



Deposited via The University of Sheffield.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/id/eprint/190835/>

Version: Published Version

Article:

Chimplee, S., Smythe, C., Tipmanee, V. et al. (2022) Anticancer mechanism of 7- α -hydroxyfrullanolide on microtubules and computational prediction of its target binding in triple-negative breast cancer cells. PeerJ, 10. e13508. ISSN: 2167-8359

<https://doi.org/10.7717/peerj.13508>

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:

<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Anticancer mechanism of 7- α -hydroxyfrullanolide on microtubules and computational prediction of its target binding in triple-negative breast cancer cells

Siriphorn Chimplee¹, Carl Smythe², Varomyalin Tipmanee¹, Suchada Sukrong³ and Kanyanatt Kanokwiroon¹

¹ Department of Biomedical Sciences and Biomedical Engineering, Faculty of Medicine, Prince of Songkla University, Hat Yai, Songkhla, Thailand

² School of Biosciences, University of Sheffield, Sheffield, United Kingdom

³ Center of Excellence in DNA Barcoding of Thai Medicinal Plants, Department of Pharmacognosy and Pharmaceutical Botany, Faculty of Pharmaceutical Sciences, Chulalongkorn University, Bangkok, Thailand

ABSTRACT

Background. Triple-negative breast cancer (TNBC) responds poorly to the available drugs; thus, the mortality rate associated with TNBC remains high. 7- α -Hydroxyfrullanolide (7HF) possesses anticancer properties and arrests cells in the G2/M-phase via modulation of several proteins involved in the G2/M-phase transition, as well as the mitotic checkpoint in MDA-MB-468 (TNBC) cells. Microtubules (MTs) dynamically regulate cell division in the G2/M phase and are related to cancer cell stress response. However, antimitotic drug cytotoxicity to multiple cancer resistance developed in response to drugs are obstacles faced to date. Here, the activity and mechanism via which 7HF controls MTs dynamics was investigated in MDA-MB-468 cells.

Methods. 7HF uptake by MDA-MB-468 cells was assessed using spectrophotometry. The drug-like properties of 7HF were predicted using the Swiss-absorption, distribution, metabolism, and excretion (ADME) webtool. Then, the effect of 7HF treatment (6, 12, and 24 μ M) on the dynamic arrangement of MTs was assessed for 1, 12, and 24 h using indirect immunofluorescence. Polymerization of α - and β -tubulin was assessed using different 7HF concentrations in a cell-free system for 1 h. Cell proliferation assay with bromodeoxyuridine plus propidium iodide staining and flow cytometry was performed at different 7HF concentrations and time points. The mechanism of action was assessed by detecting the expression of proteins, including Bub3, cyclin B1, p-Cdk1 (Tyr15), Rb, p-Rb (Ser780), Chk1, p-Chk1 (Ser345), Chk2, p-Chk2 (Ser516), and p-H2AX (Ser139), using western blotting. Molecular docking was used to predict the molecular interactions between 7HF and tubulins in MTs.

Results. We observed that 7HF was able to enter the MDA-MB-468 cells. The ADME webtool analysis predicted that it possesses the high passive permeation and gastrointestinal absorption properties of drugs. Various concentrations of 7HF disrupted the dynamic arrangement of spindle MTs by causing radial spindle array shrinkage and expansion of fibrous spindle density and radial array lengths in a time-dependent manner. 7HF reduced polymerization of α -, β -tubulin in dose-dependent manner.

Submitted 17 February 2022

Accepted 6 May 2022

Published 27 May 2022

Corresponding author
Kanyanatt Kanokwiroon,
kanyanatt.k@psu.ac.th

Academic editor
Vladimir Uversky

Additional Information and
Declarations can be found on
page 17

DOI 10.7717/peerj.13508

© Copyright
2022 Chimplee et al.

Distributed under
Creative Commons CC-BY 4.0

OPEN ACCESS

7HF also triggered DNA damage response by inducing G2/M and G1 phase arrests in a concentration and time-dependent manner, which occurred due to the upregulation of Bub3, Chk1, p-Chk1 (Ser345), p-Cdk1 (Tyr15), and cyclin B1. According to molecular docking analysis, 7HF preferred to bind to β -tubulin over α -tubulin. The lactone, ketone, and hydroxyl groups of 7HF supported the 7HF-tubulin interactions. Hydrogen bonding with a hydrocarbon ring and salt bridge attractive forces were responsible for the binding versatility of 7HF.

Conclusions. This is the first study to investigate the molecular mechanism, MTs interacting sites, and the internalization and drug-like properties of 7HF in TNBC cells. The findings will be useful for developing 7HF-based treatment for patients with TNBC.

Subjects Biochemistry, Cell Biology, Computational Biology, Molecular Biology

Keywords 7- α -Hydroxyfrullanolide, Sesquiterpene lactones, Natural product, Triple-negative breast cancer, Microtubules, G2/M arrest, G1 arrest, DNA damage response, Molecular docking

INTRODUCTION

Female breast cancer is the most common (2.3 million new cases) and the fifth leading cause of cancer-related mortality (0.68 million cases) worldwide, according to GLOBOCAN 2020 (Ferlay et al., 2021; Sung et al., 2021). Triple-negative breast cancer (TNBC) accounts for 15–20% of all breast cancer cases (Nunez Abad et al., 2021). TNBC exhibits poor biological behavior, with more aggressiveness, earlier recurrence, and ability to metastasize to distant organs than non-TNBC subtypes because of the lack of available therapeutic targets, such as estrogen receptor, progesterone receptor, and human epidermal growth factor receptor 2 (Nunez Abad et al., 2021). Therefore, the development of targeted therapies for patients with TNBC has been challenging (Nunez Abad et al., 2021). The high response rate to standard chemotherapy combinations (anthracycline taxanes with vinca alkaloid or targeted drugs) appears to improve the pathological complete response of a neoadjuvant or adjuvant regimen; however, an early recurrence of TNBC is high in such cases due to poor efficacy of prolonged chemotherapy (Bianchini et al., 2021; Nunez Abad et al., 2021; Wang et al., 2018). Thus, chemotherapy in TNBC is associated with low survival, high rate of relapse, high risk of secondary cancers, and strong adverse effects (De Ruijter et al., 2011; Liao, Apaya & Shyur, 2013; Nunez Abad et al., 2021; Sharma, 2016; Sharma, 2018).

