



This is a repository copy of *Prevalence and risk factors for *Staphylococcus aureus* colonisation among healthy individuals in low- and middle-income countries: a systematic review and meta-analysis*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/224177/>

Version: Published Version

---

**Article:**

Locke, T.E. orcid.org/0000-0001-7984-0659, Keeley, A.J. orcid.org/0000-0001-9386-1157, Laundry, N. et al. (5 more authors) (2025) Prevalence and risk factors for *Staphylococcus aureus* colonisation among healthy individuals in low- and middle-income countries: a systematic review and meta-analysis. *Journal of Infection*, 90 (4). 106462. ISSN 0163-4453

<https://doi.org/10.1016/j.jinf.2025.106462>

---

**Reuse**

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:  
<https://creativecommons.org/licenses/>

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>



## Bacteria and Bacterial Diseases

# Prevalence and risk factors for *Staphylococcus aureus* colonisation among healthy individuals in low- and middle-income countries: A systematic review and meta-analysis



Thomas E. Locke <sup>a,b,\*</sup>, Alexander J. Keeley <sup>a,b,c,d</sup>, Nicholas Laundry <sup>e</sup>, Christopher Keil <sup>f</sup>, Jean Hamilton <sup>g</sup>, Abdullah Pandor <sup>g</sup>, Thushan I de Silva <sup>a,b,c</sup>, Thomas C. Darton <sup>a,b</sup>

<sup>a</sup> Division of Clinical Medicine, School of Medicine and Population Health, The University of Sheffield, UK

<sup>b</sup> The Florey Institute of Infection, The University of Sheffield, UK

<sup>c</sup> Vaccines and Immunity Theme, MRC Unit The Gambia at the London School of Hygiene and Tropical Medicine, The Gambia

<sup>d</sup> Clinical Research Department, London School of Hygiene and Tropical Medicine, London, UK

<sup>e</sup> The Royal Hobart Hospital and University of Tasmania, Tasmania, Australia

<sup>f</sup> Department of Medical Microbiology, Sheffield Teaching Hospitals NHS Foundation Trust, Sheffield, UK

<sup>g</sup> Sheffield Centre for Health and Related Research (SCHARR), School of Medicine and Population Health, The University of Sheffield, UK

## ARTICLE INFO

### Article history:

Accepted 28 February 2025

Available online 5 March 2025

### Keywords:

*Staphylococcus aureus*

Low income populations

Antimicrobial drug resistance

## SUMMARY

**Background:** *Staphylococcus aureus* is capable of asymptomatic colonisation, which can progress to opportunistic and potentially life-threatening infection. The data on *S. aureus* colonisation in low- and middle-income countries (LMIC) are limited. This systematic review and meta-analysis estimates the prevalence of *S. aureus* colonisation in asymptomatic individuals in LMIC, with secondary objectives of assessing antimicrobial resistance, colonisation risk factors, and the molecular epidemiology of colonising strains.

**Methods:** Articles published up to July 2023 were identified by searching four electronic databases. Studies that presented *S. aureus* colonisation prevalence in healthy individuals from a community setting in LMIC were included. Data extraction was performed independently by two reviewers with disagreement resolved through consensus. Studies were critically appraised using the Joanna Briggs Institute Prevalence tool. Random effects meta-analysis was conducted where appropriate. This study was registered in advance with PROSPERO (CRD42019147780).

**Findings:** A total of 16 610 citations were identified of which 138 studies (59 732 participants) met the eligibility criteria. The majority of studies had a low risk of bias. The pooled prevalence of *S. aureus* colonisation at nose and/or throat sites was 26.4% (95% CI 23.8 - 29.1%). The prevalence of methicillin-resistance in colonising *S. aureus* strains was 15.0% (95% CI: 11.8 to 18.6%), with a higher prevalence observed in Africa compared to Asia and South America (22.5% vs. 13.1% vs. 5.4% respectively). Panton-Valentine leukocidin genes were present in 26.4% (95% CI: 17.1% to 32.8%) of 2531 isolates.

**Interpretation:** While the prevalence of asymptomatic *S. aureus* colonisation in LMIC mirrors that found in high-income countries, there was a higher prevalence of antimicrobial resistance and other virulence factors. Variability in study methods and sparsity of data from many LMIC, underscore the need for a global approach to *S. aureus* surveillance. This will be critical for informing effective infection prevention strategies.

© 2025 The Authors. Published by Elsevier Ltd on behalf of The British Infection Association. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

\* Correspondence to: Division of Clinical Medicine, School of Medicine and Population Health, The University of Sheffield, Sheffield S10 2RX, UK.

E-mail address: [mnda05tel@sheffield.ac.uk](mailto:mnda05tel@sheffield.ac.uk) (T.E. Locke).

## Impact statement

*Staphylococcus aureus* is the leading cause of bacterial infection related deaths worldwide, with treatment often complicated by antimicrobial resistance (AMR). Asymptomatic colonisation increases the risk of infection, often with an individual's own colonising strain. Community-based transmission perpetuates cycles of colonisation and infection, which intersect with healthcare settings. Targeted interventions, such as topical decolonisation therapies and future vaccines, will play a key role in addressing the global burden of *S. aureus* infection but require robust surveillance data to maximise their efficacy. While epidemiological data of this nature is readily available from high-income countries (HIC), there is a lack of equivalent data from low- and middle-income countries (LMIC). Existing LMIC studies are often limited by a narrow geographical scope or a focus on high-risk populations, such as healthcare workers or those with active infection. The lack of comprehensive global data on *S. aureus* colonisation in healthy individuals from community settings is a critical gap that limits our understanding of this pathobiont and our ability to address its burden of disease.

This systematic review and meta-analysis provides the most comprehensive overview to date of *S. aureus* colonisation in healthy individuals from LMIC, including over 50 000 participants from 40 countries. Our data reveals that *S. aureus* colonisation is a global phenomenon, with similar prevalence in LMIC and HIC. However, colonising strains in LMIC show higher levels of AMR and putative virulence factors, raising concerns about their potential to cause more severe infections. This review identifies significant disparities in the quality and quantity of *S. aureus* colonisation data between LMIC and HIC, highlighting critical gaps in global surveillance.

By mapping *S. aureus* colonisation epidemiology across LMIC, this study provides essential insights into this pathobiont, laying the groundwork for strategies to reduce the global burden of *S. aureus* infection. Future efforts should focus on establishing a standardised, collaborative approach to global *S. aureus* surveillance. This approach will be crucial for capacity building and informing localised infection prevention strategies that ultimately help to address the morbidity and mortality associated with *S. aureus*.

## Introduction

*Staphylococcus aureus* is a human pathobiont which frequently colonises the anterior nares and predisposes to infection, ranging from localised skin infection to systemic, life-threatening complications of bloodstream infection. Host- and pathogen-related factors affecting human colonisation dynamics have been reviewed recently<sup>1</sup> and are key to developing interventions for minimising or preventing *S. aureus* infection risk, especially in higher-risk groups or populations.

The prevalence of human colonisation by *S. aureus* is not well known. Whilst robust *S. aureus* colonisation surveillance data and multinational publications are available, there is bias towards high-income countries (HIC); a more comprehensive description of *S. aureus* colonisation prevalence within and across low- and middle-income countries (LMIC) is required. These data could highlight important epidemiological trends and also pave the way to localised infection transmission and prevention strategies. For example, *S. aureus* is highly clonal with certain strains exhibiting distinct virulence profiles (including antimicrobial resistance [AMR]) and becoming established in specific geographic or environmental niches.<sup>2</sup> Furthermore, there has been a focus on *S. aureus* colonisation related

to healthcare exposure, but networks of community transmission in healthy populations likely also play a key role in maintaining circulation.<sup>3</sup> Finally, *S. aureus* colonisation is associated with a number of chronic non-communicable diseases, and the prevalence of these varies but is known to be increasing globally.<sup>4,5</sup>

The primary objective of this systematic review and meta-analysis was to determine the pooled prevalence of *S. aureus* colonisation among healthy individuals in community settings by LMIC. Secondary objectives included determining the prevalence of AMR, identifying risk factors for colonisation and characterising the molecular epidemiology of colonising strains.

## Methods

### Search strategy and search criteria

We performed a systematic review and meta-analysis following a protocol registered with PROSPERO (CRD42019147780)<sup>6</sup> in accordance with PRISMA guidelines.<sup>7</sup> We systematically searched electronic databases, including Medline, Scopus, Web of Science, and the Cochrane Library until 31 July 2023 [AJK], for free text, thesaurus terms and combined synonyms relating to *Staphylococcus aureus*, colonisation, and LMIC as defined by the OECD (appendix I).<sup>8</sup> No language or date restrictions were used. All records were downloaded to EndNote (Clarivate Analytics, Philadelphia) for deduplication and initial screening [TL or AJK], with any citations clearly not meeting eligibility criteria excluded.

Eligible studies: 1) presented extractable data relating to the prevalence of *S. aureus* colonisation in healthy individuals in a community setting (i.e. non-healthcare related) in a LMIC, 2), where relevant, presented colonisation data collected before any intervention was performed, 3) were available in the English language, and 4) had the full text available.

Studies were excluded if they: 1) reported data for subjects who were hospitalised, were attending healthcare facilities (unless as part of a healthy review, for example, pregnant women attending antenatal care or people living with HIV attending for antiretroviral medication review), were healthcare workers; 2) did not make it clear that the subjects were asymptomatic; 3) did not report original data; 4) reported data from a single case; 5) did not describe the body site sampled to assess for colonisation; 6) did not use at least one of microbiological culture, targeted antigen or PCR testing to detect *S. aureus*; or 7) only reported methicillin-resistant *S. aureus* (MRSA) prevalence.

Eligible studies were imported to Covidence systematic review software (Veritas Health Innovation, Melbourne,) which was used for all subsequent selection and extraction steps. Eligibility was confirmed by full text review [TL, AJK, NL, or CK]. Studies were included if only a subpopulation met the eligibility criteria, but only subpopulation data was extracted.

### Data extraction

Data extraction was performed independently by two reviewers [TL, AJK, NL, CK], with disagreement resolved through consensus or arbitration [TD]. Only original data presented in the publicly available publication or associated supplementary materials were extracted (appendix II). Missing data for binary measures (e.g. sex) were calculated and missing values for the Human Development Index (HDI) were imputed using data from the nearest available year. Otherwise, values were marked as missing. Electronic tools such as digitisers were not used. For longitudinal studies, data from the earliest time-point was extracted. Reviewers did not evaluate their own publications.

## Risk of bias assessment

We assessed full texts for methodological quality and risk of bias using the Joanna Briggs Institute (JBI) Prevalence Critical Appraisal Tool.<sup>9</sup> Assessments were performed at the time of data extraction. Scoring of nine criteria, for responses including “yes” (one point), “no or unclear” (zero points) and “not applicable” (denominator reduced by one), produced a total score for each study. These are presented as percentages and categorised into three arbitrary levels: high risk of bias as ≤33%, moderate risk 34–66% and low risk >66%.<sup>10</sup>

## Data synthesis and analysis

Sampled body sites were categorised as nose and/or throat (NT, see [appendix III](#) for a list of NT sites), skin, other single site, or a combination where >1 site sampled without site-specific data reported separately. A single colonisation prevalence figure was included from each eligible study. If a study reported data from multiple distinct body sites, then data relating to NT sites, and specifically the anterior nares were preferentially selected, being the most high-yield site for *S. aureus* colonisation. The compiled dataset including all body sites is termed “all sites”. To minimise heterogeneity, a subset of the data were created including only studies reporting colonisation of ≥1 NT site. This “NT only” dataset was used to calculate pooled *S. aureus* colonisation prevalence and associated subgroup, meta-regression, and risk factor analyses. The “all sites” dataset was used to assess risk of bias, AMR and typing analyses as the specific body site sampled was deemed less important for these purposes.

Analysis was performed using R version 4.3.1.<sup>11</sup> Prevalence data (*S. aureus* colonisation, AMR, *pvl* gene presence, and *tst* gene presence) was transformed using the Freeman-Tukey double arcsine method and synthesised using a random effects model with the DerSimonian estimator and associated 95% confidence intervals calculated by the Clopper-Pearson method (“meta”<sup>12</sup> package). Methods for exploring heterogeneity between studies included the  $I^2$  statistic, prediction intervals, leave-one-out analysis and externally studentised residuals (“meta”<sup>12</sup> and “dmetar”<sup>13</sup> packages). Where appropriate, between study heterogeneity in colonisation prevalence was explored through subgroup analysis and meta-regression. Risk factors for *S. aureus* colonisation included variables reported at the individual participant level from more than one study. A pooled prevalence ratio (PR) for each risk factor was calculated using a meta-analysis of binary data (“meta”<sup>12</sup> package). A random effects model was used unless fewer than five studies reported data for a risk factor, in which case a fixed model (Mantel-Haenszel method without continuity correction) was used. MLST typing data were grouped by continent and the proportion of each MLST type was calculated. Strain diversity was quantified using Simpson’s Diversity Index (SDI).<sup>14</sup>

## Role of the funding source

The funder had no role in the study design, data collection, analysis, nor in the writing of the manuscript or the decision to submit for publication.

