



This is a repository copy of *A laboratory-scale physical model for freeze–thaw studies in soil columns under simulated climate change conditions.*

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/223606/>

Version: Accepted Version

---

**Article:**

Sahoo, M. [orcid.org/0000-0003-3552-4691](https://orcid.org/0000-0003-3552-4691), Bentley, P., Smith, A. et al. (4 more authors) (2025) A laboratory-scale physical model for freeze–thaw studies in soil columns under simulated climate change conditions. *Environmental Science and Pollution Research*, 32 (9). pp. 5293-5301. ISSN 0944-1344

<https://doi.org/10.1007/s11356-025-36053-8>

---

© The Author(s). Except as otherwise noted, this author-accepted version of a journal article published in *Environmental Science and Pollution Research* is made available via the University of Sheffield Research Publications and Copyright Policy under the terms of the Creative Commons Attribution 4.0 International License (CC-BY 4.0), which permits unrestricted use, distribution and reproduction in any medium, provided the original work is properly cited. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>

**Reuse**

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:

<https://creativecommons.org/licenses/>

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>



29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53

## Abstract

Responses of soils to climate warming during winter can be studied by monitoring soil temperature variations in the vadose zone. Freeze-thaw cycles during winter can have a significant impact on soil biogeochemical and physical processes. Observing soil temperature at different depths offers important insights into heat availability, which influences biogeochemical activity and solute movement driven by temperature gradients. However, it is difficult to replicate the relevant processes in soil columns without maintaining freezing temperatures with an unfrozen soil layer below the freezing interface. This paper describes the development and experimental verification of a laboratory-scale physical model to assess the effect of freeze-thaw cycles on biogeochemical processes in unsaturated soil. The results show that the experimental design can (i) induce cyclical freezing of soil down to desirable depths, (ii) maintain vertical temperature gradients and (iii) ensure the rest of the column remains unfrozen below the freezing interface for pore water sampling. The rate of freezing is suitable for quick freeze-thaw cycles (0.19°C/minute). This setup can be further developed to observe solute transport and attenuation in variably-saturated soil and soil moisture status for different freeze-thaw regimes under simulated climate change scenarios.

*Keywords* – Freeze-thaw cycle, freezing jacket, soil column, soil temperature, unsaturated soil

## 54 1. INTRODUCTION

55 Freeze-thaw cycles (FTCs) are increasingly recognized as a critical process  
56 influencing soil systems in temperate and high-latitude regions. Warming winters and an  
57 increase in snow-free days, driven by climate change, have resulted in frequent, intense, and  
58 rapid FTCs these regions (Kreyling et al., 2008; Schmitt et al., 2008; Sang et al., 2021). Soil  
59 FTCs can affect the movement of solutes (Man et al., 2022; Xu, 2022; Zong et al., 2023),  
60 pore and soil structure (Leuther and Schlüter, 2021; Miranda-Velez et al., 2023; Zhang et  
61 al., 2021), and plant and microbial activity (Koponen et al., 2006; Kreyling et al., 2010;  
62 Sorensen et al., 2018). FTCs have been found to increase nutrient fluxes from soil, which  
63 can be lost via leaching or as upward accumulation on soil surface (Yanai et al., 2004;  
64 Hentschel et al., 2008; Matzner and Borken 2008; Gao et al., 2018; Sun et al., 2021; Liu et  
65 al., 2023). Understanding factors which affect the development of FTCs in variably-  
66 saturated soil and simulating future FTC patterns is therefore important in managing  
67 potential impacts of nutrient loads and pollutant export from to soil and water resources  
68 under these conditions.

69 Observation of FTCs in the field is difficult, due to uncertainties associated with the  
70 occurrence and frequency of these phenomena. Long-term seasonal monitoring over several  
71 years provides important observations at the catchment scale (Zhang et al., 2022; Liu et al.,  
72 2023), but it is unrealistic to use these observations for accurate simulation and prediction.  
73 Laboratory-based studies have been undertaken to overcome the uncertainties related to  
74 field studies, by collecting soil samples from a field site and inducing FTCs in a controlled-  
75 temperature environment (Sharma et al., 2006; Vestgarden and Austnes, 2009; Stres et al.,  
76 2010; Li et al., 2024). FTCs in soil can be induced by keeping the samples (e.g. undisturbed  
77 soil cores) in a temperature-controlled cold room (Rooney et al., 2022), incubators  
78 (Koponen and Martikainen, 2004; Wu et al., 2014), or laboratory freezers (Sharma et al.,

79 2006). Soil cores were packed in PVC cylinders or glass jars and kept in controlled-  
80 temperature environments. Arrangements for different chemical sampling were made  
81 destructively, i.e., air-drying and sieving, water extraction for chemical analyses (Wu et al.,  
82 2014; Wu et al., 2021; Rooney et al., 2022) or non-destructively for microbial sampling  
83 (Sharma et al., 2006), at the end of the freezing/thawing. This procedure, however, cannot  
84 be used for instantaneous measurements of soil temperature and moisture movement. Soil  
85 column experiments which can be used for unsaturated freezing-thawing experiments are  
86 rare. To overcome these limitations, soil column experiments have been conducted to  
87 experimentally simulate field conditions (Fitzhugh et al., 2001; Hermansson, 2004; Evirgen  
88 and Tuncan, 2019; Xu et al., 2019; Libby et al., 2020). Fitzhugh et al. (2001) and Libby et  
89 al., (2020) attempted to simulate field conditions using soil columns in outdoor and indoor  
90 sites, respectively. The experiment by Hermansson (2004) experiment involved freezing a  
91 soil column from the top and observing the frost penetration. However, this experiment, is  
92 not reliable due to poor reproducibility and may result in inadequate freezing depths. The  
93 experiment by Xu et al. (2019) involved circulating warm and cold fluids at the top and  
94 bottom of a soil column to develop FTCs. Evirgen and Tuncan's (2019) experiment  
95 circulated coolant fluid through pipes which were buried within soil to freeze it. The  
96 objectives for each experiment were different from the objectives of the present study. The  
97 main objective of the present study was to develop a mechanism for freezing/thawing  
98 experiments which can be useful in studying unsaturated soil behaviour (both chemically  
99 and physically) during quick and frequent FTCs. Previous experimental designs used to  
100 investigate FTCs in unsaturated soil have important limitations, in that (i) they are unsuitable  
101 to create rapid FTCs (as is observed in the field), (ii) conducted at unrealistically low (< -  
102 10°C), and (iii) they cannot create a warmer soil mass beneath the frozen soil zone, as occurs  
103 naturally. Topsoil temperatures <-10°C are not observed during quick and frequent FTCs in

104 temperate regions. Existing studies using laboratory freezers typically use a minimum  
105 temperature of -20°C, thus killing soil organisms responsible for microbial activity during  
106 FTCs at deeper levels (Henry, 2007). As not all soil water freezes during winter, active solute  
107 transport may still occur through the soil profile, driven by the developed temperature  
108 gradients. Therefore, maintaining a warm soil zone beneath the frozen layer is important to  
109 observe instantaneous changes occurring during FTCs in soil.