7- α -Hydroxyfrullanolide (7HF), a natural product, is a eudesmanolide sesquiterpene lactone (SL) and is isolated from the flowering plants of the Asteraceae family, such as *Grangea maderaspatana* L. Poir. and *Sphaeranthus indicus* Linn. (Nahata et al., 2013; Pandey et al., 2019; Ruangrungsi et al., 1989). 7HF is used to produce its bioactive derivatives via fungal transformation (Atta ur et al., 1994). It also inhibits DNA topoisomerase I (Top I) in yeast cells and possesses anti-inflammatory and anticancer activities (Fonseca et al., 2010; Nahata et al., 2013; Pandey et al., 2019; Uppatanpreecha, 2009). It inhibits the proliferation of certain types of cancer cells, such as small cell lung cancer, oral cavity cancer, colon cancer, and breast cancer cells (Fonseca et al., 2010; Nahata et al., 2013; Pandey et al., 2019; Uppatanpreecha, 2009). A previous study on the

anti-breast cancer activity of 7HF showed that TNBC cells (MDA-MB-468, MDA-MB-231, and Hs578T) are more sensitive to 7HF than non-TNBC cells (MCF-7) (Chimplee et al., 2022). 7HF has been reported to be associated with G2/M arrest and apoptosis signaling pathways in TNBC (MDA-MB-468) and colon cancer cells (Chimplee et al., 2022; Nahata et al., 2013; Pandey et al., 2019). Notably, 7HF triggers G2/M arrest by modulating the expression of several proteins involved in the regulation of G2/M phase transition, mitotic chromosome segregation, mitotic checkpoint regulation, and mitotic spindle organization in MDA-MB-468 cells (Chimplee et al., 2022). Mitotic associated proteins of tubulins and Bub3 mitotic checkpoint were significantly upregulated by 7HF in the TNBC cells indicating through controlling of MTs formation (Chimplee et al., 2022). Hence, the additional molecular mechanism of the anticancer activity of 7HF on MTs in TNBC cells should also be elucidated.

The microtubule (MT) cytoskeleton is a highly dynamic structure (Avila, 1992). MTs consist of α -, β -tubulin subunits and play a critical role in cell division, cell movement, and intracellular signaling (Mollinedo & Gajate, 2003). MT stress induces G2/M phase cell cycle arrest, which is one of the mechanisms of action of antimetabolic drugs used in cancer treatment, such as taxanes and vinca alkaloids (Gascoigne & Taylor, 2009; Parker, Kavallaris & McCarroll, 2014; Tischer & Gergely, 2019). In addition, sesquiterpene lactones (SLs) exhibit anti-MT activity by regulating MT dynamics, and the proposed mechanism of action involves a target protein associated with an arresting of G2/M or M-phases in many cancer cells (Bosco & Golsteyn, 2017). SLs, including parthenolide, costunolide, artesunate, santamarine, calein C, 6-O-angeloylplenolin, coronopilin, pulchelloid A, and hymenoratin, have shown promising anti-MT activity by inducing G2/M or M phase arrest in tested cancer cell lines (Bosco & Golsteyn, 2017; Bosco et al., 2021; Caldas et al., 2018; Cotugno et al., 2012; Dumontet & Jordan, 2010; Liu et al., 2011a; Liu et al., 2011b; Manchado, Guillamot & Malumbres, 2012; Molina et al., 2021; Steinbruck, Pereira & Efferth, 2010; Whipple et al., 2013). The anti-MT activity of SLs has been studied extensively and their mechanism of action has been reported; however, the specific overcome to target MT-binding is not sufficient (Bosco & Golsteyn, 2017; Gallagher & Brian, 2007). Therefore, studies for the development of new MT-binding agents that can be used for the development of TNBC-targeted drugs are in progress. Internalization and accumulation of drugs in cancer cells are also crucial for improving the chemotherapeutic efficacy (Nunez Abad et al., 2021; Piccolo et al., 2019). Hence, the stability of drugs in TNBC cells should also be assessed.

In this study, we aimed to investigate the effects of 7HF on MT dynamics and its molecular mechanism of action. Cellular uptake of 7HF, MT-7HF interaction, and the drug-like properties of 7HF were also determined.

MATERIALS AND METHODS

Plant material preparation and 7HF isolation

Dried *G. maderaspatana* (L.) Poir plants were purchased from the Chaokromper Drug Store (Bangkok, Thailand). The plant material was prepared, and 7HF was isolated as described previously (Chimplee et al., 2022). The chemical structure of 7HF is shown in Fig. 1A. 7HF was stored at -20°C .

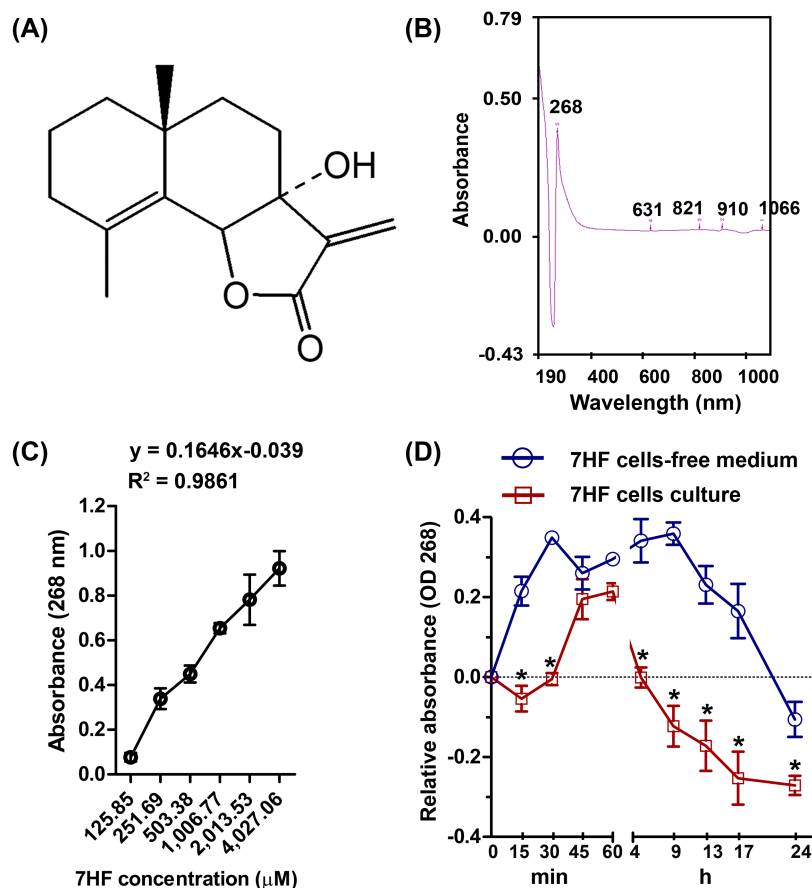


Figure 1 7- α -Hydroxyfrullanolide (7HF) uptake by MDA-MB-468 cells. (A) Chemical structure of 7HF. (B) The identical peaks of 7HF at 268 nm and 1,006.77 μ M. (C) 7HF calibration curve at 268 nm. 7HF concentration ranged from 125.85 to 4,027.06 μ M. (D) 7HF uptake profile of MDA-MB-468 cells compared to that of the control (7HF-cells-free medium). Mean differences were analyzed using Two-way ANOVA and Bonferroni posttests multiple comparisons between cells- and cells-free cultures ($*p < 0.001$). The amount of 7HF was subtracted from that present in only medium and DMSO-medium (in cells-free medium) and in cells-medium and DMSO-cells medium (in cells culture).

Full-size DOI: 10.7717/peerj.13508/fig-1

Cancer cell culture

The MDA-MB-468 cells were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) and cultured in Dulbecco's modified Eagle's medium (DMEM) (Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA) with condition as described previously (Chimplee *et al.*, 2019; Chimplee *et al.*, 2022).

