

This is a repository copy of Longitudinal lung function assessment of patients hospitalised with COVID-19 using 1H and 129Xe lung MRI.

White Rose Research Online URL for this paper: <u>https://eprints.whiterose.ac.uk/197911/</u>

Version: Published Version

Article:

Saunders, L.C. orcid.org/0000-0002-1080-9861, Collier, G.J., Chan, H.-F. orcid.org/0000-0002-5382-2097 et al. (41 more authors) (2023) Longitudinal lung function assessment of patients hospitalised with COVID-19 using 1H and 129Xe lung MRI. Chest, 164 (3). pp. 700-716. ISSN 0012-3692

https://doi.org/10.1016/j.chest.2023.03.024

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Check for updates

Longitudinal Lung Function Assessment of Patients Hospitalized With COVID-19 Using ¹H and ¹²⁹Xe Lung MRI

Laura C. Saunders, PhD; Guilhem J. Collier, PhD; Ho-Fung Chan, PhD; Paul J. C. Hughes, PhD; Laurie J. Smith, PhD; J. G. R. Watson, MBBS; James E. Meiring, DPhil; Zoë Gabriel, MBBS; Thomas Newman, MB BChir; Megan Plowright, BMBS; Phillip Wade, MBChB; James A. Eaden, MBChB, PhD; Siby Thomas, BSc; Scarlett Strickland, MBChB; Lotta Gustafsson, MBChB; Jody Bray BSc; Helen Marshall, PhD; David A. Capener, MSc; Leanne Armstrong; Jennifer Rodgers, BA; Martin Brook, MSc; Alberto M. Biancardi, PhD; Madhwesha R. Rao, PhD; Graham Norquay, PhD; Oliver Rodgers, BEng; Ryan Munro, MEngHons; James E. Ball, MSci; Neil J. Stewart, PhD; Allan Lawrie, PhD; R. Gisli Jenkins, MD, PhD; James T. Grist, PhD; Fergus Gleeson, MD, PhD; Rolf F. Schulte, PhD; Kevin M. Johnson, PhD; Frederick J. Wilson, MSc; Anthony Cahn, MRCP PhD; Andrew J. Swift, MD, PhD; Smitha Rajaram, MD; Gary H. Mills, PhD; Lisa Watson, PhD; Paul J. Collini, PhD; Rod Lawson, PhD; A. A. Roger Thompson, PhD; and Jim M. Wild, PhD

BACKGROUND: Microvascular abnormalities and impaired gas transfer have been observed in patients with COVID-19. The progression of pulmonary changes in these patients remains unclear.

RESEARCH QUESTION: Do patients hospitalized with COVID-19 without evidence of architectural distortion on structural imaging exhibit longitudinal improvements in lung function measured by using ¹H and ¹²⁹Xe MRI between 6 and 52 weeks following hospitalization?

STUDY DESIGN AND METHODS: Patients who were hospitalized with COVID-19 pneumonia underwent a pulmonary ¹H and ¹²⁹Xe MRI protocol at 6, 12, 25, and 51 weeks following hospital admission in a prospective cohort study between November 2020 and February 2022. The imaging protocol was as follows: ¹H ultra-short echo time, contrast-enhanced lung perfusion, ¹²⁹Xe ventilation, ¹²⁹Xe diffusion-weighted, and ¹²⁹Xe spectroscopic imaging of gas exchange.

RESULTS: Nine patients were recruited (age 57 ± 14 [median \pm interquartile range] years; six of nine patients were male). Patients underwent MRI at 6 (n = 9), 12 (n = 9), 25 (n = 6), and 51 (n = 8) weeks following hospital admission. Patients with signs of interstitial lung damage were excluded. At 6 weeks, patients exhibited impaired ¹²⁹Xe gas transfer (RBC to membrane fraction), but lung microstructure was not increased (apparent diffusion coefficient and mean acinar airway dimensions). Minor ventilation abnormalities present in four patients were largely resolved in the 6- to 25-week period. At 12 weeks, all patients with lung perfusion data (n = 6) showed an increase in both pulmonary blood volume and flow compared with 6 weeks, although this was not statistically significant. At 12 weeks, significant improvements in ¹²⁹Xe gas transfer were observed compared with 6-week examinations; however, ¹²⁹Xe gas transfer remained abnormally low at weeks 12, 25, and 51.

INTERPRETATION: ¹²⁹Xe gas transfer was impaired up to 1 year following hospitalization in patients who were hospitalized with COVID-19 pneumonia, without evidence of architectural distortion on structural imaging, whereas lung ventilation was normal at 52 weeks.

CHEST 2023; 164(3):700-716

KEY WORDS: ¹²⁹Xe; COVID-19; gas transfer; hyperpolarized gas; imaging; MRI; xenon MRI

Take-home Points

Study Question: Do patients hospitalized due to COVID-19 with no evidence of architectural distortion exhibit longitudinal improvements in ¹²⁹Xe gas transfer to within a normal range between 6 and 52 weeks following hospitalization?

Results: At 12 weeks, significant improvements in ¹²⁹Xe gas transfer were observed compared with 6-week examinations. However, ¹²⁹Xe gas transfer remained abnormally low at weeks 12, 25, and 51. **Interpretation:** In a cohort of patients with moderate severity disease, ¹²⁹Xe gas transfer improved but did not return to within a normal range within 1 year following hospitalization.

In patients hospitalized with pneumonia caused by infection with SARS-CoV-2, the existing literature and clinical experience suggest that there is considerable overlap in clinical presentation with typical pneumonia and ARDS in patients exhibiting hyperinflammation and progressive hypoxemia. However, patients with severe COVID-19 also display evidence of an inflammatory and thrombotic vasculopathy with endothelial dysfunction and excessive blood flow to collapsed lung tissue.¹⁻³ Abnormal pulmonary vasoregulation has been observed in patients in the acute phase of COVID-19¹ and may be a pathophysiologic mechanism contributing to the progressive hypoxemia seen in these patients.

Abnormalities on chest radiograph or CT scan imaging at 12 weeks following hospitalization due to COVID-19 are present in some patients, particularly those with more severe disease who require ICU treatment.⁴ However, for patients without radiographic abnormalities, sensitive techniques for monitoring longitudinal change in lung function are needed.

Lung MRI with hyperpolarized ¹²⁹Xe gas allows direct, regionally sensitive measurements of lung ventilation and function, and it is an emerging method that is used both clinically and in clinical research, alongside ¹H MRI, to evaluate lung function and abnormalities.⁵⁻¹¹ In addition, ¹²⁹Xe can image gas diffusion within the lung airspace (diffusion-weighted MRI [DW-MRI]), and the derived apparent diffusion coefficient (ADC) and mean diffusive length scale (Lm_D) provide threedimensional in vivo information of the underlying lung microstructure that is highly sensitive to changes in patients with emphysema¹² and fibrotic lung disease.¹³ In addition, ¹²⁹Xe is soluble in the interstitium/ membrane (M) and in the RBCs, and the signal from ¹²⁹Xe in these dissolved compartments can be distinguished spectroscopically. The ratio of the ¹²⁹Xe MRI signal observed in the lung airspaces (gas), the lung M, and bound to the RBCs can thus be determined with magnetic resonance spectroscopic imaging. In particular, the fractions of the ¹²⁹Xe signal in the RBC to M fraction (RBC:M), RBC to gas fraction (RBC:gas), and M to gas fraction (M:gas) ratios have been used to probe gas transfer^{14,15} and are highly sensitive to gas transfer limitation and longitudinal change in interstitial, emphysematous, and pulmonary vascular diseases.¹⁶⁻¹⁸ RBC:M has been shown to correlate highly with the transfer factor for carbon monoxide (TL_{CO}).^{6,19}