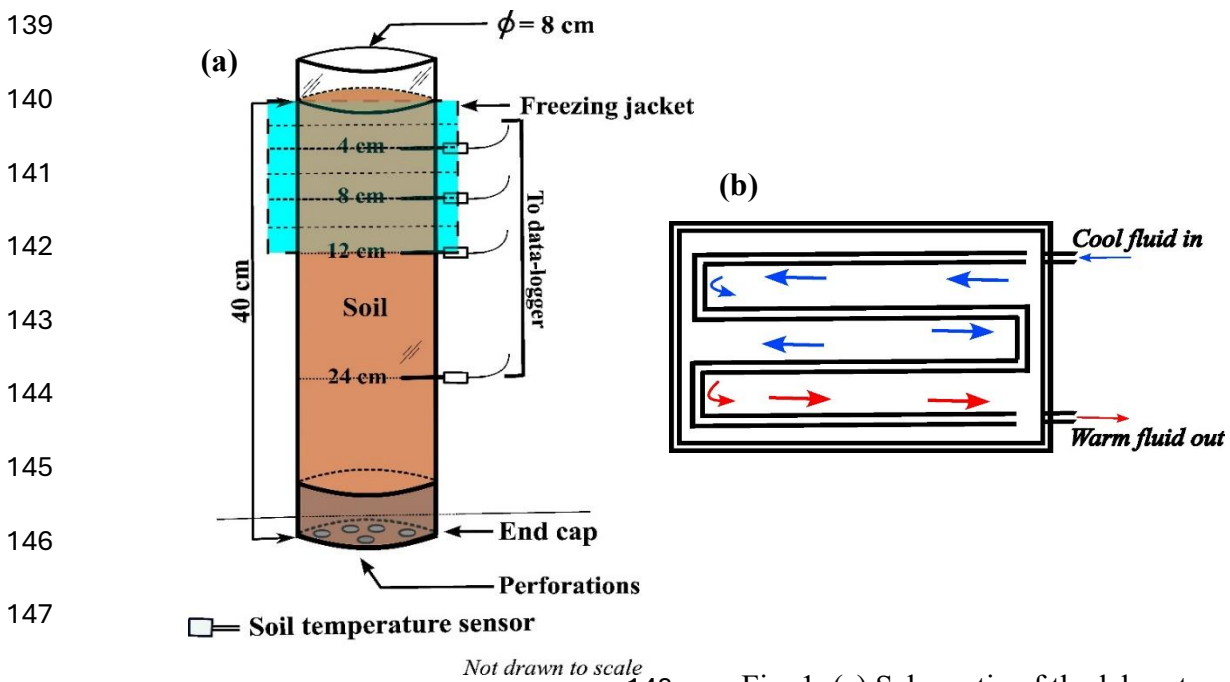
110 The present study describes the development of a novel laboratory-scale physical  
111 model to experimentally determine the effect of FTCs on biogeochemical and solute  
112 transport processes in variably-saturated soil under simulated climate change conditions.  
113 The specific objectives of the research are to: (i) develop the experimental model to maintain  
114 freezing temperatures and unfrozen soil simultaneously in a soil column, and (ii) verify the  
115 performance of the physical model in replicating FTC propagation and temperature  
116 gradients in soil columns.

## 117 2. MATERIALS AND METHODS

### 118 2.1 Conceptual design of physical model

119 Soil acts as a heat reservoir and can retain heat during winter to support microbial activity  
120 at deeper depths (Li et al., 2020; Sahoo, 2022). The effective plant and microbial activity  
121 can be found within the top 30 cm of soil (Hao et al., 2021). Soils across the temperate zone  
122 and high latitudes experience multiple and quick freeze-thaw cycles during winter  
123 (Kreyling, 2010). Frequent FTCs can have a significant impact on solute movement in soil,  
124 which may result in leaching of soil nutrients to groundwater (Matzner and Borken, 2008),  
125 upward accumulation of solutes on soil surface (Liu et al., 2022), and seasonal elevation of  
126 nutrient content in stream water during early spring (Creed et al., 1996; Chen et al., 2020).  
127 Observing the movement of solutes in unsaturated soil due to imposed temperature gradients  
128 can be challenging at the field-scale due to uncertainties related to replication. A simplified

129 laboratory-scale physical model was conceptualized to replicate FTCs in a soil column and  
 130 changes in critical soil physical-chemical properties resulting from them. Assuming an  
 131 average root-zone soil depth of 100 cm and active depth of soil microbial activity as 30 cm,  
 132 the occurrence FTCs was hypothesized within the top 30 cm soil depth. An efficient freezing  
 133 mechanism which can freeze only the top 30 cm and create precise temperature gradients  
 134 within the soil profile, while ensuring the remaining depth remains unfrozen was necessary.  
 135 A jacket-like mechanism was proposed, which remains wrapped around a soil column  
 136 extracting the heat from the soil down to the desirable depth. With the aim of testing the  
 137 efficiency of a freezing jacket to freeze the soil to a desirable depth, a 1/4<sup>th</sup> scaled-down  
 138 version of laboratory set up was constructed (Fig. 1).



148 Fig. 1. (a) Schematic of the laboratory soil  
 149 column setup, showing temperature gradients (a) soil column; (b) Coolant circulation  
 150 mechanism through the freezing jacket.

