

This is a repository copy of Dynamic changes of key metabolites in Longjing green tea during processing revealed by widely targeted metabolomic profiling and sensory experiments.

White Rose Research Online URL for this paper: <u>https://eprints.whiterose.ac.uk/212645/</u>

Version: Accepted Version

Article:

Zeng, L., Fu, Y.-Q., Gao, Y. et al. (6 more authors) (2024) Dynamic changes of key metabolites in Longjing green tea during processing revealed by widely targeted metabolomic profiling and sensory experiments. Food Chemistry, 450. 139373. ISSN 0308-8146

https://doi.org/10.1016/j.foodchem.2024.139373

© 2024 Elsevier Ltd. This is an author produced version of an article published in Food Chemistry. Uploaded in accordance with the publisher's self-archiving policy. This manuscript version is made available under the CC-BY-NC-ND 4.0 license http://creativecommons.org/licenses/by-nc-nd/4.0/.

Reuse

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND) licence. This licence only allows you to download this work and share it with others as long as you credit the authors, but you can't change the article in any way or use it commercially. More information and the full terms of the licence here: https://creativecommons.org/licenses/

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

1	Dynamic changes of key metabolites in Longjing green tea during processing
2	revealed by widely targeted metabolomic profiling and sensory experiments
3	
4	Lin Zeng ^{a, 1} , Yan-Qing Fu ^{a, 1} , Ying Gao ^a , Fang Wang ^a , Shuang Liang ^a , Jun-Feng
5	Yin ^a , Marie-Laure Fauconnier ^b , Lijing Ke ^c , Yong-Quan Xu ^{a *}
6	^a Tea Research Institute Chinese Academy of Agricultural Sciences, Key Laboratory
7	of Tea Biology and Resources Utilization, Ministry of Agriculture, 9 South Meiling
8	Road, Hangzhou 310008, China
9	^b Laboratory of Chemistry of Natural Molecules, Gembloux Agro-Bio Tech, University
10	of Liege, 5030 Gembloux, Belgium
11	^c School of Food Science and Nutrition, University of Leeds, Leeds LS2 9JT, UK

¹ The authors contribute equally to this paper.

13

14 Corresponding Authors

15	*Yong-Quan	Xu,	Tel:	+86-571-86650594.	Fax:	+86	571	86650056.	Email:
16	yqx33@126.c	com.							
17									
18									
19									
20									
21									
22									

23 Abstract:

24 In this study, widely targeted metabolomics and chemometrics were utilized to 25 comprehensively analyse the formation of taste compounds in Longjing green tea. A 26 total of 580 non-volatile metabolites were identified by using ultra-performance liquid 27 chromatography-electrospray ionization-tandem mass spectrometry, and alterations in 28 three metabolic pathways were investigated. Notably, the fixation process reduced 29 phosphatidic acid levels, resulting in the formation of lyso-phosphatidylcholine and 30 lyso-phosphatidylethanolamine, as well as the release of esterified polyunsaturated fatty acids. Baiye No.1 had high levels of L-glutamic acid and L-glutamine, while 31 32 Longjing 43 showed elevated levels of flavones. Correlation analysis and sensory 33 verification indicated that the specific concentration of L-leucine could decrease the 34 umami of the tea. These findings advance our understanding of Longjing green tea 35 quality improvement and cultivar development.

36 **Keywords:** Longjing tea; Cultivar; Processing; Taste; Widely targeted metabolomics

37 1. Introduction

38 Green tea, a non-fermented tea, is the most produced and consumed tea in China. 39 It is renowned for its health-enhancing properties, including antioxidant, anti-40 inflammatory, anticancer effects and other functional activities (Schneider et al., 2009). 41 Manufactured from the fresh new shoots of tea plant (Camellia sinensis), the typical 42 green tea process involves picking, spreading, fixing, rolling, and drying. Sensory 43 evaluations of green tea reveal a variety of aroma attributes, including faint, floral, 44 chestnut-like, and other categories. Green tea infusion exhibits a bright green colour 45 and the taste is usually bitter, astringent, sweet, and umami. Importantly, taste is a key 46 determinant of consumer preference and acceptance, making it a critical aspect of the 47 sensory characteristics of green tea. The quality of tea depends on the secondary 48 metabolites it contains. For instance, the astringency of tea is primarily due to the 49 presence of polyphenols, and alkaloids contribute significantly to its bitter taste (Ye et 50 al., 2022). Amino acids constitute approximately 70% of the umami taste intensity of 51 green tea, while soluble sugars are the primary source of sweetness (Yue et al., 2017). 52 Taste compounds in tea leaves could be influenced by the quality of fresh tea 53 leaves (cultivars, growing conditions, and picking tenderness), processing technique, 54 and storage condition. Firstly, storage time is one of the quality evaluation criteria for 55 tea, especially for dark tea and white tea (Zhou et al., 2023). However, green tea is usually not recommended for long-term storage because its taste and aroma deteriorate 56 57 quickly. Secondly, tea plant cultivar is closely associated with tea quality (Wang et al., 58 2021). For instance, Longjing 43 is considered to be the most suitable cultivar for

59 producing Longjing tea (Dragon well tea). The fresh and mellow taste of Anji bai tea processed from Baiye No.1 is profited from its high L-theanine content and low tea 60 61 polyphenol levels (Zeng, Lin, Liu, & Liu, 2019). Finally, tea processing technologies 62 affected the quality of final tea product through regulating metabolism. Many bioactive 63 compounds vary significantly during the tea manufacturing process (Liao, Zhou, & 64 Zeng, 2022). In the fixation or roasting process, amino acids could react with carbonyl 65 compounds to form Strecker aldehydes that contributed to the formation of the tea aroma (Rizzi, 2008). The fixation stage was primarily associated with chlorophyll 66 67 decomposition, phosphatidic acids reduction and glycolipids degradation (Li et al., 68 2021). However, previous studies have mostly focused on the changes within a single cultivar during the processing or the differences between different cultivars, with few 69 70 investigating the differences in metabolite changes during the processing of different 71 cultivars. The formation of tea flavour and quality is very complicated, and different 72 cultivars may exhibit varying trends during processing. Therefore, comprehensive 73 studies are necessary to explore the dynamic changes of green tea used fresh tea leaves 74 from different cultivars as raw materials during the entire manufacturing process.

