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Article:

Khamis, K, Brown, LE, Hannah, DM et al. (1 more author) (2015) Experimental evidence that predator range expansion modifies alpine stream community structure. *Freshwater Science*, 34 (1). 66 - 80. ISSN 2161-9549

<https://doi.org/10.1086/679484>

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1 LRH: Predation in alpine stream mesocosms K. Khamis et al.

2 RRH: Volume 34 March 2015

3

4 **Experimental evidence that predator range expansion modifies alpine stream**
5 **community structure**

6

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17 Received 11 December 2013; Accepted 27 May 2014; Published online XX Month 2014.

18 **Abstract:** Climate change is projected to facilitate altitudinal range expansions of ‘lowland’
19 taxa, creating novel species interactions. However, how range shifts will alter biotic
20 interactions and community structure in alpine streams is not well understood. In the
21 Pyrénées, climate-induced physicochemical habitat change is hypothesized to facilitate the
22 colonization of high-altitude streams by *Perla grandis*, a carnivorous stonefly. A field-based
23 experiment was done in mesocosm channels beside a hillslope spring (2000 m asl) in the
24 Taillon-Gabiétous catchment, French Pyrénées. The influence of *P. grandis* predation on
25 community structure, feeding trait composition, body-size spectrum, and algal chlorophyll a
26 concentration was examined. Gut contents were analyzed and used to identify consumed
27 prey. Total invertebrate density was not significantly reduced by *P. grandis*, but *Baetis* spp.
28 densities were depressed in the treatment channels through a combination of direct
29 consumption and predator avoidance (emigration/drift). However, despite fewer grazers in
30 the predator treatment channels, the magnitude of the trophic cascade effect on basal
31 resources (measured as chlorophyll a density) was comparable between treatment and control
32 channels. The results of this experiment suggest that size/species specific predation,
33 intraguild predation, and interference competition are the likely mechanisms that altered the
34 body-size spectrum in treatment channels. In synergy with climate-driven physicochemical
35 habitat change, the extinction risk of some range-restricted taxa (prey and other predators)
36 could be increased where *P. grandis* colonization occurs. Hence, conservation efforts are
37 required to ensure that additional anthropogenic stressors (e.g., nutrient enrichment, cattle
38 trampling, hydropower development, ski runs, and tourism) are limited to minimize further
39 pressures on these unique and sensitive habitats.

40 **Key words:** climate change, mesocosm, intraguild predation, *Perla grandis*, Pyrénées, food
41 web, body size, enemy release

42

43 Future climate warming will alter ecosystem processes, biotic patterns, and
44 interactions across a range of spatial and temporal scales (Parmesan 2006, Bellard et al.
45 2012), which in turn, will influence a wide variety of habitat types and taxonomic groups
46 (Dirzo and Raven 2003, Thomas et al. 2004, Xenopoulos et al. 2005, Engler et al. 2011,
47 Sauer et al. 2011). Mountain environments support unique biota (Brown et al. 2009, Engler et
48 al. 2011) that are often at their range limits. These taxa are particularly vulnerable to warming
49 because current and predicted warming rates in high-altitude environments are typically
50 higher than the global mean (Beniston 2012). Altitudinal range shifts or expansions, although
51 less well documented than latitudinal expansions (Parmesan 2006), have been observed for a
52 variety of species (Pauli et al. 2007, Chen et al. 2011). However, in freshwater ecosystems,
53 the spatial and temporal variability in trophic interactions make predictions about range shifts
54 based on species–environment relationships potentially problematic (Woodward et al. 2010).

55 The sensitivity of alpine benthic stream ecosystems to climate change and range
56 expansions has been highlighted increasingly in recent years (Muhlfeld et al. 2011, Sauer et
57 al. 2011, Jacobsen et al. 2012, Khamis et al. 2014). Many range-restricted benthic organisms
58 will be the ‘losers’ of future climate change (Somero 2010) because they will be unable to
59 respond spatially or physiologically to water temperature increase (Bellard et al. 2012).
60 Synergistic effects of climate/hydrological change and altered biotic interactions are likely to
61 promote extinctions in pristine alpine river habitats (Tierno de Furoa et al. 2010). Warming
62 will cause changes in meltwater dynamics, and thus, stream discharge (magnitude and
63 variability), water temperature, and stream channel stability will be altered (Brown et al.
64 2007a, Jacobsen et al. 2012) making habitats more favorable to some species, e.g., upstream
65 colonization by more downstream or ‘lowland’ taxa (Brown et al. 2007a).

66 The type and strength of interactions between ‘invaders’ and ‘native’ taxa can have
67 important implications for both biotic interactions and foodweb properties (Woodward et al.

68 2005). Woodward and Hildrew (2001) presented one of the few examples of (invertebrate)
69 predator invasion effects on a low-order stream system and showed an increase in biotic
70 interactions and foodweb trophic height after the invasion of a large-bodied dragonfly
71 (Woodward and Hildrew 2001). In the context of low-order alpine streams, perturbations
72 caused by predator invasion are likely to have significant effects because these food webs are
73 characterized by high levels of omnivory and dietary overlap (Zah et al. 2001), with relatively
74 high connectance and short food-chain lengths (Clitherow et al. 2013). Hence, factors
75 affecting one node may propagate quickly through the network, with potential for cascading
76 effects (Shurin et al. 2002). Therefore, in light of the rapid rate of environmental change
77 predicted for alpine river ecosystems, the potential implications of predator range expansion
78 on biotic interactions in these systems needs to be quantified (Milner et al. 2009).

79 *Perla grandis* (Plecoptera:Perlidae), a rheophilous mesothermal predatory stonefly, is
80 widely distributed across the mountain ranges of southern and central Europe (Fenoglio et al.
81 2008) and is common throughout mid-altitude Pyrenean streams where it is the top
82 invertebrate predator (Lavandier and Decamps 1984, Vincon 1987). In the alpine zone, its
83 distribution appears to be limited by stream water-source contribution and altitude because
84 higher-altitude streams, fed predominantly by melting snow or ice in spring, have a habitat
85 template that is unsuitable (i.e., low water temperature, unstable beds, and high turbidity;
86 Brown et al. 2007b). Sparse data exist regarding the feeding habits and ecology of *P. grandis*
87 nymphs. The only study is from a mid-elevation stream (800 m asl) in the Appenines,
88 northwestern Italy. Fenoglio et al. (2007) reported that the principal prey were chironomids,
89 *Baetis* spp., and other Ephemeroptera. However, work on a similar species (*Dinocras*
90 *cephalotes*:Perlidae) identified other large-bodied, predatory invertebrates as a dietary
91 component (Bo et al. 2008). As glacier and snow packs recede, *P. grandis* is expected to
92 expand its range upwards into lower-order, higher-altitude streams (Brown et al. 2007a).

93 However, brown trout (*Salmo trutta*), a potential competitor/predator of *P. grandis*, will be
94 unable to track such range expansion because of migratory barriers, such as waterfalls.
95 Hence, the likely decoupling of this historical trophic interaction suggests that *P. grandis* will
96 experience ‘enemy release’, with significant effects on prey and predators/competitors in the
97 invaded community (Gilman et al. 2010, Sih et al. 2010).

98 A field-based experimental approach was taken to assess potential effects of climate-
99 driven range expansion (i.e., change in river water-source dynamics) of the stonefly *P.*
100 *grandis* on stream benthic communities. Artificial through-flow mesocosm channels fed by a
101 1st-order stream were used to test 4 complementary hypotheses:

102 H₁: The abundance of key prey taxa will be reduced in *P. grandis*-invaded systems
103 through either direct consumption or nonconsumptive effects (e.g., increased drift).

104 H₂: Community feeding-guild structure will be altered by *P. grandis* invasion through
105 prey selection of active grazing taxa and interference competition with other large bodied
106 predators.

107 H₃: *Perla grandis* immigration will alter population and community body size
108 distributions through increased predation pressure and competition with resident
109 predators.

110 H₄: *Perla grandis* will increase the magnitude of the trophic cascade by decreasing prey
111 abundance (H₁), particularly active grazing taxa (H₂).

112

113 **METHODS**

114 **Study site, experimental channels, and experimental organisms**

115 Our study was conducted in the Taillon-Gabiétous catchment, Cirque de Gavarnie,
116 French Pyrénées (lat 43°6'N, long 0°10'W) between 4 July and 27 July 2011. A detailed
117 description of the study basin was provided by Hannah et al. (2007). Briefly, the catchment

118 lies above the tree line (i.e., alpine zone), and has steep slopes (30–70°) and a sedimentary
119 geology. On south-facing slopes a number of groundwater-fed streams are sourced from
120 hillslope, alluvial aquifers. Here, allochthonous inputs are limited to grasses and sedges.
121 Thus, the basal resources in these systems consist of primarily diatoms and benthic algae.
122 *Perla grandis* is currently absent from streams >1900 m asl.

