

This is a repository copy of *Memory at your fingertips: how viscoelasticity affects tactile neuron signaling*.

White Rose Research Online URL for this paper: <u>https://eprints.whiterose.ac.uk/id/eprint/227905/</u>

Version: Preprint

### **Preprint:**

Saal, H.P. orcid.org/0000-0002-7544-0196, Birznieks, I. orcid.org/0000-0003-4916-1254 and Johansson, R.S. orcid.org/0000-0003-3288-8326 (Accepted: 2023) Memory at your fingertips: how viscoelasticity affects tactile neuron signaling. [Preprint - eLife Sciences Publications, Ltd] (Submitted)

https://doi.org/10.7554/eLife.89616.1.sa2

© 2023, Saal et al. This article is distributed under the terms of the Creative Commons Attribution License, (http://creativecommons.org/licenses/by/4.0/) which permits unrestricted use and redistribution provided that the original author and source are credited.

### Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

### Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.





#### **Reviewed Preprint**

Published from the original preprint after peer review and assessment by eLife.

#### About eLife's process

**Reviewed preprint posted** September 5, 2023 (this version)

Sent for peer review June 23, 2023

#### Posted to bioRxiv May 17, 2023

#### Neuroscience

# Memory at your fingertips: how viscoelasticity affects tactile neuron signaling

### Hannes P. Saal 🐸, Ingvars Birznieks, Roland S. Johansson

Active Touch Laboratory, Department of Psychology, University of Sheffield, Sheffield S1 2LT, UK • School of Biomedical Sciences, UNSW Sydney, Sydney, New South Wales, Australia • Neuroscience Research Australia, Sydney, New South Wales, Australia • Physiology Section, Department of Integrative and Medical Biology, Umeå University, SE-901 87 Umeå, Sweden

https://en.wikipedia.org/wiki/Open\_access
https://creativecommons.org/licenses/by/4.0/

# Abstract

Human skin and its underlying tissues constitute a viscoelastic medium, implying that any deformation depends not only on the currently applied force, but also the recent loading history. The extent to which this physical memory influences the signaling of first-order tactile neurons during natural hand use is not well understood. Here, we examined the effect of past loading on the responses of fast-adapting (FA-1) and slowly-adapting (SA-1 and SA-2) first-order tactile neurons innervating the human fingertip to loadings applied in different directions representative of object manipulation tasks. We found that variation in the preceding loading affected neurons' overall signaling of force direction. Some neurons kept signaling the current direction, while others signaled both the current and preceding direction, or even primarily the preceding direction. In addition, ongoing impulse activity in SA-2 neurons between loadings signaled information related to the fingertip's viscoelastic deformation state between loadings. We conclude that tactile neurons at the population level signal continuous information about the fingertip's viscoelastic deformation state, which is shaped by both its recent history and current loading. Such information might be sufficient for the brain to correctly interpret current force loading and help in computing accurate motor commands for interactions with objects in manipulation and haptic tasks.

### eLife assessment

The **fundamental** findings reported here provide insight into how the viscoelasticity of the fingertip skin influences the activity of mechanoreceptive afferents and thus the neural coding of force in humans. The basic principle studied was whether and to what extent the previous applied force directions impact the firing of FA-1, SA-1 and SA-2 neurons during the current applied force directions. The data and analyses are **compelling** and will be **important** for modeling the neural representations of force in the context of object grasping and manipulation.



# Introduction

To enable successful skilled object manipulation and haptic object exploration, the brain must have access to information related to the forces acting on the fingertips (Johansson and Westling, 1984 C; Westling and Johansson, 1984 C; Robles-De-La-Torre and Hayward, 2001 C). Experimental evidence indicates that populations of first-order tactile neurons with end-organs distributed throughout the fingertip skin provide sensory information about the distribution, magnitude, and direction of such fingertip forces (see Johansson and Flanagan, 2008 2, 2009 2, for reviews). However, these neurons do not signal contact forces per se, but local tissue deformations at the site of their receptor organs. This means that the relationship between fingertip forces and neuronal signaling can be very complex, since the deformation patterns resulting from a force applied to the fingertip depends on its geometry and its non-linear, viscoelastic and anisotropic material properties (Serina et al., 1997 C; Pawluk and Howe, 1999 C; Nakazawa et al., 2000 C; Jindrich et al., 2003 2; Pataky et al., 2005 2; Wang and Hayward, 2007 2). Concerning the viscoelastic properties in particular, the recent loading history of the fingertip might influence a neuron's response to a given contact force. That is, since tissue viscoelasticity causes deformation changes to lag force changes, residual deformations from previous loadings will affect how the fingertip reacts mechanically under a given loading, and thus affect the deformation changes to which neurons' receptor organs are exposed.

Despite indications in early animal studies that stimulation history via tissue viscoelasticity may indeed affect the responsiveness of first-order tactile neurons (Lind-blom, 1965<sup>•</sup>); Werner and Mountcastle, 1965<sup>•</sup>; Beitel et al., 1977<sup>•</sup>; Pubols, 1982<sup>•</sup>), the issue has subsequently received little attention in studies of tactile mechanisms. Because the viscoelasticity of human fingertips exhibits time constants of up to several seconds (D'Angelo et al., 2016<sup>•</sup>; Kumar et al., 2015<sup>•</sup>; Pawluk and Howe, 1999<sup>•</sup>; Wu et al., 2003<sup>•</sup>), we hypothesized that loading history would interfere with the signaling of first-order tactile neurons to rapidly fluctuating fingertip forces on a similar time scale to those experienced naturally (Kunesch et al., 1989<sup>•</sup>; Callier et al., 2015<sup>•</sup>; Morley et al., 1983<sup>•</sup>).

Here, we tested the effect of loading history on neural information transmission in human firstorder tactile neurons about the direction of fingertip forces during repetitive loadings that mimic those occurring during natural object manipulation. We examined information conveyed in the three types of neurons: fast-adapting type I (FA-1), slowly adapting type I (SA-1), and slowly adapting type II (SA-2) (Birznieks et al., 2001 **C**; Johansson and Birznieks, 2004 **C**; Birznieks et al., 2009 **C**; Saal et al., 2009 **C**). These neuron types most likely supply Meissner corpuscles, Merkel cell neurite complexes, and Ruffini-like end-organs, respectively. For neurons of all three types, we show that variations in loading history not only affect fingertip deformation, but also reduce information about the force direction in the prevailing loading. Although most neurons of each type continued to preferentially signal information about the current force direction, a minority signaled more information about the preceding force direction than the current one. For those SA-2 neurons that exhibit ongoing activity without external stimulation (Birznieks et al., 2009 **C**; Johansson, 1978 **C**; Knibestöl, 1975 **C**; Chambers et al., 1972 **C**), we found that they could signal information about the viscoelastic state of the fingertip even when unloaded.

# Results

We recorded action potentials in the median nerve of individual first-order human tactile neurons innervating the glabrous skin of the fingertip (Vallbo and Hagbarth, 1968 .). Sixty of the neurons were fast adapting type I (FA-1), 73 were slowly adapting type I (SA-1) and 41 were slowly adapting type II (SA-2) neurons (Vallbo and Johansson, 1984 .). The receptive fields of the neurons within each class were widely distributed over the glabrous skin of the distal phalanx (**Fig. 1A** .). The

# 🍪 eLife

fourth type of tactile neurons in the human glabrous skin, fastadapting type II neurons (FA-2) supplied by Pacinian corpuscles, were not considered because our stimuli did not contain mechanical events with frequency components high enough to reliably excite them. Fingertip forces were applied in 5 different directions with a flat surface (referred to as the contactor), which was always oriented parallel to the skin surface at the primary site of object contact in dexterous tasks, i.e., in the middle of the flat portion of the volar surface of the fingertip. Because forces were applied to a standardized site, the neurons could vary widely in their responsiveness depending on where in the mechanically complex fingertip their transduction sites were located. Forces were applied normal (N) to the skin and at 20 degrees to the normal direction in the radial (R), distal (D), ulnar (U), or proximal (P) direction, respectively (**Fig. 1B** <sup>C2</sup>). All force stimuli consisted of a force protraction phase (125 ms), a plateau phase at 4 N normal force (250 ms), and a force retraction phase (125 ms); inter-stimulus periods were 250 ms.

