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Trait networks reveal turnover in Caribbean corals and changes in community resilience through the Cenozoic

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Abstract

The present-day climate crisis is transforming coral reef communities, potentially undermining ecosystem functioning. Evolutionary trade-offs between species traits result in diverse life-history strategies, enabling corals to survive disturbance events through specific adaptive mechanisms. Trait–trait relationship networks offer insights into trait turnover and changing life-history strategies during environmental changes. Paleoecological insights from the fossil records can further illustrate how species adapt to environmental shifts, highlighting resilience traits.

We highlight coral traits that promote resilience in the Caribbean based on fossil occurrences and morphological traits, examining biological determinants of species and trait turnover across the Cenozoic. We use traits that underpin the survival of corals during disturbances, for example, corallite diameter, colony growth form, corallite integration, and budding type. We analyzed species turnover and extinctions with a bipartite network and explored trait turnover with trait–trait co-occurrence networks based on 4268 species records at 421 sites over ~40 Myr.

Our findings support existing evidence that species turnover coincided with major environmental and biogeographic changes across the Cenozoic. Additionally, our results provide new insight into functional changes throughout the Cenozoic. Past cooler climates favored corals with a fast growing and reproducing (competitive) life-history strategy, which boosts short-term success, but also increases susceptibility to diseases and thermal stress. Cenozoic species and trait turnover occurred during environmental change, corroborating expectations of such turnover in the future. We found trait co-occurrence modules associated with competitive and stress-tolerant life-history strategies. The transition from the “greenhouse” (Paleogene) to the “icehouse” (Neogene) climate over ~40 Myr favored competitive traits, which supported fast-growing, shallow reefs. With rising temperatures and declining *Acropora* in the Caribbean, future reefs may resemble Eocene reefs: dominated by stress-tolerant, slow-growing corals adapted to marginal environments.

Non-technical Summary

Coral reefs are among the most diverse ecosystems on Earth and are facing significant changes due to the current climate crisis. These changes threaten the many important functions these reefs perform. Corals have different survival strategies, adapting to changes in their environment through various traits. By studying the relationships between these traits, we can understand how coral species have changed over time and how they might respond to future environmental changes. The fossil record provides valuable insights into how coral communities have adapted to past climate changes and highlight traits that helped them survive.

In our research, we looked at the fossil records of Caribbean corals and their traits, such as the size and shape of their skeletons, to identify which traits have helped them persist through past climatic changes. We analyzed data from 4268 coral record occurrences from 421 sites over 40 million years, using network analysis to explore how coral species and their traits have changed.

Our findings suggest that corals of the late Cenozoic possessed traits adapted to cooler climates, which may now be detrimental to survival in a warming world. Cooler climates in the recent past favored corals that are now more vulnerable to diseases and heat stress. The turnover of coral species and traits in the past occurred during environmental change, indicating that similar shifts are likely in the future. We discovered that branching corals with small corallites, moderate corallite integration, and asexual reproduction through extracalicular budding thrived in cooler intervals, allowing them to form fast-growing, shallow reefs.

As temperatures rise and certain coral species decline in the Caribbean, future coral reefs may resemble those from the Eocene epoch, dominated by slow-growing, stress-tolerant corals that thrive in less favorable environments.

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Introduction

The current climate crisis is substantially transforming coral reef ecosystems globally (Pandolfi and Jackson 2007; Hughes et al. 2018; Cook et al. 2022), with altered and reduced ecosystem functioning due to species losses and community and trait turnover beyond natural rates (Hodapp et al. 2023). The ecological mechanisms and consequences of such changes in ecosystem functioning remain unknown, but past patterns of species, community, and trait turnover can provide insight into future shifts in community diversity and structure (Payne and Clapham 2012; Pandolfi and Kiessling 2014; O'Dea et al. 2017; Raja et al. 2021).

Ecosystem functioning is defined as the flux of energy, nutrients, and organic matter through the environment (Brandl et al. 2019). Differences in species traits enable species to occupy different functional niches, influencing ecosystem functioning (Hooper et al. 2005; Hughes et al. 2017; Brandl et al. 2019). Coral reefs, with their high diversity, exhibit niche partitioning (Loreau and Hector 2001), but also significant functional redundancy due to species sharing similar niches (Yachi and Loreau 1999; Fonseca and Ganade 2001). High functional redundancy can buffer against species loss, therefore maintaining ecosystem functions despite species turnover (Yachi and Loreau 1999; Fonseca and Ganade 2001).

As environmental change intensifies, the magnitude and rate of community turnover will likely increase, due to the increasing likelihood of species range expansions (Vergés et al. 2014, 2019; Gilson et al. 2021; Clay et al. 2023) and contractions (Stuart-Smith et al. 2021), along with regional and global extinctions (Harnik et al. 2012). The resulting transformed communities may harbor fewer species, but retain the main functions, essentially functioning with a “skeleton crew,” much like communities in the geological past after mass extinction events (e.g., Foster and Twitchett 2014; Dunhill et al. 2018).

Examining trait turnover in coral reef communities, especially in response to environmental changes, provides insights into community functional resilience. There are often trade-offs between different traits, for example, branching corals have an increased growth rate that allows them to outcompete competitors for space on the reef (Baird and Hughes 2000); however, this strategy requires a high energy investment, meaning less energy is available for stress responses (Pinzón et al. 2014). These evolutionary trade-offs lead to different life-history strategies, which can be thought of as existing on a spectrum from “fast” (r-selected) and “slow” (K-selected) life histories (Pianka 1970; Darling et al. 2012; Padma et al. 2021). Changes in environmental conditions may favor one strategy over another. Corals with an r-selected or competitive life-history strategy are fast growing with high fecundity and high mortality (e.g., *Acropora*) and the K-selected or stress-tolerant corals are slow growing and long-lived but with lower fecundity (e.g., *Porites*) (Sleeman et al. 2005; Darling et al. 2012).

Co-occurrence networks allow for the visualization of how species are associated based on their presence or absence, and therefore the turnover in ecological interactions between species (Ulrich et al. 1991; Araújo et al. 2011; Harris 2016; Morueta-Holme et al. 2016; Matich et al. 2017). Combining these networks with trait-based analysis allows for quantifying trait co-occurrences, linking to community resilience or vulnerability (He et al. 2020; Clay et al. 2024a). Network metrics, such as modularity, edge density, degree centrality, and node degree, in trait co-occurrence networks quantify a community's ability to recover from a disturbance (Flores-Moreno et al. 2019; Gao et al. 2022), making both species and trait co-occurrence networks valuable for understanding community turnover and resilience post-environmental disturbance.

A variety of modeling techniques can predict coral community change in response to future climate change (Donner et al. 2018). These models are often based on the physiological, ecological, and evolutionary responses of corals to current or very recent climate change; however, there is still a great deal of uncertainty concerning how reefs will respond to the degree of climate change predicted (Pandolfi 2015). The understanding required to predict future responses to environmental change can come from paleoecology, as the fossil record provides evidence of how communities have responded to the scale of change predicted over the coming decades to centuries, which is as yet unprecedented in human history (O'Dea et al. 2017). The Cenozoic (66 Ma to present) (Cohen et al. 2013) has seen temporal fluctuations in biodiversity and functional patterns within shallow reef ecosystems. The Cenozoic Era may provide insight into the future impacts of climate change, as conditions vary frequently throughout the interval, with some reflecting current and predicted future conditions (Norris et al. 2013).

Fluctuating environmental conditions and regional biogeographic changes in the Caribbean led to corals' diversity and abundance variations throughout the Cenozoic (Johnson et al. 2008, 2009; Roff 2021; Yasuhara et al. 2022; Fig. 1). There were multiple periods of environmental change between the end of the Eocene and the late Pleistocene that may provide insight into coral trait changes in response to environmental change. In this paper, we will examine three of these periods of change: (1) Reductions in sea-surface temperature (SST) and sea level across the Eocene/Oligocene boundary (~33.9 Ma) caused previously algal-dominated reefs (Scheibner and Speijer 2008; Mutti et al. 2011; Takayanagi et al. 2012) to become large architectural reefs (Johnson et al. 2008, 2009; Miller et al. 2020; Yasuhara et al. 2022). (2) Throughout the Miocene, the isolation of the Caribbean from other reef areas, along with tectonic and ocean current changes (Yasuhara et al. 2022), contributed to species turnover. This isolation led to shared ecological characteristics across the Caribbean and ultimately the evolution of endemic Caribbean corals, which are the ancestors of today's Caribbean species (Johnson and Kirby 2006). (3) The formation of the Isthmus of Panama in the Pliocene triggered a regional extinction event (Roff 2021), reducing coral diversity by up to 80% (Budd et al. 2011), but paving the way for a surge in reef growth during the late Pleistocene (O'Dea et al. 2007; Johnson et al. 2008; Klaus et al. 2012). The recovery of reef growth in the late Pleistocene was mainly driven by the genus *Acropora* (Klaus et al. 2012; Renema et al. 2016), and its associated traits such as asexual reproduction and fast growth rates (Roff 2021). Today, the Caribbean is threatened by environmental and anthropogenic change, with declining coral abundance, diversity, functional redundancy (Jackson et al. 2014), and low resilience to disturbance (Connell 1997; Jackson et al. 2014; Cramer et al. 2020). Exploring species and community turnover on Caribbean coral reefs may help determine whether there is an evolutionary historic reason for their current low resilience in comparison to Indo-Pacific reefs (Roff 2021).

We can predict how modern coral reefs might respond to environmental change at the trait level by assessing when environmental change pushed species turnover into trait turnover in the past. Here, we quantify to what degree large-scale environmental change, of similar magnitude to modern-day climate change, has transformed coral communities, changed the complexity of trait networks, and thus affected ecosystem function. We test the following hypotheses: (1) Periods of species turnover and extinctions across the Cenozoic Era coincide with changing environmental conditions. (2) In addition, species turnover is associated with trait

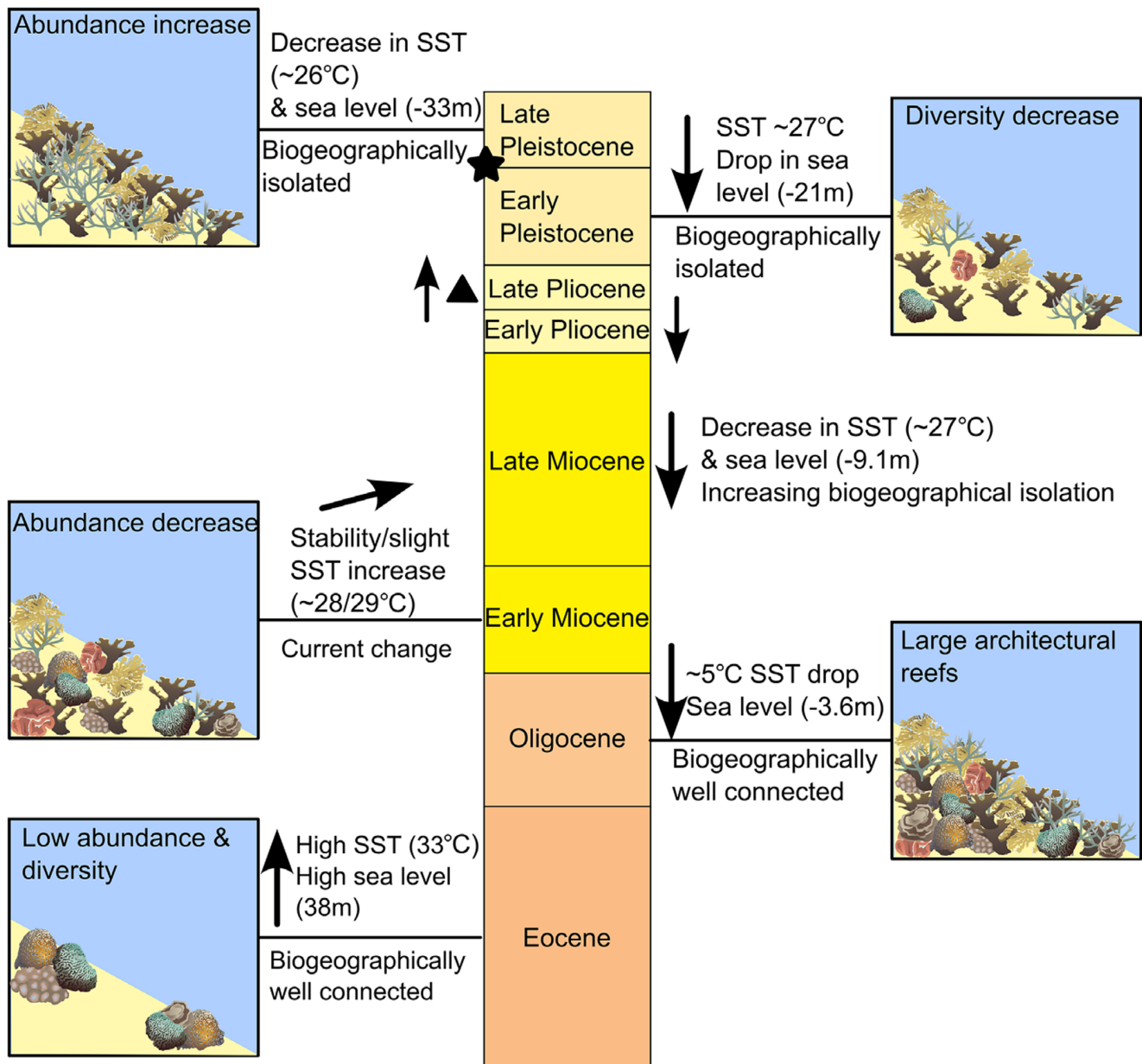


Figure 1. Environmental and biogeographic change across the Cenozoic Era (~56 Ma to ~0.0117 Ma) and how these changes affected coral abundance and diversity. Central column = epochs/sub-epochs, size relates to the length of time (Cohen et al. 2013). Boxes reflect changes in abundance and diversity of corals (Johnson et al. 2008, 2009; Budd et al. 2011; Steinthorsdottir et al. 2021). Arrows indicate the direction of change in sea-surface temperatures (SSTs) (°C) and sea level (SL) (m) (Miller et al. 2020; Rae et al. 2021). Caribbean regional biogeographic changes are also indicated (Roff 2021; Yasuhara et al. 2022). The triangle indicates when the closure of the Panama Isthmus occurred (O'Dea et al. 2016), and the star indicates the early/late Pleistocene regional extinction event (Budd 2000). Coral icons from vecta.io (<https://vecta.io>).

turnover across the Cenozoic Era. (3) Particular traits or trait relationships were integral to surviving major periods of change, such as those associated with a competitive or stress-tolerant life-history strategy. Testing these hypotheses will enhance our understanding of the mechanisms underpinning coral survival and extinction.

Methods

Data Acquisition

We downloaded coral occurrence data from the Paleobiology Database (PBDB; <https://www.paleobiodb.org>, accessed 1 October 2022), using the following parameters: taxa = Anthozoa; time

interval = Cenozoic; longitude min. = -105.249, max. = 40.5176; latitude min = 7.1881, max = 30.6001 (to capture the western equatorial Atlantic, including the Caribbean Sea, the Gulf of Mexico, and the Bahamas). The Cenozoic Era is divided into seven epochs (Paleocene, Eocene, Oligocene, Miocene, Pliocene, Pleistocene, and Holocene), three of which (Miocene, Pliocene, and Pleistocene) are further divided into sub-epochs (Cohen et al. 2013). We filtered the data to remove occurrence data from the Paleocene, middle Miocene, middle Pleistocene, and Holocene, due to a lack of data availability for these epochs/sub-epochs. This left only the epochs and sub-epochs of interest: Eocene (56.0–33.8 Ma), Oligocene (33.9–23.03 Ma), early Miocene (23.03–15.98 Ma), late Miocene (11.63–5.33 Ma), early Pliocene (5.33–3.60 Ma), late

Pliocene (3.60–2.58 Ma), early Pleistocene (2.58–0.77 Ma), and late Pleistocene (0.129–0.0117 Ma).