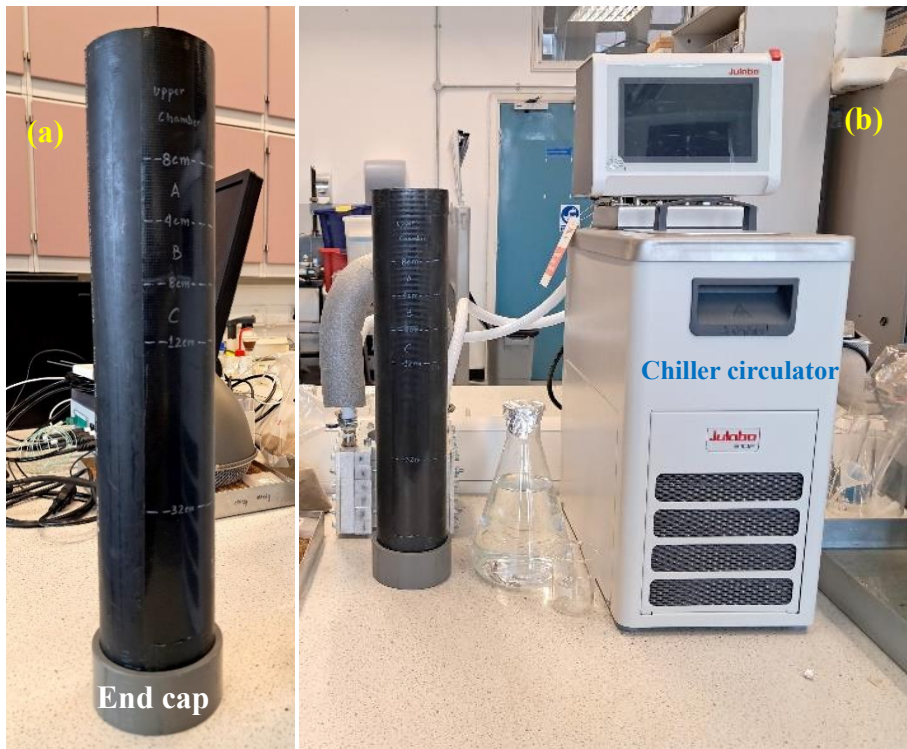
151 At this stage, only the freezing mechanism and freezing efficiency of the jacket was  
 152 tested without adding any other arrangements to observe solute movement. Soil temperature  
 153 sensors (thermocouples [Fig. 1]) were placed within the depths corresponding to the layers

154 of freezing jacket. One thermocouple was installed at a deeper depth of soil column away  
155 from the jacketed layer to test the soil temperature in the unfrozen layer.

## 156 2.2 Soil column

157 An unplasticized polyvinyl chloride (UPVC) tube (8 cm internal diameter and 48 cm  
158 height) was used to construct a soil column (Fig. 2). UPVC has a thermal conductivity of  
159 the same order of magnitude as a natural soil and therefore a weak temperature gradient will  
160 exist between soil and the tube. Lateral soil moisture movement towards the tube is likely

161  
162  
163  
164  
165  
166  
167  
168  
169  
170



171 Fig. 2. (a) UPVC column used for the FT experiments, and (b) laboratory set up for FT  
172 studies.

173 to be negligible (Gray and Granger, 1986), thus inducing a one-dimensional flow within the  
174 column. The side wall of the tube was roughened to restrict any side wall flow (Lewis and  
175 Sjöstrom, 2010). Air-dried soil was packed in the tube by regular tamping to obtain a  
176 uniform packing throughout the column. The soil column was maintained at a height of 40  
177 cm, leaving the upper 8 cm for the accumulation of any water/snow during the FT  
178 experiments. The tube was fitted with a hollow end cap [Fig. 2(a)]. The end cap contained



179 a 100 $\mu$ m stainless steel mesh to retain soil particles, while allowing samples of free draining  
180 water to be collected from the base of the column. The soil total porosity was maintained  
181 between 55 – 57%, with a corresponding bulk density of 1.18 – 1.2 g/cm<sup>3</sup>.

### 182 2.3. Freezing jacket

183 A controlled depth of freezing in a soil column can be achieved by wrapping the  
184 column with a freezing jacket down to the required depth. Ideally, the freezing jacket should  
185 remain tightly held to the column to remove the heat from the soil at any given depth. In the  
186 present study, the freezing jacket was constructed from aluminium alloy for its good heat  
187 conductance, even distribution of heat, and resistance to corrosion [Fig. 3(a)] and a coolant  
188 (ethylene glycol) circulated through it to achieve the desired freezing temperature in soil.  
189 The freezing jacket consists of three detachable layers (each 4 cm in width) for controlling  
190 freezing depth up to 12 cm depth.



191 Fig. 3. (a) Freezing jacket, (b) front-view of freezing jacket with detachable layers, and (c)  
192 internal circuit for coolant circulation.

193 Each layer was further machined to obtain two sub-layers (each 2 cm in width) to  
194 achieve freezing temperatures within the desired depth [Fig. 3(b)]. The internal plates were  
195 fitted with a groove to form a circuit for coolant circulation [Fig. 3(c)]. A chiller circulator  
196 [Fig. 2(b)] was used to circulate chilling fluid (i.e. glycol in the present study) through the  
197 freezing jacket. The detachable layers can be added or removed depending on the required  
198 freezing depth. One or more 4 cm-layers (or two sub-layers together) was either added or

199 removed to test the performance of the jacket across different simulated freezing depths.  
200 The unjacketed part of the soil column should always remain above freezing temperature  
201 during the experiment.