Each neuron was subject to two different sequences of force stimuli, first the 'regular sequence' and then the 'irregular sequence'. As stimuli were force controlled, the contactor's movement and position producing identical reactive force may differ depending on stimulation history, due to viscoelastic properties of the fingertip tissue. In the regular sequence, the five force directions were repeatedly presented in a fixed relative order (R, D, U, P, N), such that each loading in each direction received the same immediate stimulation history (**Fig. 2A** C<sup>\*</sup>). Differences in the path of the contactor between different force directions were clearly visible in the tangential plane, where the changes in the force stimulation took place between trials (**Fig. 2B** C<sup>\*</sup>). In the regular sequence, the contactor path was practically identical across the five test trials in each direction (**Fig. 2B** C<sup>\*</sup>), indicating that the fingertip deformed similarly during the repetitions. However, the contactor path deviated from the direction of the force due to anisotropic mechanical properties of the fingertip and, importantly, it differed between force protraction and retraction due to the viscoelasticity of the fingertip (**Fig. 2B** C<sup>\*</sup>). Likewise, the viscoelastic properties were reflected as a pronounced hysteresis between the force and the contactor position, creep during the force plateaus, and creep recovery during the interstimulus periods (**Fig. 2A** C<sup>\*</sup>).

In the irregular sequence, also including five trials in each force direction, the stimulation history for trials in each direction varied systematically, such that trials in each stimulation direction were preceded by loading in each of the five directions once (**Fig. 2C C**). As in the regular sequence, the contactor moved broadly in the direction of the force regardless of the previous loading direction, but its precise path differed markedly depending on the previous loading direction (**Fig. 2D C**). This variability was most evident during the protraction phase (**Fig. 2E C**) but could be discerned throughout the trial (**Fig. 2D C**). Hence, deduced from the contactor's behavior, these observations indicated that variations in the immediate loading history caused a greater intertrial variation in the fingertip deformation than an unchanging loading history. This is illustrated in **Fig. 3A C**, which shows the contactor path for both regular and irregular sequences from a single participant for five test trials in the distal direction as well as the corresponding preceding trials.

# Effects of loading history on neural responses

We asked whether the greater intertrial variability in fingertip deformation during the irregular stimulation sequence would be reflected in the responses of tactile neurons.

First, we observed greater intertrial variability in the firing rate profiles of the neurons in the irregular compared to the regular sequence during test trials (see examples in **Fig. 3B** <sup>□</sup> –**D** <sup>□</sup>). This was most evident during the force protraction phase where neurons tended to respond most intensely. The responses evoked during the irregular sequence generally showed greater intertrial temporal jitter and variation in firing rate. For SA-1 and SA-2 neurons, which typically generated nerve impulses also during the force plateau, and for FA-1 neurons, which often responded during force retraction, we noted a tendency for greater variability in the intensity of these responses as



### Figure 1.

### Experimental setup.

**A**. Receptive field center locations shown on a standardized fingertip for all first-order tactile neurons included in the study, divided by neuron type. **B**. The flat stimulus surface was centered at the standard site of stimulation and oriented such that its tangential plane was parallel to the flat portion of skin on the fingertip. The surface maintained contact with the skin at a force of 0.2 N in intertrial periods. Force stimuli were superimposed on this background contact force and were delivered in the normal direction (N), and at an angle of 20 degrees to the normal with tangential components in the distal (D), radial (R), proximal (P) and ulnar (U) directions, as indicated by the five arrows in the lower panel.



### Figure 2.

### Stimulation sequence exposes fingertip viscoelasticity.

**A**. Trial order for the entire regular sequence, which repeats fingertip loadings in five different force directions in a fixed order, implying that loadings in each direction received the same stimulation history. Force (red lines) and contactor position (black lines) are shown along the normal (*z*), distal/proximal (*y*), and ulnar-radial (*x*) axes, while recording action potentials from a single exemplary SA-1 neuron (bottom trace). **B**. Average contactor position in the tangential plane for all trials in the regular sequence across all recorded neurons (and thus fingertips). The colored segments of the curves indicate the protraction phase for each of the five force directions, while other phases of the fingertip loading (plateau, retraction) and the interstimulus period are shown in grey. Dashed lines show the directions in which the tangential force components were applied. **C**. Trial order for the entire irregular sequence, where force directions are varied such that trials in each stimulation direction were preceded by loading in each of the five directions once. Same neuron (and fingertip) as in A. **D**. Average contactor position in the tangential plane for the tangential plane at the start of (filles circles) and during the force protraction phase (colored lines) per force direction, referenced to the fingertip position at rest (gray marker). Same trials as in D, but different force directions are shown in separate panels for better visibility.



### Figure 3.

### Influence of preceding loading direction on fingertip deformation and neural responses.

A. Contact position along the x, y, and z axes (see **Fig. 1B** <sup>C</sup>) as a function of time super-imposed for all five trials with loading in the distal direction ('test trial') as well as the respective previous loading ('preceding trial') in the regular (left column) and the irregular (right column) sequence. Vertical dashed lines mark transitions between loading phases (Pr: protraction, Pl: plateau, Re: retraction phase, Int: intertrial period). The yellow shaded area indicates the protraction phase of the test trials. Each trace is colored according to the force direction of the previous loading. Data was recorded from a neuron whose response is shown in B. **B**. Dots (top) represent action potentials recorded from an FA-1 neuron for each trial, whose contactor movements are shown in A. The superimposed traces below represent the corresponding firing rate profiles, defined as the recip-rocal of the interval between subsequent action potentials. Color coding as in A. **C,D**. Exemplary responses of one SA-1 (C) and two SA-2 neurons (D) to force loadings corresponding to those in A. All neurons show higher variability in their firing rate profiles during test trials in the irregular compared to the regular sequence.

# 🍪 eLife

well. Finally, the variation in previous loading direction during the irregular sequence could also modify impulse activity in SA-2 neurons generated during interstimulus periods, the implications of which will be further addressed below.

We quantified the effect of the stimulation history on a neuron's response by first calculating for each stimulation sequence the time-varying standard deviation of the instantaneous firing rate during the test trials in each of the five force directions. Averaged across neurons and loading directions, firing rate variability during the force protraction phase was just over twofold higher for FA-1s in the irregular sequence compared to the regular sequence, almost twofold higher for SA-1 and about 70% higher for SA-2 neurons (**Fig. 4A** <sup>(2)</sup>). In addition to the protraction phase, variability was elevated in the irregular sequence for both SA types during the plateau phase and for FA-1 neurons during the retraction phase (all  $p_{corrected} \leq 0.001$ , paired Wilcoxon signed rank tests). FA-1 neurons did not respond during the plateau phase (see Fig. 4B 🗹 ), while both SA types responded only weakly during the retraction phase with no apparent difference between the sequences ( $p \ge 0.3$ ). Thus, in all phases where neurons responded reliably, their response variability in test trials increased when the force direction in the preceding trial varied. Notably, neurons' overall firing rates did not differ between the regular and irregular sequence when averaged over test trials in all force directions (Fig. 4B 22) other than for SA-1s during the protraction phase ( $p_{corrected}$  = 0.002), where the difference in firing rate was less than 1 imp/s. This suggested that the greater variability was linked to the stimulation history and not a change in the neurons' overall responsiveness.

