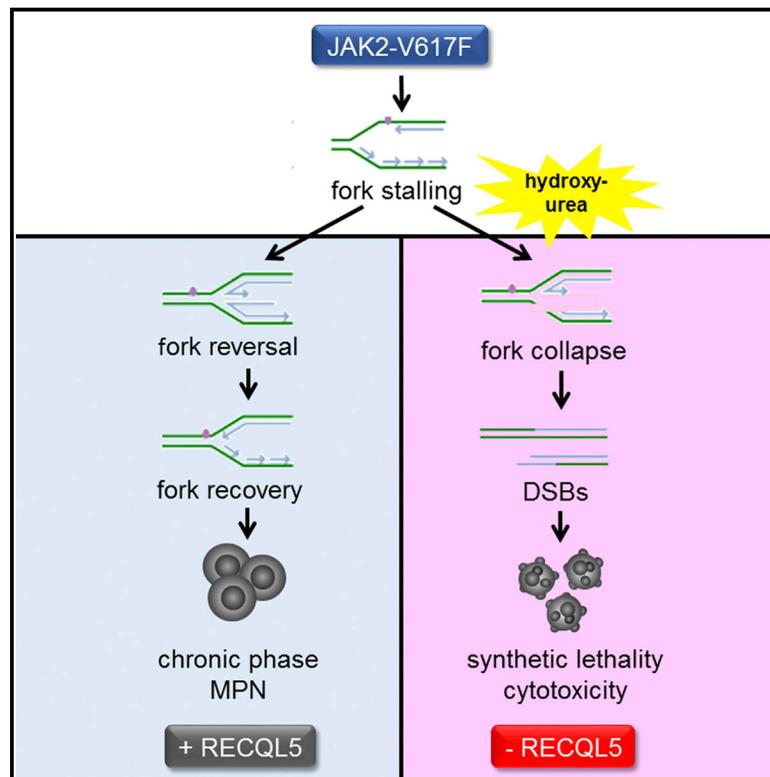


RECQL5 Suppresses Oncogenic JAK2-Induced Replication Stress and Genomic Instability

Graphical Abstract



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In Brief

Oncogenic JAK2 signaling in MPN patients leads to DNA damage, yet MPNs are characterized by genomic stability. Chen et al. show that the DNA helicase RECQL5 maintains genomic integrity in response to JAK2V617F-associated replication stress. Moreover, RECQL5 depletion sensitizes JAK2V617F-mutant cells to hydroxyurea, the most common treatment for MPN.

Highlights

- RECQL5 is upregulated in JAK2V617F mutant erythroblasts in MPN patients
- RECQL5 is a target of activated JAK2-PI3K signaling
- RECQL5 stabilizes stalled forks during JAK2V617F-associated replication stress
- RECQL5 depletion sensitizes JAK2V617F-expressing cells to hydroxyurea

RECQL5 Suppresses Oncogenic JAK2-Induced Replication Stress and Genomic Instability

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SUMMARY

JAK2V617F is the most common oncogenic lesion in patients with myeloproliferative neoplasms (MPNs). Despite the ability of JAK2V617F to instigate DNA damage *in vitro*, MPNs are nevertheless characterized by genomic stability. In this study, we address this paradox by identifying the DNA helicase RECQL5 as a suppressor of genomic instability in MPNs. We report increased RECQL5 expression in JAK2V617F-expressing cells and demonstrate that RECQL5 is required to counteract JAK2V617F-induced replication stress. Moreover, RECQL5 depletion sensitizes JAK2V617F mutant cells to hydroxyurea (HU), a pharmacological inducer of replication stress and the most common treatment for MPNs. Using single-fiber chromosome combing, we show that RECQL5 depletion in JAK2V617F mutant cells impairs replication dynamics following HU treatment, resulting in increased double-stranded breaks and apoptosis. Cumulatively, these findings identify RECQL5 as a critical regulator of genome stability in MPNs and demonstrate that replication stress-associated cytotoxicity can be amplified specifically in JAK2V617F mutant cells through RECQL5-targeted synthetic lethality.

INTRODUCTION

The propensity of cancer cells to undergo clonal evolution is enabled by a heightened state of genomic instability wherein cancer cells are continuously accumulating and repairing DNA damage. This increase in genomic flux allows cancer cells to accumulate somatic mutations that can drive disease progres-

sion. However, heightened genomic instability can also activate DNA damage-associated checkpoints, which can lead to apoptosis or cellular senescence. Cancer cells continuously tread a fine balance between cell death and survival in response to DNA damage (Negrini et al., 2010).

Chronic myeloproliferative neoplasms (MPNs) encompass a spectrum of clonal hematological disorders with an inherent tendency to transform into a more aggressive disease in the form of acute myeloid leukemia (AML). MPNs provide a window into cancer early during its ontogeny and give insights into the processes that regulate genome stability during malignant clonal evolution. The most common recurrent lesion in MPN patients is an activating V617F mutation in the JAK2 non-receptor tyrosine kinase (JAK2V617F) that causes hyperactive JAK-STAT signaling and confers a capacity for cytokine-independent growth (Baxter et al., 2005; James et al., 2005; Kralovics et al., 2005; Levine et al., 2005). Recently, a growing body of work has suggested that JAK2V617F is associated with increased DNA damage: (1) increased numbers of γ H2Ax-marked double-strand breaks (DSBs) have been detected in Ba/F3 pro-B cells overexpressing JAK2V617F (Marty et al., 2013) and in lineage-negative, Sca1-positive, c-Kit-positive (LSK) cells (enriched for hematopoietic stem cell [HSC] activity) from 6-month-old JAK2V617F-heterozygous knockin mice (Li et al., 2010); (2) JAK2V617F expression is associated with increased levels of DNA-damaging reactive oxygen species (Marty et al., 2013); (3) RAD51-positive foci indicative of increased DSB repair have been observed in CD34⁺ hematopoietic cells obtained from JAK2V617F-positive MPN patients (Plo et al., 2008); and (4) JAK2V617F expression in both human diploid fibroblasts and primary erythroblasts from MPN patients leads to higher rates of stalled replication forks, with improper processing of stalled replication intermediates representing a potential source of DSBs (Chen et al., 2014).

Given the genome-destabilizing functionalities of JAK2V617F and the inherent tendency for leukemic transformation in patients with MPN, a reasonable supposition is that oncogenic

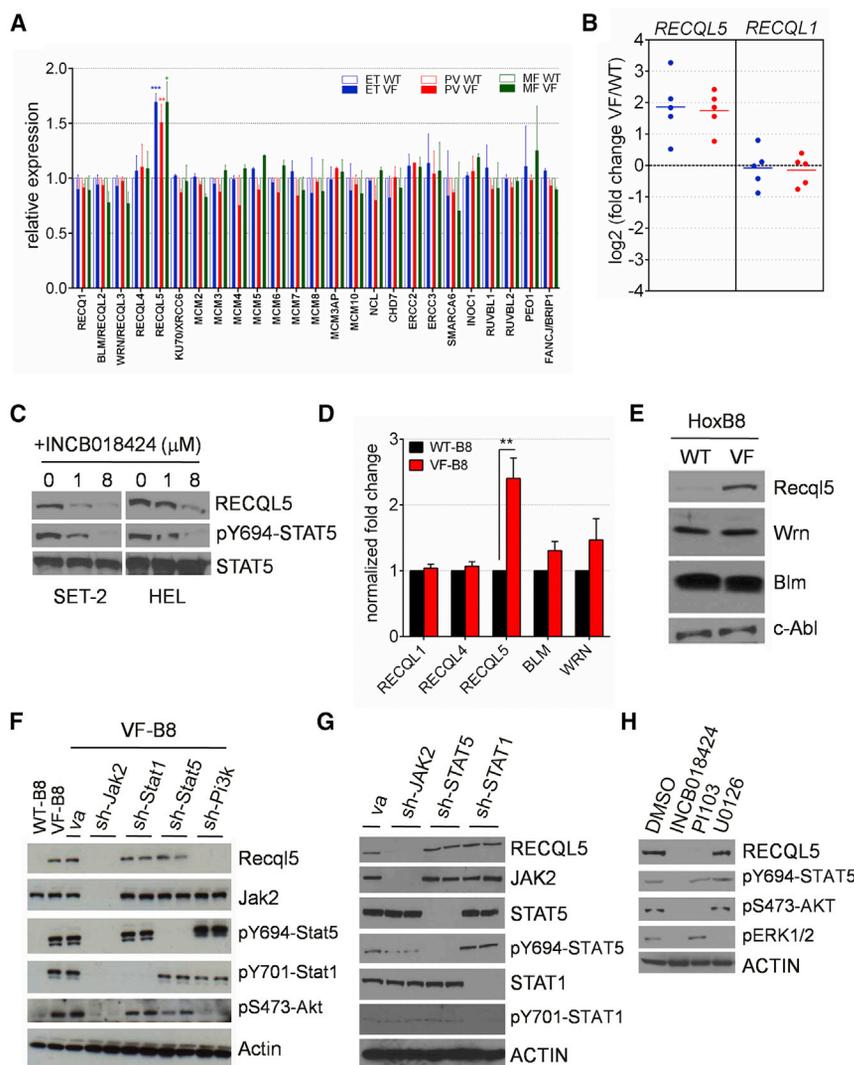


Figure 1. JAK2V617F Increases the Expression of RECQL5

(A) Gene expression profiles depicting the expression of 25 DNA helicases in 40 MPN patients (20 ET patients, 16 PV patients, and 4 MF patients). (B) qPCR validation of RECQL5 and RECQL1 expression in ET (blue) and PV (red) patients. (C) RECQL5 expression in SET-2 and HEL cells following treatment with 0–8 mM of the JAK2 inhibitor INCB18424. (D and E) Expression of Recq family members in WT-B8 and VF-B8 cells by qPCR (D) and western immunoblot (E). * $p < 0.01$. (F) Recq5 expression in WT-B8 and VF-B8 cells following knockdown of Jak2 (sh-Jak2), the p85 subunit of PI3K (sh-Pi3k), Stat1 (sh-Stat1), or Stat5 (sh-Stat5). (G) RECQL5 expression in HEL cells following knockdown of JAK2 (sh-JAK2), STAT5 (sh-STAT5), or STAT1 (sh-STAT1). (H) RECQL5 expression in HEL cells following treatment for 16 hr with INCB018424 (1 μ M), the PI3K inhibitor PI103 (1 μ M), or the ERK1/2 inhibitor U0126 (5 μ M). See also [Figure S1](#).