ABBREVIATIONS: ADC = apparent diffusion coefficient; DCE = dynamic contrast enhanced; DW-MRI = diffusion-weighted MRI; IQR = interquartile range; Lm_D = mean diffusive length scale; M = membrane; M:gas = membrane to gas fraction; PFT = pulmonary function test; RBC:gas = RBC to gas fraction; RBC:M = RBC to membrane fraction; SPGR = spoiled gradient echo; T_2^* = transverse relaxation time; TL_{CO} = transfer factor for carbon monoxide; UTE = ultra-short echo time; VDP = ventilation defect percentage; VP = ventilation percentage

of Radiology (J. T. G. and F. G.), Oxford University Hospitals, Oxford, England; GE Healthcare (R. F. S.), Munich, Germany; Department of Medical Physics (K. M. J.), University of Madison, Madison, WI; and GSK (F. J. W. and A. C.), Stevenage, England. Current affiliation for H.-F. C.: Auckland Bioengineering Institute, University of Auckland, Auckland, New Zealand. Current affiliation for A. L.: Faculty of Medicine, National Heart & Lung Institute, Imperial College London, London, England. Current affiliation for F. J. W.: Blue Earth Diagnostics Ltd, Oxford, England.

DOI: https://doi.org/10.1016/j.chest.2023.03.024

AFFILIATIONS: From the Department of Infection, Immunity and Cardiovascular Disease (L. C. S., G. J. C., H.-F. C., P. J. C. H., L. J. S., J. A. E., S. T., J. B., H. M., D. A. C., L. A., J. R., M. B., A. M. B., M. R. R., G. N., O. R., R. M., J. E. B., N. J. S., A. L., A. J. S., G. H. M., P. J. C., A. A. R. T., and J. M. W.), University of Sheffield, Sheffield, England; Sheffield Teaching Hospitals (J. G. R. W., J. E. M., Z. G., T. N., M. P., P. W., S. S., L. G., S. R., L. W., and R. L.), NHS Foundation Trust, Sheffield, England; National Heart and Lung Institute (A. L. and G. J.), Imperial College London, London, England; Department of Oncology (J. G. and F. G.), Oxford NHS Foundation Trust, Oxford, England; Oxford Centre for Clinical Magnetic Resonance Research (J. T. G.), University of Oxford, Oxford, England; Department of Physiology, Anatomy, and Genetics (J. T. G), University of Oxford, Oxford, England; Department

Part of this article has been presented in abstract form at the International Society for Magnetic Resonance in Medicine, May 12, 2021-May 20, 2021, virtual meeting, the International Society for Magnetic Resonance in Medicine, July 5, 2022-July 12, 2022, London, England, and the European Respiratory Society, August 4, 2022-August 6, 2022, Barcelona, Spain).

CORRESPONDENCE TO: Jim M. Wild; email: j.m.wild@sheffield.ac.uk Copyright © 2023 The Author(s). Published by Elsevier Inc under license from the American College of Chest Physicians. This is an open access article under the CC BY license (http://creativecommons.org/ licenses/by/4.0/).

Previous studies have reported reduced RBC:M in patients with COVID-19,²⁰⁻²³ including in patients with normal chest CT scan imaging but ongoing dyspnea.²¹ In patients with residual lung abnormalities on CT scans, decreased RBC:M may be due to an increase of xenon uptake in the interstitial lung tissue. However, in the absence of CT scan abnormalities, we propose that a decreased RBC:M instead indicates microvascular (capillary) abnormalities. Therefore, RBC:M in particular may be a sensitive metric suitable for longitudinal assessment of regional gas exchange abnormalities in patients who have had COVID-19 and have normal structural imaging. It is currently unknown whether RBC:M improves longitudinally following COVID-19 pneumonia. Age-related reductions in the RBC:M ratio may be relevant in other cohorts,²⁴ and control cohorts well-matched for age are therefore needed for accurate interpretation of RBC:M in post-COVID-19 studies. It is also unclear whether patients with abnormal RBC:M have concurrent abnormalities in lung perfusion or ventilation.

¹H lung MRI is able to assess changes in lung structure and perfusion. Ultra-short echo time (UTE) imaging enables good visualization of the lung parenchyma and has shown excellent agreement with CT scan imaging in the visualization of lesions in

Study Design and Methods *Participants*

Patients with acute COVID-19 pneumonia and no previously diagnosed respiratory disease (excluding mild asthma) were recruited from Sheffield Teaching Hospital's pulmonology and infectious diseases wards from November 2020 to February 2022 for this prospective cohort study, prior to or shortly following discharge. Follow-up ¹²⁹Xe and ¹H lung MRI examinations were acquired at approximately 6, 12, 24, and 52 weeks following COVID-19 infection. Patients were required to meet the following criteria: (1) a positive SARS-CoV-2 result from a nasal/pharyngeal or respiratory sample; (2) hospitalization with a diagnosis of pneumonia (chest radiograph or CT scan consistent with COVID-19 infection); (3) development of impaired oxygenation (pulse oximetry saturation \leq 93% on room air) requiring additional oxygen; and (4) no evidence of interstitial lung damage on CT scan or MRI structural imaging at 12 weeks following hospital admission, as judged by a clinical chest radiologist.

Patients with evidence of interstitial lung damage at 12 weeks following hospital admission were recruited into the parallel UK Interstitial Lung Disease Consortium (UKILD) study.²⁹ Standard MRI exclusion criteria were applied to all subjects. In addition, patients were excluded if they were unable to tolerate a test inhalation of ¹²⁹Xe gas according to the supervising clinicians' judgment or if they had a chest size exceeding the ¹²⁹Xe chest coil circumference (76 cm).

patients with COVID-19.²⁵ Dynamic contrastenhanced (DCE) ¹H lung MRI allows the assessment of lung perfusion, with high sensitivity and specificity in detecting perfusion defects without exposing the participant to ionizing radiation,²⁶ and it is therefore well suited for patient follow-up studies. Increased lung perfusion transit times (time to peak) have been reported in both an acute hospitalized patient with COVID-19 and in nonhospitalized male patients with breathlessness who have had COVID-19.^{27,28}

The current study used a ¹H and ¹²⁹Xe MRI protocol that combines hyperpolarized ¹²⁹Xe imaging methods sensitive to ventilation, lung microstructure (DW-MRI), and gas exchange (dissolved xenon spectroscopic imaging) alongside ¹H DCE perfusion and UTE lung structural imaging to assess pathophysiologic changes in patients who had been hospitalized with COVID-19 pneumonia during the postacute period. The primary hypothesis of this work was that abnormal imaging and pulmonary function test (PFT) markers of lung function would increase to within a normal range over the course of 1 year in patients without structural abnormalities seen on CT scan or proton structural imaging. Patients underwent up to four follow-up MRI examinations at approximately 6, 12, 24, and 52 weeks following hospitalization.

Where possible, PFTs were acquired on the same day as the MRI examination at each visit. Spirometry and transfer factor were performed, and from these tests, the metrics FEV_1 , FVC, FEV_1/FVC , TL_{CO} , and carbon monoxide transfer coefficient were calculated and presented as *z* scores and % predicted using Global Lung Function Initiative reference ranges.^{30,31} This study was approved by the London-Hampstead Research Ethics Committee (REC reference: 9/LO/1115).

MRI Acquisition

Patients underwent scanning on either an HDx 1.5T (N = 7) or a 450w 1.5T (N = 2) (GE Healthcare) MRI scanner.³² The ¹²⁹Xe images were acquired with the patient in a flexible quadrature transmit/receive vest coil (Clinical MR Solutions). Patients' vital signs were monitored throughout the MRI examination. Each patient underwent MRI examinations on the same scanner for baseline visits and follow-up. Figure 1 presents an illustrative diagram of the lung MRI methods used in this study.