7HF uptake assay

7HF (125.85–4,027.06 μ M) dilutions in dimethyl sulfoxide (DMSO) were scanned to identify the maximum absorbance (λ_{\max}) between 190 and 1,100 nm, using a UV-visible spectrophotometer (Shimadzu, Kyoto, Japan). The λ_{\max} was obtained at 268 nm and at a concentration of 1,006.77 μ M (Fig. 1B and Fig. S1). To test whether 7HF was able to pass through the TNBC cells, the cells were treated with 1,006.77 μ M 7HF. MDA-MB-468 cells

(3×10^4 cells/well) were seeded on 96-well plate and incubated with the DMEM medium overnight. Then, 100 μ L of 1,006.77 μ M 7HF was used to treat the TNBC cells for 0.25–24 h at 37 °C in a 5% CO₂ incubator. In case of 7HF cells-free medium, 1,006.77 μ M of 7HF was prepared with only medium. After incubation, the remaining of 7HF supernatant in cells culture medium and cells-free medium was immediately collected without washing with buffer. The absorbance of 7HF was then measured at 268 nm and the absorbance was plotted *versus* time (Fig. 1D and Table S2). The mean absorbance values of 7HF were subtracted from the absorbance value of only medium and DMSO-supplemented medium (in cells-free medium), and cells medium and cells-DMSO medium (in cells-culture). The experiment was performed in triplicate. In addition, a concentration curve of 7HF was plotted on a linear scale at 268 nm and concentrations ranging from 125.85 to 4,027.06 μ M (Fig. 1C and Table S1). The linear regression coefficients were calculated using the general equation of the calibration curve ($y = ax + b$; $a = 0.1646$, $b = -0.039$, and $R^2 = 0.9861$) (Fig. 1C).

Immunofluorescence (IF) assay

The effect of 7HF on MT dynamics in the TNBC cells was visualized using indirect IF, with certain modifications (Duangmano *et al.*, 2012). In brief, MDA-MB-468 cells (1.5×10^5 cells/well) were seeded on cover slips and incubated at 37 °C in a 5% CO₂ incubator overnight. After washing with phosphate-buffered saline (PBS), the TNBC cells were treated with 6, 12, and 24 μ M 7HF for 1, 12, and 24 h. The 7HF concentrations were obtained from 0.5X, 1X, and 2X its IC₅₀ by MTT assay at 72 h, as reported previously (Chimlee *et al.*, 2022). Paclitaxel (PTX), vincristine (VCR), and doxorubicin (DOX), 200 nM each, were also used to treat the TNBC cells for 24 h, with minor adaptation (Blajeski *et al.*, 2002; Mukhtar, Adhami & Mukhtar, 2014). The control was untreated cells with indicated concentrations of 7HF and chemotherapies. Then, the cells were fixed with 4% formaldehyde, permeabilized with 0.5% Triton X-100, and blocked in 1% bovine serum albumin (BSA) in PBS. After washing the cover slips, the untreated and treated cells were stained overnight with anti- α -tubulin antibody (Cell Signaling Technology, Inc., Danvers, MA, USA; 1:50) and DAPI (Sigma-Aldrich, St. Louis, MO, USA; 1:1,000), followed by incubation with Alexa Fluor 488-conjugated goat anti-IgG rabbit secondary polyclonal antibody (GE Healthcare Ltd., Little Chalfont, Bu, UK; 1:200) for 1 h. Finally, the stained coverslips were mounted with 50% glycerol and sealed on a glass slide. The slides were visualized under an inverted fluorescence microscope (EVOS, Thermo Fisher Scientific Inc.).

Tubulin polymerization assay in cell-free system

The effects of 7HF on tubulin polymerization were analyzed using the tubulin polymerization assay kit (Cytoskeleton Inc., Denver, CO, USA). The assay was performed following the manufacturer's instructions. Briefly, 2 mg/mL tubulin was diluted using tubulin polymerization buffer. After pre-warming of empty dark 96-well plate at 37 °C in a Spark microplate reader (Tecan, Männedorf, Switzerland) for 5 min, 5 μ L 7HF (6, 12, and 24 μ M), PTX (3 μ M), VCR (3 μ M), DOX (3 μ M), and tubulin control were added,

and the plate was incubated for 1 min, after which 50 μL of tubulin reaction reagent was manually injected into the microplate reader immediately. The effect of reagents on tubulin polymerization was then measured at 360 and 450 nm at 37 °C, every minute up to 1 h in kinetic mode.

Cell proliferation assay using fluorescence-activated cell sorting (FACS)

To study the effect of MT dynamics on DNA damage in 7HF-treated cells, we first treated TNBC cells with anti-bromodeoxyuridine (BrdU) and then incubated them with propidium iodide (PI) to stain the newly synthesized DNA. The protocol for this experiment was adapted from *Cecchini, Amiri & Dick (2012)* and *Zhu (2012)*, with some modifications. The MDA-MB-468 cells (1×10^6 cells/plate) were cultured and pulsed with 25 μM BrdU (Cayman Chemical, Ann Arbor, MI, USA) for 30 min. Untreated and 7HF-treated cells (6, 12, and 24 μM) were harvested after 12 and 24 h. Then, 1×10^6 cells were counted. After centrifugation (7,000 rpm, 5 mins), the cell pellets were resuspended in 2 mM EDTA, fixed with 70% EtOH, and stored overnight at -20 °C. Then, the cells were harvested and resuspended in washing buffer (0.5% BSA in PBS). After centrifugation (10,000, 2 mins), the cell pellets were resuspended in 2 M HCl and sodium borate (pH 8.5) solution and incubated for 30 min. The cell pellets were washed with washing buffer containing 0.2% Tween-20. After centrifugation (10,000 rpm, 2 mins), anti-BrdU primary antibody (20 μL) and RNase (200 $\mu\text{g}/\text{mL}$) were added, and the cells were incubated overnight at 4 °C. Then, Alexa Fluor 488-conjugated goat-anti-IgG mouse secondary antibody (Thermo Fisher Scientific, Inc.; 50 μL of 50 $\mu\text{g}/\text{mL}$) and PI (Sigma-Aldrich; 20 $\mu\text{g}/\text{mL}$) were added, and the cells were incubated for 1 h prior to injection in the FACS machine (Becton Dickinson, San Jose, CA, USA). Density plots were generated using the CellQuestPro software (Becton Dickinson).

Western blot analysis

To determine the effect of 7HF on the expression of proteins involved in MT dynamics and DNA damage response (DDR) pathways, MDA-MB-468 cells (3×10^5 cells/well or 1×10^6 cells/plate) were grown at 37 °C in a 5% CO_2 incubator overnight. The cells were treated with 6 μM 7HF for 0, 12, 24, and 48 h. For optimizing 7HF concentration, the cells were incubated for 24 h in the presence of 0, 6, (or 12), and 24 μM 7HF. Chemiluminescence-based western blotting was performed as described previously (*Chimblee et al., 2022*). Briefly, 50 μg protein was separated electrophoretically 12% sodium dodecyl sulfate (SDS)-polyacrylamide gel electrophoresis (PAGE) and transferred onto nitrocellulose membrane (*Chimblee et al., 2022*). The blots were blocked and incubated overnight with 1;1,000 dilutions of various primary antibodies (anti-cyclin B1, anti-p-Cdk1 (Tyr15), anti-Bub3, and anti- β -actin; Cell Signaling Technology, Inc.) at 4 °C. After washing, the blots were incubated with horseradish peroxidase-conjugated anti-rabbit IgG (GE Healthcare Ltd; 1:5,000) for 1 h.