JAK2 signaling imposes a mutator phenotype on MPN cells, accelerating the accumulation of mutations and promoting clonal evolution and disease progression. However, longitudinal studies of MPN patients indicate that JAK2V617F-positive polycythemia vera (PV) and essential thrombocythemia (ET) patients (i.e., chronic-phase MPN) typically remain clinically and cytogenetically stable over decades ([Tefferi et al., 2014](#)). A recent copy-number analysis of the genome of chronic-phase MPN patients has shown that cytogenetic abnormalities are rare ([Klampfl et al., 2011](#)), and an analysis of the mutational landscape of PV and ET patients revealed that each MPN patient harbors a modest number of mutations per exome (approximately 6.5) ([Nangalia et al., 2013](#)).

To reconcile the apparent paradox of JAK2V617F-induced DNA damage with the clinical and cytogenetic stability characteristic of chronic-phase MPN, we hypothesized that JAK2V617F, in addition to instigating a state of increased DNA damage, could also, in parallel, activate protective pathways that counteract and prevent DNA damage-induced apoptosis. In this report, we

identify increased expression of the DNA repair helicase RECQL5 in JAK2V617F-expressing cells and characterize its role in constraining JAK2V617F-induced replication stress and maintaining genomic integrity in MPNs.

RESULTS

Activated JAK2 Signaling Regulates Expression of the RECQL5 Helicase in MPN Cells

We analyzed the gene expression profiles of autologous normal and JAK2V617F-

heterozygous burst-forming unit erythroid (BFU-E) colonies from 40 MPN patients ([Chen et al., 2010](#)). To explore the hypothesis that JAK2V617F may activate genes that counteract DNA damage, we evaluated the expression of DNA helicases in JAK2 mutant and autologous normal cells in this dataset. Of 25 DNA helicases expressed, RECQL5 was the sole DNA helicase enzyme whose expression was elevated significantly in mutant-JAK2 BFU-Es relative to autologous BFU-Es in ET, PV, and myelofibrosis (MF) patients after multiple hypothesis testing ($q < 0.05$) ([Figure 1A](#)). Real-time qPCR analysis of 10 MPN patients (five PV patients and five ET patients) demonstrated increased expression of RECQL5 (but not the related RECQ family member RECQL1) in mutant JAK2 colonies relative to autologous wild-type colonies in both PV and ET patients ([Figure 1B](#)). We next assessed RECQL5 levels following JAK2 inhibition in two independent cell lines that harbor the JAK2V617F mutation: HEL and SET-2 cells. We found that treatment of the erythroleukemic cell line HEL and the megakaryoblastic cell line SET-2 with the JAK2 inhibitor INCB018424 reduced RECQL5 expression

concomitant with decreased STAT5 phosphorylation (Figure 1C). In aggregate, these data indicate that RECQL5 expression is regulated by activated JAK2 signaling in human disease-relevant contexts.

Physiological Levels of Jak2V617F Lead to Increased Recq15 Expression

To investigate the role of Recq15 under conditions that more closely recapitulate chronic-phase MPN in patients, we generated disease-relevant cell lines from Jak2V617F knockin mice that we developed previously and characterized extensively (Mullally et al., 2010, 2013). This mouse model closely recapitulates the features of human MPN, and Jak2V617F expression is physiological in the model, being driven from the endogenous Jak2 promoter. To generate cell lines, we engineered bone marrow progenitors from littermate wild-type or Jak2V617F knockin mice to express a β -estradiol-regulated Hoxb8 homeodomain-containing protein. Following 3 weeks of serial passaging and antibiotic selection, immortalized myeloid progenitor cells were generated from both wild-type mice (WT-B8) and Jak2V617F knockin mice (VF-B8) (Wang et al., 2006). Both the WT-B8 and VF-B8 lines resemble cells at a granulocyte-macrophage progenitor (GMP) stage of myeloid maturation by cell surface immunophenotype analysis (Figure S1A). Immunoblotting showed equivalent levels of Jak2 expression in both cell lines but increased levels of phosphorylated Stat5 in VF-B8 cells relative to WT-B8 cells (Figure S1B). Additionally, VF-B8 cells exhibited an increased proliferative capacity and survival under reduced serum conditions (Figures S1C–S1D), indicating that VF-B8 cells recapitulate this key pathognomonic feature of MPN biology.

We next determined whether the expression of Recq15 was modulated by mutant Jak2 in these Hoxb8-immortalized cell lines. In accordance with the primary human data, VF-B8 cells exhibited elevated expression of Recq15 relative to WT-B8 cells by qPCR analysis (Figure 1D) and western immunoblotting (Figure 1E). No significant differential expression of other Recq family members (Recq1, Recq4, Blm, and Wrrn) was observed. These data demonstrate that mutant JAK2 increases expression of the DNA repair helicase RECQL5 in both human and murine cells.

RECQL5 Is a Target of JAK2-PI3K Signaling

To determine which signaling pathways were necessary for the regulation of Recq15 by Jak2V617F, we used small hairpin RNAs (shRNAs) to knock down Jak2 and downstream effector molecules of Jak2 in VF-B8 cells. Expression of shRNAs targeting Jak2 led to diminution of Recq15 levels relative to cells transduced with an empty vector control (Figure 1F), which is consistent with the finding that Recq15 is a downstream target of Jak2 signaling. Next, we assessed Recq15 levels in VF-B8 cells following transduction of shRNAs targeting Stat1, Stat5, or the p85 subunit of phosphatidylinositol 3-kinase (PI3K). The levels of Recq15 in VF-B8 cells were reduced following knockdown of PI3K but not Stat1 or Stat5 (Figure 1F). Similarly, RECQL5 levels were also attenuated following knockdown of JAK2 in HEL cells but were not attenuated following knockdown of STAT5 or STAT1 (Figure 1G), and treatment of HEL cells with the PI3K inhibitor PI103 (but not the ERK1/2 inhibitor U0126) led to attenuated levels of RECQL5, accompanied by decreased AKT

phosphorylation (Figure 1H). Taken together, these data indicate that RECQL5 expression is increased upon JAK2 activation in a PI3K-dependent manner.

Recq15 Depletion Sensitizes Jak2V617F-Expressing Cells to Replication Stress

We next sought to clarify the function of RECQL5 upregulation in JAK2V617F-expressing cells. Previously, we had demonstrated that overexpression of JAK2V617F in human diploid fibroblasts resulted in increased replication fork stalling and replication stress (Chen et al., 2014). Because RECQL5 has been linked functionally to regulating stalled replication forks in normal cells, we hypothesized that the upregulation of RECQL5 may function to mitigate the deleterious consequences of replication stress in JAK2 mutant cells.

To test this, we knocked down Recq15 in VF-B8 cells using three independent shRNAs (Figure S2A). Densitometric analysis revealed a knockdown efficiency of 61%–95% for the three hairpins. Under normal growth conditions, Recq15 knockdown in VF-B8 cells did not affect the proliferation rate (Figure S2B) or apoptosis (Figure S2C) relative to either equivalently modified WT-B8 cells or VF-B8 cells expressing empty vector alone (VA) controls. To simulate replication stress, we subjected WT-B8 and VF-B8 cells to low-serum conditions to deprive cells of deoxynucleotide triphosphate (dNTP) (Bester et al., 2011). We observed that VA-expressing VF-B8 cells exhibited greater viability relative to WT-B8 cells under low-serum conditions (Figure 2A, left), which is in accordance with our previous data on non-genetically perturbed cells. In contrast, Recq15-depleted VF-B8 cells exhibited decreased viability relative to equivalently modified WT-B8 cells (Figure 2A, right). Hypersensitivity of Recq15-depleted VF-B8 cells to low-serum conditions was abrogated completely upon repletion of deoxynucleotide (dNTPs) into the culture medium (Figure 2B). The results shown represent the average of three independent Recq15-targeting shRNAs and are consistent with data obtained for each individual hairpin (Figure S2D). Collectively, these data demonstrate that Recq15 protects VF-B8 cells from endogenous replication stress instigated by low dNTP levels.