¹²⁹Xe doses were polarized to approximately 30% using a home-built high-performance spin-exchange optical pumping polarizer.³² This had regulatory approval for manufacture of hyperpolarized ¹²⁹Xe for clinical lung MRI by the UK Medicines and Healthcare Products Regulatory Agency.

MR imaging was conducted as summarized in the following text (parameters are detailed in e-Table 1).



Figure 1 – A-B, Illustrative diagram showing how the lung MRI techniques used in this article measure lung perfusion, ventilation, lung microstructure (diffusive length scale), and xenon gas transfer (the transfer of xenon between the airspace, membrane, and RBCs). A, Techniques in a healthy alveolus. B, Possible interpretation of the findings of this article in patients who have had COVID-19, with reduced RBC:M due to damage to pulmonary microcirculation but preserved acinar airway dimensions. RBC:M = RBC to membrane fraction.

A structural ¹H scan was acquired following inhalation of a bag of air to match the lung inflation state of the subsequent xenon sequences. ¹²⁹Xe ventilation images were acquired using a three-dimensional imaging sequence with whole-lung coverage following inhalation of a 1 L maximum mixture of ¹²⁹Xe and nitrogen (titrated if subject height < 160 cm³³) and inhaled from functional residual capacity; patients were coached in the required breathing maneuver prior to their MRI examination.³⁴

 $^{129}\mathrm{Xe}$ DW-MRI to assess alveolar microstructural change was acquired following inhalation of a maximum 1 L mixture of $^{129}\mathrm{Xe}$ and nitrogen (three-dimensional spoiled gradient echo [SPGR] multiple b-value sequence with compressed sensing with whole-lung coverage). 35

Three-dimensional spectroscopic imaging of the gas and dissolved phase xenon resonances (dissolved xenon in lung M and in blood

RBCs) was acquired by using a maximum 1 L dose of hyperpolarized ¹²⁹Xe (repetition time = 15 milliseconds, flip angle = 22° [three-dimensional acquisition with whole-lung coverage]).⁶

¹H MRI was acquired by using an eight-element cardiac array (GE Healthcare). UTE images were acquired with a three-dimensional radial sequence during 8 min of free-breathing with prospective respiratory bellows gating on expiration.³⁶

Three-dimensional variable flip angle SPGR images^{37,38} were acquired (flip angle = 2°, 4°, 10°, and 30°) to allow for the correction of lung T₁ and proton density. DCE lung perfusion MRI was acquired (three-dimensional volumetric time-resolved SPGR acquisition). A half dose (0.05 mL/kg) of Gadovist (Bayer) was administered at an injection rate of 4 mL/s followed by a 20 mL saline flush at 4 mL/s. Patients were advised to hold their breath for as long as possible and breathe shallowly thereafter.

Image Analysis

Qualitative assessments of the UTE ¹H structural, ¹²⁹Xe ventilation, and DCE lung perfusion images were made by two radiologists with 10 and 14 years of experience, respectively. UTE images were assessed for parenchymal changes, and ventilation and perfusion images were assessed for defects.

Metrics of ventilation defect percentage (VDP), low ventilation percentage (VP), normal VP, and hyper VP for each patient were calculated by using linear binning (see the Methods section of e-Appendix 1). The coefficient of variation of the segmented lung ventilation images was also calculated from the ¹²⁹Xe ventilation images as a marker of ventilation heterogeneity.

Maps of ^{129}Xe ADC and $\text{Lm}_{\rm D}$ from a stretched exponential model of ^{129}Xe gas diffusion in the lungs were calculated on a voxel-by-voxel basis. 39

Maps of gas transfer ratios (RBC:M, RBC:gas, and M:gas) were calculated from three-dimensional spectroscopic imaging. The transverse relaxation time (T_2^*) of the RBC and M spectroscopic peaks was also calculated.

Mean values of all global metrics were calculated for each patient. A sample size calculation was not performed because this was an exploratory study.

Global MRI metrics from visits 1, 2, 3, and 4 were compared by using a Skillings-Mack test due to the presence of missing data⁴⁰ with pairwise

Results

Of the 16 recruited patients, 14 showed no signs of interstitial lung damage at 12 weeks and were therefore

Wilcoxon tests and a correction for multiple testing,⁴¹ implemented by using R software.⁴² Data are presented as median (range), unless otherwise stated.

Mixed linear effect models were set up using a random intercept model to test the relationship between RBC:M and the following: (1) pulmonary blood volume; (2) pulmonary blood flow; (3) mean transit time; (4) VDP; and (5) $TL_{CO} z$ score. IBM SPSS Statistics 27 (IBM SPSS Statistics, IBM Corporation) was used for analysis. A *P* value < .05 was considered statistically significant.

Age- and Sex-Matched Healthy Volunteer Metrics

Median ADC and Lm_D values for an age- and sex-matched control cohort were determined by retrospective analysis of previously published data.⁴³ Eleven subjects from this previously published work were selected based on matching median and interquartile range (IQR) of age and sex ratio from a cohort of 23 subjects while blinded to MRI metrics; the control cohort had a median age of 63 (40-70) years, and 73% were male.

Median RBC:M, RBC:gas, and M:gas for an age- and sex-matched control cohort were determined by retrospective analysis of a healthy cohort data set, with control subjects chosen based on matching median and IQR of age and sex ratio while blinded to MRI metrics. Twelve subjects were selected (median age, 57 [41-68] years), and 67% were male.

included as part of this study. Nine of 14 patients had follow-up examinations and were included for analysis (Fig 2).



Figure 2 – Flow chart of patient recruitment. UKILD = UK Interstitial Lung Disease Long-COVID-19 study.

TABLE 1	Patient Demographic Data
---------	--------------------------

Characteristic	Group	No. of Patients
Demographic charac	teristics	
Age, y	< 50	2
	50-59	3
	60-69	3
	70-79	1
Sex	Male	6
	Female	3
BMI, kg/m ²	25-29.9	3
	30-39.9	5
	≥ 40	1
Comorbidities 4C score ⁴⁴	0	4
	1	4
	3	1
Tobacco use history	No tobacco use	4
	Former tobacco use	5
Clinical characteristi	cs on admission	
Admission SF ratio	< 200	0
	200-299	3
	300-399	3
	≥ 400	3
Clinical characteristi	cs during admission	
Maximum oxygen requirement during hospital stay	< 28%	1
	28%-35%	4
	40%	2
	> 60%	2
	CPAP	1
ISARIC 4C score ⁴⁴	1-4	3
	5-8	5
	> 8	1
Length of stay, d	1-5	4
	6-9	4
	> 10	1
Maximum National Early Warning Score 2 score ⁴⁵	5-6	4
	≥ 7	5
Medication during stay	Oral antibiotics	1
	IV antibiotics	2
		(Continued)

 TABLE 1] (Continued)

Characteristic	Group	No. of Patients
	Dexamethasone	9
	Remdesivir (Gilead Sciences)	5
	Immunomodulation therapy	3
	Convalescent plasma	2
	Colchicine	2
	Aspirin	3
	Included in an interventional study?	7
Lowest SF ratio during stay	< 200	2
	200-299	3
	300-399	4
	≥ 400	0
Maximum FIO2 during stay	< 28%	1
	28%-35%	4
	40%	2
	> 60%	2
	CPAP	1
Clinical characteristi	cs postdischarge	
MRC Dyspnoea Scale (1-5), 6 wk	1	6
	2	2
	Not available	1
MRC Dyspnoea Scale (1-5), 3 mo	1	7
	2	1
	3	1
	Not available	0
Readmittance	Yes	1 for general surgery unrelated to COVID- 19
	No	8

Individuals who formerly used tobacco use all reported \leq 15 pack y. The pulse oximetry saturation/Fio₂ ratio (SF ratio) was calculated by using estimated Fio₂ based on flow rate when delivered by nasal cannulae. ISARIC = International Severe Acute Respiratory and Emerging Infection Consortium; MRC = Medical Research Council; pulse oximetry saturation/Fio₂ (SF).