We also analyzed the effect of stimulation sequence on the fingertip deformation, again by using the contactor behavior as a proxy. For each neuron examined and stimulation sequence, we calculated the time-varying standard deviation in the tangential plane of the contactor position and of its velocity across the five test stimuli in each of the five force directions. As results were similar across the different test directions, we then averaged these data to arrive at a single variability measure per neuron. Importantly, the variability in contactor position was markedly higher in the irregular than in the regular sequence (**Fig. 4C** , solid vs. dashed lines). The variability in the irregular sequence decreased over time and did so especially rapidly during the force protraction phase. Averaged across all neurons, at the beginning of the force protraction the variability in contactor position was about 20 times greater in the irregular than in the regular sequence. Variability was 7 times greater during the plateau phase and still about 4 times so at the end of the retraction phase. During all phases, including the interstimulus period before and after the test trial, the variability in the irregular sequence was significantly higher than in the regular sequence (*p<sub>corrected</sub>* < 0.001, paired Wilcoxon signed rank tests). The effect of stimulation sequence present even at the end of the force retraction phase indicates that the viscoelastic memory trace of the previous loading direction lingered to some extent even beyond the test trials. Notably, the time-varying variation in contactor position was virtually identical for data pertaining to each of the three neuron types (Fig. 4C <sup>C2</sup>), demonstrating that the differences in neural response behavior could not be explained by variability in the skin responses across the different experimental runs.

Regarding the effect of viscoelasticity variability on contactor velocity, we found that the variability in contactor velocity was significantly greater in the irregular than in the regular sequence during the initial interstimulus period, the force protraction phase, and the plateau phase (**Fig. 4D**  $\bigcirc$ , black curves;  $p_{corrected} < 0.001$ ), but not during the retraction phase or the subsequent interstimulus period (p > 0.4). However, even during the irregular sequence, the variability in contactor speed appeared rather modest compared to the absolute contactor speed (**Fig. 4D**  $\bigcirc$ , cf. black and gray curves), which likely primarily drove the dynamic neural response during the force protraction for all classes (cf. **Fig. 4B**  $\bigcirc$ ).



### Figure 4.

#### Increased variability in neural responses and fingertip deformations in the irregular sequence.

**A**. Standard deviation of neuronal instantaneous firing rates as a function of time during test trials in each loading direction in the regular (dashed lines) and irregular (solid lines) sequences. Data averaged across neurons of each type and all loading directions. Shaded areas indicate SEM. Vertical dashed lines mark transitions between loading phases as in **Fig. 3A**  $\square$  and stars indicate phases where there was a significant difference between regular and irregular sequences at *p* < 0.05. **B**. Average instantaneous firing rates for the regular and irregular sequence as a function of time. Dashed black lines indicate the force profile of the fingertip loading. Note that the average firing rates are almost identical in the regular and irregular sequence. **C**. Standard deviation of tangential (2D) contactor position for the regular and irregular sequences. **D**. Standard deviation of contactor velocities (black lines) and average contactor velocity (purple lines) for the regular and irregular sequences.



Taken together, the variable loading history in the irregular sequence affected the neurons' firing rates and most so during their dynamic responses elicited by the force protraction. Further, the effects on the neurons' responses reasonably matched the influence of the variation in previous force direction on the state of the fingertip deformation and its change during the test trials, which was also most pronounced during the force protraction phase.

# Information transmission about past and present loading

To assess whether the increased firing rate variability affected neural information transmission, we calculated a lower bound on the mutual information transmitted about both the current and the preceding force direction based on the neural spiking responses of individual neurons (see Methods). We focused on protraction phase only, during which firing rates and their variability were highest. Averaged across all neurons of each type, information about the current force direction tended to accumulate throughout the force protraction phase, as shown previously for FA-1 and SA-1 neurons (Saal et al., 2009 C). However, for all types, the rate of information increase was considerably lower and cumulative information tended to plateau at a much lower value in the irregular than in the regular sequence (compare black dashed with orange lines in **Fig. 5A** C). At the end of the protraction phase, both FA-1 and SA-1 neurons signaled on average only 50% of the information in the irregular compared to the regular sequence (p < 0.001 for both types, paired Wilcoxon signed rank tests). For the SA-2 neurons the corresponding information transfer was reduced by only 20% (p = 0.02). However these neurons signaled far less information about force direction to begin with. As a result, average information transmission in the three classes of neurons ended up comparable in the presence of viscoelastic effects.

We also assessed information about the preceding force direction contained in the neural responses during the protraction phase in the irregular sequence. We reasoned that if the preceding loading systematically affected contactor position in the subsequent trial, then neurons might carry information about past stimulation in their responses. Consistent with this idea, such information was present, albeit at a relatively low level. Averaged across all neurons of each type, information about the preceding force direction increased for about 60–70 ms into the protraction phase after which it appeared to plateau or decrease (yellow traces in **Fig. 5A**  $\square$ ; p < 0.001 for each type, Wilcoxon one-sample signed rank tests based on information values halfway through the protraction phase compared against zero information). Irrespective of neuron type, signaling of the preceding as well as the current force direction could vary substantially between neurons, with some carrying information mostly about the current direction, and others about the preceding one (see example information traces for individual neurons in **Fig. 5B** <sup>C</sup>). We quantified the diversity amongst neurons in this respect based on whether they primarily conveyed information about the present force direction, the preceding force direction, or a mix of both (including 53 out of 67 FA-1, 67 out of 73 SA-1 and 35 out of 41 SA-2; see Methods). Most neurons primarily signaled information about the current force direction (46 SA-1, 35 FA-1 and 18 SA-2 neurons, see **Fig. 5C** <sup>(2)</sup>). Fewer showed mixed tuning and those that did signaled preceding force direction early during the protraction phase and then switched to information about the current loading direction (16 SA-1, 9 FA-1 and 13 SA-2). Finally, some neurons (9 FA-1, 4 SA-1 and 4 SA-2) primarily signaled information about preceding force direction. We found no significant differences in the relative frequency of FA-1, SA-1 and SA-2 neurons that responded to the current or previous stimulation, or both ( $\chi^2$ (3, 155) = 6.95, p = 0.14). Notably, information transmission in both FA-1 and SA-1 neurons decreased between the regular and irregular sequence even when only considering those neurons that predominantly signaled information about the current force direction (Fig. 5D <sup>C2</sup>), confirming that information transmission about the ongoing stimulus was still affected by the fingertip's viscoelastic memory (p < 0.001 for both types, p = 0.18 for SA-2, paired Wilcoxon signed rank tests).

Our findings indicate that past loading reduces the information conveyed about the direction of prevailing fingertip forces in these neurons. However, responses in neurons of all three types can carry information about the previous force direction, though with considerable heterogeneity



### Figure 5.

### Information about current and previous force direction during the force protraction phase.

**A**. Average mutual information about force direction for FA-1 (left), SA-1 (middle), and SA-2 (right) neurons as a function of time during the protraction phase in the irregular sequence. Information is shown for the current trial (solid orange line) or the preceding trial (solid yellow line) and is compared to the regular sequence (dotted black line). Grey dashed lines denote the stimulus force profile. **B**. Examples of mutual information curves (top) and spike trains (bottom) for three individual neurons with different response behaviors. Information curves as in A. Spike trains are split by current force direction with spikes colored by previous force direction. Examples are of a neuron with mixed tuning (left), a neuron that predominantly signals information about the previous stimulus (middle), and a neuron that primarily signals information about the current force direction (right). **C**. Proportion of FA-1 (blue), SA-1 (green), and SA-2 (purple) neurons showing different response behaviors during the protraction phase. Most neurons of either class signaled predominantly the current force direction, but around 35% of neurons either signaled the previous force direction or showed mixed response behavior. **D**. Information transmitted about force direction for neurons tuned to the current force direction for the regular and the irregular sequence. Information decreases considerably for the irregular sequence, even in neurons responding strongly to the current direction.



among individual neurons.

