



Deposited via The University of Leeds.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/231932/>

Version: Accepted Version

Article:

Świrski, M.I., Tierney, J.A.S., Albà, M.M. et al. (2025) Translon: a single term for translated regions. *Nature Methods*. ISSN: 1548-7091

<https://doi.org/10.1038/s41592-025-02810-3>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

Translon: a single term for translated regions.

by

Michał I. Świrski^{1,*}, Jack A. S. Tierney^{2,£,*}, M. Mar Albà^{3,4}, Dmitry E. Andreev^{5,6}, Julie L. Aspden^{7,8,9}, John F. Atkins^{2,10}, Michal Bassani-Sternberg^{11,12}, Marla J. Berry¹³, Stefano Biffo^{14,15}, Kathleen Boris-Lawrie¹⁶, Mark Borodovsky^{17,18}, Ian Brierley¹⁹, Matthew Brook²⁰, Marie A. Brunet^{21,22}, Janusz M. Bujnicki²³, Neva Caliskan^{24,25}, Lorenzo Calviello²⁶, Anne-Ruxandra Carvunis^{27,28}, Jamie H. D. Cate²⁹, Can Cenik³⁰, Kung Yao Chang³¹, Yiwen Chen³², Sonia Chothani³³, Jyoti S. Choudhary³⁴, Patricia L. Clark³⁵, Jim Clauwaert³⁶, Lynn Cooley³⁷, Erik Dassi³⁸, Kellie Dean², Jean-Jacques Diaz³⁹, Christoph Dieterich^{40,41}, Rivka Dikstein⁴², Jonathan D. Dinman^{43,44}, Sergey E. Dmitriev^{6,45}, Olga A. Dontsova^{5,46,47}, Christine M. Dunham⁴⁸, Sandeep M. Eswarappa⁴⁹, Philip J. Farabaugh⁵⁰, Pouya Faridi^{51,52}, Ivo Fierro-Monti⁵³, Andrew E. Firth¹⁹, David Gatfield⁵⁴, Fátima Gebauer^{55,56}, Mikhail S. Gelfand⁴⁷, Nicola K. Gray⁵⁷, Rachel Green⁵⁸, Chris H. Hill^{59,60,61}, Ya-Ming Hou⁶², Norbert Hübner^{63,64,65,66}, Zoya Ignatova⁶⁷, Pavel Ivanov^{68,69}, Shintaro Iwasaki^{70,71}, Rory Johnson⁷², Ahmad Jomaa^{73,74}, Marko Jovanovic⁷⁵, Irwin Jungreis^{76,77}, Manolis Kellis^{76,77}, Jeffrey S. Kieft⁷⁸, Alex V. Kochetov⁷⁹, Eugene V. Koonin⁸⁰, Andrei A. Korostelev⁸¹, Joanna Kufel¹, Ivan V. Kulakovskiy^{82,83}, Leo Kurian^{84,85,86}, Denis L.J. Lafontaine^{87,88}, Ola Larsson⁸⁹, Gary Loughran⁹⁰, Julius Lukeš^{91,92}, Marco Mariotti⁹³, Elena S. Martens-Uzunova⁹⁴, Thomas F. Martinez^{95,96,97}, Akinobu Matsumoto⁹⁸, Joel McManus^{99,100}, Jan Medenbach¹⁰¹, Sergey V. Melnikov¹⁰², Gerben Menschaert¹⁰³, Catharina Merchante¹⁰⁴, Martin Mikl¹⁰⁵, W. Allen Miller¹⁰⁶, Oliver Mühlemann¹⁰⁷, Olivier Namy¹⁰⁸, Danny D. Nedialkova^{109,110}, Jozef Nosek¹¹¹, Sandra Orchard⁵³, Petar Ozretić¹¹², Mihaela Perlea¹¹³, Dmitri D. Pervouchine⁴⁷, Luísa Romão^{114,115}, David Ron¹¹⁶, Xavier Roucou^{117,118}, Maria P. Rubtsova^{5,46}, Jorge Ruiz-Orera⁶³, Alan Saghatelian^{119,120}, Steven L. Salzberg¹²¹, Lucia A. Seale¹³, Cathal Seoighe¹²², Petr V. Sergiev⁶, Premal Shah^{123,124}, Nikolay Shirokikh¹²⁵, Sarah A. Slavoff¹²⁶, Nahum Sonenberg^{127,128}, Timothy J. Stasevich¹²⁹, Roman J. Szczesny¹³⁰, Tiina Tamm¹³¹, Marek Tchorzewski¹³², Ivan Topisirovic^{127,133}, Michel L. Tremblay^{127,128}, Tamir Tuller¹³⁴, Igor Ulitsky¹³⁵, Leoš Shivaya Valášek¹³⁶, Petra Van Damme¹³⁷, Gabriella Viero¹³⁸, Juan Antonio Vizcaino⁵³, Christine Vogel¹³⁹, Edward W. J. Wallace¹⁴⁰, Jonathan S. Weissman^{141,142,143}, Eric Westhof^{144,145}, Nicola Whiffin^{146,147}, Daniel N. Wilson⁶⁷, Zhi Xie¹⁴⁸, Jonathan W. Yewdell¹⁴⁹, Martina M. Yordanova², Chien-Hung Yu^{150,151}, Vyacheslav Yurchenko¹⁵², Bojan Zagrovic¹⁵³, TRANSLACORE, Eivind Valen^{154,155,*}, Pavel V. Baranov^{2,*}