123 To emulate groundwater-fed stream habitat, experimental channels were situated
124 beside a 1st-order stream, ~2000 m asl (Fig. 1A, B), where *P. grandis* was absent. Water was
125 diverted from the stream via a feeder pipe (diameter = 10 cm) into 2 plastic header tanks (0.6
126 m × 0.4 m × 0.3 m). Each tank fed a block of 4 channels directly through equally spaced gate
127 valves. Channels were made from rectangular plastic gutter (1 m × 0.13 m × 0.17 m) with a
128 0.05-m-diameter outflow pipe (Fig. 1A) through which water flowed into a drainage pipe that
129 returned water to the main stream channel. Drift nets (250- μ m mesh) attached to the outflow
130 pipes captured emigrating *P. grandis*, which were then returned to the channels, and other
131 emigrating taxa. Gravel (5–25 mm) and pebbles (25–45 mm) were collected from the feeder
132 stream, elutriated thoroughly to ensure attached organisms and eggs were removed, and then
133 used to fill the channels to a depth of 5 cm. Five cobbles (intermediate axis > 100 mm) were
134 then placed in each channel, and the attached biofilm was inspected, and macroinvertebrates
135 removed with forceps.

136 Flow velocity was recorded on 5 occasions in the center of each channel (0.6 depth
137 from the water surface) using a Sensa RC2 electro-magnetic flow meter (Sensa, Aqua Data
138 Services, Calne, UK). The velocity across all channels and dates was similar (mean \pm SE,
139 0.13 ± 0.03 m/s, $n = 40$). Flow velocity was low, but the channels represented riffled habitat
140 because at the low water depth and water volume in channel, broken water was observed over
141 the substrate. Electrical conductivity (EC) and pH (measured using a Hanna HI 98129
142 handheld probe; Hanna Instruments, Woonsocket, USA), was also recorded on 5 dates. Both

143 EC ($177 \pm 2 \mu\text{S}/\text{cm}$, $n = 40$) and pH (8.62 ± 0.3 , $n = 40$) were similar between channels and
144 dates. Water temperature was recorded continuously (15-min intervals for the duration of the
145 experiment) in 1 channel from each block using a Gemini Tinytag Plus (instrument error \pm
146 0.2°C ; Gemini, Chichester, UK). Mean water temperature was comparable ($10.2^\circ\text{C} \pm 0.9$, $n =$
147 2208) between blocks and was lower than the Tourettes stream ($11.4^\circ\text{C} \pm 2.7$, $n = 2208$; KK,
148 unpublished data) where the experimental *P. grandis* were collected. Water depth was similar
149 throughout all channels and ranged from 6 cm above fine gravel to 2 cm above cobbles.
150 Before initiating the experiment, flow in the channels was established for 3 d with the inflow
151 pipe covered by 100- μm mesh, to inhibit colonization by drifting invertebrates while
152 allowing algal colonization (Ledger et al. 2006).

153 *Perla grandis* ($n = 33$) were collected from the nearby Tourettes stream (1800 m asl;
154 Fig. 1B) from which 8 well developed nymphs of equal body length (mean \pm SE, $22.05 \pm$
155 0.66 mm) were selected for the experiment. The remaining nymphs ($n = 25$) were retained for
156 gut-content analysis and stored in 70% ethanol.

157

158 **Experimental design**

159 The experiment followed a systematically balanced design with 2 treatments
160 replicated 4 times: 1) a predator-free control, and 2) a *P. grandis* treatment (2 individuals
161 [ind]/channel), corresponding to a density of $15 \text{ ind}/\text{m}^2$. The ambient density of *P. grandis* in
162 the Tourettes stream was between 10 and $30 \text{ ind}/\text{m}^2$. All *P. grandis* nymphs were placed in
163 the treatment channels on 4 July 2011, and the mesh was removed from the inlet pipe to
164 enable colonization from the proximal stream species pool. Drift nets attached to the outflow
165 pipes were inspected at least every 48 h for *P. grandis*. Invertebrate emigration via drift was
166 sampled on 3 occasions (10–11, 18–19 and 26–27 July) for 24-h periods. Drift nets (250- μm
167 mesh) were emptied into Whirlpak[®] bags and stored in 70% ethanol. The experiment was

168 terminated after 23 d on 27 July 2011.

169 At the end of the experiment, 3 cobbles were selected from each channel for
 170 chlorophyll a (chl a) analysis, and all attached invertebrates were removed with forceps.
 171 Epilithon was removed from the surface of cobbles in the laboratory with a stiff toothbrush in
 172 50 mL of deionized water. The resultant slurry was drawn through a Whatman GF/F filter
 173 paper (0.7 μm ; Whatman, Maidstone, UK) and frozen immediately. Samples were analyzed
 174 for chl a with the trichromatic spectroscopy method outlined in ASTM D3731 (ASTM 2004).
 175 Pigment extractions were made in 90% acetone. Periphyton concentrations were converted to
 176 mass per unit area (mg/m^2) as

$$177 \quad \text{Chl } a = \frac{CaE}{A} \quad (\text{Eq. 1})$$

178 where Ca = concentration of chlorophyll a in the extract (mg/L), E = extract volume (L), and
 179 A = substrate area sampled (m^2).

180 All substrate from each experimental channel was transferred into a bucket, sieved,
 181 and the remaining organic material and invertebrates transported to the laboratory where
 182 macroinvertebrates were sorted from gravel under a bench lamp and stored in 70% industrial
 183 methylated spirit (IMS). Individuals from both the drift and end-point community were
 184 identified to the lowest practical taxonomic level (i.e., 60% of all individuals were identified
 185 to genus level or below) with the aid of a selection of taxonomic keys (Müller-Liebenau
 186 1969, Tachet et al. 2000, Zwick 2004) and assigned to functional feeding groups following
 187 Moog (1995). For the most abundant taxa ($>15 \text{ ind}/\text{m}^2$) and larger bodied predators, body
 188 length (mm) of all individuals in each taxon was measured on a dissecting microscope at 6.5
 189 to 75 \times magnification (Stemi 2000-C; Ziess, Oberkochen, Germany) fitted with an eye-piece
 190 graticule. Gut contents of the mesocosm P. grandis (n = 8) and the individuals collected from
 191 the Tourettes stream (n = 25) were assessed. The entire gut was dissected and placed on a
 192 slide, and the contents were dispersed and identified on a dissecting microscope (6.5–75 \times) or

193 a stage microscope at 100 to 1000× magnification (Optiphot-2; Nikon, Tokyo, Japan) as
194 necessary. Partial specimens of consumed prey were counted only if the head was attached to
195 the thorax.

196

197 Data analysis

198 All abundance data were standardized before statistical analysis to ind/m² and the
199 relative abundance (%) of functional feeding groups was calculated. Taxonomic richness
200 (i.e., number of taxa) and community dominance (Berger–Parker dominance index) were
201 calculated for each channel using nontransformed abundance data. Drift propensity was
202 calculated for the final sampling date (24–25 July) by dividing the number of drifting
203 individuals of a given taxon by the total number of individuals of that taxon recorded in the
204 channels.

205 Chl a, macroinvertebrate abundance, drift propensity, and functional feeding group
206 data were tested for normality using a combination of QQ plots and the Shapiro–Wilk test.
207 Variances were tested for homogeneity with Levene’s test (Zuur et al. 2010). Subsequently,
208 Chl a and abundance data were $\sqrt{(x + 0.5)}$ -transformed, relative abundance data were
209 $\arcsin\sqrt{(x)}$ -transformed, and drift propensity data were $\log_{10}(x + 1)$ -transformed to meet
210 assumptions of parametric analysis.

211 Because of the large number of statistical tests reported herein, null hypothesis testing
212 was complimented by the calculation of standardized effect size (SES; McCabe et al. 2012).
213 Unlike the application of stringent Bonferroni corrections, which reduce power and increase
214 type II errors to unacceptable levels (Nakagawa 2004), calculation of SES enables biological
215 importance and significance to be assessed simultaneously and reduces publication bias of
216 selective reporting of results (Garamszegi 2006). Cohen’s d (SES) for each test was
217 calculated as:

218
$$d = \frac{\bar{X}_1 - \bar{X}_2}{S^2} \quad (\text{Eq. 2})$$

219 where \bar{X}_1 is the mean of the control, \bar{X}_2 is the mean of the treatment, and S^2 is the pooled
 220 standard deviation (SD) calculated as:

221
$$S^2 = \frac{(n_1 - 1)S_1^2 + (n_2 - 1)S_2^2}{n_1 + n_2} \quad (\text{Eq. 3})$$

222 where S_i^2 is the SD of the of the i^{th} group, and n_i is the sample size. Confidence intervals
 223 (95%) were calculated for d in the R environment (version 2.14.1; R Project for Statistical
 224 Computing, Vienna, Austria) using script created by Smithson (2011) that are freely available
 225 for download (<http://psychology3.anu.edu.au/people/smithson/details/CIstuff/CI.html>).

226 To test H_1 (the abundance of key prey taxa will be reduced in *P. grandis*-invaded
 227 systems through either direct consumption or nonconsumptive effects), prey selection was
 228 estimated using the raw abundance data from the treatment channels. The Log of the Odds
 229 Ratio (LOR) was calculated following Brodner (1998) as

230
$$LOR = \ln \frac{g_i(100 - g_i)}{c_i(100 - c_i)} \quad (\text{Eq. 4})$$

231 where g_i is the relative abundance of taxon i found in the gut of *P. grandis* and c_i is the
 232 relative abundance of taxon i found in the treatment channels. This ratio varies from $-\infty$ to
 233 $+\infty$, and positive values represent preference.

