000; Seehausen 2004; Mallet 2007; Seehausen *et al.* 2008; Abbott *et al.* 2013), the population
95 genetic mechanisms operating from secondary contact to the emergence of new hybrid types

96 remain vague (Nolte & Tautz 2010; Abbott *et al.* 2013 but see Buerkle *et al.* 2000). Thus,
97 theoretical considerations notwithstanding, there is a need to identify systems appropriate for the
98 study of the incipient stages of hybrid lineage formation and subsequent speciation (Buerkle *et al.*
99 2000; Nolte & Tautz 2010; Seehausen *et al.* 2014). The identification of newly formed hybrid
100 lineages can not only provide key insights into the formation of new hybrid species, but also
101 answer important questions related to the pace of hybrid lineage stabilization and the associated
102 extent of genomic integration (Nolte & Tautz 2010; Abbott *et al.* 2013; Seehausen *et al.* 2014;
103 Williams *et al.* 2014).

104
105 The threespine stickleback (*Gasterosteus aculeatus* species complex) has repeatedly colonized
106 freshwater environments throughout its natural range from marine ancestors shortly after the last
107 glacial retreat (~ 10 000 years ago). In many newly colonized freshwater habitats, stickleback
108 have formed distinct ecotypes (McPhail 1984; Schluter 1993; Thompson *et al.* 1997; Kaeuffer *et*
109 *al.* 2012; Ravinet *et al.* 2013) mostly through recurrent selection on standing genetic variation
110 maintained at low frequencies in marine populations (Schluter & Conte 2009; Deagle *et al.* 2012;
111 Jones *et al.* 2012). Many of the studied marine-to-freshwater stickleback colonizations have been
112 associated with genetic bottlenecks, reducing genetic variation and likely, the adaptive potential
113 within freshwater habitats (Reusch *et al.* 2001; Mäkinen *et al.* 2006; Deagle *et al.* 2012). While
114 stickleback are common in many parts of Europe (Bertin 1925; Munzing 1963; Mäkinen *et al.*
115 2006), their distribution within Switzerland was initially restricted to the tributaries of the Rhine
116 near Basel prior to 1870 (Lucek *et al.* 2010; Fig. 1). Following several introductions and the
117 channelization of Swiss waterways (Heller 1870; Fatio 1882; Bertin 1925), stickleback
118 underwent a range expansion and now occupy a wide range of different habitats throughout the

119 country (Berner *et al.* 2010; Lucek *et al.* 2010; Lucek *et al.* 2013; Lucek *et al.* 2014). The Swiss
120 midlands are characterized by many large lakes linked by a vast network of streams and canals
121 allowing gene flow among different lake systems, which enables adaptation to distinct habitats
122 (e.g., shallow rivers and streams versus deep large lakes; Berner *et al.* 2010; Lucek *et al.* 2010;
123 Lucek *et al.* 2013; Lucek *et al.* 2014). A mitochondrial DNA survey of populations across the
124 country revealed the colonization of Switzerland by three distant genetic stickleback lineages
125 (five mtDNA haplotypes) from different parts of Europe (Lucek *et al.* 2010). The Lake
126 Constance area is dominated by an eastern European lineage from the Baltic region (haplotype
127 EU27; Mäkinen & Merilä 2008; Fig. 1; Table S1), whereas the Lake Geneva area is dominated
128 by a lineage typical of the Rhône (haplotypes EU09, EU10 and EU36). A third lineage
129 dominates the Basel region, and may have been native to that small part of Switzerland (CH01;
130 Lucek *et al.* 2010). Over the last 140 years, all three lineages have expanded into the Swiss
131 midlands. In places such as lakes Neuchâtel, Biel, Wohlen, and in their drainages, populations
132 are a mix of several mitochondrial lineages associated with elevated haplotype richness (Lucek
133 *et al.* 2010). An amplified fragment length polymorphism (AFLP) analysis suggested
134 considerable admixture between lineages in the Aare river region (near GIP and WOH; Fig. 1),
135 wherein individuals display increased phenotypic variation (Lucek *et al.* 2010).

136
137 Here, we use a suite of microsatellite markers to infer genetic relationships among stickleback
138 collected across Switzerland, substantially expanding on previous work relying on AFLPs
139 (Lucek *et al.* 2010), by adding samples collected within zones showing coexistence of multiple
140 mitochondrial lineages. First, we assess the population structure of stickleback in Switzerland in
141 the context of known introductions. We next determine the sizes and connectivity among

142 recovered populations assessing both their contemporary gene flow and that which has occurred
143 in the coalescent. Finally, and in the context of previous work in the system, we examine the
144 likelihood that some populations originate from the hybridization among main colonizing
145 lineages as determined by Lucek *et al.* (2010). Overall, we show that hybridization can lead to
146 the development of new populations whose connectivities are quickly reduced. These nascent
147 populations may thus represent important initial steps by which colonization and hybridization
148 work together to promote speciation, and potentially catalyze adaptive radiations over very short
149 time.

150

151 **Material and Methods**

152 *Sample collection & genotyping*

153 Stickleback were collected from 17 different sampling sites across Switzerland, between summer
154 2007 and autumn 2008 (Fig. 1; Table S1). The sampling sites included lakes, streams and ponds.
155 Two additional samples collected outside Switzerland were taken, representing populations to
156 the North and South of the invaded range (Lucek *et al.* 2010; i.e., a Mediterranean freshwater
157 population from Corsica and a North Sea derived freshwater population from Northern
158 Germany; Fig. 1 Table S1). DNA was extracted from each individual, using a Qiagen BioSprint
159 96 robot with the Qiagen Blood Extraction kit (Qiagen, Switzerland). The genotype of 634
160 individuals was assessed at 17 microsatellite loci using a CEQ 8000 (Beckman Coulter,
161 Switzerland) following manufacturer instructions. The 17 microsatellites are located on 15 of 26
162 linkage groups determined by Peichel *et al.* 2001 and were amplified in each individual using
163 five multiplexing sets (Table S2). Previous work has shown association between 7 of these
164 markers and the quantitative traits of spine lengths, the number of lateral plates and gill rakers

165 (Table S2). No evidence of null alleles, scoring errors or large allele dropouts was detected at
166 any loci in any sampling site after checking all genotypes using MICRO-CHECKER 2.2.3 (van
167 Oosterhout *et al.* 2004).

168

169 *Population Genetic Structure*

170 An iterative approach was used to get an unbiased, best estimate of the statistically supported
171 number of distinguishable genetic clusters adhering to population genetic criteria (i.e., satisfying
172 HWE and showing acceptable levels of linkage among loci). Population structure among all
173 genotyped individuals was first assessed using STRUCTURAMA 2.0 (Huelsenbeck *et al.* 2011)
174 which searches parameter space for the most likely number of genetic clusters using a Bayesian
175 framework. Population number was set to a random variable but allowed to vary using a
176 Dirichlet Process Prior (DPP). STRUCTURAMA searches used an unsupervised mode with
177 DPPs set to 1-10, 12, 15, 17, and 20. Each search ran for 10 000 000 iterations run over three
178 Markov Chain Monte Carlo (MCMC) sampling chains. The number of populations was collected
179 every 100th iteration collecting 100 000 values overall where the first 50 000 were discarded as
180 burnin (Huelsenbeck & Andolfatto 2007; Huelsenbeck *et al.* 2011). The most likely number of
181 genetic clusters recovered was determined either by consensus among searches or by selecting
182 results of the search(es) with maximized marginal likelihoods. STRUCTURAMA analyses were
183 performed hierarchically by first using the entire dataset to get an overall assessment of the
184 number of populations. All individuals were then assigned to a particular recovered cluster by
185 their largest posterior probabilities assessed by STRUCTURE (see below), regardless of location
186 and STRUCTURAMA analyses were then re-run on each cluster. This process was repeated
187 until no further sub-division of clusters was observed or even genotype splitting of all

188 individuals occurred (see Fig. 2). At each step of the hierarchical search, STRUCTURE 2.3.4
189 (Hubisz *et al.* 2009) was used to visualize recovered genetic clusters estimated from
190 STRUCTURAMA and assess individual admixture proportions outlining their probabilities of
191 belonging to recovered clusters. In STRUCTURE, the probability of each individual's
192 assignment to recovered clusters was assessed through 10 permutations of the number of clusters
193 recovered from STRUCTURAMA, with each permutation running over 1 000 000 iterations
194 with an additional 100 000 used as burnin. STRUCTURE analyses allowed admixture and used
195 correlated allele frequencies in the population structuring models. Results of all STRUCTURE
196 permutations assessed for each hierarchical step were combined into a single individual-based
197 clustering assignment probability using CLUMMP 1.1.2 (Jakobsson & Rosenberg 2007) and
198 plotted using DISTRUCT 1.1 (ROSENBERG 2004).

199 *Marker Neutrality*

200 Because the population structure recovered using markers under selection can differ from that
201 determined using neutral markers, (e.g., Jakobsdóttir *et al.* 2011; Bradbury *et al.* 2013; Roy *et al.*
202 2014) all loci were assessed for either balancing or diversifying selection. Markers were
203 subjected to both the stepwise mutation and the infinite allele models (SMM and IAM,
204 respectively) of microsatellite mutation and tested for neutrality using an F_{ST} outlier test (FDIST)
205 as applied in LOSITAN 2.0 (Antao *et al.* 2008). The application of both models used 1 000 000
206 permutations to establish 95% confidence intervals and used a sample size reflecting the smallest
207 genetic population under consideration. Selection affecting our markers was also tested using
208 Bayescan 2.1 (Foll & Gaggiotti 2008) which applies a Bayesian framework to determine whether
209 differentiation at a given locus is best explained by a model including a locus-specific
210 component (evidence of selection) or one that is strictly related to population(s) (i.e., neutral).

211 Bayescan assessments were set to collect every 100th iteration over a total of 1 000 000 steps for
212 a total of 10 000 recorded iterations. An additional 1 000 000 iterations were used as burnin.
213 Priors for each assessment were adjusted using 20 pilot runs, each running 50 000 iterations. All
214 three loci selection tests (FDIST-IAM/FDIST-SMM and Bayescan) were initially applied at the
215 base of the recovered population structure hierarchy but also applied at deeper levels within it.