¹University of Warsaw, Faculty of Biology, Institute of Genetics and Biotechnology, Warsaw, Poland

²School of Biochemistry and Cell Biology, University College Cork, Cork, T12 CY82, Ireland

³Hospital del Mar Research Institute, Dr. Aiguader 88, Barcelona 08003, Spain

⁴Catalan Institution for Research and Advanced Studies, Passeig Lluís Companys 23, Barcelona 08010, Spain

⁵Shemyakin-Ovchinnikov Institute of Bioorganic Chemistry, Moscow, 117997, Russia

⁶Belozersky Institute of Physico-Chemical Biology, Lomonosov Moscow State University, Moscow, 119234, Russia.

⁷School of Molecular and Cellular Biology, Faculty of Biological Sciences, University of Leeds, Leeds, LS2 9JT, UK

- ⁸Leeds Omics, University of Leeds, Leeds, LS2 9JT, UK
- ⁹Astbury Centre for Structural Molecular Biology, University of Leeds, Leeds LS2 9JT, UK,
- ¹⁰Department of Human Genetics, University of Utah, Salt Lake City, Utah 84112, USA
- ¹¹University Hospital of Lausanne, Lausanne, 1005, Switzerland
- ¹²Ludwig Institute for Cancer Research, Lausanne, 1005, Switzerland
- ¹³Pacific Biosciences Research Center, University of Hawaii at Manoa, Honolulu, HI, 96822, USA
- ¹⁴National Institute of Molecular Genetics, Milan, 20132, Italy
- ¹⁵University of Milan, 20126 Milan, Italy
- ¹⁶Department of Veterinary and Biomedical Sciences, University of Minnesota, Saint Paul, MN 55108, USA
- ¹⁷Wallace H. Coulter Department of Biomedical Engineering, Georgia Tech, Atlanta, Georgia, 30332, USA
- ¹⁸School of Computational Science and Engineering, Georgia Tech, Atlanta, Georgia, 30332, USA
- ¹⁹Division of Virology, Department of Pathology, University of Cambridge, Tennis Court Road, Cambridge, CB2 1QP, UK
- ²⁰Centre for Cardiovascular Sciences, Institute for Neuroscience and Cardiovascular Research, University of Edinburgh, Edinburgh, EH16 4TJ, UK
- ²¹University of Sherbrooke, Sherbrooke, J1E 4K8, Canada
- ²²Cancer Research Institute of the University of Sherbrooke (IRCUS), Sherbrooke, J1E 4K8, Canada
- ²³International Institute of Molecular and Cell Biology in Warsaw, Trojdena 4, 02-109 Warsaw, Poland
- ²⁴Regensburg Center for Biochemistry (RCB), University of Regensburg, 93053 Regensburg, Germany
- ²⁵Helmholtz Institute for RNA-Based Infection Research, Helmholtz Centre for Infection Research (HIRI-HZI), Josef-Schneider-Strasse 2, 97080 Würzburg, Germany
- ²⁶Human Technopole, 20157 Milan, Italy
- ²⁷Department of Computational and Systems Biology, School of Medicine, University of Pittsburgh, Pittsburgh, PA 15213, USA
- ²⁸Pittsburgh Center for Evolutionary Biology and Medicine (CEBaM), School of Medicine, University of Pittsburgh, Pittsburgh, PA 15213, USA
- ²⁹University of California Berkeley; Berkeley, CA 94720, USA
- ³⁰Department of Molecular Biosciences, University of Texas at Austin, Austin, TX, 78712, USA
- ³¹Graduate Institute of Biochemistry, NCHU, Taiwan

³²The Department of Bioinformatics & Computational Biology, The University of Texas MD Anderson Cancer Center, Houston, TX, 77030, USA

