



This is a repository copy of *Collective behavior of urease pH clocks in nano- and microvesicles controlled by fast ammonia transport*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/230621/>

Version: Published Version

Article:

Miele, Y., Jones, S.J., Rossi, F. orcid.org/0000-0002-1854-532X et al. (2 more authors) (2022) Collective behavior of urease pH clocks in nano- and microvesicles controlled by fast ammonia transport. *The Journal of Physical Chemistry Letters*, 13 (8). pp. 1979-1984. ISSN: 1948-7185

<https://doi.org/10.1021/acs.jpclett.2c00069>

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:

<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Collective Behavior of Urease pH Clocks in Nano- and Microvesicles Controlled by Fast Ammonia Transport

Ylenia Miele,[#] Stephen J. Jones,[#] Federico Rossi,^{*} Paul A. Beales,^{*} and Annette F. Taylor^{*}



Cite This: *J. Phys. Chem. Lett.* 2022, 13, 1979–1984



Read Online

ACCESS |



Metrics & More

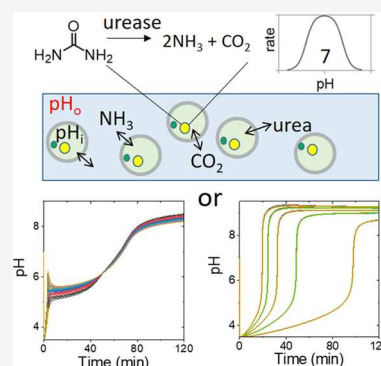


Article Recommendations



Supporting Information

ABSTRACT: The transmission of chemical signals via an extracellular solution plays a vital role in collective behavior in cellular biological systems and may be exploited in applications of lipid vesicles such as drug delivery. Here, we investigated chemical communication in synthetic micro- and nanovesicles containing urease in a solution of urea and acid. We combined experiments with simulations to demonstrate that the fast transport of ammonia to the external solution governs the pH–time profile and synchronizes the timing of the pH clock reaction in a heterogeneous population of vesicles. This study shows how the rate of production and emission of a small basic product controls pH changes in active vesicles with a distribution of sizes and enzyme amounts, which may be useful in bioreactor or healthcare applications.



Lipid vesicles, or liposomes, are employed in enzyme bioreactors^{1,2} and healthcare applications³ and are also used for the construction of artificial cells with bioinspired dynamics.^{4–6} The lipid membranes provide a protective layer with reduced permeability to large molecules and ionic species, and the release of chemicals from the vesicles can be used for molecular communication.^{7–9} Here, we investigated the role of the emission of base on the urease reaction when the enzyme was confined in synthetic nano- or microvesicles. Urease catalyzes the hydrolysis of urea, producing ammonia.¹⁰ In aqueous-phase experiments the reaction displays pH-dependent feedback and a rapid switch, referred to as a pH clock, after an induction period where the pH increases slowly.¹¹ The reaction is widely used in materials and sensing applications,^{12–17} and urease has been encapsulated in vesicles and polymersomes;^{18–22} however, the influence (if any) of compartmentalization and chemical communication on the process is not well understood.

Collective behavior has been mainly investigated with the inorganic Belousov–Zhabotinsky oscillating reaction in emulsion microdroplets or particles and vesicles.^{23–26} The period of the reaction depended on the catalyst loading and the particle size, and the products diffused between compartments, synchronizing the oscillations or driving more complex responses.^{27–31} In the encapsulation of more biologically relevant DNA and RNA transcriptional oscillators³² and protein oscillators,³³ all the reactive species were confined to the microdroplets or vesicles; however, with urease-encapsulated vesicles, neutral acidic (CO₂) and basic products (NH₃) can diffuse into the surrounding solution. The methods for producing vesicles typically result in a distribution of sizes and

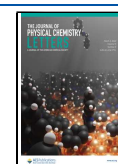
enzyme content and so a variation in the pH clock time in individual vesicles might be expected in the absence of a collective response.^{34,35} Theoretical work also suggested that autonomous pH oscillations may occur in urease vesicles providing there is the sufficiently fast transport of acid from the external solution ($P_{H^+} > 10^{-5} \text{ m s}^{-1}$); to date, however, these have not been observed in experiments.^{36–38} We show that the fast transport of ammonia controls the pH–time profile and synchronizes the pH change in the vesicles; here, the term synchronization is used to refer to a change in behavior (low to high pH) occurring at the same time in a heterogeneous population.

Nanovesicles were prepared using phospholipid film hydration and extrusion and encapsulated a solution of urease, pyranine, and HCl (SI 1.2–1.3 and Figure 1a). As the urease (Sigma-Aldrich type III) is not pure, we report enzyme concentrations in units per milliliter rather than micromoles. The reaction was initiated by adding a urea/HCl solution to a vesicle solution in a microcuvette. The ratio of the absorbance of pyranine at 450 and 405 nm was used to estimate the (apparent) pH using a calibration curve with a fitted theoretical relation (SI 1.4 and Figure S1),^{39,40} and the total number of vesicles in the 500 μL sample was on the order of $N \sim 10^{11}$ (SI 1.5).