We downloaded species-level occurrence records as collections, which represent records collected during the same paleontological survey and are used here to represent communities. We excluded any collections with fewer than five occurrences to ensure sufficient data for community-level analysis. We carried out analysis at the species level, as although both modern and paleontological coral taxonomy is highly debated (Lathuilière 1996; Kitahara et al. 2016; Ramírez-Portilla et al. 2022), analysis at the genus level would mean a great loss in trait variation and would not capture the full range of functional diversity. We checked coral species' names against the World Register of Marine Species (WoRMS) using the R package *worms* (Chamberlain and Vanhoorne 2023) to account for recent updates in coral taxonomy (Hoeksema and Cairns 2025). We only included reef-building coral species occurrences, due to the lack of available trait data for non-reef building corals. We identified azooxanthellate species and any other non-reef building species using a combination of WoRMS (Chamberlain and Vanhoorne 2023), PBDB (<https://www.paleobiodb.org>, accessed 1 October 2022), Encyclopedia of Life (EOL 2022), and data from Raja et al. (2021) and removed them from the database. The final scleractinian coral occurrence database includes 421 collections from a range of sites across the western equatorial Atlantic, hereafter referred to as the Caribbean, and a range of epochs within the Cenozoic Era (Fig. 2; Supplementary Table 1).

We collated coral traits from the Neogene Marine Biota of Tropical America (NMITA) (Budd et al. 2001), Raja et al. (2021),

and CoralTraits.org (Madin et al. 2016) databases, along with primary literature (Duncan 1864, 1873; Vaughan 1899, 1919; Vaughan and Hoffmeister 1926; Wells 1934a,b, 1935, 1945; Vaughan and Wells 1943; Durham 1946; Weisbord 1971, 1973, 1974; Frost and Langenheim 1974; Cairns and Wells 1987; Budd et al. 1992, 2019; Bosellini 1995; Cairns 1995; Filkorn et al. 2005; Locke et al. 2007; Pandolfi 2007; Budd and Wallace 2008; de Araújo Távora 2016; Flórez et al. 2019). Species traits are often variable, with intraspecies variation seen across different environments (e.g., colony growth form plasticity along depth gradients) (Smith et al. 2007; Todd 2008; Doszpot et al. 2019); however, we have focused on interspecies variation to capture broad community-level changes across the Caribbean over such a large time frame (~ 54 Myr) and therefore used the most common trait for each species. Life-history strategies are linked to survival during environmental disturbances (Darling et al. 2012; van Woesik et al. 2012), as are traits related to extinction risk, such as colony growth form, corallite integration, maximum corallite diameter, and budding type (Raja et al. 2021; Table 1). Colony growth form indicates the morphology of corals; we created four categories based on data availability (Table 1). It is also worth noting that coral morphology influences preservation bias, with branching corals more susceptible to fragmentation reducing their likelihood of fossilization (Webb 1996); therefore, the presence of branching corals may be higher than the fossil record indicates. Corallite integration refers to the arrangement of corallites within the colony (Raja et al. 2022). We split corallite integration into the same eight categories used by Raja et al. (2021; Table 1). Maximum corallite diameter was split into four categories,

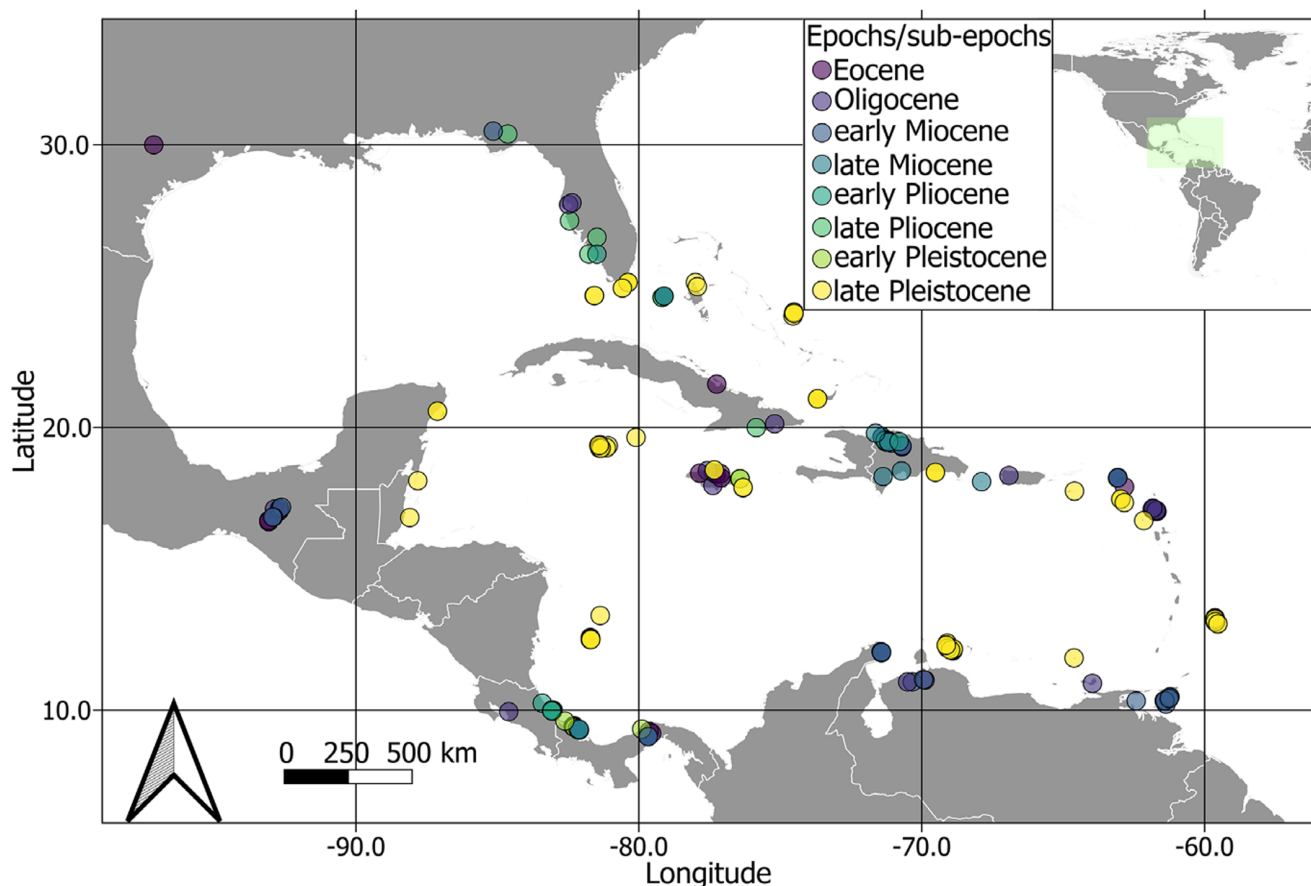


Figure 2. Sites of the 421 collections within the western equatorial Atlantic, obtained from the Paleobiology Database and used in this study. Colors indicate which epoch/sub-epoch the collection belongs to.

Table 1. The four morphological traits of corals used explore trait turnover during the Cenozoic, with category information, life-history strategy group, and short comments on trait relationships with the two life-history strategy groups: stress-tolerant (K-selected) and competitive (r-selected)

Trait	Categories	Life-history strategy	Relation to life-history strategy
Colony growth form (G)	Branching	Competitive	Branching species grow faster than other growth forms, making them likely to outcompete for space and light on reefs (Baird and Hughes 2000).
	Massive	Stress tolerant	Higher growth rates require more energy investment, which means less energy is available for stress response; whereas slow-growing, massive growth-type corals are more stress tolerant (Pinzón et al. 2014).
	Encrusting	NA	Fall within the “domed” category used by Darling et al. (2012), which also included massive and submassive growth forms. Likely stress tolerant, weedy, or generalists, but without further reproductive information (i.e., brooding or not) they cannot be correctly placed in a category.
	Platy	NA	Darling et al. (2012) associated plating corals with a competitive life strategy; however, platy corals can also be stress-tolerant due to their ability to survive in deeper waters and turbid areas with low light conditions (Rosen et al. 2000). Therefore, they are difficult to place in a category.
Corallite integration (C)	0, Solitary	Stress tolerant	Solitary corals are non-colonial and recover quickly following extinction events; for example, they were the first to reappear in the fossil record in Argentina following the End-Triassic Mass Extinction (Echevarría et al. 2017). Solitary corals have a large corallite diameter and are massive or encrusting in our database.
	1, Dendroid	NA	In dendroid colonies, the corallites branch irregularly, with spaces between each corallite (Raja et al. 2021, 2022). As dendroid colonies are branching, they could be considered competitive, but due to their low level of corallite integration, they likely have lower photosymbiotic ability (Stanley et al. 2018), making them more likely to be generalists.
	2, Phaceloid	NA	Phaceloid colonies also have branching corallites but with coenosteum present at the wall base. Corallites are separated and subparallel (Raja et al. 2021, 2022). As with dendroid colonies, they are associated with branching corals but are likely generalists rather than competitive.
	3, Plocoid	Competitive	In plocoid colonies, corallites have their own walls and are separated by coenosteum (Raja et al. 2021, 2022). Sixty-seven percent of coral species in our trait database classed as branching had plocoid coral integration, as did all the <i>Acropora</i> species; therefore, we have classified them as competitive.
	4, Cerioid	NA	Cerioid colonies have juxtaposed corallites that share a common wall (Stanley et al. 2018). Associated with genera such as <i>Porites</i> , <i>Favites</i> , and <i>Goniastrea</i> in our database, so they could be stress tolerant, weedy, or generalist (Darling et al. 2012) but cannot be placed without further reproductive information.
	5, Asteroid/ thamnasteroid	NA	Asteroid and thamnasteroid colonies are separated by walls, but the boundaries are not well defined, and septa are not joined (Stanley et al. 2018; Raja et al. 2021). Associated with the genera <i>Leptoseris</i> , <i>Mycetophyllia</i> , and <i>Pavona</i> in our database, so they are likely weedy or generalist.
	6, Meandroid	NA	Corallites are arranged in multiple series, often valley-like (Stanley et al. 2018; Raja et al. 2021). Flabello-meandroid colonies are similar to meandroid, but the corallites are arranged in a single series (Raja et al. 2022). High integration levels are correlated with photosymbiosis (Stanley et al. 2018). Highly integrated coral species might be more vulnerable to thermal stress (Swain et al. 2018) and less likely to survive extinction events (Stanley et al. 2018). In our database, meandroid and flabello-meandroid colonies had large-diameter corallites and were often massive or encrusting, usually traits associated with stress tolerance, so we were unable to assign them a life-history strategy.
	7, Flabello-meandroid	NA	
Maximum corallite diameter (D)	Small (0–3 mm)	Competitive	Small corallites are linked to higher photosynthetic autotrophy (Conti-Jerpe et al. 2020; Dimitrijević et al. 2024) and are usually associated with branching corals. All the branching <i>Acropora</i> fall into this category.
	Medium (4–7 mm)	NA	Intermediate corallite diameter, associated with a range of genera such as <i>Favia</i> , <i>Montastraea</i> , and <i>Oculina</i> , as well as a range of colony and growth-form types, thus making it difficult to assign a life-history strategy.
	Large (8–15 mm)	Stress tolerant	Larger corallites indicate heterotrophic feeding (Crabbe and Smith 2006) and may support better survival during thermal stress via heterotrophy (Hughes and Grottoli 2013). The large corallite diameter category covers mostly massive species. The very large category covers most <i>Mycetophyllia</i> and some massive growth-type corals, as well as the solitary corals.
	Very large (>15 mm)	Stress tolerant	
Budding type (B)	None	Stress tolerant	No budding occurring. This is generally associated with solitary corals, which generally do not reproduce asexually, although some species, such as those of the genus <i>Fungia</i> , can reproduce asexually (Gilmour 2004). We have assigned budding type none as stress-tolerant due to its association with solitary corals.
	Intracalicular	Stress tolerant	Budding occurs within the tentacle ring of the parent polyp (Raja et al. 2022). Intracalicular polyps are sexually mature when they bud, allowing rapid repopulation of communities following a disturbance, but extracalicular polyps are developmentally young (Sakai 1998).

(Continued)

Table 1. (Continued)

Trait	Categories	Life-history strategy	Relation to life-history strategy
	Extracalicular	Competitive	Budding occurs outside the tentacle ring of the parent polyp (Raja et al. 2022). Extracalicular budding has been linked to more effective light attenuation increasing photosynthesis rates and therefore increasing energy intake and growth (Enriquez et al. 2017).
	Both	NA	Both intra- and extracalicular budding can occur, meaning the life-history strategy cannot be determined.

as categorical data were required for trait network construction (Table 1). We included maximum corallite diameter even though it had a low importance for modern corals in Darling et al. (2012), because it was important for predicting extinction risk in fossil corals (Raja et al. 2021), and here we want to create comparable analyses. Budding type refers to where new buds form in relation to the corallite wall of the parent polyp during asexual reproduction (Raja et al. 2022). The lack of reproductive trait data for extinct species may have reduced the power of the study. Therefore, we included budding type in our analysis, even though it was the least important variable for extinction risk in Raja et al. (2021). This is because it is the only morphological trait we have related to reproduction and reproductive traits and their evolutionary history likely drives the survival of coral communities (Roff 2021). Budding type was split into four categories (Table 1). These traits capture the main factors in coral morphology that underscore coral resilience to extinction and were used to split coral species broadly into the two ends of the r-K continuum: competitive and stress tolerant (Table 1). It is worth noting that life-history strategies exist as a continuum (Stearns 1977), and so our categorization of traits as competitive or stress-tolerant is likely an oversimplification. Combinations of different traits place species along this continuum, with very few species fitting perfectly into any one life-history strategy, as highlighted by the “generalist” strategy identified in Darling et al. (2012).

Species and Trait Turnover across the Cenozoic Era

To test our first hypothesis, that species turnover coincided with environmental drivers, we constructed a bipartite network. We created a bipartite network spanning from the Eocene to the Pleistocene using a time bin by species matrix to show species turnover across the Cenozoic (Fig. 3A). We applied the *igraph* (v. 1.4.1) package function *bipartite.mapping* to ensure bipartite projection. We applied the layout “*layout.fruchterman.reingold*” to visualize all nodes within the network. The final network was edited using *Inkscape* (v. 1.2.2) to improve the visibility of all nodes and edges. We visualized the numbers of epoch/sub-epoch-specific and shared species via a heat map using the R package *ggplot2* (v. 3.4.2) (Wickham 2016; Fig. 3B). Sampling bias is an issue for all paleontological studies, with heterogeneous sampling effort and differential preservation leaving our understanding of spatiotemporal species distributions incomplete (Kiessling 2005). We chose to focus on fossil occurrence (i.e., presence/absence) rather than abundance due to the difficulty in distinguishing between true abundance and preservational artifacts (Kiessling 2008). Some species may be missing from the bipartite network, potentially skewing our turnover results; however, as the bipartite projection is not weighted by abundance, it is not influenced by uneven preservation of the species sampled in each time interval.

