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Rivu, S., Smith, E., Stafford, G. orcid.org/0000-0002-5600-2465 et al. (1 more author) (2024) Draft genome sequence of Listeria aquatica strain SG_BD1, isolated from a cow dung sample in Bangladesh. Microbiology Resource Announcements, 13 (11). e00729-24. ISSN 2576-098X

https://doi.org/10.1128/mra.00729-24

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Draft genome sequence of *Listeria aquatica* strain SG_BD1, isolated from a cow dung sample in Bangladesh

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ABSTRACT Here, we present the genome of *Listeria aquatica* strain SG_BD1, isolated from cow dung in Dhaka, Bangladesh, and assembled after Oxford Nanopore sequencing. The genome is 2,690,148 bp with 2,855 predicted coding DNA sequences, G + C content of 39.6%, and displays a putative virulence gene *clpP* and 9 CRISPRs.

KEYWORDS Listeria aquatica, genome analysis

L isteria aquatica is a non-pathogenic species of the genus Listeria, which is usually found in aquatic sources. With the ability to ferment D-tagatose and the inability to ferment maltose, this species is unique in the Listeria genus (1). Moreover, multiple virulence and resistance genes with close homology to the pathogenic Listeria monocytogenes have been detected in this species (2). In Bangladesh, the presence of L. monocytogenes is evident in food and environmental samples, though there is no information regarding other species (3, 4). Here, we share the draft genome sequence of L. aquatica strain SG_BD1, which was isolated from cow dung in Dhaka, Bangladesh in 2023.

The cow dung sample was collected from a cattle farm in Dhaka (23°43'55.2972" N, 90°28'32.0916" E) and transferred immediately to the Department of Microbiology, University of Dhaka. The sample was enriched in Listeria Enrichment Broth (Oxoid, England), followed by streaking on Listeria Selective Agar (Oxoid, England) and incubation at 37°C for 48 h. A black colony from Listeria selective agar was confirmed as a potential *Listeria* spp. microscopically. Using the boiled DNA method, genomic DNA was extracted from an overnight culture of the isolate grown in nutrient broth and incubated at 37°C (5) and was subjected to Oxford Nanopore MinION genome sequencing using the Oxford Nanopore Sequencing Technologies, UK-Native Barcoding Kit 24 V14 and R10.4.1 SpotON flow cells. No shearing or size selection of DNA was performed before the sequencing.

Sequencing generated 2.81 M reads with N_{50} of 3.59 kb. Reads below the minimum quality score value of 8 were classified as failed reads. Sequencing was analyzed using the MinKNOW GUI 5.9.17 followed by assembly using Raven Galaxy version 1.8.0 and annotation utilizing NCBI Prokaryotic Genome Annotation Pipeline version 6.7 (PGAP) (6–8). CRISPRCasFinder web server version 4.2.20 (9) was used to detect CRISPR arrays, while virulence and antibiotic resistance genes were detected using the Virulence Factor Database and Comprehensive Antibiotic Resistance Database in ABRicate 1.0.1 on Galaxy, respectively (10–12). All these tools were applied using default parameters.

Assembly yielded a linear genome, 2,690,148 bp in length, consisting of three contigs, a G + C content of 39.6%, and an N_{50} value of 2,599,825 bp (Table 1). The sequence coverage of the assembled genome was 99×. Whole genome phylogenetic analysis using the Type Strain Genome Server (TYGS) (13) grouped SG_BD1 with the *L. aquatica* strain isolated from water in Florida, USA (Fig. 1) (1). Analysis using the BLAST Ring Image Generator (BRIG) tool version 0.95 revealed higher similarity of the study genome with

Editor John J. Dennehy, Queens College Department of Biology, Queens, New York, USA

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The authors declare no conflict of interest.

See the funding table on p. 3.