In recent years, there has been significant progress in the development and refinement of precision instruments, enabling the application of accurate and powerful techniques for food analysis. Metabolomics studies in the field of tea research have utilized various approaches, including targeted, untargeted, and widely targeted metabolomics. These methodologies have been employed in investigating different aspects of tea, such as evaluating the grade of *Tieguanyin* tea (Zeng et al., 2023),

81	assessing the impact of processing on green tea (Shi et al., 2022), and discriminating
82	tea cultivars in oolong tea (Zeng et al., 2022). Targeted metabolomics is mainly used
83	for the accurate quantification of known metabolites, which has the characteristics of
84	strong specificity and high sensitivity. This approach has been extensively utilized for
85	several decades in the field of metabolism research. In contrast, untargeted
86	metabolomics aims to detect and identify as many metabolites as possible in the
87	samples, including known and unknown metabolites (Zeng et al., 2022). However, the
88	identification of metabolites in untargeted metabolomics is complex and time-
89	consuming (Hertzog et al., 2022). A recent advancement in metabolomics analysis is
90	the widely targeted approach, which combines the advantages of untargeted and
91	targeted methods. Widely targeted metabolomics integrates the generality of untargeted
92	metabolomics with the accuracy of targeted metabolomics, offering a valuable tool for
93	comprehensive detection of metabolites (Zhou et al., 2022). Therefore, in this study, we
94	employed the widely targeted metabolomics approach as the analysis tool.

95 The objective of this research is to investigate the influence of cultivars and 96 processing on the metabolite profile of Longjing green tea, utilizing ultra-performance 97 liquid chromatography-electrospray ionization-tandem mass spectrometry (UPLC-ESI-MS/MS) and chemometrics. The hypothesis was that different cultivars had varying 98 99 metabolite profiles and exhibited different changes during the processing stage, which 100 might be the primary cause of the differences in the quality of Longjing green teas. Hence, three mainly-planted tea cultivars were selected, including albino tea cultivar 101 "Baiye No.1," current Longjing cultivars "Longjing 43" and the traditional cultivar 102

"Quntizhong." Fresh tea leaves from these cultivars were processed to Longjing green tea by using the same manufacturing technology. This study will investigate the differences in metabolite changes during the processing stage of different cultivars, and validate the results through sensory evaluation. The findings of this study are helpful in the understanding of quality improvements for Longjing green tea and high-quality tea cultivars development.

109 2. Materials and methods

110 *2.1. Chemicals*

The main chemicals used in this study included pure water (HangZhou Wahaha Group Co., Ltd., Hangzhou, China); methanol (MeOH, HPLC grade), and acetonitrile (HPLC, \geq 99.9%) (Merck, Darmstadt, Germany); formic acid (FA), (–)-Epigallocatechin (EGC), dihydromyricetin, and L-leucine (Aladdin, Shanghai, China). 2.2. Tea samples

116 In 2020, young shoots consisting of one bud and two leaves from Camellia sinensis cultivars 'Longjing 43 (LJ)', 'Quntizhong (QTZ)', and 'Baiye No.1 (BY)' were 117 118 collected from Pan'an County in Zhejiang Province, China. These leaves were utilized 119 in Longjing tea processing experiments. The tea processing procedure involved 120 sequential steps, starting with the picking of fresh leaves and followed by natural 121 withering at temperatures ranging from 18 to 24 °C for a duration of 8 hours. 122 Subsequently, the first drying (fixation) was carried out using an automatic flat tea stirfrying machine (6CCB-100ZD type, Zhejiang Hengfeng Technology Development Co., 123 124 Ltd., Shaoxing, China) at temperatures between 180 and 195 °C for a period of 1 hour, 125 with a rotational speed of 395 xg and a leaf load of 80-100g. After the first drying, the shaping process was conducted for 30 min in the automatic, flat tea stir-frying machine 126 for shaping at 180 °C for 60 min. Finally, the second drying was conducted for 30 min 127 in a tea roasting machine (6CH-3.0 type, Zhejiang Hengfeng Technology Development 128 Co., LTD, Shaoxing, China) with a temperature of 90 °C. Samples were collected and 129 130 preserved for analysis during the processing stages, including fresh tea leaves (FTL), 131 withering leaves (Wi), fixation/the first drying leaves (Fix), shaping leaves (Sha), and the second drying leaves/final tea products (Dry). The samples were carefully obtained 132 and stored to ensure their integrity before being subjected to further analysis. 133

134 2.3. Sensory evaluation

Each tea infusion was prepared by brewing 3.0 g of dried tea leaves with 150 mL 135 of boiling water for a duration of 4 min at room temperature (RT, $25 \pm 2^{\circ}$ C), according 136 137 to the standard method for Longjing green tea brewing outlined in Chinese standard 138 GB/T 23776-2018. The evaluation and scoring of the tea infusion were conducted by a trained panel of experts from the Tea Research Institute, Chinese Academy of 139 140 Agricultural Sciences. The panellists, aged between 25 and 48 years, possessed 141 certifications for tea quality evaluation issued by the Tea Scientific Society of China. 142 Prior to the evaluation, all panellists signed a consent form and willingly underwent 143 comprehensive training to develop their ability to discern various taste attributes, 144 including bitterness, astringency, umami, and total score. Each member of the panel 145 assigned scores to the different taste attributes, using a 0-10 scale to indicate the 146 intensity. The scoring scale ranged from 0-2 (very weak/just perceptible) to 8-10 (very

strong intensity). The mean values of the scores were calculated and reported (Zeng etal., 2023).