## Results

We identified 16 610 citations of which 138 studies met the eligibility criteria and were included in further analysis ([Fig. 1](#) and [appendix IV](#)).<sup>15–152</sup> The “all sites” dataset included 138 studies with 59 732 participants ([appendix V](#)). Most studies assessed *S. aureus* colonisation at a single body site (128 studies, 93%) with NT sites being the most commonly sampled (121 studies, 88%).

The “NT only” dataset included 121 studies comprising 54 527 participants (91% of participants represented in the “all sites” dataset). Publication year ranged from 1975 to 2023, with 114 studies (94%) performed since 2005. Most studies used a cross-sectional design (117 studies, 97%). Studies were conducted in four continents, with 68 studies (56%) from Asia, 41 studies (34%) from Africa, 11 studies (9.1%) from South America and one study (0.8%) from Oceania. All publications reported data from a single country only, comprising 40 distinct countries. Five countries (India, Nigeria, Iran, China, and Brazil) accounted for 56 publications (46%) and 59% of participants (32 361 of 54 527).

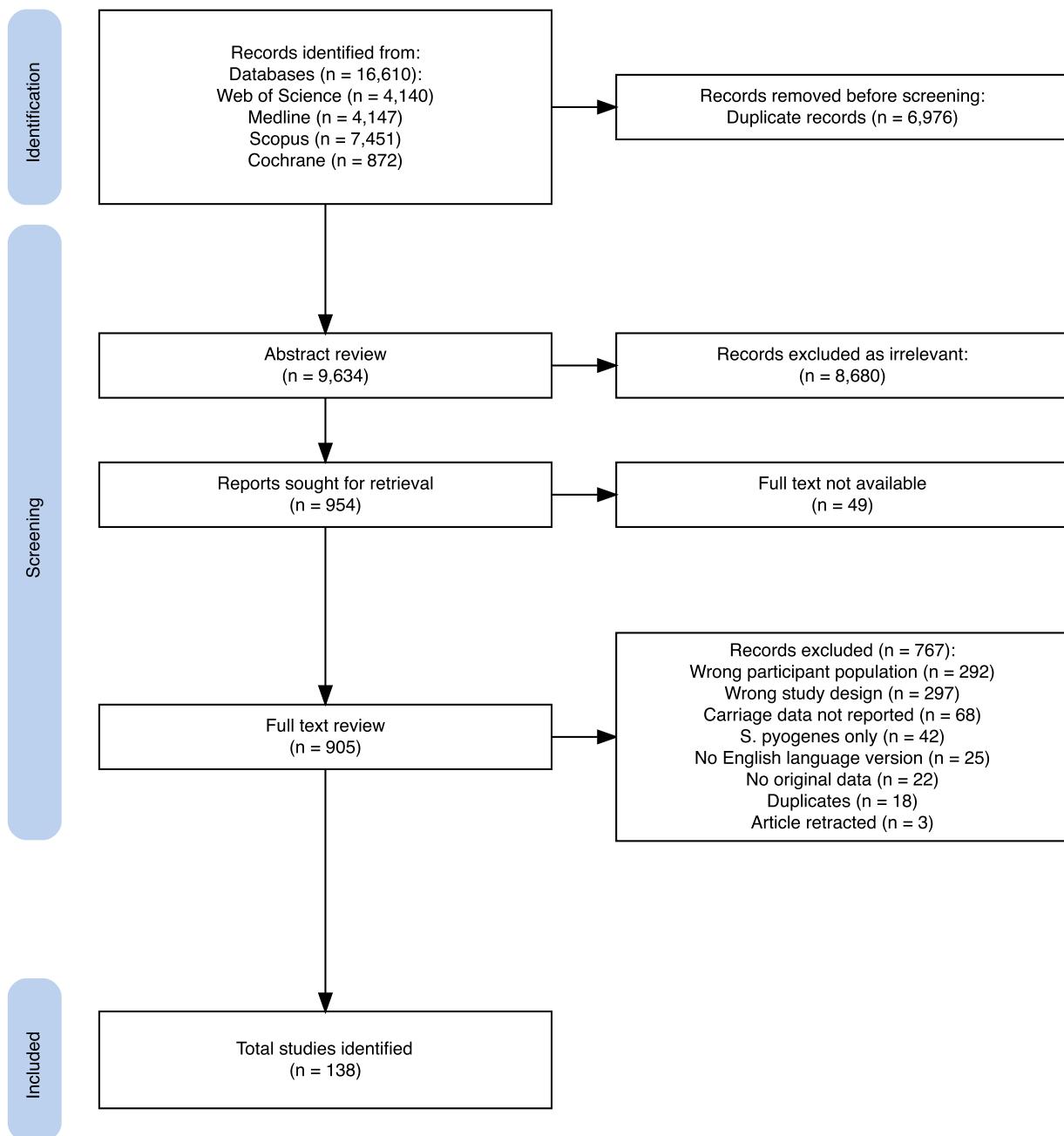
The median percentage of male participants was 52% although sex was only reported in 84 studies (69%). Studies described colonisation by age group including 44 adult only studies (36%), 41 children only (34%), 29 adult and children (24%), and seven in which age-related data were not reported (5.8%). Study populations were diverse in setting and demography, but most commonly included: being resident within a defined geographical region (32 studies, 26%), attending healthcare facilities as part of a health review (22, 18%), school children (17, 14%), university students (14, 12%), specific occupations (13, 11%), and childcare attendees (10, 8%).

Across the 121 studies in the “NT only” dataset there were 124 NT sites sampled, as three studies sampled both anterior nares and oropharynx. A single NT site was selected from each publication which resulted in the following distribution of sites sampled: anterior nares (104 sites, 86%), nasopharynx (9, 7%), oropharynx (3, 2.5%), oral rinse (2, 1.7%), and combinations of NT sites (3, 2.5%). Bacterial culture was the most common method used for the detection of *S. aureus* colonisation (92 studies, 76%), followed by a combination of culture and molecular (28, 23%) or molecular methods alone (one study, 0.8%). Skin sites were sampled in 12 studies including 2264 participants ([appendix VI](#)).

Methodological quality assessment of the 138 included studies (“all sites”) demonstrated a low risk of bias (120 studies, 87%) with the remaining 18 studies (13%) categorised as moderate risk ([appendix VII](#)). Performance tended to be poorer in domains related to participant selection. Specifically, 43 studies (31%) scored zero points for lower quality recruitment methods such as convenience sampling or due to insufficient methodological detail. Similarly, sample size was deemed inadequate in 30 studies (22%), with limited description of the population studied in 52 studies (38%).

From 121 studies included in the “NT only” dataset, 12 933 of 54 527 subjects were colonised with *S. aureus* in the nose and/or throat. The pooled prevalence was estimated at 26.4% (95% CI 23.8–29.1%) ([Fig. 2](#)). Between study heterogeneity in *S. aureus* colonisation prevalence was high ( $I^2=98.0\%$ , 95%CI 97.8–98.1%, prediction interval of 4.3–58.2%). However, sensitivity analysis yielded consistent results with no notable change in the pooled *S. aureus* colonisation prevalence estimate ([appendix VIII](#)).

Factors explored in subgroup analysis of NT *S. aureus* colonisation prevalence included continent the study was performed in, specific NT site sampled, participant age group, HDI category, laboratory methods for *S. aureus* detection, and type of population sampled. Significant differences were observed for continent ( $p < 0.001$ ), NT site sampled ( $p = 0.036$ ) and laboratory method ( $p < 0.001$ ; [appendix IX](#)). The highest prevalence was found in South America (36.7%, 95% CI: 24.6–49.8%), followed by Africa (31.0%, 95% CI: 25.7–36.7%), while studies from Asia (22.4%, 95% CI: 19.5–25.4%) reported lower prevalence rates ([Fig. 3](#)). Combined culture and molecular methods detected a higher prevalence (27.2%, 95%CI: 23.2–31.4%) of *S. aureus* colonisation compared to culture alone (26.4%, 95% CI: 23.2–29.7%). A lower prevalence of *S. aureus* colonisation was detected from nasopharyngeal sampling (14.1%, 95% CI: 6.0–24.9%) compared to the anterior nares (27.0%, 95% CI: 24.3–29.8%), although the number of participants in the nasopharyngeal sampling group was limited.

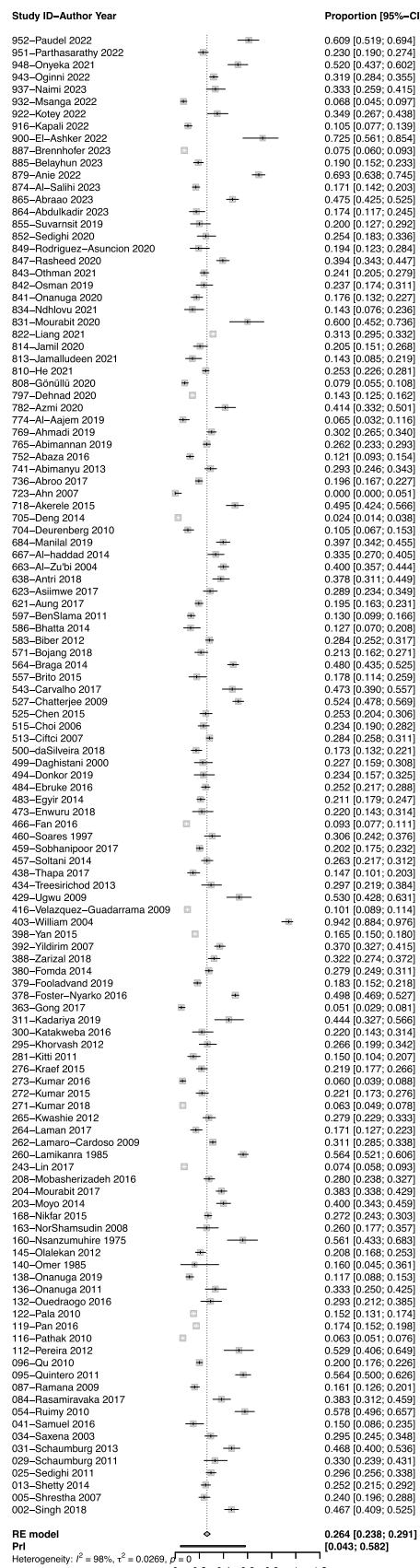


**Fig. 1.** PRISMA flowchart (aside from the 'Included' step, all numbers reported here include studies relating to *S. aureus* and *S. pyogenes*).

Using multiple meta-regression we explored the relationship between *S. aureus* colonisation prevalence at NT sites and study world sub-region, year of study, laboratory methods for *S. aureus* detection, HDI score, and specific NT site sampled (Table 1). Heterogeneity between studies remained high ( $I^2$  97·6%), although variance in colonisation prevalence was comparably low ( $\tau^2$  0·02). East Asian countries, namely China, exhibited significantly lower *S. aureus* colonisation prevalence compared to Sub-Saharan African countries (coefficient -0·2341,  $p$ =0·003). Lower prevalence was also estimated when sampling the nasopharynx compared to the anterior nares (coefficient -0·2105,  $p$ =0·0019). Both findings remained significant after adjusting for study year, HDI, and laboratory method of *S. aureus* detection. Missing data necessitated the exclusion of sex and average age as covariates in the meta-regression model, with data only available from 84 studies (69%) and 50 studies (41%), respectively.

However, no significant relationship between either variable and colonisation was found in univariate analysis (average age: coefficient -0·0017,  $p$ =0·31; proportion of male participants: coefficient 0·53,  $p$ =0·96).

Most studies (85%) described AST data, but these were not extractable from eight of 117 studies due to limitations in reporting, including the absence of raw data. Testing methods included phenotypic techniques such as disc diffusion and gradient strip testing (52 studies, 47%), molecular methods (4 studies, 3·7%), or both (52 studies, 47%). Overall, 56 studies (51%) included molecular testing, with all testing for MRSA by PCR amplification of the *mecA* gene. Additionally, six studies (11%) tested for *mecC*, and one study (1·8%) for VRSA using *vanA* and *vanB* gene amplification. Among the 104 studies using phenotypic methods, 79% (82 studies) cited Clinical and Laboratory Standards Institute guidance.



**Fig. 2.** Forest plot showing the pooled prevalence of *S. aureus* colonization from nose and/or throat sites.

Pooled prevalence of *S. aureus* resistance to 14 antimicrobials is detailed in [Table 2](#); the majority of *S. aureus* isolates were resistant to penicillin (90%), whereas resistance to gentamicin, mupirocin, rifampicin, or linezolid was uncommon (from 0.8 to 9.4%).