#### 202 **2.4. Soil and ancillary materials**

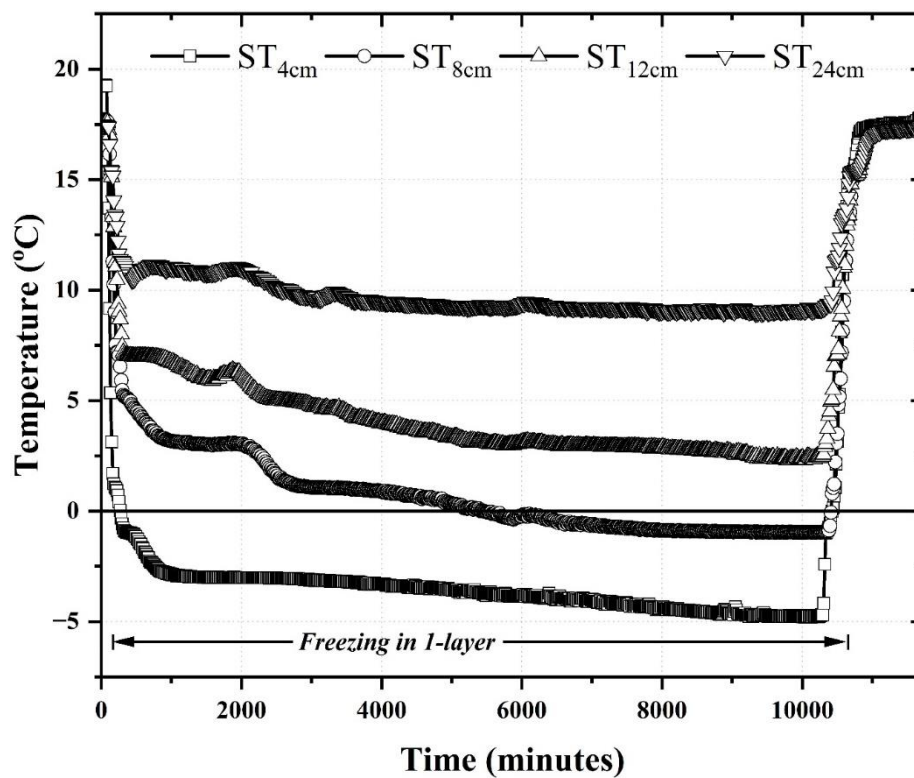
203 Soil samples were collected from an agricultural farm in Wakefield, United Kingdom.  
204 The soil was predominantly weathered sandstone with impermeable layer starting  
205 approximately at 15 cm depth. Large stones, debris or litter material were removed before  
206 the experiment by sieving the soil sample. Screened and homogenized soil from this  
207 preparation was used to create a soil column for good method reproducibility. The sample  
208 has sandy loam texture (71% sand, 18% silt, and 11% clay) and the column contained soil  
209 with particle sizes 3.35 mm or finer in grain size. The soil drains rapidly and has an organic  
210 matter content of 7.2 wt %. The initial soil saturation was completed from the base upwards,  
211 allowing the moisture to wick upwards by capillary action (Lewis and Sjöstrom, 2010).  
212 Subsequent saturation with water was done from the top. Freezing of the soil was started  
213 once the soil reached a soil moisture content of 35 – 40% (field capacity of the soil used). A  
214 constant head permeability test determined the hydraulic conductivity of the soil as 26.36  
215 cm/day.

216 Thermocouples were installed at 4 cm, 8 cm, and 12 cm from the soil surface (three  
217 depths within the full jacketed zone) and 24 cm (unjacketed zone) of the soil column to  
218 observe temperature changes within the soil profile during FT experiments. The  
219 thermocouples monitored and recorded temperature changes within the centre of the column  
220 at 15-minute intervals. The duration of the experiments varied between 1 – 9 days, to  
221 examine the performance of the overall set up for representative and realistic durations of  
222 FTCs. The laboratory set up was operated at a room temperature of 22°C.

223

224 **3. RESULTS AND DISCUSSION**

225 The freezing jacket decreased the soil temperature from 20.4°C to 0.08°C (near  
226 freezing temperature) in 105 minutes (1 hour 45 minutes) at a rate of 0.19°C/minute. The  
227 freezing temperature can be maintained until the chiller circulator begins operating. Once  
228 the chiller circulator was switched off, the soil started thawing at a rate of 0.05°C/minute.  
229 The coolant was circulated through the freezing jacket at -10°C. The chiller circulator could  
230 operate continuously for long durations (up to 7.06 days in the present study), maintaining  
231 freezing temperatures within the jacketed region of the soil column (Fig. 4). Freezing  
232 temperatures beyond the unjacketed region started appearing after 4 days, reaching a  
233 minimum temperature of -0.95°C. The minimum temperature within the jacketed region

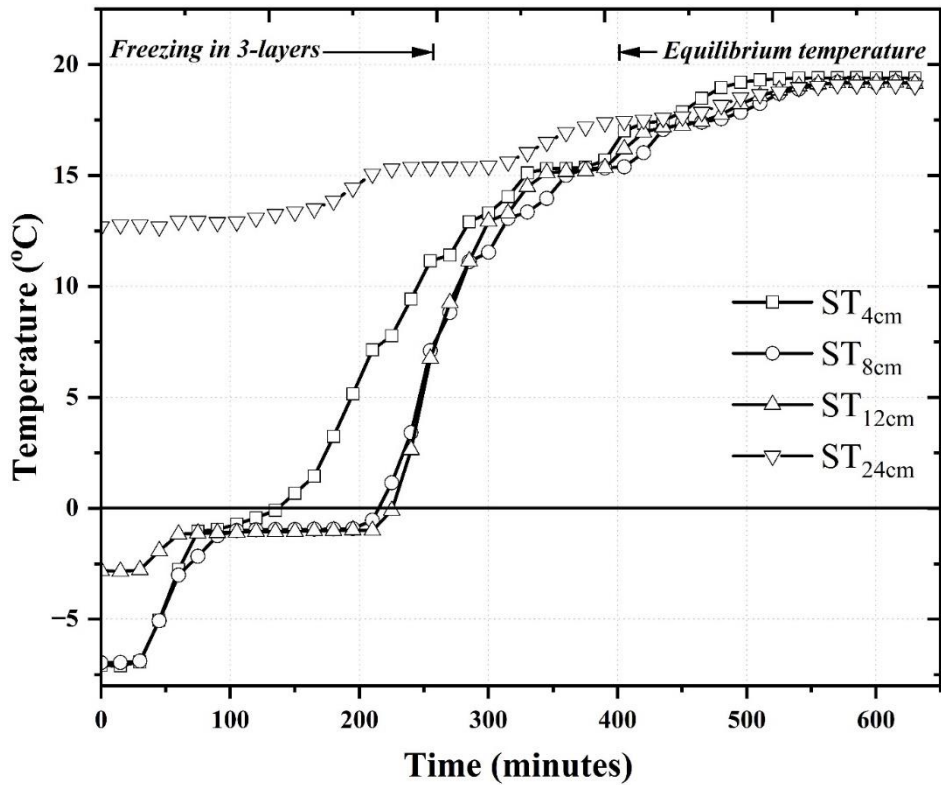


243 Fig. 4. Freezing soil column with 1-layer of jacket (4 cm depth).

244 varied between -7.10°C and -4.79°C. The temperature at 24 cm depth always remained  
245 above +10°C (Fig. 4-6).

