

# Separating the genetic and environmental drivers of body temperature during the development of endothermy in an altricial bird

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## Abstract

When altricial birds hatch, they are unable to regulate their own temperature, but by the time they fledge they are thermally independent. Early-life conditions have been shown to be an important factor contributing to fitness. However, it is currently unknown to what extent body temperature during endothermy development is driven by genetic variation or by the early environment. We use thermal images of cross-fostered house sparrows (*Passer domesticus*) throughout the nestling period to separate genetic and environmental drivers of body temperature. We estimated negligible heritability of body temperature at all ages. We further found that there are effects from the natal environment that carry over into the late nestling stage. A correlation between the early- and mid-nestling periods was explained by the natal environment, and during this period body temperature and growth followed independent developmental trajectories. Furthermore, higher body temperature was under viability selection, independent of body mass. We, therefore, demonstrate that the natal environment influences future offspring phenotype via a novel measure; body temperature. Our study provides a novel investigation into the environmental and genetic drivers of body temperature variation in a wild bird, furthering our understanding of how traits evolve.

**Keywords:** endothermy, infrared thermography, nestling development, quantitative genetics

## Introduction

Passerine birds are heterothermic endotherms, whose body temperature is influenced by both intrinsic and extrinsic factors, such as body condition (Nord et al., 2013), time of day (Barrett & Takahashi, 1995; Binkley et al., 1971) and exposure to stressors (Jerem et al., 2019). As adults, optimisation of body temperature has fundamental benefits for the individual, such as energy conservation (Tattersall et al., 2016) and improved ability to remain active (Torre-Bueno, 1976). Altricial birds start to develop their own body temperature regulation as nestlings and complete this development before fledging. Nestlings are therefore sensitive to fluctuations in their thermal condition (reviewed in Nord & Giroud, 2020) and to ambient temperature, which have both been shown to impact their survival (Andreasson et al., 2018; Bourne et al., 2020; Marques-Santos & Dingemanse, 2020). Previous studies on ambient temperature studied its effects at the brood level. We therefore know little about how an individual's body temperature is determined, and this is yet to be considered in an evolutionary context.

Early-life conditions affect the phenotype of developing individuals with effects persisting into adulthood (Metcalfe & Monaghan, 2001; Saino et al., 2018; Spagopoulou et al., 2020; Zwaan et al., 2020). To date, the literature on body tem-

perature in birds has largely focussed on the adaptive response to environmental conditions, mostly in experimental studies. Very few studies have been conducted in the wild and most of the literature on body temperature in birds comes from studies on poultry that have been selected for production. The rate of endothermy development in wild nestling blue tits (*Cyanistes caeruleus*) has been shown to vary between individuals and with brood size (Andreasson et al., 2016), and nestlings in experimentally heated nests were better able to thermoregulate at later nestling ages (Page et al., 2022).

Early environments, such as the nest condition, may affect the thermal characteristics of juvenile birds, with future fitness implications. Prehatching (e.g., egg yolk composition) and posthatching (e.g., parental incubation behaviour) conditions have been shown to affect nestling phenotypes, such as growth (Mousseau, 1998; Nord & Nilsson, 2011; Saino et al., 2003). However, we do not know what determines an individual's development of body temperature regulation, and if the drivers of body temperature regulation are the same or separate from the drivers of growth, especially in the wild.

Metabolic rate is a trait that is studied more commonly than body temperature, but both probably share physiological mechanisms. Metabolic rate is a key trait for determining an individual's energy expenditure (Biro & Stamps, 2010).

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time of measurement) were corrected for during image extraction in either Thermimage or FLIR Tools.

## Data analysis

LAW performed all statistical analyses for this study with guidance from JLP, who also reviewed the analysis code. LAW determined the heritability of body temperature for each age (2, 5, 10, and 12 days) in separate “animal models”, which use the relatedness structure of a pedigree to estimate additive genetic effects (Henderson, 1988; Kruuk, 2004). Our pruned pedigree contained 1,242 individuals for the age 2 dataset with 1,143 maternities and 1,159 paternities (see [Supplementary Information S1](#) for further pedigree statistics and for the other ages). We included fixed effects of air temperature (at time of image capture; continuous), year (two-level factor), and brood size (at given age; continuous) in the models. In addition to additive genetic effects, we estimated random effects of the natal brood (brood in which the nestling was born into), the rearing brood (brood in which the nestling was raised) and date. As the additive genetic effect variance at all ages was low,  $p$ -values were calculated to determine if there was statistical support for this effect. Random effects of rearing brood were not included in the model for day 2, as temperature measures were taken at the point of cross-fostering. All covariates were mean-centred to aid interpretation. We also calculated evolvability (Houle, 1992) of body temperature at each age; to do this we used the same models described above but converted body temperature from ( $^{\circ}\text{C}$ ) to Kelvin (traits need a true 0 to calculate evolvability).