149 2.4. Sample preparation and extraction

150 2.4.1. Dry sample extraction

Biological samples were subjected to vacuum freeze-drying using a Scientz-100F 151 152 lyophilizer (Ningbo Xinzhi Biotechnology Co., Ltd., Ningbo, China). Subsequently, the 153 dried samples were ground to a powder form using a MM-400 Retsch grinder (Verder 154 Shanghai Instruments and Equipment Co., Ltd., Shanghai, China) operating at a frequency of 30 Hz for 1.5 min. For further analysis, 50 mg of the powdered sample 155 156 was weighed using an MS105DM electronic balance (Mettler Toledo Technology Co., Ltd., Shanghai, China). Subsequently, 1200 µL of a 70% methanolic aqueous internal 157 standard extract, pre-cooled to -20 °C, was added to the sample. The sample and extract 158 159 were vortexed once every 30 min for a duration of 30 seconds, repeating this process 160 six times. Following centrifugation at a rotation speed of 1152 xg for 3 minutes, the 161 supernatant was aspirated, and the sample was filtered using a microporous membrane 162 (ANPEL, Shanghai, China) with a pore size of 0.22 µm. The filtered sample was then stored in an injection vial (Beijing Castemer Technology Development Co., Ltd., 163 164 Beijing, China) for subsequent UPLC-MS/MS analysis.

165 2.4.2. UPLC Conditions

166 The sample extracts were subjected to analysis using a UPLC-ESI-MS/MS system 167 (UPLC, ExionLCTM AD) coupled with tandem mass spectrometry. The analytical 168 conditions included an Agilent SB-C18 UPLC column (1.8 μ m particle size, 2.1 mm ×

169 100 mm); the mobile phase consisted of solvent A, which was composed of pure water with 0.1% formic acid, and solvent B, which was composed of acetonitrile with 0.1% 170 171 formic acid. Sample measurements were carried out using a gradient program. Initially, the composition was 95% A and 5% B. Within 9 minutes, a linear gradient to 5% A and 172 173 95% B was applied, and this composition was maintained for 1 minute. Subsequently, 174 within 1.1 minutes, the composition was adjusted to 95% A and 5.0% B, and this 175 composition was maintained for 2.9 minutes. The flow velocity was set at 0.35 mL per minute, and the column oven temperature was maintained at 40 °C. The injection 176 volume was 2 µL. The effluent from the UPLC system was directed to an ESI-triple 177 178 quadrupole-linear ion trap (QTRAP)-MS for analysis.

179 *2.4.3. ESI-Q TRAP-MS/MS*

180 The electrospray ionization source operation parameters were set as follows: the source temperature was maintained at 500 ° C and the ion spray voltage was set to 5500 181 182 V for positive ion mode and -4500 V for negative ion mode. The ion source gases I and II and the curtain gas were set at 50, 60, and 25 psi, respectively. The collision-activated 183 184 dissociation was set to high. Pre-selection of qualitative ion pairs was conducted using 185 the characteristic ion fragmentation information and quasi-molecular ion compositions 186 of each metabolite for quantitative multiple reaction monitoring (MRM) analysis. MRM scans were acquired using a triple quadrupole mass spectrometer, with the 187 188 collision gas (nitrogen) set to medium. The declustering potential and collision energy 189 for each MRM transition were optimized through further declustering potential and 190 collision energy optimization. A specific set of MRM transitions was monitored for

191 each period based on the eluted metabolites within that period.

192 2.4.4. Principles of metabolite qualitative and quantitative analysis

193 In our study, we employed the self-built Metware Database (MetWare, Wuhan, 194 China) for compounds qualification, utilizing secondary spectral information. During 195 the analysis, we implemented a filtering process to eliminate duplicate signals originating from isotopes, as well as ions such as K⁺, Na⁺, NH₄⁺, and fragment ions that 196 197 are inherent components of larger molecular weight metabolites. For metabolite 198 quantification, we utilized the MRM mode of a triple quadrupole mass spectrometer. In 199 this mode, the quadrupole initially screened the precursor ions (parent ions) specific to 200 the target metabolites, thereby excluding ions corresponding to other molecular weight 201 compounds, and effectively minimizing interference. Subsequently, the precursor ions 202 were induced to undergo ionization within the collision chamber, resulting in 203 fragmentation into multiple ion fragments. These fragment ions were further filtered 204 through the triple quadrupole to select a characteristic fragment ion, thereby eliminating non-target ion interferences. This approach significantly enhanced the accuracy and 205 reproducibility of quantification. 206

Upon acquisition of mass spectrometry data for metabolomic analysis from diverse samples, peak area integration was performed for all chromatographic peaks corresponding to the compounds of interest. Subsequently, integration correction was applied to the mass spectrometry peaks of the same metabolite across different samples.

- 211 Metabolites exhibiting a matching score of 0.7 or higher for retention time and spectral
- 212 fragmentation ions in the database were selected and retained for further analysis.
- 213 2.5. Sensory verification experiment
- 214 We prepared 450 mL of tea according to the method described in section 2.3 of the 215 study. We divided the tea into 10 equal parts, with each part containing 40 mL, while 216 reserving 50 mL as the control sample (CK). Design different concentration gradients for the sensory verification experiment based on the desired concentrations of L-217 218 phenylalanine, EGC, dihydromyricetin, and L-leucine in the tea. The additive amount 219 for each compound should be as follows: 220 L-phenylalanine: 8 mg and 24 mg 221 EGC: 8 mg and 24 mg Dihydromyricetin: 200 mg and 400 mg 222 L-leucine: 12 mg and 36 mg 223 224 Add the appropriate amount of each compound, according to the designed 225 concentration gradients, to the respective 40 mL portions of tea. Ensure thorough 226 mixing to achieve homogeneity. Ask panellists to evaluate the corresponding taste 227 attributes of the tea samples. Each member will receive a sample with a specific 228 concentration of the compound (s) for evaluation. The taste attributes to be assessed 229 include bitterness, astringency, umami, and the overall flavour score. The CK should serve as a reference for comparison. Collect the evaluation scores and feedback from 230

231 panellists for further analysis and interpretation.

232 2.6. Data processing and statistical analysis

233 Partial least squares discriminant analysis (PLS-DA) modelling was conducted using SIMCA 13.0 software (Umetric, Umea, Sweden). Heat map analysis was 234 235 performed using TBtools v2.003 software (Chen et al., 2020). Hierarchical cluster analysis and radar map visualization were conducted using Origin 2023b software 236 237 (Origin Lab Corp., Northampton, MA). Pearson correlation analysis between taste 238 attributes and chemical compounds was performed using SPSS software version 20.0 239 (SPSS, Chicago, IL, USA), and the resulting network diagram was generated using 240 Cytoscape software version 3.9.1 (https://cytoscape. Org). K-means clustering analysis 241 was performed using R software version 3.5.1 (https://www.r-project.org/).