216

217 *Population genetic indices and statistics*

218 Linkage disequilibrium among loci (LD) and their adherence to Hardy-Weinberg expectations
219 (HWE) was assessed in each genetic cluster recovered from the
220 STRUCTURAMA/STRUCTURE analyses (hereafter populations) using Arlequin version
221 3.5.1.2 (Excoffier & Lischer 2010). LD tests used 10 000 permutations and deviations from
222 HWE were tested using 1 000 000 MCMC iterations with 100 000 dememorization steps.
223 Significance of both LD and HWE tests were assessed using sequential Bonferonni corrections
224 (Rice 1989). Arlequin was also used to estimate population-specific observed and expected
225 heterozygosities (H_o and H_e , respectively). Population-specific allelic richness (with rarefaction;
226 A_R) and inbreeding coefficients (F_{IS}) were estimated in FSTAT 2.9.3.2 (Goudet 1995). The
227 number of private alleles (Pa) per population was also calculated (with rarefaction) using
228 GenalEx 6.5 (Peakall & Smouse 2006). Levels of genetic differentiation among all possible
229 population pairs was evaluated using the classic F_{ST} index (calculated as θ ; Weir & Cockerham
230 1984) supported by 1000 bootstraps and derived from 100 000 permutations of the MCMC
231 algorithm implemented in MSA 4.05 (Dieringer & Schlötterer 2003). The pairwise D_{Jost} index of
232 genetic differentiation was also estimated with using DEMETics (Gerlach *et al.* 2010) using 1000
233 bootstrapping iterations to calculate significance. To test whether loci putatively linked to

234 quantitative traits (see above) exhibited significantly different population genetic indices relative
235 to unlinked ones, global locus-specific A_R , H_o , H_e , F_{IS} and F_{ST} s were compared using Wilcoxon
236 sum rank tests. A_R , H_o , H_e , F_{IS} and Pa were also compared between Swiss populations (as
237 inferred by STRUCTURAMA) exhibiting mtDNA haplotypes consistent with a single main
238 colonizing lineage (hereafter MCL) versus those exhibiting the presence of multiple lineages
239 (see Fig. 1, Table S1) using Welch's Two-sample t -tests.

240

241 *Population Size and Connectivity*

242 Contemporary effective population sizes (N_e) were estimated for each population using the
243 linkage disequilibrium model (LDNe) based on single moment data as implemented in
244 N_e Estimator v2 (Do *et al.* 2014). LDNe uses the weighted average level of expected random
245 linkage disequilibrium among alleles over loci pairs within a given population to estimate its
246 effective size (Waples & England 2011). Estimates of N_e based on linkage disequilibrium
247 assume selective neutrality, no physical linkage among loci and a closed but randomly mating
248 population. Because our data could not identify differently aged individuals, and likely combined
249 several year classes, our estimates most likely reflect something between the effective number of
250 breeding individuals N_b and N_e (i.e., \widehat{N}_e) within each population rather than the true population-
251 specific N_e (Hare *et al.* 2011). These estimates may nevertheless be useful in gauging the relative
252 size of populations (Hare *et al.* 2011; Do *et al.* 2014). \widehat{N}_e estimates were made using allele
253 frequencies greater than 0.01 and 95% credible limits were established from jackknifing over all
254 loci pairs. Contemporary gene flow among populations was estimated using BayesAss 3.0
255 (Wilson & Rannala 2003) with 10 000 000 MCMC iterations used as burnin and sampling an

256 additional 100 000 000 iterations at an interval of 1000. This procedure used mixing parameters
257 of 0.3, 0.5 and 0.1 for allele frequencies, inbreeding coefficients and migration rates, respectively,
258 and led to a total sample size 100 000 from which estimates were derived.

259

260 *Coalescent-based Size and Connectivity*

261 To generate time-integrated estimates of N_e that also consider historical influences among
262 populations, including migration rates (m), we applied isolation with migration (IM) models
263 estimating the long term N_e and m of each population in the coalescent (Hey & Nielsen 2004;
264 Hey 2010). IM models search parameter space for the most likely estimates using a Bayesian
265 framework assuming random mating within populations and that populations are each other's
266 closest relatives not exchanging genes with other nonsampled populations (Hey & Nielsen 2004;
267 Hey 2010). We used IMA2 on a subsample of 9-35 individuals from each population combining
268 their microsatellite genotypes with 436 bp of mitochondrial control region (CR) and 965 bp of
269 cytochrome B (CytB) sequences determined by Lucek et al (2010). Although we recognize that
270 our data may violate some of the IM model assumptions, previous work has shown that IM
271 models as applied in IMA2, are generally robust to random mating violations and those involving
272 small to moderate levels of introgression among considered taxa (Strasburg & Rieseberg 2010).
273 IM analyses were run pairwise between populations following recommendations concerning the
274 information (i.e., number of marker loci) needed for reliable parameter estimation in studies
275 involving more than two populations (IMa2 manual; Hey 2010). Searches used priors determined
276 from preliminary runs and were iterated using between 6 000 000 - 26 000 000 steps to reach
277 stationary distributions before sampling. Once stationarity was achieved, all searches ran for an
278 additional 10 000 000 steps, sampling every 100th step for a total of 100 000 recorded

279 genealogies from which parameters were assessed. All analyses used 100 metropolis-coupled
 280 MCMC chains with heating terms ensuring high swap rates among them (<0.70). Long-term N_e
 281 and m were calculated from generated population-specific θ estimates using mutation rates of 1
 282 $\times 10^{-4}$, 9.6×10^{-6} , and 1.97×10^{-5} for microsatellites, CR and CytB sequences, respectively.
 283 These mutation rates were used in previous studies implementing IM based analyses in other
 284 stickleback populations (Caldera & Bolnick 2008; Mäkinen & Merilä 2008; Berner *et al.* 2009).
 285 Final population-specific long-term N_e was calculated by taking the geometric mean of all values
 286 determined from pairwise comparisons including the focal population. The proportion of
 287 migrants per generation emanating from a focal population was also recovered from all pairwise
 288 comparisons ($C \times V$; see IMA2 manual). We then used all comparisons including a focal
 289 population to estimate weighted migration rates to all other populations using the following
 290 formula:

$$291 \quad m_{i \rightarrow j} = \frac{\overline{m_{i \rightarrow j}} m_{i \rightarrow j}}{\sum_{j=1}^n m_{i \rightarrow j}}, \quad j \neq i. \quad (1)$$

292
 293 where $m_{i \rightarrow j}$ is the per generation migration estimate from population i into population j
 294 determined from the IM model, $\overline{m_{i \rightarrow j}}$ is the mean per generation migration rate over all
 295 comparisons including population i , and n is the number of populations considered. Although we
 296 recognize the simplistic nature of our conversion, which likely fails to consider how migration
 297 rates among all populations can interact, it nevertheless makes some concessions for the uneven
 298 distribution of migrants to the different populations and generates per generation migration rates
 299 qualitatively comparable to those generated using contemporary methods as implemented in

300 BayesAss 3.0. The advantage of using IM models, however, is that determined parameters are
301 estimated in the coalescent, or over the time frame since populations split (Hey 2010).

302

303 *Tests of hybrid origin*

304 Because four of the recovered populations within Switzerland corresponded to the MCL, we
305 tested whether the remaining three populations were of hybrid origin among them. First, the
306 genetic variance-covariance matrix (VCV) of MCL populations, likely representing parental
307 lines, are expected to be less variable and more constrained relative to those of putative hybrid
308 populations (Grant 1994; Steppan *et al.* 2002; Jones *et al.* 2003; Eroukhmanoff & Svensson
309 2011; Seehausen *et al.* 2014). To test this we performed a principal coordinates analysis (PCoA)
310 in GenAlEx on the genetic distances calculated among all individuals. Resulting individual scores
311 along the first two PCo axes were plotted by population in common genotypic space and the area
312 and eccentricity of population-specific 95% confidence ellipses was estimated. The area of the
313 ellipse surrounding a population outlines its genetic variance, while ellipse eccentricity reflects
314 the degree of constraint applied to this variance (Steppan *et al.* 2002; Jones *et al.* 2003;
315 Eroukhmanoff & Svensson 2011; Seehausen *et al.* 2014). High eccentricities (i.e., $\epsilon \sim 1$) indicate
316 high covariance in genetic signals among loci and thus narrow genetic trajectories, while low
317 eccentricities ($\epsilon \sim 0$; i.e., a more rounded ellipses) imply less genetic covariance among loci and
318 thus fewer genetic constraints (Steppan *et al.* 2002; Jones *et al.* 2003; Eroukhmanoff & Svensson
319 2011). PCoAs were also conducted on each Swiss population separately to recover eccentricities
320 in global genotypic space unconstrained by the variance of other populations. Population-
321 specific ellipse construction and determination of area and eccentricities were performed in R
322 3.1.2 (R Core Development Team 2014)

323
324 Next, we tested whether the genetic composition of the three putative hybrid populations was of
325 some combination among all MCLs, and whether their admixture proportions was predictable by
326 their spatial arrangement among and/or geographic proximities to MCLs. Alternatively, these
327 populations could trace their ancestries to other lineages outside Switzerland, in which case our
328 predictions would not apply. To test this we simulated an independent hybrid scenario where the
329 genotypes of 50 individuals at 17 loci in 3 populations were generated using EASYPOP 2.0.1
330 (Balloux 2001). Simulations assumed random mating among diploid individuals with equal
331 proportion of both sexes and where all loci were assumed to evolve at similar rates and following
332 a similar evolutionary model ($\mu = 1 \times 10^{-4}$, combined 85% stepwise mutation, 15% infinite
333 allele models). The number of alleles at each locus was set using levels found in Swiss
334 populations. Simulated populations were connected through a strict island model with relatively
335 low migration rates (0.01 migrants per generation) and allowed to interact for 140 generations.
336 Resulting populations were considered representative of the MCLs and used to generate 3
337 additional but different hybrid populations (of equal size) using Hybridlab 1.0 (Nielsen *et al.*
338 2006). The hybrids reflected the anticipated mix among simulated MCLs with the last cross (last
339 population added to the mix) exerting the strongest influence. A list of expected hybrids among
340 simulated MCLs is available (Table S3). Shortest waterway distances (SWD) between each
341 population pairs was also calculated using Google Earth (Google Inc. MountainView CA, USA)
342 measuring distances between the closest sampling locations between populations (see Figs. 1 and
343 2). In situations where populations were not connected by waterways, shortest overland distances
344 (max < 1 km) between connecting waterways were incorporated in SWD estimates. Both

345 linearized F_{ST} and D_{Jost} estimates of genetic differentiation were compared to log transformed
346 SWDs (to account for multiple dispersal directions and dimensions; Rousset 1997) and to
347 expected genetic differentiation within a hybrid scenario by linear regression analyses supported
348 by 10 000 Mantel randomizations. The combined effects of both SWD and the hybrid scenario
349 were also tested (Revell 2012). Changes to the Akaike information criteria (corrected for small
350 sample sizes; ΔAIC_c) were used to determine the model that best explained genetic
351 differentiation among populations. Mantel regressions were performed in R, where the
352 multivariate versions used the phytools package (Revell 2012).

353

354 Finally, we determined whether the genotypes of the putative hybrid populations were consistent
355 with possible combinations of genotypes found in the MCLs, and whether or not they were
356 consistent with a hybrid swarm. We first used all individuals assigned to the MCL populations
357 by STRUCTURAMA/STRUCTURE and tested how successfully they reassigned to their
358 respective populations using exclusion-based assignments in GeneClass2.0 (Rannala & Mountain
359 1997; Piry *et al.* 2004). Individuals were treated as unknowns and either excluded ($P < 0.05$) or
360 considered likely residents of populations using 1 000 000 simulated individuals calculated as
361 per Paetkau *et al.* 2004 (i.e., assuming random mating and based on observed genotypic
362 frequencies within populations). Here, resident/reassignment is defined as the failure to be
363 excluded from a population ($P > 0.05$)—that is, an individual cannot be excluded from a
364 population at the 95% level. The successful reassignment of MCL individuals as residents to
365 their respective populations implies that these make good reference populations useful for
366 excluding individuals of unknown origin (Piry *et al.* 2004; Taylor *et al.* 2006). Next, actual MCL
367 populations were used to generate 50 individuals of various hybrid classes among them including

368 F1s (F1), F1-backcrosses (F1B), F2s (F2), and complex F2s and F2 backcrosses combining all
369 three MCLs (F2C). In all, 17 different hybrid classes were generated from the MCL populations
370 using Hybridlab (Table S4). We then used the MCL populations and the different hybrid classes
371 as reference populations to assign all individuals from the three putative hybrid populations
372 using the same exclusion method described above with the same parameters. Individuals that
373 cannot be excluded entirely from various hybrid classes support a hybrid origin of these
374 populations while assignments to complex F2 hybrids and backcrosses is consistent with an
375 origin from within a hybrid swarm combining more than two lineages. We also included
376 individuals collected from the COR and NGG locations as controls to test whether individuals
377 tracing their ancestry outside the MCLs would be excluded from them and their simulated
378 hybrids.