We next tested whether Recq15 depletion would also sensitize VF-B8 cells to exogenous instigators of replication stress, such as pharmacological agents. We tested hydroxyurea (HU) and camptothecin (CPT), which impair DNA replication by limiting production of dNTPs and inhibiting topoisomerase I activity, respectively. Strikingly, contemporaneous knockdown of Recq15 and exposure to either HU or CPT led to significantly decreased viability of VF-B8 cells compared with like-treated WT-B8 cells after 24 hr of drug treatment (Figures 2C, 2D, and 2G; Figure S2D). Decreased cell viability was noted as early as 12 hr post-drug treatment (Figures S2E and S2F). In contrast, Recq15-depleted VF-B8 cells were not hypersensitive to the double-stranded breaking agents doxorubicin (DOX) and etoposide (ETP) (Figures 2E and 2F; Figure S2D). Concordant with these data, shRNA depletion of human RECQL5 also enhanced cytotoxicity in HEL and SET2 cells, but only to pharmacological instigators of replication stress (HU, CPT, aphidicolin, and irinotecan) and not to pro-oxidants or DSB-generating drugs (Figures S2G and S2H).

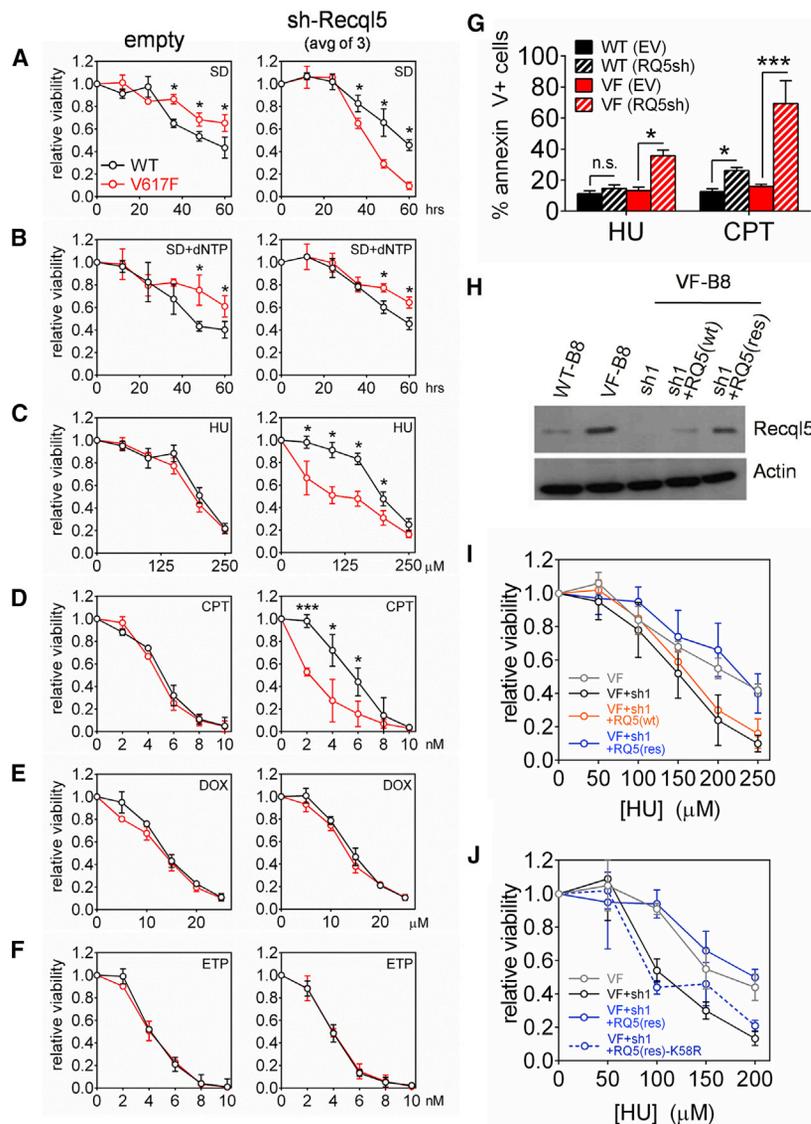


Figure 2. Recq15 Depletion Sensitizes Jak2V617F-Expressing Cells to Replication Stress

(A–F) Viability of WT-B8 (black) and VF-B8 (red) cells transduced with VA or Recq15-targeting shRNAs following serum deprivation (SD) in 0.5% fetal calf serum (FCS) (A), SD with 100 μM dNTPs (SD + dNTPs) (B), HU (0–250 μM) (C), CPT (0–10 nM) (D), DOX (0–25 μM) (E), or ETP (0–10 nM) (F). For VA-transduced cultures, each point represents the mean of three independent cultures. For Recq15-knockdown cultures, each point represents the mean of three separate Recq15-targeting shRNAs.

(G) Quantitation of annexin V positivity in cells treated with 100 μM HU or 4 nM CPT.

(H–J) VF-B8 cells were transduced with Recq15-targeting shRNA simultaneously with either a wild-type Recq15 cDNA (RQ5wt), shRNA-resistant Recq15 cDNA (RQ5res), or RQ5res cDNA harboring a K58R mutation (RQ5res-K58R). Immunoblotting for Recq15 protein levels was performed (H), and cell viability was assessed (I and J).

Testing for statistical significance was performed using Student's t test (*p < 0.05, **p < 0.01, ***p < 0.001). See also Figure S2.

Given the potential off-target effects of shRNAs that may potentially influence the observed phenotypes, we next confirmed the specificity of the Recq15 shRNAs. We designed a Recq15 cDNA, RQ5(res), that was mutated at every third nucleotide to disrupt each of the shRNA-binding sites while retaining the correct amino acid encoded at each triplet codon. Expression of a wild-type Recq15 (RQ5(WT)) in VF-B8 cells together with a Recq15-targeting shRNA did not rescue Recq15 expression (Figure 2H) and failed to abrogate the increased sensitivity of Recq15-depleted VF-B8 cells to HU (Figure 2I). In contrast, expression of RQ5(res) resulted in high levels of Recq15 expression

that were maintained despite expression of a Recq15 shRNA (Figure 2H) and successfully abrogated the hypersensitivity to HU of the VF-B8 cells co-expressing the Recq15 shRNA (Figure 2I). Moreover, a RQ5(res) cDNA harboring a K58R mutation within the helicase domain is incapable of abrogating HU hypersensitivity, revealing the essentiality of Recq15 helicase activity in protecting against replication stress (Figure 2J). Together, these data demonstrate that the effects of the shRNAs were on target and that Recq15 depletion was the critical factor for conferring increased sensitivity of VF-B8 cells to replication stress.

Replication stressors such as HU and CPT can also induce DSBs at sufficiently high doses. We therefore validated whether the HU and CPT doses associated with preferential cytotoxicity of Recq15-depleted VF-B8 cells were causing increased replication stress or excessive formation of DSBs. To differentiate between these two phenomena, we treated parental WT-B8 and VF-B8 cells with HU (6 μM) and CPT (4 nM) and performed immunocytochemical staining for foci containing the RPA protein (which marks stalled replication forks) or 53BP1 (which localizes to DSBs) (Figures S3A and S3B). At these HU and CPT dosages, we observed a marked increase in foci containing RPA (Figure S3C), with only a marginal increase in numbers of 53BP1-positive foci (Figure S3D). In contrast, etoposide (4 nM) generated both RPA-positive and 53BP1-positive foci (Figures S3C and S3D). In aggregate, these findings verify that the dosage of HU and CPT used to enhance the cytotoxicity of Recq15-depleted VF-B8 cells causes enhanced replication stress with only a slight elevation of DSBs.

that were maintained despite expression of a Recq15 shRNA (Figure 2H) and successfully abrogated the hypersensitivity to HU of the VF-B8 cells co-expressing the Recq15 shRNA (Figure 2I). Moreover, a RQ5(res) cDNA harboring a K58R mutation within the helicase domain is incapable of abrogating HU hypersensitivity, revealing the essentiality of Recq15 helicase activity in protecting against replication stress (Figure 2J). Together, these data demonstrate that the effects of the shRNAs were on target and that Recq15 depletion was the critical factor for conferring increased sensitivity of VF-B8 cells to replication stress.

Recq15 Depletion Increases the Severity of Replication Fork Stalling in Jak2V617F-Expressing Cells Exposed to Exogenous Replication Stress

To understand the molecular mechanisms by which Recq15 depletion in VF-B8 cells leads to increased cytotoxicity by exogenous replication stressors (such as HU), we performed

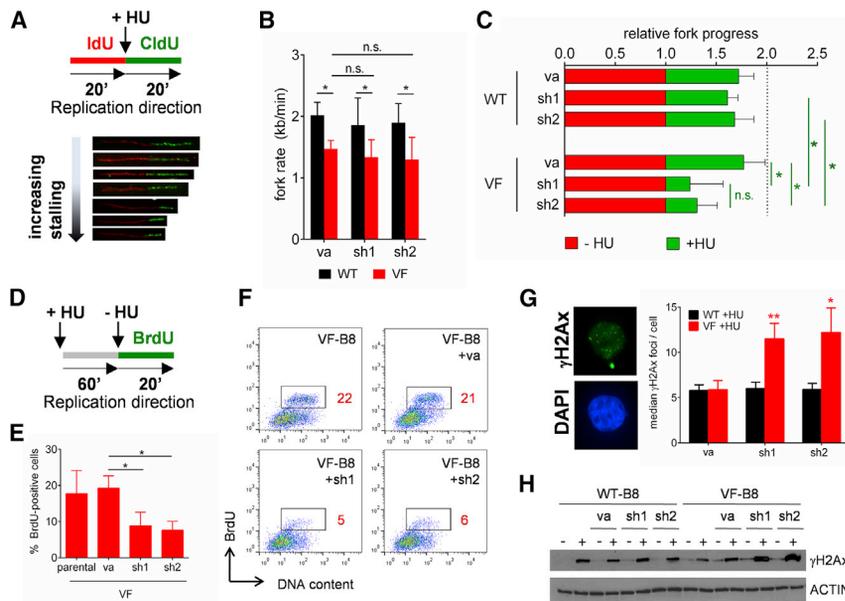


Figure 3. Recq15 Protects against the Collapse of Stalled Replication Forks in Jak2V617F-Expressing Cells Exposed to Replication Stress

(A) Schematic of the time course for chromosome combing experiments (top). Shown are representative replication structures of a single combed DNA molecule labeled with IdU (red) and CldU (green) (bottom).