Received: January 10, 2022

Accepted: February 17, 2022

Published: February 21, 2022



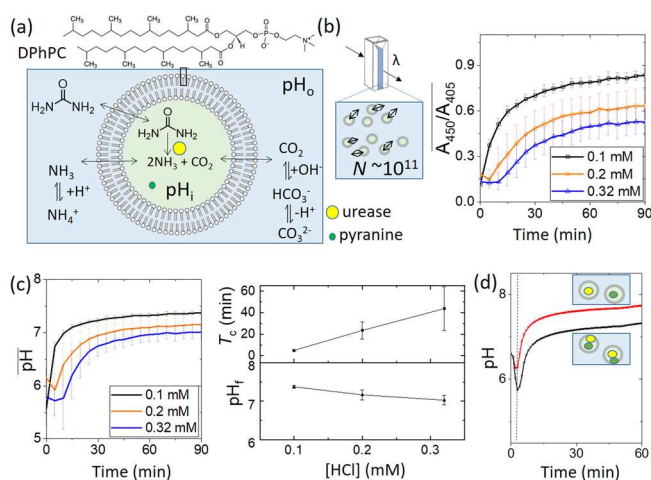


Figure 1. Urease pH clock reaction in nanovesicles of diameter $D \sim 200$ nm that were prepared using 1,2-diphytanoyl-*sn*-glycero-3-phosphocholine (DPhPC), urease (220 U mL^{-1}), HCl (0.1–0.32 mM), and pyranine (20 mM) and placed in a solution of urea (50 mM) and HCl (0.1–0.32 mM). (a) Schematic of the reaction with urease and pyranine confined to the vesicle (pH_i), with urea in the outer solution (pH_o) and the relevant equilibria. (b) Average ratio of absorbance from vesicles in a microcuvette in a urea/acid solution and the estimated total number of vesicles, N , in $500 \mu\text{L}$. (c) The average pH in time obtained from the ratio of the absorbance in panel b, showing the average clock time T_c (here to pH 6.75) and the final pH as a function of initial acid concentration. (d) Comparison of the pH in time in experiments with $[\text{HCl}] = 0.2 \text{ mM}$ where pyranine was included in the same vesicles as the enzyme (lower black curve) and where pyranine and the enzyme were in separate vesicles (upper red curve). The dotted vertical line corresponds to mixing time. Error bars indicate the standard error from three independent experiments.

The ratio, A_{450}/A_{405} , is shown in time for different initial acid concentrations in Figure 1b, and the corresponding pH is in Figure 1c. In the aqueous phase, the clock time, T_c , was defined as the time to reach pH 7, where the urease reaction rate was at a maximum, and depended on the acid concentration, the urea, and the amount of enzyme present in solution.¹¹ The switch in pH was generally less sharp in the nanovesicles compared to that of the pH clocks in the aqueous phase, with a higher initial pH (after mixing) and a lower final pH (SI 1.6). However, the average clock time increased with the initial acid concentration as expected (Figure 1c).

Evidence for the increase in the amount of ammonia in the outer solution was obtained through experiments in which two populations of vesicles were prepared, one with pyranine but no enzyme and one with enzyme but no pyranine. When the two were mixed and urea solution was added, the pH increased, demonstrating the transfer of ammonia to the urease-free vesicles via the outer solution (Figure 1d). To rule out the possibility that enzymes from burst vesicles could contribute to the overall pH change in the cuvette, Triton-X was added to the solution to rupture all the vesicles. No increase in absorbance was observed in time (Figure S3).

The slow increase in pH in the nanovesicles obtained using absorbance measurements may arise from the average of a broad distribution in clock times in a diverse population or the increase may be slow and synchronized in all vesicles. In order to monitor the pH clock in individual vesicles, the urease reaction was performed in microvesicles prepared by the droplet transfer method (SI 2.1–2.2).^{19,41,42} A sample of vesicles was added to a reaction chamber, and the ratio of the

fluorescence intensity (F_{458}/F_{405}) obtained using confocal microscopy was used to determine the (apparent) pH, with a fitted theoretical relationship (SI 2.3–2.4). The sizes of the vesicles ranged from 2–40 μm (Figure 2a), and the total number of vesicles in the chamber was $N \sim 10^4$ (SI 2.5).

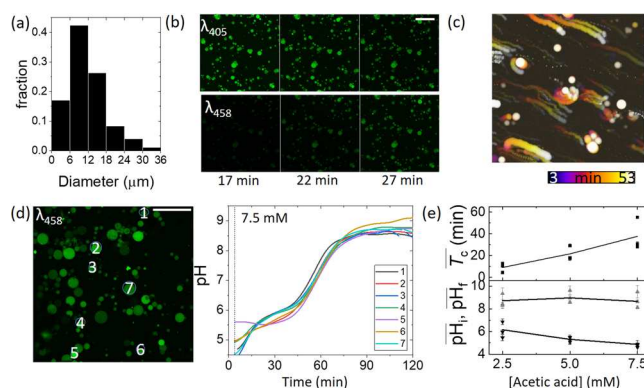


Figure 2. Urease pH clock reaction in synthetic microvesicles prepared using 1-palmitoyl-2-oleoyl-*sn*-glycero-3-phosphocholine (POPC), urease (80 U mL^{-1}), pyranine (50 μM), and acetic acid (2.5–7.5 mM) in a solution of urea (32 mM) and acetic acid (2.5–7.5 mM). (a) Fraction of vesicles with a given diameter taken from reaction data with nine runs. (b) Confocal images in time of the urease pH clock reaction in vesicles with 5 mM acetic acid for two different excitation wavelengths. (c) Image overlay ($\lambda_{458\text{nm}}$) for the entire time series in an experiment with an acid concentration of 2.5 mM showing vesicle motion. The colored bar indicates time. (d) Confocal image of the seven vesicles used in data analysis for 7.5 mM acid and the pH in time in each vesicle, where the dotted vertical line corresponds to the mixing time. (e) Clock time T_c and initial (black) and final pH (gray) levels from experiments with different initial acid concentrations. Data points are the mean and standard error from seven vesicles in a single experiment, and the line shows the average from the three independent experiments. The white scale bar on the images is equal to 100 μm .