To account for selective disappearance when estimating the variance components of body temperature (i.e., there is a potential to underestimate variances at later ages when viability selection is acting on the traits), LAW then modelled body temperature (for all ages combined) using a multivariate linear mixed effects model (Hadfield, 2008; Hadfield et al., 2013), in a character state approach. This approach uses data augmentation to estimate data for missing individuals at older ages through their values at younger ages. It also means that we did not have to make any assumption about the function by which temperature changes over age. We included a fixed effect of age (categorical), as well as brood size (at given age; continuous), air temperature (continuous), and year (categorical), which were treated separately for each age inside the model. Date of image capture was modelled as random effects in the model. We modelled natal brood and residual effects with  $4 \times 4$  unstructured covariance matrixes, specifying a variance for each age, and the covariances between ages. We modelled rearing brood effects for ages 5, 10, and 12 with a  $3 \times 3$  unstructured covariance matrix. We did not have sufficient power to model genetic effects in this analysis, most likely due to the very low genetic variance across ages. When natal brood is separated from the rearing brood in our models, the genetic variance will be confounded with the natal brood effects; however, given the small additive genetic effects in the models for each age separately, the additive genetic effect will be negligible. We repeated this analysis but included a fixed effect of body mass (again separately for each age) to determine to what extent body temperature is an independent trait from body mass.

The analyses described above were run in MCMCglmm version 2.36 (Hadfield, 2010) using R version 4.4.0 (R Development Core Team, 2009). The burn-in period was

45,000 for all analyses and a chain length of 195,000 iterations and a thinning interval of 150. Parameter expanded priors were used for the random effect variances ( $V = 1$ ,  $\nu = 1$ ,  $\alpha.\mu = 0$ , and  $\alpha.V = 1,000$ ), and inverse wishart priors were used for the residual variances ( $V = 1$  and  $\nu = 0.002$  for the univariate models, and  $V = 1e-6$  and  $\nu = 5$  for the multivariate models).  $p$ -Values were deemed statistically significant if the 95% credible intervals do not overlap zero. To infer statistical support for the genetic component in the day 2 univariate model, JLP generated a null distribution of posterior medians for the univariate models at each age via simulation following Pick et al. (2023).  $p$ -Values were generated as the proportion of samples from the null distribution that were greater than the posterior median estimated from the real data. The minimum detectable heritabilities (posterior medians) were 0.072, 0.081, 0.151, and 0.107 for days 2, 5, 10, and 12, respectively. When model estimates are presented, we use the posterior median (unless otherwise stated).

LAW analysed whether body temperature predicts survival to the next age using generalised linear mixed-effect models with a binomial error distribution in lme4 (Bates et al., 2015), using the BOBYQAO optimiser. For age 12, survival was determined by whether an individual was sighted as an adult (i.e., over winter or during the following breeding season). All ages were analysed in separate models, and included fixed effects of body temperature, body mass, and year, and random effects of natal brood (all ages) and rearing brood (age 5, 10, and 12 only).  $p$ -Values were estimated from likelihood ratio tests, comparing nested models.

## Results

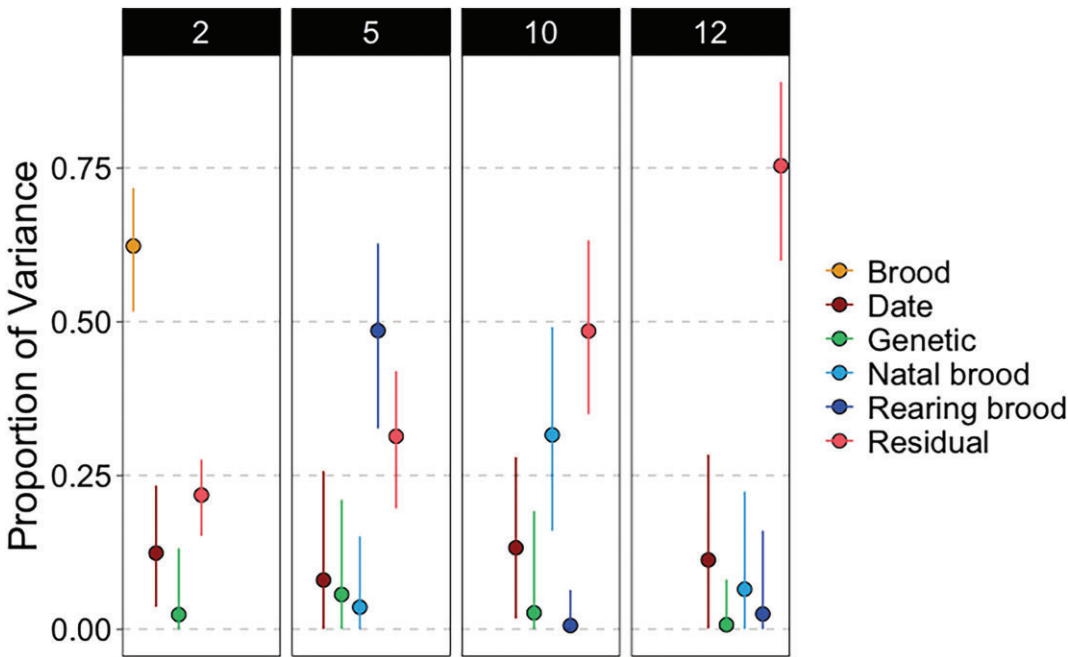
### Variance explaining body temperature at each age