³³Centre for Computational Biology, Duke-NUS medical school, Singapore

³⁴Functional Proteomics team, Chester Beatty Laboratories, The Institute of Cancer Research, London SW3 6JB, UK

³⁵Department of Chemistry & Biochemistry, University of Notre Dame, Notre Dame, IN, 46556, USA

³⁶University of Michigan, Ann Arbor, 48109, USA

³⁷Department of Genetics, Yale University School of Medicine, New Haven, CT, 06510, USA

³⁸Department of Cellular, Computational and Integrative Biology (CIBIO), University of Trento, 38123, Trento, Italy

³⁹Centre de Recherche en Cancérologie de Lyon, Inserm U1052, CNRS UMR5286, Université de Lyon, Université Claude Bernard Lyon, Centre Léon Bérard, LabEx Dev2CAN, CEDEX 08, Lyon, France

⁴⁰Klaus Tschira Institute for Integrative Computational Cardiology, Heidelberg, 69120, Germany

⁴¹German Center for Cardiovascular Research (DZHK) - Partner site Heidelberg/Mannheim, Germany

⁴²Department of Biomolecular Sciences, Weizmann Institute of Science, Rehovot, 76100, Israel

⁴³Department of Cell Biology and Molecular Genetics, College Park MD, 20742, USA

⁴⁴Institute for Bioscience and Biotechnology Research, Rockville MD, 20850, USA

⁴⁵Faculty of Bioengineering and Bioinformatics, Lomonosov Moscow State University, Moscow, 119234, Russia

⁴⁶ Faculty of Chemistry, Lomonosov Moscow State University, Moscow, 119991, Russia

⁴⁷Center of Life Sciences, Skolkovo Institute of Science and Technology, Moscow, 143025, Russia

⁴⁸Department of Chemistry, Emory University, Atlanta, Georgia, USA

⁴⁹Department of Biochemistry, Indian Institute of Science, Bangalore, 560012, India

⁵⁰University of Maryland Baltimore County, Baltimore, Maryland 21250, USA

⁵¹Centre for Cancer Research, Hudson Institute of Medical Research, Melbourne, Australia, 3168

⁵²Monash Proteomics and Metabolomics Platform, Department of Medicine, School of Clinical Sciences, Monash University, Melbourne, 3168, Australia

⁵³European Molecular Biology Laboratory-European Bioinformatics Institute (EMBL-EBI), Wellcome Genome Campus, Hinxton, Cambridge. CB10 1SD. UK

⁵⁴Center for Integrative Genomics, University of Lausanne, 1015 Lausanne, Switzerland

⁵⁵Centre for Genomic Regulation (CRG), The Barcelona Institute of Science and Technology, Barcelona, 08003, Spain

- ⁵⁶Universitat Pompeu Fabra (UPF), Barcelona, 08003, Spain
- ⁵⁷Centre for Reproductive Health, Institute for Regeneration and Repair, University of Edinburgh, Edinburgh, EH16 4UU, UK
- ⁵⁸HHMI, Johns Hopkins University School of Medicine, Baltimore, Maryland, 21210 USA
- ⁵⁹Department of Biology, University of York, York, YO10 5DD, UK
- ⁶⁰York Structural Biology Laboratory, University of York, York, YO10 5DD, UK
- ⁶¹York Biomedical Research Institute, University of York, York, YO10 5DD, UK
- ⁶²Department of Biochemistry and Molecular Biology, Thomas Jefferson University, Philadelphia, PA, USA
- ⁶³Max Delbrück Center for Molecular Medicine in the Helmholtz Association (MDC), 13125, Berlin, Germany
- ⁶⁴Charite - Iniversitätsmedizin Berlin, Germany
- ⁶⁵Helmholtz-Institute for Translational AngioCardioScience (HI-TAC) of the Max Delbrück Center for Molecular Medicine in the Helmholtz Association (MDC) at Heidelberg University, Heidelberg 69117, Germany
- ⁶⁶German Center for Cardiovascular Research (DZHK) - Partner site Berlin, Germany
- ⁶⁷Institute for Biochemistry and Molecular Biology, University of Hamburg, 20146, Hamburg
- ⁶⁸Division of Rheumatology, Inflammation and Immunity, Brigham and Women's Hospital, Boston, MA, 02115, USA
- ⁶⁹Department of Medicine, Harvard Medical School, Boston, MA 02115, USA
- ⁷⁰RNA Systems Biochemistry Laboratory, Pioneering Research Institute, Wako, Saitama 351-0198, Japan
- ⁷¹Department of Computational Biology and Medical Sciences, Graduate School of Frontier Sciences, The University of Tokyo, Kashiwa, Chiba 277-8561, Japan
- ⁷²School of Biology and Environmental Science, University College Dublin, Dublin, Ireland.
- ⁷³Department of Molecular Physiology and Biological Physics, University of Virginia, Charlottesville, VA, 22903, USA
- ⁷⁴Department of Biochemistry and Molecular Genetics, University of Virginia, Charlottesville, VA 22903, USA
- ⁷⁵Columbia University, New York, NY 10025, USA
- ⁷⁶MIT Computer Science and Artificial Intelligence Laboratory, Cambridge, MA, 02139 USA
- ⁷⁷Broad Institute of MIT and Harvard, Cambridge, MA, 02142, USA
- ⁷⁸New York Structural Biology Center, New York, NY, 10027, USA
- ⁷⁹Institute of Cytology & Genetics, Novosibirsk, Russia