A series of confocal images obtained from a typical experiment are shown in Figure 2b. There was a gradual increase in the fluorescence intensity in all vesicles following excitation at 458 nm, corresponding to an increase in pH as the reaction progressed. The coordinated transport of some vesicles, particularly at lower initial acid concentrations, was observed after the pH clock, possibly due to convection in the external solution (Figure 2c).⁴³

The pH–time profile in seven individual vesicles is shown in Figure 2d. The rate of increase of pH in the vesicles was more gradual than that in aqueous-phase experiments. There was little evidence of a correlation between the vesicle diameter and the clock time (Figure S6) or between the spatial position of the vesicles and the clock time; the timing of the switch from pH 6 to 8 was similar in each vesicle. Overall, there was an increase in the clock time and a decrease in the initial pH with an increase in the initial acid concentration, as expected (SI 2.6 and Figure S7). The average clock time varied between repeats, but the standard deviation was small in each experiment, suggesting the possibility of a synchronized switch in pH in the vesicles mediated by the emission of ammonia (Figure 2e).

Propagating reaction–diffusion fronts were not observed in either the nano- or microvesicles under these conditions, probably as a result of the low concentration of vesicles.

Hence, some insight can be gained from simulations with a simple ODE model of the vesicles and the external solution (SI 3.4). The reaction can be modeled by taking into account stochastic effects; however, in other work it was determined that population-level behavior was retained in the ODE models.^{38,44} The rate of change of the concentration of a species A_i in a vesicle was determined by the reaction and mass transfer rate as follows:^{45–47}

$$\frac{dA_i}{dt} = f(A_i) + \frac{3P_i}{r}(A_o - A_i) \quad (1)$$

where $f(A_i)$ contains the enzyme reaction and solution equilibria terms (SI 3.1–3.2) taken from earlier work,^{10,11,48} A_o is the concentration of the species in the outer solution, P_i is the permeability coefficient of species i , and r is the radius of the vesicle. In the outer solution, the rate of change of the concentration of each species, A_o , was given by the reaction rate ($g(A_o)$) and the mass transfer rate (for identical vesicles) as follows:

$$\frac{dA_o}{dt} = g(A_o) + \phi \frac{3P_i}{r}(A_i - A_o) \quad (2)$$

where $\phi = NV_i/V_o$ = the vesicle volume fraction, which takes into account dilution as a result of the volume change from vesicle to solution. The permeability coefficients for the neutral species were $P_{\text{NH}_3} = 1 \times 10^{-4} \text{ m s}^{-1}$, $P_{\text{CO}_2} = 1 \times 10^{-6} \text{ m s}^{-1}$, and $P_{\text{Urea}} = 1 \times 10^{-8} \text{ m s}^{-1}$ are broadly in line with literature values (see SI 3.3).^{47,49} We assumed the permeability of the membrane to all ions (NH_4^+ , CO_3^{2-} , HCO_3^- , H^+ , OH^- , and pyranine) was negligible.

In experiments with nanovesicles of diameters $\sim 200 \text{ nm}$, the volume fraction of vesicles was estimated as $\phi = NV_i/V_o \sim 2 \times 10^{-3}$ (see SI 1.6). A similar pH–time profile was obtained in the simulations with the enzyme concentration of $[E] = 55 \text{ U mL}^{-1}$, thus assuming an encapsulation efficiency of 25%.^{50,51} Initially, the pH increased rapidly in the vesicles, reaching a steady value around $\text{pH} = 5.5$ (Figure 3a, black curve). The pH switch in the vesicles at 15 min was accompanied by an increase in the pH of the outer solution (Figure 3a, red curve). The final pH in the vesicles was lower than that of the external solution as a result of the buffering effect of pyranine (Figure S8a) and the fact that the reaction was not at equilibrium at $T = 90 \text{ min}$, as less than 2% of the urea was consumed (Figure S8b). The model was able to reproduce the experimental trends with changes in the initial acid concentration (Figure S9).

The pH–time profile was mainly controlled by the transfer of ammonia to the outer solution (Figure S10). With a lower permeability coefficient of NH_3 , the pH increased rapidly in the vesicle to a high pH with no change in the outer solution, whereas for greater P_{NH_3} the pH in the vesicle and the external solution were the same (Figure 3a). The clock time increased to 90 min with $P_{\text{NH}_3} = 1 \times 10^{-2} \text{ m s}^{-1}$, and the effect of encapsulation was eliminated. The same result could be obtained by having the enzyme dispersed in the external solution and ammonia diffusing into empty vesicles. This illustrates that compartmentalization played an important role in the pH–time profile in the vesicles, as the partial entrapment of ammonia raised the internal pH of the vesicles and enhanced the rate. Nevertheless, the switch was less sharp than that in aqueous-phase experiments because of the relatively fast loss of ammonia to the outer solution.