We calculated the Jaccard dissimilarity of coral species using the *Vegan* package (v. 2.6.4) (Oksanen et al. 2020) to quantify species turnover between each of the neighboring epochs and sub-epochs using presence/absence data.

To test our second hypothesis that trait turnover coincided with species turnover, we calculated Jaccard dissimilarity of coral traits using the same methods as for species Jaccard dissimilarity. Species and trait dissimilarity was plotted for comparison (Fig. 4, Supplementary Fig. 1).

Trait Co-occurrences across Major Periods of Change

To test our third hypothesis, that particular traits or trait relationships were integral to surviving major periods of change, we used trait–trait co-occurrence networks (Clay et al. 2024a). We constructed trait–trait co-occurrence networks for three major periods of environmental change that coincided with periods of high species and/or trait turnover identified during the testing of our first and second hypotheses (Eocene–Oligocene, early Miocene–late Miocene, late Pliocene–early Pleistocene). To account for the prevalence of each trait within each collection, we calculated community weighted means (CWMs) for each collection within each epoch/sub-epoch by combining a presence/absence site-by-species matrix with a species-by-trait matrix, using the *FD* package (v. 1.0.12.1) in R (Laliberté et al. 2014). To account for the differing numbers of collections between epochs/sub-epochs, we randomly subsampled each group to match the smallest available number of collections (e.g., 13 of the 23 Oligocene collections, as only 13 collections were available from the Eocene; early Miocene = 40, late Miocene = 38, late Pliocene = 36, early Pleistocene = 98). We repeated the random sampling 10 times, with networks constructed for each iteration. We calculated a trait–trait relationship matrix by applying Pearson’s correlation to the collection-by-CWM matrix. We set a significance threshold of $p < 0.05$, with correlations below the threshold assigned as one and those above assigned as zero (He et al. 2020; Clay et al. 2024a). This approach allowed us to analyze a network of significant trait–trait relationships in *igraph* (v. 1.4.1) (Csardi and Nepusz 2006). All 10 randomly sampled networks were constructed for each sub-epoch (Supplementary Figs. 2–4). The networks presented in the “Results” represent the closest to the mean values for edge density and modularity (Figs. 5–7). It is also possible that important trait–trait relationships have been overlooked due to missing species or trait information, but restricting our study to the well-sampled Caribbean region means spatial sampling bias is likely not a major issue, as it would be with a global study (Vilhena and Smith 2013).

Following network construction, we calculated three network metrics using *igraph* (v. 1.4.1): node degree, edge density, and modularity. Node degree measures the importance of a node within a network, a trait with a high node degree is integral to the network and can be thought of as a “keystone” trait (Flores-Moreno et al. 2019;

Gao et al. 2022; Clay et al. 2024a). A higher node degree is ecologically preferable, as networks that are made up of one or two keystone traits may be functionally vulnerable, with the loss of one or both those traits leading to the breakdown of the network and thus the loss of ecosystem functioning (Mouillot et al. 2014; Gao et al. 2022). Edge density measures how well connected a network is by quantifying the number of edges within a network. Highly connected networks can infer high environmental marginality (Flores-Moreno et al. 2019), potentially indicating high trait matching and an increased abundance of generalist species. Modularity refers to the presence of distinct subgroups or neighborhoods, where groups of traits have more connections with one another than with the rest of the network (Flores-Moreno et al. 2019). High modularity within a network likely indicates high functional diversity of the ecosystem. Modules may indicate different life strategies or functional niches, meaning high modularity within a network likely indicates high functional diversity, that is, more modes of life and, therefore, higher resilience to disturbance (Liu et al. 2021). We incorporated metrics into the visualization of the network, where modules are indicated by gray ellipses around nodes within the same module, and node degree is indicated by the size of the nodes (Figs. 5–7).

As trait–trait co-occurrence networks do not include all possible traits, but only those traits that are significantly correlated, we also created simple weighted edge networks to visualize all possible trait connections. We calculated the edge density for the weighted edge networks and compared it with the edge density of the trait–trait co-occurrence networks to explore how many of the possible trait connections were deemed significant in the trait–trait co-occurrence networks.

To quantify community overlap between the epochs/sub-epochs compared through trait–trait networks and to identify which traits drive the separation of communities we used principal component analysis (PCA). We calculated a Euclidean distance matrix using the numerical CWM for each collection within each epoch/sub-epoch using the function *prcomp* and visualized it in multidimensional space using PCA (Figs. 5C, 6C, 7C). In addition to exploring community changes, we quantified changes in trait space between the neighboring epochs/sub-epochs, using principal coordinate analysis (PCoA). Comparing trait space allows visualization of the functional richness of each epoch/sub-epoch. We used convex hulls to visualize this, placing species in multidimensional space based on their functional dissimilarity (Magneville et al. 2022). A larger trait space indicates higher functional richness (Cornwell et al. 2006; McWilliam et al. 2018). We calculated Gower's (1983) dissimilarity matrices for all time bins using *funct.diss* (de Bello et al. 2021). To calculate and assess the quality of functional trait space, we used the mFD package (v. 1.0.4) to calculate functional richness across four axes (Magneville et al. 2022). We constructed a global hull that included all species and their traits and then created regional hulls by subsetting species by each time bin (Laliberté et al. 2014; Floyd et al. 2020; Clay et al. 2024b). To explore which traits were driving the functional separation of species, we plotted trait vectors on the regional hulls using the *vegan* package (v. 2.6.4) from R (Oksanen et al. 2020; Supplementary Figs. 5d, 6d, 7d).

Results

Turnover of Species and Traits across the Cenozoic

The bipartite network revealed a continuous species turnover across the Cenozoic (Fig. 3A). Species richness increased between

the Eocene ($N = 69$) and Oligocene ($N = 86$), before falling during the early Miocene ($N = 70$), late Miocene ($N = 71$), and early Pliocene ($N = 72$), rose slightly in the late Pliocene ($N = 82$) before falling again in the early ($N = 74$) and late Pleistocene ($N = 40$). Fifty-one of the species recorded are currently extant today, with 4 originating in the Oligocene, 5 in the early Miocene, 17 in the late Miocene, 6 in the early Pliocene, 8 in the late Pliocene, 10 in the early Pleistocene, and 1 in the late Pleistocene (Fig. 3A). There were also five species identified as regionally extinct in the Caribbean (*Gardineroseris planulata*, *Leptoseris gardineri*, *Leptoseris glabra*, *Meandrine brasiliensis*, and *Mussismilia hispida*) but extant elsewhere. All five of the regionally extinct species originate in the Miocene (1, early Miocene; 4, late Miocene) (Fig. 3A). The largest turnover in species occurs between the Eocene and Oligocene, with only 28% of species from the Eocene surviving through to the Oligocene (Fig. 3B) (Oligocene–early Miocene = 53%, early Miocene–late Miocene = 46%, late Miocene–Pliocene = 77%, early Pliocene–late Pliocene = 78%, late Pliocene–early Pleistocene = 62%, early Pleistocene–late Pleistocene = 88%).

Jaccard dissimilarity indicates species turnover to be the greatest between the Eocene and the Oligocene (0.86) (Fig. 4). Dissimilarity decreases between the Oligocene and early Miocene (0.58) before increasing during the early Miocene/late Miocene turnover (0.71). There is another decrease in Jaccard dissimilarity between the late Miocene and early Pliocene (0.38) (Fig. 4, Supplementary Fig. 1). Dissimilarity then increases gradually across the rest of the Cenozoic (early Pliocene–late Pliocene = 0.43, late Pliocene–early Pleistocene = 0.51, early Pleistocene–late Pleistocene = 0.56) (Fig. 4, Supplementary Fig. 1).

Trait Co-occurrences across the Eocene–Oligocene Cooling Period

Across the Eocene–Oligocene cooling period, edge density is reduced in the trait–trait co-occurrence networks for both epochs (Eocene = 0.12, Oligocene = 0.14 [mean $\pm$ 0.04 SD]), in comparison to the weighted edge networks (Eocene = 0.74, Oligocene = 0.96 [mean $\pm$ 0.03 SD]). Several traits present in the Eocene communities are not present in the trait–trait relationship network: *corallite integration 3 (plocoid)*, *corallite integration 7 (flabello-meandroid)*, *small corallite diameter*, and *encrusting colony growth form* (Supplementary Fig. 3A,C). *Platy colony growth form* is missing from the Eocene communities completely (Supplementary Fig. 3A,C). In the Oligocene, *corallite integration 1 (dendroid)* and *7 (flabello-meandroid)* are missing from the communities (Supplementary Fig. 3B,D). Several traits do not appear in all 10 randomly sampled weighted networks (numbers in brackets indicate how many networks they do appear in): *budding type none* (8), *corallite integration 0 (solitary)* (9), *corallite integration 2 (phaceloid)* (7), and *colony growth form encrusting* (9). All traits present in the Oligocene weighted edge network are present in the trait–trait relationship network (Supplementary Fig. 3B,D), but many of these traits are not present in all of the 10 randomly sampled networks: *budding types both* (7), *extra* (7), and *none* (8); *corallite integration 0 (solitary)* (9), *2 (phaceloid/reptoid)* (5), *3 (plocoid)* (6), and *4 (cerioid)* (9); *corallite diameters large* (7) and *very large* (4); and *colony growth forms encrusting* (9) and *massive* (9) (Table 2).

Trait–trait relationships become more complex following the Eocene–Oligocene cooling event, with the slight increase in edge density (Eocene = 0.12, Oligocene = 0.14 [mean $\pm$ 0.04 SD]) that indicates more connections in the Oligocene than in the Eocene

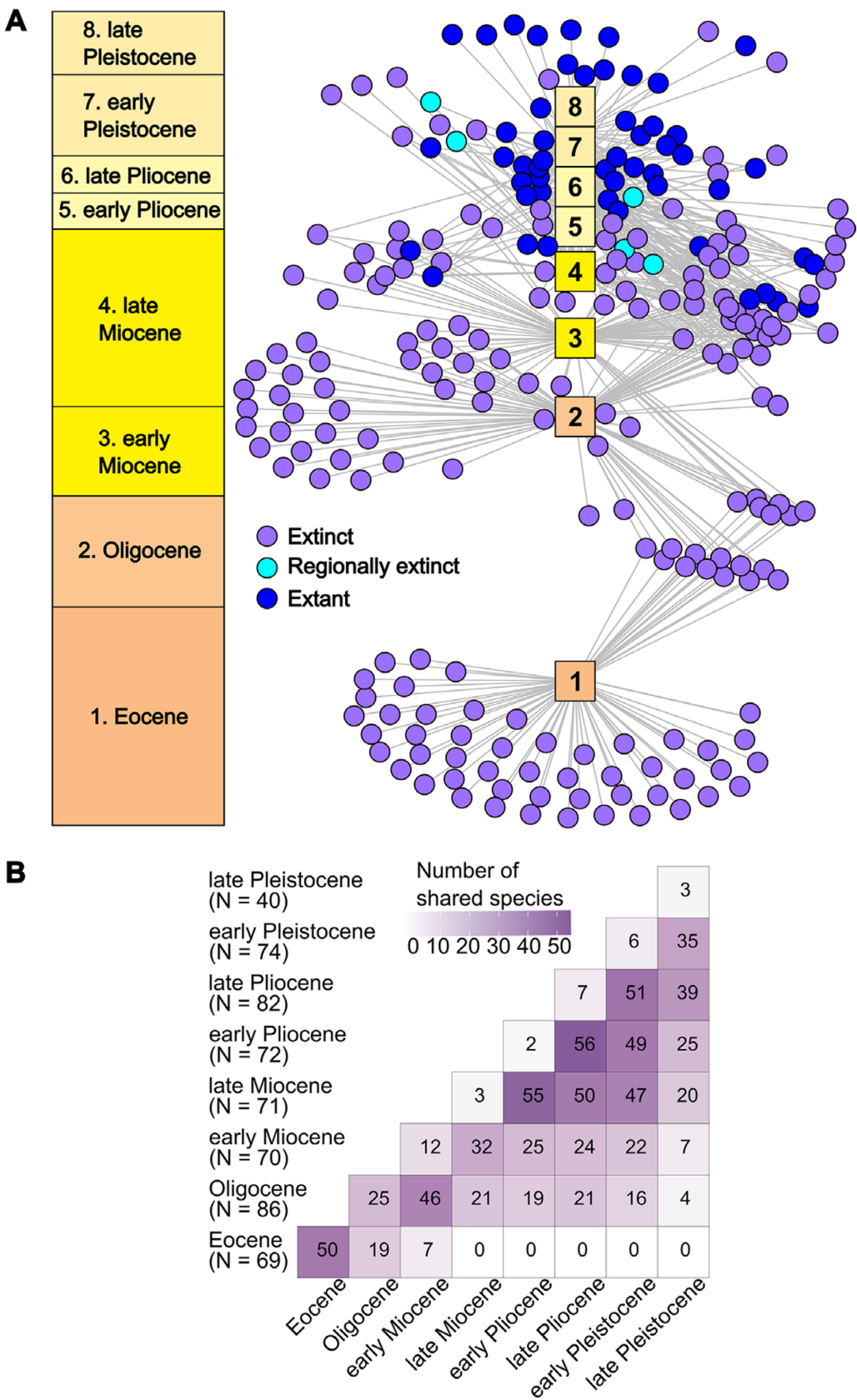


Figure 3. Coral species turnover throughout the Cenozoic Era (Eocene–Pleistocene) within the Caribbean. **A**, Bipartite affiliation network. Circles represent different coral species: purple, extinct species; light blue, extant species now regionally extinct in the Caribbean; dark blue, extant species. **B**, Heat map showing species overlap between epochs/sub-epochs; *N* indicates the overall number of species within each epoch/sub-epoch.