Six of nine patients were male. Median patient age, height, and weight were 57 (42-72) years, 173 (170-191) cm, and 101 (84-112) kg, respectively. Visit 1 (N = 9)



Figure 3 – Example of UTE images, RBC:M maps, ¹²⁹Xe ventilation images, and maps of pulmonary blood flow at visit 1 and visit 2, for each patient. The white arrow indicates a segmental perfusion defect visible at visit 1, which improves at visit 2. M = membrane; PBF = pulmonary blood flow; RBC:M = RBC to membrane fraction; UTE = ultra-short echo time.

occurred 6 (4-12) weeks following hospital admission; visit 2 (N = 9) occurred 12 (11-22) weeks following hospital admission; visit 3 (n = 7) occurred 25 (23-28) weeks following hospital admission; and visit 4 (n = 8) occurred 51 (49-62) weeks following hospital admission. Patients had been admitted to the hospital with COVID-19 for 6 (2-15) days. Further patient demographic data are presented in Table 1.^{44,45} No patients received any trial of pharmacologic treatment for post-COVID-19 symptoms following discharge. Two of the patients commenced treatment for diabetes during the follow-up period.

UTE and ¹²⁹Xe MRI were successfully acquired in all patients at all visits. DCE lung perfusion imaging was

successfully acquired in six of nine patients at visit 1, eight of nine patients at visit 2, six of seven patients at visit 3, and five of eight patients at visit 4. The reasons for unsuccessful lung perfusion imaging were patients failing screening (6 visits), patient motion (2 visits), and technical issues (1 visit).

Figure 3 shows representative slices from the UTE images, RBC:M maps, ¹²⁹Xe ventilation images, and DCE pulmonary blood flow maps for each patient at visit 1 and visit 2. Figure 4 displays plots of ventilation, dissolved phase ¹²⁹Xe, and DCE lung perfusion metrics for each patient at each visit. Median metrics and statistical comparisons of metrics at each visit are presented in Table 2.



Figure 4 – Spaghetti plots of ventilation, dissolved phase xenon, and dynamic contrast-enhanced lung perfusion metrics at visits 1 to 4. CV = coefficient of variation of lung ventilation; M = membrane; M:gas = membrane to gas fraction; RBC:gas = RBC to gas fraction; RBC:M = RBC to membrane fraction; VDP = ventilation defect percentage.

¹²⁹Xe MRI

Ventilation: At visit 1, small ventilation defects were visible in the lung periphery in four patients (1, 3, 4, and 6). No other patients had visible lung ventilation defects. At visits 2 and 3, the ventilation defects observed in patients 1, 3, 4, and 6 had improved, with small defects still visible, particularly in patient 3. At visit 4, small peripheral ventilation defects were observed in patients 1, 3, and 6 (e-Fig 1, Fig 3).

Whole lung VDP was calculated for each patient. At visit 1, median VDP was 1.6% (0.6%-3.9%); at visit 2, VDP was 1.3% (0.7%-2.6%); at visit 3, VDP was 1.2% (0.4%-2.1%); and at visit 4, VDP was 0.8 % (0.4%-3.7%).

Quantitative metrics of ventilation improved at visits 2, 3, and 4 compared with visit 1; however, this was not statistically significant following adjustment for multiple corrections (Table 2).

DW-MRI (Alveolar Microstructure): Median ADC and Lm_D at each visit are reported in Table 2. No significant longitudinal changes in ADC and Lm_D were seen between visits. Median ADC and Lm_D were within the median \pm IQR of age- and sex-matched control data (age- and sex-matched control data: median ADC, 0.0360 cm²/s [IQR, 0.005 cm²/s]; median Lm_D , 289 µm [IQR, 27 µm]) at all visits (e-Fig 2).

TABLE 2	Median Metrics	for All MRI	Parameters a	at Visits	1, 2,	3, and	4
---------	----------------	-------------	--------------	-----------	-------	--------	---

MRI Parameter	Visit 1	Visit 2	Visit 3	Visit 4	P Value	Adjusted P Value
No.	9	9	7	8		
ADC, cm ² /s	0.0344 (0.309-0.0373)	0.0327 (0.0281-0.0386)	0.0340 (0.310-0.364)	0.0338 (0.307-0.0357)		
Lm _D , μm	281 (260-300)	273 (251-301)	278 (263-290)	279 (263-288)		
RBC:M	0.22 (0.15-0.37)	0.25 (0.18-0.41)	0.25 (0.19-0.44)	0.23 (0.19-0.44)	V1-V2, <i>P</i> = .004 V1-V3, <i>P</i> = .047 V1-V4, <i>P</i> = .039	V1-V2, <i>P</i> = .023 V1-V3, <i>P</i> = .094 V1, V4, <i>P</i> = .094
RBC:gas	0.0026 (0.0018-0.0039)	0.0030 (0.0019-0.0038)	0.0026 (0.0020-0.0040)	0.0024 (0.0017-0.0038)		
M:gas	0.0113 (0.0091-0.0125)	0.0114 (0.0081-0.0179)	0.0101 (0.0091-0.0113)	0.0094 (0.0082-0.0104)	V1-V4, <i>P</i> = .031 V2-V4, <i>P</i> = .023 V3-V4, <i>P</i> = .031	V1-V4, <i>P</i> = .063 V2-V4, <i>P</i> = .063 V3-V4, <i>P</i> = .063
M T ₂ *, ms	2.58 (2.46-2.68)	2.47 (2.38-2.58)	2.42 (2.36-2.56)	2.22 (1.94-2.40)	V1-V2, P = .044 V1-V3, P = .023 V1-V4, P = .008 V2-V4, P = .008 V3-V4, P = .031	V1-V2, <i>P</i> = .053 V1-V3, <i>P</i> = .031 V1-V4, <i>P</i> = .023 V2-V4, <i>P</i> = .023 V3-V4, <i>P</i> = .047
RBC T_2^* , ms	2.20 (2.05-2.48)	2.16 (2.01-2.49)	2.16 (2.06-2.32)	2.27 (2.10-2.47)		
DCE PBV, mL/ 100 mL	37.8 (11.7-53.5)	47.6 (15.0-60.2)	45.3 (36.3-58.4)	53.8 (52.46-60.72)		
SD PBV, mL/ 100 mL	18.0 (7.5-28.5)	21.3 (13.0-24.8)	23.8 (17.9-25.7)	21.1 (20.8-22.0)		
IQR PBV, mL/ 100 mL	25.1 (7.8-34.9)	27.8 (10.5-36.1)	35.9 (24.2-41.5)	29.5 (27.0-31.9)		
DCE PBF, mL/ 100 mL/min	76.9 (19.6-107.2)	90.2 (30.7-109.5)	71.1 (60.3-117.7)	98.3 (93.4-116.2)		
SD PBF, mL/ 100 mL/min	45.8 (11.5-58.8)	54.5 (32.0-75.0)	48.6 (35.6-69.5)	54.2 (41.0-65.4)		
IQR PBF, mL/ 100 mL/min	54.0 (14.1-61.3)	59.2 (25.5-102.9)	59.0 (43.2-78.7)	64.8 (54.2-72.4)		
Median MTT, s	6.5 (5.6-8.0)	7.3 (6.4-8.0)	7.1 (3.5-9.9)	6.9 (2.3 – 7.6)		
SD MTT, s	1.3 (0.9-1.6)	1.2 (0.6-2.6)	1.3 (0.5-2.2)	0.7 (0.6-0.9)		
IQR MTT, s	1.3 (1.1-2.0)	1.6 (0.7-3.1)	1.3 (0.6-3.5)	0.7 (0.5-1.1)		
VDP, %	1.6 (0.6-3.9)	1.3 (0.7-2.6)	1.2 (0.4-2.1)	0.8 (0.4-3.7)	V1-V3, <i>P</i> = .016	V1-V3, <i>P</i> = .094
Normal VP, %	76.4 (62.5-77.7)	76.9 (72.3-86.2)	78.9 (67.4-81.5)	81.1 (69.0-82.6)	V1-V2, <i>P</i> = .027 V1-V3, <i>P</i> = .031	V1-V2, <i>P</i> = .093 V1-V3, <i>P</i> = .093
Low VP, %	12.5 (10.0-15.4)	11.9 (8.9-13.1)	10.9 (9.1-14.8)	10.6 (10.0-13.7)		