(B) Fork rate in WT-B8 and VF-B8 cells transduced with empty vector alone (VA) or with two different Recq15-targeting shRNAs (sh1 and sh2). The results represent the mean \pm SD for at least 50 fibers. (C) Quantification of HU-induced effects on fork dynamics. Relative fork progress is depicted as the normalized ratio of the second (HU-treated) labeling step relative to the first (HU-free) labeling step.

(D–F) Schematic of the timeline for fork restart experiments (D). Also shown are quantification of BrdU-positive cells (E) and representative flow cytometric plots of BrdU staining (F).

(G and H) Quantification of γ H2Ax-marked double-stranded breaks as assessed by immunofluorescence detection (G) and western immunoblot (H). (D–H) Testing for statistical significance was performed using Student's t test (* $p < 0.05$; ** $p < 0.01$; n.s., not significant).

See also Figure S2.

chromosome combing, which allows the direct visualization and analysis of replication tracts on individual, bromodeoxyuridine (BrdU)-labeled DNA fibers. We subjected WT-B8 or VF-B8 cells transduced with Recq15-targeting shRNAs or VA controls to this procedure. All cultures underwent a first labeling step with the BrdU analog iododeoxyuridine (IdU) under normal growth conditions, followed by a second labeling step with another BrdU analog, chlorodeoxyuridine (CldU), in culture medium supplemented with HU (Figure 3A). In this way, the extent of fork progression can be measured both in the absence and presence of HU to determine any differential effects of HU on the cultures.

We focused our initial analysis on fibers generated solely from the first labeling step to ascertain whether Recq15 depletion in the absence of HU altered DNA replication kinetics. For this analysis, we measured the length of these fibers and calculated the average fork rate because decreased fork processivity is a robust indicator of fork stalling. In control cells, we observed a significant decrease in the mean replication rate in VF-B8 cells compared with WT-B8 cells (1.47 ± 0.18 kb/min [$n = 88$] in VF-B8 cells versus 1.97 ± 0.21 kb/min [$n = 94$] in WT-B8 cells [$p < 0.01$]) (Figure 3B), consistent with reports published previously indicating a replication processivity impairment in JAK2V617F-expressing cells (Chen et al., 2014). However, the mean replication rate was not altered significantly by Recq15 knockdown in either WT-B8 or VF-B8 cells (WT-B8+sh1, 1.88 ± 0.44 kb/min [$n = 80$]; WT-B8+sh2, 1.91 ± 0.28 kb/min [$n = 55$]; VF-B8+sh1, 1.33 ± 0.31 kb/min [$n = 101$]; VF-B8+sh2, 1.32 ± 0.39 kb/min [$n = 50$]). Critically, no difference was observed in fork rate between VF-B8 cells depleted for Recq15 relative to those transduced with an empty vector (Figure 3B). These data indicate that physiological levels of Jak2V617F expression in myeloid cells gives rise to a replication stress

phenotype, as evidenced by decreased fork processivity, but that Recq15 depletion alone has no additional affect.

We next explored the possibility that depletion of Recq15 in WT-B8 and VF-B8 cells could lead to altered replication dynamics in the presence of HU. Because replication fork progress in VF-B8 cells is impaired globally relative to WT-B8 cells, to facilitate comparison, the extent of fork progress during the first (HU-free) labeling step was normalized and arbitrarily designated at 1, and the second (HU-treated) labeling step was depicted relative to the normalized first label. Using this analysis, we observed that the presence of HU impaired fork progression in all cultures tested (Figure 3C). However, the impairment in fork progression caused by exposure to HU was significantly greater in Recq15-depleted VF-B8 cells relative to control VF-B8 cells and Recq15-depleted WT-B8 cells (Figure 3C). This indicates that Recq15 depletion differentially impairs replication fork progression in VF-B8 cells following HU exposure compared with WT-B8 cells.

The exacerbation in impairment of fork progression may be due to an increased frequency of fork collapse. To test for this, we measured the restart efficiency of a stalled replication fork, a process that is highly impaired following fork collapse. VF-B8 cells transduced with a control shRNA or shRNAs targeting Recq15 were initially exposed to HU for 60 min, followed by removal of HU and addition of BrdU (Figure 3D). Flow cytometric detection of BrdU-positive cells reflected cells that had efficiently restarted stalled forks. We observed significantly fewer BrdU-positive cells in HU-treated Recq15-depleted VF-B8 cells compared with cells expressing control hairpins (Figures 3E and 3F). Consistent with a role for Recq15 in protecting VF-B8 cells against HU-induced fork collapse, we noted that, following HU exposure, Recq15-depleted VF-B8 cells exhibited

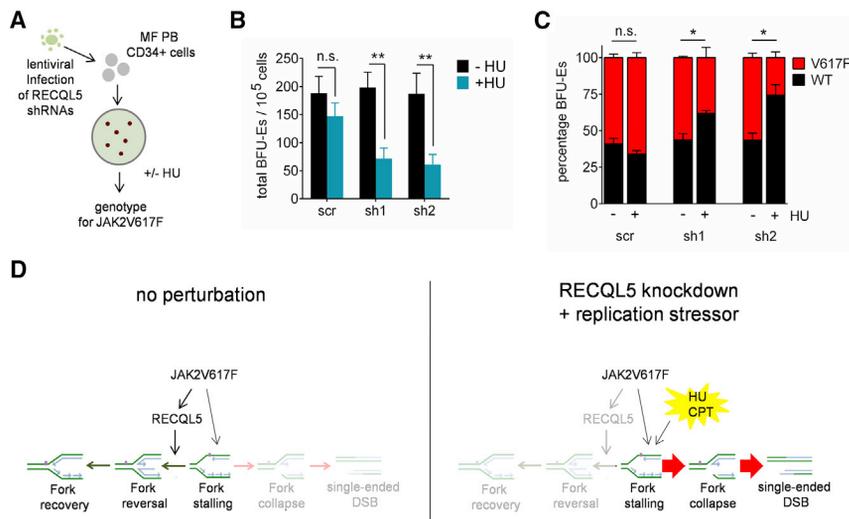


Figure 4. RECQL5 Depletion Sensitizes JAK2V617F-Positive Cells from MPN Patients to Replication Stressors

(A) Workflow for determining the effect of RECQL5 knockdown on the HU sensitivity of primary MPN samples.

(B and C) CD34⁺ peripheral blood mononuclear cells from three myelofibrosis patients were transduced with a non-targeting shRNA (scr) or RECQL5-targeting shRNA (sh1 and sh2) and cultured on methylcellulose in the presence and absence of 2 μ M HU. After 14 days, total BFU-Es were counted (B) and genotyped for JAK2V617F (C). * $p < 0.05$, ** $p < 0.01$.

(D) Model depicting interactions between JAK2V617F, RECQL5 depletion, and exogenous pharmacological replication stress.

See also Figure S4.

an increase in DSBs, as indicated by higher numbers of γ H2A-positive foci (Figure 3G) and higher levels of γ H2Ax by immunoblot analysis (Figure 3H). Together, these data demonstrate that Recq15 is essential to maintain fork stability in mutant JAK2-expressing cells following exposure to HU.

RECQL5 Depletion Increases the Sensitivity of JAK2V617F-Positive Cells from MPN Patients to HU

Finally, we tested whether modulation of RECQL5 could also increase the sensitivity of JAK2V617F-positive cells from primary MPN patients to HU. Following depletion with RECQL5, CD34-positive peripheral blood mononuclear cells from JAK2V617F-positive myelofibrosis patients were grown in semi-solid medium supplemented with HU for 14 days (Figure 4A). Strikingly, following RECQL5 depletion, there was a preferential eradication of total BFU-E colonies relative to a control shRNA (Figure 4B). Moreover, genotyping of the colonies for JAK2V617F positivity revealed a preferential elimination of JAK2V617F-positive BFU-E colonies compared with autologous wild-type colonies following exposure to HU (Figure 4C; Figure S4). Collectively, this indicates that RECQL5 knockdown preferentially sensitizes JAK2V617F-positive BFU-E colonies from MPN patients to pharmacological induction of replication stress with HU.

DISCUSSION

Chronic MPNs represent a model of the early stages of leukemogenesis and can provide insights into the balance between oncogene-associated DNA damage and the mechanisms that act to constrain it. JAK2V617F is the most common molecular driver of MPNs, and, in PV and ET, it is frequently the sole genetic driver identified. Although there is experimental evidence demonstrating that JAK2V617F induces DNA damage, clinical evidence indicates that chronic-phase MPNs follow a relatively indolent course over decades and that MPN genomes remain generally stable over time. In this study, we help resolve this apparent conundrum by demonstrating a role for the DNA repair helicase RECQL5 in regulating the balance between

JAK2V617F-induced DNA replication stress and genomic integrity in patients with MPNs.