3. Results

243 3.1. Determination of non-volatile metabolites in three varying cultivars

A total of 2615 metabolites were obtained after peak-picking and alignment in the analysis. Subsequently, the metabolites were matched with the database based on their secondary mass spectra (containing all fragment ions of the substances) and retention time, resulting in 580 non-volatile metabolites with a matching degree of 70% or above. These metabolites comprised various classes, including 36 alkaloids, 64 amino acids and derivatives, 138 flavonoids, 29 lignans and coumarins, 78 lipids, 27 nucleotides and derivatives, 33 organic acids, 70 phenolic acids, 22 tannins, 19 terpenoids, and 64 251 other metabolites (Figure 1A).

To better understand the changes in non-volatile metabolites during Longjing tea 252 253 processing, the hierarchical cluster analysis was performed on these three different cultivars (Figure 1B). The analysis revealed significant differences among three 254 255 cultivars, indicating distinct metabolic profiles. When comparing variations in 256 processing within the same cultivar, it was observed that samples taken before and after 257 fixation process showed noticeable discrepancies. Furthermore, through a K-means plot 258 analysis of the processing stages, four different change tendencies were identified for 259 these metabolites (Figure 1C). Among them, a total of 271 non-volatile metabolites 260 exhibited lower content in the Dry compared to the FTL, while 309 metabolites 261 displayed higher content. This observation strongly indicated that the manufacturing 262 process influenced the transformation and generation of metabolites.

263

3 3.2. Alteration of metabolites in fixation process

264 The hierarchical cluster analysis results demonstrated significant disparities in 265 non-volatile metabolites subsequent to fixation. To find the essential difference 266 metabolites, a PLS-DA model was established, which is shown in Figure 2A and 2B. 267 This model effectively distinguished tea samples before and after fixation, and further screened 30 key metabolites with variable importance in projection (VIP) values 268 269 exceeding 2, as visualized in Figure 2C, 12 of which decreased post-fixation, while the 270 rest increased. These key metabolites contained 10 lipid compounds, indicating that the lipids changed greatly during fixation process. Further observation of lipid metabolite 271

alterations following fixation revealed that most glycerol esters displayed a downward
tendency, while lyso-phosphatidylcholine (LPC) and lyso-phosphatidylethanolamine
(LPE) increased, as illustrated in Figure 2D.

275 *3.3. Differences in sensory and metabolites among three cultivars Longjing teas*

276 Different tea plant cultivars contain varying biochemical compositions, such as 277 large-leaf cultivars with a high level of polyphenols, and albino cultivars with a high content of amino acids (Zhao et al., 2022). In this experiment, Longjing teas made from 278 279 three cultivars were analyzed and well discriminated, as shown in Figure 3A, and 3B. 280 Based on the criterion of VIP value greater than 2, key differential metabolites were identified and visualized in a heat map (Figure 3C), with a total of 35 metabolites 281 282 identified. The BY cultivar exhibited higher levels of (-)-Epigallocatechin (EGC), 3-283 methylellagic acid, and dihydromyricetin. In the LJ cultivar, isoorientin-7-O-glucoside, 284 orientin-2"-O-galactoside, 3-isopropylmalic acid, 2-propylmalic acid, (-)-Epicatechin 285 gallate (ECG), cryptochlorogenic acid (4-O-caffeoylquinic acid), morin-3-arabinoside, and quercetin-3-O-arabinoside were found to be more abundant than the other two 286 cultivars. The QTZ cultivar exhibited higher levels of naringenin-7-O-glucoside 287 (flavanones), kaempferol-3-O(2"-galloyl) galactoside, and epicatechin-3-(3"-O-288 289 methyl) gallate.

To quantify specific sensory factors attributes (i.e., bitterness, astringency, and umami) among three cultivars, an organoleptic test was conducted. As shown in Figure 3D, the BY cultivar achieved the highest total and umami scores, the LJ cultivar scored 293 highest in bitterness, and the QTZ cultivar exhibited a pronounced astringency. 294 Previous studies have reported that the superior performance of the BY cultivar was 295 attributed to its high levels of amino acids and low levels of catechins and caffeine, 296 which reduced astringency and bitterness while enhanced the umami taste (Feng et al., 297 2014). This observation was further supported by the dynamic changes in 12 amino 298 acids (Figure 3E), where the content of L-glutamic acid and L-glutamine were highest 299 in the BY cultivar. These compounds are known to be the primary contributors to the umami taste in green tea. Therefore, the L-glutamic acid and L-glutamine might be the 300 301 reason for the intense umami taste of the BY cultivar. Additionally, previous studies 302 have identified caffeoyl- or feruloyl-substituted quinides as contributors to the bitter taste in roasted coffee (Frank et al., 2006). Hence, it was possible that cryptochlorogenic 303 304 acid (4-O-caffeoylquinic acid) contributed to the strong bitterness of the LJ cultivar. Similarly, naringenin-7-O-glucoside (a flavanone), kaempferol-3-O(2"-galloyl) 305 306 galactoside, and epicatechin-3-(3"-O-methyl) gallate (polyphenols), which are known 307 to present bitter taste, were enriched in the QTZ cultivar. However, these results were 308 only tentative and based on previous studies. To further demonstrate these findings, 309 correlation analyses between sensory results and key metabolites, as well as sensory 310 verification experiment, were conducted.

311 3.4. Correlation analysis and sensory verification

The data analysis workflow, illustrated in Figure 4A, involved the utilization of a PLS-DA model to identify 35 metabolites exhibiting differential expression (VIP > 2). Subsequently, these 35 metabolites and 12 amino acids were assessed for correlation

315 with the sensory evaluation results. The significance was set as the absolute value of the correlation coefficients surpassing 0.8. The findings revealed significant 316 317 correlations between these metabolites and the sensory attributes of umami and 318 astringency, whereas no correlations were observed with bitterness. Astringency, as the 319 characteristic taste of green tea, is primarily influenced by hydrolysable and condensed 320 tannins (Granato et al., 2014). Among the 7 metabolites showing strong correlations 321 with astringency, compounds such as N-(sulfonyl) phenylalanine, jaceosidin-7-Ogalactoside, epicatechin-3-(3"-O-methyl) gallate, and 4-hydroxy-3-methoxyphenyl 1-322 323 $O-\beta$ -D-(6'-O-galloyol)-glucopyranoside displayed the significantly positive 324 correlations with the astringency score. Conversely, dihydromyricetin (ampelopsin), epigallocatechin, and L-phenylalanine exhibited notably negative correlations with the 325 326 astringency score.