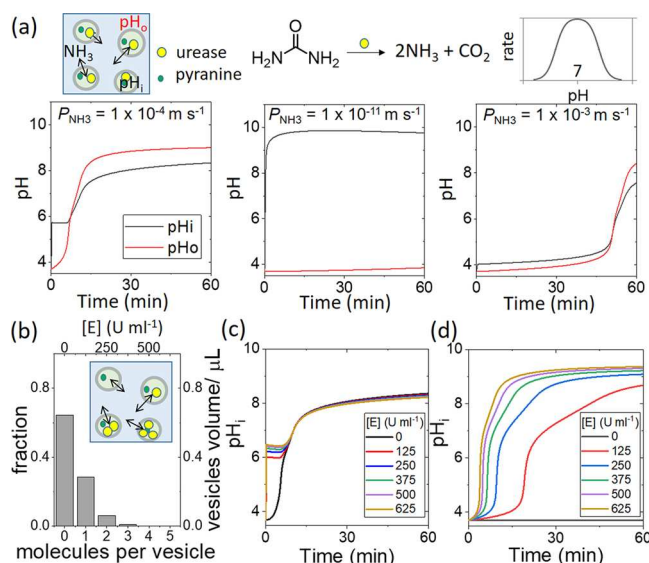


Figure 3. Simulations of the pH clock with a population of nanovesicles, $D = 200 \text{ nm}$, with $[E] = 55 \text{ U mL}^{-1}$, $[\text{pyranine}] = 5 \text{ mM}$, and $[\text{HCl}] = 0.2 \text{ mM}$ in a solution of $[\text{urea}] = 50 \text{ mM}$ and $[\text{HCl}] = 0.2 \text{ mM}$ with a vesicle volume fraction $\phi \sim NV_i/V_o = 2 \times 10^{-3}$. (a) pH in time in identical vesicles (black curve, pH_i) and the outer solution (red curve, pH_o) and effect of the permeability coefficient of ammonia on the pH clock reaction. (b) Fraction of vesicles with n molecules of urease (Poisson distribution) and the equivalent enzyme concentration (U mL^{-1}) for a given volume of vesicles (where $\langle E \rangle = 55 \text{ U mL}^{-1}$, including empty vesicles, and a total vesicle volume of $1 \mu\text{L}$). (c) Synchronized switch in pH for each volume fraction of vesicles given in panel b. (d) Range of pH clock times for each volume fraction of vesicles given in panel b with reduced ammonia permeability and enzyme turnover number ($P_{\text{NH}_3} = 1 \times 10^{-11} \text{ m s}^{-1}$ and $k_{\text{cat}}' = k_{\text{cat}}/500$).

Simulations were performed with a distribution of the enzyme amount in the vesicles. On average, there was less than one enzyme molecule per vesicle (see SI 3.5.2). The probability of n molecules per vesicle was determined from a Poisson distribution ($P(X = n)$, $\lambda = 0.44$), and the equivalent enzyme concentration in units per milliliter was determined from the total number of enzyme molecules in a given volume of the vesicles (Figure 3b). The average enzyme concentration for the heterogeneous population (including vesicles with no enzyme) was $\langle E \rangle = 55 \text{ U mL}^{-1}$, and the clock time of $T_c = 9.6 \text{ min}$ was similar to that of the homogeneous population with $[E] = 55 \text{ U mL}^{-1}$ ($T_c = 9.3 \text{ min}$). Vesicles with different enzyme loadings in the heterogeneous population have the same clock time despite the differences in their internal pH and enzyme concentration (Figure 3c). The potential impact of heterogeneity on the reaction can only be determined if we reduce both the membrane permeability to ammonia and the enzyme turnover number ($k_{\text{cat}}' = k_{\text{cat}}/500$ to give the same average clock time of $\sim 9 \text{ min}$); then, a broad range of clock times can be observed (Figure 3d).

We also determined the influence of a distribution in enzyme loading and vesicle diameter on the pH clock reaction in the microvesicles. The vesicle volume fraction was $\phi = NV_i/V_o = 0.018$, and the number of enzyme molecules per vesicle was $\sim 10^5$ (SI 2.4). The encapsulation efficiency is generally assumed to be close to 100% using the droplet transfer method; however, significant differences in the macromolecular content have been reported.^{34,35} The simulations

were undertaken using the experimental probability mass function along with a normal distribution for the enzyme concentration to obtain a bivariate histogram (SI 3.6) with a range of $[E] = 60\text{--}100\text{ U mL}^{-1}$ and a diameter of $6\text{--}36\text{ }\mu\text{m}$ (Figure 4a). The membrane permeability to acetic acid (P_{HA}) was also included in these simulations.