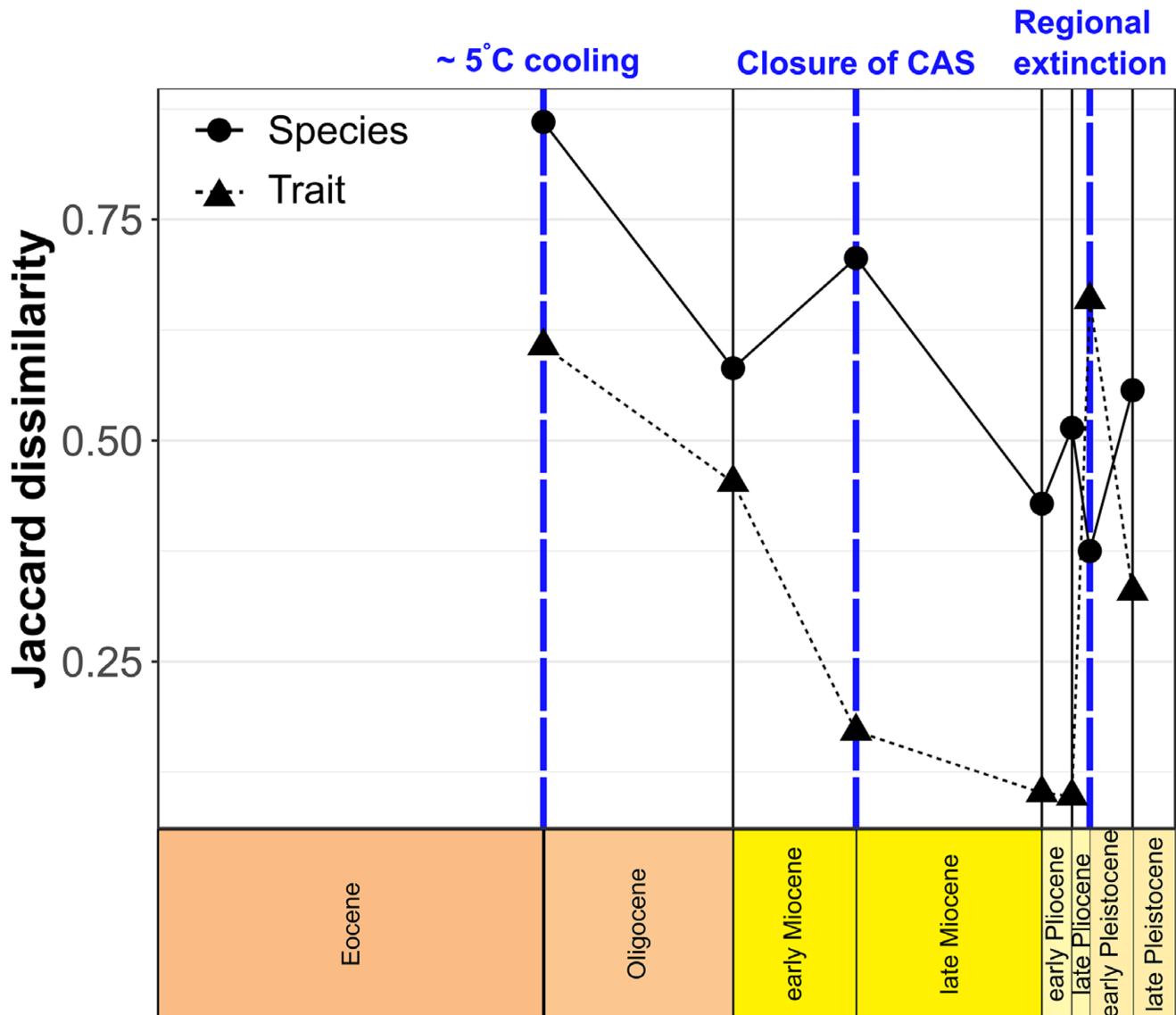


Figure 4. Jaccard dissimilarity scores between neighboring epochs/sub-epochs. Solid lines and circles indicate species dissimilarity and dotted lines with triangles indicate trait dissimilarity. Blue vertical dashed lines represent major environmental or biogeographic changes, with the first indicating the ~5°C Eocene-Oligocene cooling event, the second the closure of the central American Seaway (CAS) and the third the Caribbean regional coral extinction seen following the closure of the Panama isthmus.

network (Fig. 5A,B). In the Eocene network, the only two traits associated with a competitive life-history strategy are *branching colony growth form* and *extracalicular budding* (Fig. 5A), whereas in the Oligocene, this module increases in size and often contains three to four of the competitive traits: *small diameter corallites*, *branching colony growth form*, *corallite integration 3*, and *extracalicular budding* (Fig. 5B, Supplementary Fig. 1). The node degree of colony growth form branching remains stable between the Eocene and the Oligocene, whereas the node degree of *extracalicular budding* is reduced (Table 2). Traits related to a stress-tolerant life-history strategy in corals vary between the Eocene and Oligocene, with some co-occurring within the same module (i.e., *colony growth form massive*, *intracalicular budding*, and *very large diameter corallites* in the Eocene (Fig. 5A), and other traits being outside of the main stress-tolerant module. The co-occurrence of stress-tolerant traits across the Oligocene's randomly sampled 10 networks varies greatly (Supplementary Fig. 1). Across the Eocene–Oligocene epochs,

three stress-tolerant traits increase in node degree (*corallite integration 0* (*solitary*), *large diameter corallites* and *very large diameter corallites*) and two decrease in node degree (*intracalicular budding* and *colony growth form massive*) (Table 2).

The change in trait communities between the Eocene and Oligocene is illustrated by the PCA results (Fig. 5C). The first two PCA axes accounted for 41.62% of variation (PC 1, 23.77%; PC 2, 17.85%) (Fig. 5C). There was a decrease in trait community hull area from the Eocene (35.15) to the Oligocene (16.63) (Fig. 5C). The first two PCoA axes account for 58.72% of variation (PCoA 1, 37.45%; PCoA 2, 21.27%) (Fig. 5D). There was a decrease in functional richness between the two epochs, with the Eocene occupying 99% and the Oligocene occupying 79% of the overall functional richness (Fig. 5C). The decrease in functional richness was driven by a decrease in *corallite integration traits 1* (*dendroid*), 6 (*meandroid*), and 7 (*flabello-meandroid*) and *intracalicular budding* in the Oligocene (Fig. 5C).

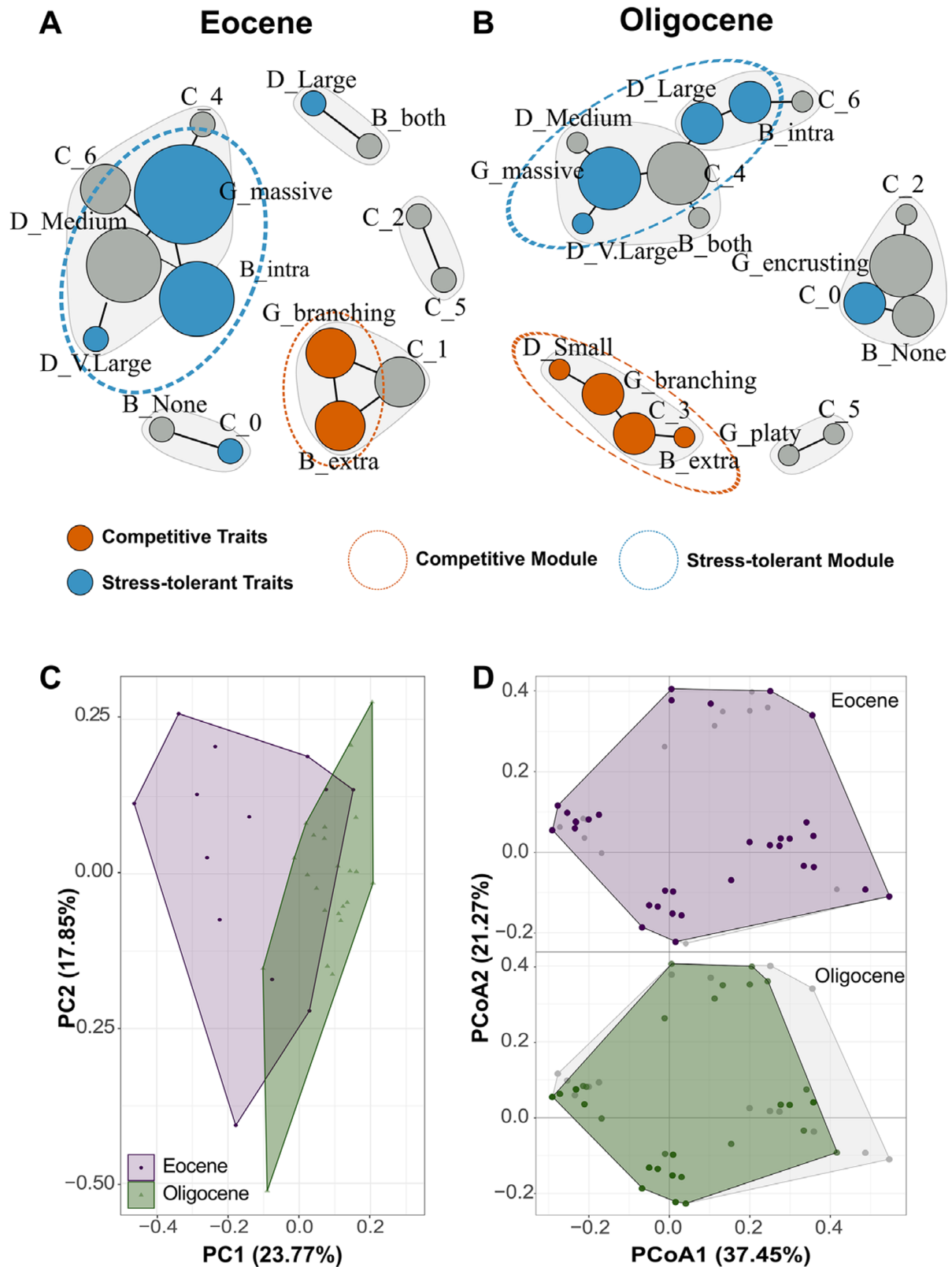


Figure 5. Trait turnover of Caribbean corals across the Eocene–Oligocene cooling period. Eocene (**A**) and Oligocene (**B**) trait–trait relationship networks based on the community weighted mean (CWM) trait values. Circles indicate network nodes (traits), node size relates to node degree, edges indicate significant co-occurrences, and gray shading indicates network modules. Orange nodes indicate traits associated with a competitive life-history strategy in corals, and blue nodes indicate traits related to a stress-tolerant life-history strategy in corals. For the Oligocene, the network with metrics closest to the mean is shown out of a possible 10 networks, created from randomly sampling 13 collections from each epoch 10 times. **C**, Principal component analysis (PCA) of CWMs for Caribbean coral trait communities for each epoch (Eocene and Oligocene). G = colony growth form, C = corallite integration, D = maximum corallite diameter, B = budding type. **D**, Gower distance–based principal coordinate analysis (PCoA) for Eocene and Oligocene. Gray convex hull represents overall trait space, with gray points indicating species in both the Eocene and Oligocene epochs. Purple convex hull and purple points indicate species from the Eocene. Green convex hull and green points indicate species from the Oligocene.

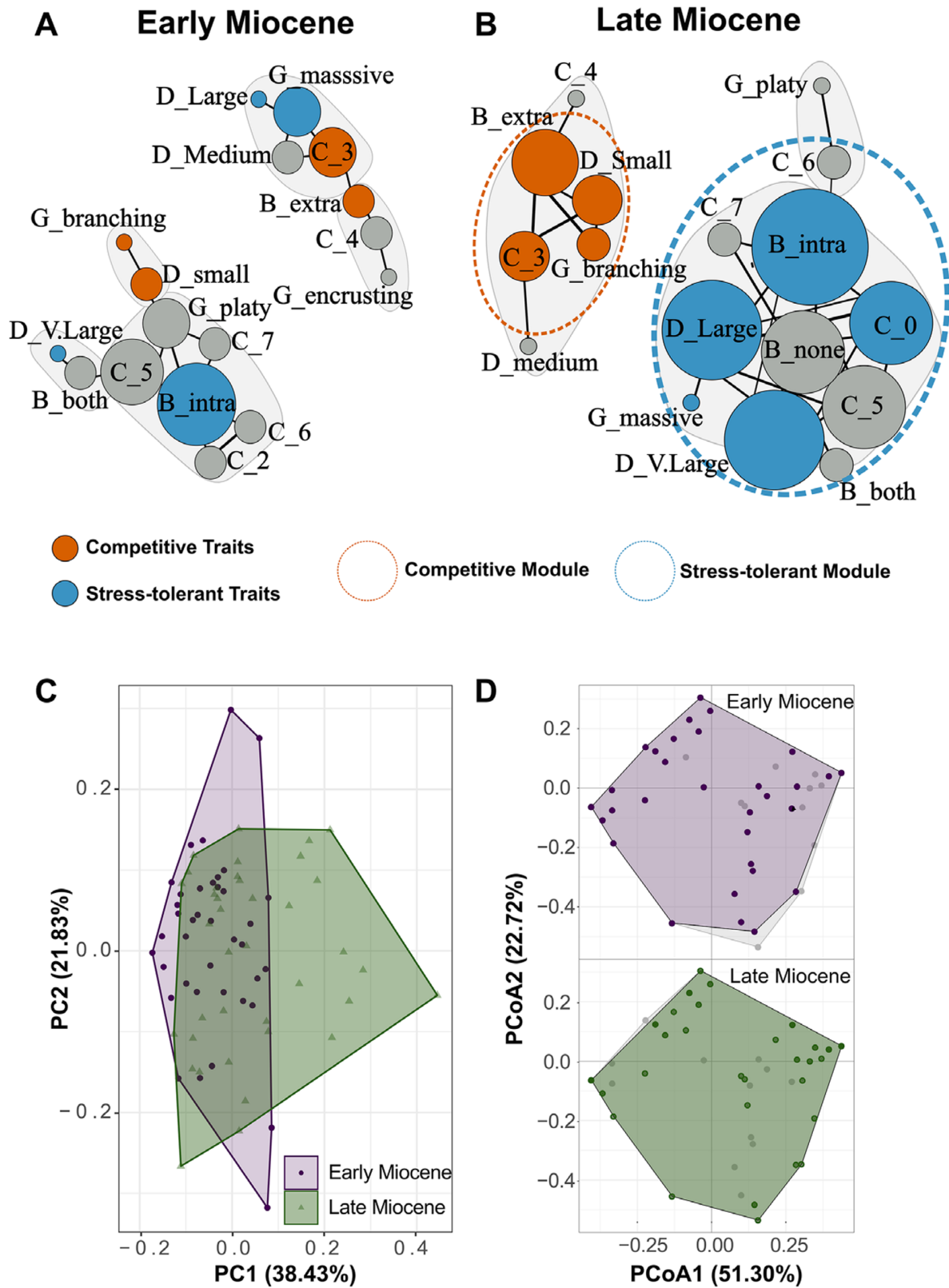


Figure 6. Trait turnover of Caribbean corals across the early Miocene–late Miocene interval of increased biogeographic isolation. Early Miocene (**A**) and late Miocene (**B**) trait–trait relationship networks based on the community weighted mean (CWM) trait values. Circles indicate network nodes (traits), node size relates to node degree, edges indicate significant co-occurrences, and gray shading indicates network modules. Orange nodes indicate traits associated with a competitive life-history strategy in corals, and blue nodes indicate traits related to a stress-tolerant life-history strategy in corals. For the early Miocene, we show the network with metrics closest to the mean out across 10 subsampled networks. **C**, Principal component analysis (PCA) of CWMs for Caribbean coral trait communities for each epoch (early Miocene, late Miocene). G = colony growth form, C = corallite integration, D = maximum corallite diameter, B = budding type. **D**, Gower distance–based principal coordinate analysis (PCoA) for the early and late Miocene. Gray convex hull represents overall trait space, with gray points indicating species in both sub-epochs. Purple convex hull and purple points indicate species from the early Miocene. Green convex hull and green points indicate species from the late Miocene.