To reliably estimate the environmental and genetic variance components explaining body temperature at each age, we accounted for air temperature, which as expected was significantly positively correlated with surface body temperature at all ages (Table 1). As our study took place over two consecutive years, we also accounted for differences in the environmental conditions between years, by including year as a two-level factor in the model. In 2019, birds were warmer at ages 2 and 5 compared to those recorded in 2018 (although this was not statistically supported). However, birds were cooler at ages 10 (not significant) and 12 compared to 2018 (Table 1).

We found that body temperature had a small (and not statistically supported) heritability at all ages [Age 2,  $h^2 = 0.02$  (95% CrI: 0.00–0.13,  $p = .333$ ); Age 5,  $h^2 = 0.06$  (95% CrI: 0.00–0.21,  $p = .105$ ); Age 10,  $h^2 = 0.03$  (95% CrI: 0.00–0.19,  $p = .562$ ); Age 12,  $h^2 = 0.01$  (95% CrI: 0.00–0.08,  $p = .990$ ); Figure 1, Table 1]. We calculated evolvability ( $I_A$ ) of body temperature (in Kelvin) to be 0.000003 (95% CrI: 0.00000001–0.00001) for age 2, 0.000004 (95% CrI: 0.00000001–0.00001) for age 5, 0.000003 (95% CrI: 0.00000002–0.00001) for age 10, and 0.0000004 (95% CrI: 0.000000007–0.000002) for age 12. At early ages (ages 2 and 5), body temperature was driven by the current conditions that the individual experienced in the nest (i.e., the combined natal and rearing brood effect for age 2 and rearing brood for age 5). At age 10, when the nestlings become thermally independent, temperature was not driven by their

**Table 1.** Variance components of body temperature by age. Values for fixed effects and random effect variances are the posterior median (95% credible intervals). Statistically significant values ( $p_{\text{MCMC}} < 0.05$ ) are in bold.

	Age (days)			
	2	5	10	12
Fixed				
Intercept	30.79 (30.12–31.35, $p < .001$ )	34.24 (33.75–34.72, $p < .001$ )	37.65 (37.11–38.17, $p < .001$ )	38.56 (38.28–38.82, $p < .001$ )
Air temperature	0.55 (0.45–0.64, $p < .001$ )	0.44 (0.37–0.52, $p < .001$ )	0.22 (0.14–0.29, $p < .001$ )	0.19 (0.14–0.25, $p < .001$ )
Year (2019)	0.64 (–0.12–1.47, $p = .134$ )	0.51 (–0.04–1.17, $p = .10$ )	–0.47 (–1.11–0.12, $p = .142$ )	–0.54 (–0.92– –0.14, $p = .006$ )
Brood size	–0.04 (–0.39–0.30, $p = .818$ )	0.61 (0.33–0.91, $p < .001$ )	0.33 (0.09–0.58, $p = .006$ )	–0.06 (–0.23–0.11, $p = .538$ )
Random				
Genetic	0.20 (0.00–1.12, $p = .333$ )	0.27 (0.00–1.08, $p = .105$ )	0.15 (0.00–1.12, $p = .562$ )	0.02 (0.00–0.20, $p = .990$ )
Natal brood	5.25 (4.18–6.59)	0.18 (0.00–0.76)	1.76 (0.85–3.09)	0.16 (0.00–0.55)
Rearing brood		2.37 (1.58–3.34)	0.03 (0.00–0.35)	0.06 (0.00–0.38)
Date	1.04 (0.29–2.15)	0.38 (0.00–1.43)	0.74 (0.10–1.70)	0.26 (0.00–0.75)
Residual	1.85 (1.38–2.19)	1.55 (1.03–1.92)	2.72 (2.12–3.34)	1.80 (1.50–2.15)



**Figure 1.** The proportion of variance in body temperature at each age (2, 5, 10, and 12 days posthatching) over the nestling period for the random effects in the model. Points are the proportion of variance from posterior medians and whiskers represent the 95% Crl's. Note that birds were cross-fostered at age 2 and brood is the combination of rearing and natal brood (which are both the same at this age).