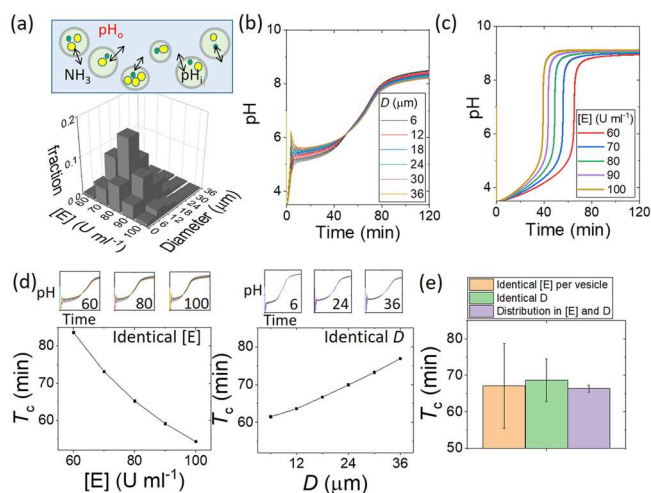


Figure 4. Effect of the urease concentration $[E]$ and the vesicle diameter D on the pH clock in simulations of a population of microvesicles with $[\text{pyranine}] = 50\text{ }\mu\text{M}$ in a solution of $[\text{urea}] = 80\text{ mM}$ and $[\text{acetic acid}] = 7.5\text{ mM}$ with a vesicle volume fraction $\phi = 0.018$. (a) Bivariate distribution in the enzyme concentration with $[E] = 80\text{ U mL}^{-1}$ and the vesicle diameter (D). (b) Synchronized switch in pH in vesicles with the distribution given in panel a. (c) Range of pH clock times in vesicles with both the distribution given in panel a and lower permeability and enzyme turnover number ($P_{\text{NH}_3} = P_{\text{HA}} = 1 \times 10^{-11}\text{ m s}^{-1}$ and $k_{\text{cat}}' = k_{\text{cat}}/50$). (d) Effect of $[E]$ or D on the clock time T_c in simulations with a population of vesicles with either identical $[E]$ per vesicle and the distribution in D given in panel a or identical D and the distribution in $[E]$ given in panel a. (e) Comparison of the average and standard deviation (error bars) in the clock time for simulations in a population of vesicles with identical $[E]$, identical D , or a distribution in both $[E]$ and D .

The change in pH in the vesicles is shown in Figure 4b. The profiles are similar to those observed in experiments; there was a rapid increase in the internal pH to ~ 5.5 , then a transition to high pH at around 50 min that was accompanied by an increase in pH in the surrounding solution as a result of the fast transport of ammonia into the outer solution (Figure 4b(ii)). The initial pH was influenced by both the enzyme content and the diameter, with a smaller diameter and smaller enzyme concentration favoring low pH (Figure S11). Again, the synchronized switch in pH did not occur if the permeability coefficient of ammonia or acid and the enzyme turnover number were reduced; in that case, a range of clock times was obtained (Figure 4c).

The effect of the concentration of the enzyme, $[E]$, on the pH clock with a high ammonia permeability was determined in simulations in which the enzyme concentration in each vesicle was identical but increased from 60 to 100 U mL^{-1} in separate runs. The clock time T_c decreased from 85 to 52 min as a result of the increased total amount of catalyst in the vesicles (Figure 4c). In simulations in which the diameter, D , of each vesicle was identical and increased from 6 to $26\text{ }\mu\text{m}$ in separate runs, the clock time increased from 60 to 80 min as a result of

the reduced rate of transport of ammonia to the external solution (Figure 4c). In Figure 4d, the average clock times are shown for all the simulations with identical $[E]$ or identical D (from Figure 4c), compared to the simulation with a distribution in both $[E]$ and D (from Figure 4b). The standard deviation in T_c is small for the population with a bivariate distribution, as the pH switch is governed by communication between vesicles via the ammonia in the surrounding solution rather than the internal enzyme concentration or the diameter of each vesicle.

In conclusion, we have shown that the pH–time profile and the synchronization of the pH clock in heterogeneous vesicles was controlled by the relatively fast transport and increase in the amount of ammonia in the external solution. The behavior was observed in nano- and microvesicles with different phospholipids and acids, thus demonstrating the universal nature of the response. In natural systems, micro-organisms such as bacteria and yeast use extracellular signaling to overcome population diversity and change behavior.^{52,53} Ammonia is important in cell–cell communication and the multicellular structures that form in yeast and bacterial colonies. It has been implicated in complex functionalities such as metabolic oscillations and colony survival.⁵⁴ Further control of the membrane permeability would provide a useful platform for the investigation of more complex collective behavior in populations of synthetic vesicles driven by acid or base changes.⁵⁵

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jpclett.2c00069>.

Preparation and analysis of the nanovesicles and microvesicles, including materials, urease encapsulation, calibration curve for pH, estimation of the number of vesicles, and kinetic data analysis with plots; modeling of the urease–urease reaction in vesicles, including the enzyme-catalyzed reaction, equilibria, transfer rates, equations and parameters; and model files (PDF)

■ AUTHOR INFORMATION

Corresponding Authors

Paul A. Beales – School of Chemistry and Astbury Centre for Structural Molecular Biology, University of Leeds, Leeds LS2 9JT, U.K.; orcid.org/0000-0001-9076-9793;

Email: P.A.Beales@leeds.ac.uk

Federico Rossi – Department of Earth, Environmental and Physical Sciences, University of Siena, 53100 Siena, Italy;

orcid.org/0000-0002-1854-532X;

Email: federico.rossi2@unisi.it

Annette F. Taylor – Chemical and Biological Engineering, University of Sheffield, Sheffield S1 3JD, U.K.; orcid.org/0000-0003-0071-8306; Email: A.F.Taylor@sheffield.ac.uk

Authors

Ylenia Miele – Department of Chemistry and Biology, University of Salerno, 84084 Fisciano, Salerno, Italy

Stephen J. Jones – School of Chemistry and Astbury Centre for Structural Molecular Biology, University of Leeds, Leeds LS2 9JT, U.K.