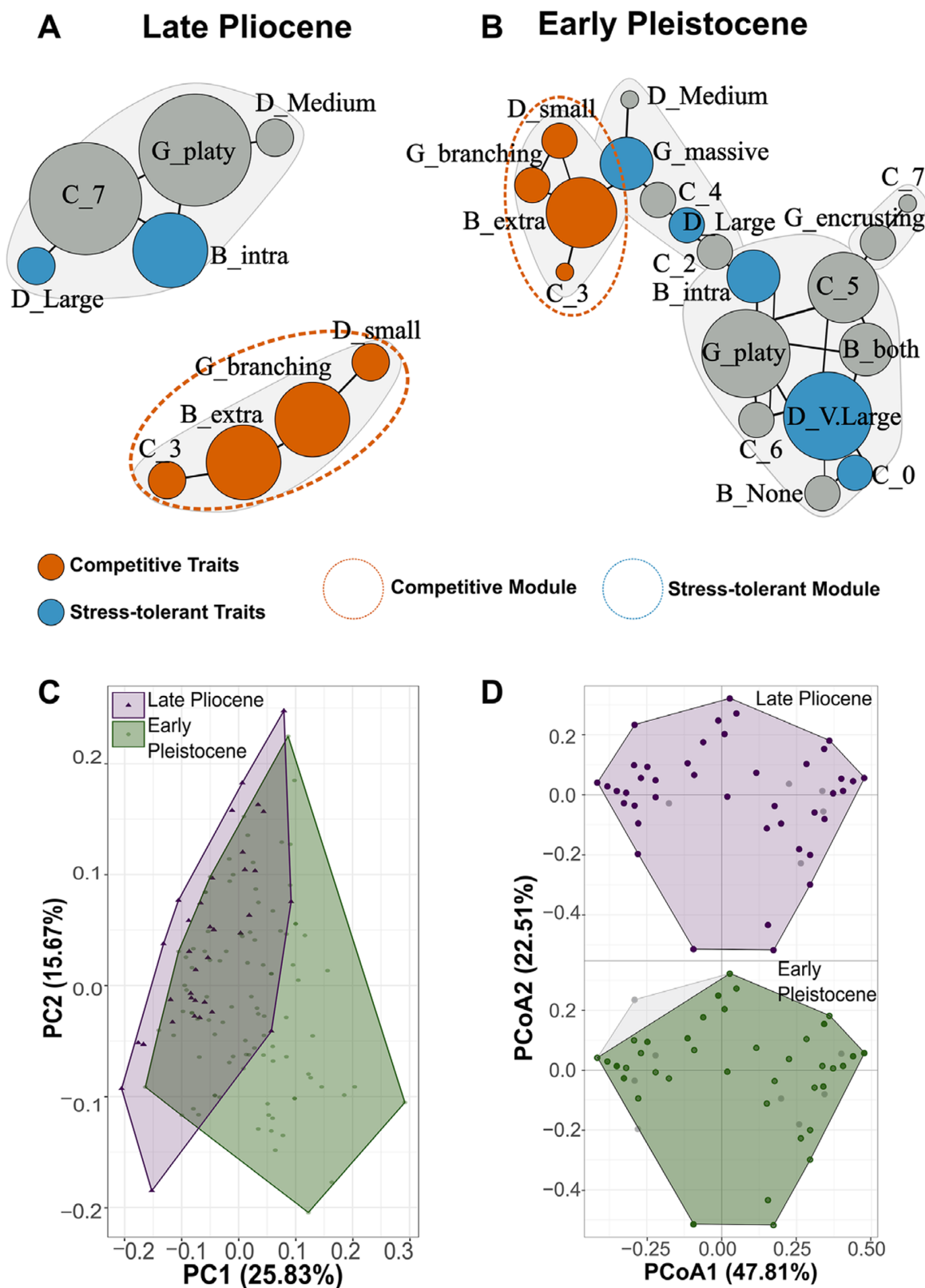


Figure 7. Trait turnover of Caribbean corals across the late Pliocene–early Pleistocene regional extinction event. Late Pliocene (**A**) and early Pleistocene (**B**) trait–trait relationship networks based on the community weighted mean (CWM) trait values. Circles indicate network nodes (traits), node size relates to node degree, edges indicate significant co-occurrences, and gray shading indicates network modules. Orange nodes indicate traits associated with a competitive life-history strategy in corals, and blue nodes indicate traits related to a stress-tolerant life-history strategy in corals. For the early Pleistocene, the network with metrics closest to the mean is shown out of a possible 10 networks, created from randomly sampling 36 collections from each sub-epoch 10 times. G = colony growth form, C = corallite integration, D = maximum corallite diameter, B = budding type. **C**, Principal component analysis (PCA) of CWMs for Caribbean coral trait communities for each epoch (late Pliocene, early Pleistocene). **D**, Gower distance–based principal coordinate analysis (PCoA) for the late Pliocene and early Pleistocene. Gray convex hull represents overall trait space, with gray points indicating species in both sub-epochs. Purple convex hull and purple points indicate species from the late Pliocene. Green convex hull and green points indicate species from the early Pleistocene.

Table 2. Node degree for Caribbean coral traits following trait–trait relationship network construction. Mean node degree, standard deviation (SD), and number of times trait appears (*N*) for Oligocene, early Miocene, and early Pleistocene networks where we created 10 randomly sampled networks to account for uneven collection numbers across neighboring epochs. G = colony growth form, C = corallite integration, D = maximum corallite diameter, B = budding type

		Eocene	Oligocene			Early Miocene			Late Miocene		Late Pliocene	Early Pleistocene		
Life-history strategy	Trait	Node degree	Mean node degree	SD	N	Mean node degree	SD	N	Node degree	Node degree	Mean node degree	SD	N	
Competitive	B_Extra	0.14	0.09	0.04	7	0.10	0.03	10	0.25	0.25	0.17	0.05	10	
	C_3	NA	0.13	0.06	6	0.19	0.00	10	0.19	0.125	0.10	0.04	10	
	D_Small	NA	0.10	0.06	10	0.13	0.02	10	0.19	0.125	0.15	0.03	10	
	G_Branching	0.14	0.14	0.05	10	0.06	0.00	10	0.13	0.25	0.15	0.06	9	
Stress tolerant	B_Intra	0.21	0.14	0.09	10	0.30	0.03	10	0.44	0.25	0.16	0.07	10	
	C_0	0.07	0.18	0.08	9	NA	NA	10	0.31	NA	0.11	0.04	9	
	D_Large	0.07	0.12	0.07	7	0.06	0.00	10	0.34	0.125	0.14	0.05	10	
	D_V.Large	0.07	0.12	0.07	4	0.06	0.00	10	0.38	NA	0.26	0.07	10	
	G_Massive	0.29	0.16	0.06	9	0.19	0.00	10	0.06	NA	0.14	0.05	10	
Other	B_None	0.07	0.16	0.03	8	NA	NA	10	0.31	NA	0.11	0.04	9	
	B_Both	0.07	0.12	0.06	7	0.15	0.03	10	0.13	NA	0.19	0.04	10	
	C_1	0.14	NA	NA	NA	NA	NA	10	NA	NA	0.12	NA	1	
	C_2	0.07	0.18	0.13	5	0.13	0.03	10	NA	NA	0.14	0.03	10	
	C_4	0.07	0.13	0.05	9	0.10	0.03	10	0.06	NA	0.08	0.03	10	
	C_5	0.07	0.12	0.06	10	0.26	0.02	10	0.31	NA	0.22	0.04	10	
	C_6	0.14	0.12	0.08	10	0.11	0.03	10	0.13	NA	0.13	0.02	10	
	C_7	NA	NA	NA	NA	0.13	0.00	10	0.13	0.375	0.19	0.10	6	
	D_Medium	0.21	0.12	0.05	10	0.13	0.00	10	0.06	0.13	0.08	0.04	9	
	G_Encrusting	NA	0.18	0.04	9	0.06	0.00	10	NA	NA	0.08	0.03	5	
	G_Platy	NA	0.15	0.10	10	0.22	0.03	10	0.06	0.38	0.24	0.04	10	

Trait Co-occurrences across the early Miocene–late Miocene Period of Increased Biogeographic Isolation

Edge density was reduced in the co-occurrence networks (early Miocene = 0.14 [mean $\pm$ 0.004], late Miocene = 0.21), in comparison to the weighted edge networks (early Miocene = 0.94 [mean $\pm$ 0.005], late Miocene = 0.87). Three traits are missing from the early Miocene community (*corallite integration 0 (solitary)*, *1 (dendroid)*, and *budding type none*), and all the traits present in the community are present in the trait–trait relationship networks (Supplementary Fig. 4a,c). All the traits present in the early Miocene communities appear in all 10 of the random samples (Table 2). The late Miocene community is missing two traits (*corallite integration 1 (dendroid)* and *colony growth form encrusting*), and *corallite integration 2 (phaceloid)* is present in the community but missing from the trait–trait relationship networks (Supplementary Fig. 4b,d).

Turnover in trait–trait relationships is seen between the early and late Miocene (Fig. 6A,B). In the early Miocene trait–trait relationship networks, there is no clear grouping of competitive or stress-tolerant traits (Fig. 6A). By the late Miocene, distinct modules have formed, with competitive traits clustered into an abundant group, while dominant stress-tolerant traits formed a second well-defined module but were intermixed with other traits (Fig. 6B). Three of the competitive traits (*budding type extra*, *small corallite diameter*, and *colony growth form branching*) increased in node degree from the early to the late Miocene, and *corallite*

integration 3 (plocoid) stayed the same (Table 2). Three of the stress-tolerant traits (*intracalicular budding*, *large*, and *very large corallite diameter*) increased in node degree from the early to the late Miocene, whereas *colony growth form massive* was reduced in node degree and *corallite integration 0 (solitary)* was only present in the late Miocene (Table 2).

The PCA results illustrate the change in trait communities, with the first two PCA axes accounting for 60.26% (PC 1, 38.43%; PC 2, 21.82%) of the variation (Fig. 5C). There was an increase in community hull area from the early (0.74) to the late Miocene (1.12), which was driven by increases in the traits for *large diameter corallites* and *intracalicular budding* (Fig. 5C). The first two PCoA axes accounted for 74.02% of the variation (PCoA 1, 51.30%; PCoA 2, 22.72%) (Fig. 5D). Functional richness increased slightly from the early (97%) to the late Miocene (99%) (Fig. 5D).

Trait Co-occurrences across the late Pliocene–early Pleistocene Regional Extinction Event

Edge density is reduced in the trait–trait co-occurrence networks for both the late Pliocene and early Pleistocene sub-epochs (late Pliocene = 0.22, early Pleistocene = 0.15 [mean $\pm$ 0.01 SD]), in comparison to the weighted edge networks (late Pliocene = 0.96, early Pleistocene = 0.98 [mean $\pm$ 0.02 SD]). *Corallite integration 0 (solitary)* and *budding type none* are missing from the late Pliocene community, and eight traits (*corallite integration 1 (dendroid)*, *2*

(*phaceloid/reptoid*), 4 (*cerioid*), 5 (*asteroid/thamnasteroid*), 6 (*meandroid*); very large diameter corallites, colony growth form encrusting, and budding type both) are present in the communities but missing from the trait–trait relationship networks (Supplementary Fig. 5a,c). All possible traits are present in the early Pleistocene communities (Supplementary Fig. 5b,d). Three traits do not appear in all 10 randomly sampled weighted networks: *budding type none* (9), *corallite integration 0 (solitary)* (9), and *corallite integration 1 (dendroid)*. All traits present in the early Pleistocene weighted edge network are present in the trait–trait relationship network (Supplementary Fig. 3B,D); however, many of these traits are not present in all of the 10 randomly sampled networks: *budding type none* (9), *colony from 0 (solitary)* (9), 1 (*dendroid*) (1), and 7 (*flabello-meandroid*); *corallite diameter medium* (9); and *colony growth form encrusting* (5) (Table 2).

Trait–trait relationships increase in complexity between the late Pliocene and early Pleistocene, indicated by the increase in traits, modules, and module connectedness seen in the early Pleistocene (Fig. 7A,B). All four traits related to a competitive life-history strategy are connected as a module in the early Pliocene and all but two of the early Pleistocene randomly sampled networks (Fig. 7A,B, Supplementary Fig. 2). The node degree of three of the competitive traits decreases from the late Pliocene to the early Pleistocene (*corallite integration 3 (plocoid)*, *extracalicular budding*, and *branching colony growth form*). In contrast, the *small diameter corallites* increased in node degree (Table 2). Only two stress-tolerant traits are present in the late Pliocene trait–trait relationship network (*large diameter corallites* and *intracalicular budding*) (Fig. 6A). In contrast, all five appear in the early Pleistocene networks (Fig. 7B, Supplementary Fig. 2). Co-occurrence of stress-tolerant traits across the early Pleistocene randomly sampled 10 networks varies, but stress-tolerant traits are present across all the modules, apart from the competitive module (Supplementary Fig. 2). Between the late Pliocene and early Pleistocene, *large diameter corallites* increased in node degree, while *intracalicular budding* decreases (Table 2), both traits are related to stress tolerance.

The change in trait communities between the late Pliocene and early Pleistocene is substantial (Fig. 7C). The first two PCA axes accounted for 41.50% of the variation (PC 1, 25.83%; PC 2, 15.67%) (Fig. 7C). There was an increase in trait community hull area from the late Pliocene (33.55) to the early Pleistocene (56.46) (Fig. 7C). The first two PCoA axes account for 70.32% of the variation in trait communities (PCoA 1, 47.81%; PCoA 2, 22.51%) (Fig. 7D). There was a decrease in functional richness between the two epochs, with the late Pliocene occupying 100% and the early Pleistocene occupying 94% of the overall functional richness (Fig. 7C). The decrease in functional richness was driven by a decrease in *corallite integration traits 1 (deandroid)* and *extracalicular budding* in the early Pleistocene (Fig. 7C).

Discussion

Our results support previous findings that periods of species turnover were evident throughout the Cenozoic (Eocene–Pleistocene), with the largest of these occurring between the Eocene and Oligocene, early and late Miocene, and late Pliocene to early Pleistocene, following environmental cooling and biogeographic changes (Budd 2000; Scheibner and Speijer 2008; Budd et al. 2011; Yasuhara et al. 2022; Figs. 3, 4). Turnover in traits occurred at two of the three boundaries: the Eocene/Oligocene transition and the late Pliocene/early Pleistocene transition (Fig. 4, Supplementary Fig. 1). Changes in trait abundances and trait–trait relationships were observed in all three transitions, indicating ecosystem-wide transformations of

coral reef composition and function (Figs. 5–7). The changing environmental conditions and increasing isolation of the Caribbean Sea selected for competitive (r-selected) life-history traits within reef-building corals, due to the variable environment, favoring morphological characteristics associated with faster growth and photosymbiosis over heterotrophy (Figs. 5–7). This view suggests a similar expansion of photosymbiosis in response to cooling climates, as observed in the Triassic (Kiessling 2010), potentially making their modern descendants less resilient to current increases in SST (Purvis et al. 2000; Roff 2021).

Species and Trait Turnover across the Eocene to Oligocene Cooling Period

All metrics indicate the highest turnover in species occurred between the Eocene and the Oligocene (Fig. 3). The high SST, sea level, and primary productivity in the Eocene favored algal reefs (Scheibner and Speijer 2008; Mutti et al. 2011; Takayanagi et al. 2012), which would have outcompeted corals for space (Sandin and McNamara 2012). As described by Yasuhara et al. (2022), the move toward cooler conditions in the Oligocene caused a decrease in the diversity of larger benthic foraminifera but allowed for coral reef species diversification (Fig. 3). It is, however, important to consider that the Eocene occurrence records contained fewer collections than the Oligocene (PBDB, <https://www.paleobiodb.org>, accessed 1 October 2022), potentially influencing the lower number of species we reported in the Eocene. However, most corals currently live less than 2°C below their thermal limits, with temperatures between 27°C and 37°C leading to coral bleaching and mortality (Anton et al. 2020). Given that SSTs were ~33°C during the Eocene (Cramer et al. 2011), fewer species were likely present in the Caribbean during the Eocene. High species dissimilarity (Fig. 4, Supplementary Fig. 1) indicates that the large architectural reefs of the Oligocene were not solely the result of increased growth rates of all existing Caribbean species (Johnson et al. 2008, 2009), but a turnover in species from those adapted to extreme heat to cooler-adapted species.