For fluorescence-based western blotting, the protein samples were extracted using radioimmunoprecipitation assay buffer (Tris pH 7.5, NaCl, EDTA, EGTA, NP-40, sodium

deoxycholate, sodium pyrophosphate, β -glycerophosphate, and protease inhibitor cocktail) and quantified using the Bradford assay (Bio-Rad Laboratories, Hercules, CA, USA). Then, 50 μ g protein was separated on SDS-PAGE gels (8%, 12%, and 15% variation with molecular weight of protein) and blotted onto nitrocellulose membranes at 100 V for 2 h. Apart from phosphoproteins for which BSA was used for blocking, the blots were blocked with blocking buffer (5% low-fat dry milk in Tris buffered saline) for 1 h. After washing, the blots were incubated with various primary antibodies for Rb, p-Rb (Ser780), Chk2, p-Chk2 (Ser516), and β -actin (Cell Signaling Technology, Inc.), Chk1, p-Chk1 (Ser345) (kindly provided by Prof. Dr. Carl Smythe), and p-H2AX (Ser139) (EMD Millipore Corp., Billerica, MA, USA) overnight 4 °C. The following antibody concentrations were used: 1:500 for anti-p-Chk2, 1:1,000 for anti-p-Rb, anti-Chk1, anti-p-Chk1, and anti-p-H2AX, 1:2,500 for anti-Rb and anti-Chk2, and 1:5,000 for anti- β actin. The blots were incubated for 1 h in 1:1,000 dilutions of secondary IgGs with various hosts and fluorophore labels, including anti-sheep (for 680 nm), anti-mouse (for 800 nm), and anti-rabbit (for 680 nm) antibodies (LI-COR Biosciences, Lincoln, NE, USA). Finally, the protein bands were observed using a chemiluminescent detection kit (Thermo Fisher Scientific, Inc.) and the Fusion FX western blot imaging system (Vilber Lourmat Sté, Collégien, France). The relative protein expression was quantified using the Fusion Capt Advanced Quantitation Analysis program (Vilber Lourmat Sté). Fluorescence-based protein bands were observed using the Odyssey imaging system (LI-COR Biosciences) and relative expression was calculated using the Image Studio Lite 5.2 program (LI-COR Biosciences). Two-three independent experiments were performed.

Prediction of 7HF drug-likeness

To predict the 7HF drug design, 7HF structure (PubChem CID 11983230) was submitted to the Swiss-absorption, distribution, metabolism and excretion-(ADME) web tool (<http://www.swissadme.ch>, accessed on 07 December 2021) (Daina, Michielin & Zoete, 2017).

Molecular docking analysis

Molecular docking was done to predict whether 7HF bound to the target protein, as per a previous study with certain modifications (Saechan *et al.*, 2021). The PDB ascension number 5SYF represents the three-dimensional (3D) protein structure of α -, β -tubulin (Kellogg *et al.*, 2017). The AutoDock Tool (ADT) 4.1 was used to eliminate all water molecules. The other co-crystallized ligands were retained. The PDB file format contains all polar hydrogen atoms that emulate hydrogen bond interactions. The 3D structures of 7HF (CID 11983230), pironetin (CID 6438891), PTX (CID 36314), and VCR (ID 5758) were obtained from the PubChem and ChemSpider databases. The SMILES Translator and Structure File Generator (<https://cactus.nci.nih.gov/translate/>) created PDB files for 7HF, pironetin, PTX, and VCR. All polar hydrogen atoms were added using ADT. Finally, the protein and ligand structures were saved as PDB and Partial Charge (Q) and Atom Type (T) or PDBQT files.

All ligands were docked with the tubulin proteins using AutoDock4 version 4.2 (Morris *et al.*, 2009). Using the SG atom of 315th-cysteine residue (Cys315) as the interaction

point with pironetin, the grid box was centered at an x - y - z position of “329.387 449.699 378.122” with a grid size of $90 \times 90 \times 90$ cubic angstrom (\AA^3) (Prota *et al.*, 2016; Steinmetz & Prota, 2018; Yang *et al.*, 2016). The grid box center of PTX and VCR was as per previous reports (Field Jessica, Díaz José & Miller John, 2013; Gigant *et al.*, 2005). In total, 50 docking runs with a population of 200 were required. The default parameters of the AutoDock4 program were used. The quintuple independent samplings reported the lowest binding energy (ΔG_{bind}). The drug-tubulin complex structure was visualized using the Visual Molecular Dynamics (VMD) package (Humphrey, Dalke & Schulten, 1996). The ligand-protein interactions were then studied and visualized using the Protein-Ligand Interaction Profiler (PLIP) webtool (Adasme *et al.*, 2021).

Statistical analysis

Each experiment was performed thrice. Data were analyzed using one-way analysis of variance (ANOVA) and two-way ANOVA using GraphPad Prism 5 (GraphPad Software, San Diego, CA, USA). The differences were considered significant at $p < 0.05$.

RESULTS

7HF uptake in MDA-MB-468 cells

In cells-free medium, 7HF was stable from 15 min to 17 h, and the stability gradually reduced up to 24 h (Fig. 1D and Table S2). In contrast, 7HF could not be detected between 5 h and 17 h in the cells culture medium (Fig. 1D and Table S2). 7HF may have penetrated MDA-MB-468 cells within 5 h and was internalized by 12 h.

Effect of 7HF on microtubule dynamics, tubulin polymerization, and protein expression

To determine the effect of 7HF on spindle MT arrangement, α -tubulin staining (shown in green) was visualized using IF (Fig. 2A). We observed that the spindle MT arrangement differed from that of the control when treated with 6–24 μM 7HF, excepting at 24 h. The radial array of the spindle gradually reduced after 1 h. The fibrous density along the spindle length was differed between control and 12 h 7HF. Radial lengthening of the spindle array was insignificantly observed from that of control at 24 h. We also observed time-dependent changes (1–24 h), *i.e.*, the spindle MT length ranged from short radial arrays to long fiber when treated with 6–24 μM 7HF. Notably, maximum reduction in the number of α -tubulin-presenting cells was observed at 12 h for all 7HF concentrations. Spindle length increased in PTX- and DOX-treated cells in both the fibrous dense bipolar poles and radial arrays (Fig. 2B). The VCR-treated cells showed decrease in spindle length in radial array arrangements. A reduction in α -tubulin-presenting cell aggregation was observed after treatment with all chemotherapeutics (PTX, VCR, and DOX). Additionally, structural changes in nuclear DNA (blue) were observed, with more expansion in 12 μM 7HF treated-cells and minimal shrinkage in PTX- and DOX treated-cells after 24 h (Figs. 2A–2B)