Complete contact information is available at:

<https://pubs.acs.org/10.1021/acs.jpclett.2c00069>

Author Contributions

#Joint first author.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

Y.M., F.R., and A.F.T. acknowledge the support through the COST Action CM1304 (Emergence and Evolution of Complex Chemical Systems). S.J. received an EPSRC studentship, and P.B. was supported by EPSRC Grant EP/M027929/1.

REFERENCES

- (1) Küchler, A.; Yoshimoto, M.; Luginbühl, S.; Mavelli, F.; Walde, P. Enzymatic reactions in confined environments. *Nat. Nanotechnol.* **2016**, *11*, 409–420.
- (2) Maiti, S.; Fortunati, I.; Ferrante, C.; Scrimin, P.; Prins, L. J. Dissipative self-assembly of vesicular nanoreactors. *Nat. Chem.* **2016**, *8*, 725–731.
- (3) Samad, A.; Sultana, Y.; Aqil, M. Liposomal drug delivery systems: An update review. *Curr. Drug Del.* **2007**, *4*, 297–305.
- (4) Tang, T. Y. D.; Cecchi, D.; Fracasso, G.; Accardi, D.; Coutable-Pennarun, A.; Mansy, S. S.; Perriman, A. W.; Anderson, J. L. R.; Mann, S. Gene-Mediated Chemical Communication in Synthetic Protocell Communities. *ACS Synth. Biol.* **2018**, *7*, 339–346.
- (5) Trantidou, T.; Fridin, M.; Elani, Y.; Brooks, N. J.; Law, R. V.; Seddon, J. M.; Ces, O. Engineering Compartmentalized Biomimetic Micro- and Nanocontainers. *ACS Nano* **2017**, *11*, 6549–6565.
- (6) Rideau, E.; Dimova, R.; Schwill, P.; Wurm, F. R.; Landfester, K. Liposomes and polymersomes: a comparative review towards cell mimicking. *Chem. Soc. Rev.* **2018**, *47*, 8572–8610.
- (7) Soldner, C. A.; Socher, E.; Jamali, V.; Wicke, W.; Ahmadzadeh, A.; Breiting, H. G.; Burkovski, A.; Castiglione, K.; Schöber, R.; Sticht, H. A Survey of Biological Building Blocks for Synthetic Molecular Communication Systems. *IEEE Communications Surveys and Tutorials* **2020**, *22*, 2765–2800.
- (8) Rampioni, G.; Leoni, L.; Stano, P. Molecular Communications in the Context of 'Synthetic Cells' Research. *IEEE Trans. NanoBiosci.* **2019**, *18*, 43–50.
- (9) Tomasi, R.; Noël, J. M.; Zenati, A.; Ristori, S.; Rossi, F.; Cabuil, V.; Kanoufi, F.; Abou-Hassan, A. Chemical communication between liposomes encapsulating a chemical oscillatory reaction. *Chemical Science* **2014**, *5*, 1854–1859.
- (10) Krajewska, B. Ureasas I. Functional, catalytic and kinetic properties: A review. *J. Mol. Catal. B: Enzym.* **2009**, *59*, 9–21.
- (11) Hu, G.; Pojman, J. A.; Scott, S. K.; Wrobel, M. M.; Taylor, A. F. Base-catalyzed feedback in the urea-urease reaction. *J. Phys. Chem. B* **2010**, *114*, 14059–14063.
- (12) Jagers, R. W.; Bon, S. A. F. Independent responsive behaviour and communication in hydrogel objects. *Materials Horizons* **2017**, *4*, 402–407.
- (13) Panja, S.; Boháčová, K.; Dietrich, B.; Adams, D. J. Programming properties of transient hydrogels by an enzymatic reaction. *Nanoscale* **2020**, *12*, 12840–12848.
- (14) Gao, N.; Li, M.; Tian, L.; Patil, A. J.; Pavan Kumar, B. V. S.; Mann, S. Chemical-mediated translocation in protocell-based micro-actuators. *Nat. Chem.* **2021**, *13*, 868–879.
- (15) Lvov, Y.; Antipov, A. A.; Mamedov, A.; Möhwald, H.; Sukhorukov, G. B. Urease Encapsulation in Nanoorganized Microshells. *Nano Lett.* **2001**, *1*, 125–128.
- (16) Muzika, F.; Růžicka, M.; Schreiberová, L.; Schreiber, I. Oscillations of pH in the urea-urease system in a membrane reactor. *Phys. Chem. Chem. Phys.* **2019**, *21*, 8619–8622.
- (17) Maity, I.; Sharma, C.; Lossada, F.; Walther, A. Feedback and Communication in Active Hydrogel Spheres with pH Fronts: Facile Approaches to Grow Soft Hydrogel Structures. *Angewandte Chemie - International Edition* **2021**, *60*, 22537–22546.
- (18) Madeira, V. M. C. Incorporation of urease into liposomes. *BBA - General Subjects* **1977**, *499*, 202–211.