234 The most abundant prey items (Fig. 2) were identified based on gut contents from our
 235 study (Table 1) and other relevant studies (Fenoglio et al. 2007, Bo et al. 2008). One-way
 236 analysis of variance (ANOVA), with block included as an additive variable (c.f. model 2;
 237 Newman et al. 1997), was used to identify treatment effect on the density of the most
 238 abundant potential prey taxa and the drift propensity. Hence, the error term for treatment
 239 effect in both these tests was the residual mean square (Newman et al. 1997).

240 Nonmetric dimensional scaling (NMDS), based on Bray–Curtis dissimilarity, was
 241 used to assess the effect of treatment on prey community structure. Analysis of similarity

242 (ANOSIM) was adopted to test whether the 2 treatments had different taxonomic
243 compositions (i.e., dissimilarities between treatments greater than within treatment).

244 To explore H₂, 1-way ANOVA (with block as an additive effect) was carried out to
245 identify treatment effects on functional feeding group density and functional feeding group
246 relative abundance. Predatory taxa that colonized the channels were split into 2 groups.
247 Individuals >10 mm body length were classified as large-bodied predators and those <10 mm
248 as small bodied (c.f. Ilg and Castella 2006).

249 To explore the first component of H₃ (P. grandis would alter population-level body-
250 size distributions) Student's t-tests were used to test for treatment effect on mean body length
251 of the most abundant prey taxa (Baetis gemellus, Baetis muticus, Simulium sp., and
252 Orthocladiinae). Raw body-size data were used for statistical analysis because they were
253 normally distributed and displayed homogeneity of variance. Violin plots (a combined box
254 plot and kernel density plot) were used to assess body length differences between treatments
255 because they display more information on data spread than box plots alone. The probability
256 density function of the data at different values is shown, much like a histogram, but each
257 block is centered at each data point rather than fixed in the form of class bins (Hintze and
258 Nelson 1998).

259 To test the 2nd component of H₃ (P. grandis would alter community-level body-size
260 distributions), body mass (ash free dry mass [AFDM], mg) was calculated for the measured
261 taxa using published length–mass regressions (Table S1). The body mass data for all taxa
262 were then pooled and log₁₀(x)-transformed to explore the effect of treatment on community-
263 level body-size distribution (Brown et al. 2011). Transformed biomass data were sorted into 5
264 size classes (< -1.3, -1.3 to -0.7, -0.7 to -0.1, -0.1 to 0.5, > 0.5 mg). Treatment response was
265 calculated as the % difference between treatment and control for each size class. Pairwise
266 comparisons were used to calculate the mean difference and standard error (SE) for each size

267 class. Each treatment channel was compared to all control channels, hence $n = 16$ for the
268 mean and SE calculations. A Kruskal–Wallis test was used to test for treatment effect on the
269 number of individuals in each body-mass class.

270 To test H_4 , 1-way ANOVA (with block as an additive effect) was carried out to
271 identify treatment effect on Chl a density. All tests were considered significant at $p < 0.05$.
272 All plots, t-tests, ANOVA, and multivariate analyses were made using the base and vegan
273 (Oksanen et al. 2012) packages in R (version 2.14.1).

274

275 **RESULTS**

276 **Taxonomic composition and community structure of the mesocosm channels**

277 *Perla grandis* individuals acclimatized rapidly to the channels, and only 2 individuals
278 were returned from the drift nets to the channels (i.e., no mortality recorded). A total of 33
279 aquatic invertebrate taxa (excluding Collembola) colonized the experimental channels (Table
280 S2). These taxa were representative of the source stream community (KK, unpublished data),
281 and included predominantly several Diptera, Ephemeroptera, and Coleoptera (larvae and
282 adults). Trichoptera were more abundant in the control than in the treatment channels, and
283 Plecoptera taxa were rare in both treatment and control channels. Oligochaeta and the triclad,
284 *Polycelis* sp., were the only noninsect taxa to colonize the channels.

285 *Baetis gemellus* was the most abundant taxon in control channels, whereas
286 Orthocladiinae dominated in treatment channels (Table S2). Eight insect taxa (*B. gemellus*, *B.*
287 *muticus*, *Baetis alpinus*, *Simulium* sp., *Elmis* sp., Orthocladiinae, Diamesinae, and
288 Tanypodinae) were recorded at mean densities >10 ind/m² (Table S2), whereas other taxa
289 were recorded in low densities (<10 ind/m²). Large-bodied species, other than *P. grandis*,
290 were rare and consisted of 5 taxa: 2 dipteran taxa (Tipulidae and Rhagionidae, <1 ind/m²,
291 across all channels) and 3 predatory trichopterans, *Rhyacophila* spp., *Rhyacophila*

292 intermedia, and *Plectrocnemia* sp. *Rhyacophila* spp. were found exclusively in the treatment
 293 channels (mean \pm SE, 2 ± 2 ind/m²), whereas *R. intermedia* and *Plectrocnemia* sp. were
 294 recorded in higher densities (14 ± 5 ind/m² and 8 ± 3 ind/m², respectively) but exclusively in
 295 the control channels. The body sizes of *R. intermedia* (12.87 ± 0.75 mm) and *Plectrocnemia*
 296 sp. (11.21 ± 0.67 mm) were markedly smaller than *P. grandis* (22.05 ± 0.66 mm).

297 Total macroinvertebrate density was greater on average in control (1043 ± 65 ind/m²)
 298 than in treatment channels (812 ± 73 ind/m²). The treatment effect, though marginally
 299 nonsignificant (ANOVA, $F_{1,5} = 5.44$, $p = 0.058$) was large ($d = -1.56$, CI = -3.18 – 0.001).
 300 Mean species richness of the treatment and control channels was comparable (15.25 ± 0.75
 301 and 14.75 ± 0.68 , respectively). Community dominance was lower in treatment (0.29 ± 0.01)
 302 than control channels (0.39 ± 0.01). This effect was large ($d = -4.7$, CI = -7.4 to -2.0) and
 303 significant (ANOVA, $F_{1,5} = 34.38$, $p = 0.0006$).

304

305 **H₁: The effect of *P. grandis* on invertebrate prey abundance and drift**

306 The average number of prey/*P. grandis* gut was similar in the experimental channels
 307 (3.6 ± 0.5) and the Tourettes stream (3.4 ± 0.4), and the diet was entirely carnivorous. All
 308 prey items were insects, and *Baetis* spp. and *Orthocladiinae* were the most frequent.
 309 However, *Baetis* spp. were proportionally more abundant in the guts of *P. grandis* in the
 310 experimental channels (Table 1). Large-bodied prey items also were recorded, and entire
 311 *Rhyacophila* spp. and *Limnephilidae* larvae were found in the foreguts of *P. grandis* from the
 312 Tourettes stream. *Perla grandis* had a positive LOR and selectivity for *Protonemura* spp. in
 313 the channels ($+2.85$) and the Tourettes stream ($+1.62$), *Baetis* spp. ($+0.26$) in the
 314 experimental channels, and *Orthocladiinae* ($+1.35$) in the Tourettes stream (Table 1). High
 315 positive LOR values also were apparent for *Rhyacophila* spp ($+1.37$) and *Limnephilidae*
 316 ($+1.13$) in the Tourettes stream. *Perla grandis* displayed no selectivity (negative LOR values)

317 for Simuliidae in either the experimental channels or Tourettes stream (Table 1).

318 The abundances of the mayflies, *B. gemellus* and *B. muticus*, were significantly lower
319 in the treatment than in the control channels (Table 2) and the effect size was large (Fig. 2).
320 Abundance of *B. alpinus* was significantly greater in the treatment than in the control
321 channels (Table 2, Fig. 2). Other common prey taxa (i.e., Orthocladiinae and Diamesinae)
322 showed no significant response to the treatment (Table 2, Fig. 2). The most stable solution
323 (stress = 0.04) of the NMDS analysis of the abundant prey taxa (>15 ind/m²) consisted of 2
324 dimensions. The treatment and control channels were divided along axis 1 (Fig. 3), and the
325 stress plot indicated the loss of a negligible amount of the variation in the original data set (R^2
326 = 0.99). Treatment significantly affected prey community structure (ANOSIM, $R = 0.47$, $p =$
327 0.02). Control channels were characterized by greater abundance of *B. gemellus*, *B. muticus*,
328 and *Elmis* sp. (Fig. 3).

329 *Baetis alpinus* and *Protonemura* spp. were the most abundant taxa in the drift
330 compared to their channel densities (Fig. 4). Drift propensity rates for all other key prey taxa
331 (Orthocladiinae, *B. gemellus* and *B. muticus*) were low (<1) (Fig. 4). A positive treatment
332 effect was recorded for the drift propensity rate of *B. gemellus*, and a negative treatment
333 effect was recorded for *B. alpinus* (Table 3). No significant treatment effect was detected for
334 *B. muticus* and *Protonemura* spp. (Table 3).

335

336 **H₂: Effects of *P. grandis* on invertebrate feeding guild structure**

337 Grazers were the most abundant feeding guild and large predators were the least
338 abundant in control and treatment channels (Table 4). Filter feeders (predominantly *Simulium*
339 spp.) were numerically and relatively more abundant in treatment channels, as were predators
340 (predominantly Tanypodinae). The density and relative abundance of grazers was
341 significantly lower in treatment than in control channels (Table 4, Fig. 5). Shredder and

342 collector abundances were similar between treatment and control channels (Fig. 5). When
343 predators (excluding *P. grandis*) were divided into small- (5–10 mm) and large-bodied (>10
344 mm) classes, a significant negative effect of treatment was observed for the large-bodied
345 predators (Table 4, Fig. 5). The most common of these large-bodied predators, *R. intermedia*,
346 was more abundant in control (13 ± 5 ind/m²) than in treatment channels (2 ± 2 ind/m²), but
347 this effect was not significant (ANOVA, $F_{1,5} = 3.89$, $p = 0.096$).