RECQ helicases are a family of highly conserved genome surveillance enzymes. Humans possess five RECQ helicases—RECQL1, BLM, WRN, RECQL4, and RECQL5—that have both unique and overlapping roles in the regulation of DNA replication, repair, and transcription (Larsen and Hickson, 2013). Strikingly, we observed that the increased expression of RECQL5 in JAK2V617F mutant cells was specific and not seen with other RECQ helicases. This finding precludes the possibility that increased RECQL5 levels are due to an excess of S phase cells in cultures of JAK2V617F-expressing cells or that they represent an epi-phenomenon of increased DNA damage load. Rather, using shRNA knockdown and various pharmacological agents to inhibit individual signaling pathways, we demonstrate that JAK2V617F increases RECQL5 expression through the PI3K-AKT signaling axis. In terms of the effectors downstream of PI3K-AKT signaling responsible for mediating RECQL5 activation, it is possible that a direct transcriptional mechanism, such as modulating the activity of the FOXO family of transcription factors, is involved. Indeed, a similar pathway has been shown previously to be active in JAK2 mutant cells to modulate expression of the antioxidant protein catalase (Marty et al., 2013).

The mechanisms by which RECQL5 maintains genomic integrity in normal and cancer cells are ongoing areas of investigation. Germline loss-of-function mutations in RECQ family members have been associated with a predisposition to developing cancer, and a key role has been described recently for RECQL4 in hematopoiesis (Smeets et al., 2014). However, RECQL5 dysregulation has not been implicated previously in human disease. Our data identify increased expression of RECQL5 as a means by which oncogene-induced replication stress is counteracted in JAK2-mutated MPNs. Concomitant induction of replication stress and RECQL5 expression downstream of JAK2-PI3K signaling potentially forms a feedback loop that regulates DNA damage accumulation. Depleting RECQL5 in JAK2 mutant cells disrupts this homeostasis and exposes a synthetic lethal vulnerability of JAK2 mutant cells with pharmacological inducers of

replication stress. Following depletion of RECQL5, JAK2V617F-expressing cells exposed to replication stressors (such as HU or CPT) exhibit more severe fork stalling and impaired fork restarting in comparison with isogenic wild-type cells. Moreover, RECQL5-depleted JAK2 mutant cells exhibited higher numbers of DSBs and underwent apoptosis at lower doses of HU compared with RECQL5-depleted wild-type cells (Figure 4D).

We envision two mutually non-exclusive scenarios to explain why JAK2 mutant cells are more sensitive than isogenic wild-type cells to contemporaneous RECQL5 depletion and exogenous replication stress. First, JAK2V617F-expressing cells are known to have more replication fork stalling and replication stress (Chen et al., 2014). Increased RECQL5 expression in these cells may be required to mitigate replication-associated genomic instability and maintain an appropriate balance of DNA damage/repair to ensure cell viability. RECQL5 depletion in JAK2V617F-expressing cells may therefore be more deleterious than to equivalently modified wild-type cells because JAK2 mutant cells may have less leeway to cope with the additional replication stress in the form of exogenous administration of HU. Second, JAK2 mutant cells may exhibit specific replication fork structures whose resolution is especially dependent on RECQL5 and, therefore, would be particularly sensitive to RECQL5 depletion. The mechanism of JAK2V617F-induced replication stress remains unresolved, but is likely to involve unscheduled euchromatinization during S phase by JAK2 directly (Dawson et al., 2009) or following activation of downstream STATs (Shi et al., 2006), leading to physical collisions between the replication machinery and transcriptional apparatus on the same competing DNA template. The resulting DNA:RNA hybrid structures (called R loops) are potentially mutagenic and genome-destabilizing. RECQL5 may play an important role in resolving R loops in JAK2 mutant cells. Future studies to distinguish between these possibilities will be interesting.

Finally, our data may have broader pharmacological implications for cancer therapy. Although HU remains the frontline treatment for patients with ET and PV (Harrison et al., 2005), most studies have demonstrated that HU does not preferentially target the V617F-positive cell fraction in ET and PV patients (Antonionioli et al., 2010), and, as a result, HU does not alter the natural history of MPNs. Our findings highlight a potential approach to amplify replication stress-associated cytotoxicity through helicase-targeted synthetic lethality. Indeed, there is precedent for this approach. Inhibition of the Werner helicase using small molecules has been shown to sensitize HeLa and U2OS cells to the replication stressor topotecan (Aggarwal et al., 2011). Nonetheless, the development of small-molecule inhibitors of RECQL5 in MPNs would need to be approached with caution. In particular, the safety of RECQL5 inhibition with respect to its potential to provoke more genomic insults would need to be examined carefully. However, exploiting the differential dependencies of cancer cells on DNA repair pathways has already demonstrated clinical efficacy through the use of PARP1 inhibitors in BRCA-mutant cancers (Bryant et al., 2005). Identifying and leveraging synthetic lethal relationships with DNA helicases may represent another similar approach for improved anti-cancer treatment strategies more broadly.

EXPERIMENTAL PROCEDURES

Generation of Myeloid Progenitor Cell Lines from Jak2V617F Knockin Mice

Myeloid progenitor cell lines were generated from a C57Bl/6 Jak2V617F-expressing mouse (Mullally et al., 2010) and from a Jak2 wild-type littermate control by retroviral overexpression of an estrogen-dependent Hoxb8 transcription factor, as described previously (Wang et al., 2006). Conditionally immortalized myeloid progenitor cell lines from wild-type mice (WT-B8) and Jak2V617F knockin mice (VF-B8) were maintained in myeloid medium (RPMI medium + 10% fetal bovine serum (FBS) supplemented with 50 ng/ml murine stem cell factor (mSCF) and 1 μ M β -estradiol (Sigma).

Cell Viability Assays

WT-B8 and VF-B8 cells were counted, and 5×10^4 cells were seeded in triplicate in each well of a 96-well plate in 100 μ l of myeloid medium. For drug treatments, agents were added freshly. For serum deprivation studies, cells were rinsed with 1 \times PBS twice, and 1×10^5 cells/well were seeded in a 96-well plate in RPMI medium + 0.5% FBS supplemented with 1 μ M β -estradiol. Cell growth was measured using a standard Alamar blue assay (Life Technologies).

Chromosome Combing

Chromosome combing was performed as described previously (Chen et al., 2014). Briefly, 1×10^5 WT-B8 or VF-B8 cells were seeded in a well of a 96-well plate in a volume of 100 μ l. Replicating DNA was first labeled with 25 μ M IdU (Sigma), followed by 250 μ M CldU (Sigma) for 20 min each. Cells were then harvested, genomic DNA was extracted, and individual DNA molecules were stretched on glass slides. The slides were immunostained with fluorescently labeled anti-IdU (1:300) and anti-CldU (1:150) and analyzed under oil immersion on a Zeiss AxioScope 2 fluorescence microscope.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Supplemental Experimental Procedures and four figures and can be found with this article online at <http://dx.doi.org/10.1016/j.celrep.2015.11.037>.

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Cell Reports

Supplemental Information

RECQL5 Suppresses Oncogenic JAK2-Induced Replication Stress and Genomic Instability

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SUPPLEMENTAL INFORMATION

RECQL5 suppresses JAK2V617F-induced replication stress and genomic instability

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E.C., J.S.A., D.B.S., J.B. and A.K. performed experiments. A.L.G., J.N., D.J.D., A.R.G. and A.M. provided patient samples. E.C., J.S.A., A.R.G. and A.M. designed experiments. E.C. and A.M. wrote the paper.

Supplemental Data

1. **Figure S1** (Related to Figure 1). Characterization of wild-type and Jak2V617F-expressing Hoxb8-immortalized myeloid progenitor cells.
2. **Figure S2** (Related to Figure 2). Recql5 depletion sensitizes Jak2V617F-expressing cells to replication stress.
3. **Figure S3** (Related to Figure 2). RECQL5 depletion sensitizes human JAK2V617F-expressing cell lines to replication stress.
4. **Figure S4** (Related to Figure 4). RECQL5 depletion sensitizes JAK2V617F-positive cells from MPN patients to replication stressors.