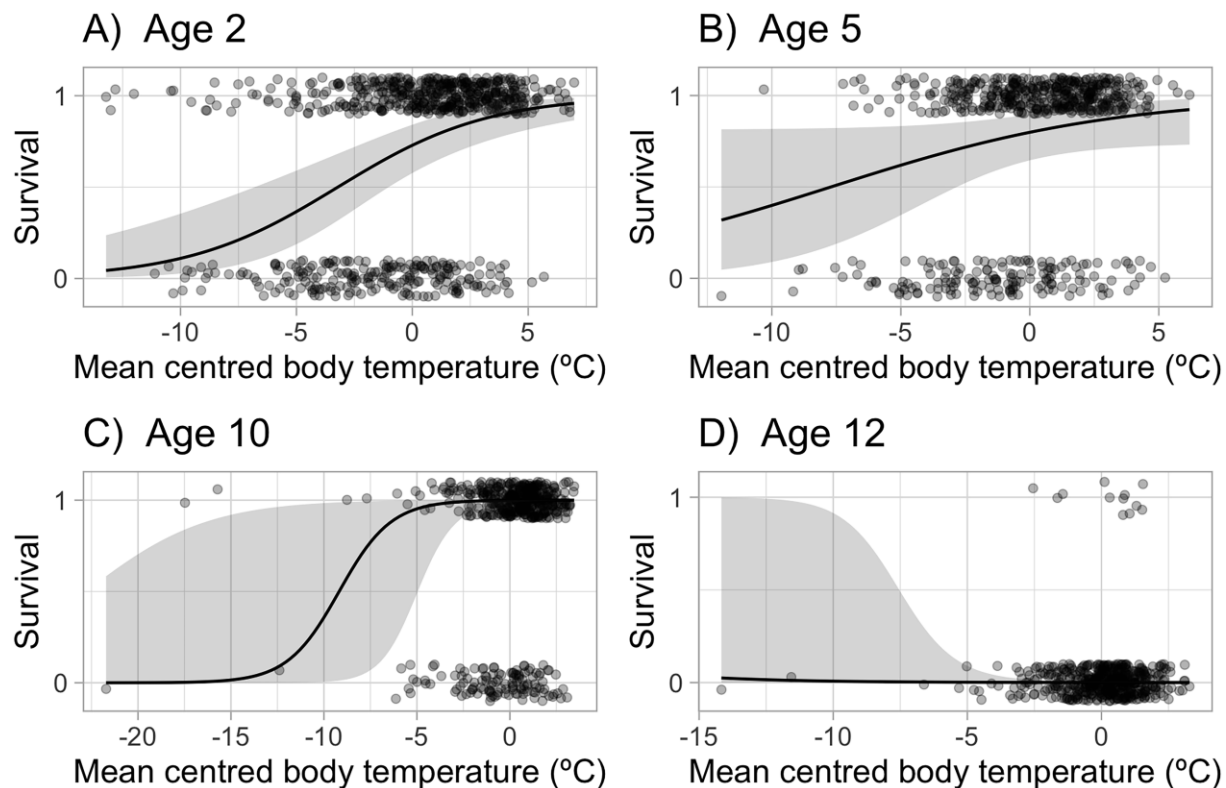
current environment (rearing) but by their prior natal environment. The amount of residual variation in body temperature increased as the nestlings aged, suggesting that environmental variables not captured in our models became increasingly determinative of body temperature. By age 12, only a small proportion of variation was driven by the rearing brood (and negligible variation was driven by genetic or the natal environment) and most variation was explained by residual variation (Table 1, Figure 1).

**Covariance in body temperature across ages**

We found that chicks that hatched in the same nest who have higher temperatures just after hatching (2 days) also have higher temperatures at later ages (5 and 10 days) (Table 2). This relationship weakened slightly when body mass was included in the model, but remained significant between ages 2 and 10 (Table 3). However, by age 12 the correlation with earlier ages disappears (Tables 2 and 3; although note the large confidence intervals for these estimates, likely driven by the







**Figure 2.** Survival given mean centred maximum head temperature ( $^{\circ}\text{C}$ ) for nestlings aged (A) 2, (B) 5, (C) 10, and (D) 12 days old. Points show the raw data and are jittered to aid interpretation. Solid lines are survival predictions from models and shaded regions are the 95% confidence intervals.

brood explained more variation in body temperature than the current environment—the rearing brood. This was the case when body temperature was modelled for each age individually and additionally when selective disappearance was accounted for, where body temperature was largely explained by covariances associated with natal brood effects between ages 2–5 and 2–10.

However, when body mass, as well as selective disappearance, were accounted for, early-stage body temperature did correlate with mid-stage nestling temperature (i.e., ages 2–10 only) but the effects of natal and rearing brood could not be separated for these ages. This does still, importantly, show that early-life effects had significant carry-over effects to the later stages of the nestling period and that this occurred independently from the effects of growth, though further work is needed to determine if it is the natal parents or social parents that drive this variation. Thomson et al. (2017) found in blue tit (*Cyanistes caeruleus*) nestlings, a small effect of the nest of origin and heritability of body mass, and concluded that as nestlings age, body mass is driven more by the social parents. However, our results do not provide evidence for the same trend, as rearing brood explained little variation in any of our models, though this variation was confounded with that of the natal brood for the age 2 effects.