348

349 **H₃: Effects of *P. grandis* on body-size spectrum**

350 The predator treatment had no significant effect on the body size of Orthoclaadiinae or
351 *Simulium* spp. individuals (Table 5). Mean body lengths of *B. muticus* and *B. gemellus* were
352 0.15 mm and 0.25 mm smaller in treatment than in control channels, respectively (Table 5).
353 However, effect sizes were smaller for both taxa ($d < 0.5$, $p < 0.1$; Table 5). Violin plots of *B.*
354 *gemellus* body lengths revealed an asymmetric density distribution, with fewer 5- to 6-mm
355 individuals and smaller median body size in the predator treatment channels (Fig. 6).

356 The body mass distributions for the control and treatment (including *P. grandis*)
357 channels were broadly similar but displayed some important differences: 1) distributions in
358 the treatment channels displayed strong right skew because *P. grandis* individuals were
359 considerably larger than any other taxa; and 2) there were fewer taxa between 1–10 mg in the
360 treatment than in the control channels (Fig. 7). When all taxa (excluding *P. grandis*) were
361 pooled and allocated to body-mass size classes, a significant negative treatment response was
362 observed for the largest class (>5 mg; Fig. 7).

363

364 **H₄: Cascading effects of *P. grandis***

365 Chl a content of the cobble biofilm was lower in the control (4.81 ± 0.86 mg/m²) than
366 in the treatment channels (7.13 ± 2.16 mg/m²), but the effect size was small ($d = 0.63$, CI =

367 0.15–0.95) and not significant (ANOVA, $F_{1,5} = 4.03$, $p = 0.092$).

368

369 **DISCUSSION**

370 Predation is a key biotic process in aquatic systems that can drive physiological and
371 behavioral responses of prey taxa (Peckarsky 1982) and alter community structure and
372 functioning (Jefferies and Lawton 1984, Sih et al. 1998). Our study identified a number of
373 community- and population-level responses to the experimental simulation of *P. grandis*
374 range expansion. The density of conspicuous grazing taxa (*Baetis* spp.) was depressed
375 through direct consumption and prey-avoidance mechanisms (Lancaster 1990). However, the
376 magnitude of the trophic cascade (basal resources estimated as Chl a) did not change,
377 probably because predators were present in the control (i.e., *R. intermedia* and *Plectrocnemia*
378 sp.). A negative shift in mean body size was apparent in the presence of *P. grandis* at the
379 population (*B. gemellus*) and community levels (avoidance by potential competitors).

380

381 **Effects of *P. grandis* on prey abundance and drift**

382 The finding that *Baetis* spp. and chironomid larvae (Orthocladiinae) were the most
383 abundant prey items in the guts of *P. grandis* was similar to findings in other studies on a
384 range of predatory stoneflies from Europe and North America (Allan 1982, Peckarsky 1985,
385 Elliott 2003, Céréghino 2006, Bo et al. 2008). In our study, *P. grandis* displayed a preference
386 for *Baetis* spp. in the experimental channels but not in the Tourettes stream and for orthoclads
387 in the Tourettes stream but not in the experimental channels. The differences in preference
388 between the Tourettes stream and experimental channels might be a consequence of the
389 different proportions of *Baetis* species in the 2 environments. *Baetis alpinus* made up >90%
390 of the Baetidae in the Tourettes stream, whereas *B. gemellus* made up >70% in the
391 experimental channel. These 2 species appear to display different antipredator behaviors.

392 *Baetis alpinus* was a more mobile, stronger swimmer than *B. gemellus* (KK, personal
393 observation) and tended to drift more frequently, a common response to plecopteran
394 predation pressure (Kratz 1996). *Calotriton asper*, the top predator in many alpine springs,
395 typically selects for more mobile prey (Montori 1992). Hence, the more sedentary behavior
396 of *B. gemellus* when compared to *B. alpinus* may reflect adaption to *C. asper* predation
397 pressure, but a degree of prey naivety in the presence of *P. grandis* (Cox and Lima 2006). On
398 the other hand, the difference may have been an artefact of the small size of the experimental
399 channels, which would have reduced in-channel predation refugia when compared to the
400 Tourettes stream (Bechara et al. 1993).

401 Intraguild predation occurred in the benthic community of the Tourettes stream.
402 Large *Rhyacophila* spp. were found in the guts of *P. grandis*. However, further work is
403 needed to ascertain the symmetry of this relationship, i.e., to learn whether *P. grandis* is
404 always the predator of *Rhyacophila* or whether the roles change depending on body size or
405 life-cycle stage (Polis et al. 1989). Both these taxa share chironomid prey (Lavandier and
406 Céréghino 1995, KK, unpublished data), thus some degree of interference competition can be
407 inferred. Bo et al. (2008) also found *Rhyacophila* and other caseless caddisfly larvae in the
408 guts of stoneflies, results suggesting that intraguild predation may be a common feature in
409 low-order streams. Hence, the range expansion of *P. grandis* is likely to intensify biotic
410 interactions by increasing the number of foodweb links with only a slight increase in the
411 number of species (Woodward and Hildrew 2001). This shift will in turn reduce niche space,
412 particularly among predators which, because of dietary overlap with *P. grandis*, will increase
413 competition for prey items (Wissinger and Mcgrady 1993, Woodward and Hildrew 2002a).

414 Despite a large effect size, the reduction in total invertebrate abundance was not
415 significant, probably because of high prey turnover rates, i.e., immigration replaced
416 consumed or emigrated individuals (Lancaster 1990), or the small number of replicates (low

417 statistical power). However, densities of 2 *Baetis* spp., which are typically important
418 components of carnivorous stonefly diets (Peckarsky 1985, Céréghino 2006), were
419 significantly reduced in our study. This finding supported H₁ (*P. grandis* would reduce prey
420 abundance) and suggests that predatory stoneflies are more efficient at catching *Baetis*
421 nymphs than other, more sedentary organisms, such as chironomids (Peckarsky and Cowan
422 1995, Elliott 2003). This greater success is primarily a result of the conspicuous swimming
423 behavior of *Baetis*, which acts as a predator stimulus (Peckarsky and Penton 1989).

424 In contrast to our results, stonefly predation reduced chironomid densities in other
425 laboratory and field experiments (Peckarsky 1985, Lancaster 1990, Elliott 2003). These
426 investigators used ‘predation arenas’ (Allan et al. 1987, Elliott 2003), with the abundance and
427 diversity of prey and predatory taxa strictly controlled, or field-based mesocosms where
428 colonization by other, nontarget, large-bodied invertebrates was limited (e.g., Woodward and
429 Hildrew 2002b). In our experiment, predators other than *P. grandis* were free to colonize all
430 channels. The most abundant other predators were *Rhyacophila* spp. (mainly *R. intermedia*)
431 and *Tanypodinae*, both of which feed predominantly on chironomids (Lavandier and
432 Céréghino 1995, Woodward and Hildrew 2002b). These predators were relatively abundant
433 in the control channels, so predation of orthoclad larvae probably occurred in treatment and
434 control channels, but by different taxa, i.e., *P. grandis* consumed both *Baetis* and
435 chironomids in the treatment channels, whereas *R. intermedia* and tanypods consumed
436 chironomids in the control channel.

437 The increased drift rate by *B. gemellus* and *B. muticus* observed in the *P. grandis*
438 treatment is a common behavioral response among *Baetidae* (Wooster and Sih 1995,
439 Peckarsky et al. 2008). This response enables increased resource acquisition when mortality
440 (predation) risk is high (Peckarsky 1996). However, *B. alpinus* exhibited a per capita drift
441 rate far greater than that of the other *Baetis* spp., most likely because its co-evolution with *P.*

442 grandis amplified this behavioral trait (McPeck 1990). These findings further support H₁ (P.
443 grandis would reduce prey abundance) because the key prey found in the guts (Baetis spp.)
444 also were more abundant in the drift from P. grandis channels. Baetis alpinus drift was
445 significantly greater from the control than from treatment channels, possibly because of
446 apparent and exploitative competitive interactions among the Baetis spp. (Holt and Lawton
447 1994), with B. gemellus and B. muticus primarily predator (P. grandis) limited and B. alpinus
448 primarily resource limited (Chase et al. 2002). Prey with superior resource acquisition
449 capabilities, in this case, B. gemellus and B. muticus, also are the most vulnerable to predation
450 (Holt and Lawton 1994).

451

452 **Macroinvertebrate feeding guild structure**

453 Distinct changes in functional feeding guild structure were observed in our study.
454 These changes appeared to support H₂ (P. grandis would alter community feeding guild
455 structure through selective predation and competition). The density of grazing taxa was lower
456 in treatment than in control channels, probably because Baetis spp., the most abundant
457 grazers recorded during the experiment, were more prone than other taxa to stonefly
458 predation (Peckarsky and Penton 1989, Elliott 2003). The significantly lower relative
459 abundance of large-bodied predators in the P. grandis channels probably was the result of
460 intraguild predation/interference competition (Polis et al. 1989). However, all P. grandis
461 nymphs used in our experiment were fully developed and the experiment ran for a relatively
462 short time interval, so ontogenetic shifts in feeding habits of P. grandis and other predatory
463 taxa were not incorporated into the study design (Lavandier and Céréghino 1995, Woodward
464 and Hildrew 2001, Céréghino 2006). Nevertheless, these findings suggest the potential for
465 future predator replacement as the physicochemical habitat template of low-order alpine
466 streams changes under a warmer climate (Khamis et al. 2013).