[164#3 CHEST SEPTEMBER 2023]

708 Original Research

(Continued)

s. P values are shown for	een at least two variable	e was a significant difference betw	is-Mack test determined that there	lable data for each visit. If a Skillinc	an (range) of all patients with avail	Data are presented as medi
V1-V2, P = .078 V1-V3, P = .078 V1-V4, P = .078	V1-V2, P = .040 V1-V3, P = .016 V1-V4, P = .040	26.1 (25.0 -33.3)	26.9 (25.6-34.1)	28.8 (22.3-32.2)	29.0 (27.5-37.1)	Lung ventilation CV, %
:	:	8.4 (6.8-15.2)	9.7 (7.9-15.8)	11.0 (4.2-13.3)	11.7 (9.5-18.3)	Hyper VP, %
Adjusted P Value	<i>P</i> Value	Visit 4	Visit 3	Visit 2	Visit 1	MRI Parameter

Wilcoxon pairwise tests. P values are shown prior to and following adjustment for multiple testing. ADC = apparent diffusion coefficient; CV = coefficient of variation; DCE = dynamic contrast enhanced; IQR = interquartile range; Lm_b = mean diffusive length scale; M = membrane; M:gas = membrane to gas fraction; MTT = mean transit time; PBF = pulmonary blood flow; PBV = pulmonary blood volume; RBC:gas = RBC to gas fraction; RBC:M = RBC to membrane fraction; T_2^* = transverse relaxation time; V = visit; VDP = ventilation defect percentage; VP = ventilation percentage Dai

Dissolved Xenon (Gas Exchange): Figure 5 presents sample RBC:M maps. The global RBC:M ratio significantly increased at visit 2 compared with visit 1 ($P_{adj} = .023$). RBC:M at visit 1 was 0.22 (0.15-0.37), and at visit 2 it was 0.25 (0.18-0.41). No subjects showed a decrease in RBC:M at visit 2 compared with visit 1 (Figs 4, 5). RBC:M at visits 3 and 4 were 0.25 (0.19-0.44) and 0.23 (0.19-0.44), respectively. At visits 3 and 4, some patients showed continued improvement (Figs 3, 4), while others maintained an abnormal RBC:M during the 25- to 51-week period. There were no significant changes between visits 2, 3, and 4.

Figure 6 shows boxplots of the RBC:M, RBC:gas, and M:gas for patients at each visit, with reference boxplots of age- and sex-matched control data (control RBC:M median, 0.39 [IQR, 0.13]; RBC:gas median, 0.0034 [IQR, 0.0006]; M:gas median, 0.0088 [IQR, 0.0021]). The number of patients who had RBC:M below the median \pm IQR of the age- and sex-matched healthy volunteers was eight of nine at visit 1, seven of nine at visit 2, five of seven at visit 3, and six of eight at visit 4.

The T_2^* of the M and RBCs was calculated. M T_2^* showed a significant longitudinal decrease across visits, with lower M T_2^* at visit 4 compared with visits 1, 2, and 3 ($P_{adj} = .023$, $P_{adj} = .023$, and $P_{adj} = .047$, respectively) and between visits 1 and 3 ($P_{adj} = .031$) (Table 2). No other significant changes in the T_2^* of the RBC or M were seen (e-Fig 3).

¹H MRI

Structural Changes: The UTE image of Patient 3 showed abnormal linear parenchymal changes at visit 1, which improved but remained abnormal at visits 2 and 3 and were resolved at visit 4. Patients 2, 6, 7, and 8 displayed air trapping on their UTE image at visit 1, which resolved at visit 2 for patients 6, 7, and 8. Patient 2 continued to have air trapping present at visits 3 and 4. The UTE images of Patients 1, 4, 5, and 9 were normal at all visits (e-Table 2).

DCE (Perfusion): Patient 1 showed a segmental perfusion defect at visit 1 that was resolved at visit 2. No other patients showed any substantial regional perfusion defects. Median pulmonary blood volume and flow increased in all patients (n = 6) at visit 2 compared with visit 1; however, the increase was not statistically significant. For the six patients with DCE MRI at visits 1 and 2, median pulmonary blood volume was 37.8 (11.7-53.5) mL/100 mL at visit 1 and 47.6 (15.0-60.2) mL/100 mL at visit 2, and pulmonary blood flow was

TABLE 2 Continued



Figure 5 – Lung RBC:M maps in three patients with four MRI visits at 6, 12, 25, and 51 weeks following hospital admission. Mean RBC:M at each visit is shown. M = membrane; RBC:M = RBC to membrane fraction.

76.9 (19.6-107.2) mL/100 mL/min at visit 1 and 91.1 (30.7-109.5) mL/100 mL/min at visit 2 (Fig 4).

Pulmonary Function Tests

Data were available on PFTs for six of nine patients at visit 1, six of nine patients at visit 2, seven of seven patients at visit 3, and seven of eight patients at visit 4; all *z* scores and % predicted data are shown in Figure 7.

There was a median of 0 days (mean, 2.8 days; range, 0-23 days) between MRI and PFTs.

Median TL_{CO} z score was -1.66 (-1.96 to 0.66) at visit 1, -0.88 (-1.49 to 0.68) at visit 2, -0.47 (-1.51 to 0.90) at visit 3, and -0.31 (-1.67 to 1.05) at visit 4. Three of six patients had an abnormal TL_{CO} z score (< 1.64) at visit 1. No patients had an abnormal TL_{CO} z score at visit 2 or 3. One patient had an abnormal TL_{CO} z score at visit 4.



Figure 6 – Boxplots of xenon gas transfer ratios from patients at visits 1 to 4 as well as metrics from an age- and sex-matched healthy cohort. Open circles denote data > 1.5 interquartile range; star denotes data > 3 interquartile range. M = membrane.; M:gas = membrane to gas fraction; RBC:gas = RBC to gas fraction; RBC:M = RBC to membrane fraction.



Figure 7 – Spaghetti plots of FEV₁ z score, FVC z score, FEV₁/FVC z score, K_{CO} z score, and TL_{CO} z score. K_{CO} = carbon monoxide transfer coefficient; TL_{CO} = transfer factor for carbon monoxide.

One patient (Patient 5) had abnormally low FVC at visit 1 and visit 4. No other forced lung volume metrics were abnormal at any visits.