327 To further validate their actual taste contributions, L-Phenylalanine, EGC, and 328 dihydromyricetin were selected to be added into tea infusion due to the difficulty in 329 obtaining other compounds (Figure 4B). The results showed that dihydromyricetin enhanced the astringency, while L-phenylalanine reduced it. EGC displayed varying 330 331 effects depending on its concentration. Despite exhibiting a negative correlation in the 332 correlation analysis, dihydromyricetin was found to enhance the astringency score. On 333 the other hand, the bitterness of L-phenylalanine, which was proven to be a major 334 contributor to bitterness in bamboo shoots (Gao et al., 2019). However, the interaction 335 between L-phenylalanine and astringency remains unknown and required further 336 investigation to better understand their relationship.

337 Focused on the umami taste, although 8 metabolites exhibited strong correlations with umami taste. The majority of metabolites did not show a significant correlation 338 339 with umami, except for L-leucine and N-(sulfonyl) phenylalanine, which presented a 340 negative correlation. To validate this finding, we added L-leucine into tea infusion, and 341 the results indicated that the umami score decreased upon the addition of L-leucine 342 (Figure 4C), consistent with previous findings that L-leucine has a negative effect on 343 umami taste. As a branched-chain amino acid, L-leucine itself has a bitter taste, which could explain the observed reduction in umami flavour. In conclusion, it was found that 344 345 L-phenylalanine has a negative effect on astringency taste, while L-leucine can reduce 346 the umami taste.

347 Based on the results, L-phenylalanine, epigallocatechin, dihydromyricetin, and L-348 leucine were identified as essential taste metabolites in Longjing tea. Furthermore, heat 349 map was conducted to visualize the distinct variations in these compounds during 350 processing in three cultivars (Figure 4D). Specifically, QTZ exhibited the highest content of L-leucine, whereas BY had the highest level of L-phenylalanine, 351 epigallocatechin, and dihydromyricetin. These findings can help to explain the sensory 352 353 evaluation results, with QTZ exhibiting the lowest umami flavour and BY having the 354 lowest astringency taste. Notably, L-phenylalanine and L-leucine displayed an 355 increasing trend across the three cultivars during processing. The increase in amino 356 acids was attributed to the protein hydrolysis promoted by high temperature treatment during processing. Similar observations have been reported in other studies on the 357 358 processing of green tea (Wang et al., 2021).

359 3.5. Dynamic changes in the main taste metabolites of three cultivars Longjing360 teas during processing

The primary taste constituents of tea are amino acids (umami), flavonoid (bitterness and astringency), and alkaloids (bitterness) (Zhang et al., 2020). In order to comprehensively understand the transformations of the non-volatile compounds during Longjing tea processing, we focused on three salient metabolic pathways: the flavonoid pathway, the amino acid pathway, and the alkaloid pathway. The modifications in these metabolic pathways were discussed in detail blow.

367 3.5.1 Modifications in the flavonoid metabolic pathway

368 The major components of the flavonoid metabolic pathway in tea include flavones 369 and flavone glycosides, flavonol glycosides, and flavanols, as depicted in Figure 5. Flavones and flavonols are predominantly present as O-glycosides, with a glycoside 370 371 moiety attached to the C-3 position of the aglycones. These compounds played the crucial role in contributing to the bitter taste of tea (Fang et al., 2019). During 372 373 processing, the concentrations of flavones (such as vitexin, apigenin, and isovitexin) and most apigenin glycosides generally exhibited an upward trend, particularly after 374 375 the fixation process. However, exceptions were observed for apigenin-4'-O-glucoside, 376 apigenin-7-O-glucoside (cosmosiin), and isovitexin-7-O-glucoside (saponarin). 377 Although the abundance of flavones and flavone glycosides differed among the three tea cultivars, the overall change tendency during processing was similar. 378

379 The concentration of quercetin significantly increased after the fixation process.

Quercetin was known to contribute to the green colour of tea infusion (Wang et al., 2004), which could explain the enhancement of tea infusion colour observed after fixation. The detected flavonol glycosides were categorized as kaempferol glycosides, quercetin glycosides, and myricetin glycosides. The abundance of quercetin glycosides displayed a notable decrease trend during processing, particularly after fixation, whereas there were no significant changes observed in the trends for kaempferol glycosides and myricetin glycosides.

387 Flavanols, the most characteristic and abundant metabolites in tea, were 388 considered the primary contributors to the astringency and bitterness taste of tea (Zhang 389 et al., 2020). In this study, different types of flavanols exhibited diverse changes. (-)-390 Epigallocatechin gallate (EGCG) and (-)-catechin gallate (CG) decreased during the 391 whole processing. Previous research has indicated that catechins undergo various transformations, including isomerization, optical isomerization, hydrolysis, thermal 392 393 polymerization, and pyrolysis, during the processing of green tea (Wang et al., 2021), 394 These transformations may be the reason for the observed decrease in EGCG and CG.

395 *3.5.2 Modifications in the amino acid metabolic pathway*

396 Dynamic alterations in amino acids during processing are illustrated in Figure 6A. 397 Throughout the tea manufacturing process, the abundance of amino acids varied 398 significantly. Initially, fresh tea leaves contained high concentrations of L-theanine and 399 L-glutamic acid, which gradually diminished during processing. Following withering, 400 the concentrations of L-valine, L-aspartic acid, L-tyrosine, and L-tryptophan 401 significantly increased. These amino acids changes likely represented differentially402 expression responsible for the distinctive taste profile of Longjing green tea.