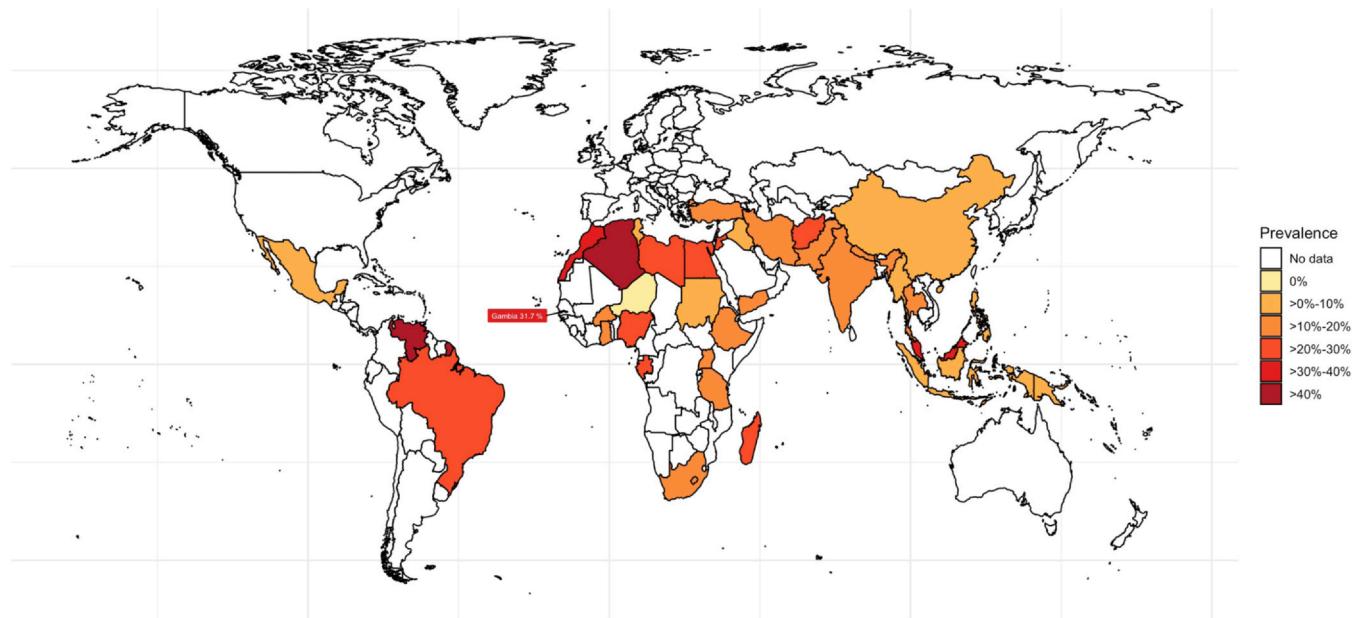
Nearly all studies where AST data were reported (103 studies, 95%) included testing for methicillin resistance, using phenotypic methods (47 studies, 46%), molecular methods (four studies, 4%), or both (52 studies, 50%). Among studies using molecular methods, only 25 (45%) tested all isolates while the remainder used it for confirmatory testing. Overall, 12 234 isolates were tested for MRSA, yielding a pooled prevalence of 15.0% (95% CI: 11.8 to 18.6%). Subgroup analysis and meta-regression were used to further explore the relationship between MRSA prevalence and continent the study was performed in, AST methodology, and study end year. MRSA prevalence varied by continent with higher prevalence in Africa (22.5%, 95%CI: 13.3–33.3%), compared to Asia (13.1%, 95%CI: 9.9–16.8%), and South America (5.4%, 95%CI: 2.9–8.4%) ([appendix X](#)). Subgroup analysis of MRSA prevalence by study country is presented in [Fig. 4](#). The lower prevalence of methicillin resistance in Asian countries remained statistically significant in a meta-regression model that included AST testing methodology and study end year ([appendix X](#)). Since 2000, African and Asian studies have shown an increasing trend in MRSA prevalence (Africa:  $R=0.39$ ,  $p=0.026$ , Asia:  $R=0.31$ ,  $p=0.018$ ) although this was not seen in studies from South America ( $R=0.041$ ,  $p=0.92$ ; [appendix X](#)).

Vancomycin resistance was assessed in 4 385 isolates from 43 of 109 studies. All studies used phenotypic methods, and one also used molecular techniques. The pooled prevalence estimate for VRSA was 0.5% (95%CI 0 to 1.8%). The prevalence of VRSA was highest in African countries (4.1%) although this was not statistically significant ([appendix XI](#)).

We performed a meta-analysis to explore potential risk factors for *S. aureus* colonisation in LMIC using the “NT only” dataset ([Table 3](#)). Aside from participant sex, this analysis was hindered by the limited data available for each risk factor resulting in small sample sizes. Household animal contact (PR 1.35,  $p=0.03$ , fixed effects model) and attendance at childcare facilities (PR 1.38,  $p < 0.001$ , fixed effects model) were significantly associated with *S. aureus* colonisation at NT sites. There was a non-significant trend towards less colonisation among smokers (PR 0.73,  $p=0.14$ , random effects model). No difference in colonisation was observed between males and females (PR 1.03,  $p=0.58$ , random effects model; [appendix XII](#)).

Panton-Valentine leukocidin is a *S. aureus* toxin putatively associated with virulence and transmission. In 37 studies comprising 2 531 isolates, the pooled prevalence of *pvl* genes was 24.6% (95% CI: 17.1% to 32.8%). Additionally, 13 studies comprising 1 181 isolates assessed the presence of toxic shock syndrome toxin (*tst*) gene, with a pooled prevalence of 10.8% (95% CI: 4.4% to 19.1%).

Multi-locus sequencing typing (MLST) was the most frequently reported *S. aureus* typing method. A total of 129 unique MLST types were identified from 911 bacterial isolates across 20 studies. Only 14 countries reported MLST data, with China accounting for nearly a third (279 of 911 isolates, 31%). The top 20 most common MLST types from each continent are presented in [Fig. 5](#). Diversity was higher in studies performed in Africa and Asia compared to South America, although this difference was not statistically significant (SDI: 0.92, 0.93, and 0.88, respectively,  $p=0.36$ ). Although there was strain diversity within each continent, types ST1, ST5, ST15, ST188, and ST30 were among the most commonly encountered in all three continents. In contrast, several types were only associated with a single continent, such as ST22 and ST398 in Asia and ST1223 in South America.



**Fig. 3.** World map with pooled prevalence of *S. aureus* colonisation by country from nose and/or throat sites.

## Discussion

In this systematic review of healthy community populations living in LMIC, including over 50 000 participants from 40 LMIC, we estimate the prevalence of *S. aureus* colonisation at nose and/or throat sites to be 26.4%. Notably, we found a high prevalence of bacterial factors putatively associated with virulence, including methicillin-resistance and PVL-toxin presence. Colonisation is a major risk factor for invasive infection, and with *S. aureus* being a leading cause of death due to bacterial infection frequently complicated by AMR<sup>153,154</sup>, the data presented in this review highlight the critical need for public health interventions suitable to LMIC community settings.

Our estimation of *S. aureus* colonisation prevalence specifically addresses healthy individuals in community settings. Despite the

ubiquity of *S. aureus*, global data from these settings are scarce. Instead, most studies focus on those with infection, or populations at higher risk of exposure such as healthcare contact. Previous meta-analyses from individual LMIC report *S. aureus* colonisation prevalence between 21.2% and 30.9%<sup>155,156</sup>. While multi-country estimates have been generated, these are limited by methodological weaknesses that make direct comparison difficult. These include a focus on MRSA only, limited patient populations, uncertainty regarding sites sampled, incorporating estimates from studies with less strict inclusion criteria, and the inclusion of infection cases or non-LMIC settings.<sup>157–160</sup> In contrast, large-scale *S. aureus* prevalence studies are more common in HIC. In Europe, the pooled nasal colonisation prevalence from nine countries was 21.6% (range 12.7–29.4%, 32 206 samples).<sup>161</sup> Similarly, a nationwide study in the US estimated nasal colonisation prevalence at 28.6% (9 004

**Table 1**

Multiple meta-regression of *S. aureus* colonisation prevalence at nose and/or throat sites.

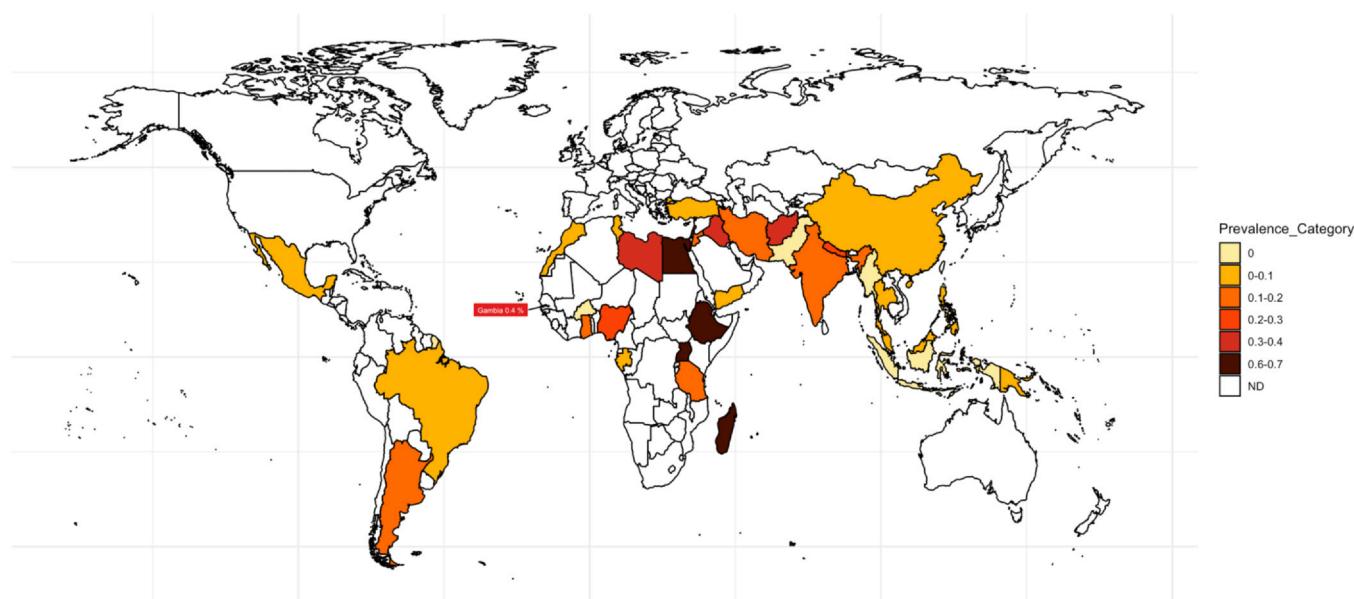
Factor	Coefficient	95% CI (Coefficient)	OR	P-value
Sub-region				
Sub-Saharan Africa (reference)	NA	NA	1.0000	NA
Northern Africa	0.0068	-0.1521 to 0.1657	1.0068	0.9328
Southern Asia	-0.1155	-0.2245 to -0.0066	0.8909	<b>0.0379</b>
South-Eastern Asia	-0.1012	-0.2517 to 0.0493	0.9037	0.1852
Eastern Asia	-0.2341	-0.3868 to -0.0813	0.7913	<b>0.0030</b>
Western Asia	-0.1278	-0.2706 to 0.015	0.8800	0.0788
Latin America and the Caribbean	0.0033	-0.1571 to 0.1636	1.0033	0.9678
Melanesia	-0.2165	-0.5607 to 0.1277	0.8053	0.2151
Other				
Study end year	-0.0035	-0.0084 to 0.0015	0.9965	0.1667
HDI	0.0165	-0.4292 to 0.4621	1.0166	0.9417
Laboratory detection method				
Culture (reference)	NA	NA	1.0000	NA
Culture & molecular	-0.0357	-0.1209 to 0.0496	0.9649	0.4084
Molecular	-0.1298	-0.4887 to 0.2291	0.8783	0.4749
Nose/throat site				
Anterior nares (reference)	NA	NA	1.0000	NA
Nasopharynx & throat	0.1220	-0.2329 to 0.477	1.1298	0.4970
Anterior nares & throat	0.1793	-0.0696 to 0.4282	1.1964	0.1562
Nasopharynx	-0.2105	-0.3414 to -0.0797	0.8102	<b>0.0019</b>
Oral rinse	0.1739	-0.0877 to 0.4356	1.1900	0.1903
Oropharynx	-0.1626	-0.3822 to 0.0569	0.8499	0.1449

Statistically significant results ( $P < 0.05$ ) are indicated in bold.

HDI = Human Development Index.

**Table 2**Pooled prevalence of antimicrobial resistance to 14 antimicrobials for *S. aureus* isolates from all body sites.