Trait turnover occurs alongside species turnover during the Eocene–Oligocene cooling period (Fig. 5). Traits related to stress tolerance appear more important in the Eocene (Fig. 5A), with stress-tolerant coral species being better equipped to withstand the heat of the Eocene and survive in deeper water (Lesser et al. 2021). One way stress-tolerant coral achieve this is by having larger diameter corallites, which are better adapted for heterotrophy, making them less reliant on their symbionts, which would be vulnerable to such high temperatures (Crabbe and Smith 2006; Conti-Jerpe et al. 2020; Dimitrijević et al. 2024). As temperatures decreased during the Oligocene, corals benefited from adopting a competitive life-history strategy, facilitating the formation of large architectural reefs in shallow waters (Johnson et al. 2008, 2009; Yasuhara et al. 2022). Competitive corals thrive in such shallow, high-light conditions (Baird and Hughes 2000; Darling et al. 2012). Although many competitive traits have been linked to increased extinction risk (Raja et al. 2021) in a warming world, the opposite is likely true in the Eocene–Oligocene cooling period.

The decrease in both trait community hull space and trait space between the Eocene and Oligocene epochs indicates that communities became more cosmopolitan in the Oligocene (Fig. 7), a parallel with contemporary homogenization of coral communities due to climate change (Stuart-Smith et al. 2021). Increases in both competitive and stress-tolerant traits are correlated with the change in community hull space between the Eocene and Oligocene, along

with reductions in encrusting solitary corals (Fig. 7A), indicating a general increase in functional diversity across the Eocene/Oligocene transition.

Species and Trait Turnover across the early Miocene–late Miocene Period of Increased Biogeographic Isolation

Species turnover is also apparent between the early and late Miocene (Fig. 3), which coincides with decreases in carbon dioxide (CO₂) and SST and a jump in community dissimilarity (Fig. 1) (Edinger and Risk 1994; Miller et al. 2020; Roff 2021). Tectonic changes throughout the Miocene leading to the closure of the Central American Seaway (CAS), which previously separated North and South America, shifted currents and increased upwelling in the Caribbean, restricting corals to marginal environments and reducing reef-building (Roff 2021; Yasuhara et al. 2022). Caribbean corals also became increasingly isolated during the Miocene, due to the closure of the Tethys Sea (now the Mediterranean) (Yasuhara et al. 2022), with species turnover at this time leading to the evolution of many modern Caribbean coral ancestors (Johnson and Kirby 2006).

Our results indicate that rather than the expected simplification of coral communities, the biogeographic changes of the Miocene became the force that drove the separation of the two life-history strategies: competitive and stress tolerant (Fig. 6A,B). Although there was minimal trait turnover across the early to late Miocene (Fig. 4), there was a change in trait abundances (Fig. 6D) during species turnover, leading to a change in trait–trait relationships. A lack of data availability for the middle Miocene means that part of the story may be missing, where trait turnover potentially may have occurred between the middle and late Miocene.

Species and Trait Turnover across the late Pliocene–early Pleistocene Regional Extinction Event

The coral communities during final four sub-epochs (early Pliocene, late Pliocene, early Pleistocene, and late Pleistocene) of the Cenozoic appear to be very similar, as seen from their proximity to each other in the bipartite network (Fig. 3A). The decrease in temperature and sea level across the early Pliocene to the late Pleistocene (Fig. 1) potentially pushed some of the remaining coral species below their thermal optima, with the increased isolation also contributing to community simplification (O’Dea et al. 2016; Roff 2021). Many extant species first appeared during the Pliocene and Pleistocene epochs (Fig. 3A), caused by the increasing isolation of the Caribbean, restricting gene flow from other seas (Jackson et al. 2001). As expected, a reduction in species richness can also be seen across the Pliocene/Pleistocene boundary, due to the regional extinction event between the late Pliocene and early Pleistocene (O’Dea et al. 2007). Although there is little evidence of a global coral extinction event between the late Pliocene and early Pleistocene, there were other marine extinction events, such as the Pliocene megafauna extinctions (Pimiento et al. 2017) and possibly wider marine invertebrate extinctions (Pimiento et al. 2020). Therefore, a combination of environmental factors, increasing isolation, and tectonic activity appears to have shaped the Caribbean coral community into the communities with low species richness of today.

Our findings confirm that *Acropora* became the dominant coral genus during the late Pliocene–early Pleistocene regional extinction event, as evidenced by competitive modules in the trait–trait networks from both periods (Fig. 7A,B). This aligns with existing research (Klaus et al. 2012; Renema et al. 2016). Stress-tolerant

traits are still present within the late Pliocene and early Pleistocene networks, but they no longer form stress-tolerant modules in either epoch (Fig. 7A,B), indicating a shift toward versatile communities with mixed types of traits. The reduction in CO₂, SST, and sea level throughout the Cenozoic, along with the closing of the Isthmus of Panama and isolation of the Caribbean Sea, appears to have selected for traits that are less resilient to environmental change in comparison to other reefs (e.g., Indo-Pacific reefs) (Roff 2021). The *Acropora* species that are still extant in the Caribbean today (*A. palmata* and *A. cervicornis*) are large and branching but also very long-lived (Johnson et al. 1995). Both *Acropora* species rely on asexual reproduction, which although useful, as fragmentation allows for quick repopulation of a reef following storm events, has contributed to lower genetic diversity on Caribbean reefs (Baums et al. 2006, 2013; Roff 2021). This low genetic diversity may have made them more susceptible to disease and other stressors (Baums et al. 2006; Brown et al. 2022). The majority of *Acropora* species suffered losses during the 1970s and 1980s, leaving Caribbean reefs dominated by “weedy” brooding species whose low reproductive outputs make repopulating the reef difficult (McWilliam et al. 2018). By contrast, in the Indo-Pacific, functional redundancy of *Acropora* species is much higher (Roff and Mumby 2012), and Indo-Pacific reefs appear to have better recovery rates following bleaching than their Caribbean counterparts (Roff 2021). Similarly, we posit that species with these traits will struggle to survive or recover from the current climate crisis (Renema et al. 2016; Enríquez et al. 2017).

The dominant *Acropora* species in the Pleistocene, which were the ancestors of many modern Caribbean corals, had small corallite diameters and a moderately high level of corallite integration, associated with a reliance on photosynthetic hypertrophy (Crabbe and Smith 2006; Stanley et al. 2018; Dimitrijević et al. 2024). These species have a low heat tolerance because, during periods of extreme heat, the concentration of endosymbiotic algae within a coral is reduced, potentially leading to the coral’s mortality if it cannot switch to heterotrophy for carbon assimilation (Hughes and Grotzli, 2013). They were also branching and, therefore, fast growing (Renema et al. 2016; Roff 2021), which allowed them to survive the Pleistocene by outcompeting for space on the reef, but may now be detrimental, as fast growth requires a lot of energy, making them less able to respond to stress (Pinzón et al. 2014).

The increase in community dissimilarity and trait diversity (Fig. 7C,D) across the late Pliocene and early Pleistocene may help to explain why we observe trait turnover, but not species turnover (Fig. 4, Supplementary Fig. 1), as this indicates species abundances changed rather than the loss or gain of species occurring (Fig. 7C, D). However, the trait–trait networks for the early Pleistocene show trait modules to be more interconnected, indicating greater similarity across communities. This is supported by the known reduction in species richness following the late Pliocene–early Pleistocene extinction event (Budd et al. 2011). Four often co-occurring traits drive the increase in community dissimilarity, suggesting an increase in corals that are platy growing, form colonies with asteroid or thamnasterioid integration, reproduce asexually via both budding types, and have large-diameter corallites (Fig. 7B,C). The four traits are only found together in *Mycetophyllia* species. Two species of *Mycetophyllia* were recorded in the late Pliocene occurrence records and six in the early Pleistocene, which potentially explains the increase in these traits driving the change in community dissimilarity. Another factor to consider is the lack of species abundance data available in the fossil record, as our analysis uses coral species presence/absence and *Mycetophyllia*

species are currently rare (Moulding and Ladd 2022). The lack of abundance information makes these traits (platy colony growth form, budding both, corallite integration 5 (asteroid/thamnasteroid)) appear more important in the communities than they are.

Implications for Coral Paleobiology

Our results indicate very little change in species richness across the Cenozoic, with some increase toward the end of the Cenozoic, which supports previous findings (Budd 2000). Budd (2000) found increases in species diversity to coincide with species turnover across the middle–late Eocene, late Oligocene–early Miocene, and Pliocene–Pleistocene. As we did not split up the Eocene, we were not able to detect turnover between the middle and late Eocene. We also found turnover between the Oligocene and early Miocene (Fig. 4, Supplementary Fig. 1); this period of turnover is well documented (Edinger 1991; Edinger and Risk 1994; Budd 2000; Johnson et al. 2008, 2009; Budd et al. 2011; Wallace and Bosellini 2015). However, we detected higher species dissimilarity between the early and late Miocene. The drop in dissimilarity following the Miocene supports the theory that biogeographic isolation was a major driver of coral diversity reduction due to a loss of gene flow from the Mediterranean (Rosen 1984; Budd 2000; O’Dea et al. 2017; Roff 2021; Steinhorsdottir et al. 2021). Our results also support high turnover across the Pliocene/Pleistocene boundary, where the regional extinction event reduced coral abundance and diversity (Johnson et al. 1995; Budd 2000). Although the species turnover across the Pliocene/Pleistocene boundary was not as high as across the Eocene/Oligocene boundary, trait turnover was higher, supporting theories that regional extinction events are not random but trait-based (van Woesik et al. 2012).

Implications for Modern Caribbean Corals

We have shown that as biogeographic isolation and global cooling increased throughout the Cenozoic, selection pressure favored a competitive life-history strategy, leading to the subsequent dominance of *Acropora* in the Pleistocene. This selection pressure may have caused the lack of resilience in modern Caribbean corals (Roff 2021). *Acropora* and other similar corals have high disease susceptibility and high sensitivity to bleaching, making their survival in a now-warming world challenging (McWilliams et al. 2005; Roff 2021). Modern Caribbean reefs lack the abundance and diversity of *Acropora* species due to heat stress and anthropogenic disturbance (Cramer et al. 2020, 2021).

With temperatures predicted to rise by up to 4°C by 2100 (IPCC 2023) and with coral communities shifting toward stress tolerance (Cramer et al. 2021), reefs of the future might resemble Eocene ones: stress tolerant, homogenized, and non-architectural. As the world cooled from “greenhouse” to “icehouse” over the last ~40 Myr, trait selection and evolution occurred slowly over time, with only a few intense transient phases (Norris et al. 2013). With modern environmental changes occurring rapidly and the corals’ ability to adapt to rising temperatures highly uncertain (Cramer et al. 2021), ongoing and future severe declines in Caribbean coral communities are almost inevitable.

Conclusion

In conclusion, our findings support existing knowledge on coral species diversity changes and provide new insight into trait turnover across the Cenozoic. We identified trait co-occurrence modules

integral to coral survival throughout the Cenozoic: competitive and stress tolerant. These two alternative life-history strategies are recognized as successful approaches for coral communities to persist in the face of predicted disturbance events (Kubicek et al. 2019). We have also shown that environmental and biogeographic changes often leads to the simplification and homogenization of coral communities, a phenomenon that continues in the Caribbean today in response to climate change and anthropogenic disturbance (Pawlik and Loh 2017; Estrada-Saldivar et al. 2019). The protracted fall in ocean temperatures and increasing geographic isolation throughout the Cenozoic appear to have selected for traits less resilient to heat stress (e.g., branching, small diameter, extracalicular budding), setting up the Caribbean’s coral reefs for failure in the current and predicted climate crisis, where species with competitive life histories will struggle to survive or recover.

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Literature Cited

- Anton, A., J. L. Randle, F. C. Garcia, S. Rossbach, J. I. Ellis, M. Weinzierl, and C. M. Duarte. 2020. Differential thermal tolerance between algae and corals may trigger the proliferation of algae in coral reefs. *Global Change Biology* 26: 4316–4327.
- Araújo, M. B., A. Rozenfeld, C. Rahbek, and P. A. Marquet. 2011. Using species co-occurrence networks to assess the impacts of climate change. *Ecography* 34:897–908.
- Araújo Távora, V. de. 2016. New records and redescrptions of Brazilian Scleractinia corals (Itamaracá, Maria Farinha and Pirabas Formations). *Paleontologia Mexicana* 5:71–86.
- Baird, A., and T. Hughes. 2000. Competitive dominance by tabular corals: an experimental analysis of recruitment and survival of understory assemblages. *Journal of Experimental Marine Biology and Ecology* 251:117–132.
- Baums, I. B., M. W. Miller, and M. E. Hellberg. 2006. Geographic variation in clonal structure in a reef-building Caribbean coral, *Acropora palmata*. *Ecological Monographs* 76:503–519.
- Baums, I., M. Devlin-Durante, N. Polato, D. Xu, S. Giri, N. Altman, D. Ruiz, J. Parkinson, and J. Boulay. 2013. Genotypic variation influences reproductive success and thermal stress tolerance in the reef building coral, *Acropora palmata*. *Coral Reefs* 32:703–717.
- Bello, F. de, Z. Botta-Dukat, J. Leps, and P. Fibich. 2021. Towards a more balanced combination of multiple traits when computing functional differences between species. *Methods in Ecology and Evolution* 12:443–448. Package ‘gawdis’.
- Bosellini, F. 1995. The scleractinian genus *Actinacis*. Systematic revision and stratigraphic record of the Tertiary species with special regard to Italian occurrences. *Rivista Italiana di Paleontologia e Stratigrafia* 101. <https://doi.org/10.13130/2039-4942/8575>.
- Brandl, S. J., D. B. Rasher, I. M. Côté, J. M. Casey, E. S. Darling, J. S. Lefcheck, and J. E. Duffy. 2019. Coral reef ecosystem functioning: eight core processes and the role of biodiversity. *Frontiers in Ecology and the Environment* 17:445–454.
- Brown, A. L., D.-E. Anastasiou, M. Schul, S. MacVittie, L. J. Spiers, J. L. Meyer, C. Manfrino, and T. K. Frazer. 2022. Mixtures of genotypes increase disease resistance in a coral nursery. *Scientific Reports* 12:19286.
- Budd, A. F. 2000. Diversity and extinction in the Cenozoic history of Caribbean reefs. *Coral Reefs* 19:25–35.
- Budd, A. F., and C. C. Wallace. 2008. First record of the Indo-Pacific reef coral genus *Isopora* in the Caribbean region: two new species from the Neogene of Curaçao, Netherlands Antilles. *Palaeontology* 51:1387–1401.