Our findings also show that the drivers of body temperature are fundamentally different to those of body mass, despite being correlated (Supplementary Information S2). Previous studies have shown that nest temperature affects survival (Andreasson et al., 2018; Berntsen & Bech, 2016). Our study, importantly, goes beyond the collective nest effect and explores body temperature at the individual level. Determining the survival effects of body temperature at the individual

level also allowed us to separate this effect from other physiological effects that might impact survival. We concluded that selection acts on body temperature; body temperature predicts survival to the next age, with warmer birds being more likely to survive. The selection on body temperature was independent of body mass, showing the selection effect of body temperature was not simply a result of larger nestlings being selected for. Interestingly, at age 10, body mass did not significantly predict survival, but body temperature did. Numerous other studies have shown that lower nestling body mass is associated with reduced survival (e.g., Monrós et al., 2002; Tinbergen & Boerlijst, 1990; and Ronget et al., 2018, however also see Linden et al., 1992). However, few studies have assessed selection of body mass across ontogeny. Hadfield et al. (2013), showed that selection of larger blue tit nestlings remained strong across ontogeny. Sauve et al. (2024), however, found that selection on larger body masses was stronger in early ontogeny than later. This is consistent with our study, which also showed stronger selection on body mass earlier in the nestling period.

This demonstrates the potential for body temperature to be a key indicator of individual quality during the nestling period. By age 12, this trend disappeared, possibly because by this age the cooler, poor-quality nestlings had already died (i.e., selective disappearance). It is also possible that, after fledging, that many other extrinsic factors lead to the death of an individual and, therefore, the body temperature becomes negligible in explaining mortality. Indeed, very few individuals survived to adulthood, and so the results of survival to adulthood should be treated with caution. Body temperature therefore indicates the quality of a nestling that is independent of growth, although this may not translate into adulthood. It

**Table 4.** Model outputs for survival to the next age class predicted by body temperature at each age. Survival for age 12 was determined by being resighted as an adult. *p*-Values are from likelihood ratio tests.

	Estimate	SE	<i>z</i>	<i>p</i>
<b>Age 2</b>				
<i>Fixed</i>				
(Intercept)	0.99	0.35	2.86	.004
Body temperature	0.31	0.07	4.25	<.001
Body mass	0.91	0.19	4.83	<.001
Year (2019)	3.49	0.66	5.26	<.001
<i>Random</i>				
Variance		SD		
Natal brood	8.88	2.98		
<b>Age 5</b>				
<i>Fixed</i>				
(Intercept)	1.38	0.40	3.45	<.001
Body temperature	0.18	0.10	1.90	.054
Body mass	0.47	0.10	4.66	<.001
Year (2019)	3.65	0.70	5.23	<.001
<i>Random</i>				
Variance		SD		
Natal brood	0.36	0.60		
Rearing brood	7.22	2.69		
<b>Age 10</b>				
<i>Fixed</i>				
(Intercept)	6.57	1.19	5.51	<.001
Body temperature	0.72	0.22	3.32	<.001
Body mass	0.11	0.09	1.25	.21
Year (2019)	5.16	1.90	2.71	<.001
<i>Random</i>				
Variance		SD		
Natal brood	9.40	3.07		
Rearing brood	121.30	11.01		
<b>Age 12</b>				
<i>Fixed</i>				
(Intercept)	−7.84	1.66	−4.72	<.001
Body temperature	−0.28	0.43	−0.64	.45
Body mass	0.10	0.15	0.67	.50
Year (2019)	−18.84	34669.44	−0.001	.12
<i>Random</i>				
Variance		SD		
Natal brood	33.61	5.80		
Rearing brood	6.28	2.51		

would also be interesting, though challenging in terms of the sample size required, to follow these effects throughout adulthood to determine if there are subsequent fitness consequences of nestling body temperature.

## Conclusion

Our results provide insight into what drives variation in body temperature throughout the nestling period and that body temperature is somewhat independent of the effects of growth. This study also demonstrates the importance of determining what drives variation in phenotypic traits in early life, as in the absence of cross-fostering, it would be difficult to separate rearing effects from prehatching incubation effects. Importantly, our study shows that the rearing environment explains very little of the variation in body temperature after endothermy has developed. We have also shown that body temperature is selected for independently of body mass and, therefore, body temperature could provide a useful tool for determining differences in individual quality.

## Supplementary material

Supplementary material is available at *Journal of Evolutionary Biology* online.

## Data availability

Data and R code are available on Zenodo <https://doi.org/10.5281/zenodo.17534342>.

## Author contributions

Lucy A. Winder (Conceptualization [lead], Data curation [equal], Formal Analysis [lead], Investigation [lead], Methodology [lead], Project administration [lead], Visualization [lead], Writing – original draft [lead], Writing – review & editing [equal]), Jacob Hogger Gadsby (Data curation [equal]), Eleanor Wellman (Data curation [equal]), Joel L. Pick (Formal Analysis [supporting], Methodology [supporting], Supervision [equal], Validation [equal], Writing – review & editing [equal]), Julia Schroeder (Data curation [supporting], Resources [equal]), Mirre J. P. Simons (Supervision [equal], Writing – review & editing [equal]), and Terry Burke (Supervision [lead], Writing – review & editing [equal]).