379

380 **Results**

381 *Population genetic structure*

382 The most probable number of genetic populations recovered from unsupervised
383 STRUCTURAMA searches considering the entire dataset, was six (Table 1, Fig. S1). Using
384 STRUCTURE to visualize this result showed that most individuals could be assigned to one of
385 these populations with high certainty, with only 5% of individuals assigned to their most
386 probable population with less than 60% probability (32/634) (Fig. 2a). Recovered genetic
387 clusters did not correspond to river drainages, lake systems or sampling sites but rather grouped
388 several sites and certain lake systems, some within different drainages, into the same genetic
389 population (Fig. 2a). One population in particular spanned two different drainages (i.e., the
390 Rhône and the Aare; Orange cluster). Populations at the base of the hierarchy showed some

391 association with colonizing maternal lineages in different areas (Figs. 1 and 2a). Individuals
392 collected from ALL, STS, GLA, GUP, YVB, YVM and WBB showed genetic affiliation with
393 mtDNA lineages found in the Rhône (hereafter Rhône). Individuals collected from MOE, in the
394 upper Rhine, showed genetic affiliation with the purported native Swiss lineage (hereafter MOE),
395 while those collected from GIP, CLA and CUP (hereafter Rhine) showed affiliation with the
396 eastern European lineage present in the lower Rhine (Fig. 2a). Individuals collected from the
397 Lakes Biel/Wohlen region (MOR, GOL, WOH, EYM, GAE, and CHR) formed a genetically
398 distinct population (hereafter WOH; Figs. 1 and 2a). The individuals collected in Corsica and
399 northern Germany also formed genetically distinct populations (hereafter COR and NGG), but
400 we also recognize some level of uncertainty in assignment present among all recovered
401 populations likely reflecting allele sharing due to incomplete lineage sorting and/or admixture
402 (Fig. 2a).

403
404 Subsequent STRUCTURAMA analyses performed on all six populations showed variable levels
405 of internal sub-structure. Whereas neither WOH nor COR showed further sub-division, the
406 Rhône, MOE, Rhine, and NGG populations showed additional structure (Table S5). Assignments
407 of individuals within respective populations as determined in STRUCTURE, largely confirmed
408 STRUCTURAMA results (Fig. 2b-g). In the Rhône population, assignments predominantly
409 grouped individuals collected from Lake Geneva, its tributaries and those at WBB into a
410 population (hereafter RHO) separate from another population (hereafter NEU) made up of
411 individuals mostly collected in Lake Neuchâtel but also present in Lake Geneva and its
412 tributaries (Table S5; Fig. 2b). This likely reflects the higher and more consistent levels of
413 admixture of NEU individuals, with some genetic similarities with individuals in the Rhine and

414 in the distant NGG populations (Fig. 2a-b). More importantly however, this also implies the
415 sympatric coexistence of two genetically distinguishable populations within the Lakes Geneva/
416 Neuchâtel systems. Additional testing performed on either RHO and NEU revealed no further
417 structure within them. Assignments in the Rhine population separated individuals collected from
418 GIP from those collected in the Lake Constance area (CLA and CUP) (Fig. 2e), likely reflecting
419 the higher admixture levels observed between MOE and GIP (Fig. 2a and e). No further structure
420 was recovered in GIP but additional tests on the Lake Constance area samples recovered two
421 additional populations; one associated with the lake (CLA) and another associated with its
422 upstream tributary (CUP), with substantial admixture between them (Fig. 2e). No further sub-
423 structure was evident in the CUP population but the CLA population exhibited still further
424 structure (Table S5), which was generated from the even split of individual genotypes rather than
425 subdivision among individuals (Fig. 2e). Such results are not indicative of population structure
426 but rather likely indicate the programs inability to distinguish between genotypes at sites with
427 low genetic differentiation (i.e., low F_{ST} ; Pritchard *et al.* 2000; Falush *et al.* 2007; Hubisz *et al.*
428 2009). Similarly, although STRUCTURAMA indicated substantial internal genetic structure in
429 MOE and NGG populations (Table S5), more detailed individual assignments tests showed both
430 cases were examples of genotype splitting (Fig. 2d and f). The overall hierarchical search for
431 population structure therefore, recovered nine genetically distinguishable populations among the
432 634 sampled individuals. Of these, two were outside of Switzerland (COR and NGG), four were
433 consistent with the main colonizing lineages (RHO, MOE and CLA-CUP), and the last three
434 (NEU, WOH and GIP), although genetically distinguishable by microsatellite allele frequencies,
435 exhibited various mtDNA haplotypes (Figs. 1 and 2).

436

437 *Neutrality tests*

438 None of the markers used to recover population genetic structure at the different hierarchical
439 levels showed evidence of selection using the FDIST algorithm as applied in LOSITAN,
440 regardless of the applied mutational model (Fig. S2). Similarly, selection tests using Bayescan
441 2.1 also failed to detect signs of selection in any used markers (Fig. S2). These results indicate
442 that neutral processes largely governed allele frequencies and population genetic differentiation
443 at the markers used.

444

445 *Population genetic statistics*

446 Descriptive statistics of genetic diversity over the nine populations and 17 loci are available
447 (Table S6). No evidence of linkage disequilibrium was detected between any pair of loci ($p >$
448 0.05). Eight population-loci combinations deviated from genotypic frequencies expected under
449 HWE, out of a possible 153 comparisons, a number very close to that expected by chance ($n =$
450 7.65). None of these deviations involved the same locus in different populations consistent with
451 their random nature. The 17 loci showed variable levels of polymorphism in the different
452 populations. The allelic richness (A_R) ranged between 1.00 and 9.80 with a mean of 3.24, and the
453 number of private alleles (Pa) ranged from 0.00 to 1.29 with a mean of 0.38, over all populations
454 and loci. Large and significant levels of genetic differentiation estimated as F_{ST} and D_{Jost} were
455 detected among all possible pairwise population comparisons, indicating strong support for
456 genetic differences among them (Table S7). These differences were generally greater among
457 populations reflecting the MCLs. No significant differences were found in population genetic
458 diversity indices or global F_{ST} estimated using putatively QTL linked versus unlinked loci ($W \geq$
459 25, $p \geq 0.216$), consistent with marker neutrality. No significant differences were observed in

460 genetic diversity indices among the MCL populations versus those exhibiting mixed
461 mitochondrial lineages ($t \leq 2.00$, $d.f.$ range = 3.01-4.95, $p \geq 0.164$).

462

463 *Population sizes and connectivity*

464 All nine recovered populations exhibited comparable contemporary \widehat{N}_e except WOH and COR,
465 which had estimates near an order of magnitude greater (Fig. 3). The WOH population was by
466 far the largest within Switzerland while CLA was the smallest. These results were similar when
467 considering a greater minimum allele frequency of 0.02, except that the estimates for COR
468 became indeterminate (Fig. S3). Only three populations were connected by contemporary
469 migration rates greater than 0.01 (Fig. 3). These higher migration rates showed high
470 unidirectional migration from CUP to CLA and more restricted unidirectional migration from
471 CUP to GIP. Thus, CUP acts as a source population to both GIP and CLA. All other populations
472 appear contemporarily isolated. To eliminate the possibility that low contemporary migration
473 rates are an artifact of the way we grouped individuals within populations (i.e., by assignment
474 probability), we also estimates them using individuals grouped by sample location. Here,
475 individuals were assigned to populations based on the predominant genetic cluster recovered at
476 each site. Contemporary migrations rates produced in this way were nearly identical except that
477 we also recovered some low migration (0.014) from RHO into NEU (see Fig S4).

478

479 Coalescent-based N_e estimates tended to be smaller and less variable than contemporary ones
480 ($\sigma_{\text{contemporary}} = 490.5$, $\sigma_{\text{coalescent}} = 137.4$) and showed that most populations were of comparable
481 size (Fig. 4). Unlike estimates of contemporary gene flow, coalescent based per generation

482 migration rates showed extensive ($\gg 0.01$) multidirectional gene flow among populations within
483 Switzerland (Fig. 4). Notably, most Swiss populations consistent with MCLs (i.e., RHO, MOE,
484 CLA and CUP) tended to export more and import fewer migrants than did the populations of
485 putative hybrid origins (NEU, WOH, GIP). We found no indications of historical gene flow
486 between any Swiss population and the Corsican one, and the possibility of low historical gene
487 flow between a single Swiss population (RHO) and the North German one.

488

489 *Tests of hybrid origin*

490 PCo analyses performed on the genetic distances among individuals collected within Switzerland
491 showed distinct clustering of individuals belonging to the seven Swiss populations with variable
492 degrees of overlap (Fig. 5). MCL populations tended to occupy the periphery of the genotypic
493 space outlined by the first 2 PCo axes (accounting for nearly 70% of the genetic variation among
494 individuals), while the remaining three populations (NEU, WHO, GIP) were encompassed
495 entirely within the range defined by the MCL populations. The area of the 95% confidence
496 ellipses calculated for the MCL populations were significantly smaller than those calculated for
497 the remaining three consistent with greater genetic variation in the latter group and with their
498 hybrid origin ($t = 3.391$, $d.f. = 4.16$, $p = 0.013$). The ellipses of the three remaining populations
499 were also less eccentric relative to those of the MCLs when compared both in common ($t =$
500 3.883 , $d.f. = 2.03$, $p = 0.029$) and global ($t = 2.231$, $d.f. = 4.01$, $p = 0.047$) genotypic spaces,
501 consistent with relaxed genetic constraints and increased evolutionary potential expected in
502 hybrids.

503

504 Results of the AIC_c model comparisons of F_{ST} and D_{JOST} based Mantel regressions showed
505 similar results (Table 2). In both cases, the most likely model explaining genetic differentiation
506 among Swiss populations was one based solely on the hybrid scenario, while that using shortest
507 waterway distances exclusively, or in combination with the hybrid scenario were less likely
508 and/or not significant (Table 2). These results imply the uneven and variable contribution of the
509 different MCLs to the various possible hybrid populations, and that this contribution is more
510 likely related to the spatial arrangement of the MCLs within Switzerland, rather than to the strict
511 distances between them.