⁸⁰Computational Biology Branch, Division of Intramural Research, National Library of Medicine, National Institutes of Health, Bethesda, MD 20894, USA

⁸¹RNA Therapeutics Institute, University of Massachusetts Chan Medical School. Worcester, MA 01605, USA

⁸²Institute of Protein Research, Russian Academy of Sciences, Pushchino, 142290, Russia

⁸³Vavilov Institute of General Genetics, Russian Academy of Sciences, Moscow, 119991, Russia

⁸⁴Institute for Cardiovascular Physiology, Goethe University Frankfurt, Germany

⁸⁵German Center for Cardiovascular Research (DZHK), Partner Site Rhein-Main, D-60590 Frankfurt, Germany.

⁸⁶Cardio-Pulmonary Institute (CPI), Goethe University Frankfurt, Frankfurt, Germany.

⁸⁷RNA Molecular Biology, Université libre de Bruxelles, Biopark campus, B-6041 Gosselies, Belgium

⁸⁸Fonds de la Recherche Scientifique (F.R.S./FNRS), B-1000 Brussels, Belgium

⁸⁹Department of Oncology-Pathology, Science for Life Laboratory, Karolinska Institutet, 17165 Stockholm, Sweden

⁹⁰EIRNA Bio, Cork, Ireland

⁹¹Institute of Parasitology, Biology Centre, 37005 České Budějovice, Czech Republic

⁹²Faculty of Science, University of South Bohemia, 37005 České Budějovice, Czech Republic

⁹³Department of Genetics, Microbiology and Statistics, Universitat de Barcelona, Barcelona, Catalonia, 08024, Spain

⁹⁴Erasmus MC Cancer Institute, University Medical Center Rotterdam, Department of Urology, 3000CA Rotterdam, The Netherlands

⁹⁵Department of Pharmaceutical Sciences, University of California, Irvine, Irvine, CA 92617, USA

⁹⁶Department of Biological Chemistry, University of California, Irvine, Irvine, CA 92617, USA

⁹⁷Chao Family Comprehensive Cancer Center, University of California, Irvine, Irvine, CA 92617, USA

⁹⁸Division of Biological Science, Graduate School of Science, Nagoya University, Nagoya 464-8602, Japan

⁹⁹Department of Biological Sciences, Carnegie Mellon University, Pittsburgh, PA, 15213, USA

¹⁰⁰Computational Biology Department, Carnegie Mellon University, Pittsburgh, PA, 15213, USA

¹⁰¹University Hospital RWTH Aachen, 52074 Aachen, Germany

¹⁰²Biosciences Institute, Newcastle University, Newcastle upon Tyne, NE2 4HH, UK

¹⁰³Faculty of Bioscience Engineering, Department of Data Analysis and Mathematical Modelling, Ghent University, Ghent, 9000, Belgium