To determine the effect of 7HF on spindle MT formation, α -, β -tubulin polymerization was examined in a cell-free system for 1 h (Fig. 2C and Spreadsheet S1). All concentrations

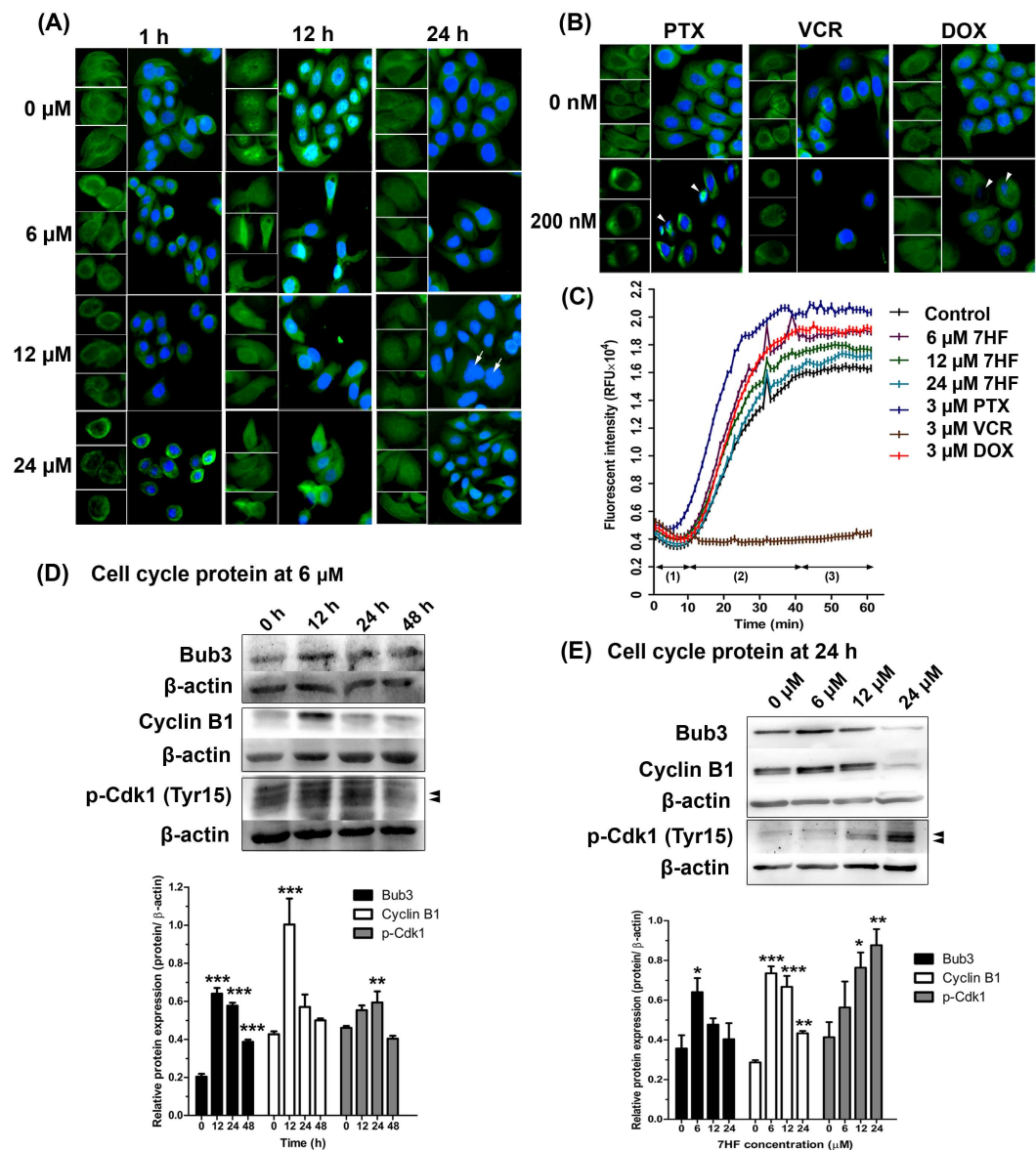


Figure 2 Effect of 7- α -hydroxyfrullanolide (7HF) on microtubule dynamics and protein expression in MDA-MB-468 cells. (A) Immunofluorescence (IF) of cells treated with 0, 6, 12, and 24 μM 7HF for 1, 12, and 24 h. (B) IF of cells treated with 0 and 200 nM PTX, VCR, and DOX for 24 h. PTX, paclitaxel; VCR, vincristine; DOX, doxorubicin; green, α -tubulin; blue, nuclear DNA; white arrow, likely to be M-phase cells; white head arrow, apoptotic cells; scale, 100 μm . (C) Results of tubulin polymerization assay performed using different 7HF concentrations (6 μM ; purple, 12 μM ; green, and 24 μM ; light blue), and 3 μM chemotherapeutics (PTX, dark blue; VCR, brown, and DOX, red) for 1 h in cell-free system. (1) nucleation, (2) growth, and (3) steady state equilibrium during tubulin formation. RFU: Relative fluorescence units. (D) Bub3, cyclin B1, and p-Cdk1 (Tyr15) levels in cells treated with 6 μM 7HF for 0, 12, 24, and 48 h. (E) Bub3, cyclin B1, and p-Cdk1 (Tyr15) levels in cells treated with 0, 6, 12, and 24 μM 7HF for 24 h. Mean differences were statistically analyzed using one-way ANOVA and Bonferroni's multiple comparison test. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ compared to control (0 h or 0 μM). Black arrowhead, an indicated protein band.

Full-size DOI: 10.7717/peerj.13508/fig-2

of 7HF (6, 12, and 24 μM) affected tubulin formation. For all 7HF concentrations, tubulin nucleation occurred for 10 min, followed by tubulin growth till 40 min and tubulin equilibrium. After 10 min of nucleation, all 7HF concentrations triggered faster tubulin polymerization than the control. However, tubulin polymerization and relative fluorescence intensity was reduced in a 7HF concentration-dependent manner. Tubulin polymerization was relatively increased by DOX, and PTX and reduced by VCR treatments. PTX stimulated tubulin nucleation within 5 min (fastest among the three drugs), followed by tubulin growth till 40 min, while maximum suppression of tubulin formation was observed in case of VCR.

The expression of proteins involved in MT dynamics at the G2/M phase arrest was examined in 7HF-treated cells both with respect to 7HF concentration and time of incubation (Figs. 2D–2E). For assessing concentration dependence, the cells were incubated with 0, 6, 12, and 24 μM 7HF for 24 h (Fig. 2E and Fig. S2B). Bub3 was significantly upregulated in 6 μM 7HF-treated cells compared to that in the control. Similarly, upregulation of cyclin B1 and p-Cdk1 (Tyr15) was observed in concentration-independent and -dependent manners, respectively. For assessing time dependence, the cells were treated with 6 μM 7HF for 0, 12, 24, and 48 h (Fig. 2D and Fig. S2A). Bub3 was significantly upregulated between 12–24 h. Cyclin B1 was significantly upregulated at 12 h. In addition, p-Cdk1 (Tyr15) was significantly upregulated between 12 and 24 h and gradually downregulated at 48 h. All comparisons were made with the control group.