512
513 Nearly 90% of individuals from each MCL population could not be excluded from their
514 respective population at the 0.05 level (Fig. 6). In all cases, only exclusion errors were made and
515 no individual was incorrectly reassigned to one of the other MCL populations, indicating that the
516 MCLs were suitable reference populations for exclusion analyses of unknown individuals (Fig.
517 6). Using the MCLs and simulated hybrid classes in exclusion analyses performed on individuals
518 tracing their ancestry in populations located outside Switzerland (COR and NGG) showed that
519 all individuals were excluded from both the MCLs and their expected hybrid classes (Fig. 6d, e).
520 Performing the same analyses on NEU individuals, however, showed that over 25% could not be
521 excluded from the RHO population (Fig. 6f). This result is not surprising given the similarity
522 between RHO and NEU (see Figs. 2 and 5). Moreover, a substantial proportion of NEU
523 individuals could also not be excluded from possible hybrid classes with a general increase in
524 assignment probabilities as the hybrid class complexity increased (Fig. 6f). Similar exclusion
525 tests performed on WOH and GIP showed that all individuals were excluded from all MCL
526 populations (Fig. 6g and h). On the other hand, a substantial proportion of both WOH and GIP

527 individuals could not be excluded from possible hybrid classes, and the same general pattern of
528 increasing assignment probabilities with increasing hybrid complexity was observed.

529

530 **Discussion**

531 Here, we show that the recent range expansion of threespine stickleback in Switzerland is
532 associated with the formation of a hybrid ‘superswarm’ among three distinct lineages that
533 colonized Switzerland about 140 years ago (Heller 1870; Fatio 1882; Bertin 1925; Lucek *et al.*
534 2010). This massive hybridization likely gave rise to three genetically distinguishable novel
535 populations. We demonstrate that current populations are genetically stable and all but the most
536 closely related ones are nearly isolated with low levels of contemporary gene flow. Coalescent-
537 based analyses on the same populations, however, show clear connectivity with extensive
538 multidirectional gene flow among them in the past. If our inferences are correct, backcrossing to
539 the source populations is less than expected from geographical distances, and migration between
540 areas that currently host genetically differentiated populations of hybrid origin is lower now than
541 it was during colonization. Thus, secondary contact among three distant lineages during the
542 colonization of Swiss waterways initially led to formation of a hybrid ‘superswarm’, followed by
543 stabilization of genetically differentiated populations. Whether or not this hybridization among
544 main colonizing lineages and stabilization of hybrid populations has facilitated ecological range
545 expansion into various habitats remains to be determined but appears to be the case (Lucek *et al.*
546 2010; Lucek *et al.* 2014)

547

548 *Population Structure*

549 We recovered seven Swiss stickleback populations from our hierarchical analyses. The
550 population structure determined here differs from many previous population based stickleback
551 studies. Rather than assigning population status to different sampling sites by default, we used an
552 approach based on individual admixture proportions. Although both methods are effective, they
553 are useful in addressing different hypotheses. In the context of reconstructing a biological
554 invasion from the multiple introductions of distantly related lineages, a more quantitative based
555 approach using a population genetics framework (i.e., individuals assigned to population in
556 HWE with low linkage among loci) may be more appropriate (Darling *et al.* 2008).

557
558 The recovered population structure groups several geographically distant locations together
559 within the same genetic population, irrespective of habitat type. This indicates substantially
560 greater gene flow among sampling locations and habitat types within recovered genetic
561 populations relative to that between them. On the other hand, our analysis also assigns
562 individuals within single sampling sites into two genetically distinguishable groups, suggesting
563 that distinct stickleback populations coexist at some sites in the Lakes Neuchâtel and Geneva
564 systems, and that the development of these populations is relatively recent.

565
566 The population structure recovered here cannot be explained by local adaptation but rather
567 reflects structure imposed by drift and gene flow. First, two outlier loci detection approaches
568 (LOSITAN-FDIST and Bayescan) found no evidence of diversifying or balancing selection at
569 any loci. Second, even though some of our markers were shown to be linked to known QTLs in
570 studies of other stickleback populations (Peichel *et al.* 2001; Mäkinen *et al.* 2008), these loci did
571 not behave differently from neutral markers.

572
573 *Population Connectivity and Size*
574 Extensive contemporary gene flow among populations would likely result in violations of HWE
575 and/or LD among loci within populations greater than expected by chance alone (e.g.,
576 heterozygote deficiencies). This could result in Wahlund effects within populations or in signs of
577 recombination or epistatic linkage among loci (Slatkin 2008; Excoffier & Lischer 2010). Without
578 exception, however, no departures from HWE or evidence of excessive LD are evident in our
579 populations. Moreover, our populations are significantly differentiated, often showing high
580 F_{ST}/D_{Jost} indices, with next to no contemporary gene flow among them. The only contemporary
581 gene flow observed occurs in a unidirectional manner from CUP into both CLA and GIP. These
582 results are in accordance with previous work showing substantial gene flow among stickleback
583 collected from stream and lake locations within the Lake Constance region (Berner *et al.* 2010;
584 Moser *et al.* 2012; Lucek *et al.* 2013; Lucek *et al.* 2014) and between Constance region
585 stickleback and those in the upper Rhine (i.e., GIP; Lucek *et al.* 2010). Lucek *et al.* 2014 suggest
586 that stickleback from the Constance region are becoming locally adapted with decreasing gene
587 flow between lake and stream populations. So, the gene flow observed between CLA and CUP is
588 likely occurring in primary contact between diverging stream and lake ecotypes that originated
589 within the past 140 years from a common gene pool. Coalescent-based analyses support the gene
590 flow reduction in the Constance region in particular, but also more generally throughout
591 Switzerland. IM based coalescent analyses show extensive multidirectional gene flow among
592 most Swiss populations and recovers much larger migration estimates than contemporary ones.
593 The differences between estimated per generation migration rates are likely due to methods for
594 assessing contemporary gene flow only taking current allele frequencies into account and thus

595 only resolving recent migration among populations (Wilson & Rannala 2003; Piry *et al.* 2004).
596 Coalescent-based analyses as implemented in IMA2, instead, estimate migration rates over the
597 divergence time between and among considered taxa (Hey & Nielsen 2004; Hey 2010; Strasburg
598 & Rieseberg 2010). The latter are essentially averages over the coalescent and do not make
599 concessions for migration rates that may be temporally dynamic. Thus, coalescent-based
600 migration rate estimates can be quite different from those using contemporary methods, which
601 reflect more current population connectivity. Here, we combined both approaches allowing us to
602 conclude that, although gene flow among Swiss populations was likely extensive in the past, it
603 has been substantially reduced relatively recently. Coalescent-based estimates show that
604 populations corresponding to the three MCLs (RHO, MOE, CLA/CUP) exhibit much larger
605 outgoing than incoming migration rates while the opposite pattern holds for the remaining three
606 populations (NEU, WOH and GIP). These findings suggest that the MCLs, geographically
607 restricted to the northeast, northwest and far west parts of Switzerland, acted as genetic sources
608 seeding other populations that subsequently expanded across the Swiss midlands and now show
609 variable levels of complex admixture among MCLs.

610

611 *Hybrid superswarm*

612 Given the high level of gene flow that the putative hybrid populations (NEU, WOH and GIP)
613 received from the MCLs in the past, a plausible scenario for their origin is genetic admixture
614 among the MCLs. As expected, the putative hybrid populations occupy intermediate and less
615 constrained (more variable) genotypic space than the MCLs, consistent with the breakdown and
616 reshuffling of genetic constraints established in parental lineages (Buerkle *et al.* 2000; Mallet
617 2007; Schluter & Conte 2009; Abbott *et al.* 2013; Seehausen *et al.* 2014). Assignment tests also

618 showed improving assignments of individuals in hybrid populations to increasingly complex
619 simulated hybrid classes. Exclusion-based assignments allow individuals to remain unclassified
620 if their genotype is too dissimilar from the reference populations (Paetkau *et al.* 2004; Piry *et al.*
621 2004). Consequently, finding an increasing number of individuals assigned to increasingly
622 complex hybrid classes implicates admixture among all three MCLs in the formation of these
623 three populations. It is important to note that while assignment to hybrid classes may be
624 relatively low, we tested only 17 of a diverse array of hybrid classes potentially produced by the
625 MCLs and included only formative F1s and F2s and their backcrosses. Consequently, tests
626 including more complex hybrid classes may find greater hybrid assignment. Moreover, relatively
627 low assignment rates may also reflect past hybridization with ensuing decreasing gene flow,
628 genetic stabilization and recombination within newly established populations possibly eroding
629 more obvious hybridization signals (Currat *et al.* 2008; Seehausen *et al.* 2008). This is supported
630 by the NEU population, which is the least differentiated among the hybrid populations showing
631 the highest hybrid assignments. This may indicate that, all else being equal, and in light of the
632 limited contemporary gene flow (see above), the NEU population is the most recently formed
633 hybrid. On the other hand, NEU is also the only hybrid population sympatrically distributed in
634 many sites with the RHO MCL population. Consequently, its greater assignments to hybrid
635 classes may also be related to its continued physical contact with a seeding MCL versus GIP and
636 WOH who are currently entirely allopatric from all MCLs as determined here.

637

638 The hybrid origin of NEU, WOH and GIP is also consistent with modeling results showing the
639 best model explaining genetic differentiation among populations is one explicitly assigning
640 intermediate genetic makeup to putative hybrid populations relative to simulated MCLs.

641 However, we found no relationship between genetic differentiation and distance either in
642 combination with the hybrid scenario or by itself. These results contrast those of Lucek *et al.*
643 2013 who showed significant isolation by distance (IBD) and by adaptation (IBA; based on
644 phenotypic dissimilarity) contributing to extensive genetic differences observed in stickleback
645 within different Swiss lake systems. Lucek *et al.* (2013)'s patterns were likely the result of
646 parallel adaptive differentiation of populations into lake and stream ecotypes contributing to
647 among population divergence. Lucek *et al.* (2013) suggested that increasing local adaptation is
648 associated with gene flow reduction and increased reproductive isolation among different
649 sampling location within lake systems. Because our current study and that of Lucek *et al.* (2013)
650 did not use the same population units (Lucek *et al.* 2013-location based; here-genetic cluster
651 based), it is difficult to determine whether IBA also contributes to the differentiation of the
652 populations recovered here. The proximity of different habitats to one another, however,
653 suggests that this is not a factor as many adjacent populations, although highly genetically
654 divergent, occur in similar habitats not separated by habitat transitions (e.g., GIP-MOE, both
655 stream habitats) while other populations occur in the same location and hence occupy the same
656 habitat (e.g., both RHO and NEU individuals recovered from STS, ALL, GLA, GUP, YVB and
657 YVM). Thus, although parallel habitat based divergence seems evident at a finer, more lake-
658 specific level, the nature of genetic divergence between the geographically more inclusive
659 populations identified in the present work is less obvious. Clarifying the causes of among
660 population divergence and the reduction/cessation of gene flow among genetic populations
661 recovered here, and the mechanisms of their local coexistence in several sites, is a logical next
662 step for future work. Irrespective of the mechanisms, the hybrid origins of NEU, WOH and GIP
663 populations is consistent with previous reports implicating hybridization as an important driver

664 of population divergence in some regions of Switzerland (Lucek *et al.* 2010) and to successful
665 invasions more generally (Lockwood *et al.* 2007; Prentis *et al.* 2008; Lack *et al.* 2012; Parepa *et*
666 *al.* 2014; Williams *et al.* 2014). An important distinction from many previous reports, however,
667 is that we show evidence of three populations originating from a hybrid ‘superswarm’ involving
668 complex crosses and backcrosses among more than two distant lineages. Whether this
669 ‘superswarm’ was the result of one major hybridization event or established from several
670 pairwise hybridizations in stages is difficult to determine from current analyses. Future work
671 using many more incorporated into more complex isolation with migration (IM) analyses could
672 better estimate coalescent based demographic parameters (i.e., m and N_e) and better resolve the
673 timing and number of hybridization events as well as testing the inferred recent cessations of
674 gene flow (Hey 2010).