- ¹⁰⁴Instituto de Hortofruticultura Subtropical y Mediterránea La Mayora (IHSM-UMA-CSIC). Boulevard Louis Pasteur, 49. 29010 Málaga, Spain
- ¹⁰⁵Department of Human Biology, University of Haifa, Haifa, 3498838, Israel
- ¹⁰⁶Plant Pathology, Entomology & Microbiology Department, Iowa State University, Ames, Iowa, 50011, USA
- ¹⁰⁷Department of Chemistry, Biochemistry and Pharmaceutical Sciences, University of Bern, 3012 Bern, Switzerland
- ¹⁰⁸CEA, CNRS, Institute for Integrative Biology of the Cell (I2BC), Université Paris-Saclay, Gif-sur-Yvette, 91198, France
- ¹⁰⁹Max Planck Institute of Biochemistry, 82152 Martinsried, Germany
- ¹¹⁰Department of Bioscience, TUM School of Natural Sciences, 85748 Garching, Germany
- ¹¹¹Comenius University Bratislava, Faculty of Natural Sciences, 842 15 Bratislava, Slovakia
- ¹¹²Laboratory for Hereditary Cancer, Division of Molecular Medicine, Ruđer Bošković Institute, 10000 Zagreb, Croatia
- ¹¹³Department of Biomedical Engineering, Johns Hopkins University, Baltimore, Maryland 21211, USA
- ¹¹⁴Instituto Nacional de Saúde Doutor Ricardo Jorge, 1649-016 Lisboa, Portugal
- ¹¹⁵BioISI-Biosystems Integrative Sciences Institute, Faculty of Sciences, University of Lisboa, 1749-016 Lisboa, Portugal
- ¹¹⁶Cambridge Institute for Medical Research, Department of Clinical Biochemistry, University of Cambridge, Cambridge CB2 0XY, UK
- ¹¹⁷Department of biochemistry and functional genomics, Université de Sherbrooke, 3201 Jean Mignault, Sherbrooke, QC J1E 4K8, Canada
- ¹¹⁸Centre de Recherche du Centre Hospitalier Universitaire de Sherbrooke (CRCHUS), Sherbrooke, QC J1H 5N4, Canada
- ¹¹⁹Salk Institute, La Jolla, CA, 92037, USA
- ¹²⁰UCSD, La Jolla, CA, 92037, USA
- ¹²¹Center for Computational Biology, Johns Hopkins University, Baltimore, Maryland 21211, USA
- ¹²²School of Mathematical and Statistical Sciences, University of Galway, Galway, Ireland
- ¹²³Convergence Bio, MA USA 02139
- ¹²⁴Rutgers University, NJ USA 08807
- ¹²⁵School of Human Sciences, University of Western Australia, Crawley, M309, Perth, 6009, Western Australia, Australia
- ¹²⁶Department of Chemistry, Yale University, New Haven, CT 06520, USA
- ¹²⁷Department of Biochemistry, McGill University, Montréal, QC, H3G 1Y6, Canada

- ¹²⁸Rosalind and Morris Goodman Cancer Research Centre, McGill University, Montréal, QC, H3A 1A3, Canada
- ¹²⁹Department of Biochemistry and Molecular Biology, Colorado State University, Fort Collins, CO, 80525, USA
- ¹³⁰Laboratory of RNA Biology, Institute of Biochemistry and Biophysics Polish Academy of Sciences, Warsaw, 02-106, Poland
- ¹³¹Institute of Molecular and Cell Biology, University of Tartu, Tartu 51010, Estonia
- ¹³²Department of Molecular Biology, Institute of Biological Sciences, Maria Curie-Skłodowska University, Lublin, Poland
- ¹³³Lady Davis Institute, Gerald Bronfman Department of Oncology, Division of Experimental Medicine, McGill University, Montréal H3A 1A3, Canada
- ¹³⁴Department of Biomedical Engineering, Tel-Aviv University, Tel Aviv, Israel
- ¹³⁵Department of Immunology and Regenerative Biology, Weizmann Institute of Science, Rehovot 76100, Israel
- ¹³⁶Laboratory of Regulation of Gene Expression Institute of Microbiology of the Czech Academy of Sciences Videnska 1083, 142 20, Prague 4, Czech Republic
- ¹³⁷iRIP Unit, Laboratory of Microbiology, Department of Biochemistry and Microbiology, Ghent University, 9000 Ghent, Belgium
- ¹³⁸Institute of Biophysics CNR, Via Sommarive 18 38123 Trento, Italy
- ¹³⁹Department of Biology, New York University, New York, 10003, USA
- ¹⁴⁰School of Biological Sciences, University of Edinburgh, Edinburgh, EH9 3FF, UK
- ¹⁴¹Whitehead Institute, Cambridge, MA 02139, USA
- ¹⁴²Department of Biology, MIT, Cambridge, MA 02139 USA
- ¹⁴³Howard Hughes Medical Institute, Chevy Chase, MD 20815, USA
- ¹⁴⁴Université de Strasbourg, Institut de Biologie Moléculaire et Cellulaire, Architecture et Réactivité de l'ARN, CNRS UPR9002, 2 allée Konrad Roentgen, F-67084 Strasbourg, France
- ¹⁴⁵Engineering Research Center of Clinical Functional Materials and Diagnosis & Treatment Devices of Zhejiang Province, Wenzhou Institute, University of Chinese Academy of Sciences, Wenzhou, China
- ¹⁴⁶Big Data Institute, University of Oxford, Oxford, OX3 7LF, UK
- ¹⁴⁷Centre for Human Genetics, University of Oxford, Oxford, OX3 7BN, UK
- ¹⁴⁸State Key Laboratory of Ophthalmology, Zhongshan Ophthalmic Center, Sun Yat-sen University, Guangzhou, 510060, China
- ¹⁴⁹Cellular Biology Section, Laboratory of Viral Diseases, Division of Intramural Research, NIAID, Bethesda, 20892, USA