467

468 **Body-size spectrum**

469 The smaller individuals of *B. gemellus* in the treatment channels (highlighted by the
470 skewed violin plots) may a consequence of size-selective predation by *P. grandis*, with a
471 preference for larger individuals as prey. Few investigators have studied size-selective
472 predation in Plecoptera, but both Allan et al. (1987) and Peckarsky (1985) found
473 experimentally that developed stonefly nymphs (>25 mg) preferred medium-sized prey (>0.2
474 mg). A 2nd explanation for the *B. gemellus* body-size distribution would agree with the
475 findings of Lancaster (1990), who found that larger *Baetis* nymphs drifted more frequently
476 than small nymphs from channels in the presence of a predatory stonefly.

477 When body size was considered at the community level (individual based rather than
478 taxon averaged) individuals in the larger body-size classes were less abundant in the
479 treatment than in the control channels, supporting H₃. This difference probably was a result of
480 intraguild predation/interference competition, as evidenced by the presence of *Rhyacophila*
481 spp. in the diet of *P. grandis* and associated positive LOR scores. This result suggests
482 significant niche overlap between *Rhyacophila* spp. and *P. grandis* resulting from shared
483 common prey (Lavandier and Céréghino 1995). Hence, the pattern of reduced body size in
484 the treatment channels was caused by consumption of other large-bodied invertebrate
485 predators by *P. grandis* or by avoidance of *P. grandis* by other large-bodied predators
486 (Woodward and Hildrew 2002a, Vanak and Gompper 2010). This result has important
487 implications for foodweb stability and structure because changes in the body-size spectrum
488 (see Fig. 7), particularly an increase in size of the apex predator, can reduce refugia
489 associated with gape-limited predation (Woodward et al. 2005). However, predicting how
490 such changes will propagate through the food web in alpine systems is difficult because
491 changes in interaction strength and omnivory can destabilize and stabilize food webs,

492 respectively (Borrvall et al. 2000).

493

494 **Cascading effects of *P. grandis***

495 Top-down, predation-driven, trophic cascades are more prevalent and often more
496 pronounced in aquatic systems than terrestrial settings, primarily because of larger consumer
497 to producer biomass ratios (Shurin et al. 2002). However, our study did not reveal any
498 significant change in the magnitude of the trophic cascade caused by the presence of *P.*
499 *grandis* (H₄ not supported). This lack of support may have been a result of a number of
500 factors. First, the reduction in grazer densities observed in the treatment channels was
501 relatively small (Table 4). Second, the high feeding plasticity of macroinvertebrates in alpine
502 streams (Zah et al. 2001, Füreder et al. 2003, Clitherow et al. 2013) may increase ecological
503 redundancy among feeding guilds (i.e., detritivores/predators also consume algae), thus
504 dampening the trophic cascade (Polis et al. 2000). Third, the presence of other large-bodied
505 predatory taxa (e.g., *Rhyacophila* spp.) probably would have affected grazing taxa in the
506 control channels, potentially reducing grazing efficiency (Wooster and Sih 1995).

507

508 **Conclusions and implications**

509 As physicochemical habitat characteristics of alpine streams become more benign
510 because of climate warming/river water-source changes, upstream migration of *P. grandis* is
511 likely to increase the trophic height of invaded communities as current invertebrate predators
512 (e.g., *Rhyacophila* spp.) become prey. Our study highlighted interference competition and
513 interguild predation as important structuring mechanisms with the potential to alter the body-
514 size spectrum and foodweb interactions in *P. grandis*-invaded systems (Woodward and
515 Hildrew 2002b). Therefore, the predicted range expansion is likely to intensify biotic
516 interactions, reduce niche breadth, and increase competition among predators). Furthermore,

517 certain grazing taxa, primarily *B. gemellus* and *B. muticus*, are likely to be selectively
518 predated, which could alter community structure and, potentially, its ecological functioning.

519 Our study has further emphasised the need to consider biotic interactions in species-
520 abundance models (Araújo and Luoto 2007) because both community structure and body-size
521 structure were altered under experimental predator range expansion. These findings have
522 important implications for ecosystem stability because predator and prey body-size ratios
523 control trophic interaction strengths (Emmerson and Raffaelli 2004, Woodward et al. 2005).
524 Thus, ignoring or treating biotic interactions as constant is likely to cause erroneous
525 predictions regarding future distributions and extinction vulnerability. This increased
526 extinction vulnerability is particularly poignant in alpine river networks because, despite
527 having low α diversity, 1st-order streams are important sites for regional biodiversity (Finn et
528 al. 2011). Upstream movement by predatory taxa will create synergistic feedbacks among
529 biotic interactions and climate-driven physicochemical habitat change, which are likely to
530 increase the risk of species extinctions to the detriment of both β and γ diversity (Brown et al.
531 2007a). Therefore, work is urgently needed to ensure that additional anthropogenic pressures
532 (e.g., water abstraction, hydropower schemes, nutrient enrichment, and cattle trampling) are
533 limited to prevent more pressure on this unique and fragile habitat (Hannah et al. 2007,
534 Khamis et al. 2013).

535

536 **ACKNOWLEDGMENTS**

537 This research was funded by an EU-FP7 grant awarded to the authors through the
538 Assessing Climate Impacts on the Quantity and quality of Water (ACQWA) Project
539 (<http://www.acqwa.ch>). We are grateful to the Parc National des Pyrénées for permission to
540 work in the study area. We thank Richard Johnson for logistical support during the field
541 season and for his assistance in the design and construction of the mesocosm channels, Mel
542 Bickerton for laboratory support, and Lawrence Bird, Faye Jackson, and Robert Senior for
543 field assistance. We also appreciate the time and thoughts of Associate Editor B. L. Kerans
544 and 2 anonymous referees, whose constructive comments greatly improved the manuscript.
545