675

676 **Conclusion**

677 Our findings supports the formation stickleback hybrid populations that have contributed to the
678 extensive genetic and likely phenotypic (Lucek *et al.* 2010; Lucek *et al.* 2013) diversity observed
679 within Switzerland. This is consistent with secondary contact among distant lineages converting
680 interpopulation genetic diversity into intrapopulation genetic variation by hybridization
681 (Lockwood *et al.* 2007; Dlugosch & Parker 2008; Prentis *et al.* 2008; Seehausen *et al.* 2008). We
682 show that this process can occur between more than just two distant lineages, likely providing
683 extensive standing genetic variation from which several newly formed genetic combination can
684 emerge to establish viable populations expressing decreasing levels of gene flow over time. Here,
685 three new populations of hybridogenic origin have likely emerged within Swiss inland waters in
686 the span of 140 years. This work is thus consistent with a growing body of work implicating

687 range expansion and hybridization as potent drivers of new populations, potentially leading to
688 speciation (Mallet 2007; Nolte & Tautz 2010; Abbott *et al.* 2013; Seehausen *et al.* 2014) and as a
689 likely catalyst for adaptive radiations over very short time scales.

690

691 **Acknowledgments**

692 Sampling help was provided by the Fish Ecology group at the EAWAG, and in particular by
693 Pascal Vonlanthen, Guy Périat, Alan Hudson and Isabel Magalhaes. We thank the Swiss
694 Cantonal authorities of Aargau, Basel Land, Bern, St.Gallen, Thurgau, Valais and Vaud as well
695 as the Corsican fishery authorities for collection permits. The EAWAG Action Field Grant
696 ‘AquaDiverse’ and The Swiss National Science Foundation Grants to OS funded this work.

697

698 **Data accessibility**

699 The raw genotypes for all individuals used in this study are stored and accessible through
700 Labarchives.com and can be accessed at the following link: <http://dx.dio.org/> (provided when
701 available)

702

703 **References**

- 704 Abbott R, Albach D, Ansell S, *et al.* (2013) Hybridization and speciation. *J. Evol. Biol.* **26**, 229-
705 246.
- 706 Antao T, Lopes A, Lopes RJ, Beja-Pereira A, Luikart G (2008) LOSITAN: A workbench to
707 detect molecular adaptation based on a Fst-outlier method. *BMC Bioinformatics* **9**, 1-5.
- 708 Balloux F (2001) EASYPOP (Version 1.7): A Computer Program for Population Genetics
709 Simulations. *J. Heredity* **92**, 301-302.
- 710 Berner D, Grandchamp AC, Hendry AP (2009) Variable progress toward ecological speciation
711 in parapatry: Sticklebacks across eight lake-stream transitions. *Evolution* **63**, 1740-1753.
- 712 Berner D, Roesti M, Hendry AP, Salzburger W (2010) Constraints on speciation suggested by
713 comparing lake-stream stickleback divergence across two continents. *Mol. Ecol.* **19**,
714 4963-4978.

- 715 Bertin L (1925) Recherches bionomiques, biométriques et systématiques sur les épinoches
716 (*Gastérostéidés*). *Ann. Inst. Océano.* **II**, 205.
- 717 Bradbury IR, Hubert S, Higgins B, *et al.* (2013) Genomic islands of divergence and their
718 consequences for the resolution of spatial structure in an exploited marine fish. *Evol Appl*
719 **6**, 450-461.
- 720 Buerkle CA, Morris RJ, Asmussen MA, Rieseberg LH (2000) The likelihood of homoploid
721 hybrid speciation. *Heredity* **84**, 441-451.
- 722 Caldera EJ, Bolnick DI (2008) Effects of colonization history and landscape structure on genetic
723 variation within and among threespine stickleback (*Gasterosteus aculeatus*) populations
724 in a single watershed. *Evol. Ecol. Res.* **10**, 575-598.
- 725 Currat M, Ruedi M, Petit RJ, Excoffier L (2008) The hidden side of invasions: Massive
726 introgression by local genes. *Evolution* **62**, 1908-1920.
- 727 Darling JA, Bagley MJ, Roman J, Tepolt CK, Geller JB (2008) Genetic patterns across multiple
728 introductions of the globally invasive crab genus *Carcinus*. *Mol. Ecol.* **17**, 4992-5007.
- 729 Deagle BE, Jones FC, Chan YF, *et al.* (2012) Population genomics of parallel phenotypic
730 evolution in stickleback across stream-lake ecological transitions. *Proc. R. Soc. B-Biol.*
731 *Sci.* **279**, 1277-1286.
- 732 Dieringer D, Schlötterer C (2003) Microsatellite Analyser (MSA): a platform independent
733 analysis tool for large microsatellite data sets. *Mol. Ecol. Notes* **3**, 167-169.
- 734 Dlugosch KM, Parker IM (2008) Founding events in species invasions: genetic variation,
735 adaptive evolution, and the role of multiple introductions. *Mol. Ecol.* **17**, 431-449.
- 736 Do C, Waples RS, Peel D, *et al.* (2014) NeEstimator v2: re-implementation of software for the
737 estimation of contemporary effective population size (Ne) from genetic data. *Mol. Ecol.*
738 *Res.* **14**, 209-214.
- 739 Eroukhmanoff F, Svensson EI (2011) Evolution and stability of the G-matrix during the
740 colonization of a novel environment. *J. Evol. Biol.* **24**, 1363-1373.
- 741 Excoffier L, Lischer HEL (2010) Arlequin suite ver 3.5: a new series of programs to perform
742 population genetics analyses under Linux and Windows. *Mol. Ecol. Res.* **10**, 564-567.
- 743 Falush D, Stephens M, Pritchard J (2007) Inference of population structure using multilocus
744 genotype data: dominant markers and null alleles. *Mol. Ecol. Notes* **7**, 574-578.
- 745 Fatio V (1882) *Faune des vertébrés de la Suisse*, 1st edn. H. Georg, Genève.
- 746 Foll M, Gaggiotti O (2008) A Genome-Scan method to identify selected loci appropriate for both
747 dominant and codominant markers: A Bayesian perspective. *Genetics* **180**, 977-993.
- 748 Gerlach G, Jueterbock A, Kraemer P, Deppermann J, Harmand P (2010) Calculations of
749 population differentiation based on GST and D: forget GST but not all of statistics! *Mol.*
750 *Ecol.* **19**, 3845-3852.
- 751 Goudet J (1995) FSTAT (Version 1.2): A Computer Program to Calculate F-Statistics. *J.*
752 *Heredity* **86**, 485-486.
- 753 Grant PR (1994) Population Variation and Hybridization - Comparison of Finches from 2
754 Archipelagos. *Evolutionary Ecology* **8**, 598-617.
- 755 Hare M, Nunney L, Schwartz MK, *et al.* (2011) Understanding and estimating effective
756 population size for practical application in marine species management. *Conserv. Biol.*
- 757 Heller C (1870) Die Fishes Tirols und Vorarlbergs. *Z. Ferdinandeums Tirol* **5**, 295-369.
- 758 Hey J (2010) Isolation with Migration Models for More Than Two Populations. *Mol. Biol. Evol.*
759 **27**, 905-920.

- 760 Hey J, Nielsen R (2004) Multilocus methods for estimating population sizes, migration rates and
761 divergence time, with applications to the divergence of *Drosophila pseudoobscura* and *D-*
762 *persimilis*. *Genetics* **167**, 747-760.
- 763 Hubisz MJ, Falush D, Stephens M, Pritchard JK (2009) Inferring weak population structure with
764 the assistance of sample group information. *Mol. Ecol. Res.* **9**, 1322-1332.
- 765 Huelsenbeck JP, Andolfatto P (2007) Inference of population structure under a Dirichlet process
766 model. *Genetics* **175**, 1787-1802.
- 767 Huelsenbeck JP, Andolfatto P, Huelsenbeck ET (2011) Structurama: Bayesian inference of
768 population structure. *Evol. Bioinf.* **7**, 55.
- 769 Jakobsdóttir KB, Pardoe H, Magnússon Á, *et al.* (2011) Historical changes in genotypic
770 frequencies at the Pantophysin locus in Atlantic cod (*Gadus morhua*) in Icelandic waters:
771 evidence of fisheries-induced selection? *Evol. Appl.* **4**, 562-573.
- 772 Jakobsson M, Rosenberg NA (2007) CLUMPP: a cluster matching and permutation program for
773 dealing with label switching and multimodality in analysis of population structure.
774 *Bioinformatics* **23**, 1801-1806.
- 775 Jones AG, Arnold SJ, Borger R (2003) Stability of the G-matrix in a population experiencing
776 pleiotropic mutation, stabilizing selection, and genetic drift. *Evolution* **57**, 1747-1760.
- 777 Jones FC, Grabherr MG, Chan YF, *et al.* (2012) The genomic basis of adaptive evolution in
778 threespine sticklebacks. *Nature* **484**, 55-61.
- 779 Kaeuffer R, Peichel CL, Bolnick DI, Hendry AP (2012) Parallel and Nonparallel Aspects of
780 Ecological, Phenotypic, and Genetic Divergence across Replicate Population Pairs of
781 Lake and Stream Stickleback. *Evolution* **66**, 402-418.
- 782 Lack JB, Greene DU, Conroy CJ, *et al.* (2012) Invasion facilitates hybridization with
783 introgression in the *Rattus rattus* species complex. *Mol. Ecol.* **21**, 3545-3561.
- 784 Lockwood JL, Hoopes M, Marchetti MP (2007) *Invasion Ecology* Blackwell Publishing.
- 785 Lucek K, Roy D, Bezault E, Sivasundar A, Seehausen O (2010) Hybridization between distant
786 lineages increases adaptive variation during a biological invasion: stickleback in
787 Switzerland. *Mol. Ecol.* **19**, 3995-4011.
- 788 Lucek K, Sivasundar A, Roy D, Seehausen O (2013) Repeated and predictable patterns of
789 ecotypic differentiation during a biological invasion: lake-stream divergence in
790 parapatric Swiss stickleback. *J Evol Biol* **26**, 2691-2709.
- 791 Lucek K, Sivasundar A, Seehausen O (2014) Disentangling the role of phenotypic plasticity and
792 genetic divergence in contemporary ecotype formation during a biological invasion.
793 *Evolution* **68**, 2619-2632.
- 794 Mäkinen HS, Cano JM, Merilä J (2006) Genetic relationships among marine and freshwater
795 populations of the European three-spined stickleback (*Gasterosteus aculeatus*) revealed
796 by microsatellites. *Mol. Ecol.* **15**, 1519-1534.
- 797 Mäkinen HS, Cano M, Merilä J (2008) Identifying footprints of directional and balancing
798 selection in marine and freshwater three-spined stickleback (*Gasterosteus aculeatus*)
799 populations. *Mol. Ecol.* **17**, 3565-3582.
- 800 Mäkinen HS, Merilä J (2008) Mitochondrial DNA phylogeography of the three-spined
801 stickleback (*Gasterosteus aculeatus*) in Europe - Evidence for multiple glacial refugia.
802 *Mol. Phylogenet. Evol.* **46**, 167-182.
- 803 Mallet J (2007) Hybrid speciation. *Nature* **446**, 279-283.