Linear Mixed-Effect Model of RBC:M

A significant increase in RBC:M was found with increasing pulmonary blood volume, pulmonary blood flow, decreasing VDP, and increasing $TL_{CO} z$ score, using data from all 4 visits (Table 3). No statistically

significant relationship was found between RBC:M and mean transit time.

Discussion

This study used a comprehensive ¹H and ¹²⁹Xe MRI protocol to assess pathophysiologic pulmonary changes in hospitalized patients with COVID-19 for up to 1 year following hospitalization. At 6 weeks following hospitalization, four of nine patients had small

Independent Variable	Estimated Coefficient	P Value	Lower CI	Upper CI
Pulmonary blood volume (mL/100 mL)	0.0016	.002	0.0007	0.0025
Pulmonary blood flow (mL/100 mL/min)	0.00067	.015	0.00014	0.00120
Mean transit time (s)	0.0082	.076	-0.00093	0.01729
VDP (%)	-0.025	.009	-0.0427	-0.007
TL _{CO} z score	0.048	< .001	0.027	0.069

 TABLE 3] Effect of Pulmonary Blood Volume, Pulmonary Blood Flow, Mean Transit Time, VDP, and TL_{CO} z Score on RBC:M Tested Using Linear Mixed-Effect Model Analysis With a Random Intercept

M = membrane; RBC:M = RBC to membrane fraction; $TL_{CO} =$ transfer factor for carbon monoxide; VDP = ventilation defect percentage.

ventilation defects, $TL_{CO} z$ score was abnormal in three of nine patients, and xenon gas transfer (RBC:M) was outside the median \pm IQR of age- and sex-matched healthy subjects in eight of nine patients. At 12 weeks, improvements were seen in lung ventilation and xenon gas transfer. However, there was no longitudinal change in xenon gas transfer between 12 and 52 weeks, and median ¹²⁹Xe gas transfer in these patients remained lower than expected. This indicates that some of the patients with COVID-19 exhibited continued abnormalities in ¹²⁹Xe gas transfer at 12 to 51 weeks following hospitalization, despite normal lung structural imaging and ventilation, with six of eight patients outside the median \pm IQR of normal age- and sexmatched patients at 51 weeks.

Several studies have reported reduced gas transfer to the RBCs in patients hospitalized due to COVID-19.²¹⁻²³ Because xenon gas transfer depends on both the xenon uptake in the lung tissue and the xenon uptake in the RBCs, a combination of lung perfusion abnormalities and/or alveolar/interstitial endothelial changes may be mechanistically driving the reduced xenon gas transfer seen in patients following COVID-19. Although not directly comparable to the results from those studies due to differences in imaging parameters, our findings are in accordance with the reporting of significantly lower RBC:M values between hospital discharge and 24 weeks postdischarge in previous studies.²⁰⁻²² In the current study, the inclusion of data from age- and sex-matched healthy control subjects shows that these changes are not due to age or sex differences between control subjects and patients in this study. RBC:gas and M:gas did not show significant longitudinal change once adjusted for multiple comparisons, implying that the change in RBC:M was a combined effect of changes in both M and RBC. A significant reduction in M T_2^* at visit 2 was also found. The physiological mechanisms behind changes in M T2* are not well established and are

discussed further in the Discussion section of e-Appendix 1

We also found that changes in xenon gas transfer increased significantly with increased $TL_{CO} z$ score, VDP, and lung perfusion metrics (pulmonary blood volume and pulmonary blood flow). All patients with DCE data available displayed an increase in regional pulmonary blood flow and volume between visits 1 and 2, despite only one having a substantial perfusion defect. This may indicate microvascular improvements at 12 weeks, and that microvascular recovery may be partially driving changes in RBC:M in these patients. In parallel, a concomitant reduction in M signal due to resolution of postinfection endothelial inflammation could contribute to the increase in RBC:M with time.

Although we see global correlations between RBC:M, ventilation, and perfusion, regional heterogeneity in RBC:M did not visually agree with ventilation or perfusion heterogeneity; for example, as shown in Figure 3, Patient 8 has a visually heterogeneous RBC:M map but no visual concordance with pulmonary blood flow heterogeneity and homogeneous ventilation on the similar slices presented. Further work assessing regional distributions seen in the different functional magnetic resonance images available here is warranted to evaluate regional correlations quantitatively.

In this study, most patients (seven of nine) did not report significant breathlessness at visit 2 (12 weeks), despite lower RBC:M than the control reference data. The two patients who reported breathlessness at visit 2 had the two lowest RBC:M values at that visit. Larger studies in symptomatic patients are needed to further investigate links between RBC:M and breathlessness or other post-COVID-19 symptoms. Fully recovered post-COVID-19 control groups will be important in further studies investigating post-COVID-19 breathlessness with these imaging techniques. Median patient ADC and Lm_D were within the ageand sex-matched reference range at visits 1 and 2,³⁵ with no significant change at visit 2, indicating that airway dimensions were not increased in these nine patients who had COVID-19 but no signs of interstitial lung damage on structural imaging. This study excluded patients with signs of interstitial lung damage, as previous work has shown that patients with interstitial lung diseases can have reduced xenon gas transfer,⁶ alterations in lung microstructure measured using ¹²⁹Xe MRI,⁶ reductions in lung ventilation,⁴⁶ and reductions in lung perfusion.⁴⁷ Although this means that there is considerable promise for lung MRI to provide longitudinal biomarkers in patients with signs of interstitial lung damage, it also suggests that persistent perfusion, ventilation, gas transfer, and lung microstructure abnormalities may be mechanistically related to the visible tissue changes within a cohort with structural lung abnormalities. Further work using a ¹H and ¹²⁹Xe protocol in patients with established pulmonary fibrosis due to COVID-19 on CT scan imaging is the subject of an ongoing study (UKILD).²⁹

Minor ventilation heterogeneity and defects were present in this cohort shortly following acute illness; these defects improved over time, which is consistent with the findings of Grist et al²¹ and of Li et al.²⁰ Overall, the current study and the findings from previous literature suggest it is unlikely that impaired lung ventilation is the primary cause of ongoing symptoms following the acute stage of COVID-19 and that the pathophysiology is not primarily of the airways.

The main limitation of the current study is the limited number of participants, which was largely caused by the challenging nature of recruiting patients for scanning directly following a recent hospitalization due to COVID-19 in the first wave of the pandemic. In addition, not all patients had DCE lung perfusion imaging or PFTs at all examinations (due to aerosolization constraints). The numbers recruited limit correlations with symptoms, activity, and lung function, as well as the statistical tests used to test for change. All ¹²⁹Xe acquisitions were acquired at functional residual capacity plus 1 L, resulting in some variability between patients in the lung inflation state. A final potential source of bias in this study is that five patients who were potentially eligible for the study were excluded due to chest size exceeding the size of the xenon MRI coil.

Interpretation

This study found that in a cohort of patients who were hospitalized with COVID-19 pneumonia of moderate severity who had normal CT scan/lung structural imaging, ¹²⁹Xe gas transfer improved at 12 weeks but did not return to within a normal range within 1 year following hospitalization. Improvements in ¹²⁹Xe gas transfer were associated with an increased lung perfusion on DCE-MRI and increased TL_{CO} *z* score; therefore, abnormalities in ¹²⁹Xe gas transfer may be a marker of ongoing microvascular abnormalities post-COVID-19.

 TL_{CO} *z* score was within a normal range for seven of eight patients with available data at 51 weeks posthospitalization. This indicates that ¹²⁹Xe gas transfer may be a more sensitive measure of gas exchange in this population and that it may be able to identify abnormalities that routine clinical tests overlook.