246 Temperature gradients occur at different depths in soil at the field-scale (Contosta et  
247 al., 2016; Sahoo, 2022). The freezing jacket was able to maintain a consistent temperature

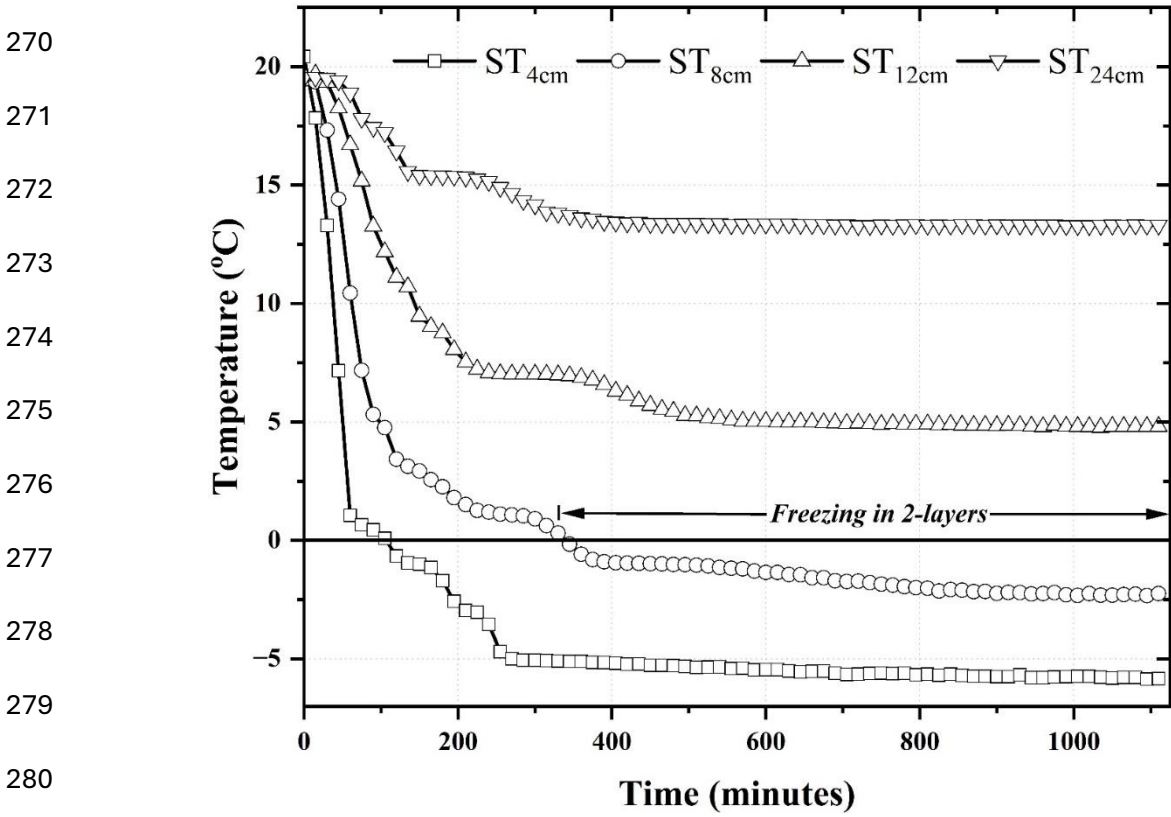
248 difference within the jacketed region for significant time periods (Fig. 4-6). Soil at deeper  
249 depths was warmer than at the soil surface. After thawing, soil temperatures within the entire  
250 column reach an equilibrium temperature (Fig. 5).



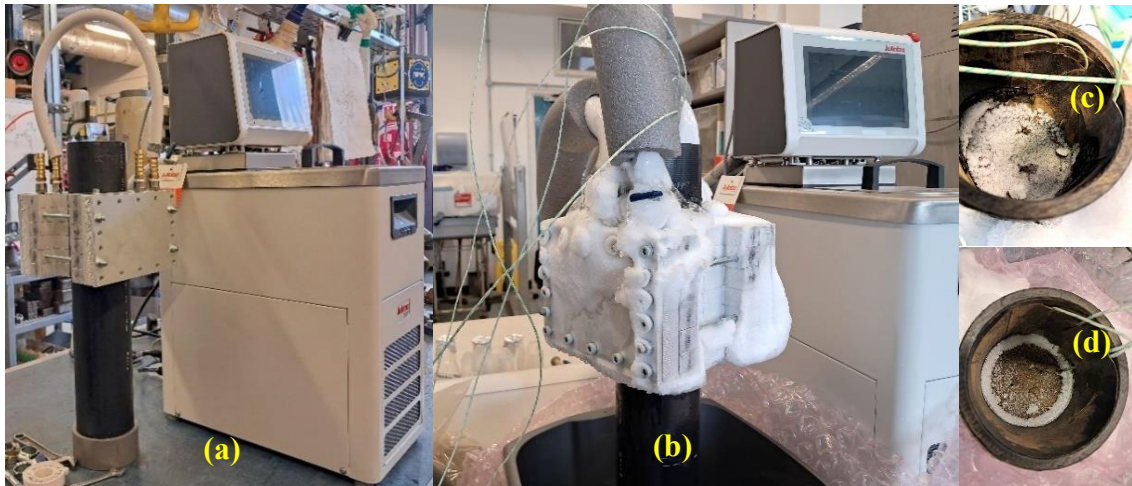
262 Fig. 5. Freezing soil column with 3-layers of jacket (12 cm depth).

263 The efficiency of the jacket in restricting the freezing depth was precise and could be  
264 achieved down to 4 cm depth with each aluminium layer (Fig. 6). The addition or removal  
265 of layers suggests that the freezing jacket is effective in cooling the soil down to the desired  
266 depth and duration and can maintain soil temperature differences within the frozen soil  
267 depth. With changes in the number of layers in the jacket, the amount of accumulated ice on

268 the soil surface is significantly reduced (Fig. 7). This change suggests that the freezing jacket  
269 layers were able to replicate mild and severe winter scenarios at the laboratory-scale.



281 Fig. 6. Freezing soil column with 2-layers of jacket (8 cm depth).



282 Fig. 7. (a) Column with the freezing jacket, (b) frozen column with 3-layers of  
283 jacket, and (c) and (d) accumulation of ice on frozen soil surface, with 3-layers and 1-layer  
284 of jacket, respectively.

285           The unjacketed portion of the soil column was not affected by the environmental  
286 temperature changes. UPVC is a poor conductor and heat does not affect the temperature of  
287 soil within the column. The temperature changes within the unjacketed column were due to  
288 the movement of the freezing front from above the soil column, the heat retained during  
289 packing the column, and the movement of soil water during the saturation stage.