The dynamic changes observed in L-valine, L-aspartic acid, L-theanine, Lglutamic acid, L-tyrosine, and L-tryptophan were similar among the three tea cultivars. However, the dynamic change trend of L-arginine during processing was different. During processing L-arginine kept increasing in QTZ cultivar, but showed a trend of first decrease and then increase in BY and LJ cultivars. The variability in L-arginine levels could be attributed to varietal specificity.

409 *3.5.3 Modifications in the alkaloid metabolic pathway.*

410 Caffeine and theobromine are the primary alkaloids found in tea, and contributed 411 to the characteristic bitter taste of tea infusions. As depicted in Figure 6B, tea processing 412 had significant impact on the levels of caffeine and theobromine. Specifically, the 413 content of theobromine consistently decreased throughout the processing stages. 414 However, the content of caffeine showed an overall increasing trend and varied among 415 the three cultivars during the processing. This observed phenomenon could be attributed to the conversion of theobromine, an intermediate compound in the 416 417 biosynthesis of caffeine, into caffeine during the processing steps (Xia et al., 2017). 418 Furthermore, the fluctuation in caffeine content throughout the processing stages could 419 potentially be attributed to sublimation of caffeine caused by exposure to high 420 temperature. It is worth noting that the content of both theobromine and caffeine was found to be lower in BY than that in LJ and QTZ. These findings highlighted the 421 422 dynamic changes in caffeine and theobromine levels during tea processing and

423 demonstrated the influence of processing on the composition and taste characteristics424 of tea.

425 **4. Discussion**

426 4.1. Sensory and metabolomic profiling differences among three different tea427 cultivars

428 According to previous research, tea quality can be greatly influenced by various factors, such as cultivar, ecological environment, cultivation method, processing 429 430 technology, and storage condition. Among these factors, cultivar plays a key role (Qin 431 et al., 2023). The metabolite levels vary significantly among different cultivars, resulting in different sensory qualities of the final tea product. In this experiment, BY 432 433 cultivar, also known as Anji Baicha, was chosen. It is a low-temperature-sensitive, periodic albino variant that produces albino young shoots in early spring when the 434 environmental temperature is below 20-22 °C. Previous reports have indicated that 435 436 albino tea contains high levels of amino acids and low levels of chlorophyll, catechins, 437 and caffeine, which contribute to a favorable taste profile by reducing bitterness and astringency, and increasing umami. Similarly, in our study BY cultivar presented high 438 439 umami characteristics and notably higher levels of L-glutamic acid and L-glutamine than the other two cultivars (Figure S1). L-glutamic acid could reduce the bitterness of 440 441 ester catechins, and it was one of the main contributors to the umami taste in green tea 442 (Liu et al., 2023). Additionally, the sensory experiment revealed that L-phenylalanine could reduce the astringency of tea leaves, and BY cultivar contained higher levels of 443

444 L-phenylalanine, which may contribute to its lowest astringency taste.

LJ and QTZ are typical cultivars used in the production of Longjing tea. QTZ is a 445 446 sexual cultivar reproduced from seeds under specific natural conditions. LJ is a superior asexual cultivar bred from QTZ. In this experiment, LJ exhibited a pronounced 447 448 bitterness, while QTZ tea had a more noticeable astringency and lowest umami flavour. 449 Further correlation analysis and sensory validation experiment revealed that L-leucine might be the main cause of the umami of QTZ tea, while the bitterness of LJ tea may 450 451 be attributed to cryptochlorogenic acid (4-O-caffeoylquinic acid). Currently, the 452 research on the sensory and metabolomic differences between sexual and asexual tea 453 cultivars is rarely reported, highlighting the need for further study in this area

454 *4.2. Metabolite changes during the processing of Longjing tea*

455 The metabolites' changes in tea processing mainly come from the different 456 chemical reactions that occur in the processing, including hydrolysis, substitution, isomerization, oxidation-reduction, and other thermo-physical and chemical reactions. 457 458 The changes in metabolites have an important impact on the quality of tea. Different kinds of tea have different processing technologies, and green tea processing includes 459 460 withering, fixation and drying. Withering is a crucial step in the formation of tea quality, 461 primarily aimed at reducing the moisture content of fresh tea leaves. This process 462 involves chemical transformations, increased activity of leaf enzymes, and the oxidation of certain polyphenols, degradation of chlorophyll, hydrolysis of 463 polysaccharides and disaccharides into monosaccharides, and breakdown of proteins 464

465	into amino acids. These transformations play a key role in establishing the substance
466	foundation for the formation of tea quality. In this experiment, significant increases
467	were observed in the levels of L-valine, L-aspartic acid, L-tyrosine, phenylalanine, and
468	L-tryptophan after the withering stage (Figure S1). This accumulation of amino acids
469	during the withering stage might be due to the degradation of proteins by endogenous
470	peptidases (Chen et al., 2020). Fixation is the most critical step in Longjing tea
471	processing. The processing aims to inhibit the oxidation of polyphenols by inactivating
472	the oxidase, and promote the volatilization of low-boiling-point aroma compounds,
473	resulting in a unique aroma (Yu et al., 2023). In our study, it was found that the content
474	of L-theanine decreased after fixation. Previous studies had reported that the decrease
475	in L-theanine content was due to the Maillard reaction between theanine and glucose,
476	leading to the formation of Amadori rearrangement products, such as methylpyrazine
477	and 2,5-dimethylpyrazine, which are commonly found in various teas (Han et al., 2022).
478	In addition to amino acids, the levels of lipid metabolites also underwent
479	significant changes after the fixation stage. Most glycerol esters showed a decrease
480	trend, while LPC and LPE increased (Figure S1). Previous studies have demonstrated
481	that LPC and LPE could induce lipid degradation under heat treatment to form carbonyl
482	metabolites, such as hexanal, 2,4-dienal, and 1-octen-3-one (Lin & Blank, 2003).
483	Therefore, it can be inferred that thermally-induced processes potentially decreased the
484	content of phosphatidic acids, leading to the formation of LPC and LPE and the release
485	of esterified forms of polyunsaturated fatty acids. Besides, in this study, flavonoids,
486	important bitter compounds in tea, also showed some characteristic changes during

487 processing. The levels of apigenin, quercetin, and vitexin increased after fixation, while 488 myricetin decreased. Previous studies have suggested that the increase in these 489 flavonoids was related to the hydrolysis of flavonoid glycosides (Wang et al., 2021). 490 However, in this experiment, there was no corresponding decrease in the metabolites 491 of *O*-glycosides, which requires further investigation.