# Neural responses in the absence of loading

SA-2 neurons can exhibit ongoing activity without external stimulation and sense tension states in collagenous fiber strands in dermal and subdermal tissues (Knibestöl, 1975 🖒; Birznieks et al., 2009 🖒; Johansson, 1978 Ć); Chambers et al., 1972 Ć). Since the deformation of the fingertip by the force stimuli was mostly absorbed by such tissues, we hypothesized that SA-2 neurons active during the interstimulus periods might convey ongoing information about the viscoelastic state of the fingertip during the recovery from the recurrently applied loadings. A subset of our SA-2 neurons (20 out of 41) exhibited such activity (see Fig. 6A Č) for three examples). Calculation of mutual information indicated that some individual neurons were highly informative about the preceding stimulus direction, but the time of maximal information transmission could occur at different points (see examples in Fig. 6B C). On average, SA-2 neurons provided low but continuous information about the preceding force direction throughout the interstimulus period, which was highest at the start of the interstimulus period and tended to decrease slightly over time (Fig. 6C C), yellow trace). This decrease was likely driven by the gradual relaxation of the fingertip.

We next asked whether SA-2 neural activity at the population level could track the deformation state of the fingertip during the interstimulus period. We calculated a low-dimensional representation of the SA-2 population activity to compare with the fingertip deformation at three different time points: in the middle and at the end of the inter-stimulus period, and again at the end of the protraction phase. Specifically, based on activity recorded during the irregular sequence we calculated pairwise spike distances across all 25 trials (5 force directions × 5 trials) for each neuron, providing a measure for how distinct the activity of this neuron was across different trials. This process yielded a matrix, which was averaged across all active SA-2 neurons. We then used multi-dimensional scaling to place each of the 25 types of trials into a twodimensional space. Using Procrustes analysis, finally we rotated and scaled the responses to match the recorded tangential contactor positions, which were calculated for the same 25 types of trial at the corresponding time points and averaged across fingertips (Fig. 7A<sup>CC</sup>). Notably, there was a good match between the neural representation and the fingertip deformation. For both measures, different trial types were clustered according to the force direction of the preceding loading throughout the interstimulus period, but cluster separation decreased as time progressed. Then trial types diverged during the protraction phase, and at the end of the phase they were clustered according to the current force direction. To quantify the similarity of the two representations, for each we calculated two measures. The 'total variance' across all 25 trial types indicates the general level of variability (see black dashed ellipse in the top panel of **Fig. 7A** <sup>C</sup>). The 'direction variance', which was calculated over trials representing the same preceding force direction and then averaged, indicates variability within clusters of trials with the same preceding direction (see dashed orange ellipse in the top panel of **Fig. 7A** <sup>(2)</sup>). Skin positions and SA-2 population activity displayed a similar variance pattern (Fig. 78 <sup>C2</sup>). During the interstimulus period the total variance was much larger than the direction variance, signifying a marked clustering according to the preceding force direction, but this difference tended to decrease with time. In contrast, both variances were large and roughly equal at the end of the protraction phase, indicating little clustering based on the preceding force direction. Taken together, these results suggest that SA-2 neurons can continuously signal the mechanical state of the fingertip even in the absence of fingertip loading.



### Figure 6.

### SA-2 neurons signal information about previous forces in the absence of loading.

**A**. Spike raster plots for three SA-2 neurons recorded during the interstimulus periods of the irregular sequence. Each dot represents an action potential, and the colors indicate the preceding force direction. Note that the responses differ systematically based on the previous force direction. Bottom panel: To illustrate the effect of previous force direction on the deformation state of the fingertip during the interstimulus period, the average contactor position in the ulnar-radial (U-R) direction during the corresponding irregular stimulation sequences is shown (on the same time scale as upper panels). Colors indicate the force direction in the preceding trial corresponding to the color coding in A. **B**. Average mutual information about force direction in the test trial (orange line) or the preceding trial (yellow line) during the interstimulus period and during the subsequent protraction phase for the same three neurons shown in panel A. **C**. Average mutual information across all SA-2 neurons active in the absence of load during the interstimulus period.



### Figure 7.

### Continuous representation of fingertip viscoelastic state in the SA-2 population.

**A**. Colored dots indicate tangential contactor positions (top) and their representation in the SA-2 population signal (bottom) at three different times: the middle of the interstimulus period (–0.125 s), the end of the interstimulus period (0 s) and the end of the subsequent protraction phase (0.125 s). The colors of the markers denote the force direction in the directly preceding trial. Contactor position is the two-dimensional position in the tangential plane. SA-2 representations are derived from average spike trains distances across the different trials, visualized in a two-dimensional space using multidimensional scaling and aligned with the contactor positions using Procrustes analysis (see Methods for details). **B**. Averaged total variance and within-direction variance for contactor position and SA-2 population signal representations at the same three time points as in A. As illustrated by the dashed ellipses in A, total variance denotes the two-dimensional variance across all trials, while within-direction variance denotes the variance for trials belonging to the same preceding force direction. Higher total than within-direction variance indicates that data for trials in the same preceding direction are more clustered than data for trials in all preceding directions, which is required for discrimination of preceding force direction.



# **Discussion**

We found that the viscoelasticity of the fingertip affects signals in first-order tactile neurons when responding to fingertip loadings mimicking those experienced in everyday object manipulation tasks. Such tasks involve applying forces of different magnitudes and directions in rapid succession, such as during grasping and transporting objects, handicraft, cooking, cleaning, and food gathering. The neurons' signaling of force direction was significantly influenced by the direction of the preceding loading, even if it varied by only 20 degrees relative to the perpendicular direction. This effect was most prominent during the force protraction phase, which is when neurons typically signal the most information about force direction. However, there was heterogeneity in how individual neurons behaved. Some neurons primarily signaled information about the current direction, while others signaled both the current and previous direction, or even primarily the preceding direction. Our results also indicate that neurons can signal information related to the fingertip's viscoelastic deformation state even between loadings: SA-2 neurons' ongoing impulse activity between loadings was influenced by the previous loading direction. This diversity suggests that, at the population level, first-order neurons carry information about the fingertip's current viscoelastic state, which within it contains a memory of past stimulation.

The observed heterogeneity between neurons of the same type is expected, given that their transduction sites were distributed widely within the fingertip skin and they are sensitive to the local stresses and strains at their transduction site, rather than the global deformation of the fingertip (Birznieks et al., 2001 , Saal et al., 2009 ) Thus, the viscoelastic memory of the preceding loading would have modulated the pattern of strain changes in the fingertip differently depending on where their receptor organs are situated in the fingertip.

We also observed some differences between neuron types regarding how variation in the preceding force direction affected their signaling during the current loading. The impact of stimulation history during the force protraction phase was more pronounced in FA-1 and SA-1, compared to SA-2 neurons. It is possible that the neurons' response properties accounted for this difference: type I neurons are primarily sensitive to deformation of the fingerprint ridges (Sukumar et al., 2022 C; Jarocka et al., 2021 C), while SA-2 neurons primarily signal tension states in deeper dermal and subdermal tissues (Knibestöl, 1975 <sup>™</sup>; Birznieks et al., 2009 <sup>™</sup>; Johansson, 1978 **C**; Chambers et al., 1972 **C**). The fingertip deformation changes during the loadings can be seen as twofold. First, the tangential force component of the contactor caused bulk deformation changes of the fingertip that depended on the force direction. This is because the friction between the stimulation surface and the skin was high enough to prevent the contactor from sliding over the fingertip. Bulk deformation changes are closely linked to widespread alterations in stress and strain distribution in deeper tissues, making the SA-2 neuron population the preferred signaling source for such changes. For example, it has been demonstrated that experimentally induced compliance changes of the finger pulp using venous occlusion readily influenced activity of SA-2, but not SA-1 neurons (Hudson et al., 2015 2). However, concurrent with the bulk deformation changes, the fingertip skin would undergo direction dependent surface deformations changes involving planar tensile strain changes and partial slippage peripherally within the contact surface (Delhaye et al., 2014 2, 2016 2; Willemet et al., 2021 2). Changes in planar tensile strain per se might excite neurons of either type, given that even neurons terminating outside the contact area can respond to fingertip loadings (Bisley et al., 2000 ℃; Birznieks et al., 2001 ℃). However, partial slippages occurring during fingertip loadings, where some parts of the fingertipobject interface slip while others remain stuck, excite SA-1 and especially FA-1 neurons most intensely (Johansson and Westling, 1987 C; Srinivasan et al., 1990 C; Khamis et al., 2014 C; Delhaye et al., 2021 🔼). A substantial part of the sensitivity of FA-1 and SA-1 neurons to stimulation history could therefore be attributed to the preceding force direction affecting the location and timing of partial slips. That is, due to the viscoelastic memory of the fingertip, previous loadings in different directions could result in different patterns of planar tensile stress changes under a



given loading condition, which would affect how and where the local partial slips occur. The responsiveness of type I neurons to partial slippage might also explain their apparently higher sensitivity to stimulation history compared to SA-2s. In sum, we believe that both bulk and superficial deformation changes play a role in the activation of the tactile neurons during the fingertip loadings.