- Budd, A. F., T. A. Stemmann, and R. H. Stewart. 1992. Eocene Caribbean reef corals: a unique fauna from the Gatuncillo Formation of Panama. *Journal of Paleontology* 66:570–594.
- Budd, A. F., C. T. Foster, J. P. Dawson, and K. G. Johnson. 2001. The Neogene Marine Biota of Tropical America (“NMITA”) database: Accounting for biodiversity in paleontology. *Journal of Paleontology* 75:743–751.
- Budd, A. F., J. S. Klaus, and K. G. Johnson. 2011. Cenozoic diversification and extinction patterns in Caribbean reef corals: a review. *Paleontological Society Papers* 17:79–93.
- Budd, A. F., J. D. Woodell, D. Huang, and J. S. Klaus. 2019. Evolution of the Caribbean subfamily Mussinae (Anthozoa: Scleractinia: Faviidae): transitions between solitary and colonial forms. *Journal of Systematic Palaeontology* 17:1581–1616.
- Cairns, S., and J. Wells. 1987. Neogene paleontology in the northern Dominican Republic. 5. The suborders Caryophylliina and Dendrophylliina (Anthozoa: Scleractinia). *Bulletins of American Paleontology* 93:23–43.
- Cairns, S. D. 1995. New records of azooxanthellate stony corals (Cnidaria: Scleractinia and Stylasteridae) from the Neogene of Panama and Costa Rica. *Proceedings of the Biological Society of Washington* 108:533–550.
- Chamberlain, S., and B. Vanhoorne. 2023. worms: World Register of Marine Species (WoRMS) Client, R package version 0.4.3. <https://doi.org/10.32614/CRAN.package.worms>.
- Clay, C. G., J. D. Reimer, K. M. Cook, H. Yamagawa, E. Gravenor, L. H. Y. Theodora, and M. Beger. 2023. Temporal functional changes in coral and fish communities on subtropical coastal coral reefs. *Marine and Freshwater Research* 74:1081–1094.
- Clay, C. G., A. M. Dunhill, J. D. Reimer, and M. Beger. 2024a. Trait networks: assessing marine community resilience and extinction recovery. *iScience* 27.
- Clay, C. G., J. D. Reimer, B. Sommer, K. M. Cook, M. Mizuyama, M. Obuchi, I. Kawamura, H. Kise, and M. Beger. 2024b. Variation in functional composition of reef fishes along a tropical-to-temperate gradient. *Journal of Biogeography* 51:454–466.
- Cohen, K. M., S. M. Finnery, P. L. Gibbard, and J.-X. Fan. 2013. *The ICS International Chronostratigraphic Chart*. International Commission on Stratigraphy. www.episodes.org, accessed 5 May 2023.
- Connell, J. H. 1997. Disturbance and recovery of coral assemblages. *Coral Reefs* 16:S101–S113.
- Conti-Jerpe, I. E., P. D. Thompson, C. W. M. Wong, N. L. Oliveira, N. N. Duprey, M. A. Moynihan, and D. M. Baker. 2020. Trophic strategy and bleaching resistance in reef-building corals. *Science Advances* 6:eaa5443.
- Cook, K. M., H. Yamagawa, M. Beger, G. D. Masucci, S. Ross, H. Y. T. Lee, R. D. Stuart-Smith, and J. D. Reimer. 2022. A community and functional comparison of coral and reef fish assemblages between four decades of coastal urbanisation and thermal stress. *Ecology and Evolution* 12:e8736.
- Cornwell, W. K., D. W. Schwilk, and D. D. Ackerly. 2006. A trait-based test for habitat filtering: convex hull volume. *Ecology* 87:1465–1471.
- Crabbe, M. J. C., and D. J. Smith. 2006. Modelling variations in corallite morphology of *Galaxea fascicularis* coral colonies with depth and light on coastal fringing reefs in the Wakatobi Marine National Park (S.E. Sulawesi, Indonesia). *Computational Biology and Chemistry* 30:155–159.
- Cramer, B. S., K. G. Miller, P. J. Barrett, and J. D. Wright. 2011. Late Cretaceous–Neogene trends in deep ocean temperature and continental ice volume: reconciling records of benthic foraminiferal geochemistry ($\delta^{18}\text{O}$ and Mg/Ca) with sea level history. *Journal of Geophysical Research: Oceans* 116(C12).
- Cramer, K. L., J. B. Jackson, M. K. Donovan, B. J. Greenstein, C. A. Korpanty, G. M. Cook, and J. M. Pandolfi. 2020. Widespread loss of Caribbean acroporid corals was underway before coral bleaching and disease outbreaks. *Science Advances* 6:eaa9395.
- Cramer, K. L., M. K. Donovan, J. B. C. Jackson, B. J. Greenstein, C. A. Korpanty, G. M. Cook, and J. M. Pandolfi. 2021. The transformation of Caribbean coral communities since humans. *Ecology and Evolution* 11:10098–10118.
- Csardi, G., and T. Nepusz. 2006. The igraph software package for complex network research. *InterJournal, Complex Systems* 1695:1–9.
- Darling, E. S., L. Alvarez-Filip, T. A. Oliver, T. R. McClanahan, and I. M. Côté. 2012. Evaluating life-history strategies of reef corals from species traits. *Ecology Letters* 15:1378–1386.
- Dimitrijević, D., N. B. Raja, and W. Kiessling. 2024. Corallite sizes of reef corals: decoupling of evolutionary and ecological trends. *Paleobiology* 50:43–53.
- Donner, S. D., S. F. Heron, and W. J. Skirving. 2018. Future scenarios: a review of modelling efforts to predict the future of coral reefs in an era of climate change. Pp. 325–341 in M. J. H. van Oppen and J. M. Lough, eds. *Coral bleaching: patterns, processes, causes and consequences*. Springer International, Cham, Switzerland.
- Doszpot, N. E., M. J. McWilliam, M. S. Pratchett, A. S. Hoey, and W. F. Figueira. 2019. Plasticity in three-dimensional geometry of branching corals along a cross-shelf gradient. *Diversity* 11:44.
- Duncan, P. M. 1864. A description of, and remarks upon, some fossil corals from Sinde. *Annals and Magazine of Natural History* 13:295–307.
- Duncan, P. M. 1873. On the older Tertiary formations of the West-Indian Islands. *Quarterly Journal of the Geological Society* 29:548–565.
- Dunhill, A. M., W. J. Foster, J. Sciberras, and R. J. Twitchett. 2018. Impact of the Late Triassic mass extinction on functional diversity and composition of marine ecosystems. *Palaeontology* 61:133–148.
- Durham, J. W. 1946. *Eocene faunas from the Department of Bolivar, Colombia*, Vol. 16. Geological Society of America, New York.
- Echevarría, J., M. S. Hodges, S. E. Damborenea, G. D. Stanley Jr., and M. O. Manceñido. 2017. Recovery of scleractinian morphologic diversity during the Early Jurassic in Mendoza Province, Argentina. *Ameghiniana* 54:70–82.
- Edinger, E. N. 1991. *Mass extinction of Caribbean corals at the Oligocene–Miocene boundary: paleoecology, paleoceanography, paleobiogeography*. Master's thesis. McMaster University, Hamilton, Ont., Canada.
- Edinger, E. N., and M. J. Risk. 1994. Oligocene–Miocene extinction and geographic restriction of Caribbean corals: roles of turbidity, temperature, and nutrients. *Palaios* 9:576–598.
- [EOL] Encyclopedia of Life. 2022. Home page. <https://naturalhistory.si.edu/research/eol>.
- Enriquez, S., E. R. Méndez, O. Hoegh-Guldberg, and R. Iglesias-Prieto. 2017. Key functional role of the optical properties of coral skeletons in coral ecology and evolution. *Proceedings of the Royal Society B* 284:20161667.
- Estrada-Saldivar, N., E. Jordán-Dalhgren, R. E. Rodríguez-Martínez, C. Perry, and L. Alvarez-Filip. 2019. Functional consequences of the long-term decline of reef-building corals in the Caribbean: evidence of across-reef functional convergence. *Royal Society Open Science* 6:190298.
- Filkorn, H. F., J. Avendaño-Gil, M. A. Coutiño-José, and F. J. Vega-Vera. 2005. Corals from the Upper Cretaceous (Maastrichtian) Ocozocoautla Formation, Chiapas, Mexico. *Revista mexicana de ciencias geológicas* 22:115–128.
- Flores-Moreno, H., F. Fazayeli, A. Banerjee, A. Datta, J. Kattge, E. E. Butler, O. K. Atkin, et al. 2019. Robustness of trait connections across environmental gradients and growth forms. *Global Ecology and Biogeography* 28:1806–1826.
- Flórez, P., P. Zapata-Ramírez, and J. S. Klaus. 2019. Early Miocene shallow-water corals from La Guajira, Colombia: part I, Acroporidae–Montastraeidae. *Journal of Paleontology* 93:1–24.
- Floyd, M., M. Mizuyama, M. Obuchi, B. Sommer, M. G. R. Miller, I. Kawamura, H. Kise, J. D. Reimer, and M. Beger. 2020. Functional diversity of reef molluscs along a tropical-to-temperate gradient. *Coral Reefs* 39:1361–1376.
- Fonseca, C. R., and G. Ganade. 2001. Species functional redundancy, random extinctions and the stability of ecosystems. *Journal of Ecology* 89:118–125.
- Foster, W. J., and R. J. Twitchett. 2014. Functional diversity of marine ecosystems after the Late Permian mass extinction event. *Nature Geoscience* 7:233–238.
- Frost, S. H., and Ralph L. Langenheim. 1974. *Cenozoic reef biofacies: Tertiary larger foraminiferas and scleractinian corals from Chiapas, Mexico*. Northern Illinois University Press, DeKalb, Ill.
- Gao, D., S. Wang, F. Wei, X. Wu, S. Zhou, L. Wang, Z. Li, P. Chen, and B. Fu. 2022. The vulnerability of ecosystem structure in the semi-arid area revealed by the functional trait networks. *Ecological Indicators* 139:108894.
- Gilmour, J. P. 2004. Asexual budding in Fungiid corals. *Coral Reefs* 23:595.
- Gilson, A. R., D. A. Smale, and N. O'Connor. 2021. Ocean warming and species range shifts affect rates of ecosystem functioning by altering consumer–resource interactions. *Ecology* 102:e03341.
- Gower, J. C. 1983. Distance matrices and their Euclidean approximation. Pp. 1–19 in E. Diday, ed. *Third International Conference on Data Analysis and Informatics*. INRA, Paris, Versailles.

- Harnik, P. G., H. K. Lotze, S. C. Anderson, Z. V. Finkel, S. Finnegan, D. R. Lindberg, L. H. Liow, R. Lockwood, C. R. McClain, and J. L. McGuire. 2012. Extinctions in ancient and modern seas. *Trends in Ecology and Evolution* 27:608–617.
- Harris, D. J. 2016. Inferring species interactions from co-occurrence data with Markov networks. *Ecology* 97:3308–3314.
- He, N., Y. Li, C. Liu, L. Xu, M. Li, J. Zhang, J. He, et al. 2020. Plant trait networks: improved resolution of the dimensionality of adaptation. *Trends in Ecology and Evolution* 35:908–918.
- Hodapp, D., I. T. Roca, D. Fiorentino, C. Garilao, K. Kaschner, K. Kesner-Reyes, B. Schneider, et al. 2023. Climate change disrupts core habitats of marine species. *Global Change Biology* 29:3304–3317.
- Hoeksema, B. W., and S. D. Cairns. 2025. World List of Scleractinia. Scleractinia incertae sedis. <https://www.marinespecies.org/scleractinia>, accessed 1 March 2025.
- Hooper, D. U., F. S. Chapin III, J. J. Ewel, A. Hector, P. Inchausti, S. Lavorel, J. H. Lawton, et al. 2005. Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. *Ecological Monographs* 75:3–35.
- Hughes, A. D., and A. G. Grottoli. 2013. Heterotrophic compensation: a possible mechanism for resilience of coral reefs to global warming or a sign of prolonged stress? *PLoS ONE* 8:e81172.
- Hughes, T. P., J. T. Kerry, M. Álvarez-Noriega, J. G. Álvarez-Romero, K. D. Anderson, A. H. Baird, R. C. Babcock, et al. 2017. Global warming and recurrent mass bleaching of corals. *Nature* 543:373–377.
- Hughes, T. P., J. T. Kerry, A. H. Baird, S. R. Connolly, A. Dietzel, C. M. Eakin, S. F. Heron, et al. 2018. Global warming transforms coral reef assemblages. *Nature* 556:492–496.
- [IPCC] Intergovernmental Panel on Climate Change. 2023. *Climate Change 2022—impacts, adaptation and vulnerability: Working Group II Contribution to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge University Press, Cambridge.
- Jackson, J., M. Donovan, K. Cramer, and V. Lam. 2014. *Status and trends of Caribbean coral reefs: 1970–2012*. Global Coral Reef Monitoring Network, Gland, Switzerland.
- Jackson, J. B., M. X. Kirby, W. H. Berger, K. A. Bjorndal, L. W. Botsford, B. J. Bourque, R. H. Bradbury, R. Cooke, J. Erlandson, and J. A. Estes. 2001. Historical overfishing and the recent collapse of coastal ecosystems. *Science* 293:629–637.
- Johnson, K. G., and M. X. Kirby. 2006. The Emperor Limestone rediscovered: early Miocene corals from the Culebra formation, Panama. *Journal of Paleontology* 80:283–293.
- Johnson, K. G., A. F. Budd, and T. A. Stemmann. 1995. Extinction selectivity and ecology of Neogene Caribbean reef corals. *Paleobiology* 21:52–73.
- Johnson, K. G., J. B. Jackson, and A. F. Budd. 2008. Caribbean reef development was independent of coral diversity over 28 million years. *Science* 319:1521–1523.
- Johnson, K. G., M. R. Sánchez-Villagra, and O. A. Aguilera. 2009. The Oligocene–Miocene transition on coral reefs in the Falcón Basin (NW Venezuela). *Palaios* 24:59–69.
- Kiessling, W. 2005. Habitat effects and sampling bias on Phanerozoic reef distribution. *Facies* 51:24–32.
- Kiessling, W. 2008. Sampling-standardized expansion and collapse of reef building in the Phanerozoic. *Fossil Record* 11:7–18.
- Kiessling, W. 2010. Reef expansion during the Triassic: spread of photosymbiosis balancing climatic cooling. *Palaeogeography, Palaeoclimatology, Palaeoecology* 290:11–19.
- Kitahara, M. V., H. Fukami, F. Benzoni, and D. Huang. 2016. The new systematics of Scleractinia: integrating molecular and morphological evidence. Pp. 41–59 in S. Goffredo and Z. Dubinsky, eds. *The Cnidaria, past, present and future*. Springer, Cham, Switzerland.
- Klaus, J. S., D. F. McNeill, A. F. Budd, and A. G. Coates. 2012. Neogene reef coral assemblages of the Bocas del Toro region, Panama: the rise of *Acropora palmata*. *Coral Reefs* 31:191–203.
- Kubicek, A., B. Breckling, O. Hoegh-Guldberg, and H. Reuter. 2019. Climate change drives trait-shifts in coral reef communities. *Scientific Reports* 9:3721.
- Laliberté, E., P. Legendre, B. Shipley, and M. E. Laliberté. 2014. FD: measuring functional diversity from multiple traits, and other tools for functional ecology, R package version 1.0–12.3. <https://cran.r-project.org/web/packages/FD/FD.pdf>.
- Lathuilière, B. 1996. Is morphology a good way to understand the evolution of corals? *Paleontological Society Papers* 1:81–105.
- Lesser, M. P., M. Slattery, and C. D. Mobley. 2021. Incident light and morphology determine coral productivity along a shallow to mesophotic depth gradient. *Ecology and Evolution* 11:13445–13454.
- Liu, H., Z. Liu, M. Zhang, J. Bascompte, F. He, and C. Chu. 2021. Geographic variation in the robustness of pollination networks is mediated by modularity. *Global Ecology and Biogeography* 30:1447–1460.
- Locke, J. M., E. Weil, and K. A. Coates. 2007. A newly documented species of *Madracis* (Scleractinia: Pocilloporidae) from the Caribbean. *Proceedings of the Biological Society of Washington* 120:214–226.
- Loreau, M., and A. Hector. 2001. Partitioning selection and complementarity in biodiversity experiments. *Nature* 412:72–76.
- Madin, J. S., K. D. Anderson, M. H. Andreasen, T. C. Bridge, S. D. Cairns, S. R. Connolly, E. S. Darling, M. Diaz, D. S. Falster, and E. C. Franklin. 2016. The Coral Trait Database, a curated database of trait information for coral species from the global oceans. *Scientific Data* 3:1–22.
- Magneville, C., N. Loiseau, C. Albouy, N. Casajus, T. Claverie, A. Escalas, F. Leprieur, E. Maire, D. Mouillot, and S. Villéger. 2022. mFD: an R package to compute and illustrate the multiple facets of functional diversity. *Ecography* 2022(1). <https://onlinelibrary.wiley.com/doi/full/10.1111/ecog.05904>.
- Matich, P., J. J. Kiszka, J. Mourier, S. Planes, and M. R. Heithaus. 2017. Species co-occurrence affects the trophic interactions of two juvenile reef shark species in tropical lagoon nurseries in Moorea (French Polynesia). *Marine Environmental Research* 127:84–91.
- McWilliam, M., M. O. Hoogenboom, A. H. Baird, C.-Y. Kuo, J. S. Madin, and T. P. Hughes. 2018. Biogeographical disparity in the functional diversity and redundancy of corals. *Proceedings of the National Academy of Sciences USA* 115:3084–3089.
- McWilliams, J. P., I. M. Côté, J. A. Gill, W. J. Sutherland, and A. R. Watkinson. 2005. Accelerating impacts of temperature-induced coral bleaching in the Caribbean. *Ecology* 86:2055–2060.
- Miller, K. G., J. V. Browning, W. J. Schmelz, R. E. Kopp, G. S. Mountain, and J. D. Wright. 2020. Cenozoic sea-level and cryospheric evolution from deep-sea geochemical and continental margin records. *Science Advances* 6:eaaaz1346.
- Moruea-Holme, N., B. Blonder, B. Sandel, B. J. McGill, R. K. Peet, J. E. Ott, C. Violle, B. J. Enquist, P. M. Jørgensen, and J. Svenning. 2016. A network approach for inferring species associations from co-occurrence data. *Ecography* 39:1139–1150.
- Mouillot, D., S. Villéger, V. Parravicini, M. Kulbicki, J. E. Arias-González, M. Bender, P. Chabanet, et al. 2014. Functional over-redundancy and high functional vulnerability in global fish faunas on tropical reefs. *Proceedings of the National Academy of Sciences USA* 111:13757.
- Moulding, A., and M. Ladd. 2022. Staghorn coral (*Acropora cervicornis*), elkhorn coral (*Acropora palmata*), lobed star coral (*Orbicella annularis*), mountainous star coral (*Orbicella faveolata*), boulder star coral (*Orbicella franksi*), rough cactus coral (*Mycetophyllia ferox*), and pillar coral (*Dendrogya cylindrus*) 5-year review. National Marine Fisheries Service, Southeast Regional Office, Saint Petersburg, Fla.
- Mutti, M., W. E. Piller, and C. Betzler. 2011. *Carbonate systems during the Oligocene–Miocene climatic transition*. Wiley, Hoboken, NJ.
- Norris, R. D., S. K. Turner, P. M. Hull, and A. Ridgwell. 2013. Marine ecosystem responses to Cenozoic global change. *Science* 341:492–498.
- O'Dea, A., J. B. C. Jackson, H. Fortunato, J. T. Smith, L. D'Croz, K. G. Johnson, and J. A. Todd. 2007. Environmental change preceded Caribbean extinction by 2 million years. *Proceedings of the National Academy of Sciences USA* 104:5501–5506.
- O'Dea, A., H. A. Lessios, A. G. Coates, R. I. Eytan, S. A. Restrepo-Moreno, A. L. Cione, L. S. Collins, A. De Queiroz, D. W. Farris, and R. D. Norris. 2016. Formation of the Isthmus of Panama. *Science Advances* 2:e1600883.
- O'Dea, A., E. M. Dillon, A. H. Altieri, and M. L. Lepore. 2017. Look to the past for an optimistic future. *Conservation Biology* 31:1221–1222.
- Oksanen, J., F. G. Blanchet, M. Friendly, R. Kindt, P. Legendre, D. McGlinn, P. R. Minchin, et al. 2020. Vegan: community ecology package, R package version 2.5–7. <https://cran.r-project.org/web/packages/vegan/index.html>.