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## Conflict of interest

We have no conflict of interest to declare.

## References

- Andreasson, F., Nord, A., & Nilsson, J.-Å. (2016). Brood size constrains the development of endothermy in blue tits. *Journal of Experimental Biology*, 219, 2212–2219. <https://doi.org/10.1242/jeb.135350>
- Andreasson, F., Nord, A., & Nilsson, J.-Å. (2018). Experimentally increased nest temperature affects body temperature, growth and apparent survival in blue tit nestlings. *Journal of Avian Biology*, 49, jav-01620. <https://doi.org/10.1111/jav.01620>
- Barrett, R. K., & Takahashi, J. S. (1995). Temperature compensation and temperature entrainment of the chick pineal cell circadian clock. *The Journal of Neuroscience*, 15, 5681–5692. <https://doi.org/10.1523/JNEUROSCI.15-08-05681.1995>
- Bates, D., Mächler, M., Bolker, B., & Walker, S. (2015). Fitting linear mixed-effects models using lme4. *Journal of Statistical Software*, 67, 1–48. <https://doi.org/10.18637/jss.v067.i01>
- Berntsen, H. H., & Bech, C. (2016). Incubation temperature influences survival in a small passerine bird. *Journal of Avian Biology*, 47, 141–145. <https://doi.org/10.1111/jav.00688>
- Binkley, S., Kluth, E., & Menaker, M. (1971). Pineal function in sparrows: circadian rhythms and body temperature. *Science*, 174, 311–314. <https://doi.org/10.1126/science.174.4006.311>
- Biro, P. A., & Stamps, J. A. (2010). Do consistent individual differences in metabolic rate promote consistent individual differences in behavior? *Trends in Ecology & Evolution*, 25, 653–659. <https://doi.org/10.1016/j.tree.2010.08.003>
- Bize, P., Metcalfe, N. B., & Roulin, A. (2006). Catch-up growth strategies differ between body structures: Interactions between age and