That SA-2 neurons signaled the viscoelastic state of the fingertip in periods between loadings is consistent with previous ideas that SA-2 neurons continuously measure stresses in collagen fiber strands that run within and between dermal and subdermal tissues (Vallbo and Johansson, 1984 C; Chambers et al., 1972 C; Birznieks et al., 2009 C). Indeed, their relatively low dynamic sensitivity to externally applied loads and the well sustained response to maintained loadings suggest that SA-2 neurons are tailored to encode slow viscoelastic and quasistatic events occurring in dermal and subdermal tissues (Westling and Johansson, 1987 C; Birznieks et al., 2001 C, 2009 2). Furthermore, even in the absence of externally applied stimulation, they can exhibit ongoing impulsive activity, which suggests they are capable of monitoring inherent mechanical tension patterns in dermal and subdermal tissues in the skin in the unloaded state (Knibestöl, 1975 C; Johansson, 1978 C). In particular, changes in the tension patterns via finger and hand movement without external stimulation can also modulate this activity. We believe that by constantly transmitting information related to tissues' viscoelastic state, SA-2 neurons could help keep the brain updated about the current mechanical state of body parts. In agreement with this view, peripheral nerve blocks affect the perceived image of body parts such as the arm and fingers (Inui et al., 2011 2; Walsh et al., 2015 2; Melzack and Bromage, 1973 2). Likewise, although sensations elicited by electrical stimulation of single SA-2 neurons innervating the hand have been elusive (Kunesch et al., 1995 2; Ochoa and Torebjörk, 1983 2), a recent study indicates that they can give rise to distinct sensations that include the experience of diffuse skin deformation (Watkins et al., 2022 C). By continuously informing about viscoelastic state of the fingertips, SA-2 neurons could help the brain in computing accurate motor commands for interactions with objects in manipulation and haptic tasks by updating reference frames for interpreting information signaled by type I neurons.

Although tactile information in general is crucial for planning and executing motor actions in such tasks, to our knowledge a possible influence of fingertip viscoelasticity on task performance has not been systematically investigated. Therefore, it is unclear whether viscoelasticity limits performance or if it is compensated for in some way.

Our findings indicate that the population of tactile neurons that innervate a fingertip encode continuous information about the fingertip's viscoelastic deformation state. This information could potentially aid the brain in managing the effects of viscoelasticity on tactile coding and fingertip actions. For instance, the brain could intermittently use this information to estimate the state of the fingertip during planning and evaluation of tactile-based actions (cf. Johansson and Flanagan, 2009 C2). It is also conceivable that the brain continuously represents the current deformation state of the fingertips using online population information. However, such processing might require considerable computational resources.

# **Materials and Methods**

# General procedure and study participants

Our study is based on data obtained from 21 females and 12 males (19-30 years of age) who participated after providing written informed consent in accordance with the Declaration of Helsinki. The Umeå University ethics committee approved the study. The general experimental methodology, procedure and apparatus have been described previously (Birznieks et al., 2001 🖒), as well as other aspects of the same experimental data than those analyzed here (Birznieks et al.,



2001 C; Jenmalm et al., 2003 C; Saal et al., 2009 C; Johansson and Birznieks, 2004 C). Briefly, action potentials (spikes) in axons of single first-order tactile neurons that terminated in the distal segment of the index, middle or ring finger were recorded with tungsten needle electrodes inserted into the median nerve at the level of the upper arm 0.5-0.6 m from the fingertips (Vallbo and Hagbarth, 1968 C). For neurons with cutaneous receptive fields on the distal segment of a finger, force stimuli were applied to its fingertip in five different directions by means of a custom-built robot. The fingertip was stabilized by gluing the nail to a firmly fixed metal plate. Force was transferred through a circular plane (30 mm diameter) that was centered on the midpoint of a line extending in the proximal-distal direction from the papillary whorl to the distal end of the finger. The stimulating surface was oriented parallel to the skin at this primary contact site (see Fig. 1B C), which was located approximately at the center of the flat part of the fingertip's volar surface and serves as a primary target for object contact in fine manipulation tasks engaging 'tip-to-tip' precision grips (Christel et al., 1998 C).

# Force stimuli

### **Force parameters**

One of the five directions of force stimulation was normal (N) to the skin surface at the primary contact site and the other four were angled 20<sup>°</sup> to the normal direction in the radial (R), distal (D), ulnar (U), and proximal (P) directions, respectively. All force stimuli were superimposed on 0.2 N background force normal to the skin and consisted of a force protraction phase (125 ms), a plateau phase at 4 N normal force (250 ms), and a force retraction phase (125 ms) (Fig. 1C<sup>2</sup>). In the four trials with a tangential force component, the tangential force was 1.4 N at the force plateau. The time course of the force changes followed a half-sinusoid (sine wave frequency of 4 Hz). The position and orientation of the stimulation surface in relation to the primary contact site was maintained when the loading contained tangential force components. That is, the friction between the stimulation surface and the skin was high enough to prevent frictional slips. The interval between successive fingertip loadings was 250 ms. Thus, the frequency of recurrent fingertip loadings (1.3 Hz) was representative of the frequency at which tactile events follow each other during dexterous object manipulation tasks (see e.g. Draper, 1994<sup>™</sup>; Kunesch et al., 1989<sup>™</sup>; Teulings and Maarse, 1984 2). Likewise, in trials with a tangential force component, the magnitudes, directions and time courses of the fingertip forces were similar to those employed when people use a precision grip to lift an object weighing 250-300 g (Johansson and Westling, 1984<sup>C</sup>; Westling and Johansson, 1984<sup>C</sup>).

### **Stimulation sequences**

Two force stimulation sequences containing trials in each of the five different directions were delivered repeatedly. In the regular sequence, the trial order was the same (R, D, U, P, N) for all repetitions of the sequence. The regular sequence was presented 6 times, but to standardize the stimulation history for all trials only data obtained during the last five repetitions was included in the analysis (5 force directions × 5 trials). Immediately after completion of the regular sequence, the irregular sequence was delivered in which the trial order was systematically changed over repeats of the sequence. Each of the five loading directions (R, D, U, P and N) were presented five times in such a way that each loading was preceded once by loading in each of the five directions (5 force directions × 5 trials).

# **Neural sample**

The neurons recorded from were classified as fast-adapting type I (FA-1), fast-adapting type II (FA-2), slowly-adapting type I (SA-1), and slowly-adapting type II (SA-2) according to criteria described previously (Johansson and Vallbo, 1983 ; Vallbo and Johansson, 1984 ). Briefly, FA afferents respond only to changes in skin deformation, whereas SA afferents show an ongoing response



during periods of static skin deformation. Type I afferents (FA-1 and SA-1) possess small and welldelineated receptive fields if probed by light, pointed skin indentations, while the receptive fields of type II afferents (FA-2 and SA-2) are often large and poorly defined (see <u>Vallbo and Johansson</u>, 1984<sup>22</sup>, for further details).