Effect of 7HF on cell proliferation and expression of DDR proteins

The effect of MT dynamics on cell proliferation and DNA synthesis in 7HF-treated cells was determined using BrdU and PI staining, and FACS analysis. The cells were treated with various concentrations of 7HF (6, 12, and 24 μM) for 12 and 24 h (Figs. 3A–3C and Fig. S3); 7HF-concentration dependence analysis at 12 h (Figs. 3A–3B) revealed that the number of G1 phase cells was significantly higher at 6 μM (45.61%) than that of the control (30.23%). The number of S phase cells increased gradually in a concentration-dependent manner from 6 to 24 μM (29.32%–34.89%), but was lower than that of the control (44.76%). The number of cells in the G2 phase was significantly higher than that in the control (20.07%) and increased in a concentration-dependent manner at 6 μM (26.68%), 12 μM (31.24%), and 24 μM (32.14%). Concentration dependence analysis at 24 h (Figs. 3A and 3C) revealed that the number of cells in the G1 phase was significantly higher than that in the control (33.86%) and increased in a concentration-dependent manner from 6 to 24 μM (51.56–54.87%). The number of S phase cells was clearly lower than that in the control (49.19%) when treated with 6–24 μM 7HF (34.64–24.55%), although the number of cells treated with 24 μM 7HF (22.64%) was higher than that in the control (15.11%). The proportion of cells in the G2 phase appeared independent of time of treatment (12–24 h). In contrast, the proportion of cells in the G1 phase occurred in a time-dependent manner (Figs. 3B–3C). Hence, we concluded that 7HF triggers G2/M and G1 phase arrests in concentration- and time-dependent manner.

7HF treatment significantly increased the number of G1 phase cells at 24 h (Fig. 3C). DDR proteins such as Rb, p-Rb (Ser780), Chk1, p-Chk1 (Ser345), Chk2, p-Chk2 (Ser516),

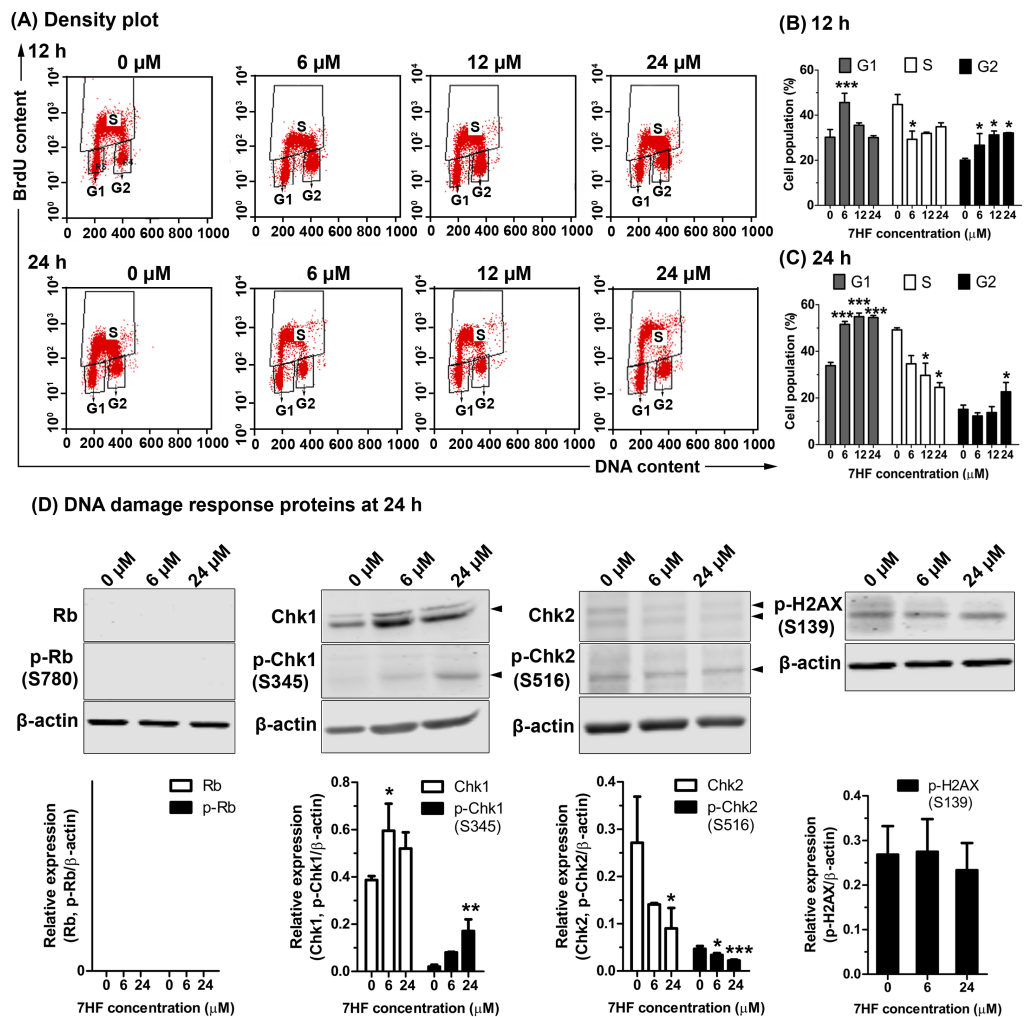


Figure 3 Effect of 7- α -hydroxyfrullanolide (7HF) on cell proliferation and expression of DNA damage response (DDR) proteins in MDA-MB-468 cells. (A) Density plots of cells treated with 0, 6, 12, and 24 μ M 7HF for 12 h and 24 h. Left-lower, middle-top, and right-lower quadrants represent the number of cells in G1, S, and G2 phases, respectively. (B–C) The histograms represent the percentages of cells in G1, S, and G2 phases after 12 and 24 h treatment. (D) Expression of DDR proteins, including Rb, p-Rb (Ser780), Chk1, p-Chk1 (Ser345), Chk2, p-Chk2 (Ser516), and p-H2AX (Ser139), in cells treated with 6 and 24 μ M 7HF for 24 h. S, serine; black arrowhead, an indicated protein band. Two or three independent experiments were performed. Mean difference was statistically analyzed using one-way ANOVA and Bonferroni's (Figs. 3B–3C) and Dunnett's (Fig. 3D) multiple comparison test. * $p < 0.05$, *** $p < 0.001$, compared to 0 μ M (control).

Full-size DOI: 10.7717/peerj.13508/fig-3

and p-H2AX (Ser139) were subsequently measured in the cells treated with 6 and 24 μ M 7HF for 24 h, using western blotting (Fig. 3D and Fig. S4). However, the relative levels of Rb and p-Rb were not detected. Compared to that in the control, Chk1 and p-Chk1 were significantly upregulated, while Chk2 and p-Chk2 were significantly downregulated in a 7HF-concentration-dependent manner in the treated cells. p-H2AX expression was insignificantly differed in 7HF-treated group of 6 and 24 μ M.