We believe this to be the first follow-up study of similar patients with such an extensive range of functional lung imaging techniques. Our findings show the sensitivity and complementary nature of functional MRI to followup post-COVID-19 lung pathophysiology in a clinical setting.

Funding/Support

This study was supported by a Medical Research Council grant (MR/M008894/1), a GSK investigator-led grant funding (R/167242-11-1) and a GE Healthcare investigator-led grant funding (R/167303-11-1) to J. M. W. A. A. R. T. was supported by a British Heart Foundation Intermediate Clinical Fellowship (FS/18/13/ 33281). The study was supported by the NIHR Sheffield Biomedical Research Centre (BRC) / NIHR Sheffield Clinical Research Facility (CRF). Grant funding from the NIHR was received by A. J. S. (AI award: AI_AWARD01706), F. G. (EXPLAIN study: COV-LT2-0049), and K. M. H. for the development of an MRI sequence used in this work (5R01HL136965).

Financial/Nonfinancial Disclosures

The authors have reported to *CHEST* the following: The following authors have declarations of support from organizations for the submitted work: P. J. C. H. receives grant funding from GSK and Bayer. A. A. R. T. is funded by British Heart Foundation Intermediate Clinical Fellowship and grant funding from Janssen-Cilag Ltd. A. L. receives grant funding from the British Heart

Foundation (fellowship award). R. G. J. receives grant funding from AstraZeneca, Biogen, Galecto, GSK, RedX, Pliant, and Genentech. A. J. S. receives grant funding from the National Institute for Health and Care Research (NIHR) (AI award), Wellcome (Innovator award), and Janssen-Cilag Ltd (project grant). J. M. W. receives grant funding from the Medical Research Council, GSK (investigator led research grant), and GE Healthcare. F. G. receives grant or contract funding from Oxford NIHR Biomedical Research Centre, NIHR (HypErpolarised Xenon Magnetic Resonance PuLmonary Imaging in PAtIeNts with Long-COVID -EXPLAIN), POLAREAN Ltd, and GE Healthcare. K. M. J. has received National Institutes of Health grant funding for the development of an MRI sequence used in this work. The following authors declare financial relationships with organizations that might have an interest in the submitted work in the previous 3 years: A. C. is an employee of GSK and is a shareholder in GSK. F. J. W. was an employee of GSK and was a shareholder in GSK at the time of the study. R F. S. is employed by, and a shareholder of, GE Healthcare. A. L. receives funding support from Janssen-Cilag Ltd for meetings/travel. A. A. R. T. receives funding from Janssen-Cilag Ltd for meetings/travel. R. G. J. receives consulting fees from Bristol Myers Squibb, Daewoong, Veracyte, Resolution Therapeutics, RedX, Pliant, and Chiesi. A. J. S. receives consultancy fees from Janssen-Cilag Ltd. R. G. J. receives payment from Chiesi, Roche, patientMpower, AstraZeneca, GSK, and Boehringer Ingelheim for lectures. A J. S. receives payment for presentations from Janssen-Cilag Ltd. F. G. receives payment for attending a research advice meeting from POLAREAN Ltd. R. G. J. is a trustee of Action for Pulmonary Fibrosis. F. G. is the president of the European Society for Thoracic Imaging. None declared (L. C. S., G. J. C., H.-F. C., L. J. S., J. G. R. W., J. E. M., Z. G., T. N., M. P., J. A. E., S. T., S. S., L. G., H. M., J. B., D. A. C., L. A., J. R., M. B., A. M. B., G. N., O. R., M. R. R., R. M., N. J. S., J. T. G., S. R., G. H. M., R. L., P. J. C., J. E. B., P. W., L. W.).

Acknowledgments

Author contributions: J. M. W. and L. C. S. had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. J. M. W., A. A. R. T., R. L., P. J. C., G. H. M., A. C., F. J. W., K. M. J., R. F. S., F. G., R. G. J., A. L., J. T. G., and G. J. C. conceived and designed the study. L. C. S., G. J. C., H.-F. C., P. J. C. H., L. J. S., J. M. W., J. E. M., Z. G., T. N., M. P., P. W., J. A. E., J. B., S. T., S. S., L. G., H. M., D. A. C., L. A., J. R., M. B., A. M. B., M. R. R., G. N., O. R., R. M., J. E. B., N. J. S., A. J. S., S. R., and L. W. obtained, prepared, and analyzed the data. L. C. S. wrote the first draft. All authors assume responsibility for the overall content and integrity of the article. All authors were involved in reviewing and shaping the manuscript, and all approved the final version prior to submission.

Role of sponsors: The sponsor had no role in the design of the study, the collection and analysis of the data, or the preparation of the manuscript.

Additional information: The e-Appendix, e-Figures, and e-Tables are available online under "Supplementary Data."

References

- Lang M, Som A, Carey D, et al. Pulmonary vascular manifestations of COVID-19 pneumonia. *Radiol Cardiothorac Imaging*. 2020;2(3): e200277.
- Loo J, Spittle DA, Newnham M. COVID-19, immunothrombosis and venous thromboembolism: biological mechanisms. *Thorax*. 2021;76(4):412-420.

- Attaway AH, Scheraga RG, Bhimraj A, Biehl M, Hatipoglu U. Severe Covid-19 pneumonia: pathogenesis and clinical management. *BMJ*. 2021;372:n436.
- 4. Robey RC, Kemp K, Hayton P, et al. Pulmonary sequelae at 4 months after COVID-19 infection: a single-centre experience of a COVID follow-up service. *Adv Ther.* 2021;38(8):4505-4519.
- Stewart NJ, Smith LJ, Chan HF, et al. Lung MRI with hyperpolarised gases: current & future clinical perspectives. Br J Radiol. 2022;95(1132):20210207.
- Collier GJ, Eaden JA, Hughes PJC, et al. Dissolved (129) Xe lung MRI with fourecho 3D radial spectroscopic imaging: quantification of regional gas transfer in idiopathic pulmonary fibrosis. *Magn Reson Med.* 2021;85(5):2622-2633.
- Wielputz MO, Puderbach M, Kopp-Schneider A, et al. Magnetic resonance imaging detects changes in structure and perfusion, and response to therapy in early cystic fibrosis lung disease. *Am J Respir Crit Care Med.* 2014;189(8): 956-965.
- Ohno Y, Koyama H, Matsumoto K, et al. Dynamic MR perfusion imaging: capability for quantitative assessment of disease extent and prediction of outcome for patients with acute pulmonary thromboembolism. J Magn Reson Imaging. 2010;31(5):1081-1090.
- 9. Thomen RP, Walkup LL, Roach DJ, et al. Regional structure-function in cystic fibrosis lung disease using hyperpolarized (129)Xe and ultrashort echo magnetic resonance imaging. *Am J Respir Crit Care Med.* 2020;202(2): 290-292.