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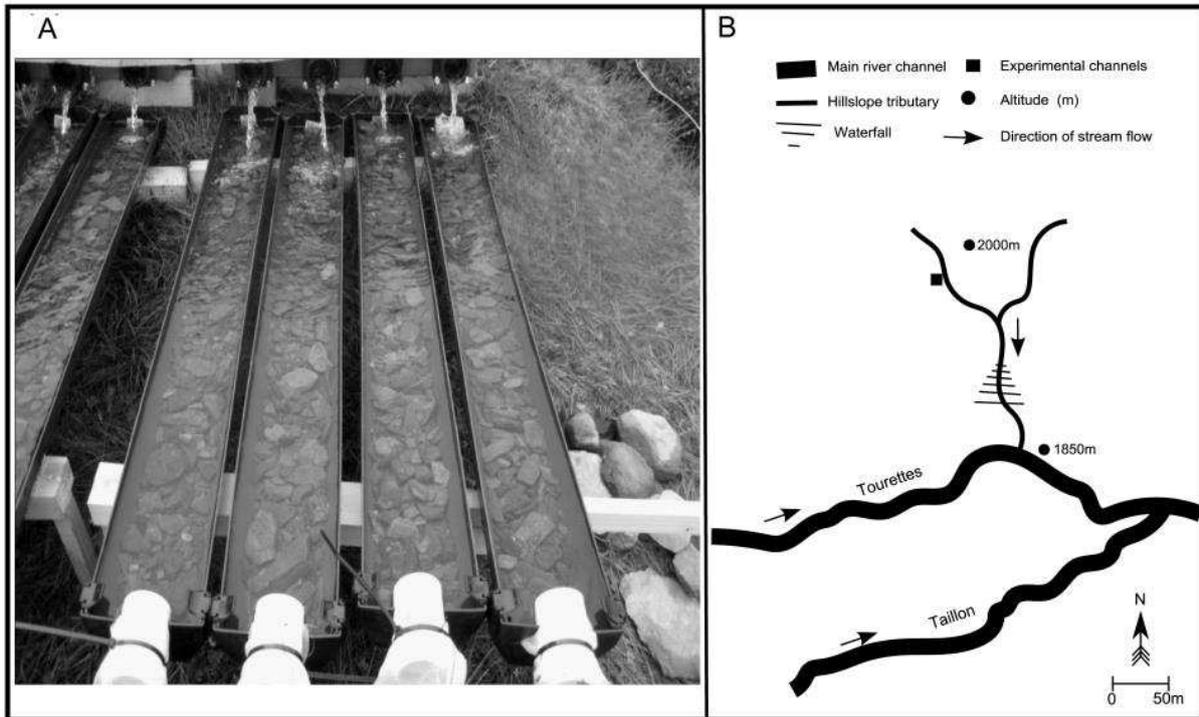
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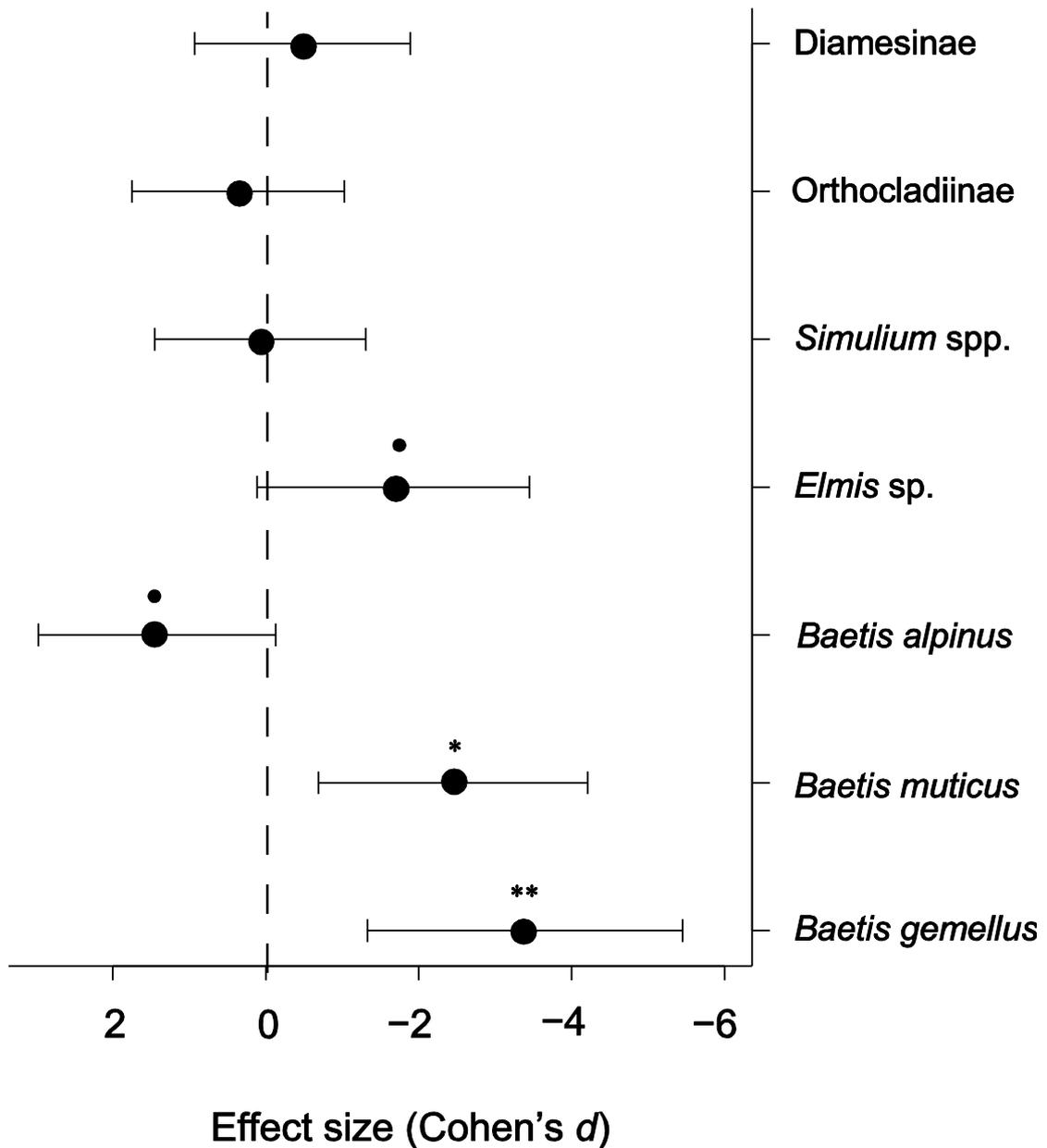
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764 Fig. 1. Experimental channels (A) and a map of the study area showing the field site and

765 location of experimental channels (B).

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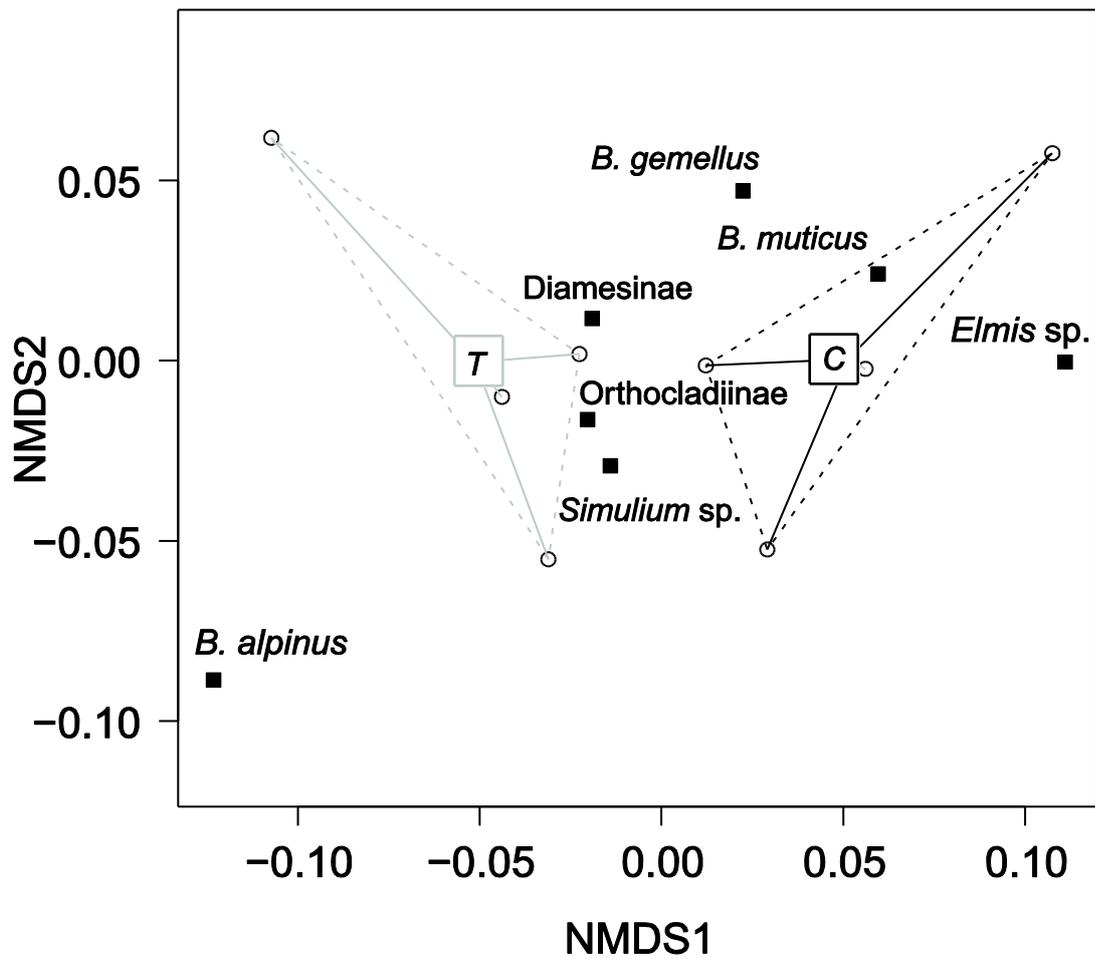
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769 Fig. 2. Standardized effect size (Cohen's *d*) for the difference between treatment and control770 channels for prey density (individuals/m²). Whiskers represent the 95% confidence771 intervals, and significant 1-way analysis of variance tests are indicated by ** = $p <$ 772 0.01, * = $p < 0.05$, • = $p < 0.1$.

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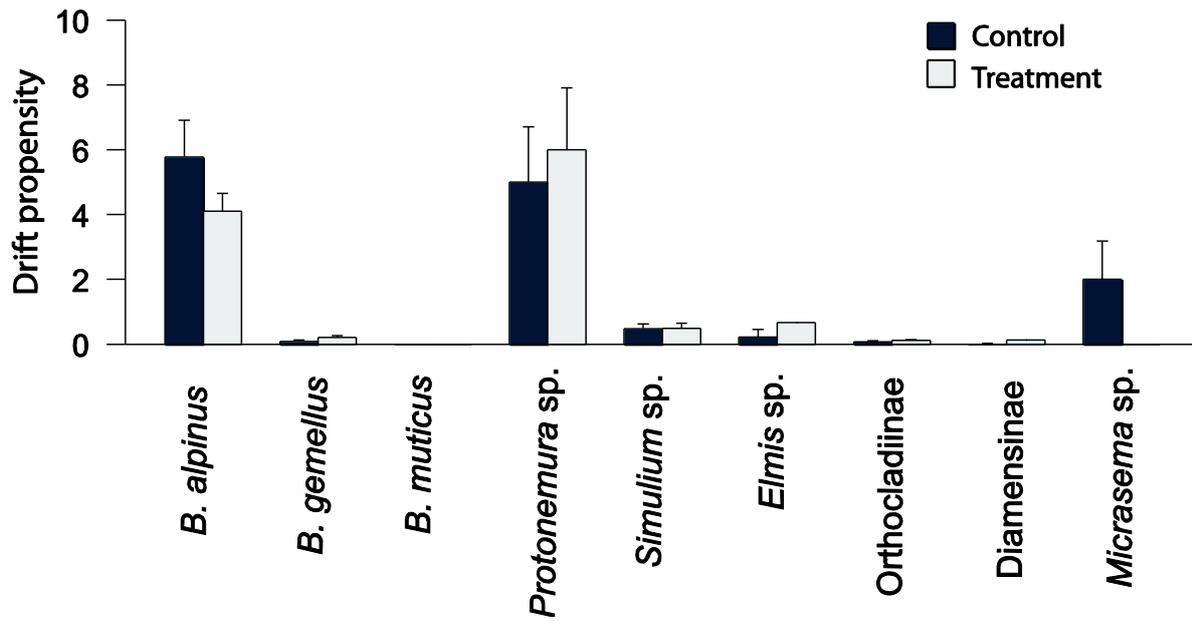


774

775 Fig. 3. Nonmetric dimensional scaling (NMDS) ordination of abundant taxa (B. = Baetis) in
 776 experimental channels. Numbers denote channels (treatment: 1–4, control: 5–8).