Antimicrobial	Number of studies	Number of isolates	Proportion resistant	95% CI
Chloramphenicol	26	2752	0.1073	0.0556 - 0.1718
Clindamycin	52	5570	0.1838	0.1199 - 0.2567
Co-trimoxazole	60	5648	0.2237	0.1537 - 0.3018
Doxycycline	9	530	0.2858	0.0876 - 0.5367
Erythromycin	68	6470	0.2954	0.2328 - 0.3618
Gentamicin	58	4870	0.0938	0.0629 - 0.1293
Linezolid	21	2337	0.0082	0 - 0.034
Methicillin (MRSA)	103	12 234	0.1502	0.1177 - 0.1857
Mupirocin	7	1130	0.0320	0 - 0.1365
Penicillin	44	4501	0.9031	0.8245 - 0.9625
Rifampicin	27	2762	0.0310	0.0128 - 0.0549
Teicoplanin	13	1339	0.0013	0 - 0.0481
Tetracycline	42	4693	0.2400	0.1832 - 0.3014
Vancomycin (VRSA)	43	4385	0.0053	0 - 0.0181

**Fig. 4.** World map with pooled prevalence of methicillin resistance in colonising *S. aureus* isolates by country from all body sites.**Table 3**Pooled prevalence ratio for eight risk factors for *S. aureus* colonisation at nose and/or throat sites.

Risk factor	Studies	Cases	Total	Model	Prevalence ratio	95% CI	P-value
Male sex	36	10 037	19 919	Random	1.0303	0.9254 - 1.1472	0.5758
Hospitalisation (12 months)	11	1244	9317	Random	1.1977	0.8498 - 1.6879	0.2686
Antibiotics (6 months)	9	2530	8090	Random	1.1988	0.7754 - 1.8533	0.3653
Smoker (active)	5	1173	5183	Random	0.7308	0.4529 - 1.1793	0.1430
HCP (household)	5	366	2042	Random	0.9175	0.6880 - 1.2235	0.4529
Animal (household)	2	287	1540	Fixed	1.3523	1.0333 - 1.7698	<b>0.0279</b>
HIV	3	485	1146	Fixed	0.9293	0.7206 - 1.1985	0.5722
Childcare (attends)	2	259	853	Fixed	1.3766	1.1454 - 1.6545	<b>0.0007</b>

Statistically significant results ( $P < 0.05$ ) are indicated in bold.

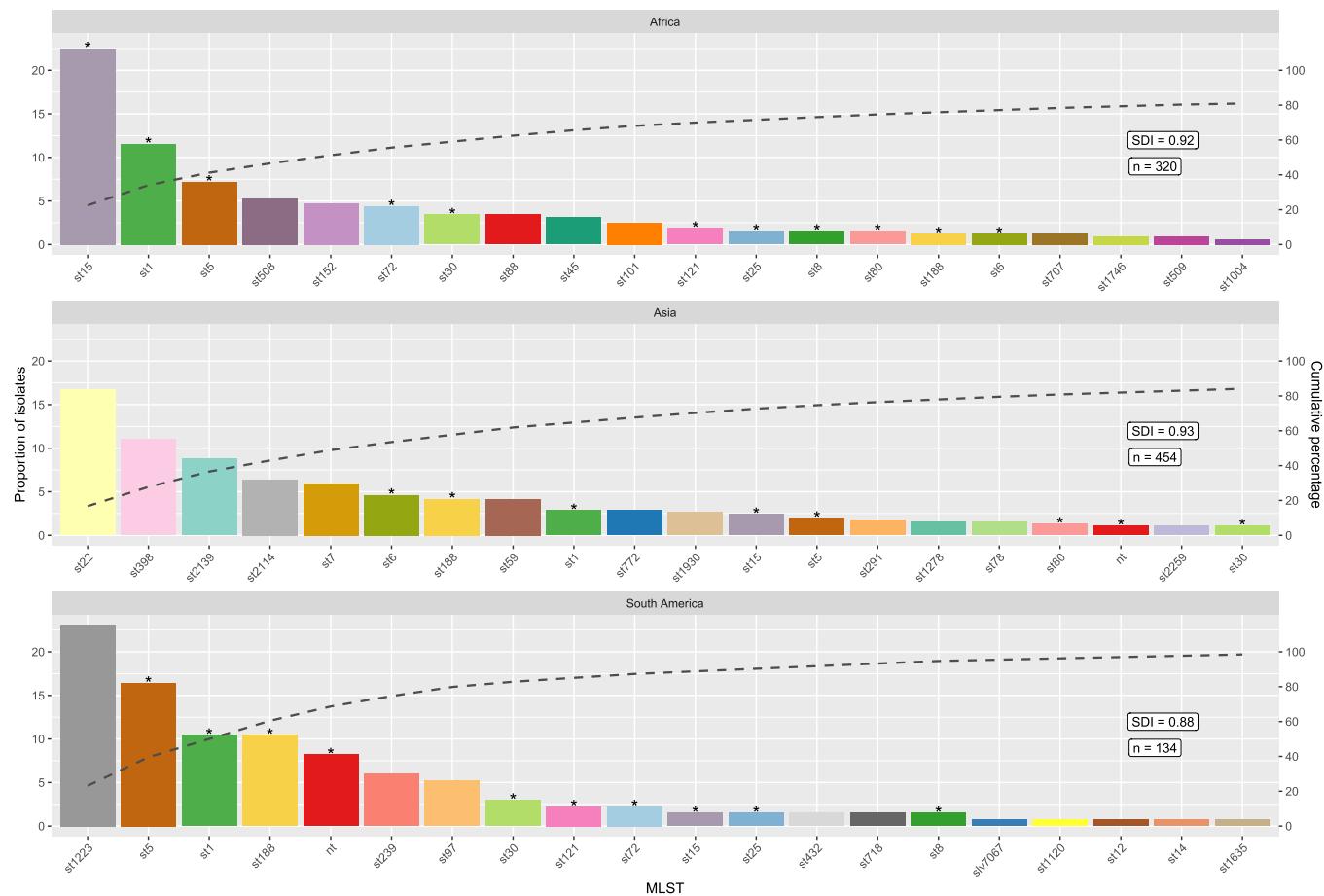
HCP = Healthcare professional.

HIV = Human immunodeficiency virus.

samples).<sup>162</sup> These two studies underscore the stark disparity in the quantity of *S. aureus* data available from HIC and LMIC, with a combined sample size of 41 210, compared to 54 527 in the 121 studies from 40 countries included in this systematic review.<sup>161,162</sup>

As a pathobiont, colonising strains of *S. aureus* are often implicated in opportunistic, invasive infections, emphasising the importance of understanding resistance profiles to guide antimicrobial selection. While we found that nearly all colonising *S. aureus* isolates exhibited resistance to penicillin, the widespread and high levels of resistance to commonly used anti-staphylococcal agents such as macrolides, clindamycin, tetracyclines, and co-trimoxazole was

frequent and concerning. These agents are often used first-line for managing skin and soft tissue infection (SSTI) and are widely available and affordable. In contrast, resistance rates among colonising *S. aureus* isolates in Europe are notably lower. For example, erythromycin (1.6% to 16.5%), tetracycline (1.8% to 7.2%), and co-trimoxazole (0% to 1%).<sup>161</sup> We also found a high prevalence of MRSA among colonising isolates in LMIC (15%), markedly surpassing that typically observed in HIC (0–2% in European and US populations).<sup>161,162</sup> Particularly concerning were the high MRSA prevalences found in the Africa and Asia regions, with increasing trends since 2000. This is in contrast to previous reports from the Americas,



**Fig. 5.** Top 20 most common MLST as proportions of all isolates for three continents (Oceania excluded as only three isolates underwent MLST). \*\* denotes an MLST identified in two or more continents. SDI = Simpson's Diversity Index.

Europe and Asia-Pacific where MRSA prevalence amongst clinical isolates appears to have peaked in the mid-2000s.<sup>163</sup> The pooled prevalence of VRSA was 0.5%, with the highest levels in African countries. Whilst the global prevalence of VRSA is low, it remains a clinical concern, particularly for LMIC where access to alternative antimicrobials may be limited.<sup>164</sup> Overall, our findings demonstrate higher rates of antimicrobial resistance among colonising *S. aureus* strains in LMIC compared to HIC. Studies from HIC settings demonstrate that infection caused by drug-resistant strains is associated with increased mortality and healthcare costs<sup>165,166</sup>, and so ongoing surveillance is crucial to systematically monitor local *S. aureus* epidemiology and to inform patient management and public health strategies.

We found that household animal exposure and attendance at childcare facilities were significantly associated with *S. aureus* colonisation, consistent with previous reports from HIC settings.<sup>3</sup> However, other commonly implicated factors found not to be associated in our review included male sex, recent antibiotic use or hospitalisation. This is likely due to limited data availability, substantial heterogeneity across the included studies and diversity within each risk factor. For example, differing types and duration of healthcare exposure. While we note some trend towards reduced *S. aureus* colonisation in association with smoking, the nature of this relationship remains unclear, with conflicting results from previous studies.<sup>167,168</sup>

PVL toxin has been associated with SSTI, particularly amongst community-acquired cases of MRSA infection.<sup>169</sup> In this meta-analysis, approximately a quarter of colonising *S. aureus* isolates were found to be PVL-positive. In general, there is limited data on the

prevalence of PVL amongst colonising strains as most work has explored a virulence role during infection. While PVL genes appear to be more commonly found in infecting *S. aureus* strains compared to colonising isolates, the exact role in infection is still debated.<sup>169,170</sup> Where data is available from HIC, PVL is infrequently detected in colonising *S. aureus* isolates (<1% in European studies).<sup>171,172</sup> A higher prevalence of PVL in colonising MRSA has been reported in the US likely due to the success of the PVL-positive USA300 clone, particularly in community settings. In contrast, the *tst* gene, associated with toxic shock syndrome, was identified in 10.8% of colonising isolates in this review, and a similar prevalence has been previously reported in European studies (17.5 to 32.5%).<sup>171,172</sup>

Multi-locus sequence typing identified 129 different sequence types from over 900 *S. aureus* isolates. Common types identified in multiple LMIC and across different continents included ST1, ST5, ST15, and ST30. However, these types are frequently reported in association with infections worldwide and are not unique to LMIC,<sup>2,173</sup> highlighting the capacity of *S. aureus* clones to spread and become established in diverse settings. For instance, in the current review, ST15 was frequently isolated in Africa, Asia and South America and is also associated with bacteraemia throughout Europe.<sup>173–175</sup> The ST22 strain was one of the most frequently identified types in this review but only in Asian studies. However, this strain is globally distributed and the observed discrepancy may be explained by the high number of isolates sequenced from Asia, specifically in China.<sup>176</sup> ST1223 was exclusively identified in South America but this MLST type has recently been linked with *Staphylococcus argenteus* which is a newly described member of the *S. aureus* clonal complex.<sup>177</sup> In general, there is a paucity of data that

characterises the molecular epidemiology of colonising *S. aureus* isolates, particularly in LMIC settings. Instead, studies have tended to focus on infecting strains and frequently those with methicillin resistance. The typing data presented here should be interpreted cautiously for three reasons. Firstly, the relevance of typing colonising *S. aureus* strains to infection is unclear. Whilst the vast majority of invasive infection is due to colonising strains, infection is relatively uncommon and therefore only a small proportion of colonising isolates will ultimately cause infection. Secondly, the establishment of dominant strains in an environmental or geographical niche is an evolving and dynamic process. Given the longitudinal nature of this meta-analysis across multiple decades, it is difficult to accurately infer the current global molecular profile of *S. aureus*. Finally, variability in typing methods and laboratory techniques can limit direct comparisons and data synthesis.

Several limitations to this study should be acknowledged. Firstly, the study populations were highly heterogeneous, encompassing a globally diverse population from multiple geographical and socio-cultural regions. This heterogeneity also extended to the anatomical sites sampled. Additionally, laboratory techniques including use of molecular methods and the use of internationally-defined standards for *S. aureus* detection were variably used and described. Our aim was to capture a global snapshot of *S. aureus* in LMIC reflecting the natural diversity of colonisation while recognising that colonisation is often a transient state.<sup>1</sup> To mitigate this, we employed strict eligibility criteria ensuring participants were asymptomatic and not from traditionally higher risk groups. Methodologically, we used a random-effects model and meta-regression analysis to explore the high levels of heterogeneity. Secondly, although this review is broad in scope, the final dataset only includes data from ~25% of all LMIC. This may in part be due to the previously mentioned eligibility criteria but is also reflective of the gaps in *S. aureus* surveillance in many LMIC. Whilst five countries account for a large proportion of the data, they are also some of the world's most populous nations, suggesting that this analysis may be representative at a population level, albeit not geographically. Thirdly, the exclusion of non-English publications may have led to under-representation of certain regions, particularly Latin America and the Caribbean, where studies are published in relevant non-English language. Fourth, the assessment of methodological quality was limited for many studies by poor quality reporting of study methodology.

In conclusion, prevalence of asymptomatic human colonisation by *S. aureus* in LMIC is similar to that observed in HIC. In LMIC community settings however, colonising strains more often demonstrate characteristics associated with a virulent phenotype including antimicrobial resistance and presence of putative virulence-associated genes. This review highlights inequalities in the quantity and quality of *S. aureus* colonisation data from LMIC compared to HIC. Given the significance of *S. aureus* as a human pathogen, our findings underscore the need for a global approach to surveillance to effectively deploy future infection prevention strategies.