¹⁵⁰Department of Biochemistry and Molecular Biology, National Cheng Kung University, Tainan, 701, Taiwan

¹⁵¹Institute of Basic Medical Sciences, College of Medicine, National Cheng Kung University, Tainan, 701, Taiwan

¹⁵²Life Science Research Centre, Faculty of Science, University of Ostrava, 710 00 Ostrava, Czech Republic

¹⁵³Department of Structural and Computational Biology, Max Perutz Labs, University of Vienna, Campus Vienna Biocenter 5, 1030, Vienna, Austria

¹⁵⁴Department of Biosciences, University of Oslo, Kristine Bonnevis hus, Blindernveien 31, 0731 Oslo, Norway

¹⁵⁵Computational Biology Unit, Department of Informatics, University of Bergen, Thormøhlensgate 55, 5008 Bergen, Norway.

‡Current affiliation: European Molecular Biology Laboratory-European Bioinformatics Institute (EMBL-EBI), Wellcome Genome Campus, Hinxton, Cambridge. CB10 1SD, UK

*To whom correspondence should be addressed: michal.swirski@uw.edu.pl, jackt@ebi.ac.uk, eivind.valen@ibv.uio.no, p.baranov@ucc.ie

During translation, ribosomes synthesise polypeptides using RNA molecules as templates. All cellular proteins are products of translation, and the identification of protein-coding regions is the primary goal of genome annotation. Beyond protein synthesis, translation has long been known to have regulatory functions independent of its products^{1,2}. However, only with the advent of ribosome profiling, was the broad scale and complexity of translated regions fully appreciated.

Recent interest in pervasive translation has exposed a lack of general terminology for translated regions that does not depend on the properties of their products or their sequence. In the absence of such terminology, a range of inconsistently defined terms are currently used to describe them. These terms are typically variations on Open Reading Frame (ORF), for example, **non-canonical**, **RiboSeq**, **alternative**, **translated**, **small**, **short**, and other **xORFs**-based labels. Such terms largely overlap and redundantly describe the same core concept, that the region in question is translated.

Denoting translated regions with ORF-based terms is problematic for two main reasons. First, in sequence analysis, ORFs are defined purely by the nucleotide sequence and the relevant genetic code (starts/stops; Fig. 1a). Thus, ORFs are found everywhere in the genome, including regions that are not transcribed. This ORF definition is useful for the prediction of potential protein-coding regions. However, using the same term to specify regions that are known or confidently predicted to be translated leads to confusion. Indeed, for annotated protein-coding regions, a different technical term, CDS (historically from ‘Coding DNA Sequence’), is commonly used.

The second problem is that, although most known protein-coding regions in the majority of model organisms conform to an ORF, starting at an AUG and terminating at the first in-frame stop, there are numerous alternatives. In addition to AUG, other triplets can also initiate translation³ (Fig. 1a), making it challenging to determine start codons purely from sequence. The incorporation of non-standard proteinogenic amino acids (selenocysteine/pyrrolysine) usually occurs at stop codons within the CDS, with over a hundred in CDSs encoding selenoproteins in some species⁴ (Fig. 1a). Furthermore, in some organisms, termination codons are defined not only by their sequence, but also by their position within mRNA⁵.

Ribosomal frameshifting (Fig. 1a), which involves translation of two different reading frames to produce a single protein, is common in viruses and also occurs in nuclear-encoded genes of most organisms, in some species, in 5-20% of the genes⁶. Because the products of translation of frameshifted CDSs cannot be derived automatically by converting nucleotide triplets to amino acids, current annotation practice is to introduce a “pseudo-intron” between two partial ORFs in the annotation of these genes (Fig. 1b). Most concerning is the practice of automatically modifying RNA sequences inferred from the genome to obtain full-length protein product via triplet translation (Fig. 1c). Such artificial fitting of well-established translation mechanisms to a simplified model reinforces a widespread fallacy that all translated regions can be represented as single ORFs. This leads to an inaccurate and oversimplified representation of genetic information and molecular composition of the cells, with potentially serious consequences for those unaware of the underlying makeshift solutions. Similar terminological confusion, such as the routine conflation of “exons” and “protein-coding regions”⁷, highlights the need for precise vocabulary relating to gene expression.