Table 1 Interaction of drugs with tubulins and their relative binding energies (ΔG_{bind}).

Compounds	ΔG_{bind} (kcal/mol)	H-bonds	Salt bridges	π - π stacking
α-tubulin				
-7HF	-6.77	Ile341, Phe343	-	-
-Pironetin	-5.45	Thr257, Val260, Cys315	-	-
β-tubulin				
-7HF	-7.12	Asp211, Ala298, Met301	Arg215	-
-PTX	-8.05	Asp226, Thr276, Gly279	His229, Arg278	His229
-VCR	-7.09	Glu71, Gly100	Glu71	-

Notes.

7HF, 7- α -Hydroxyfrullanolide; PTX, paclitaxel; VCR, vincristine.

Drug-likeness properties of 7HF

The Swiss ADME analysis of 7HF revealed its physicochemical properties, pharmacokinetics, drug-likeness, and medicinal properties. 7HF showed predicted drug-likeness, as it followed the Lipinski's rule of Five ([Lipinski et al., 2001](#); [Yang & Hinner, 2015](#)): MW 248.32, 3 H-bond acceptors, 1 H-bond donor, Clog P 2.46, and high gastrointestinal (GI) absorption.

Predicted interaction of 7HF with α - and β -tubulins

Molecular docking was used to predict whether 7HF targeted α - or β -tubulins of MTs. 7HF and pironetin bind to α -tubulin with relative binding energies (ΔG_{bind}) of -6.77 and -5.45 kcal/mol, respectively ([Table 1](#)). However, 7HF, PTX, and VCR bind to β -tubulin with ΔG_{bind} values of -7.12 , -8.05 , and -7.09 kcal/mol, respectively ([Table 1](#)). 7HF bound to both α - and β -tubulin sites, which was distinct from that observed with pironetin (a known α -tubulin inhibitor) and PTX and VCR (also known β -tubulin inhibitors), as shown in [Fig. 4A](#). [Table 1](#) and [Figs. 4B–4C](#) show 7HF-inhibitor interactions in terms of amino acid residues of various tubulin proteins. The initial binding site was at α -tubulin ([Fig. 4B](#)). A hydroxyl group (-OH) at C-7 in 7HF interacted with α -tubulin *via* hydrogen bonds (H-bond) with Ile341 and Phe343. An additional hydrophobic interaction with Lys336 and Phe343 further stabilized the 7HF binding. Compared to 7HF, pironetin formed H-bonds with Cys315, and backbone atoms of Thr257 and Val260. The neighboring amino acids such as Trp346, Ala314, and Lys352 also supported the binding *via* hydrophobic interactions.

7HF also interacted with β -tubulin ([Table 1](#) and [Fig. 4C](#)). A lactone ring established a salt bridge with Arg215. The C-12 ketone (C=O) formed H-bonds with Asp211. A hydroxyl (-OH) group at C-7 formed H-bonds with Ala298 and Met301. Some amino acids, such as Ala208, Ala298, Asp211, Asp306, and Ile212, exhibited hydrophobic interactions with 7HF. His229 and Arg278 carboxylic acids formed salt bridges to PTX. The PTX-protein complex displayed π - π stacking with His229. Furthermore, hydrophobic interactions with Leu217, Leu219, Leu230, Leu275, Leu 371, Asp226, Ala233, Phe272, and Arg278 contributed to PTX- β -tubulin binding. VCR-tubulin formed a salt bridge at Gln71 tetramine, and its

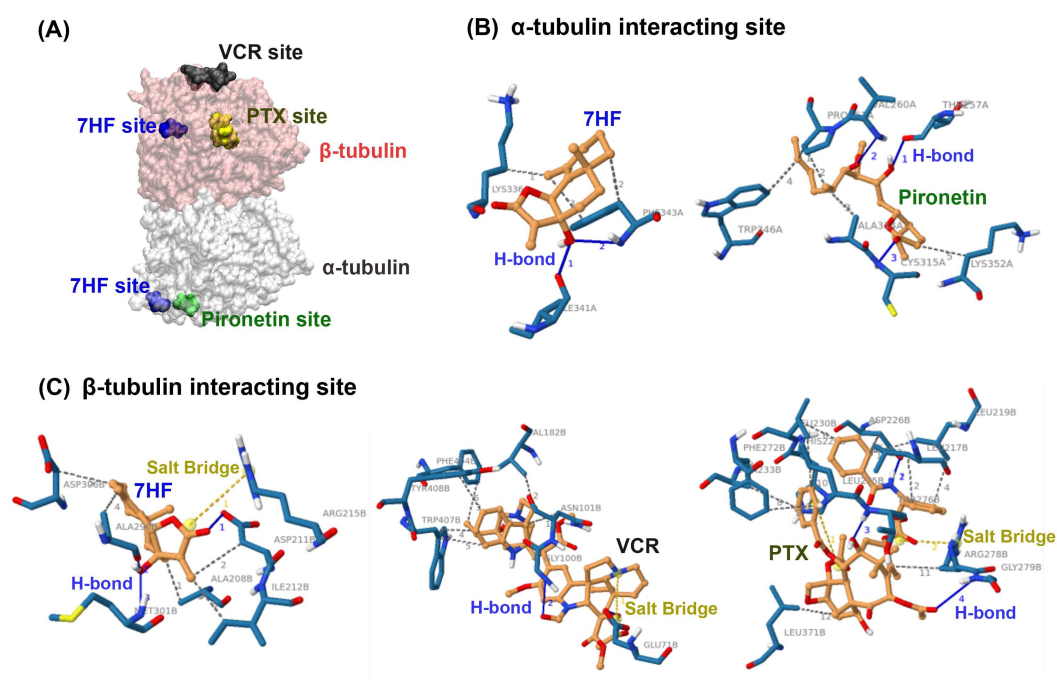


Figure 4 Predictive binding of 7- α -hydroxyfrullanolide (7HF) and the inhibitors with α - and β -tubulins. (A) Predictive binding locations of 7HF (blue), VCR (black), PTX (yellow), and pironetin (green) in the α/β -tubulin complex. The α - and β -tubulins are shown as grey and pink surface structures, respectively. (B) Interacting sites of ligands (orange): 7HF and pironetin with amino acids (navy blue) in α -tubulin structure are shown. (C) Interacting sites of ligands (orange): 7HF, PTX, and VCR with the amino acids (navy blue) in β -tubulin structure are shown. The blue, dotted grey, and dotted yellow lines depict H-bonds, hydrophobic interactions, and salt bridges, respectively. PTX, paclitaxel; VCR, vincristine.

Full-size DOI: 10.7717/peerj.13508/fig-4

H-bonds were formed by VCR. Amino acids such Asn101, Val182, Phe404, Trp407, and Tyr408 assisted in VCR binding *via* hydrophobic interactions.