Supplemental Experimental Procedures

1. Detailed description of myeloid progenitor cell line generation from Jak2V617F-knockin mice
2. Cells and reagents
3. Alamar blue assay for measurement of cell viability
4. Chromosome combing calculations and statistical analyses
5. Vectors
6. Generation of lentiviral particles and lentiviral infections
7. Real-time qPCR analysis

Figure S1

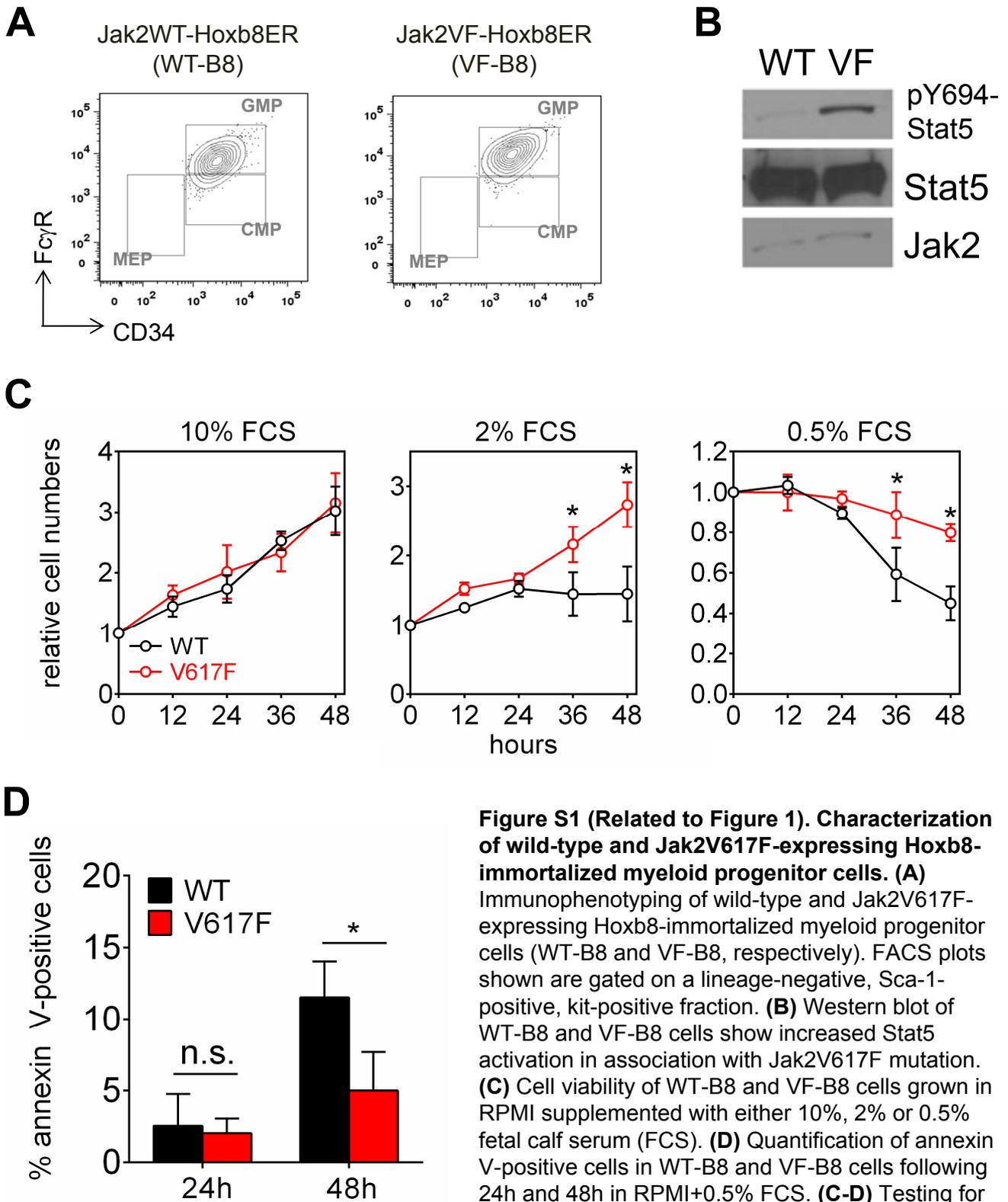


Figure S2

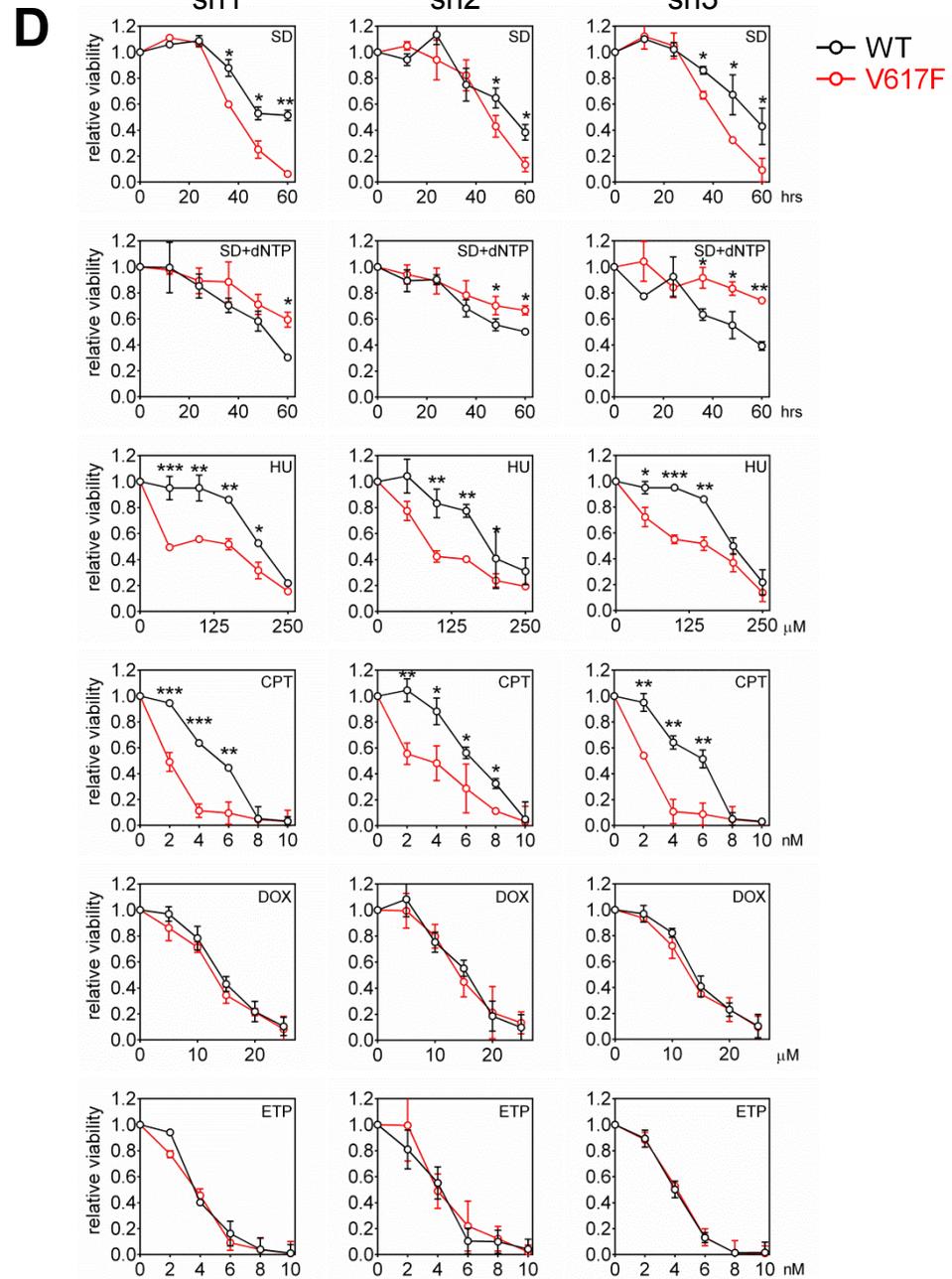
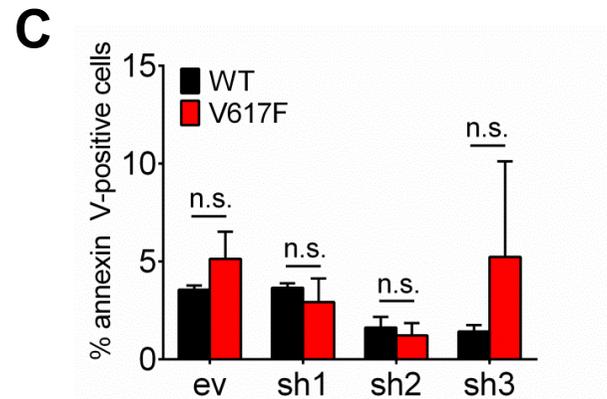
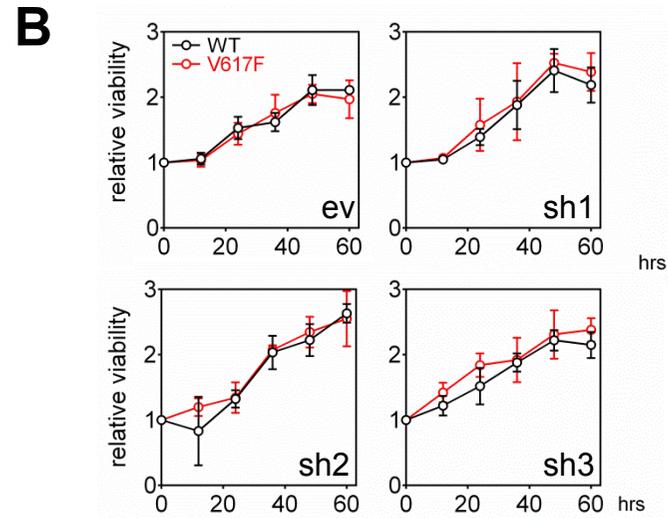
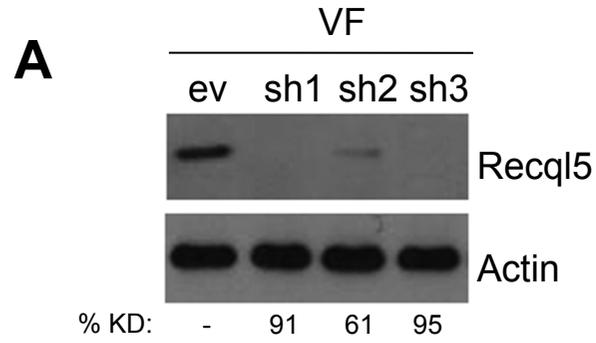