492 **5.** Conclusion

Based on the combination analysis of widely targeted metabolomics and 493 494 chemometrics, this study investigated the effects of cultivars and processing on 495 Longjing green tea's metabolite profile. A total of 580 non-volatile metabolites were identified, highlighting the alterations in flavonoids, amino acids, and alkaloids 496 497 metabolic pathways. Fixation process potentially reduced phosphatidic acid levels, 498 leading to the formation of LPC, LPE, and the release of esterified polyunsaturated fatty 499 acids. Cultivar difference brought distinct metabolites and taste profiles, with more L-500 glutamic acid and L-glutamine in BY cultivar, and accumulated flavones in LJ. 501 Correlation analysis between taste attributes and key metabolites revealed that certain concentration of L-leucine could decrease tea umami. Except for a few metabolites, the 502 503 change trends of key metabolites during processing were similar among cultivars. 504 These findings advanced our understanding of Longjing green tea quality improvement 505 and were helpful in the development of high-quality tea cultivars. However, this study still had some limitations. For example, the fixation process might cause lipid 506 507 degradation without specific measurement of the related degradation products,

508 Additionally, the correlation analysis between sensory evaluation and metabolites may

- 509 be limited due to the small sample size. Further research will focus on investigating the
- 510 degradation of lipids and their products during the fixation process.

511 Ethical statement

- 512 All the participants (healthy and nonsmokers from TRICAAS) were conducted in
- 513 accordance with the principle set forth in the Declaration of Helsinki and informed
- 514 written consent was obtained. This study was approved by the Zhejiang Gongshang
- 515 University Human Ethics Committee.

516 **CRediT authorship contribution statement**

517 Lin Zeng: Writing – original draft, Methodology, Investigation, Data curation, Formal

518 analysis, Visualization. **Yan-Qing Fu**: Writing – review & editing, Resources, Data

- 519 curation, Methodology, Supervision. Ying Gao: Writing review & editing, Resources,
- 520 Software, Visualization. Fang Wang: Writing review & editing. Shuang Liang:
- 521 Investigation, Data curation. Jun-Feng Yin: Writing review & editing. Marie-Laure
- 522 Fauconnier: Writing review & editing. Lijing Ke: Investigation, Data curation.
- 523 Yong-Quan Xu: Conceptualization, Writing review & editing, Data curation, Formal
- 524 analysis, Investigation, Project administration.

525 **Conflicts of Interest**

526 The authors declare no conflicts of interest regarding the publication of this paper.

527 Acknowledgments

528 This research was supported by the National Natural Science Foundation of China (32272368, 31872709, 32272771), the Key Research and Development Program of 529 530 Zhejiang (2022C02033), the Science and Technology Cooperation Project of Zhejiang 531 Province (2024SNJF028), and the Innovation Project for Chinese Academy of 532 Agricultural Sciences. We would also like to thank the Instrumental Analysis Center in 533 Tea Research Institute of the Chinese Academy of Agricultural Sciences. 534 References 535 Chen, C. J., Chen, H., Zhang, Y., Thomas, H. R., Frank, M. H., & He, Y. H., et al. 536 (2020). Tbtools: An integrative Toolkit developed for interactive analyses of

537 big biological data. *Molecular plant*, *13*(8), 1194-1202.

- 538 Chen, Y. Y., Zeng, L. T., Liao, Y. Y., Li, J. L., Zhou, B., & Yang, Z. Y., et al. (2020).
- Enzymatic reaction-related protein degradation and proteinaceous amino acid
 metabolism during the black tea (*Camellia sinensis*) manufacturing process. *Foods*, 9(1), 66.
- Fang, Z. T., Song, C. J., Xu, H. R., & Ye, J. H. (2019). Dynamic changes in flavonol
 glycosides during production of green, yellow, white, oolong and black teas
 from *Camellia sinensis L.* (cv. Fudingdabaicha). *International Journal of Food Science and Technology*, *54* (2), 490–498.
- 546 Feng, L., Gao, M. J., Hou, Y. R., Hu, X. Y., Zhang, L., & Wan, X. C., et al. (2014).
- 547 Determination of quality constituents in the young leaves of albino tea cultivars.

548

Food Chemistry, 155, 98-104.

- 549 Frank, O., Zehentbauer, G., & Hofmann, T. (2006). Bioresponse-guided decomposition
- of roasted coffee beverage and identification of key bitter taste compounds. *European Food Research and Technology*, 222(5-6), 492–508.
- 552 Gao, Q., Jiang, H., Tang, F., Cao, H. Q., Wu, X. W., & Qi, F. F., et al. (2019).
- Evaluation of the bitter components of bamboo shoots using a metabolomics
 approach. *Food & Function*, 10(1), 90–98.
- 555 Granato, D., Grevink, R., Zielinski, A. A. F., Nunes, D. S., & Van Ruth, S. M. (2014).
- Analytical strategy coupled with response surface methodology to maximize the extraction of antioxidants from ternary mixtures of green, yellow, and red teas (*Camellia sinensis var. sinensis*). Journal of Agricultural and Food Chemistry,
- *62*(42), 10283–10296.
- 560 Han, Z. S., Jiang, Z. D., Zhang, H., Qin, C. Y., Rong, X. Q., & Lai, G. P., et al. (2022).
- 561 Amadori reaction products of theanine and glucose: formation, structure, and 562 analysis in tea. *Journal of Agricultural and Food Chemistry*, 70(37), 11727– 563 11737.
- 564 Hertzog, A., Selvanathan, A., Devanapalli, B., Ho, G., Bhattacharya, K., & Tolun, A.
- A. (2022). A narrative review of metabolomics in the era of "-omics":
 integration into clinical practice for inborn errors of metabolism. *Translational Pediatrics*, 11(10), 1704–1716.
- Li, J., Hua, J. J., Yuan, H. B., Deng, Y. L., Zhou, Q. H., & Yang, Y. Q., et al. (2021).
 Investigation on green tea lipids and their metabolic variations during