- structure-specific growth in wild nestling alpine swifts. *Functional Ecology*, 20, 857–864. <https://doi.org/10.1111/j.1365-2435.2006.01157.x>
- Bourne, A. R., Cunningham, S. J., Spottiswoode, C. N., & Ridley, A. R. (2020). High temperatures drive offspring mortality in a cooperatively breeding bird. *Proceedings of the Royal Society B: Biological Sciences*, 287, 20201140. <https://doi.org/10.1098/rspb.2020.1140>.
- Briga, M., & Verhulst, S. (2017). Individual variation in metabolic reaction norms over ambient temperature causes low correlation between basal and standard metabolic rate. *Journal of Experimental Biology*, 220, 3280–3289. <https://doi.org/10.1242/jeb.160069>
- Dawson, D. A., Horsburgh, G. J., Krupa, A. P., ... Burkey, T. (2012). Microsatellite resources for Passeridae species: a predicted microsatellite map of the house sparrow *Passer domesticus*. *Molecular Ecology Resources*, 12, 501–523. <https://doi.org/10.1111/j.1755-0998.2012.03115.x>
- De Zwaan, D. R., Drake, A., Greenwood, J. L., & Martin, K. (2020). Timing and intensity of weather events shape nestling development strategies in three alpine breeding songbirds. *Frontiers in Ecology and Evolution*, 8, 570034. <https://doi.org/10.3389/fevo.2020.570034>
- Dunn, E. H. (1975). The timing of endothermy in the development of altricial birds. *The Condor*, 77, 288–293. <https://doi.org/10.2307/1366224>
- Giloh, M., Shinder, D., & Yahav, S. (2012). Skin surface temperature of broiler chickens is correlated to body core temperature and is indicative of their thermoregulatory status. Contribution from the Agricultural Research Organization, the Volcani Center, Bet Dagan, Israel No. 575/10. *Poultry Science*, 91, 175–188. <https://doi.org/10.3382/ps.2011-01497>
- Hadfield, J. D. (2008). Estimating evolutionary parameters when viability selection is operating. *Proceedings of the Royal Society B: Biological Sciences*, 275, 723–734. <https://doi.org/10.1098/rspb.2007.1013>
- Hadfield, J. D. (2010). MCMC methods for multi-response generalized linear mixed models: The MCMCglmm R package. *Journal of Statistical Software*, 33, 1–22. <https://doi.org/10.18637/jss.v033.i02>
- Hadfield, J. D., Heap, E. A., Bayer, F., ... Crouch, N. M. A. (2013). Disentangling genetic and prenatal sources of familial resemblance across ontogeny in a wild passerine. *Evolution; International Journal of Organic Evolution*, 67, 2701–2713. <https://doi.org/10.1111/evo.12144>
- Henderson, C. R. (1988). Theoretical basis and computational methods for a number of different animal models. *Journal of Dairy Science*, 71, 1–16. [https://doi.org/10.1016/S0022-0302\(88\)79974-9](https://doi.org/10.1016/S0022-0302(88)79974-9)
- Hill, R. W., Beaver, D. L., & Veghte, J. H. (1980). Body surface temperatures and thermoregulation in the black-capped chickadee (*Parus atricapillus*). *Physiological Zoology*, 53, 305–321. <https://doi.org/10.1086/physzool.53.3.30155793>
- Houle, D. (1992). Comparing evolvability and variability of quantitative traits. *Genetics*, 130, 195–204. <https://doi.org/10.1093/genetics/130.1.195>
- Jerem, P., Herborn, K., McCafferty, D., ... Nager, R. (2015). Thermal imaging to study stress non-invasively in unrestrained birds. *Journal of Visualized Experiments*, 105, e53184–e53184. <https://doi.org/10.3791/53184>
- Jerem, P., Jenni-Eiermann, S., McKeegan, D., ... Nager, R. G. (2019). Eye region surface temperature dynamics during acute stress relate to baseline glucocorticoids independently of environmental conditions. *Physiology & Behavior*, 210, 112627–112627. <https://doi.org/10.1016/j.physbeh.2019.112627>
- Kruuk, L. E. B. (2004). Estimating genetic parameters in natural populations using the ‘animal model’. *Philosophical Transactions of the Royal Society of London. Series B: Biological Sciences*, 359, 873–890. <https://doi.org/10.1098/rstb.2003.1437>
- Linden, M., Gustafsson, L., & Part, T. (1992). Selection on fledging mass in the collared flycatcher and the great tit. *Ecology*, 73(1), 336–343. <https://doi.org/10.2307/1938745>
- Loyau, T., Zerjal, T., Rodenburg, T. B., ... Tixier-Boichard, M. (2016). Heritability of body surface temperature in hens estimated by infrared thermography at normal or hot temperatures and genetic correlations with egg and feather quality. *Animal*, 10, 1594–1601. <https://doi.org/10.1017/S1751731116000616>
- Marques-Santos, F., & Dingemanse, N. J. (2020). Weather effects on nestling survival of great tits vary according to the developmental stage. *Journal of Avian Biology*, 51, jav.02421. <https://doi.org/10.1111/jav.02421>
- Mathot, K. J., Martin, K., Kempnaers, B., & Forstmeier, W. (2013). Basal metabolic rate can evolve independently of morphological and behavioural traits. *Heredity*, 111, 175. <https://doi.org/10.1038/hdy.2013.35>
- McCafferty, D. J. (2013). Applications of thermal imaging in avian science. *Ibis*, 155, 4–15. <https://doi.org/10.1111/ibi.12010>
- McFarlane, S. E., Ålund, M., Sirkä, P. M., & Qvarnström, A. (2021). Low heritability but significant early environmental effects on resting metabolic rate in a wild passerine. *The American Naturalist*, 198, 551–560. <https://doi.org/10.1086/715842>
- Metcalf, N. B., & Monaghan, P. (2001). Compensation for a bad start: Grow now, pay later? *Trends in Ecology & Evolution*, 16, 254–260. [https://doi.org/10.1016/S0169-5347\(01\)02124-3](https://doi.org/10.1016/S0169-5347(01)02124-3)
- Monrós, J. S., Belda, E. J., & Barba, E. (2002). Post-fledging survival of individual great tits: the effect of hatching date and fledging mass. *Oikos*, 99, 481–488.
- Mousseau, T. (1998). The adaptive significance of maternal effects. *Trends in Ecology & Evolution*, 13, 403–407. [https://doi.org/10.1016/S0169-5347\(98\)01472-4](https://doi.org/10.1016/S0169-5347(98)01472-4)
- Mueller, A. J., Miller, K. D., & Bowers, E. K. (2019). Nest microclimate during incubation affects posthatching development and parental care in wild birds. *Scientific Reports*, 9, 1–11. <https://doi.org/10.1038/s41598-019-41690-4>
- Nakagawa, S., Ivimey-Cook, E. R., Grainger, M. J., ... Burke, S. (2023). Method Reporting with Initials for Transparency (MeRIT) promotes more granularity and accountability for author contributions. *Nature Communications*, 14, 1788. <https://doi.org/10.1038/s41467-023-37039-1>
- Nilsson, J.-Å., Åkesson, M., & Nilsson, J. F. (2009). Heritability of resting metabolic rate in a wild population of blue tits. *Journal of Evolutionary Biology*, 22, 1867–1874. <https://doi.org/10.1111/j.1420-9101.2009.01798.x>
- Nord, A., & Giroud, S. (2020). Lifelong effects of thermal challenges during development in birds and mammals. *Frontiers in Physiology*, 11, 419. <https://doi.org/10.3389/fphys.2020.00419>
- Nord, A., & Nilsson, J.-Å. (2011). Incubation temperature affects growth and energy metabolism in blue tit nestlings. *The American Naturalist*, 178, 639–651. <https://doi.org/10.1086/662172>
- Nord, A., Chiriac, S., Hasselquist, D., & Nilsson, J.-Å. (2013). Endotoxin injection attenuates rest-phase hypothermia in wintering great tits through the onset of fever. *Functional Ecology*, 27, 236–244. <https://doi.org/10.1111/1365-2435.12003>
- Nord, A., Lehmann, M., MacLeod, R., ... Nager, R. G. (2016). Evaluation of two methods for minimally invasive peripheral body temperature measurement in birds. *Journal of Avian Biology*, 47, 417–427. <https://doi.org/10.1111/jav.00845>
- Ospina, E. A., Merrill, L., & Benson, T. J. (2018). Incubation temperature impacts nestling growth and survival in an open-cup nesting passerine. *Ecology and Evolution*, 8, 3270–3279. <https://doi.org/10.1002/ece3.3911>
- Page, J. L., Nord, A., Dominoni, D. M., & McCafferty, D. J. (2022). Experimental warming during incubation improves cold tolerance of blue tit (*Cyanistes caeruleus*) chicks. *Journal of Experimental Biology*, 225, jeb.243933. <https://doi.org/10.1242/jeb.243933>
- Pick, J. L., Kasper, C., Allegue, H., ... Araya-Ajoy, Y. G. (2023). Describing posterior distributions of variance components: Problems and the use of null distributions to aid interpretation. *Methods in Ecology and Evolution*, 14, 2557–2574. <https://doi.org/10.1111/2041-210X.14200>
- R Development Core Team (2009). R: A language and environment for statistical computing. R Foundation for Statistical Computing.
- Ronget, V., Gaillard, J.-M., Coulson, T., & Gueyffier, F. (2018). Causes and consequences of variation in offspring body mass: Meta-