## Funding

Funding was received from the University of Sheffield Institutional Open Access Fund to cover journal open access fees.

## Author contributions

The project was developed by AJK and TL with supervision provided by TdS and TCD. The methodology was developed by TL, AJK, TdS, TCD, AP, and JH. Data extraction was performed by TL, AJK, NL, and CK. The data reported here has been independently accessed and verified by TL, AJK, NL and CK. Formal data analysis and generation of figures was performed by TL with input from JH and AP. TL wrote the first draft of the manuscript and TCD, TdS, AJK, NL, CK, AP, and JH

were all involved in reviewing and editing subsequent drafts. All authors have read the final version of the manuscript, had full access to all the data in the study, and had final responsibility for the decision to submit for publication.

## Data availability

Data from this study will be made available with publication to researchers who provide a methodologically sound proposal to TL. Where appropriate this may include the raw collated data, a data dictionary, and the analysis code.

## Declaration of Competing Interest

One of the authors is an Editorial Board Member/Editor-in-Chief/Associate Editor/Guest Editor for this journal and was not involved in the editorial review or the decision to publish this article.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests:

**Thushan de Silva** – Associate Editor, Journal of Infection.

**Abdullah Pandor** - Member of the NIHR HTA Programme Funding Committee (General).

**Thomas Darton:**

- **Grants:** UKRI - Future Leaders Fellowship awarded to TCD (grant number MR/X032736/1) to study host-pathogen interactions with *S. aureus*, GSK Vaccines – Collaborative partnership award (CA/9211) awarded to the University of Sheffield to study immune responses to *S. aureus*, and National Institute for Health and Care Research (NIHR) Sheffield Biomedical Research Centre (NIHR203321).
- **Support for attending meetings and/or travel:** Wellcome Trust – Travel and accommodation reimbursement for attending a meeting in London (Vaccines and AMR, including *S. aureus*) and WHO – Travel and accommodation reimbursement for attending a Technical Advisory Group meeting in Geneva (Vaccines and AMR, including *S. aureus*).
- **Participation on a Data Safety Monitoring Board or Advisory Board:** GSK – Consulting fees for Research Advisory Board participation (on *S. aureus* vaccines).
- **Other:** GSK – Chief Investigator for *S. aureus* vaccine study (study 208833), with payment to the University of Sheffield.

## Appendices

Supplementary data associated with this article can be found in the online version at doi:[10.1016/j.jinf.2025.106462](https://doi.org/10.1016/j.jinf.2025.106462).

## References

1. Pievngam P, Otto M. *Staphylococcus aureus* colonisation and strategies for decolonisation. *Lancet Microbe* 2024;5:e606–18.
2. Lakhundi S, Zhang K. Methicillin-resistant *Staphylococcus aureus*: molecular characterization, evolution, and epidemiology. *Clin Microbiol Rev* 2018;31:e00020–18.
3. Mork RL, Hogan PG, Muenks CE, Boyle MG, Thompson RM, Sullivan ML, et al. Longitudinal, strain-specific *Staphylococcus aureus* introduction and transmission events in households of children with community-associated meticillin-resistant *S. aureus* skin and soft tissue infection: a prospective cohort study. *Lancet Infect Dis* 2020;20:188–98.
4. Phelps NH, Singleton RK, Zhou B, Heap RA, Mishra A, Bennett JE, et al. Worldwide trends in underweight and obesity from 1990 to 2022: a pooled analysis of 3663 population-representative studies with 222 million children, adolescents, and adults. *Lancet* 2024;403:1027–50.
5. Zhou B, Bennett JE, Phelps NH, NCD Risk Factor Collaboration. Worldwide trends in diabetes prevalence and treatment from 1990 to 2022: a pooled analysis of 1108 population-representative studies with 141 million participants. *Lancet* 2024;404:2077–93.
6. Keeley A, de Silva T, Johnston P, Darton T. *Staphylococcus aureus* and *Streptococcus pyogenes* carriage in low and middle income countries: a systematic review and meta-analysis of risk factors and prevalence. PROSPERO:

- International prospective register of systematic reviews; 2019. Available at: ([https://www.crd.york.ac.uk/prospero/display\\_record.php?ID=CRD42019147780](https://www.crd.york.ac.uk/prospero/display_record.php?ID=CRD42019147780)).
7. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. *The PRISMA 2020 statement: an updated guideline for reporting systematic reviews*. *Syst Rev* 2021; **10**:89.
  8. OECD. DAC List of ODA Recipients, effective for reporting on 2018, 2019 and 2020 flows 2019 OECD. Available at: ([https://www.oecd.org/dac/financing-sustainable-development/development-finance-standards/DAC\\_List\\_ODA\\_Recipients2018to2020\\_flows\\_En.pdf](https://www.oecd.org/dac/financing-sustainable-development/development-finance-standards/DAC_List_ODA_Recipients2018to2020_flows_En.pdf)). Accessed: 11/09/2019.
  9. Munn Z, Moola S, Lisy K, Rüttano D, Tufanaru C. *Methodological guidance for systematic reviews of observational epidemiological studies reporting prevalence and cumulative incidence data*. *Int J Evid Based Healthc* 2015; **13**:147–53.
  10. Algarni M, Hadi MA, Yahyouche A, Mahmood S, Jalal Z. *A mixed-methods systematic review of the prevalence, reasons, associated harms and risk-reduction interventions of over-the-counter (OTC) medicines misuse, abuse and dependence in adults*. *J Pharm Policy Pract* 2021; **14**:76.
  11. R Core Team. R: A Language and Environment for Statistical Computing; 2023. <https://www.R-project.org/>. Accessed: 01/01/2020.
  12. Balduzzi S, Rücker G, Schwarzer G. *How to perform a meta-analysis with R: a practical tutorial*. *Evid Based Ment Health* 2019; **22**:153–60.
  13. Harrer M, Cuijpers P, Furukawa T, Ebert DD. Dmetar: Companion R package for the guide 'doing meta-analysis in R'. Available at: (<http://dmetar.protectlab.org/>). Accessed: 01/12/2023.
  14. Smeesters PR, de Crombrugghe G, Tsai SK, Leclercq C, Baker C, Osowicki J, et al. *Global Streptococcus pyogenes strain diversity, disease associations, and implications for vaccine development: a systematic review*. *Lancet Microbe* 2024; **5**:181–93.
  15. Singh AK, Agarwal L, Kumar A, Sengupta C, Singh RP. *Prevalence of nasal colonization of methicillin-resistant Staphylococcus aureus among schoolchildren of Barabanki district, Uttar Pradesh, India*. *J Family Med Prim Care* 2018; **7**:162–6.
  16. Shrestha B. *Study of beta lactamase activity of Staphylococcus aureus isolated from healthy nasal carriers and hospital isolates*. *Nepal Med Coll J* 2007; **9**:107–10.
  17. Shetty V. *Prevalence of community-acquired methicillin-resistant Staphylococcus aureus nasal colonization among children*. *J Clin Diagn Res* 2014; **8**:DC12–5.
  18. Sedighi I, Moez HJ, Alikhani MY. *Nasal carriage of methicillin resistant Staphylococcus aureus and their antibiotic susceptibility patterns in children attending day-care centers*. *Acta Microbiol Immunol Hung* 2011; **58**:227–34.
  19. Schaumburg F, Köck R, Friedrich AW, Soulanoudjinga S, Ngoa UA, von Eiff C, et al. *Population structure of Staphylococcus aureus from remote African Babongo Pygmies*. *PLoS Negl Trop Dis* 2011; **5**:1150.
  20. Schaumburg F, Biallas B, Alabi AS, Grobisch MP, Feugap EN, Lell B, et al. *Clonal structure of Staphylococcus aureus colonizing children with sickle cell anaemia and healthy controls*. *Epidemiol Infect* 2013; **141**:1717–20.
  21. Saxena S, Singh K, Talwar V. *Methicillin-resistant Staphylococcus aureus prevalence in community in the east Delhi area*. *Jpn J Infect Dis* 2003; **56**:54–6.
  22. Samuel AB, Arabi Mohammed Saleh MA. *Analysis of drug resistant Staphylococcus aureus present in healthy human carriers in the community of Ambur Town, Tamil Nadu*. *Res J Pharm Biol Chem Sci* 2016; **7**:6–11.
  23. Ruimy R, Angebault C, Djossou F, Dupont C, Epelboin L, Jarraud S, et al. *Are host genetics the predominant determinant of persistent nasal Staphylococcus aureus carriage in humans?* *J Infect Dis* 2010; **202**:924–34.
  24. Rasamiravaka T, Andriatsitohana TT, Rasamindrakotroka A. *Evaluation of methicillin-resistant Staphylococcus aureus nasal carriage in Malagasy pig and poultry non-industrial farmers*. *J Infect Dev Ctries* 2017; **11**:129–35.
  25. Ramana KV, Mohanty SK, Wilson CG. *Staphylococcus aureus colonization of anterior nares of school going children*. *Indian J Pediatr* 2009; **76**:813–6.
  26. Quintero B, Araque M, van der Gaast-de Jongh C, Escalona F, Correa M, Morillo-Puente S, et al. *Epidemiology of Streptococcus pneumoniae and Staphylococcus aureus colonization in healthy Venezuelan children*. *Eur J Clin Microbiol Infect Dis* 2011; **30**:7–19.
  27. Qu F, Cui E, Guo T, Li H, Chen S, Liu L, et al. *Nasal colonization of and clonal transmission of methicillin-susceptible Staphylococcus aureus among Chinese military volunteers*. *J Clin Microbiol* 2010; **48**:64–9.
  28. Pereira DFA, Pinheiro MM, Silva PNF, Teodoro GR, Brighenti FL, Koga-Ito CY. *Influence of TNF-α blockers on the oral prevalence of opportunistic microorganisms in ankylosing spondylitis patients*. *Clin Exp Rheumatol* 2012; **30**:679–85.
  29. Pathak A, Marothi Y, Iyer RV, Singh B, Sharma M, Eriksson B, et al. *Nasal carriage and antimicrobial susceptibility of Staphylococcus aureus in healthy preschool children in Ujjain, India*. *BMC Pediatr* 2010; **10**:100.
  30. Pan H, Cui B, Huang Y, Yang J, Ba-Thein W. *Nasal carriage of common bacterial pathogens among healthy kindergarten children in Chaoshan region, southern China: a cross-sectional study*. *BMC Pediatr* 2016; **16**:161.
  31. Pala K, Oezakin C, Akis N, Sinirtas M, Gedikoglu S, Aytekin H. *Asymptomatic carriage of bacteria in food workers in Nilufer district, Bursa, Turkey*. *Turk J Med Sci* 2010; **40**:133–9.
  32. Ouedraogo A-S, Dunyach-Remy C, Kissou A, Sanou S, Poda A, Kyelem CG, et al. *High nasal carriage rate of Staphylococcus aureus containing Panton-Valentine leukocidin- and EDIN-encoding genes in community and hospital settings in Burkina Faso*. *Front Microbiol* 2016; **7**:1406.
  33. Onanuga A, Temedie TC. *Nasal carriage of multi-drug resistant Staphylococcus aureus in healthy inhabitants of Amassoma in Niger delta region of Nigeria*. *Afr Health Sci* 2011; **11**:176–81.
  34. Onanuga A, Onaolapo JA. *Antimicrobial susceptibility of community-associated Staphylococcus aureus isolates from healthy women in Zaria, Nigeria*. *Trop J Pharm Res* 2008; **7**:929–34.
  35. Onanuga A, Eboh RD, Okou GT. *Antibiogram and virulence characteristics of multi-drug resistant Staphylococcus aureus from nasal cavity of healthy students of Niger Delta University, Amassoma, Bayelsa State, Nigeria*. *J Clin Diagn Res* 2019; **13**:176–81.
  36. Omer EF, Hadi AE, Sakhi ES. *Bacteriology of sore throats in a Sudanese population*. *J Trop Med Hyg* 1985; **88**:337–41.
  37. Olalekan AO, Schaumburg F, Nurjadi D, Dike AE, Ojurongbe O, Kolawole DO, et al. *Clonal expansion accounts for an excess of antimicrobial resistance in Staphylococcus aureus colonising HIV-positive individuals in Lagos, Nigeria*. *Int J Antimicrob Agents* 2012; **40**:268–72.
  38. Nsanzumuhire H, Masawe AJ, Mhalu FS. *The bacteriological ecosystem of the skin of children in an African tropical environment (Tanzania)*. *Br J Dermatol* 1975; **92**:77–84.
  39. Nor Shamsudin M, Sekawi Z, van Belkum A, Neela V. *First community-acquired methicillin-resistant Staphylococcus aureus in Malaysia*. *J Med Microbiol* 2008; **57**:1180–1.
  40. Nikfar R, Shamsizadeh A, Kajbaf TZ, Panah MK, Khaghani S, Moghaddam M. *Frequency of methicillin-resistant Staphylococcus aureus nasal carriage in healthy children*. *Iran J Microbiol* 2015; **7**:67–71.
  41. Ateba Ngoa U, Schaumburg F, Adegnika AA, Kösters K, Möller T, Gaus E, et al. *Epidemiology and population structure of Staphylococcus aureus in various population groups from a rural and semi urban area in Gabon, Central Africa*. *Acta Trop* 2012; **124**:42–7.
  42. Ndiaye C, Bassene H, Lagier J-C, Raoult D, Sokhna C. *Asymptomatic carriage of Streptococcus pneumoniae detected by qPCR on the palm of hands of populations in rural Senegal*. *PLoS Negl Trop Dis* 2018; **12**:e0006945.
  43. Moyo SJ, Aboud S, Blomberg B, Mkopi N, Kasubi M, Manji K, et al. *High nasal carriage of methicillin-resistant Staphylococcus aureus among healthy Tanzanian under-5 children*. *Microb Drug Resist* 2014; **20**:82–8.
  44. Mourabit N, Arakrak A, Bakkali M, Laglaoui A. *Nasal carriage of sequence type 22 MRSA and livestock-associated ST398 clones in Tangier, Morocco*. *J Infect Dev Ctries* 2017; **11**:536–42.
  45. Mobasherizadeh S, Shojaei H, Havaei SA, Mostafavizadeh K, Davoodabadi F, Khorvash F, et al. *Nasal carriage screening of community-associated methicillin resistant Staphylococcus aureus in healthy children of a developing country*. *Adv Biomed Res* 2016; **5**:144.
  46. Lin J, Xu P, Peng Y, Lin D, Ou Q, Zhang T, et al. *Prevalence and characteristics of Staphylococcus aureus and methicillin-resistant Staphylococcus aureus nasal colonization among a community-based diabetes population in Foshan, China*. *J Diabetes Investig* 2017; **8**:383–91.
  47. Lamikanra A, Paul BD, Akinwole OB, Paul MO. *Nasal carriage of Staphylococcus aureus in a population of healthy Nigerian students*. *J Med Microbiol* 1985; **19**:211–6.
  48. Lamaro-Cardoso J, de Lencastre H, Kipnis A, Pimenta FC, Oliveira LS, Oliveira RM, et al. *Molecular epidemiology and risk factors for nasal carriage of Staphylococcus aureus and methicillin-resistant S. Aureus in infants attending day care centers in Brazil*. *J Clin Microbiol* 2009; **47**:3991–7.
  49. Laman M, Greenhill A, Coombs GW, Robinson O, Pearson J, Davis T, Manning L. *Methicillin-resistant Staphylococcus aureus in Papua New Guinea: a community nasal colonization prevalence study*. *Trans R Soc Trop Med Hyg* 2017; **111**:360–2.
  50. Kwasie ANA, Muibat AF, Modupe AI, Adejumoke BJM, Kehinde AO. *A survey of bacterial isolates cultured from apparently healthy individuals in South-Western Nigeria*. *Int J Trop Med* 2012; **7**:130–7.
  51. Kumar H, Zahoor U, Rana R, Mahajan T, Premeshwari Devi M, Garg RK, et al. *Prevalence of community-acquired methicillin-resistant Staphylococcus aureus among tribal population of north-western Himalayas, India*. *J Environ Biol* 2018; **39**:419–25.
  52. Kumar H, Palaha R, Kaur N, Ratnakar WS, Sodi A, Kaur M, et al. *Prevalence of multidrug-resistant, coagulase-positive Staphylococcus aureus in nasal carriage, food, wastewater and paper currency in Jalandhar city (north-western), an Indian state of Punjab*. *Environ Monit Assess* 2015; **187**:4134.
  53. Kumar H, Kaur A, Kishor N, Goutam U, Palaha R. *Prevalence of multiple antibiotic resistant nasal carriage MRSA among healthy population of border villages in Amritsar region, Punjab, India*. *J Clin Diagn Res* 2016; **10**:DL1–2.
  54. Krishnamurthy V, Saha A, Renushri BV, Nagaraj ER. *Methicillin resistant Staphylococcus aureus carriage, antibiotic resistance and molecular pathogenicity among healthy individuals exposed and not exposed to hospital environment*. *J Clin Diagn Res* 2014; **8**:DC04–8.
  55. Kraef C, Alabi AS, Peters G, Becker K, Kremsner PG, Rossatanga EG, et al. *Co-detection of Panton-Valentine leukocidin encoding genes and cotrimoxazole resistance in Staphylococcus aureus in Gabon: implications for HIV-patients' care*. *Front Microbiol* 2015; **6**:60.
  56. Kittip T, Boonyonying K, Sitthisak S. *Prevalence of methicillin-resistant Staphylococcus aureus among university students in Thailand*. *Southeast Asian J Trop Med Public Health* 2011; **42**:1498–504.
  57. Kigbu A, Orimadegun AE, Tongoo OO, Odaibo GN, Olaleye DO, Akinyinka OO. *Intestinal bacterial colonization in the first 2 weeks of life of Nigerian neonates using standard culture methods*. *Front Pediatr* 2016; **4**:139.
  58. Khorvash F, Abdi F, Ataei B, Fattahai Neisiyan H, Hasanzadeh Kashani H, Nariman T. *Nasal carriage of Staphylococcus aureus: frequency and antibiotic resistance in healthy adults*. *J Res Med Sci* 2012; **17**:S229–32.
  59. Khamidah N, Ervianti E, Suktanto H. *Staphylococcus aureus colonization on antecubital non-exacerbated atopic dermatitis patient compared to healthy children*. *Dermatol Rep* 2019; **11**:109–11.
  60. Katakweba AS, Muhairwa AP, Espinosa-Gongora C, Guardabassi L, Mtambo MM, Olsen JE. *Spa typing and antimicrobial resistance of Staphylococcus aureus from healthy humans, pigs and dogs in Tanzania*. *J Infect Dev Ctries* 2016; **10**:143–8.