290           Existing methods of chilling soil samples for studies of solute biogeochemistry require  
291 freezing the soil at  $< -10^{\circ}\text{C}$  inside a laboratory refrigerator or chilling incubator (Bochove et  
292 al., 2000; Henry, 2007; Patel et al., 2021; Edwards et al., 2024). However, these low  
293 temperatures are deleterious to soil microorganisms and are not usually encountered in  
294 temperate regions (Henry, 2007). The present setup was able to achieve and maintain  
295 freezing temperatures  $> -10^{\circ}\text{C}$ , that can support biogeochemical reactions within the soil  
296 profile. Soil temperature can be regulated in the chiller circulator to the desired temperature  
297 and maintained consistently over extended periods.

298           Hence the laboratory-scale physical model developed in this study can be used to  
299 reproduce field-scale scenarios of climate change-induced FTCs to understand the effect of  
300 this phenomena on the status and movement of solutes within the soil profile. In particular,  
301 the methodology can enable solute fluxes to surface water and groundwater resources to be  
302 determined for specific climate-change scenarios. This is important to predict and manage  
303 the impact of nutrients and other potential contaminants to these receiving waters from  
304 different sources in catchments.

#### 305   **4. CONCLUSIONS**

306           This study has introduced a novel laboratory-scale physical model designed to  
307 investigate the effects of FTCs on the physico-chemical processes occurring in variably-  
308 saturated soils. The model was used to create temperature gradients arising from FTCs  
309 within a required depth at the laboratory-scale for extended periods. This setup can be used

310 for experiments which consider the influence of climate change on FTCs and which require  
311 the creation of precise *in situ* soil temperature variations with depth. Modelling of  
312 unsaturated zone processes are essentially considered as one-dimensional unless stated  
313 otherwise. This setup was tested successfully for unsaturated soil processes and can be  
314 further used to examine soil moisture distribution and solute transport induced by FTCs (or  
315 winter warming) and the effect of this phenomena on solute (e.g. nutrient and potential  
316 pollutant) fluxes to surface water and groundwater. In contrast to soil freezing, the jacket  
317 can be used for soil heating experiments (or to increase soil temperature within specific  
318 depths) by circulating warm fluid in a similar way. This jacket-column arrangement can be  
319 used for one column at a time, therefore, studies which require simultaneous testing of  
320 multiple samples may require multiple jackets for each soil column. Further studies will be  
321 undertaken with this methodology, using a scaled-up version of the column and freezing  
322 jacket, with sensor arrangements to observe solute transport and other biogeochemical  
323 processes induced by FTCs under simulate climate-change scenarios.

324

## 325 REFERENCES

- 326 Chen, X., Tague, C.L., Melack, J.M., Keller, A.A., 2020. Sensitivity of nitrate  
327 concentration-discharge patterns to soil nitrate distribution and drainage  
328 properties in the vertical dimension. *Hydrological Process.* 34 (11), 2477–2493.  
329 <https://doi.org/10.1002/hyp.13742>.
- 330 Contosta, A. R., Burakowski, E. A., Varner, R. K., and Frey, S. D., 2016. Winter soil  
331 respiration in a humid temperate forest: The roles of moisture, temperature, and  
332 snowpack. *Journal of Geophysical Research: Biogeosciences*, 121(12), pp.3072  
333 – 3088.

334 Creed, I.F., Band, L.E., Foster, N.W., Morrison, I.K., Nicolson, J.A., Semkin, R.S.,  
335 Jeffries, D.S., 1996. Regulation of Nitrate-N Release from Temperate Forests:  
336 A Test of the N Flushing Hypothesis. *Water Resources Research*. 32 (11), 3337–  
337 3354. <https://doi.org/10.1029/96WR02399>

338 Edwards, J.D., Love, S.J., Phillips, R.P., Fei, S., Domke, G., Parker, J.D., McCormick,  
339 M., LaRue, E.A., Schweitzer, J.A., Bailey, J.K. and Fordyce, J., 2024. Long-and  
340 short-term soil storage methods other than freezing can be useful for DNA-  
341 based microbial community analysis. *Soil Biology and Biochemistry*, 191,  
342 p.109329. <https://doi.org/10.1016/j.soilbio.2024.109329>

343 Evirgen, B. and Tuncan, M., 2019. A physical soil freezing model for laboratory  
344 applications. *Cold Regions Science and Technology*, 159, pp.29-39.  
345 <https://doi.org/10.1016/j.coldregions.2018.12.005>

346 Fitzhugh, R.D., Driscoll, C.T., Groffman, P.M., Tierney, G.L., Fahey, T.J. and Hardy,  
347 J.P., 2001. Effects of soil freezing disturbance on soil solution nitrogen,  
348 phosphorus, and carbon chemistry in a northern hardwood ecosystem.  
349 *Biogeochemistry*, 56, pp.215-238. <https://doi.org/10.1023/A:1013076609950>

350 Gao, D., Zhang, L., Liu, J., Peng, B., Fan, Z., Dai, W., Jiang, P. and Bai, E., 2018.  
351 Responses of terrestrial nitrogen pools and dynamics to different patterns of  
352 freeze-thaw cycle: A meta-analysis. *Global Change Biology*, 24(6), pp.2377-  
353 2389. <https://doi.org/10.1111/gcb.14010>

354 Gray, D.M. and Granger, R.J., 1986. In situ measurements of moisture and salt  
355 movement in freezing soils. *Canadian Journal of Earth Sciences*, 23(5), pp.696-  
356 704. <https://cdnsiencepub.com/doi/pdf/10.1139/e86-069>

357 Hao, J., Chai, Y.N., Lopes, L.D., Ordóñez, R.A., Wright, E.E., Archontoulis, S. and  
358 Schachtman, D.P., 2021. The effects of soil depth on the structure of microbial



359 communities in agricultural soils in Iowa (United States). Applied and  
360 Environmental Microbiology, 87(4), pp. e02673-20.  
361 <https://journals.asm.org/doi/10.1128/aem.02673-20>

362 Henry, H.A., 2007. Soil freeze–thaw cycle experiments: trends, methodological  
363 weaknesses and suggested improvements. Soil Biology and Biochemistry,  
364 39(5), pp.977-986. <https://doi.org/10.1016/j.soilbio.2006.11.017>

365 Hentschel, K., Borken, W. and Matzner, E., 2008. Repeated freeze–thaw events affect  
366 leaching losses of nitrogen and dissolved organic matter in a forest soil. Journal  
367 of Plant Nutrition and Soil Science, 171(5), pp.699-706.  
368 <https://doi.org/10.1002/jpln.200700154>