Received 10 July 2024 Accepted 29 August 2024 Published 26 September 2024

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FIG 1 (a) Phylogenetic analysis by TYGS shows that the study genome had the highest similarity with *L. aquatica* isolates. The whole genome phylogenetic tree was constructed based on the Genome Blast Distance Phylogeny (GBDP) approach. It inferred with FastME version 2.1.6.1 (15) from GBDP distances calculated from genome sequences. The branch lengths are scaled in terms of GBDP distance formula d5. The numbers above branches are GBDP pseudo-bootstrap support values >60% from 100 replications, with an average branch support of 64.3%. The tree was rooted at the midpoint (16). Here, leaf labels are annotated by affiliation to species clusters, subspecies clusters, percent G + C, delta statistics, genome size (in bp), and protein count (from left to right); (b) BRIG tool analysis of the study genome *L. aquatica* strain SG_BD1 (yellow ring) with *L. aquatica* strain FSL S10-1188, *L. aquatica* FSL L7-1507 (green and blue rings, respectively), and the type strain *L. monocytogenes* strain ATCC 19115 (the inner red circle). All sequences were downloaded from the NCBI database.

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Isolate ID	L. aquatica strain SG_BD1	
Sample source	Cow dung	
Year of isolation	2023	
Genome length (bp)	2,690,148	
No. of contigs	3	
GC content (%)	39.6	
N ₅₀ (bp)	2,599,825	
Coverage (×)	99	
Coding DNA sequence	2,855	
tRNA	67	
rRNAs (5S, 16S, 23S)	6, 6, 6	
Noncoding RNA	4	
Antibiotic resistance gene	None	
Virulence gene	1 (<i>clpP</i>)	
CRISPR array	9	
Accession No.	JBDILX00000000	

TABLE 1 Assembly statistics and important genomic features of the *L. aquatica* strain SG_BD1 from cow dung sample in Bangladesh.

two available *L. aquatica* genomes in the NCBI (accession numbers AOCG00000000 and JAARRM000000000), compared to the *L. monocytogenes* strain ATCC 19115 (accession number JARWJJ000000000) (Fig. 1b) (14).

PGAP revealed the presence of 2,855 coding DNA sequences, 67 tRNAs, 18 rRNAs, and 4 noncoding RNAs (Table 1). A putative virulence gene *clpP* was demonstrated, which aids in protection from heat stress and intracellular survival in *Listeria* spp. No antibiotic resistance gene was found in the genome, while nine CRISPR arrays were detected (Table 1).

ACKNOWLEDGMENTS

This work was funded by the National Institute for Health and Care Research (NIHR) Biomedical Research Centre (BRC) grant to E.S. and G.S. The authors acknowledge funding from the Charles Wallace Trust to S.A.

The authors wish to thank Mr. Nayeemur Rahman for extending his support to this research work.

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FUNDING

Funder	Grant(s)	Author(s)
National Institute for Health and Care Research (NIHR)		Graham Stafford
		Elspeth Smith

AUTHOR CONTRIBUTIONS

Supantha Rivu, Formal analysis, Investigation, Methodology, Writing – original draft, Writing – review and editing | Elspeth Smith, Funding acquisition, Methodology, Software | Graham Stafford, Funding acquisition, Methodology, Software, Supervision, Writing – review and editing | Sangita Ahmed, Conceptualization, Formal analysis, Investigation, Methodology, Supervision, Writing – review and editing

DATA AVAILABILITY

The whole-genome sequencing effort for *L. aquatica* strain SG_BD1 has been submitted to GenBank with accession number JBDILX00000000 under the BioProject number PRJNA1104809 (BioSample accession number SAMN41088195 and SRA accession number SRR29761655). Additional details can be found here. Accession number of the genomes downloaded from the NCBI: *L. aquatica* FSL S10-1188: AOCG00000000, *L. aquatica* strain FSL L7-1507: JAARRM000000000, and *L. monocytogenes* strain ATCC 19115: JARWJJ000000000.