61. Kadariya J, Thapaliya D, Bhatta S, Mahatara RL, Bempah S, Dhakal N, Smith TC. Multidrug-resistant *Staphylococcus aureus* colonization in healthy adults is more common in Bhutanese refugees in Nepal than those resettled in Ohio. *BioMed Res Int* 2019; **2019**:5739247.
62. Inyang-Etch PC, Udofiga GC, Alaribe AAA, Udonwa NE. Asymptomatic bacteruria in patients on antiretroviral drug therapy in Calabar. *J Med Sci* 2009; **9**:270–5.
63. Gong Z, Shu M, Xia Q, Tan S, Zhou W, Zhu Y, Wan C. *Staphylococcus aureus* nasal carriage and its antibiotic resistance profiles in children in high altitude areas of Southwestern China. Portación nasal de *Staphylococcus aureus* y sus perfiles de resistencia a antibióticos en niños que viven en zonas de gran altitud del sudeste de. *Arch Argent Pediatr* 2017; **115**:274–7.
64. Gardella N, Murzicato S, Di Gregorio S, Cuirolo A, Desse J, Crudo F, et al. Prevalence and characterization of methicillin-resistant *Staphylococcus aureus* among healthy children in a city of Argentina. *Infect Genet Evol* 2011; **11**:1066–71.
65. Foster-Nyarko E, Kwambana B, Aderonke O, Ceesay F, Jarju S, Bojang A, et al. Associations between nasopharyngeal carriage of Group B Streptococcus and other respiratory pathogens during early infancy. *BMC Microbiol* 2016; **16**:97.
66. Fooladvand S, Sarmadian H, Habibi D, van Belkum A, Ghaznavi-Rad E. High prevalence of methicillin resistant and enterotoxin gene-positive *Staphylococcus aureus* among nasally colonized food handlers in central Iran. *Eur J Clin Microbiol Infect Dis* 2019; **38**:87–92.
67. Fomda BA, Thokar MA, Khan A, Bhat JA, Zahoor D, Bashir G, et al. Nasal carriage of Methicillin-resistant *Staphylococcus aureus* among healthy population of Kashmir, India. *Indian J Med Microbiol* 2014; **32**:39–43.
68. Zarizal S, Yeo CC, Faizal GM, Chew CH, Zakaria ZA, Jamil Al-Obaidi MM, et al. Nasal colonisation, antimicrobial susceptibility and genotypic pattern of *Staphylococcus aureus* among agricultural biotechnology students in Besut, Terengganu, east coast of Malaysia. *Trop Med Int Health* 2018; **23**:905–13.
69. Yıldırım M, Şahin I, Başak S, et al. The investigation of nasal MRSA carriage and colonization of nasopharyngeal pathogens at a primary school in Düzce. *Turk J Med Sci* 2007; **37**:359–65.
70. Yan X, Song Y, Yu X, Tao X, Yan J, Luo F, et al. Factors associated with *Staphylococcus aureus* nasal carriage among healthy people in Northern China. *Clin Microbiol Infect* 2015; **21**:157–62.
71. William JL, Radu S, Aziz SA, Rahim RA, Cheah YK, Liwan A, Lihan S. Prevalence of *Staphylococcus aureus* carriage by young Malaysian footballers during indoor training. *Br J Sports Med* 2004; **38**:12–4.
72. Velazquez-Guadarrama N, Martinez-Aguilar G, Galindo JA, Zuniga G, Arbo-Sosa A. Methicillin-resistant *S. aureus* colonization in Mexican children attending day care centres. *Clin Invest Med* 2009; **32**:E57–63.
73. Ugwu MC, Odimegwu DC, Ibezim EC, Esimone CO. Antibiotic resistance patterns of *Staphylococcus aureus* isolated from nostrils of healthy human subjects in a southeastern Nigeria locality. *Maced J Med Sci* 2009; **2**:294–300.
74. Treesirichod A, Hantagoor S, Prommalikit O. Nasal carriage and antimicrobial susceptibility of *Staphylococcus aureus* among medical students at the HRH Princess Maha Chakri Sirindhorn Medical Center, Thailand: a cross sectional study. *J Infect Public Health* 2013; **6**:196–201.
75. Thapa S, Gokhale S, Sharma AL, Sapkota LB, Ansari S, Gautam R, et al. Burden of bacterial upper respiratory tract pathogens in school children of Nepal. *BMJ Open Respir Res* 2017; **4**:e000203.
76. Soltani B, Taghavi Ardakan A, Moravveji A, Erami M, Haji Rezaei M, Moniri R, Namazi M. Risk factors for methicillin resistant *Staphylococcus aureus* nasal colonization of healthy children. *Jundishapur J Microbiol* 2014; **7**:e20025.
77. Sobhanipoor MH, Ahmadrajabi R, Karmostaj A, Saffari F. Molecular characterization of nasal methicillin resistant *Staphylococcus aureus* isolates from workers of an automaker company in southeast Iran. *APMIS* 2017; **125**:921–6.
78. Soares MJ, Tokumaru-Miyazaki NH, Noleto AL, Figueiredo AM. Enterotoxin production by *Staphylococcus aureus* clones and detection of Brazilian epidemic MRSA clone (III::B:A) among isolates from food handlers. *J Med Microbiol* 1997; **46**:214–21.
79. Fan YP, Wang XL, Li L, Yao ZJ, Chen SD, Ye XH. Potential relationship between phenotypic and molecular characteristics in revealing livestock-associated *Staphylococcus aureus* in Chinese humans without occupational livestock contact. *Front Microbiol* 2016; **7**:1517.
80. Enwuru NV, Adesida SA, Enwuru CA, Ghebremedhin B, Mendie UE, Coker AO. Genetics of bi-component leukocidin and drug resistance in nasal and clinical *Staphylococcus aureus* in Lagos, Nigeria. *Microb Pathog* 2018; **115**:1–7.
81. Eleje GU, Adinma JI, Ghasi S, Ikechebelu JI, Igwegbe AO, Okonkwo JE, et al. Antibiotic susceptibility pattern of genital tract bacteria in pregnant women with preterm premature rupture of membranes in a resource-limited setting. *Int J Gynaecol Obstet* 2014; **127**:10–4.
82. El Farran CA, Sekar A, Balakrishnan A, Shammugam S, Arumugam P, Gopalswamy J. Prevalence of biofilm-producing *Staphylococcus epidermidis* in the healthy skin of individuals in Tamil Nadu, India. *Indian J Med Microbiol* 2013; **31**:19–23.
83. Egyir B, Guardabassi L, Esson J, Nielsen SS, Newman MJ, Addo KK, Larsen AR. Insights into nasal carriage of *Staphylococcus aureus* in an urban and a rural community in Ghana. *PLoS One* 2014; **9**:e96119.
84. Ebruke C, Dione MM, Walter B, Worwui A, Adegbola RA, Roca A, Antonio M. High genetic diversity of *Staphylococcus aureus* strains colonising the nasopharynx of Gambian villagers before widespread use of pneumococcal conjugate vaccines. *BMC Microbiol* 2016; **16**:38.
85. Donkor ES, Kotey FCN, Dayie N, Duodu S, Tetteh-Quarcoo PB, Osei MM, Tette E. Colonization of HIV-infected children with methicillin-resistant *Staphylococcus aureus*. *Pathogens* 2019; **8**:35.
86. Daghistani HI, Issa AA, Shehabi AA. Frequency of nasal and wound isolates of *Staphylococcus aureus* associated with TSST-1 production in Jordanian population. *FEMS Immunol Med Microbiol* 2000; **27**:95–8.
87. da Silveira M, Cunha MLRS, de Souza CSM, Correa AAF, Fortaleza CMCB. Nasal colonization with methicillin-resistant *Staphylococcus aureus* among elderly living in nursing homes in Brazil: risk factors and molecular epidemiology. *Ann Clin Microbiol Antimicrob* 2018; **17**:18.
88. Ciftci IH, Koken R, Bokulmez A, Ozdemir M, Safak B, Cetinkaya Z. Nasal carriage of *Staphylococcus aureus* in 4-6 age groups in healthy children in Afyonkarahisar, Turkey. *Acta Paediatr* 2007; **96**:1043–6.
89. Choi CS, Yin CS, Bakar AA, Sakewi Z, Naing NN, Jamal F, Othman N. Nasal carriage of *Staphylococcus aureus* among healthy adults. *J Microbiol Immunol Infect* 2006; **39**:458–64.
90. Chen B, Dai X, He B, Pan K, Li H, Liu X, et al. Differences in *Staphylococcus aureus* nasal carriage and molecular characteristics among community residents and healthcare workers at Sun Yat-Sen University, Guangzhou, Southern China. *BMC Infect Dis* 2015; **15**:303.
91. Chatterjee SS, Ray P, Aggarwal A, Das A, Sharma M. A community-based study on nasal carriage of *Staphylococcus aureus*. *Indian J Med Res* 2009; **130**:742–8.
92. Carvalho SP, Almeida JB, Andrade YMFS, Silva L, Oliveira AC, Nascimento FS, et al. Community-acquired methicillin-resistant *Staphylococcus aureus* carrying SCCmec type IV and V isolated from healthy children attending public daycares in north-eastern Brazil. *Braz J Infect Dis* 2017; **21**:464–7.
93. Capriotti JA, Pelletier JS, Shah M, Caivano DM, Ritterband DC. Normal ocular flora in healthy eyes from a rural population in Sierra Leone. *Int Ophthalmol* 2009; **29**:81–4.
94. Butcher RMR, Sokana O, Jack K, Kalae E, Sui L, Russell C, et al. Active trachoma cases in the Solomon Islands have varied polymicrobial community structures but do not associate with individual non-chlamydial pathogens of the eye. *Front Med* 2017; **4**:251.
95. Brito CS, Queiroz LL, Campos PA, Batistão DW, Silva Hde A, de Agostini GG, et al. The nares as a CA-MRSA reservoir in the healthy elderly. *Rev Soc Bras Med Trop* 2015; **48**:614–6.
96. Braga ED, Aguiar-Alves F, de Freitas Mde F, de e Silva MO, Correia TV, Snyder RE, et al. High prevalence of *Staphylococcus aureus* and methicillin-resistant *S. aureus* colonization among healthy children attending public daycare centers in informal settlements in a large urban center in Brazil. *BMC Infect Dis* 2014; **14**:538.
97. Bojang A, Camara B, Jagne Cox I, Oluwalana C, Lette K, Usuf E, et al. Long-term impact of oral azithromycin taken by Gambian women during labor on prevalence and antibiotic susceptibility of *Streptococcus pneumoniae* and *Staphylococcus aureus* in their infants: follow-up of a randomized clinical trial. *Clin Infect Dis* 2018; **67**:1191–7.
98. Bibar A, Abuelaish I, Rahav G, Raz M, Cohen L, Valinsky L, et al. A typical hospital-acquired methicillin-resistant *Staphylococcus aureus* clone is widespread in the community in the Gaza strip. *PLoS One* 2012; **7**:e42864.
99. Bhatta DR, Gokhale S, Sharma AL, Gupta U, Gaur A, Gowda S, et al. Carrier state of *Haemophilus influenzae* type b (Hib), *Streptococcus pneumoniae*, *Streptococcus pyogenes*, *Neisseria meningitidis* and *Corynebacterium diphtheriae* among school children in Pokhara, Nepal. *Asian Pac J Trop Dis* 2014; **4**:45–9.
100. Ben Slama K, Gharsa H, Klibi N, Jouini A, Lozano C, Gómez-Sanz E, et al. Nasal carriage of *Staphylococcus aureus* in healthy humans with different levels of contact with animals in Tunisia: genetic lineages, methicillin resistance, and virulence factors. *Eur J Clin Microbiol Infect Dis* 2011; **30**:499–508.
101. Aung MS, San T, Aye MM, Mya S, Maw WW, Zan KN, et al. Prevalence and genetic characteristics of *Staphylococcus aureus* and *Staphylococcus argenteus* isolates harboring Panton-Valentine leukocidin, enterotoxins, and TSST-1 genes from food handlers in Myanmar. *Toxins* 2017; **9**:241.
102. Asiimwe BB, Baldan R, Trovato A, Cirillo DM. Molecular epidemiology of Panton-Valentine leukocidin-positive community-acquired methicillin resistant *Staphylococcus aureus* isolates in pastoral communities of rural south western Uganda. *BMC Infect Dis* 2017; **17**:24.
103. Antri K, Akkou M, Bouchiat C, Bes M, Martins-Simoes P, Dauwalder O, et al. High levels of *Staphylococcus aureus* and MRSA carriage in healthy population of Algiers revealed by additional enrichment and multisite screening. *Eur J Clin Microbiol Infect Dis* 2018; **37**:1521–9.
104. Al-Zubri E, Bdour S, Shehabi AA. Antibiotic resistance patterns of *mecA*-positive *Staphylococcus aureus* isolates from clinical specimens and nasal carriage. *Microb Drug Resist* 2004; **10**:321–4.
105. Al-haddad OH, Zorgani A, Ghengesh KS. Nasal carriage of multi-drug resistant Panton-Valentine leukocidin-positive methicillin-resistant *Staphylococcus aureus* in children in Tripoli-Libya. *Am J Trop Med Hyg* 2014; **90**:724–7.
106. Manial A, Shewangizaw M, Mama M, Gezmu T, Merdekios B. Methicillin-resistant *Staphylococcus aureus* colonization in HIV patients of Arba Minch province, Ethiopia: carriage rates, antibiotic resistance, and biofilm formation. *Acta Microbiol Immunol Hung* 2019; **66**:469–83.
107. Dong D, Ni Q, Wang C, Zhang L, Li Z, Jiang C, et al. Effects of intestinal colonization by *Clostridium difficile* and *Staphylococcus aureus* on microbiota diversity in healthy individuals in China. *BMC Infect Dis* 2018; **18**:207.
108. Deurenberg RH, Beisser PS, Visschers MJ, Driessens C, Stobberingh EE. Molecular typing of methicillin-susceptible *Staphylococcus aureus* isolates collected in the Yogyakarta area in Indonesia, 2006. *Clin Microbiol Infect* 2010; **16**:92–4.
109. Deng JJ, Xiao GG, Zhu Y, Zhou W, Wan CM. *Staphylococcus aureus* nasal carriage and its antibiotic resistance profiles in Tibetan School Children in Southwest China. *Hong Kong J Paediatr* 2014; **19**:75–8.