- (19) Miele, Y.; Bánsági, T., Jr.; Taylor, A. F.; Stano, P.; Rossi, F. Engineering enzyme-driven dynamic behaviour in lipid vesicles. In *Advances in Artificial Life, Evolutionary Computation and Systems Chemistry*; Rossi, F., Mavelli, F., Stano, P., Caivano, D., Eds.; Communications in Computer and Information Science, Vol. 587; Springer International Publishing, 2016; pp 197–208.
- (20) Che, H.; Cao, S.; Van Hest, J. C. M. Feedback-Induced temporal control of "breathing" polymersomes to create self-adaptive nanoreactors. *J. Am. Chem. Soc.* **2018**, *140*, 5356–5359.
- (21) Wang, X.; Moreno, S.; Boye, S.; Wen, P.; Zhang, K.; Formanek, P.; Lederer, A.; Voit, B.; Appelhans, D. Feedback-Induced and Oscillating pH Regulation of a Binary Enzyme-Polymersomes System. *Chem. Mater.* **2021**, *33*, 6692–6700.
- (22) Miele, Y.; Medveczky, Z.; Holló, G.; Tegze, B.; Derényi, I.; Hörvölgyi, Z.; Altamura, E.; Lagzi, I.; Rossi, F. Self-division of giant vesicles driven by an internal enzymatic reaction. *Chemical Science* **2020**, *11*, 3228–3235.
- (23) Toiya, M.; Vanag, V. K.; Epstein, I. R. Diffusively coupled chemical oscillators in a microfluidic assembly. *Angew. Chem., Int. Ed. Engl.* **2008**, *47*, 7753–7755.
- (24) Tinsley, M. R.; Taylor, A. F.; Huang, Z.; Showalter, K. Emergence of collective behavior in groups of excitable catalyst-loaded particles: Spatiotemporal dynamical quorum sensing. *Phys. Rev. Lett.* **2009**, *102*, 158301.
- (25) Guzowski, J.; Gizynski, K.; Gorecki, J.; Garstecki, P. Microfluidic platform for reproducible self-assembly of chemically communicating droplet networks with predesigned number and type of the communicating compartments. *Lab Chip* **2016**, *16*, 764–772.
- (26) Gentili, P. L.; Giubila, M. S.; Germani, R.; Romani, A.; Nicoziani, A.; Spalletti, A.; Heron, B. M. Optical Communication among Oscillatory Reactions and Photo-Excitable Systems: UV and Visible Radiation Can Synchronize Artificial Neuron Models. *Angew. Chem., Int. Ed. Engl.* **2017**, *56*, 7535–7540.
- (27) Totz, J. F.; Rode, J.; Tinsley, M. R.; Showalter, K.; Engel, H. Spiral wave chimera states in large populations of coupled chemical oscillators. *Nat. Phys.* **2018**, *14*, 282–285.
- (28) Budroni, M. A.; Torbensen, K.; Ristori, S.; Abou-Hassan, A.; Rossi, F. Membrane Structure Drives Synchronization Patterns in Arrays of Diffusively Coupled Self-Oscillating Droplets. *J. Phys. Chem. Lett.* **2020**, *11*, 2014–2020.
- (29) Yoshikawa, K.; Aihara, R.; Agladze, K. Size-dependent Belousov-Zhabotinsky oscillation in small beads. *J. Phys. Chem. A* **1998**, *102*, 7649–7652.
- (30) Lawson, H. S.; Holló, G.; Horvath, R.; Kitahata, H.; Lagzi, I. Chemical Resonance, Beats, and Frequency Locking in Forced Chemical Oscillatory Systems. *J. Phys. Chem. Lett.* **2020**, *11*, 3014–3019.
- (31) Gorecki, J.; Gizynski, K.; Guzowski, J.; Gorecka, J. N.; Garstecki, P.; Gruenert, G.; Dittrich, P. Chemical computing with reaction-diffusion processes. *Philos. Trans A Math Phys. Eng. Sci.* **2015**, *373*, 20140219.
- (32) Weitz, M.; Kim, J.; Kapsner, K.; Winfree, E.; Franco, E.; Simmel, F. C. Diversity in the dynamical behaviour of a compartmentalized programmable biochemical oscillator. *Nat. Chem.* **2014**, *6*, 295–302.
- (33) Litschel, T.; Ramm, B.; Maas, R.; Heymann, M.; Schwill, P. Beating Vesicles: Encapsulated Protein Oscillations Cause Dynamic Membrane Deformations. *Angewandte Chemie - International Edition* **2018**, *57*, 16286–16290.
- (34) Stano, P.; De Souza, T. P.; Carrara, P.; Altamura, E.; D'Aguzzo, E.; Caputo, M.; Luisi, P. L.; Mavelli, F. Recent Biophysical Issues about the Preparation of Solute-Filled Lipid Vesicles. *Mechanics of Advanced Materials and Structures* **2015**, *22*, 748–759.
- (35) Saito, H.; Kato, Y.; Le Berre, M.; Yamada, A.; Inoue, T.; Yosikawa, K.; Baigl, D. Time-resolved tracking of a minimum gene