369 Hermansson, Å., 2004. Laboratory and field testing on rate of frost heave versus heat  
370 extraction. Cold Regions Science and Technology, 38(2-3), pp.137-151.  
371 <https://doi.org/10.1016/j.coldregions.2003.10.002>

372 Koponen, H.T. and Martikainen, P.J., 2004. Soil water content and freezing temperature  
373 affect freeze–thaw related N<sub>2</sub>O production in organic soil. Nutrient Cycling in  
374 Agroecosystems, 69, pp.213-219.  
375 <https://link.springer.com/article/10.1023/B:FRES.0000035172.37839.24>

376 Koponen, H.T., Jaakkola, T., Keinänen-Toivola, M.M., Kaipainen, S., Tuomainen, J.,  
377 Servomaa, K. and Martikainen, P.J., 2006. Microbial communities, biomass, and  
378 activities in soils as affected by freeze thaw cycles. Soil Biology and  
379 Biochemistry, 38(7), pp.1861-1871.  
380 <https://doi.org/10.1016/j.soilbio.2005.12.010>

381 Kreyling, J., 2010. Winter climate change: a critical factor for temperate vegetation  
382 performance. Ecology, 91(7), pp.1939-1948. <https://doi.org/10.1890/09-1160.1>

383 Kreyling, J., Beierkuhnlein, C. and Jentsch, A., 2010. Effects of soil freeze–thaw cycles  
384 differ between experimental plant communities. *Basic and Applied Ecology*,  
385 11(1), pp.65-75. <https://doi.org/10.1016/j.baae.2009.07.008>

386 Kreyling, J., Beierkuhnlein, C., Pritsch, K., Schloter, M. and Jentsch, A., 2008.  
387 Recurrent soil freeze–thaw cycles enhance grassland productivity. *New*  
388 *Phytologist*, 177(4), pp.938-945. [https://doi.org/10.1111/j.1469-](https://doi.org/10.1111/j.1469-8137.2007.02309.x)  
389 [8137.2007.02309.x](https://doi.org/10.1111/j.1469-8137.2007.02309.x)

390 Leuther, F. and Schlüter, S., 2021. Impact of freeze–thaw cycles on soil structure and  
391 soil hydraulic properties. *Soil*, 7(1), pp.179-191. [https://doi.org/10.5194/soil-7-](https://doi.org/10.5194/soil-7-179-2021)  
392 [179-2021](https://doi.org/10.5194/soil-7-179-2021)

393 Lewis, J. and Sjöström, J., 2010. Optimizing the experimental design of soil columns  
394 in saturated and unsaturated transport experiments. *Journal of Contaminant*  
395 *Hydrology*, 115(1-4), pp.1-13. <https://doi.org/10.1016/j.jconhyd.2010.04.001>

396 Li, B., Norouzi, E., Zhu, H.H. and Wu, B., 2024. A thermo-poromechanical model for  
397 simulating freeze–thaw actions in unsaturated soils. *Advances in Water*  
398 *Resources*, 184, p.104624. <https://doi.org/10.1016/j.advwatres.2024.104624>

399 Li, Z., Zeng, Z., Tian, D., Wang, J., Wang, B., Chen, H.Y., Quan, Q., Chen, W., Yang,  
400 J., Meng, C. and Wang, Y., 2020. Global variations and controlling factors of  
401 soil nitrogen turnover rate. *Earth-science reviews*, 207, p.103250.  
402 <https://doi.org/10.1016/j.earscirev.2020.103250>

403 Libby, M.D., VanderZaag, A.C., Gregorich, E.G. and Wagner-Riddle, C., 2020. An  
404 improved laboratory method shows that freezing intensity increases N<sub>2</sub>O  
405 emissions. *Canadian Journal of Soil Science*, 100(2), pp.136-149.  
406 <https://doi.org/10.1139/cjss-2019-0073>

407 Liu, S., Zhou, Z., Liu, J., Wang, K., Li, J., Wang, P., Xie, X., Jia, Y. and Wang, H., 2023.  
408 Simulation of water and nitrogen movement mechanism in cold regions during  
409 freeze–thaw period based on a distributed nonpoint source pollution model  
410 closely coupled water, heat, and nitrogen processes at the watershed scale.  
411 Environmental Science and Pollution Research, 30(3), pp.5931-5954.  
412 <https://link.springer.com/article/10.1007/s11356-022-22535-6>

413 Man, Z., Xie, C., Jiang, R. and Che, S., 2022. Freeze–thaw cycle frequency affects root  
414 growth of alpine meadow through changing soil moisture and nutrients.  
415 Scientific Reports, 12(1), p.4436. [https://www.nature.com/articles/s41598-022-](https://www.nature.com/articles/s41598-022-08500-w)  
416 [08500-w](https://www.nature.com/articles/s41598-022-08500-w)

417 Matzner, E. and Borken, W., 2008. Do freeze-thaw events enhance C and N losses from  
418 soils of different ecosystems? A review. European Journal of Soil Science, 59(2),  
419 pp.274-284. <https://doi.org/10.1111/j.1365-2389.2007.00992.x>

420 Miranda-Velez, J.F., Leuther, F., Koehne, J.M., Munkholm, L.J. and Vogeler, I., 2023.  
421 Effects of freeze-thaw cycles on soil structure under different tillage and plant  
422 cover management practices. Soil and Tillage Research, 225, p.105540.  
423 <https://doi.org/10.1016/j.still.2022.105540>

424 Patel, K.F., Tatariw, C., MacRae, J.D., Ohno, T., Nelson, S.J. and Fernandez, I.J., 2021.  
425 Repeated freeze–thaw cycles increase extractable, but not total, carbon and  
426 nitrogen in a Maine coniferous soil. Geoderma, 402, p.115353.  
427 <https://doi.org/10.1016/j.geoderma.2021.115353>

428 Rooney, E.C., Bailey, V.L., Patel, K.F., Possinger, A.R., Gallo, A.C., Bergmann, M.,  
429 SanClements, M. and Lybrand, R.A., 2022. The Impact of Freeze-Thaw History  
430 on Soil Carbon Response to Experimental Freeze-Thaw Cycles. Journal of

431 Geophysical Research: Biogeosciences, 127(5), p. e2022JG006889.  
432 <https://doi.org/10.1029/2022JG006889>

433 Sahoo, M., 2022. Winter soil temperature and its effect on soil nitrate Status: A Support  
434 Vector Regression-based approach on the projected impacts. *Catena*, 211,  
435 p.105958. <https://doi.org/10.1016/j.catena.2021.105958>