Figure S2

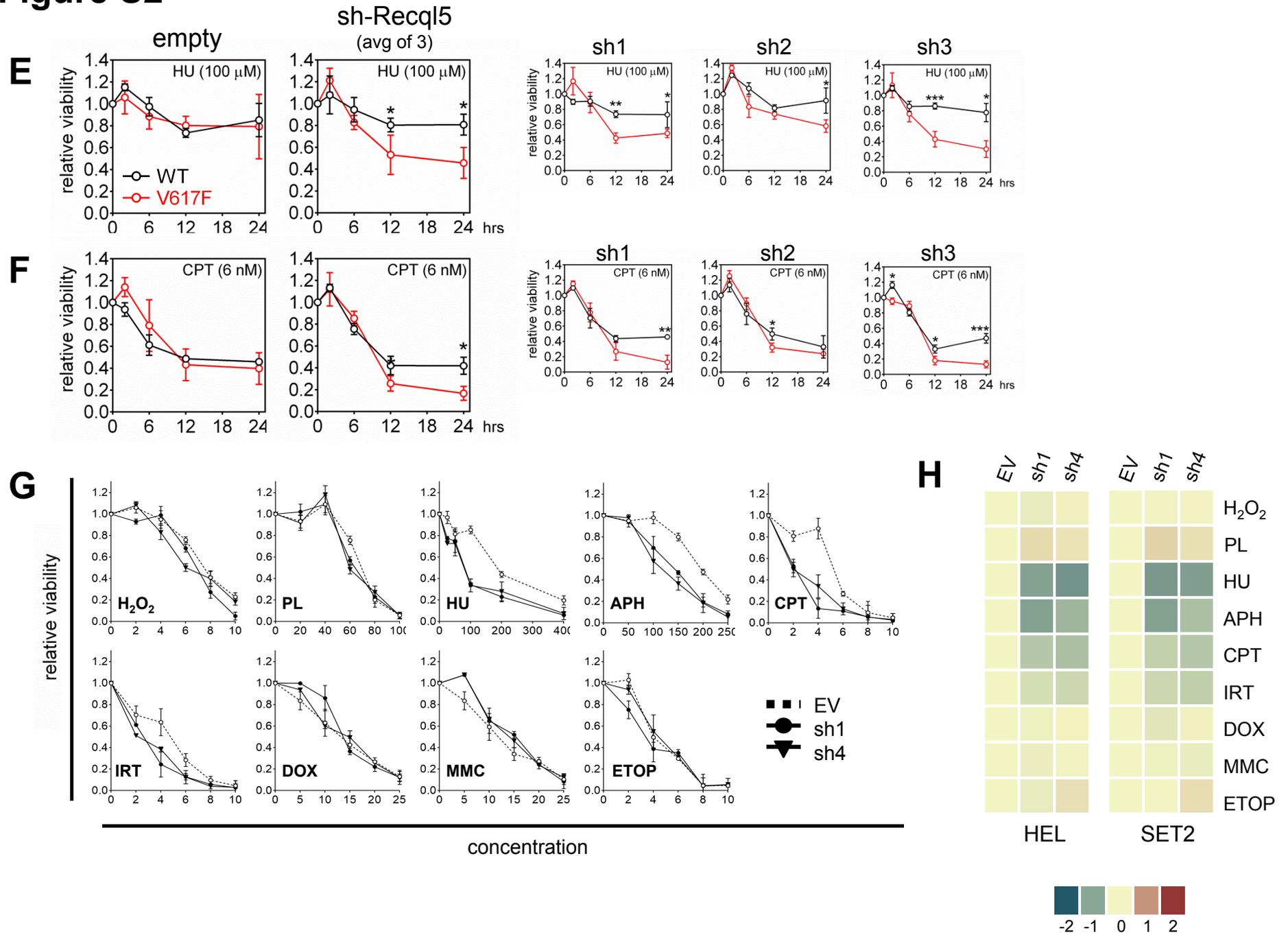


Figure S2 (Related to Figure 2). Recq15 depletion sensitizes Jak2V617F-expressing cells to replication stress. (A) Recq15 immunoblot of VF-B8 cells transduced with empty vector (ev) or 3 independent Recq15-targeting shRNAs (sh1-3). **(B)** Growth of WT-B8 and VF-B8 cells transduced with empty vector (ev) or Recq15-targeting shRNAs (sh1-3). **(C)** Quantification of annexin V-positive cells in WT-B8 and VF-B8 cells transduced with empty vector (ev) or Recq15-targeting shRNAs (sh1-3). **(D)** Assessment of viability of WT-B8 and VF-B8 cells transduced with empty vector or 3 different Recq15-targeting shRNAs following serum deprivation (SD), serum deprivation supplemented with 100 mM dNTPs (SD+dNTPs), or treatment with hydroxyurea (HU; 0-250 mM), camptothecin (CPT; 0-10 nM), doxycyclin (DOX; 0-25 mM) or etoposide (ETP; 0-10 nM). Each point represents the mean of 3 independent cultures. **(E-F)** Time-course assessment of viability of WT-B8 and VF-B8 cells transduced with empty vector or 3 different Recq15-targeting shRNAs following treatment with HU (6 mM) (Panel E) or CPT (4 nM) (Panel F). The average of the 3 Recq15-targeting shRNAs and for each individual hairpin is depicted. **(G)** Assessment of viability of SET-2 cells transduced with empty vector or 2 different RECQL5-targeting shRNAs (sh1 and sh4) following hydrogen peroxide (H₂O₂; 0-10 mM), piperlongumine (PL; 20-100 nM), hydroxyurea (HU; 0-400 mM), aphidicolin (APH; 0-250 mM); camptothecin (CPT; 0-10 nM), irinotecan (IRT; 0-10 nM); doxorubicin (DOX; 0-25 mM); mitomycin C (MMC; 0-25 nM) or etoposide (ETP; 0-10 nM). Each point represents the mean of 3 independent cultures. **(H)** Heat map of relative LC₅₀ of HEL and SET-2 cells to a panel of genotoxic agents. For each drug, the relative IC₅₀ of RECQL5-depleted cultures is calculated relative to EV-transduced cultures, and depicted as a ratio in the heat map. **(C-F)** Testing for statistical significance was performed using a student's t-test (*: p<0.05; **: p<0.01; ***: p<0.001).

Figure S3

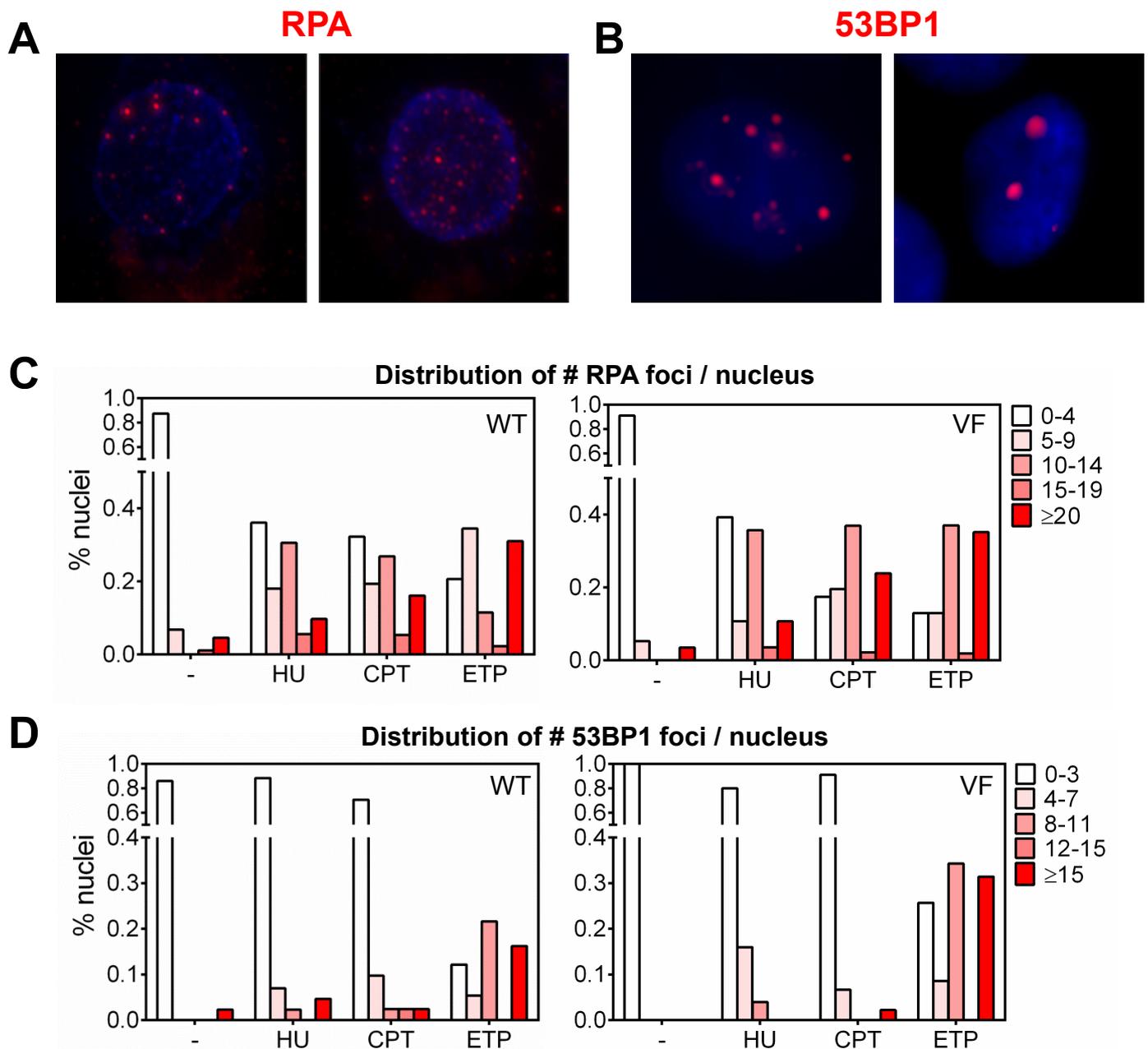


Figure S3 (Related to Figure 2). Specific doses of HU and CPT preferentially induce replication stress instead of generation of DSBs. (A-B) Representative immunofluorescent staining of RPA foci (Panel A) or 53BP1 foci (Panel B). **(C-D)** Histogram of numbers of RPA foci (Panel C) or 53BP1 foci (Panel D) of WT-B8 (WT) or VF-B8 (VF) cells treated with hydroxyurea (HU) (6 mM), camptothecin (CPT) (4 nM) or etoposide (ETP) (4 nM) for 2 hours.