The present analysis included 60 FA-1, 73 SA-1, and 41 SA-2 neurons terminating in the glabrous skin of the terminal segment of digits II, III or IV, belonging to a larger sample of 196 neurons (73 SA-1, 72 FA-1, 41 SA-2 and 10 FA-2 neurons) analyzed previously by Birznieks et al. (2001) . This sample was intentionally biased towards slowly adapting neurons to obtain a reasonably large number of SA-1 and SA-2 neurons, which exhibit a lower density in the fingertips than the FA-1 afferents (Johansson and Vallbo, 1979 .

# Analysis

### Statistics

As mechanical and neural measures were generally not normally distributed, we used nonparametric tests for all statistical analyses. Specifically, for all analyses comparing the regular and irregular sequences, we used paired Wilcoxon signed rank tests. When making multiple comparisons within the same analysis, we used Bonferroni corrections, and the resulting p values are reported as  $p_{corrected}$ .

# Quantifying mechanical and neural response variability

We assessed the variability in displacements and velocities by calculating the standard deviation of the displacement and velocity signals at each time point over the two-dimensional tangential plane across repeated trials of the same force direction, using Euclidean distance as the metric. Variability was assessed separately for trials in the regular and the irregular sequence and averaged across all fingertips recorded from. Similarly, when assessing the variability of the neural responses, we calculated the standard deviation of the instantaneous firing rate across trials in the same force direction over time. Instantaneous firing rate was calculated as the inverse of the inter-spike interval at a given time point. Again, this analysis was run separately on data from the regular and the irregular sequence. The resulting standard deviation traces were first averaged over different force directions and then over all afferents from the same class.

# **Calculation of information transmission**

To assess the amount of information about force direction conveyed in responses of individual neurons, for each neuron we calculated a lower bound on the information transmitted about both the current and the previous force direction. We used metric space analysis (Victor and Purpura, 1996 C), which employs a classifier on the neural response data to distinguish different force directions, calculates a confusion matrix from several runs of the classifier, and finally computes a lower bound on the mutual information from the confusion matrix. Details of the specific implementation used in the present study have been described previously (Saal et al., 2009 C.). In short, we first sorted the trial data either by the force direction in the current trial (to assess coding of current force direction) or by the force direction in the previous trial (to assess whether neurons responded to the viscoelastic 'memory' of the previous trial). We then computed spike distances between all pairs of spike trains according to a spike-timing based distance metric that assesses the 'cost' for transforming the first spike train in the pair into the second, by adding or removing individual spikes, or shifting existing ones in time. For analyses of the protraction phase this spike distance was computed at a temporal resolution of 8 ms, as this was determined in a previous study to be close to the optimum for maximal information transmission for the present experimental data (Saal et al., 2009 🖆 ). For the analysis of SA-2 responses during the interstimulus period, we used a lower temporal resolution of 32 ms instead, to adjust for the lower firing rates of



these neurons in the absence of stimulation. Each individual trial was then classified as originating from the force direction to which its average distance was lowest. In this way, a confusion matrix was generated for each neuron, with each entry denoting the number of times a neural response from a given force direction was classified as having originated from that direction or another one. Finally, we used the confusion matrix to compute a lower bound of the mutual information. To quantify the bias of this estimate, we reassigned neural responses from individual trials to random conditions and recalculated the mutual information. The bias term was set as the average of the outcome of 10 such random assignments and subtracted from the estimate of the mutual information. This analysis was run on successively longer time windows starting at stimulus onset (when calculating information during the protraction phase) or at the start of the interstimulus period (when calculating information for this period), respectively, and extending until the end of the considered time window in 5 ms increments. In this way, it could be assessed how neural coding of force direction evolved over time. The whole analysis was run twice on the irregular sequence data: once with stimuli grouped according to the current force direction (to assess the information conveyed about the stimulus), and a second time with stimuli grouped according to the preceding trial (to assess information conveyed in the neural responses about the past stimulus). The analysis was also run on the regular sequence; because effects due to previous and current stimulation cannot be distinguished in this data set due to the fixed trial order, this analysis yielded a single information value that represents the total amount of information available when past loading is held constant.

For the analysis covering the protraction phase, we also assigned each neuron to one of three groups, based on their information curves: those that primarily conveyed information about the current force direction ('current'), those that conveyed information about the preceding force direction ('previous'), and those that conveyed different types of information at different times during the protraction phase ('mixed'). To do this, we first excluded neurons that responded too weakly or erratically, as assessed by whether they conveyed information (above 0) about either past or present force direction for at least 10 different time windows during the protraction phase. This left 53 FA-1, 67 SA-1, and 35 SA-2 neurons. For each time window containing non-zero information, we then measured whether more information was conveyed about the present or the past force direction. Neurons that conveyed information about the current force direction in 70% or more of time windows were classed as 'current'; neurons with 30% or less were classed as 'previous'; and all other neurons were classed as 'mixed'.

### Analysis of SA-2 population responses

To derive a representation of the SA-2 population response, we took the spike train distances (see above) calculated at three different time points (in the middle and at the end of the interstimulus period at -0.125 s and 0 s relative to force onset, and at the end of the protraction phase at 0.125 s) for the irregular sequence and summed these distances across all 20 SA-2 neurons that were tonically active during the interstimulus period. This yielded a 25-by-25 matrix, with each entry denoting how dissimilar a given trial in the irregular sequence was from another one on the population level. We then employed multidi-mensional scaling, which aims to embed data points in a high-dimensional space such that their Euclidean distances adhere to those in the original distance matrix as closely as possible. We retained the first two dimensions of this space, resulting in each trial now occupying a position in a two-dimensional space. Finally, using Procrustes analysis we rotated and scaled this representation to match it to the two-dimensional skin deformations in the plane tangential to the fingertip surface at the three different time points. The main behind this analysis is that if time-varying skin deformations are encoded at the SA-2 population level, then more dissimilar skin deformations should lead to more dissimilar neural responses. The analysis tests the extent to which this idea is true.



# Acknowledgements

We would like to thank Dimitrios Dimitriou for suggesting the title phrase, and the members of the Active Touch Lab for feedback on the presentation of the results. This manuscript was typeset using the Royle Lab bioRxiv template available at *https://github.com/roylelab/manuscript* -templates



# References

Beitel R. E., Gibson J. M., Welker W. I. (1977) Functional development of mechanoreceptive neurons innervating the glabrous skin in postnatal kittens *Brain Res* **129**:213–226

Birznieks I., Jenmalm P., Goodwin A. W., Johansson R. S. (2001) **Encoding of direction of fingertip forces by human tactile afferents** *J. Neurosci* **21**:8222–8237

Birznieks I., Macefield V. G., Westling G., Johansson R. S. (2009) **Slowly adapting** mechanoreceptors in the borders of the human fingernail encode fingertip forces *J. Neurosci* **29**:9370–9379

Bisley J. W., Goodwin A. W., Wheat H. E. (2000) **Slowly adapting type I afferents from the sides and end of the finger respond to stimuli on the center of the fingerpad** *J. Neurophysiol* **84**:57–64

Callier T., Saal H. P., Davis-Berg E. C., Bensmaia S. J. (2015) **Kinematics of unconstrained tactile texture exploration** *J. Neurophysiol* **113**:3013–3020

Chambers M. R., Andres K. H., von Duering M., Iggo A. (1972) **The structure and function of the slowly adapting type II mechanoreceptor in hairy skin** *Q. J. Exp. Physiol. Cogn. Med. Sci* **57**:417–445

Christel M. I., Kitzel S., Niemitz C. (1998) **How precisely do bonobos (pan paniscus) grasp small objects?** *Int. J. Primatol* **19**:165–194

D'Angelo M. L., Caldwell D. G., Cannella F. (2016) **Fingertip recovery time depending on viscoelasticity** *In Haptics: Perception, Devices, Control, and Applications* :33–44

Delhaye B., Lefèvre P., Thonnard J.-L. (2014) **Dynamics of fingertip contact during the onset of tangential slip** *J. R. Soc. Interface* 