436 Sang, C., Xia, Z., Sun, L., Sun, H., Jiang, P., Wang, C. and Bai, E., 2021. Responses of  
437 soil microbial communities to freeze–thaw cycles in a Chinese temperate forest.  
438 *Ecological Processes*, 10(1), pp.1-18.  
439 <https://link.springer.com/article/10.1186/s13717-021-00337-x>

440 Schmitt, A., Glaser, B., Borken, W. and Matzner, E., 2008. Repeated freeze–thaw cycles  
441 changed organic matter quality in a temperate forest soil. *Journal of Plant  
442 Nutrition and Soil Science*, 171(5), pp.707-718.  
443 <https://doi.org/10.1002/jpln.200700334>

444 Sharma, S., Szele, Z., Schilling, R., Munch, J.C. and Schloter, M., 2006. Influence of  
445 freeze-thaw stress on the structure and function of microbial communities and  
446 denitrifying populations in soil. *Applied and Environmental Microbiology*,  
447 72(3), pp.2148-2154. <https://doi.org/10.1128/AEM.72.3.2148-2154.2006>

448 Sorensen, P.O., Finzi, A.C., Giasson, M.A., Reinmann, A.B., Sanders-DeMott, R. and  
449 Templer, P.H., 2018. Winter soil freeze-thaw cycles lead to reductions in soil  
450 microbial biomass and activity not compensated for by soil warming. *Soil  
451 Biology and Biochemistry*, 116, pp.39-47.  
452 <https://doi.org/10.1016/j.soilbio.2017.09.026>

453 Stres, B., Philippot, L., Faganeli, J. and Tiedje, J.M., 2010. Frequent freeze–thaw cycles  
454 yield diminished yet resistant and responsive microbial communities in two

455                   temperate soils: a laboratory experiment. *FEMS Microbiology ecology*, 74(2),  
456                   pp.323-335. <https://doi.org/10.1111/j.1574-6941.2010.00951.x>

457                   Sun, L., Chang, X., Yu, X., Jia, G., Chen, L., Wang, Y. and Liu, Z., 2021. Effect of  
458                   freeze-thaw processes on soil water transport of farmland in a semi-arid area.  
459                   *Agricultural Water Management*, 252, p.106876.

460                   van Bochove, E., Prévost, D. and Pelletier, F., 2000. Effects of freeze–thaw and soil  
461                   structure on nitrous oxide produced in a clay soil. *Soil Science Society of*  
462                   *America Journal*, 64(5), pp.1638-1643.  
463                   <https://doi.org/10.2136/sssaj2000.6451638x>

464                   Vestgarden, L.S. and Austnes, K., 2009. Effects of freeze–thaw on C and N release from  
465                   soils below different vegetation in a montane system: a laboratory experiment.  
466                   *Global Change Biology*, 15(4), pp.876-887.  
467                   <https://onlinelibrary.wiley.com/doi/epdf/10.1111/j.1365-2486.2008.01722.x>

468                   Wu, H., Xu, X., Fu, P., Cheng, W. and Fu, C., 2021. Responses of soil WEOM quantity  
469                   and quality to freeze–thaw and litter manipulation with contrasting soil water  
470                   content: A laboratory experiment. *Catena*, 198, p.105058.  
471                   <https://doi.org/10.1016/j.catena.2020.105058>

472                   Wu, X., Brüggemann, N., Butterbach-Bahl, K., Fu, B. and Liu, G., 2014. Snow cover  
473                   and soil moisture controls of freeze–thaw-related soil gas fluxes from a typical  
474                   semi-arid grassland soil: a laboratory experiment. *Biology and Fertility of Soils*,  
475                   50, pp.295-306. <https://doi.org/10.1007/s00374-013-0853-z>

476                   Xu, F., Song, W., Zhang, Y., Li, B., Hu, Y., Kim, Y. and Fu, Z., 2019. Water content  
477                   variations during unsaturated feet-scale soil freezing and thawing. *Cold Regions*  
478                   *Science and Technology*, 162, pp.96-103.  
479                   <https://doi.org/10.1016/j.coldregions.2018.11.011>

- 480 Xu, X., 2022. Effect of freeze-thaw disturbance on soil C and N dynamics and GHG  
481 fluxes of East Asia forests: Review and future perspectives. *Soil Science and*  
482 *Plant Nutrition*, 68(1), pp.15-26.  
483 <https://doi.org/10.1080/00380768.2021.2003164>
- 484 Yanai, Y., Toyota, K. and Okazaki, M., 2004. Effects of successive soil freeze-thaw  
485 cycles on soil microbial biomass and organic matter decomposition potential of  
486 soils. *Soil science and plant nutrition*, 50(6), pp.821-829.  
487 <https://doi.org/10.1080/00380768.2004.10408542>
- 488 Zhang, L., Ren, F., Li, H., Cheng, D. and Sun, B., 2021. The influence mechanism of  
489 freeze-thaw on soil erosion: a review. *Water*, 13(8), p.1010.  
490 <https://doi.org/10.3390/w13081010>
- 491 Zhang, S., Xiao, Z., Zhang, H. and Aurangzeib, M., 2022. Key factors determining soil  
492 organic carbon changes after freeze-thaw cycles in a watershed located in  
493 northeast China. *Science of the Total Environment*, 828, p.154525.  
494 <https://doi.org/10.1016/j.scitotenv.2022.154525>
- 495 Zong, R., Wang, Z., Li, W., Ayantobo, O.O., Li, H. and Song, L., 2023. Assessing the  
496 impact of seasonal freezing and thawing on the soil microbial quality in arid  
497 northwest China. *Science of The Total Environment*, 863, p.161029.  
498 <https://doi.org/10.1016/j.scitotenv.2022.161029>

499

#### 500 **Author contributions**

501 *Conceptualization, writing, reviewing, editing, data analysis, and visualization:* M.  
502 Sahoo, D. Baú, and S. Thornton; *Resources, methodology, software:* P. Bentley, A. Smith, P.  
503 Blackburn, and K. Howarth; *Supervision:* D. Baú and S. Thornton.

#### 504 **Funding**

505 This study was funded by Horizon Europe Guarantee award – UKRI (Project  
506 reference - EP/Y015843/1; Year - 2023).

507 **Competing interests**

508 The authors declare that they have no known competing interests that could influence  
509 the work reported in this paper.

510 **Ethical Approval**

511 Not applicable

512 **Consent to Participate**

513 Not applicable

514 **Consent to Publish**

515 Not applicable

516 **Data Availability**

517 Data will be made available upon request.