- 804 McPhail JD (1984) Ecology and evolution of sympatric sticklebacks (*Gasterosteus*):
805 morphological and genetic evidence for a species pair in Enos Lake, British Columbia.
806 *Can. J. Zool.* **62**, 1402-1408.
- 807 Moser D, Roesti M, Berner D (2012) Repeated Lake-Stream Divergence in Stickleback Life
808 History within a Central European Lake Basin. *PLoS ONE* **7**, e50620.
- 809 Munzing J (1963) Evolution of variation and distributional patterns in European populations of
810 3-spined stickleback, *Gasterosteus aculeatus*. *Evolution* **17**, 320-332.
- 811 Nielsen EE, Bach LA, Kotlicki P (2006) HYBRIDLAB (version 1.0): a program for generating
812 simulated hybrids from population samples. *Molecular Ecology Notes* **6**, 971-973.
- 813 Nolte AW, Tautz D (2010) Understanding the onset of hybrid speciation. *Trends in Genetics* **26**,
814 54-58.
- 815 Paetkau D, Slade R, Burden M, Estoup A (2004) Genetic assignment methods for the direct,
816 real-time estimation of migration rate: a simulation-based exploration of accuracy and
817 power. *Mol. Ecol.* **13**, 55-65.
- 818 Parepa M, Fischer M, Krebs C, Bossdorf O (2014) Hybridization increases invasive knotweed
819 success. *Evol. Appl.* **7**, 413-420.
- 820 Peakall ROD, Smouse PE (2006) Genalex 6: genetic analysis in Excel. Population genetic
821 software for teaching and research. *Mol. Ecol. Notes* **6**, 288-295.
- 822 Peichel CL, Nereng KS, Ohgi KA, *et al.* (2001) The genetic architecture of divergence between
823 threespine stickleback species. *Nature* **414**, 901-905.
- 824 Piry S, Alapetite A, Cornuet JM, *et al.* (2004) GENECLASS2: A software for genetic
825 assignment and first-generation migrant detection. *J. Heredity* **95**, 536-539.
- 826 Prentis PJ, Wilson JRU, Dormontt EE, Richardson DM, Lowe AJ (2008) Adaptive evolution in
827 invasive species. *Trends in Plant Science* **13**, 288-294.
- 828 Pritchard JK, Stephens M, Donnelly P (2000) Inference of Population Structure Using
829 Multilocus Genotype Data. *Genetics* **155**, 945-959.
- 830 Rannala B, Mountain JL (1997) Detecting immigration by using multilocus genotypes. *P Natl*
831 *Acad Sci USA* **94**, 9197-9201.
- 832 Ravinet M, Prodohl PA, Harrod C (2013) On Irish stickleback: morphological diversification in
833 a secondary contact zone. *Evol. Ecol. Res.* **15**, 271-294.
- 834 Reusch TBH, Wegner KM, Kalbe M (2001) Rapid genetic divergence in postglacial populations
835 of threespine stickleback (*Gasterosteus aculeatus*): the role of habitat type, drainage and
836 geographical proximity. *Mol. Ecol.* **10**, 2435-2445.
- 837 Revell LJ (2012) phytools: an R package for phylogenetic comparative biology (and other
838 things). *Methods in Ecology and Evolution* **3**, 217-223.
- 839 Rice WR (1989) Analyzing Tables of Statistical Tests. *Evolution* **43**, 223-225.
- 840 Rosenberg NA (2004) DISTRUCT: a program for the graphical display of population structure.
841 *Mol. Ecol. Notes* **4**, 137-138.
- 842 Rousset F (1997) Genetic differentiation and estimation of gene flow from F-statistics under
843 isolation by distance. *Genetics* **145**, 1219-1228.
- 844 Roy D, Hardie DC, Treble MA, Reist JD, Ruzzante DE (2014) Evidence supporting panmixia in
845 Greenland halibut (*Reinhardtius hippoglossoides*) in the Northwest Atlantic. *Can. J. Fish.*
846 *Aquat. Sci.* **71**, 763-774.
- 847 Sakai AK, Allendorf FW, Holt JS, *et al.* (2001) The population biology of invasive species.
848 *Annu. Rev. Ecol. Evol. Syst.* **32**, 305-332.

- 849 Schluter D (1993) Adaptive radiation in sticklebacks - size, shape, and habitat use efficiency.
850 *Ecology* **74**, 699-709.
- 851 Schluter D (1996) Adaptive radiation along genetic lines of least resistance. *Evolution* **50**, 1766-
852 1774.
- 853 Schluter D, Conte GL (2009) Genetics and ecological speciation. *Proc. Natl. Acad. Sci. USA* **106**,
854 9955-9962.
- 855 Seehausen O (2004) Hybridization and adaptive radiation. *Trends Ecol Evol* **19**, 198-207.
- 856 Seehausen O, Butlin RK, Keller I, *et al.* (2014) Genomics and the origin of species. *Nat Rev*
857 *Genet* **15**, 176-192.
- 858 Seehausen O, Takimoto G, Roy D, Jokela J (2008) Speciation reversal and biodiversity
859 dynamics with hybridization in changing environments. *Mol. Ecol.* **17**, 30-44.
- 860 Simberloff D (2009) The Role of Propagule Pressure in Biological Invasions. *Ann. Rev. Ecol.*
861 *Syst.* **40**, 81-102.
- 862 Slatkin M (2008) Linkage disequilibrium--understanding the evolutionary past and mapping the
863 medical future. *Nat Rev Genet* **9**, 477-485.
- 864 Stepan SJ, Phillips PC, Houle D (2002) Comparative quantitative genetics: evolution of the G
865 matrix. *Trends. Ecol. Evol.* **17**, 320-327.
- 866 Strasburg JL, Rieseberg LH (2010) How robust are "isolation with migration" analyses to
867 violations of the im model? A simulation study. *Mol Biol Evol* **27**, 297-310.
- 868 Taylor E, Boughman J, Groenenboom M, *et al.* (2006) Speciation in reverse: morphological and
869 genetic evidence of the collapse of a three-spined stickleback (*Gasterosteus aculeatus*)
870 species pair. *Mol. Ecol.* **15**, 343-355.
- 871 Thompson CE, Taylor EB, McPhail JD (1997) Parallel evolution of lake-stream pairs of
872 threespine sticklebacks (*Gasterosteus*) inferred from mitochondrial dna variation.
873 *Evolution* **51**, 1955-1965.
- 874 van Oosterhout C, Hutchinson WF, Wills DPM, Shipley P (2004) Micro-checker: software for
875 identifying and correcting genotyping errors in microsatellite data. *Mol. Ecol. Notes* **4**,
876 535-538.
- 877 Waples RS, England PR (2011) Estimating Contemporary Effective Population Size on the Basis
878 of Linkage Disequilibrium in the Face of Migration. *Genetics* **189**, 633-644.
- 879 Weir BS, Cockerham CC (1984) Estimating F-Statistics for the Analysis of Population Structure.
880 *Evolution* **38**, 1358-1370.
- 881 Williams WI, Friedman JM, Gaskin JF, Norton AP (2014) Hybridization of an invasive shrub
882 affects tolerance and resistance to defoliation by a biological control agent. *Evol. Appl.* **7**,
883 381-393.
- 884 Wilson GA, Rannala B (2003) Bayesian inference of recent migration rates using multilocus
885 genotypes. *Genetics* **163**, 1177-1191.
- 886 Yoder JB, Clancey E, Des Roches S, *et al.* (2010) Ecological opportunity and the origin of
887 adaptive radiations. *J. Evol. Biol.* **23**, 1581-1596.

888

889 Table 1. Population structure estimated in sampled stickleback determined from unsupervised searches (performed in
 890 STRUCTURAMA 2.0). *EK* values indicate Dirichlet Process Prior mean on which searches were centered. Marginal likelihood
 891 of searches indicates the likelihood of the resulting search performed using the corresponding *EK*.

<i>K</i>	<i>EK</i> (1)	<i>EK</i> (2)	<i>EK</i> (3)	<i>EK</i> (4)	<i>EK</i> (5)	<i>EK</i> (6)	<i>EK</i> (7)	<i>EK</i> (8)	<i>EK</i> (9)	<i>EK</i> (10)	<i>EK</i> (12)
<i>Over all sampled sites</i>											
1	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
2	0.33	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
3	0.67	0.67	0.33	0.33	0.00	0.33	0.33	0.00	0.00	0.00	0.00
4		0.33	0.00	0.34	0.33	0.00	0.34	0.33	0.00	0.33	0.00
5			0.67	0.33	0.67	0.67	0.34	0.67	0.00	0.34	0.33
6									1.00	0.33	0.67
7											
<i>Marginal likelihood of search</i>											
	-25898.19	-24585.50	-24584.81	-24584.19	-20386.27	-24584.80	-24584.79	-20386.95	-17734.47	-20385.77	-18819.18

892 Most likely number of recovered clusters is bolded
 893 Bolded marginal likelihood of searches indicate most robust and likely search results
 894
 895 Table 1. Concluded.

<i>K</i>	(<i>EK</i> 15)	<i>EK</i> (17)	<i>EK</i> (20)
<i>Over all sampled sites</i>			
1	0.00	0.00	0.00
2	0.00	0.00	0.00
3	0.00	0.00	0.00
4	0.00	0.00	0.00
5	0.67	0.00	0.33
6	0.33	1.00	0.67
7			0.00
<i>Marginal likelihood of search</i>			
	-19315.31	-17734.67	-18820.36

896
 897
 898

899 Table 2. Regression models explaining the genetic differentiation among Swiss stickleback populations. n = number of
 900 populations in the model, K = number of explanatory variables, R^2 = coefficient of determination, P_{ols} = ordinary least squared P
 901 value, P_m = Mantel permutations P values (10 000), and RSS = residual sum of squares. Variables in the models are: ln(SWD) =
 902 log transformed shortest waterway distance between populations and *Hybsc* = matrix of expected genetic differences under the
 903 hybrid scenario considering exNEU, exWOH and exGIP as hybrid populations originating from crosses among simulated main
 904 colonizing lineages (sRHO, sMOE, and sCON). Most likely models are bolded.