expression system reconstituted in giant liposomes. *ChemBioChem*. **2009**, *10*, 1640–1643.

(36) Bánsági, T.; Taylor, A. F. Role of differential transport in an oscillatory enzyme reaction. *J. Phys. Chem. B* **2014**, *118*, 6092–6097.

(37) Miele, Y.; Bánsági, T., Jr.; Taylor, A. F.; Rossi, F. Modelling Approach to Enzymatic pH Oscillators in Giant Lipid Vesicles. In *Advances in Bionanomaterials*; Piotta, S., Rossi, F., Concilio, S., Reverchon, E., Cattaneo, G., Eds.; Lecture Notes in Bioengineering; Springer International Publishing, 2018; pp 63–74.

(38) Straube, A. V.; Winkelmann, S.; Schütte, C.; Höfling, F. Stochastic pH Oscillations in a Model of the Urea-Urease Reaction Confined to Lipid Vesicles. *J. Phys. Chem. Lett.* **2021**, *12*, 9888–9893.

(39) Kano, K.; Fendler, J. H. Pyranine as a sensitive pH probe for liposome interiors and surfaces. pH gradients across phospholipid vesicles. *BBA - Biomembranes* **1978**, *509*, 289–299.

(40) Clement, N. R.; Gould, J. M. Pyranine (8-Hydroxy-1,3,6-pyrenetrisulfonate) as a Probe of Internal Aqueous Hydrogen Ion Concentration in Phospholipid Vesicles. *Biochemistry* **1981**, *20*, 1534–1538.

(41) Pautot, S.; Frisken, B. J.; Weitz, D. A. Production of unilamellar vesicles using an inverted emulsion. *Langmuir* **2003**, *19*, 2870–2879.

(42) Walde, P.; Cosentino, K.; Engel, H.; Stano, P. Giant Vesicles: Preparations and Applications. *ChemBioChem*. **2010**, *11*, 848–865.

(43) Markovic, V. M.; Bánsági, T.; McKenzie, D.; Mai, A.; Pojman, J. A.; Taylor, A. F. Influence of reaction-induced convection on quorum sensing in enzyme-loaded agarose beads. *Chaos* **2019**, *29*, 033130.

(44) Singh, A.; Marcoline, F. V.; Veshaguri, S.; Kao, A. W.; Bruchez, M.; Mindell, J. A.; Stamou, D.; Grabe, M. Protons in small spaces: Discrete simulations of vesicle acidification. *PLoS Comp. Biol.* **2019**, *15*, e1007539.

(45) Gabba, M.; Poolman, B. Physicochemical Modeling of Vesicle Dynamics upon Osmotic Upshift. *Biophys. J.* **2020**, *118*, 435–447.

(46) Missner, A.; Pohl, P. 110 years of the Meyer-Overton rule: Predicting membrane permeability of gases and other small compounds. *ChemPhysChem* **2009**, *10*, 1405–1414.

(47) Paula, S.; Volkov, A. G.; Van Hoek, A. N.; Haines, T. H.; Deamer, D. W. Permeation of protons, potassium ions, and small polar molecules through phospholipid bilayers as a function of membrane thickness. *Biophys. J.* **1996**, *70*, 339–348.

(48) Krajewska, B.; Ciurli, S. Jack bean (*Canavalia ensiformis*) urease. Probing acid–base groups of the active site by pH variation. *Plant Physiol. Biochem.* **2005**, *43*, 651–658.

(49) Lande, M. B.; Donovan, J. M.; Zeidel, M. L. The relationship between membrane fluidity and permeabilities to water, solutes, ammonia, and protons. *J. Gen. Physiol.* **1995**, *106*, 67–84.

(50) Lohse, B.; Bolinger, P. Y.; Stamou, D. Encapsulation efficiency measured on single small unilamellar vesicles. *J. Am. Chem. Soc.* **2008**, *130*, 14372–14373.

(51) Colletier, J.-P.; Chaize, B.; Winterhalter, M.; Fournier, D. Protein encapsulation in liposomes: efficiency depends on interactions between protein and phospholipid bilayer. *BMC Biotechnol.* **2002**, *2*, 9.

(52) Waters, C. M.; Bassler, B. L. Quorum sensing: Cell-to-cell communication in bacteria. *Annu. Rev. Cell Dev. Biol.* **2005**, *21*, 319–346.

(53) De Monte, S.; d'Ovidio, F.; Dano, S.; Sorensen, P. G. Dynamical quorum sensing: Population density encoded in cellular dynamics. *Proc. Natl. Acad. Sci. U. S. A.* **2007**, *104*, 18377–18381.

(54) Palkova, Z.; Janderova, B.; Gabriel, J.; Zikanova, B.; Pospisek, M.; Forstova, J. Ammonia mediates communication between yeast colonies. *Nature* **1997**, *390*, 532–536.

(55) Miller, A. J.; Pearce, A. K.; Foster, J. C.; O'Reilly, R. K. Probing and Tuning the Permeability of Polymersomes. *ACS Central Science* **2021**, *7*, 30–38.

Recommended by ACS

Development of Dual-Nanopore Biosensors for Detection of Intracellular Dopamine and Dopamine Efflux from Single PC12 Cell

Tao Zhao, Jian-Hui Jiang, *et al.*

NOVEMBER 04, 2022

ANALYTICAL CHEMISTRY

READ 

Identification of Activating Mutations in the Transmembrane and Extracellular Domains of EGFR

Anja Wagner, Michael W. Traxlmayr, *et al.*

SEPTEMBER 22, 2022

BIOCHEMISTRY

READ 

Cell-Free Synthesis Goes Electric: Dual Optical and Electronic Biosensor via Direct Channel Integration into a Supported Membrane Electrode

Zachary A. Manzer, Susan Daniel, *et al.*

JANUARY 18, 2023

ACS SYNTHETIC BIOLOGY

READ 

Flow-Cell-Based Technology for Massively Parallel Characterization of Base-Modified DNA Aptamers

Diana Wu, Hyongsok Soh, *et al.*

JANUARY 24, 2023

ANALYTICAL CHEMISTRY

READ 

Get More Suggestions >