777 Dashed line shows the convex hull for treatment (black) and control channels (gray).

778



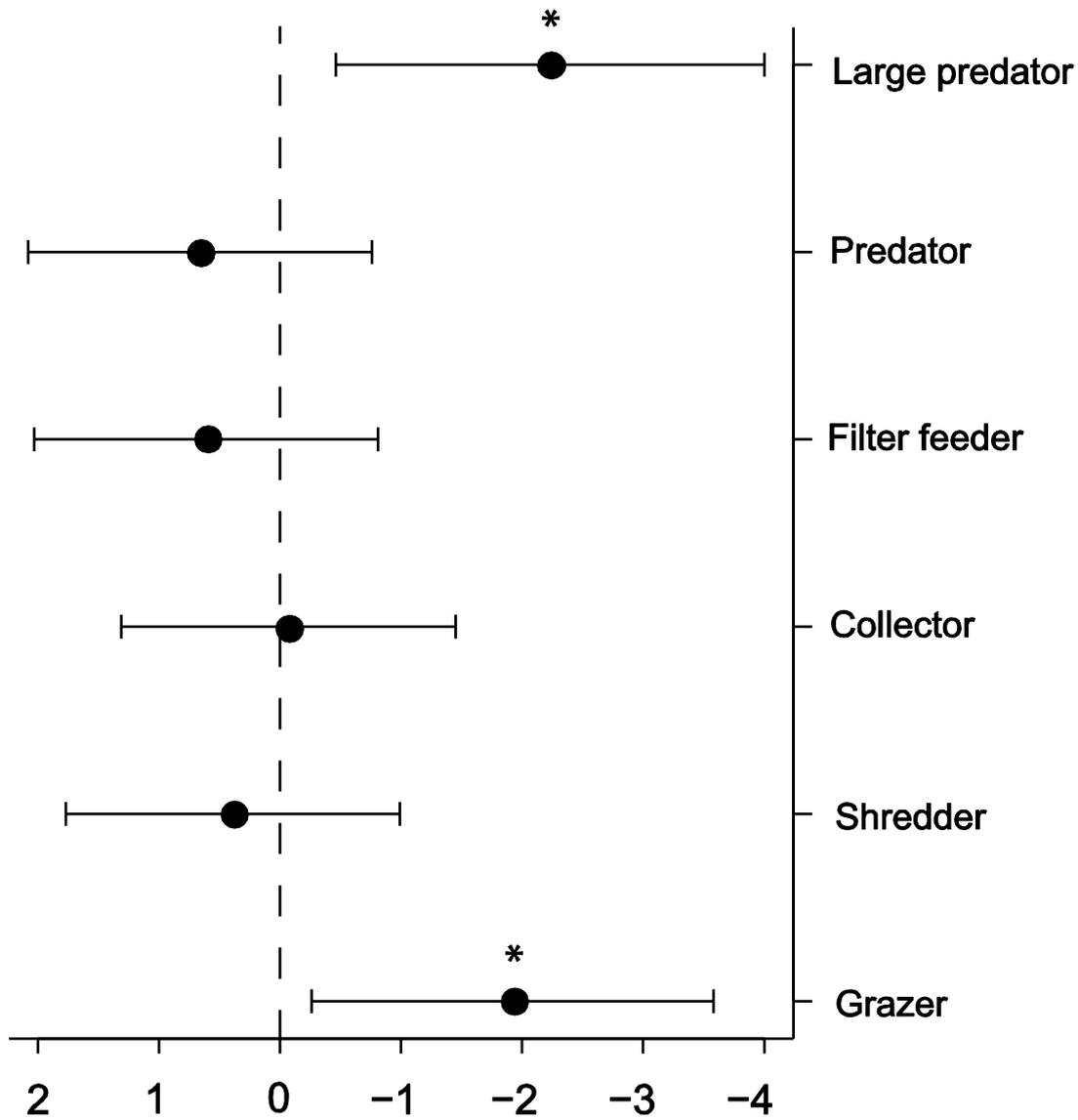
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781 Fig. 4. Drift propensity (emigration/benthic density) of the most abundant taxa recorded from

782 the mesocosm channels, 26–27 July. B. = Baetis.

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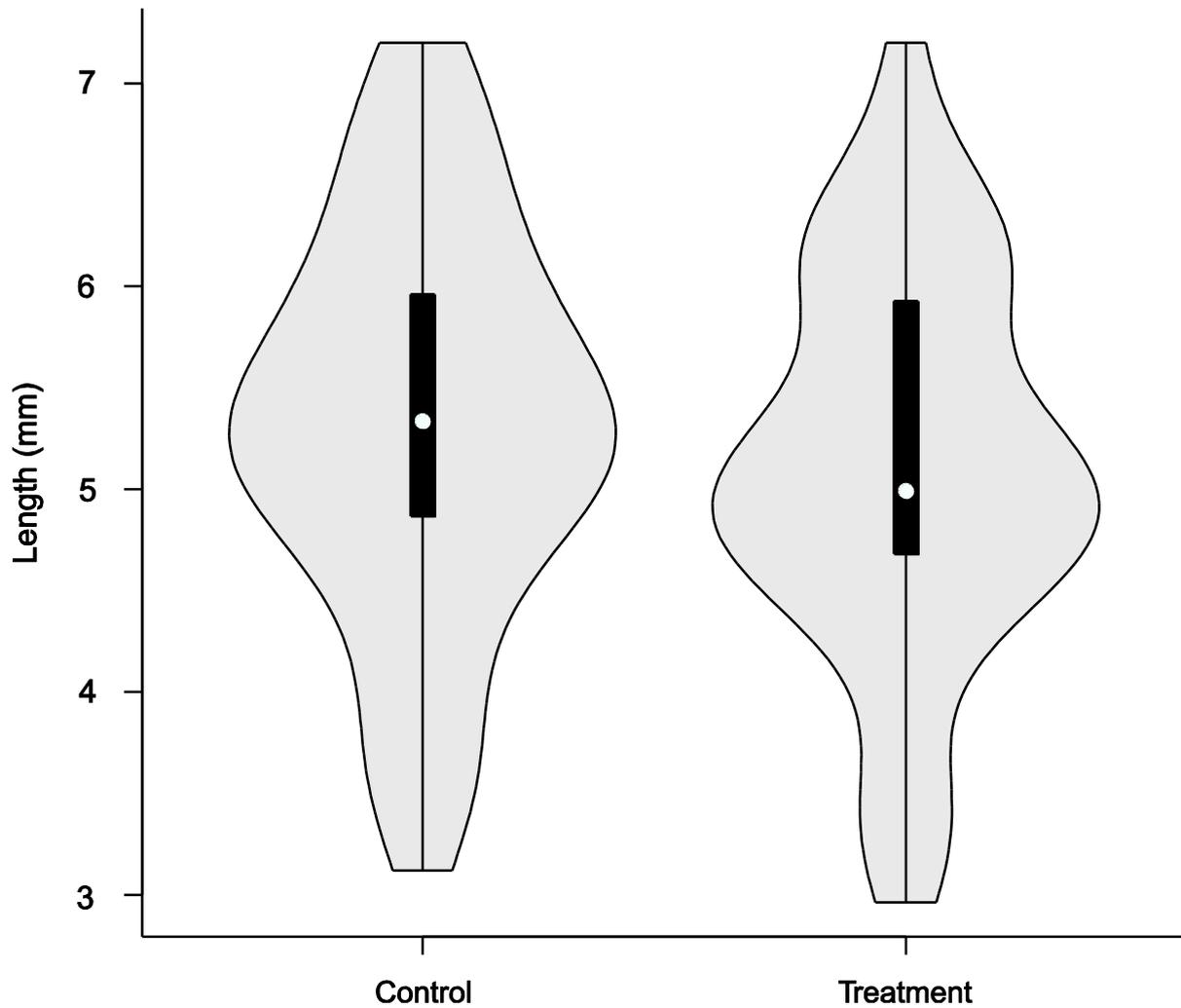


Effect size (Cohen's *d*)

784

785 Fig. 5. Standardized effect size (Cohen's *d*) for the difference between treatment and control
 786 channels for functional feeding group relative abundance. Whiskers represent the 95%
 787 confidence intervals and significant 1-way analysis of variance tests are highlighted
 788 by * = $p < 0.05$, • = $p < 0.1$.