Model	n	K	R^2	P_{ols}	P_m	RSS	AIC_c	ΔAIC_c
$F_{ST} \sim \ln(\text{SWD})$	6	1	0.153	0.149	0.072	0.629	-10.534	2.025
$F_{ST} \sim \text{Hybsc}$	6	1	0.396	0.012	0.050	0.449	-12.558	0.000
$F_{ST} \sim \ln(\text{SWD}) + \text{Hybsc}$	6	2	0.400	0.047	0.086	0.446	-7.597	4.963
$D_{Jost} \sim \ln(\text{SWD})$	6	1	0.279	0.043	0.035	3.134	-0.897	2.324
$D_{Jost} \sim \text{Hybsc}$	6	1	0.511	0.003	0.013	2.127	-3.221	0.000
$D_{Jost} \sim \ln(\text{SWD}) + \text{Hybsc}$	6	2	0.537	0.010	0.015	2.015	1.453	4.674

905

906 Figure 1. Detailed view of 17 locations within Switzerland where stickleback were sampled.
 907 Main river drainages are coloured (orange = Rhône, blue = *Aare* and green = *Rhine*) and five
 908 lake systems (Geneva, Neuchâtel, Wohlen (*not shown*), Biel and Constance). Each site code
 909 corresponds to that listed in Table S1 and shows the proportion of mtDNA haplotypes
 910 determined in Lucek et al. (2010). CHR was not assessed for mtDNA. Inset map shows
 911 Switzerland's location within mainland Europe and the location of the Corsican (COR) and the
 912 North German (NGG) sampling sites.

913

914 Figure 2. Hierarchical Bayesian posterior probability assignment of sampled stickleback. (a)
 915 Initial analysis using all individuals recovered 6 genetic clusters. Subsequent analysis run on
 916 recovered clusters (b-g), shows up to 9 genetically distinguishable clusters present in sampled
 917 data (7 within Switzerland proper). Each individual is represented by a bar whose colour
 918 corresponds to its probability of belonging to recovered genetic clusters. Locations where all
 919 genotypes are split indicate all individuals are genetically similar but admixed from multiple
 920 sources. Black and white horizontal bars above structure plots delimit main river drainage and
 921 lake systems.

922

923 Figure 3. Contemporary effective population sizes (\widehat{N}_e) and migrations rates (m) among
 924 recovered populations. Circles represent the $\ln(\widehat{N}_e)*10$ and the shading outlines their upper 95%
 925 confidence limit determined from Jackknifing over loci pairs and using allele frequencies greater
 926 than 0.01. Contemporary migration rates ($m \geq 0.01$ (i.e., $\geq 1\%$) are also shown which were
 927 determined using BayesAss3.0.

928

929 Figure 4. N_e and m estimates determined from coalescent-based analyses performed in IMA2.
 930 Circles represent the $\ln(N_e)*10$ and the shading outlines upper high probability density interval
 931 similar to 95% confidence limits for Bayesian parameter estimates (HPD95). m rates determined
 932 from multiple pairwise comparisons between populations as described in text.

933

934 Figure 5. Principal coordinates analyses of genetic distances among sampled Swiss stickleback.
 935 Ellipses encircle 95% of the individuals assigned to each genetic population as determined using
 936 STRUCTURAMA/STRUCTURE. Numbers in parentheses indicate the amount of variation
 937 determined along each axes.

938

939 Figure 6. Relative assignment probabilities of sampled stickleback to various potential source
 940 populations. Panels a-c show the reassignments of individuals from the RHO, MOE and
 941 CLA/CUP populations respectively, representing the main colonizing lineages (MCL). Panels d-
 942 h show the assignment of the control NGG and COR, and the tested NEU, WHO and GIP
 943 individuals to the main lineages and the various hybrid forms expected between them. F1 =
 944 hybrid between two main lineages, F1B = back cross between an F1 hybrid and a main lineage,
 945 F2 = the combination of two similar type hybrids and F2C = the combination of two different
 946 types of hybrids and backcrosses combining the 3 MCLs.

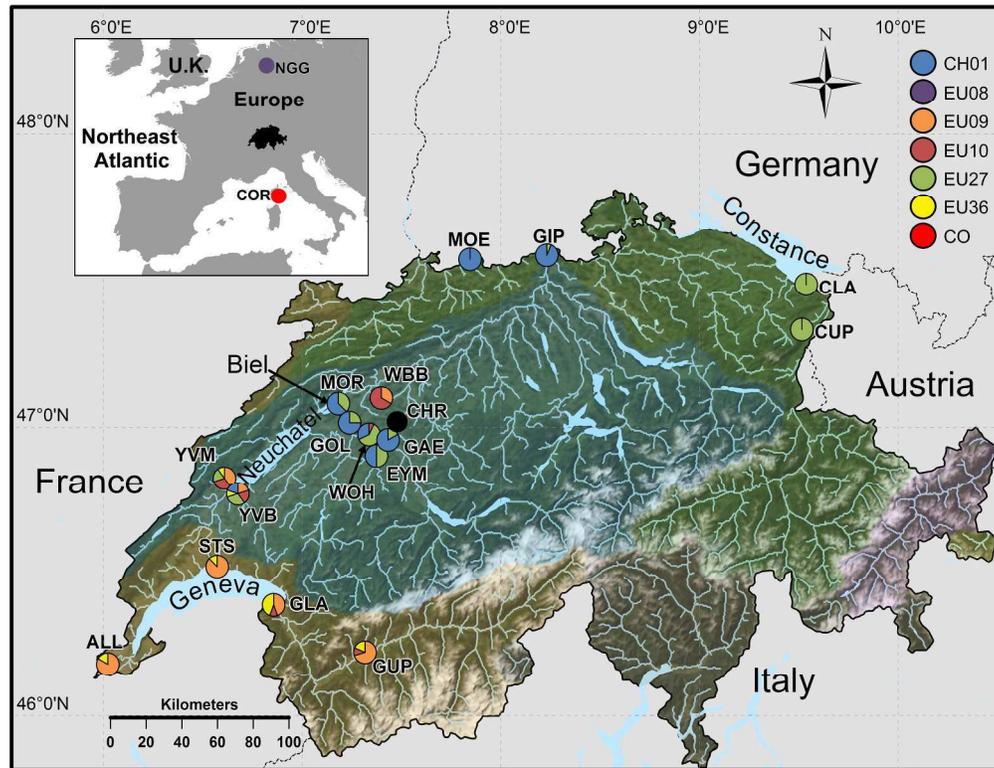


Figure 1. Detailed view of 17 locations within Switzerland where stickleback were sampled. Main river drainages are coloured (orange = Rhône, blue = Aare and green = Rhine) and five lake systems (Geneva, Neuchâtel, Wohlen (not shown), Biel and Constance). Each site code corresponds to that listed in Table S1 and shows the proportion of mtDNA haplotypes determined in Lucek et al. (2010). CHR was not assessed for mtDNA. Inset map shows Switzerland's location within mainland Europe and the location of the Corsican (COR) and the North German (NGG) sampling sites.

250x192mm (300 x 300 DPI)

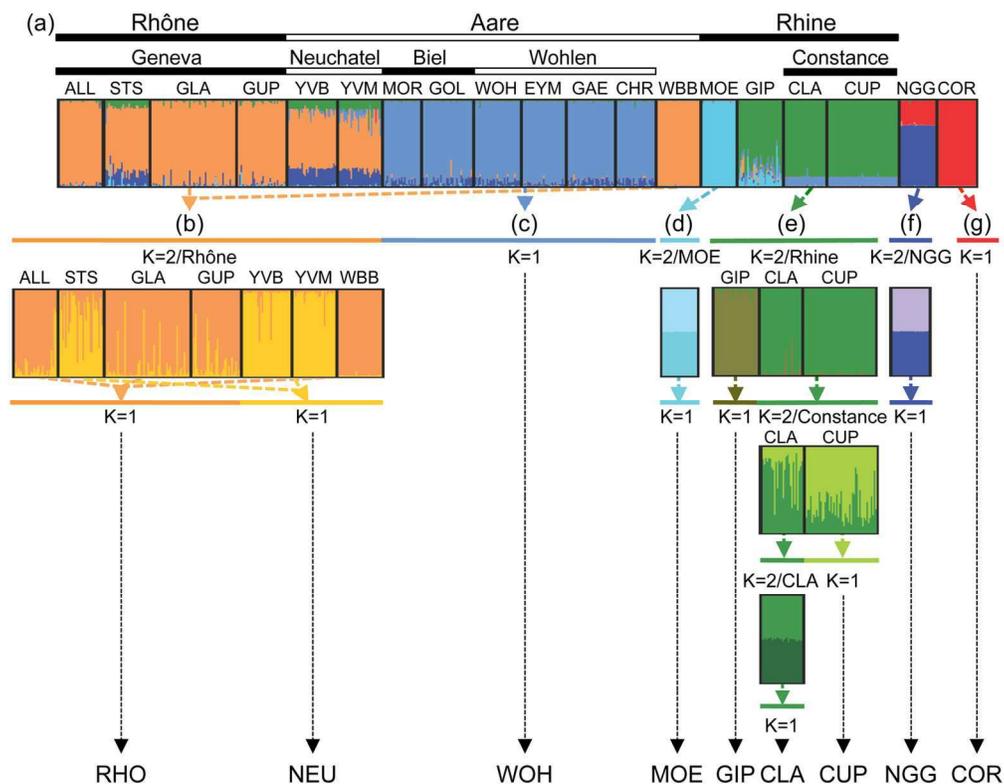


Figure 2. Hierarchical Bayesian posterior probability assignment of sampled stickleback. (a) Initial analysis using all individuals recovered 6 genetic clusters. Subsequent analysis run on recovered clusters (b-g), shows up to 9 genetically distinguishable clusters present in sampled data (7 within Switzerland proper). Each individual is represented by a bar whose colour corresponds to its probability of belonging to recovered genetic clusters. Locations where all genotypes are split indicate all individuals are genetically similar but admixed from multiple sources. Black and white horizontal bars above structure plots delimit main river drainage and lake systems.
 150x116mm (300 x 300 DPI)

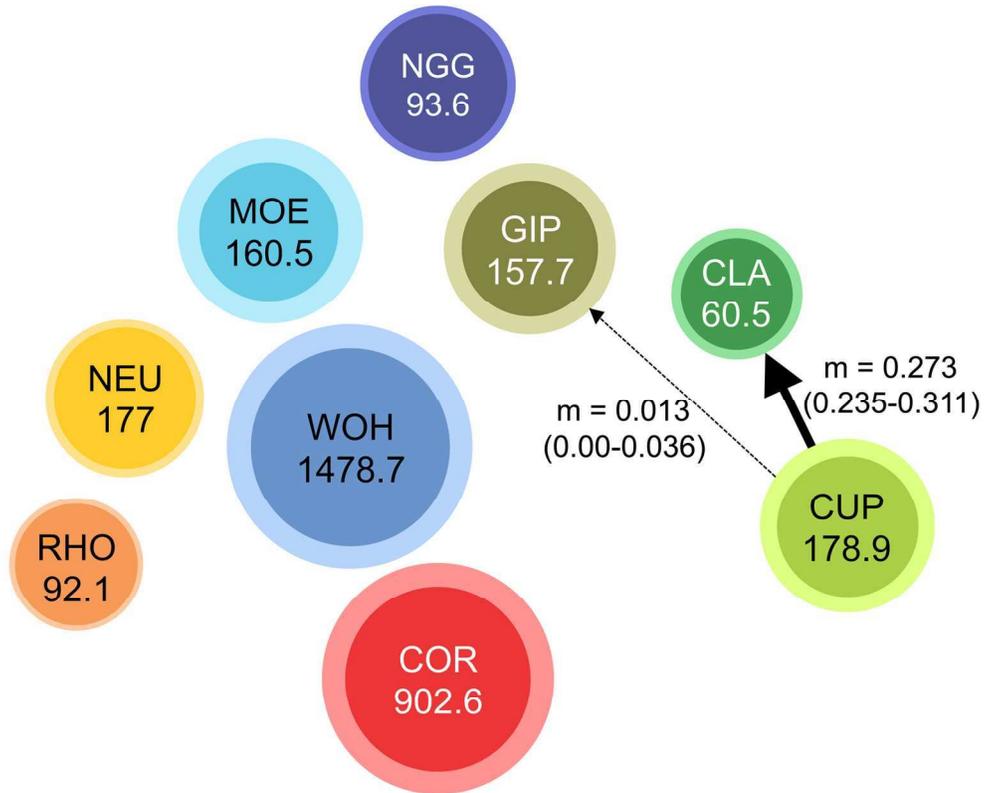


Figure 3. Contemporary effective population sizes (N_e) and migrations rates (m) among recovered populations. Circles represent the $\ln(N_e) \times 10$ and the shading outlines their upper 95% confidence limit determined from Jackknifing over loci pairs and using allele frequencies greater than 0.01. Contemporary migration rates ($m \geq 0.01$ (i.e., $\geq 1\%$) are also shown which were determined using BayesAss3.0.