110. Akerele JO, Obasuyi O, Omede D. Prevalence of methicillin-resistant *Staphylococcus aureus* among healthy residents of Ekosodin Community in Benin-City, Nigeria. *Trop J Pharm Res* 2015;14:1495–9.
111. Akerele J, Abhulimen P, Okonofua F. Prevalence of asymptomatic genital infection among pregnant women in Benin City, Nigeria. *Afr J Reprod Health* 2002;6:93–7.
112. Anh DD, Huong PLT, Watanabe K, Nguyet NT, Anh NTH, Thi NT, et al. Increased rates of intense nasopharyngeal bacterial colonization of Vietnamese children with radiological pneumonia. *Tohoku J Exp Med* 2007;213:167–72.
113. Abroo S, Hosseini Jazani N, Sharifi Y. Methicillin-resistant *Staphylococcus aureus* nasal carriage between healthy students of medical and nonmedical universities. *Am J Infect Control* 2017;45:709–12.
114. Abimanyu N, Murugesan S, Krishnan P. High prevalence of exfoliative toxins among carrier isolates of *Staphylococcus aureus* from healthy individuals from various communities in Chennai, South India. *Indian J Microbiol* 2013;53:288–90.
115. Abaza AF, Mohamed ON, El-Fiky FK, Ahmed KA. Nasal carriage of methicillin-resistant *Staphylococcus aureus* and the effect of tea extracts on isolates. *J Egypt Public Health Assoc* 2016;91:135–43.
116. Aakko J, Grzeskowiak Ł, Asukas T, Päiväsäde E, Lehto KM, Fan YM, et al. Lipid-based nutrient supplements do not affect gut bifidobacterium microbiota in malawian infants: a randomized trial. *J Pediatr Gastroenterol Nutr* 2017;64:610–5.
117. Abimannan N, Sumathi G, Krishnarajasekhar OR, Sinha B, Krishnan P. Clonal clusters and virulence factors of methicillin-resistant *Staphylococcus aureus*: evidence for community-acquired methicillin-resistant *Staphylococcus aureus* infiltration into hospital settings in Chennai, South India. *Indian J Med Microbiol* 2019;37:326–36.
118. Ahmadi E, Khojasteh M, Mortazavi SM, Khan-Mohammadi F, Kazemnia A, Beheshtipour J, Raeissadeh M. Prevalence of and risk factors for methicillin-resistant *Staphylococcus aureus* nasal carriage in the West of Iran: a population-based cross-sectional study. *BMC Infect Dis* 2019;19:899.
119. Al-Ajemi BMR. A comparative study between *S.aureus*, methecillin resistance *S. Aureus* and *Pseudomonas aeruginosa* carriage in diabetic's patients in general and those with foot ulcer in Baquba City, Diyala Province. *Indian J Public Health Res Dev* 2019;10:556–61.
120. Azmi AH, Adnan SNA, Ab Malik N. The prevalence of *Staphylococcus aureus* in the oral cavity of healthy adults in Malaysia. *Sains Malaysiana* 2020;49:583–91.
121. Dehnad A, Agdam MHG, Rahbarnia L, Naghili B, Saffarian P. Detection of hemolysine genes in methicillin-resistant *S. Aureus* isolates obtained from a healthy population in north-west of Iran. *Gene Rep* 2020;21:100874.
122. Göntüllü N, Yıldız S, Aydoğan O, Taner Z, Özdemir S, Akyol S, et al. Nasopharyngeal carriage of potential pathogenic bacteria in healthy children living in Istanbul. *Haseki Tip Bull* 2020;58:470–6.
123. He S, Lin J, Li L, Cai W, Ye J, Li Y, et al. Multidrug-resistant *Staphylococcus aureus* nasal carriage among HIV-positive outpatients in Guangzhou, China: prevalence, risk factors, phenotypic and molecular characteristics. *J Infect Chemother* 2021;27:218–25.
124. Jamalludeen NM. Nasal carriage of *Staphylococcus aureus* in healthy children and its possible bacteriophage isolates in basrah, Iraq. *Biomed Pharmacol J* 2021;14:467–75.
125. Jamil J, Zaman K, Ullah S, Ali I, Kalsoom. Colonization of *Staphylococcus aureus* in nasal cavities of healthy individuals from district Swabi, KP, Pakistan. *J Pak Med Assoc* 2020;70:1154–8.
126. Liang B, Liang X, Gao F, Long Y, Mai J, Ai X, et al. Active surveillance, drug resistance, and genotypic profiling of *Staphylococcus aureus* among school-age children in China. *Front Med* 2021;8:701494.
127. Mourabit N, Arakrak A, Bakkali M, Zian Z, Bakkach J, Laglaoui A. Nasal carriage of *Staphylococcus aureus* in farm animals and breeders in north of Morocco. *BMC Infect Dis* 2020;20:602.
128. Ndhlovu GON, Abotsi RE, Shittu AO, Abdulgader SM, Jamrozy D, Dupont CL, et al. Molecular epidemiology of *Staphylococcus aureus* in African children from rural and urban communities with atopic dermatitis. *BMC Infect Dis* 2021;21:348.
129. Onanuga A, Adamu OJ, Odetoyin B, Hamza JA. Nasal carriage of multi-drug resistant panton valentine leukocidin positive *Staphylococcus aureus* in healthy individuals of tudun-wada, Gombe State, Nigeria. *Afr J Infect Dis* 2020;15:24–33.
130. Osman M, Kamal-Dine K, El Omari K, Rafei R, Dabboussi F, Hamze M. Prevalence of *Staphylococcus aureus* methicillin-sensitive and methicillin-resistant nasal carriage in food handlers in Lebanon: a potential source of transmission of virulent strains in the community. *Access Microbiol* 2019;1:e000043.
131. Othman AM, Al-Huraibi BS, Assayaghi RM, Al-Shami HZ. Nasal carriage and methicillin resistance of *Staphylococcus aureus* among schoolchildren in Sana'a City, Yemen. *Int J Microbiol* 2021;2021:5518317.
132. Rasheed NA, Hussein NR. Prevalence of nasal carriage rate and antimicrobial susceptibility testing of *Staphylococcus aureus* strains isolated from syrian students in Kurdistan, Iraq. *Middle East J Rehabil Health Stud* 2020;7:1–5.
133. Rodriguez-Acunión K, Kanapi MPL, Wassmer GD, Cailli JC. Methicillin-resistant *Staphylococcus aureus* (*Mrsa*) colonization, risk factors, and antibiotic susceptibility profile among asymptomatic diabetes mellitus type 2 patients. *Philos J Intern Med* 2020;58:58–64.
134. Sedighi I, Faradmal J, Alikhani MY, Olfat M. The comparison of *Staphylococcus aureus* nasal colonization and its antibiotic resistance patterns in Children of Health CareWorkers (HCWs) and non-HCWs. *J Compr Pediatr* 2020;11.
135. Suvarnsit K, Kiratisin P, Bunnag C, Tantilipikorn P. Prevalence of nasal carriage of *Staphylococcus aureus* in allergic rhinitis patients and healthy controls in Thailand. *Asian Pac J Allergy Immunol* 2021;39:163–7.
136. Abdulkadir A, Kabir J, Mohammed B, Olayinka B. Characterisation and prevalence of community-associated *MRSA* among horses, dogs, cats and their human handlers: a cross-sectional study. *Trans R Soc Trop Med Hyg* 2023;117:212–8.
137. Abraão LM, Fortaleza CMCB, Camargo CH, Barbosa TA, Pereira-Franchi E, Riboli D, et al. *Staphylococcus aureus* and CA-MRSA carriage among Brazilian Indians living in peri-urban areas and remote communities. *Antibiotics* 2023;12:862.
138. Al-Salih SS, Karim GF, Al-Bayati AMS, Obaid HM. Prevalence of methicillin-resistant and methicillin sensitive *Staphylococcus aureus* nasal carriage and their antibiotic resistant patterns in Kirkuk City, Iraq. *J Pure Appl Microbiol* 2023;17:329–37.
139. Anie CO, Ibezim EC, Esimone CO, Arhewoh MI. Molecular characterization of selected nasal isolates of methicillin-resistant *Staphylococcus aureus* (MRSA) from healthy students of a tertiary institution. *Tropical J Pharm Res* 2022;21:825–31.
140. Belayhun C, Tilahun M, Seid A, Shibabaw A, Sharew B, Belete MA, Demissis W. Asymptomatic nasopharyngeal bacterial carriage, multi-drug resistance pattern and associated factors among primary school children at Debre Berhan town, North Shewa, Ethiopia. *Ann Clin Microbiol Antimicrob* 2023;22:9.
141. Brennhofer SA, Rogawski McQuade ET, Zhang J, Pholwat S, Stroup S, Platts-Mills JA, et al. Effect of biannual azithromycin to children under 5 years on the carriage of respiratory pathogens among children aged 7–11 years. *Am J Trop Med Hyg* 2023;108:428–32.
142. El-Ashker M, Monecke S, Gwida M, Saad T, El-Gohary A, Mohamed A, et al. Molecular characterisation of methicillin-resistant and methicillin-susceptible *Staphylococcus aureus* clones isolated from healthy dairy animals and their caretakers in Egypt. *Vet Microbiol* 2022;267:109374.
143. Kapali S, Pokhrel A, Bastola A, Tuladhar R, Joshi DR. Methicillin-resistant *Staphylococcus aureus* nasal colonization in people living with HIV and healthy people in Kathmandu, Nepal. *Future Sci OA* 2022;8:769.
144. Kotey FC, Awugah SA, Dayie NT, Tetteh-Quarcoo PB, Duodu S, Osei MM, et al. High prevalence of methicillin-resistant *Staphylococcus aureus* carriage among infants at the Children's Hospital, Accra, Ghana. *J Infect Dev Ctries* 2022;16:1450–7.
145. Msanga DR, Silago V, Massoza T, Kidenya BR, Balandya E, Mirambo MM, et al. High fecal carriage of multidrug resistant bacteria in the community among children in Northwestern Tanzania. *Pathogens* 2022;11:379.
146. Naimi HM, Tristan A, Bes M, Vandenesch F, Nazari QA, Laurent F, Dupieux C. Molecular characterization and antimicrobial resistance of nasal *Staphylococcus aureus* in the community of Kabul. *J Glob Antimicrob Resist* 2023;34:18–22.
147. Oginni IO, Olayinka AA. Distribution and antibiotics resistance pattern of community-acquired methicillin-resistance *Staphylococcus aureus* in Southwestern Nigeria. *Adv Exp Med Biol* 2022;1369:81–91.
148. Onyeka FI, Nwobodo DC, Umenne IC, et al. Antibiotic resistance pattern of *Staphylococcus aureus* isolated from nostrils of healthy undergraduates of Madonna University Elele Campus, Rivers State, Nigeria. *Microbes Infect Dis* 2021;2:280–5.
149. Parthasarathy AK, Babu RD, Chougale RA. Prevalence of methicillin resistant *Staphylococcus aureus* and its associated SCCmec types among healthcare workers and patient visitors from Western Maharashtra, India. *J Pure Appl Microbiol* 2022;16:834–40.
150. Paudel G, Amatya N, Saud B, Wagle S, Shrestha V, Adhikari B. Nasal colonization by potential bacterial pathogens in healthy kindergarten children of Nepal: a prevalence study. *Germs* 2022;12:86–98.
151. Saber T, Samir M, El-Mekkawy RM, Ariny E, El-Sayed SR, Enan G, et al. Methicillin- and vancomycin-resistant *Staphylococcus aureus* from humans and ready-to-eat meat: characterization of antimicrobial resistance and biofilm formation ability. *Front Microbiol* 2021;12:735494.
152. Tibebu L, Belete Y, Tigabu E, Tsegaye W. Prevalence of *Staphylococcus aureus*, methicillin-resistant *Staphylococcus aureus* and potential risk factors in selected dairy farms at the interface of animal and human in Bishoftu, Ethiopia. *Vet Med* 2021;12:241–51.
153. Ikuta KS, Swetschinski LR, Robles Aguilar G, Sharara F, Mestrovic T, Gray AP, et al. Global mortality associated with 33 bacterial pathogens in 2019: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet* 2022;400:2221–48.
154. Murray CJL, Ikuta KS, Sharara F, Swetschinski LR, Robles Aguilar G, Gray A, et al. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *Lancet* 2022;399:629–55.
155. Wu M, Tong X, Liu S, Wang D, Wang L, Fan H. Prevalence of methicillin-resistant *Staphylococcus aureus* in healthy Chinese population: a system review and meta-analysis. *PLoS One* 2019;14:e0223599.
156. Reta A, Mengist A, Tesfahun A. Nasal colonization of methicillin resistant *Staphylococcus aureus* in Ethiopia: a systematic review and meta-analysis. *Ann Clin Microbiol Antimicrob* 2019;18:25.
157. Awulachew E, Diriba K, Anja A, Wudneh F. Nasopharyngeal carriage of *Staphylococcus aureus* and its antimicrobial resistance pattern among healthy people: systematic review and meta-analysis. *J Bacteriol Parasitol* 2020;11:1–7.
158. Abdullahe IN, Lozano C, Ruiz-Ripa L, Fernández-Fernández R, Zarazaga M, Torres C. Ecology and genetic lineages of nasal *Staphylococcus aureus* and MRSA carriage in healthy persons with or without animal-related occupational risks of colonization: a review of global reports. *Pathogens* 2021;10:1000.
159. Yang L, Dharmaratne P, Zhu C, Sapugahawatte DN, Rahman N, Barua N, et al. Global epidemiology of asymptomatic colonisation of methicillin-resistant *Staphylococcus aureus* in the upper respiratory tract of young children: a systematic review and meta-analysis. *Arch Dis Child* 2024;109:267–74.
160. Al-lede M, Ayyad DM, Etoom RA, Aldameiry RH, Toubasi AA. The prevalence and risk factors of methicillin-resistant *Staphylococcus aureus* among pediatric populations: a systematic review and meta-analysis. *Eur J Pediatr* 2024;183:3679–87.
161. den Heijer CD, van Bijnen EM, Paget WJ, Pringle M, Goossens H, Bruggeman CA, et al. Prevalence and resistance of commensal *Staphylococcus aureus*, including methicillin-resistant *S. aureus*, in nine European countries: a cross-sectional study. *Lancet Infect Dis* 2013;13:409–15.