- Padda, S. S., J. R. Glass, and Z. R. Stahlschmidt. 2021. When it's hot and dry: life-history strategy influences the effects of heat waves and water limitation. *Journal of Experimental Biology* 224:jeb236398.
- Pandolfi, J. M. 2007. A new, extinct Pleistocene reef coral from the *Montastraea* "annularis" species complex. *Journal of Paleontology* 81:472–482.
- Pandolfi, J. M. 2015. Incorporating uncertainty in predicting the future response of coral reefs to climate change. *Annual Review of Ecology, Evolution, and Systematics* 46:281–303.
- Pandolfi, J. M., and J. B. Jackson. 2007. Broad-scale patterns in Pleistocene coral reef communities from the Caribbean: implications for ecology and management. Pp. 201–236 in R. B. Aronson, ed. *Geological approaches to coral reef ecology*, Vol. 192. Springer, New York.
- Pandolfi, J. M., and W. Kiessling. 2014. Gaining insights from past reefs to inform understanding of coral reef response to global climate change. *Current Opinion in Environmental Sustainability* 7:52–58.
- Pawlík, J. R., and T.-L. Loh. 2017. Biogeographical homogeneity of Caribbean coral reef benthos. *Journal of Biogeography* 44:960–962.
- Payne, J. L., and M. E. Clapham. 2012. End-Permian mass extinction in the oceans: an ancient analog for the twenty-first century? *Annual Review of Earth and Planetary Sciences* 40:89–111.
- Pianka, E. R. 1970. On r- and K-selection. *American Naturalist* 104:592–597.
- Pimienta, C., J. N. Griffin, C. F. Clements, D. Silvestro, S. Varela, M. D. Uhen, and C. Jaramillo. 2017. The Pliocene marine megafauna extinction and its impact on functional diversity. *Nature Ecology and Evolution* 1:1100–1106.
- Pimienta, C., C. D. Bacon, D. Silvestro, A. Hendy, C. Jaramillo, A. Zizka, X. Meyer, and A. Antonelli. 2020. Selective extinction against redundant species buffers functional diversity. *Proceedings of the Royal Society B* 287:20201162.
- Pinzón, J. H., L. Dornberger, J. Beach-Letendre, E. Weil, and L. D. Mydlarz. 2014. The link between immunity and life history traits in scleractinian corals. *PeerJ* 2:e628.
- Purvis, A., J. L. Gittleman, G. Cowlishaw, and G. M. Mace. 2000. Predicting extinction risk in declining species. *Proceedings of the Royal Society B* 267: 1947–1952.
- Rae, J. W. B., Y. G. Zhang, X. Liu, G. L. Foster, H. M. Stoll, and R. D. M. Whiteford. 2021. Atmospheric CO₂ over the past 66 million years from marine archives. *Annual Review of Earth and Planetary Sciences* 49:609–641.
- Raja, N. B., A. Lauchstedt, J. M. Pandolfi, S. W. Kim, A. F. Budd, and W. Kiessling. 2021. Morphological traits of reef corals predict extinction risk but not conservation status. *Global Ecology and Biogeography* 30: 1597–1608.
- Raja, N. B., D. Dimitrijević, M. C. Krause, and W. Kiessling. 2022. Ancient Reef Traits, a database of trait information for reef-building organisms over the Phanerozoic. *Scientific Data* 9:425.
- Ramírez-Portilla, C., A. H. Baird, P. F. Cowman, A. M. Quattrini, S. Harii, F. Sinniger, and J.-F. Flot. 2022. Solving the coral species delimitation conundrum. *Systematic Biology* 71:461–475.
- Renema, W., J. M. Pandolfi, W. Kiessling, F. R. Bosellini, J. S. Klaus, C. Korpany, B. R. Rosen, N. Santodomingo, C. C. Wallace, J. M. Webster, and K. G. Johnson. 2016. Are coral reefs victims of their own past success? *Science Advances* 2:e1500850.
- Roff, G. 2021. Evolutionary history drives biogeographic patterns of coral reef resilience. *BioScience* 71:26–39.
- Roff, G., and P. J. Mumby. 2012. Global disparity in the resilience of coral reefs. *Trends in Ecology and Evolution* 27:404–413.
- Rosen, B. 1984. Reef coral biogeography and climate through the late Cainozoic: just islands in the sun or a critical pattern of islands? *Special issue*, Geological Journal, no. 11, 201–262.
- Sakai, K. 1998. Effect of colony size, polyp size, and budding mode on egg production in a colonial coral. *Biological Bulletin* 195:319–325.
- Sandin, S. A., and D. E. McNamara. 2012. Spatial dynamics of benthic competition on coral reefs. *Oecologia* 168:1079–1090.
- Scheibner, C., and R. P. Speijer. 2008. Late Paleocene–early Eocene Tethyan carbonate platform evolution — A response to long- and short-term paleoclimatic change. *Earth-Science Reviews* 90:71–102.
- Sleeman, J. C., G. S. Boggs, B. C. Radford, and G. A. Kendrick. 2005. Using agent-based models to aid reef restoration: enhancing coral cover and topographic complexity through the spatial arrangement of coral transplants. *Restoration Ecology* 13:685–694.
- Smith, L. W., D. Barshis, and C. Birkeland. 2007. Phenotypic plasticity for skeletal growth, density and calcification of *Porites lobata* in response to habitat type. *Coral Reefs* 26:559–567.
- Stanley, G. D., H. M. E. Shepherd, and A. J. Robinson. 2018. Paleocological response of corals to the End-Triassic Mass Extinction: an integrational analysis. *Journal of Earth Science* 29:879–885.
- Stearns, S. 1977. The evolution of life history traits: a critique of the theory and a review of the data. *Annual Review of Ecology and Systematics* 8.
- Steinthorsdottir, M., H. K. Coxall, A. M. de Boer, M. Huber, N. Barbolini, C. D. Bradshaw, N. J. Burls, et al. 2021. The Miocene: the future of the past. *Paleoceanography and Paleoclimatology* 36:e2020PA004037.
- Stuart-Smith, R. D., C. Mellin, A. E. Bates, and G. J. Edgar. 2021. Habitat loss and range shifts contribute to ecological generalization among reef fishes. *Nature Ecology and Evolution* 5:656–662.
- Swain, T. D., E. C. Bold, P. C. Osborn, A. H. Baird, M. W. Westneat, V. Backman, and L. A. Marcelino. 2018. Physiological integration of coral colonies is correlated with bleaching resistance. *Marine Ecology Progress Series* 586:1–10.
- Takayanagi, H., Y. Iryu, M. Oda, T. Sato, S. Chiyonobu, A. Nishimura, T. Nakazawa, T. Ishikawa, and K. Nagaishi. 2012. Temporal changes in biotic and abiotic composition of shallow-water carbonates on submerged seamounts in the northwestern Pacific Ocean and their controlling factors. *Geodiversitas* 34:189–217.
- Todd, P. A. 2008. Morphological plasticity in scleractinian corals. *Biological Reviews* 83:315–337.
- Ulrich, R. S., R. F. Simons, B. D. Losito, E. Fiorito, M. A. Miles, and M. Zelson. 1991. Stress recovery during exposure to natural and urban environments. *Journal of Environmental Psychology* 11:201–230.
- Vaughan, T. W. 1899. *Some Cretaceous and Eocene corals from Jamaica*. Museum of Comparative Zoology, Harvard University, Cambridge, Mass.
- Vaughn, T. W. 1919. Contributions to the geology and paleontology of the Canal Zone, Panama, and geologically related areas in central America and the West Indies. Part 11, The biologic character and geologic correlations of the sedimentary formations of Panama in their relation to the geologic history of Central America and the West Indies. U.S. Government Printing Office, Washington, D.C.
- Vaughan, T. W., and J. E. Hoffmeister. 1926. *Miocene corals from Trinidad*. Carnegie Institution of Washington, Washington, D.C.
- Vaughan, T. W., and J. W. Wells. 1943. *Revision of the suborders families, and genera of the scleractinia*, Vol. 44. Geological Society of America, New York.
- Vergés, A., P. D. Steinberg, M. E. Hay, A. G. B. Poore, A. H. Campbell, E. Ballesteros, K. L. Heck, et al. 2014. The tropicalization of temperate marine ecosystems: climate-mediated changes in herbivory and community phase shifts. *Proceedings of the Royal Society B* 281:20140846.
- Vergés, A., E. McCosker, M. Mayer-Pinto, M. A. Coleman, T. Wernberg, T. Ainsworth, and P. D. Steinberg. 2019. Tropicalisation of temperate reefs: implications for ecosystem functions and management actions. *Functional Ecology* 33:1000–1013.
- Vilhena, D. A., and A. B. Smith. 2013. Spatial bias in the marine fossil record. *PLoS ONE* 8:e74470.
- Wallace, C. C., and F. R. Bosellini. 2015. *Acropora* (Scleractinia) from the Oligocene and Miocene of Europe: species longevity, origination and turnover following the Eocene–Oligocene transition. *Journal of Systematic Palaeontology* 13:447–469.
- Webb, G. E. 1996. Morphological variation and homoplasy: the challenge of Paleozoic coral systematics. *Paleontological Society Papers* 1:135–157.
- Weisbord, N. E. 1971. *Corals from the Chipola and Jackson Bluff formations of Florida*. State of Florida, Bureau of Geology, Tallahassee.
- Weisbord, N. E. 1973. *New and little-known corals from the Tampa Formation of Florida*, Vol. 56. Florida Department of Natural Resources, Division of Interior Resources, Bureau of Geology, Tallahassee.
- Weisbord, N. E. 1974. *Late Cenozoic corals of south Florida*. Paleontological Research Institution, Ithaca, N.Y.

- Wells, J. W.** 1934a. *Eocene corals: Part I. From Cuba. Part II. A new species of Madracis from Texas*. Paleontological Research Institution, New York.
- Wells, J. W.** 1934b. Some fossil corals from the West Indies. *Proceedings of the United States National Museum* **83**:71–110.
- Wells, J. W.** 1935. Corals from the Cretaceous and Eocene of Jamaica. *Annals and Magazine of Natural History* **15**:183–194.
- Wells, J. W.** 1945. *American old and middle Tertiary larger foraminifera and corals*, Vol. **9**. Geological Society of America, New York.
- Wickham, H.** 2016. *ggplot2: elegant graphics for data analysis*. Springer-Verlag, New York.
- Woesik, R. van, E. C. Franklin, J. O’Leary, T. R. McClanahan, J. S. Klaus, and A. F. Budd.** 2012. Hosts of the Plio-Pleistocene past reflect modern-day coral vulnerability. *Proceedings of the Royal Society B* **279**:2448–2456.
- Yachi, S., and M. Loreau.** 1999. Biodiversity and ecosystem productivity in a fluctuating environment: the insurance hypothesis. *Proceedings of the National Academy of Sciences USA* **96**:1463–1468.
- Yasuhara, M., H.-H. M. Huang, M. Reuter, S. Y. Tian, J. D. Cybulski, A. O’Dea, B. L. Mamo, L. J. Cotton, E. Di Martino, and R. Feng.** 2022. Hotspots of Cenozoic tropical marine biodiversity. Pp. 243–300 in *Oceanography and marine biology*. CRC Press, Boca Raton, Fla.