150x119mm (300 x 300 DPI)

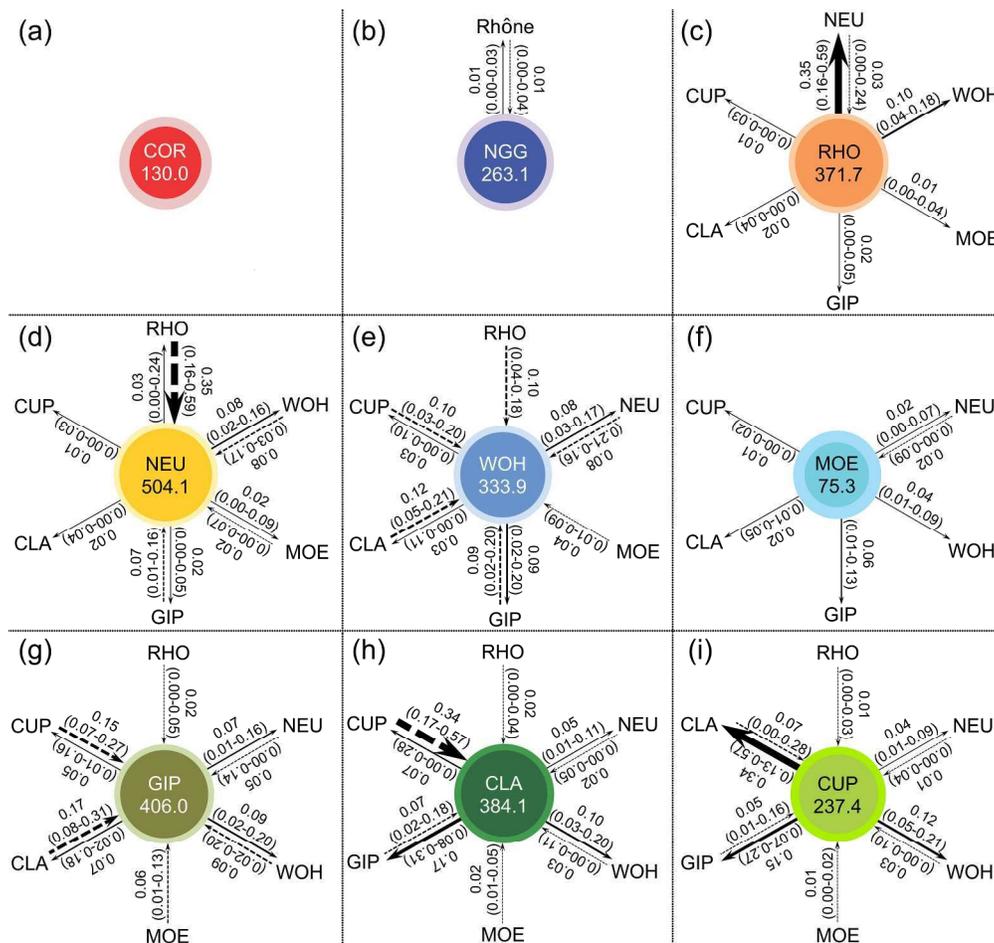


Figure 4. Ne and m estimates determined from coalescent-based analyses performed in IMA2. Circles represent the $\ln(N_e) \cdot 10$ and the shading outlines upper high probability density interval similar to 95% confidence limits for Bayesian parameter estimates (HPD95). m rates determined from multiple pairwise comparisons between populations as described in text.
250x234mm (300 x 300 DPI)

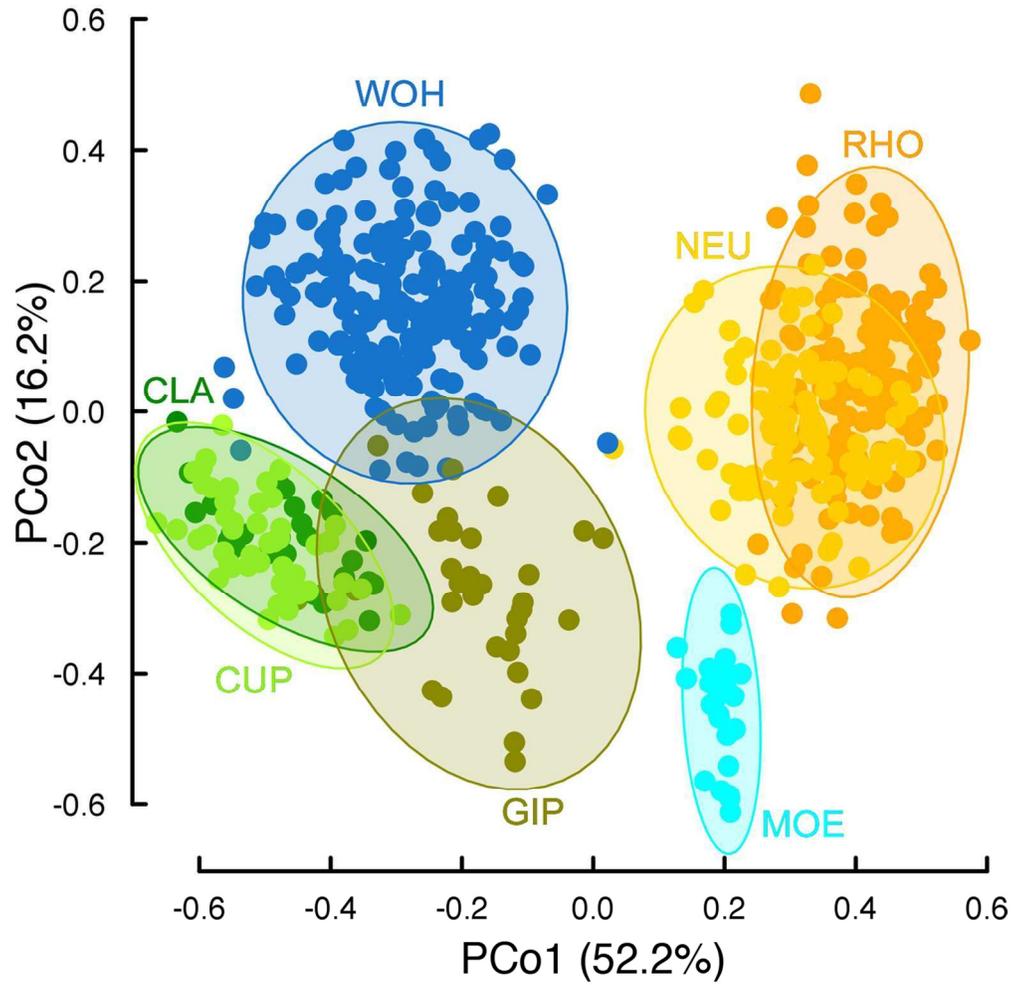


Figure 5. Principal coordinates analyses of genetic distances among sampled Swiss stickleback. Ellipses encircle 95% of the individuals assigned to each genetic population as determined using STRUCTURAMA/STRUCTURE. Numbers in parentheses indicate the amount of variation determined along each axes.

150x147mm (300 x 300 DPI)

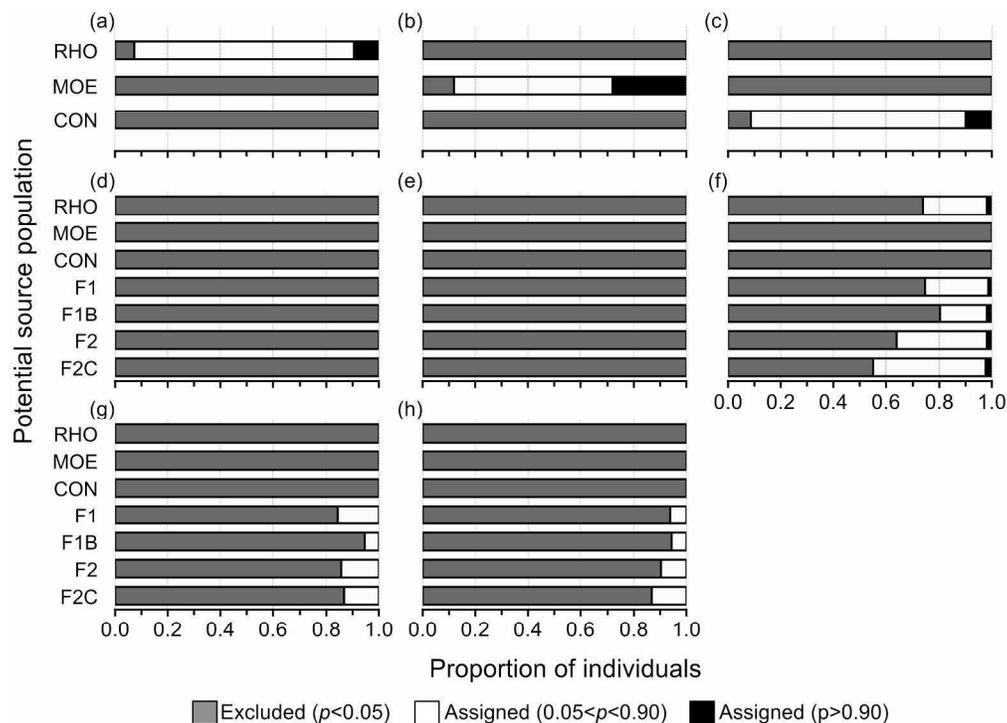


Figure 6. Relative assignment probabilities of sampled stickleback to various potential source populations. Panels a-c show the reassignments of individuals from the RHO, MOE and CLA/CUP populations respectively, representing the main colonizing lineages (MCL). Panels d-h show the assignment of the control NGG and COR, and the tested NEU, WHO and GIP individuals to the main lineages and the various hybrid forms expected between them. F1 = hybrid between two main lineages, F1B = back cross between an F1 hybrid and a main lineage, F2 = the combination of two similar type hybrids and F2C = the combination of two different types of hybrids and backcrosses combining the 3 MCLs.