Figure S4

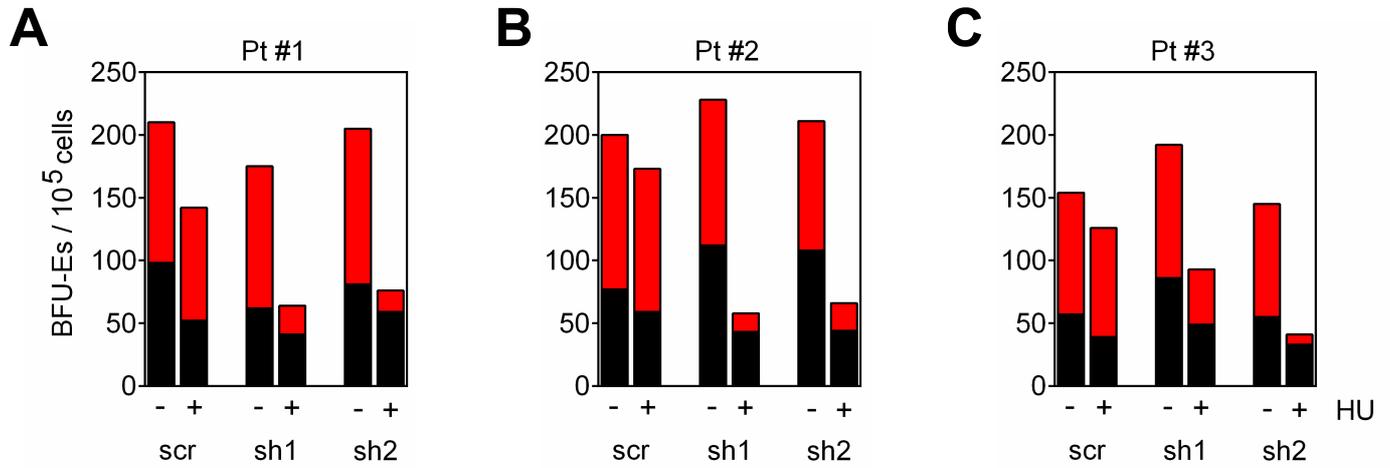


Figure S4 (Related to Figure 4). RECQL5 depletion sensitizes JAK2V617F-positive cells from MPN patients to replication stressors. (A-C) CD34-positive peripheral blood mononuclear cells from 3 myelofibrosis patients were transduced with empty vector (scr) or RECQL5-targeting shRNA (sh1 and sh2) and cultured on methylcellulose in the presence and absence of 2 mM hydroxyurea (HU). After 14 days, BFU-Es were counted, collected and genotyped for JAK2V617F.

SUPPLEMENTAL EXPERIMENTAL PROCEDURES

Detailed description of myeloid progenitor cell line generation from Jak2V617F-knockin mice

Ficoll-enrichment of bone marrow mononuclear cells was performed using a CD117-biotin antibody (BD Biosciences) and cells were expanded for 2 days in cytokine-rich media (IMDM+10% FBS supplemented with 10 ng/ml mL-3, 10 ng/ml mL-6 and 50 ng/ml mSCF). Cells were then spin-infected with packaged retroviruses, and serially passaged for 21 days in Myeloid Medium (RPMI+10% FBS supplemented with 50 ng/ml mSCF and 1 μ M β -estradiol (Sigma)), after which polyclonal, conditionally immortalized myeloid progenitor cell lines from wild-type mice (WT-B8) and Jak2V617F-knockin mice (VF-B8) were stable in culture.

Cells and reagents

293T cells were maintained in DMEM containing 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin. SET-2 and HEL cells were maintained in RPMI containing 10% FBS and 1% penicillin/streptomycin. Cells were treated with the following chemicals: INCB018424 (Incyte), hydroxyurea (Sigma), camptothecin (Selleck), doxorubicin (Sigma), etoposide (Sigma), aphidicolin (Sigma), hydrogen peroxide (Sigma), piperlongumine (Cayman Chemical Company), irinotecan (Sigma) and mitomycin C (Sigma).

Alamar blue assay for measurement of cell viability

Each sample was incubated with 10 μ L Alamar blue solution (giving final concentration of 10% v/v) and incubated at 37°C for 4 hours. After incubation, cells were centrifuged at 2,000 rpm for 10 min and the supernatants were transferred to a new 96-well plate. The fluorescence was measured by a micro-plate reader in arbitrary fluorescent units following excitation at 530-560 nm and emission at 590 nm. Fluorescence intensity of each sample is

depicted as a ratio relative to an untreated control culture and designated as the viability fold change.

Chromosome combing calculations and statistical analyses

Fibre lengths were measured in micrometres and converted to kilobases according to a constant stretching factor of $1 \mu\text{M} = 2 \text{ kb}$, as previously reported (Herrick and Bensimon, 2009). A stalled fork is defined as a >30% reduction in fork progression in the second labelling step relative to the first. An asymmetrical origin is defined as a ratio between the two oppositely moving arms of the origin structure as <0.7 . Testing for statistical significance was performed on the ratio between the two arms using a single-sample t-test using a predicted population mean of 1. Fork rate was calculated by measuring the combined red and green tracts of progressing structures and dividing by the total labelling time (40 min).

Vectors

All lentiviral pLKO.1 shRNA vectors were obtained from the Broad Institute TRC shRNA library: Jak2 (TRCN0000278124; TRCN0000278126), Stat5a (TRCN0000231566; TRCN0000231567), Stat1 (TRCN0000235836; TRCN0000235835), Pik3ca (TRCN0000234002; TRCN0000025617), JAK2 (TRCN0000315249; TRCN0000381098), STAT5A (TRCN0000232134; TRCN0000232133), STAT1 (TRCN0000280021; TRCN0000280022), PIK3CA (TRCN0000350276; TRCN0000234003); RECQL5 (TRCN0000236027; TRCN0000236028). The lentiviral vectors containing Recql5-targeting shRNAs were obtained from OriGene (Gene ID 170472; TR506050).

Generation of lentiviral particles and lentiviral infections

For generation of lentiviral particles, 3×10^6 293T cells were seeded per 10 cm dishes overnight, followed by transfection with pLKO expression vectors, the lentiviral packaging constructs psPAX2 (Gag-Pol) and pMD2.G (VSV-G) in a 7:4:4 ratio. For each 10 cm dish, 15

μ g total DNA was transfected with 45 μ L TransIT transfection reagent (Mirus), according to manufacturer's protocols. Viral supernatants were collected at 24 hours and 48 hours, pooled and concentrated using LentiX Concentrator. Viral pellets were resuspended in sterile 1X PBS and used immediately or snap-frozen in liquid nitrogen and stored in -80°C for up to 3 months.

To perform lentiviral infections, cells were spin-infected in 6-well plates as follows: 1.5×10^5 cells/well were seeded in growth media with concentrated lentivirus and 8 $\mu\text{g/ml}$ polybrene, and the cells were centrifuged at 2,000 rpm for 2 hours at 37°C . After spin-infection, supernatants were immediately aspirated off and fresh media was added. Cells were cultured for at least 24 hours prior to drug treatment experiments or validation of gene knockdown by Western immunoblot or qPCR.

Real-time qPCR analysis

Total RNA were isolated using the RNeasy RNA isolate kit with on-column DNase digestion (Qiagen). Real-time qPCR was performed using the Fast Sybr Green Master Mix (Applied Biosystems). All PCR primers were purchased from Integrated DNA Technologies: Recq1 (fwd: CTCTGTGCTATCAACTCCCTG; rev: CTCCTGTACCGAGAGCTTTG); Recq15 (fwd: CTCTGTGCTATCAACTCCCTG; rev: CTCCTGTACCGAGAGCTTTG); Blm (fwd: TGGATCAGAAAGCATCACCC; rev: GTAAAGTGTCAGCCATTGTGTC); Wrn (fwd: AACATCGACACGTACCTCATC; rev: CACCTCTTTGCTCTCCTTACAG); Recq14 (fwd: ACCTTTGGGACAAGTAGCAG; rev: GGGCAACACTAACAGAGTAGAG); RECQL5 (fwd: CCCC GAAGTACTTGGCAAT; rev: AACAAAGCATCTGATAAAGCCA); RECQL1 (fwd: GCTCTTGTTCCTTTCCGTA; rev: AAAGAATGGCGTCCGTTTC).