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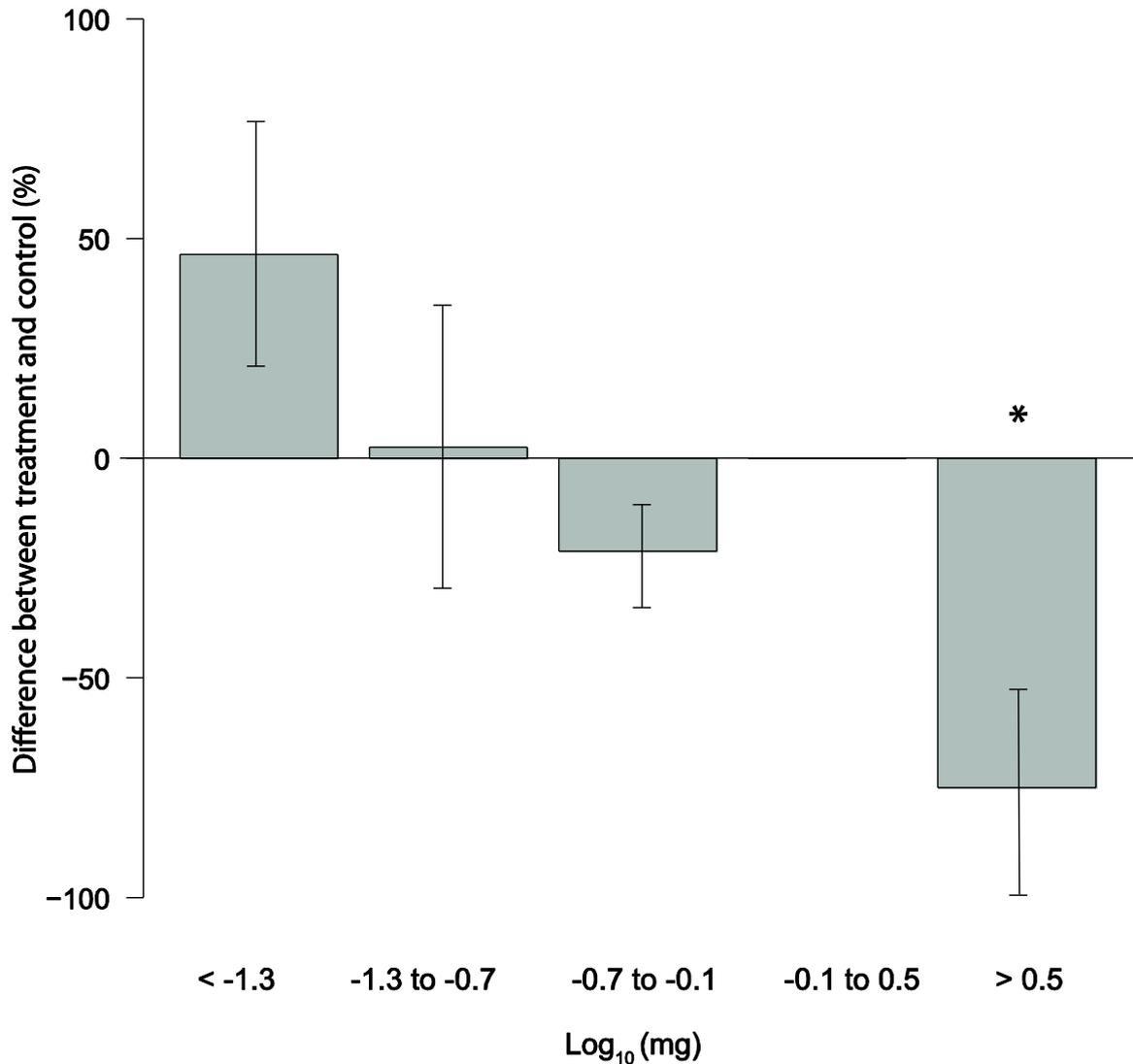
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Control

Treatment

791 Fig. 6. Violin plots for the length (mm) of *Baetis gemellus* recorded in the mesocosm
792 channels. The light grey area represents a kernel density function. The black box and
793 line represent a traditional box plot where, box ends are quartiles, whiskers are 1.5
794 multiplied by the inter-quartile range (1st and 3rd quartiles), and the white dot
795 represents the median.

796



797

798 Fig. 7. Mean (± 1 SE) % difference between treatment (not including *Perla grandis*) and
 799 control for the number of individuals in each of the 5 \log_{10} (body mass) size classes.
 800 Mean and SE were calculated from all possible pairwise comparisons of individual
 801 treatment and control channels (for each size class, $n = 16$). * denotes significant
 802 Kruskal–Wallis test ($H = 3.94$, $p < 0.05$).

803 Table 1. Diet of *Perla grandis* nymphs in the Tourettes stream ($n = 25$) and mesocosm
 804 channels ($n = 8$).

Location	Prey taxon	% in gut	% in substrate	LOR	Mean (SE) prey items/gut
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Tourettes Stream	Baetis spp.	37.6	57.1	-0.04	3.4 (0.2)
	Orthocladiinae	34.1	6.2	1.35	
	Protonemura spp.	11.8	2.1	1.62	
	Rhithrogena sp.	4.7	21.2	-1.32	
	Simulium spp.	3.5	4.9	-0.31	
	Unidentified arthropod	3.5	NA	NA	
	Limnephilidae	2.4	0.74	1.13	
	Chloroperla sp.	1.2	1.6	-0.32	
	Rhyacophila sp.	1.2	0.27	1.37	
Experimental channels	Baetis spp.	48.3	37.4	0.26	3.6 (0.3)
	Orthocladiinae	27.6	28.4	-0.01	
	Unidentified Plecoptera	10.3	NA	NA	
	Unidentified arthropod	6.9	NA	NA	
	Protonemura spp.	3.4	0.2	2.85	
	Simulium spp.	3.4	9.1	-0.96	

806 Table 2. Mean (SE) abundance (individuals/m²) of the abundant prey taxa in control and
 807 treatment channels. Results of the one-way analysis of variance examining the effect of *Perla*
 808 *grandis* (treatment) on prey abundance are also displayed

Taxa	Abundance		ANOVA		
	Control	Treatment	Source	F _{1,5}	p
Baetis. gemellus	366 (23)	233 (19)	Treatment	18.89	0.007
			Block	0.01	0.94
Baetis muticus	106 (9)	50 (12)	Treatment	10.77	0.02
			Block	0.362	0.57
Baetis alpinus	19 (2)	36 (8)	Treatment	4.57	0.08
			Block	2.28	0.19
Elmis sp.	17 (4)	6 (2)	Treatment	6.84	0.04
			Block	1.39	0.29
Simulium spp.	64 (12)	36 (35)	Treatment	0.01	0.91
			Block	1.57	0.27
Orthoclaadiinae	283 (70)	235 (24)	Treatment	0.41	0.55
			Block	1.61	0.24
Diamesinae	21 (6)	15 (3)	Treatment	0.98	0.37
			Block	7.52	0.04

809

810 Table 3. One-way analysis of variance for the effects of predator treatment on drift propensity
 811 (per capita 24-h drift rate) for date 23–24 July. The standardized effect size (Cohen’s d) is
 812 presented with associated 95% confidence intervals. No block effect was identified.

Taxon	F _{1,5}	p	d (95 % CI)
Baetis muticus	2.82	0.23	0.94 (0.42–1.48)
Baetis gemellus	36.54	0.05	3.41 (2.53–4.31)
Protonemura spp.	0.02	0.90	0.07 (–1.31–1.46)
Baetis alpinus	28.07	0.05	–2.98 (–3.81 to –2.18)

813

814 Table 4. Mean (SE) abundance (individuals/m²) and relative abundance (%) of functional
 815 feeding groups in control and treatment channels and results of 1-way analysis of variance
 816 results. Significant block effects are indicated as * = p < 0.05.

Functional feeding group	Relative abundance		Abundance		Relative abundance		Abundance	
	Control	Treatment	Control	Treatment	F _{1,5}	P	F _{1,5}	P
Grazer	77.0 (7.1)	69.3 (5.9)	804 (40)	564 (40)	7.29	0.04	7.77	0.03
Shredder	2.1 (0.7)	2.6 (1.0)	21 (1)	21 (1)	0.30	0.60	0.00	0.97
Collector	3.6 (1.0)	3.5 (0.8)	37 (3)	27 (1)	0.01	0.93	0.83	0.40
Filter feeder	5.6 (1.5)	9.2 (3.6)	60 (5)	75 (6.1)	0.74	0.42	0.17	0.69
Predator	11.7 (2.5)	15.4 (3.3)	121 (7)	125 (9)	0.88	0.38*	0.31	0.60*
Large predator	0.02 (0.05)	0.01 (0.04)	25 (4)	6 (1)	10.0	0.01	4.66	0.07

817

818

819 Table 5. Mean (SE) length of the 4 most abundant taxa recorded in the end-point community.

820 Results from Student's t-test are complimented by standardized effect sizes (unbiased

821 estimate of Cohen's d) and 95% confidence interval (CI).

Taxon	Body length (mm)		n	t	p	d (95% CI)
	Treatment	Control				
Baetis gemellus	5.06 (0.09)	5.31 (0.09)	120	2.29	0.02	-0.42 (-0.78 to -0.05)
Baetis muticus	5.99 (0.06)	6.14 (0.06)	80	1.79	0.09	-0.36 (-0.84-0.04)
Orthocladiinae	4.06 (0.16)	4.10 (0.15)	50	0.03	0.85	0.01 (-0.13-0.14)
Simulium spp.	4.25 (0.13)	4.13 (0.15)	50	0.33	0.57	0.07 (-0.07-0.21)

822