162. Gorwitz RJ, Kruszon-Moran D, McAllister SK, McQuillan G, McDougal LK, Fosheim GE, et al. Changes in the prevalence of nasal colonization with *Staphylococcus aureus* in the United States, 2001–2004. *J Infect Dis* 2008;197:1226–34.
163. Diekema DJ, Pfaller MA, Shortridge D, Zervos M, Jones RN. Twenty-year trends in antimicrobial susceptibilities among *Staphylococcus aureus* from the SENTRY Antimicrobial Surveillance Program. *Open Forum Infect Dis* 2019;6:S47–53.
164. Shariati A, Dadashi M, Moghadam MT, van Belkum A, Yaslianifard S, Darban-Sarokhalil D. Global prevalence and distribution of vancomycin resistant, vancomycin intermediate and heterogeneously vancomycin intermediate *Staphylococcus aureus* clinical isolates: a systematic review and meta-analysis. *Sci Rep* 2020;10:12689.
165. Inagaki K, Lucar J, Blackshear C, Hobbs CV. Methicillin-susceptible and methicillin-resistant *Staphylococcus aureus* bacteraemia: nationwide estimates of 30-day readmission, in-hospital mortality, length of stay, and cost in the United States. *Clin Infect Dis* 2019;69:2112–8.
166. Yaw LK, Robinson JO, Ho KM. A comparison of long-term outcomes after methicillin-resistant and methicillin-sensitive *Staphylococcus aureus* bacteraemia: an observational cohort study. *Lancet Infect Dis* 2014;14:967–75.
167. Cole AL, Schmidt-Owens M, Beavis AC, et al. Cessation from smoking improves innate host defense and clearance of experimentally inoculated nasal *Staphylococcus aureus*. *Infect Immun* 2018;86:e00912–7.
168. Olsen K, Falch BM, Danielsen K, Johannessen M, Ericson Sollid JU, Thune I, et al. *Staphylococcus aureus* nasal carriage is associated with serum 25-hydroxyvitamin D levels, gender and smoking status. The Tromsø Staph and Skin Study. *Eur J Clin Microbiol Infect Dis* 2012;31:465–73.
169. Lee AS, de Lencastre H, Garau J, Kluytmans J, Malhotra-Kumar S, Peschel A, Harbarth S. Methicillin-resistant *Staphylococcus aureus*. *Nat Rev Dis Prim* 2018;4:1–23.
170. Shallcross LJ, Fragaszy E, Johnson AM, Hayward AC. The role of the Panton-Valentine leucocidin toxin in staphylococcal disease: a systematic review and meta-analysis. *Lancet Infect Dis* 2013;13:43–54.
171. Román F, Mendez-Echevarria A, Del Rosal T, Garcia-Vera C, Escosa-Garcia L, Agud M, et al. Characterization of methicillin-resistant *Staphylococcus aureus* strains colonizing the nostrils of Spanish children. *Microbiologyopen* 2021;10:e1235.
172. Becker K, Schaumburg F, Fegeler C, Friedrich AW, Köck R. *Staphylococcus aureus* from the German general population is highly diverse. *Int J Med Microbiol* 2017;307:21–7.
173. Grundmann H, Schouls LM, Aanensen DM, Pluister GN, Tami A, Chlebowicz M, et al. The dynamic changes of dominant clones of *Staphylococcus aureus* causing bloodstream infections in the European region: results of a second structured survey. *Eur Surveill* 2014;19:20987.
174. Lawal OU, Ayobami O, Abouelfetouh A, Mourabit N, Kaba M, Egyir B, et al. A 6-year update on the diversity of methicillin-resistant *Staphylococcus aureus* clones in Africa: a systematic review. *Front Microbiol* 2022;13:860436.
175. Schaumburg F, Alabi AS, Peters G, Becker K. New epidemiology of *Staphylococcus aureus* infection in Africa. *Clin Microbiol Infect* 2014;20:589–96.
176. Monecke S, Coombs G, Shore AC, Coleman DC, Akpaka P, Borg M, et al. A field guide to pandemic, epidemic and sporadic clones of methicillin-resistant *Staphylococcus aureus*. *PLoS One* 2011;6:e17936.
177. Goswami C, Fox S, Holden M, Leanord A, Evans TJ. Genomic analysis of global *Staphylococcus argenteus* strains reveals distinct lineages with differing virulence and antibiotic resistance gene content. *Front Microbiol* 2021;12:795173.