The above problems can be solved or at least alleviated with the introduction of a specific term for a translated region that would be defined without reference to the product of translation or to the sequence of the region. For this purpose, we suggest using the term *translon* (short for **translated region**), which has previously been introduced but failed to gain traction⁸. It aligns well

with other terms describing gene structures: intron (**intragenic region**) and exon (**expressed region**)⁹.

The term translon denotes any region that is decoded by the ribosome. This ranges from minimal sequences with detectable translation (AUG followed by a stop) to sequences encoding long proteins in multiple reading frames, and even those disrupted by non-coding sequences as in translational bypassing¹⁰ (Fig. 1a). It also facilitates the efforts to characterise unannotated translation: newly identified translated regions can be described as novel translons. Their biological roles, if any, can remain enigmatic until specific information is obtained. These roles can be purely regulatory, independent of the translation products, or involving those products, such as short peptides modulating functions of other proteins, signalling peptides or antigens; some translons could encode neutral or even harmful “protein junk”.

Translon would fill the gap in the vocabulary used to describe units of genetic information (Fig. 1d). Unlike the existing terminology, translon is directly defined by the process it aims to capture instead of doing so indirectly, through sequence or function. We expect that this term will reduce confusion when discussing translated regions and facilitate development of more biologically realistic annotations.

ACKNOWLEDGMENTS

We wish to thank the participants of 2024 EMBO Workshop “Recoding and the diversity of genetic decoding” and of 2024 Dagstuhl Seminar “Deep Learning for RNA Regulation and Multidimensional Transcriptomics” as well as the members of TransCODE consortium and TRANSLACORE network (supported by COST action CA21154) for invaluable discussions that led to the development of this proposal.

AUTHOR CONTRIBUTIONS

M.I.S., J.A.S.T, E.V., and P.V.B developed the concept and drafted the correspondence. M.M.A., D. E.A., J.L.A., J.F.A., M.B.S., M.J.B., S.B., K.B.-L., M.Borodovsky, I.B., M.Brook, M.A.B., J.M.B., N.C., L.Calviello, A.-R.C., J.H.D.C., C.C., K.Y.C., Y.C., S.C., J.S.C., P.L.C, J.C., L.Cooley, E.D., K.D., J.J. D., C.D., R.D., J.D.D., S. E.D., O.A.D., C.M.D., S.M.E., P.J.F., P.F., I.F. -M., A.E.F., D.G., F.G., M.S.G., N. K.G., R.G., C.H.H., Y.-M.H., N.H., Z.I., P.I., S.I., R.J., A.J., M.J., I.J., M.K., J.S.K., A.V.K., E.V.K., A.A.K., J.K., I.V.K., L.K., D.L.J.L., O.L., G.L., J.L, M.Mariotti, E.S.M.-U., T.F.M., A.M., J.McManus, J.Medenbach, S.V.M., G.M., C.M., M.Mikl, W.A.M., O.M., O.N., D.D.N., J.N., S.O., P.O., M.P., D.D.P., L.R, D.R., X.R., M.P.R., J.R.-O., A.S., S.L.S., L.A.S., C.S., P.V.S., P.S., N.Shirokikh, S.A.S., N.Sonenberg, T.J.S., R.J.S., T. Tamm, M.T., I.T., M.L.T., T.Tuller, I.U., L.S.V., P.V.D., G.V., J.A.V., C.V., E.W.J.W., J.S.W, E.W., N.W., D.N.W, Z.X., J.W.Y., M.M.Y., C.-H.Y., V.Y., and B.Z. critically discussed the concept and/or edited the manuscript at different stages of its development.

ETHICS DECLARATION

Competing interests

A.-R.C. is on the scientific advisory board of ProFound Therapeutics. G.L. & P.V.B. are co-founders and shareholders of Eirnabio Ltd. The remaining authors declare no competing interests.

REFERENCES

1. Miller, P. F. & Hinnebusch, A. G. cis-acting sequences involved in the translational control of GCN4 expression. *Biochim Biophys Acta* 1050, 151–154 (1990).