REFERENCES

- den Bakker HC, Warchocki S, Wright EM, Allred AF, Ahlstrom C, Manuel CS, Stasiewicz MJ, Burrell A, Roof S, Strawn LK, Fortes E, Nightingale KK, Kephart D, Wiedmann M. 2014. *Listeria floridensis* sp. nov., *Listeria aquatica* sp. nov., *Listeria cornellensis* sp. nov., *Listeria riparia* sp. nov. and *Listeria grandensis* sp. nov., from agricultural and natural environments. Int J Syst Evol Microbiol 64:1882–1889. https://doi.org/10.1099/ijs.0. 052720-0
- Mao P, Wang Y, Li L, Ji S, Li P, Liu L, Chen J, Sun H, Luo X, Ye C. 2023. The isolation, genetic analysis and biofilm characteristics of *Listeria* spp. from the marine environment in China. Microorganisms 11:2166. https://doi. org/10.3390/microorganisms11092166
- Shourav AH, Hasan M, Ahmed S. 2020. Antibiotic susceptibility pattern of *Listeria* spp. isolated from cattle farm environment in Bangladesh. J Agric Food Res 2:100082. https://doi.org/10.1016/j.jafr.2020.100082
- Hasib Shourav A, Padmanon Salma K, Ahmed S, Khan MdAR. 2022. First report on *Listeria monocytogenes* detection in ready-to-eat chicken products in Bangladesh: a glimpse into their antibiotic resistance and virulence genes. SSRN J. https://doi.org/10.2139/ssrn.4153431
- Ribeiro Junior JC, Tamanini R, Soares BF, Oliveira AM de, Silva FDG, Silva FF da, Augusto NA, Beloti V. 2016 Efficiency of boiling and four other methods for genomic DNA extraction of deteriorating spore-forming bacteria from milk. Semin Cienc Agrar 37:3069. https://doi.org/10.5433/ 1679-0359.2016v37n5p3069
- Abueg LAL, Afgan E, Allart O, Awan AH, Bacon WA, Baker D, Bassetti M, Batut B, Bernt M, Blankenberg D, et al. 2024. The galaxy platform for accessible, reproducible, and collaborative data analyses: 2024 update. Nucleic Acids Res 52:W83–W94. https://doi.org/10.1093/nar/gkae410
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44:6614– 6624. https://doi.org/10.1093/nar/gkw569

- Vaser R, Šikić M. 2021. Raven: a *de novo* genome assembler for long reads. Bioinformatics. https://doi.org/10.1101/2020.08.07.242461
- Couvin D, Bernheim A, Toffano-Nioche C, Touchon M, Michalik J, Néron B, Rocha EPC, Vergnaud G, Gautheret D, Pourcel C. 2018. CRISPRCas-Finder, an update of CRISRFinder, includes a portable version, enhanced performance and integrates search for Cas proteins. Nucleic Acids Res 46:W246–W251. https://doi.org/10.1093/nar/gky425
- McArthur AG, Waglechner N, Nizam F, Yan A, Azad MA, Baylay AJ, Bhullar K, Canova MJ, De Pascale G, Ejim L, et al. 2013. The comprehensive antibiotic resistance database. Antimicrob Agents Chemother 57:3348– 3357. https://doi.org/10.1128/AAC.00419-13
- Chen L, Zheng D, Liu B, Yang J, Jin Q. 2016. VFDB 2016: hierarchical and refined dataset for big data analysis--10 years on. Nucleic Acids Res 44:D694–7. https://doi.org/10.1093/nar/gkv1239
- Seemann T. 2014. ABRicate: mass screening of contigs for antiobiotic resistance genes. Perl. Available from: https://github.com/tseemann/ abricate
- Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated highthroughput platform for state-of-the-art genome-based taxonomy. Nat Commun 10:2182. https://doi.org/10.1038/s41467-019-10210-3
- Alikhan N-F, Petty NK, Ben Zakour NL, Beatson SA. 2011. BLAST Ring Image Generator (BRIG): simple prokaryote genome comparisons. BMC Genomics 12:402. https://doi.org/10.1186/1471-2164-12-402
- Lefort V, Desper R, Gascuel O. 2015. FastME 2.0: a comprehensive, accurate, and fast distance-based phylogeny inference program. Mol Biol Evol 32:2798–2800. https://doi.org/10.1093/molbev/msv150
- 16. Farris JS. 1972. Estimating phylogenetic trees from distance matrices. Am Nat 106:645–668. https://doi.org/10.1086/282802