570

manufacturing by nontargeted lipidomics. Food Chemistry, 339, 128114

- Liao, Y. Y., Zhou, X. C., & Zeng, L. T. (2022). How does tea (*Camelliia sinensis*)
 produce specialized metabolites which determine its unique quality and
 function: a review. *Critical Reviews in Food Science and Nutrition*, 62(14),
 3751–3767.
- Liu, Z. Y., Ran, Q. S., Li, Q., Dai, Y. Q., Zhang, T., & Fang, S. M., et al. (2023).
 Interaction between major catechins and umami amino acids in green tea based
 on electronic tongue technology. *Journal of Food Science*, 88(6), 2339–2352.
- Lin, J. M., & Blank, I. (2003). Odorants generated by thermally induced degradation of
 phospholipids. *Journal of Agricultural and Food Chemistry*, 51(15), 4364–4369
- 580 Qin, X. X., Zhou, J. T., He, C., Qiu, L., Zhang, D., & Yu, Z., et al. (2023). Non-targeted
- metabolomics characterization of flavor formation of Lichuan black tea
 processed from different cultivars in Enshi. *Food Chemistry: X, 19,* 100809.
- 583 Rizzi, G. P. (2008). The Strecker degradation of amino acids: newer avenues for flavour

formation. *Food Reviews International*, 24(4), 416–435.

- 585 Schneide, C., & Segre, T. (2009). Green tea: potential health benefits. *American Family*586 *Physician*, 79(7), 591–594.
- Shi, Y. L., Zhu, Y., Ma, W. J., Shi, J., Peng, Q. H., & Lin, Z., et al. (2022).
 Comprehensive investigation on non-volatile and volatile metabolites in four
 types of green teas obtained from the same tea cultivar of LongJing 43
 (*Camellia sinensis var. sinensis*) using the widely targeted metabolomics. *Food*
- 591 *Chemistry*, *394*, 133501.

592	Wang, H. J., Hua, J. J., Yu, Q. Y., Li, J., Wang, J. J., & Deng, Y. L., et al. (2021).
593	Widely targeted metabolomic analysis reveals dynamic changes in non-volatile
594	and volatile metabolites during green tea processing. Food Chemistry, 363,
595	130131.
596	Wang, L. F., Park, S. C., Chung, J. O., Baik, J. H., & Park, S. K. (2004). The compounds
597	contributing to the greenness of green tea. Journal of Food Science, 69(8),
598	S301–S305.

Xia, E. H., Zhang, H. B. Sheng, J., Li, K., Zhang, Q. J., & Kim, C., et al (2017). The
tea tree genome provides insights into tea flavor and independent evolution of

601 caffeine biosynthesis. *Molecular Plant*, *10*(6), 866–877.

- 602 Ye, J. H., Ye, Y., Yin, J. F., Jin, J., Liang, Y. R., & Liu, R. Y., et al. (2022). Bitterness
- and astringency of tea leaves and products: Formation mechanism and reducing
 strategies. *Trends in Food Science & Technology*, *123*, 130–143.
- 605 Yu, Y. Y., Zhu, X. Z., Ouyang, W., Chen, M., Jiang, Y. W., & Wang, J. J., et al. (2023).
- Effects of electromagnetic roller-hot-air-steam triple-coupled fixation on
 reducing the bitterness and astringency and improving the flavor quality of
 green tea. *Food Chemistry-X*, *19*, 100844.
- Yue, C. N., Wang, Z. H., Mao, S. H., Liao, X. Y., & Tong, R. H. (2017). The main taste
 substances in tea research progress. *Food Research and Development, 38*(1),
 219–224.
- 612 Zeng, C. Z., Lin, H. Y., Liu, Z. X., & Liu, Z. H. (2019). Analysis of young shoots of
 613 "Anji Baicha" (*Camellia sinensis*) at three developmental stages using

- nontargeted LC-MS-based metabolomics. *Journal of Food Science*, 84(7),
 1746–1757.
- Zeng, L., Jin, S., Fu, Y. Q., Chen, L. S., Yin, J. F., & Xu, Y. Q. (2022). A targeted and 616 617 untargeted metabolomics analysis of "oriental beauty" oolong tea during processing. Beverage Plant Research, 2(20). doi: 10.48130/BPR-2022-0020 618 619 Zeng, L., Fu, Y. Q., Liu, Y. Y., Huang, J. S., Chen, J. X., & Yin, J. F., et al. (2023). 620 Comparative analysis of different grades of *Tieguanyin* oolong tea based on metabolomics and sensory evaluation. LWT-Food Science and Technology, 174, 621 622 114423. 623 Zeng, L., Fu, Y. Q., Huang, J. S., Wang, J. R., Jin, S., & Yin, J. F., et al. (2022). Comparative analysis of volatile compounds in *Tieguanyin* with different types 624 625 based on HS-SPME-GC-MS. Foods, 11(11), 1530. Zhang, L., Cao, Q. Q., Granato, D., Xu, Y. Q., & Ho, C. T. (2020). Association between 626 627 chemistry and taste of tea: A review. Trends in Food Science & Technology, 628 101, 139–149. Zhao, Z. J., Ma, Q. P., Lou, Y. G., Zhang, J., Hu, X. C., & He, J. J., et al. (2022). 629
 - 631 1" and the traditional albino tea "Huangjinya". *South African Journal of Botany*,
 632 149, 572–581.

Comparative analysis of the chloroplast genomes of a novel albino tea "Huabai

630

Zhou, S., Zhang, J. M., Ma, S. C., Ou, C. S., Feng, X. Y., & Pan, Y. N., et al. (2023).
Recent advances on white tea: manufacturing, compositions, aging
characteristics and bioactivities. *Trends in Food Science & Technology, 134*,

- 636 41–55.
- 637 Zhou, J, N., Hou, D. H., Zou, W. Q., Wang, J. H., Luo, R., & Wang, M., et al. (2022).
- 638 Comparison of widely targeted metabolomics and untargeted metabolomics of
- 639 wild *Ophiocordyceps sinensis*. *Molecules*, 27(11), 3645.