Delhaye B., Barrea A., Edin B. B., Lefèvre P., Thonnard J.-L. (2016) **Surface strain** measurements of fingertip skin under shearing *J. R. Soc. Interface* **13**:20150874–20150874

Delhaye B. P., Jarocka E., Barrea A., Thonnard J.-L., Edin B., Lefèvre P. (2021) **Highresolution imaging of skin deformation shows that afferents from human fingertips signal slip onset** *Elife* **10** 

Draper J. V. (1994) **H and acceleration impulse bandwidth during target acquisition: implications for teleoperator bandwidth requirements** *IEEE Trans. Syst. Man Cybern* **24**:931–936

Hudson K. M., Condon M., Ackerley R., McGlone F., Olausson H., Macefield V. G., Birznieks I. (2015) Effects of changing skin mechanics on the differential sensitivity to surface compliance by tactile afferents in the human finger pad *J. Neurophysiol* 

Inui N., Walsh L. D., Taylor J. L., Gandevia S. C. (2011) **Dynamic changes in the perceived posture of the hand during ischaemic anaesthesia of the arm** *J. Physiol* **589**:5775–5784



Jarocka E., Pruszynski J. A., Johansson R. S. (2021) Human touch receptors are sensitive to spatial details on the scale of single fingerprint ridges *J. Neurosci* **41**:3622–3634

Jenmalm P., Birznieks I., Goodwin A. W., Johansson R. S. (2003) **Influence of object shape on responses of human tactile afferents under conditions characteristic of manipulation** *Eur. J. Neurosci* **18**:164–176

Jindrich D. L., Zhou Y., Becker T., Dennerlein J. T. (2003) **Non-linear viscoelastic models predict fingertip pulp force-displacement characteristics during voluntary tapping** *J. Biomech* **36**:497–503

Johansson R. S., Birznieks I. (2004) First spikes in ensembles of human tactile afferents code complex spatial fingertip events *Nat. Neurosci* **7**:170–177

Johansson R. S., Flanagan J. R., Basbaum A., Kaneko A., Shepherd G., Westheimer G. (2008) **Tactile sensory control of object manipulation in humans** *The Senses: A Comprehensive Reference, pages* :136–152

Johansson R. S., Flanagan J. R. (2009) Coding and use of tactile signals from the fingertips in object manipulation tasks *Nat. Rev. Neurosci* **10**:345–359

Johansson R. S., Vallbo A. B. (1979) Tactile sensibility in the human hand: relative and absolute densities of four types of mechanoreceptive units in glabrous skin *J Physiol (Lond)* 286:283–300

Johansson R. S., Vallbo A. B. (1983) **Tactile sensory coding in the glabrous skin of the human** hand *Trends Neurosci* 6:27–32

Johansson R. S., Westling G. (1984) **Roles of glabrous skin receptors and sensorimotor memory in automatic control of precision grip when lifting rougher or more slippery objects** *Exp. Brain Res* **56**:550–564

Johansson R. S., Westling G. (1987) **Signals in tactile afferents from the fingers eliciting adaptive motor responses during precision grip** *Exp. Brain Res* **66**:141–154

Johansson R. S. (1978) Tactile sensibility in the human hand: receptive field characteristics of mechanoreceptive units in the glabrous skin area *J. Physiol* **281**:101–125

Khamis H. A., Redmond S. J., Macefield V. G., Birznieks I. (2014) **Tactile afferents encode grip safety before slip for different frictions** *In 2014 36th Annual International Conference of the IEEE Engineering in Medicine and Biology Society* :4123–4126

Knibestöl M. (1975) **Stimulus-response functions of slowly adapting mechanoreceptors in the human glabrous skin area** *J Physiol (Lond)* **245**:63–80

Kumar S., Liu G., Schloerb D., Srinivasan M. (2015) **a. Viscoelastic characterization of the primate finger pad in vivo by micro-step indentation and 3D finite element models for tactile sensation studies** *J. Biomech. Eng* 

Kunesch E., Binkofski F., Freund H. J. (1989) **Invariant temporal characteristics of** manipulative hand movements *Exp. Brain Res* **78**:539–546



Kunesch E., Knecht S., Schnitzler A., Tyercha C., Schmitz F., Freund H. J. (1995) **Somatosensory** evoked potentials elicited by intraneural microstimulation of afferent nerve fibers *J. Clin. Neurophysiol* **12**:476–487

Lindblom U. (1965) **Properties of touch receptors in distal glabrous skin of the monkey** *J. Neurophysiol* **28**:966–985

Melzack R., Bromage P. R. (1973) Experimental phantom limbs Exp. Neurol 39:261-269

Morley J. W., Goodwin A. W., Darian-Smith I. (1983) **Tactile discrimination of gratings** *Exp. Brain Res* **49**:291–299

Nakazawa N., Ikeura R., Inooka H. (2000) **Characteristics of human fingertips in the shearing direction** *Biol. Cybern* **82**:207–214

Ochoa J., Torebjörk E. (1983) **Sensations evoked by intraneural microstimulation of single mechanoreceptor units innervating the human hand** *J. Physiol* **342**:633–654

Pataky T. C., Latash M. L., Zatsiorsky V. M. (2005) **Viscoelastic response of the finger pad to incremental tangential displacements** *J. Biomech* **38**:1441–1449

Pawluk D. T., Howe R. D. (1999) **Dynamic lumped element response of the human fingerpad** *J. Biomech. Eng* **121**:178–183

Pubols B. H. (1982) Factors affecting cutaneous mechanoreceptor response. II. changes in mechanical properties of skin with repeated stimulation *J. Neurophysiol* **47**:530–542

Robles-De-La-Torre G., Hayward V. (2001) **Force can overcome object geometry in the perception of shape through active touch** *Nature* **412**:445–448

Saal H. P., Vijayakumar S., Johansson R. S. (2009) **Information about complex fingertip** parameters in individual human tactile afferent neurons *J. Neurosci* **29**:8022–8031

Serina E. R., Mote C. D., Rempel D. (1997) Force response of the fingertip pulp to repeated compression–effects of loading rate, loading angle and anthropometry *J. Biomech* **30**:1035–1040

Srinivasan M. A., Whitehouse J. M., LaMotte R. H. (1990) **Tactile detection of slip: Surface microgeometry and peripheral neural codes** *J. Neurophysiol* **63**:1323–1332

Sukumar V., Johansson R. S., Pruszynski J. A. (2022) **Precise and stable edge orientation** signaling by human first-order tactile neurons *Elife* 11

Teulings H.-L., Maarse F. J. (1984) **Digital recording and processing of handwriting movements** *Hum. Mov. Sci* **3**:193–217

Vallbo A. B., Hagbarth K. E. (1968) Activity from skin mechanoreceptors recorded percutaneously in awake human subjects *Exp. Neurol* **21**:270–289

Vallbo A. B., Johansson R. S. (1984) **Properties of cutaneous mechanoreceptors in the human hand related to touch sensation** *Hum. Neurobiol* **3**:3–14

Victor J. D., Purpura K. P. (1996) Metric-space analysis of spike trains: Theory, algorithms and application *Network-Computation In Neural Systems* 8:127–164



Walsh L. D., Hoad D., Rothwell J. C., Gandevia S. C., Haggard P. (2015) **Anaesthesia changes** perceived finger width but not finger length *Exp. Brain Res* **233**:1761–1771

Wang Q., Hayward V. (2007) **In vivo biomechanics of the fingerpad skin under local tangential traction** *J. Biomech* **40**:851–860

Watkins R. H., Amante M., Wasling H. B., Wessberg J., Ackerley R. (2022) **Slowly-adapting type II afferents contribute to conscious touch sensation in humans: evidence from single unit intraneural microstimulation** *J. Physiol* 

Werner G., Mountcastle V. B. (1965) **Neural activity in mechanoreceptive cutaneous afferents: stimulus-response relations, weber functions, and information transmission** *J. Neurophysiol* **28**:359–397