- analyses in birds and mammals. *Biological Reviews*, 93(1), 1–27. <https://doi.org/10.1111/brv.12329>
- Rønning, B., Jensen, H., Moe, B., & Bech, C. (2007). Basal metabolic rate: Heritability and genetic correlations with morphological traits in the zebra finch: Genetics of BMR and morphological traits. *Journal of Evolutionary Biology*, 20, 1815–1822. <https://doi.org/10.1111/j.1420-9101.2007.01384.x>
- Saino, N., Ambrosini, R., Rubolini, D., ... Caprioli, M. (2018). Carry-over effects of brood size on morphology, reproduction, and lifespan in barn swallows. *Behavioral Ecology and Sociobiology*, 72. <https://doi.org/10.1007/s00265-018-2446-1>
- Saino, N., Ferrari, R., Romano, M., ... Møller, A. P. (2003). Experimental manipulation of egg carotenoids affects immunity of barn swallow nestlings. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 270, 2485–2489. <https://doi.org/10.1098/rspb.2003.2534>
- Sauve, D., Charmantier, A., Hatch, S. A., & Friesen, V. L. (2024). The magnitude of selection on growth varies among years and increases under warming conditions in a subArctic seabird. *Evolution Letters*, 8(1), 56–63. <https://doi.org/10.1093/evlett/grad001>
- Schroeder, J., Nakagawa, S., Rees, M., & Burke, T. (2015). Reduced fitness in progeny from old parents in a natural population. *Proceedings of the National Academy of Sciences*, 112, 4021–4025. <https://doi.org/10.1073/pnas.1422715112>
- Spagopoulou, F., Teplitsky, C., Chantepie, S., ... Gustafsson, L. (2020). Silver-spoon upbringing improves early-life fitness but promotes re-productive ageing in a wild bird. *Ecology Letters*, 23, 994–1002. <https://doi.org/10.1111/ele.13501>
- Tattersall, G. (2017). Thermimage: Thermal image analysis. GitHub.
- Tattersall, G. J., Roussel, D., Voituren, Y., & Teulier, L. (2016). Novel energy-saving strategies to multiple stressors in birds: The ultradian regulation of body temperature. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 283, 20161551.
- Thomson, C. E., Bayer, F., Crouch, N., ... Heap, E. (2017). Selection on parental performance opposes selection for larger body mass in a wild population of blue tits: Selection on parental performance. *Evolution; International Journal of Organic Evolution*, 71, 716–732. <https://doi.org/10.1111/evo.13169>
- Tinbergen, J. M., & Boerlijst, M. C. (1990). Nestling weight and survival in individual great tits (*Parus major*). *The Journal of Animal Ecology*, 59, 1113–1127. <https://doi.org/10.2307/5035>
- Torre-Bueno, J. R. (1976). Temperature regulation and heat dissipation during flight in birds. *Journal of Experimental Biology*, 65, 471–482. <https://doi.org/10.1242/jeb.65.2.471>
- Winder, L. A., White, S. A., Nord, A., ... McCafferty, D. J. (2020). Body surface temperature responses to food restriction in wild and captive great tits. *Journal of Experimental Biology*, 223. <https://doi.org/10.1242/jeb.220046>
- Winney, I., Nakagawa, S., Hsu, Yu-H., ... Schroeder, J. (2015). Troubleshooting the potential pitfalls of cross-fostering. *Methods in Ecology and Evolution*, 6, 584–592. <https://doi.org/10.1111/2041-210X.12341>