2. Orbach, M. J., Sachs, M. S. & Yanofsky, C. The *Neurospora crassa* arg-2 locus. Structure and expression of the gene encoding the small subunit of arginine-specific carbamoyl phosphate synthetase. *J Biol Chem* 265, 10981–10987 (1990).
3. Andreev, D. E. et al. Non-AUG translation initiation in mammals. *Genome Biol* 23, 111 (2022).
4. Baclaocos, J. et al. Processive Recoding and Metazoan Evolution of Selenoprotein P: Up to 132 UGAs in Molluscs. *J Mol Biol* 431, 4381–4407 (2019).
5. Zinshteyn, B. & Green, R. When stop makes sense. *Science* 354, 1106 (2016).
6. Lobanov, A. V. et al. Position-dependent termination and widespread obligatory frameshifting in *Euplotes* translation. *Nat Struct Mol Biol* 24, 61–68 (2017).
7. Aspden, J. L., Wallace, E. W. J. & Whiffin, N. Not all exons are protein coding: Addressing a common misconception. *Cell Genom* 3, 100296 (2023).
8. Goel, S. C. Transcription Unit. *Nature* 245, 397–397 (1973).
9. Gilbert, W. Why genes in pieces? *Nature* 271, 501 (1978).
10. Nosek, J., Tomaska, L., Burger, G. & Lang, B. F. Programmed translational bypassing elements in mitochondria: structure, mobility, and evolutionary origin. *Trends Genet* 31, 187–194 (2015).

FIGURE LEGEND

Figure 1| Open Reading Frames (ORFs) and translated regions. **a.** Any nucleotide sequence can be represented as three sequences of nucleotide triplets (codons), i.e. triplets read in three different reading frames. An ORF is defined as a sequence of sense codons bounded by a stop codon (purple bar) or from a start (most commonly ATG) to a stop (light blue bar). Below are examples of how different translated regions (translons) relate to ORF. **b.** Graphical representation of human PEG10 annotation zoomed in the area of ribosomal frameshifting site. Two different transcripts are used to represent two different proteins synthesised during translation of PEG10 mRNA. The one representing the full-length protein is artificially modified with a two-nucleotide “pseudo-intron”. As a result of this modification, the product of conceptual translation differs from the full-length product of actual translation by one amino acid **c.** A fragment of macaca *OAZ1* gene transcript (XM_050770746.1) annotation illustrating automatic “correction” of mRNA sequences relative to the genomic sequence. The alignment of predicted RNA and genomic source is shown below, with the conserved stop codon at the *OAZ1* frameshifting site shown in red and deleted T in bold. **d.** Relationship of the translon to other terms used when interpreting genome information. Translon fills a missing step in the description of genetic information flow in the cells. Below is a relationship of translon to ORFs and CDSs: most ORFs are not transcribed. Of those that are, most are not translated; all CDSs are translons, but not all translons and CDSs are ORFs.

a

Open reading frames (ORFs)

Reading frame
 1 TCT TGA CAT GTT AAT CTT CTC CTG AGT GTC ACG AGC ACC CTC CGT AAA CT
 2 T CTT GAC **ATG TTA ATC TTC TCC TGA** GTG TCA CGA GCA CCC TCC GTA AAC
 3 TC TTG ACA TGT **TAA TCT TCT CCT GAG TGT CAC GAG CAC CCT CCG TAA** ACT



b

20 bases | hg38
 94,664,500 | 94,664,510 | 94,664,520 | 94,664,530 | 94,664,540
 CAAAGTC TTGCGCG **GGG**AAACTCCCGGCCCGCTGTAGAGGGACCTTCAGC
 Q S L R R R E T L P R P R C V E R G T F S A
 K V F A G G K L P G P A V E R D L Q
 S K S S P A G N S P A P L
 S K S S P G K L P G P A V E G P S A

Annotation remark: This transcript is subject to -1 ribosomal frameshifting. This has been represented in the CDS by the addition of a 2 base intron and as a consequence misses 1 amino acid.

← shift
 P A G K
 Actual: CCG GCG GGA AAC
 Annotated: CCG --G GGA AAC
 P G K

c

LOCUS XM_050770746 1142 bp mRNA linear PRI 26-FEB-2023
 DEFINITION PREDICTED: Macaca thibetana thibetana ornithine decarboxylase antizyme 1 (OAZ1), mRNA.

##RefSeq-Attributes-START##
 frameshifts: **corrected 1 indel**
 ##RefSeq-Attributes-END##

gene 1.1142
 /gene="OAZ1"
 /note="ornithine decarboxylase antizyme 1; The sequence of the model RefSeq transcript was modified relative to its source genomic sequence to represent the inferred CDS: **deleted 1 base in 1 codon**. Derived by automated computational analysis using gene prediction method: Gnomon. Supporting evidence includes similarity to: 1159 ESTs, 8 Proteins"
 /db_xref="GeneID:126942773"

"RNA" ATTCTACTCC-GATGATCGGCT XM050770746.1
 DNA ATTCTACTCC**TGA**TGATCGGCT BC100291.1