Westling G., Johansson R. S. (1984) **Factors influencing the force control during precision** grip *Exp. Brain Res* **53**:277–284

Westling G., Johansson R. S. (1987) **Responses in glabrous skin mechanoreceptors during precision grip in humans** *Exp. Brain Res* **66**:128–140

Willemet L., Kanzari K., Monnoyer J., Birznieks I., Wiertlewski M. (2021) **Initial contact shapes the perception of friction** *Proc. Natl. Acad. Sci. U. S. A* **118** 

Wu J. Z., Dong R. G., Smutz W. P., Rakheja S. (2003) **Dynamic interaction between a fingerpad and a flat surface: experiments and analysis** *Med. Eng. Phys* **25**:397–406

# **Author information**

### Hannes P. Saal

Active Touch Laboratory, Department of Psychology, University of Sheffield, Sheffield S1 2LT, UK

For correspondence: h.saal@sheffield.ac.uk ORCID iD: 0000-0002-7544-0196

### **Ingvars Birznieks**

School of Biomedical Sciences, UNSW Sydney, Sydney, New South Wales, Australia, Neuroscience Research Australia, Sydney, New South Wales, Australia ORCID iD: 0000-0003-4916-1254

### **Roland S. Johansson**

Physiology Section, Department of Integrative and Medical Biology, Umeå University, SE-901 87 Umeå, Sweden ORCID iD: 0000-0003-3288-8326

### **Editors**

Reviewing Editor **Rebecca Seal** University of Pittsburgh School of Medicine, United States of America



### Senior Editor **Tirin Moore** Howard Hughes Medical Institute, Stanford University, United States of America

# **Reviewer #1 (Public Review):**

The authors investigate how the viscoelasticity of the fingertip skin can affect the firing of mechanoreceptive afferents and they find a clear effect of recent physical skin state (memory), which is different between afferents. The manuscript is extremely well-written and well-presented. It uses a large dataset of low threshold mechanoreceptive afferents in the fingertip, where it is particularly noteworthy that the SA-2s have been thoroughly analyzed and play an important role here. They point out in the introduction the importance of the non-linear dynamics of the event when an external stimulus contacts the skin, to the point at which this information is picked up by receptors. Although clearly correlated, these are different processes, and it has been very well-explained throughout. I have some comments and ideas that the authors could think about that could further improve their already very interesting paper. Overall, the authors have more than achieved their aims, where their results very much support the conclusions and provoke many further questions. This impact of the previous dynamics of the skin affecting the current state can be explored further in so many ways and may help us to better understand skin aging and the effects of anatomical changes of the skin.

At the beginning of the Results, it states that FA-2s were not considered as stimuli and did not contain mechanical events with frequency components high enough to reliably excite them. Was this really the case, did the authors test any of the FA-2s from the larger dataset? If FA-2s were not at all activated, this is also relevant information for the brain to signal that it is not a relevant Pacinian stimulus (as they respond to everything). Further, afferent receptive fields that were more distant to the stimulus were included, which likely fired very little, like the FA-2s, so why not consider them even if their contribution was low?

One question that I wondered throughout was whether you have looked at further past history in stimulation, i.e. not just the preceding stimulus, but 2 or 3 stimuli back? It would be interesting to know if there is any ongoing change that can be related back further. I do not think you would see anything as such here, but it would be interesting to test and/or explore in future work (e.g. especially with sticky, forceful, or sharp indentation touch). However, even here, it could be that certain directions gave more effects.

Did the authors analyze or take into account the difference between receptive field locations? For example, did afferents more on the sides have lower responses and a lesser effect of history?

Was there anything different in the firing patterns between the spontaneous and non-spontaneously active SA-2s? For example, did the non-spontaneous show more dynamic responses?

Were the spontaneously active SA-2 afferents firing all the time or did they have periods of rest - and did this relate to recent stimulation? Were the spontaneously active SA-2s located in a certain part of the finger (e.g. nail) or were they randomly spread throughout the fingertip? Any distribution differences could indicate a more complicated role in skin sensing.

Did the authors look to see if the spontaneous firing in SA-2s between trials could predict the extent to which the type 1 afferents encode the proceeding stimulus? Basically, does the SA-2 state relate to how the type 1 units fire?



In the discussion, it is stated that "the viscoelastic memory of the preceding loading would have modulated the pattern of strain changes in the fingertip differently depending on where their receptor organs are situated in the fingertip". Can the authors expand on this or make any predictions about the size of the memory effect and the distance from the point of stimulation?

In the discussion, it would be good if the authors could briefly comment more on the diversity of the mechanoreceptive afferent firing and why this may be useful to the system.

Also, the authors could briefly discuss why this memory (or recency) effect occurs - is it useful, does it serve a purpose, or it is just a by-product of our skin structure? There are examples of memory in the other senses where comparisons could be drawn. Is it like stimulus adaptation effects in the other senses (e.g. aftereffects of visual motion)?

One point that would be nice to add to the discussion is the implications of the work for skin sensing. What would you predict for the time constant of relaxation of fingertip skin, how long could these skin memory effects last? Two main points to address here may be how the hydration of the skin and anatomical skin changes related to aging affect the results. If the skin is less viscoelastic, what would be the implications for the firing of mechanoreceptors?

How long does it take for the effect to end? Again, this will likely depend on the skin's viscoelasticity. However, could the authors use it in a psychophysical paradigm to predict whether participants would be more or less sensitive to future stimuli? In this way, it would be possible to test whether the direction modifies touch perception.

# **Reviewer #2 (Public Review):**

### Summary:

The authors sought to identify the impact skin viscoelasticity has on neural signalling of contact forces that are representative of those experienced during normal tactile behaviour. The evidence presented in the analyses indicates there is a clear effect of viscoelasticity on the imposed skin movements from a force-controlled stimulus. Both skin mechanics and evoked afferent firing were affected based on prior stimulation, which has not previously been thoroughly explored. This study outlines that viscoelastic effects have an important impact on encoding in the tactile system, which should be considered in the design and interpretation of future studies. Viscoelasticity was shown to affect the mechanical skin deflections and stresses/strains imposed by previous and current interaction force, and also the resultant neuronal signalling. The result of this was an impaired coding of contact forces based on previous stimulation. The authors may be able to strengthen their findings, by using the existing data to further explore the link between skin mechanics and neural signalling, giving a clearer picture than demonstrating shared variability. This is not a critical addition, but I believe would strengthen the work and make it more generally applicable.

### Strengths:

-Elegant design of the study. Direct measurements have been made from the tactile sensory neurons to give detailed information on touch encoding. Experiments have been well designed and the forces/displacements have been thoroughly controlled and measured to give accurate measurements of global skin mechanics during a set of controlled mechanical stimuli.

-Analytical techniques used. Analysis of fundamental information coding and information representation in the sensory afferents reveals dynamic coding properties to develop putative models of the neural representation of force. This advanced analysis method has been applied to a large dataset to study neural encoding of force, the temporal dynamics of this, and the variability in this.



### Weaknesses:

-Lack of exploration of the variation in neural responses. Although there is a viscoelastic effect that produces variability in the stimulus effects based on prior stimulation, it is a shame that the variability in neural firing and force-induced skin displacements have been presented, and are similarly variable, but there has been no investigation of a link between the two. I believe with these data the authors can go beyond demonstrating shared variability. The force per se is clearly not faithfully represented in the neural signal, being masked by stimulation history, and it is of interest if the underlying resultant contact mechanics are.

### Validity of conclusions:

The authors have succeeded in demonstrating skin viscoelasticity has an impact on skin contact mechanics with a given force and that this impacts the resultant neural coding of force. Their study has been well-designed and the results support their conclusions. The importance and scope of the work is adequately outlined for readers to interpret the results and significance.

### Impact:

This study will have important implications for future studies performing tactile stimulation and evaluating tactile feedback during motor control tasks. In detailed studies of tactile function, it illustrates the necessity to measure skin contact dynamics to properly understand the effects of a force stimulus on the skin and mechanoreceptors.