- Tafti S, Garrison WJ, Mugler JP 3rd, et al. Emphysema index based on hyperpolarized (3)He or (129)Xe diffusion MRI: performance and comparison with quantitative CT and pulmonary function tests. *Radiology*. 2020;297(1):201-210.
- McIntosh MJ, Kooner HK, Eddy RL, et al. CT mucus score and ¹²⁹Xe MRI ventilation defects after 2.5 years' anti-IL-5Rα in eosinophilic asthma. *Chest.* 2023;164(1):27-38.
- Kaushik SS, Cleveland ZI, Cofer GP, et al. Diffusion-weighted hyperpolarized Xe-129 MRI in healthy volunteers and subjects with chronic obstructive pulmonary disease. *Magn Reson Med.* 2011;65(4):1155-1165.
- Chan HF, Weatherley ND, Johns CS, et al. Airway microstructure in idiopathic pulmonary fibrosis: assessment at hyperpolarized He-3 diffusion-weighted MRI. *Radiology*. 2019;291(1):223-229.
- Marshall H, Stewart NJ, Chan HF, Rao M, Norquay G, Wild JM. In vivo methods and applications of xenon-129 magnetic resonance. *Prog Nucl Magn Reson Spectrosc.* 2021;122:42-62.
- Wang ZY, Rankine L, Bier EA, et al. Using hyperpolarized Xe-129 gas-exchange MRI to model the regional airspace, membrane, and capillary contributions to diffusing capacity. J Appl Physiol. 2021;130(5): 1398-1409.
- **16.** Weatherley ND, Stewart NJ, Chan HF, et al. Hyperpolarised xenon magnetic resonance spectroscopy for the longitudinal assessment of changes in gas diffusion in IPF. *Thorax.* 2019;74(5): 500-502.

- Wang JM, Robertson SH, Wang Z, et al. Using hyperpolarized (129)Xe MRI to quantify regional gas transfer in idiopathic pulmonary fibrosis. *Thorax.* 2018;73(1): 21-28.
- Myc L, Qing K, He M, et al. Characterisation of gas exchange in COPD with dissolved-phase hyperpolarised xenon-129 MRI. *Thorax*. 2021;76(2):178-181.
- **19.** Wang Z, Robertson SH, Wang J, et al. Quantitative analysis of hyperpolarized (129) Xe gas transfer MRI. *Med Phys.* 2017;44(6):2415-2428.
- 20. Li H, Zhao X, Wang Y, et al. Damaged lung gas exchange function of discharged COVID-19 patients detected by hyperpolarized (129)Xe MRI. *Sci Adv.* 2021;7(1):eabc8180.
- Grist JT, Chen M, Collier GJ, et al. Hyperpolarized (129)Xe MRI abnormalities in dyspneic patients 3 months after COVID-19 pneumonia: preliminary results. *Radiology*. 2021;301(1):E353-E360.
- 22. Grist JT, Collier GJ, Walters H, et al. Lung abnormalities depicted with hyperpolarized xenon MRI in patients with long COVID. *Radiology*. 2022;305(3): 709-717.
- Matheson AM, McIntosh MJ, Kooner HK, et al. Persistent (129)Xe MRI pulmonary and CT vascular abnormalities in symptomatic individuals with post-acute COVID-19 syndrome. *Radiology*. 2022;305(2):466-476. 220492.
- Wild JM, Collier G. (129)Xe Pulmonary MRI for individuals with post-acute COVID-19 syndrome. *Radiology*. 2022;305(2):477-478.
- 25. Yang S, Zhang Y, Shen J, et al. Clinical potential of UTE-MRI for assessing COVID-19: patient- and lesion-based comparative analysis. *J Magn Reson Imaging*. 2020;52(2):397-406.
- 26. Johns CS, Swift AJ, Rajaram S, et al. Lung perfusion: MRI vs. SPECT for screening in suspected chronic thromboembolic pulmonary hypertension. J Magn Reson Imaging. 2017;46(6):1693-1697.
- 27. Rysz S, Al-Saadi J, Sjostrom A, et al. COVID-19 pathophysiology may be driven by an imbalance in the renin-

angiotensin-aldosterone system. *Nat Commun.* 2021;12(1):2417.

- 28. Yu JZ, Granberg T, Shams R, et al. Lung perfusion disturbances in nonhospitalized post-COVID with dyspnea—a magnetic resonance imaging feasibility study. *J Intern Med.* 2022;292(6):941-956.
- 29. Wild JM, Porter JC, Molyneaux PL, et al. Understanding the burden of interstitial lung disease post-COVID-19: the UK Interstitial Lung Disease-Long COVID Study (UKILD-Long COVID). BMJ Open Respir Res. 2021;8(1):e001049.
- 30. Stanojevic S, Graham BL, Cooper BG, et al. Official ERS technical standards: Global Lung Function Initiative reference values for the carbon monoxide transfer factor for Caucasians. *Eur Respir J.* 2017;50(3):1700010.
- Quanjer PH, Stanojevic S, Cole TJ, et al. Multi-ethnic reference values for spirometry for the 3-95-yr age range: the Global Lung Function 2012 equations. *Eur Respir J.* 2012;40(6):1324-1343.
- Norquay G, Collier GJ, Rao M, Stewart NJ, Wild JM. ~{129}Xe-Rb spin-exchange optical pumping with high photon efficiency. *Phys Rev Lett.* 2018;121(15): 153201.
- 33. Smith LJ, Horsley A, Bray J, et al. The assessment of short- and long-term changes in lung function in cystic fibrosis using Xe-129 MRI. *Eur Respir J*. 2020;56: 2000441.
- 34. Stewart NJ, Norquay G, Griffiths PD, Wild JM. Feasibility of human lung ventilation imaging using highly polarized naturally abundant xenon and optimized three-dimensional steady-state free precession. *Magn Reson Med.* 2015;74(2): 346-352.
- Chan HF, Stewart NJ, Norquay G, Collier GJ, Wild JM. 3D diffusionweighted (129) Xe MRI for whole lung morphometry. *Magn Reson Med.* 2018;79(6):2986-2995.
- Johnson KM, Fain SB, Schiebler ML, Nagle S. Optimized 3D ultrashort echo time pulmonary MRI. *Magn Reson Med.* 2013;70(5):1241-1250.
- Li KL, Zhu XP, Waterton J, Jackson A. Improved 3D quantitative mapping of blood volume and endothelial

permeability in brain tumors. J Magn Reson Imaging. 2000;12(2):347-357.

- 38. Cheng HL, Wright GA. Rapid highresolution T(1) mapping by variable flip angles: accurate and precise measurements in the presence of radiofrequency field inhomogeneity. *Magn Reson Med.* 2006;55(3):566-574.
- 39. Chan HF, Collier GJ, Parra-Robles J, Wild JM. Finite element simulations of hyperpolarized gas DWI in micro-CT meshes of acinar airways: validating the cylinder and stretched exponential models of lung microstructural length scales. *Magn Reson Med.* 2021;86(1):514-525.
- Chatfield M, Mander A. The Skillings-Mack test (Friedman test when there are missing data). *Stata J.* 2009;9(2):299-305.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc Series B Stat Methodol.* 1995;57(1):289-300.
- **42.** Team RC. R: A language and environment for statistical computing. *MSOR connections*, 2014:1.
- Petersson-Sjogren M, Chan HF, Collier GJ, et al. Airspace dimension assessment (AiDA) by inhaled nanoparticles: benchmarking with hyperpolarised (129)Xe diffusionweighted lung MRI. *Sci Rep.* 2021;11(1): 4721.
- 44. Knight SR, Ho A, Pius R, et al. Risk stratification of patients admitted to hospital with covid-19 using the ISARIC WHO Clinical Characterisation Protocol: development and validation of the 4C Mortality Score. *BMJ*. 2020;370: m3339.
- 45. Williams B. Evaluation of the utility of NEWS2 during the COVID-19 pandemic. *Clin Med (Lond).* 2022;22(6):539-543.
- **46.** Tibiletti M, Eaden JA, Naish JH, et al. Imaging biomarkers of lung ventilation in interstitial lung disease from (129)Xe and oxygen enhanced (1)H MRI. *Magn Reson Imaging*. 2023;95:39-49.
- 47. Weatherley ND, Eaden JA, Hughes PJC, et al. Quantification of pulmonary perfusion in idiopathic pulmonary fibrosis with first pass dynamic contrast-enhanced perfusion MRI. *Thorax.* 2021;76(2): 144-151.