DISCUSSION

A calibration curve of 7HF at 268 nm was correlated with increasing of concentration as reported previously (Rahman *et al.*, 2016). The spectrophotometric results at 268 nm were used to examine chemical uptake in cancer cells. 7HF could pass through MDA-MB-468 cells as was apparent from the absence of 7HF in the medium at 268 nm. The passive penetration of the cancer cell membrane was also supported by Swiss ADME prediction based on Lipinski's drug likeness (Daina, Michielin & Zoete, 2017; Lipinski *et al.*, 2001). Swiss ADME predicted that 7HF preferably follows passive diffusion and possessed the high GI absorption property. These predicted features of 7HF are critical for designing and developing 7HF-based drugs.

TNBC cell lines can be used for developing better treatment options for patients with TNBC (Chavez, Garimella & Lipkowitz, 2010). The MDA-MB-468 cell line is a basal-like 1 subtype of TNBC that exhibits highly aggressive adenocarcinoma and harbors aberrations in numerous genes involved in cell division and DDR (Lehmann & Pietenpol, 2014; Yin *et*

et al., 2020). Hence, MDA-MB-468 cells may be a representative TNBC model that can be used to determine the responses of MTs and DDR pathways to 7HF treatment.

MT instability is critically involved in stress response of cancer cells (*Parker, Kavallaris & McCarroll, 2014*). Most cellular MTs exhibit cycles of growth, shortening, and regrowth (*Gudimchuk & McIntosh, 2021*). MT instability in MDA-MB-468 cells varied in 7HF concentration- and time-dependent manner (*Fig. 2A*). The density of the MTs in the spindle is high at the bipolar pole, and tubulin expression is the highest at the G2/M-phase transition (*Sato & Toda, 2004; Syred et al., 2013*). We observed that 7HF was internalized by cancer cells by 12 h, following which it disrupted MT dynamics, leading to G2/M arrest. 7HF induced G2/M phase arrest by upregulating Bub3, cyclin B1, and p-Cdk1 (Tyr15) (*Chimblee et al., 2022*), particularly at 6 μ M concentration and after incubation of 12–24 h (*Figs. 2D–2E*). Bub3 and BubR1 are ubiquitously expressed throughout the cell cycle; however, Bub3 and BubR1 production was high in the G2/M phase (*Liu & Zhang, 2016; Sudakin, Chan & Yen, 2001*). Bub3-BuR1 and Cdc20-Mad2 are evolutionarily conserved mitotic checkpoint proteins that form a mitotic checkpoint complex (MCC) only at mitosis (*Han et al., 2014; Liu & Zhang, 2016*). The active MCC is an effector of the anaphase-promoting complex/cyclosome (APC/C, an E3 ubiquitin ligase). APC/C bound to the MCC prevents cyclin B1 and securin degradation during spindle assembly checkpoint and the detachment of sister chromatids from the kinetochore (*Liu & Zhang, 2016; Matthews, Bertoli & De Bruin, 2021*). In general, cyclin B1 expression is temporally restricted to the G2 phase and early mitosis transition in human cells (*Lindqvist, Rodriguez-Bravo & Medema, 2009*). Mad2 is also found in 7HF-treated cells (*Chimblee et al., 2022*). Thus, 7HF might increase Bub3-Mad2 (MCC) formation and sustain cyclin B1 levels to check MT spindle formation during G2/M arrest. Cdk1 is inhibited when phosphorylated by Wee1 at Tyr15 (*Schmidt et al., 2017*). Phosphorylated Cdk1-cyclin B1 complex is inactive and blocks entry into mitosis (*Schmidt et al., 2017*). Indeed, high levels of p-Cdk1 (Tyr15) have been shown to be associated with G2/M phase arrest following DNA damage in different cancer cell types (*Ghelli Luserna di Rorà et al., 2020*). 7HF might result in accumulation of cyclin B1-p-Cdk1. In fact, cells in the G2 phase generally correct errors generated during DNA replication and prepare proteins before rapid growth and mitosis (*Jackman, 2011; Kousholt, Menzel & Sorensen, 2012*).

MT dynamics is a crucial factor regulating DNA damage repair during cell division; perturbations in MT dynamics lead to genomic instability (*Lottersberger et al., 2015; Petsalaki & Zachos, 2020*). Arrest in the G2/G1 phases after 12 h of 7HF treatment could be due to MT instability and persistence of DNA aberrations (*Figs. 2A and 3A–3B*). Variations in the anti-mitotic activity and disruption of MT dynamics in numerous cancer cell types have been observed due to differences in the chemical carbocyclic structure of SLs, all of which trigger G2/M arrest *via* the upregulation of p-Cdk1 (Tyr 15) and cyclin B1 (*Bosco & Golsteyn, 2017*). Santamarine (*Ma et al., 2009*), parthenolide (*Whipple et al., 2013*), and costunolide (*Bocca et al., 2004; Liu et al., 2011a; Whipple et al., 2013*) are eudesmanolides (6/6 bicyclic SLs) that are similar to 7HF. The mode of action of eudesmane SLs involves perturbation of MT stability at the levels of both tubulin polymerization and depolymerization, and upregulation of p-Cdk1 (Tyr15) and cyclin B1

in leukemia (CCRF-CEM murine), breast cancer (MCF-7), and hepatoma (HA22T/VGH) cells. 6-O-angeloylplenolin, coronopilin, pulchelloid A, and hymenoratin, 5/7-bicyclic pseudoguaianolide SLs also induce high p-Cdk1 and/or Cdk1 and cyclin B1 expression in certain cancer cell types. (Bosco et al., 2021; Cotugno et al., 2012; Liu et al., 2011b; Molina et al., 2021). However, the role of Bub3 and its partners in the modulation of MCC after SL treatment is not clear (Steinbruck, Pereira & Efferth, 2010).

The upregulation of p-Cdk1 (Tyr15) appears to maintain cyclin B1 accumulation with G2/M arrest, followed by DNA damage, after 24 h of treatment with 7HF. The G1 phase arrest was characterized by low MT dynamics, with the lengthening of the radial array in 7HF-treated cells at 24 h (Figs. 2A, 3A–3C) (Sato & Toda, 2004; Syred et al., 2013). MTs recruit trafficking proteins to repair aberrant DNA, leading to G1 phase arrest (Lotterberger et al., 2015). Previous studies have shown that 7HF inhibits DNA Top I activity; 7HF may affect pre-replicative stress or single-stranded DNA break in the G1 phase arrest by upregulating Chk1 *via* Ataxia-Telangiectasia and Rad3 related (ATR) kinase signaling (Champoux, 2001; Ju et al., 2020; Petsalaki & Zachos, 2020; Uppatanprecha, 2009). Chk1 is a key downstream regulator of ATR response and is phosphorylated by ATR at Ser345 (Ju et al., 2020; Petsalaki & Zachos, 2020). Phosphorylation of Chk1 at Ser345 is essential for its biological activity (Petsalaki & Zachos, 2020). In 7HF-treated cells, Chk1, p-Chk1 (Ser345), p-Cdk1 (Tyr15), and cyclin B1 levels increased after 24 h (Figs. 2D–2E and 3D). Chk1 plays key roles in both late G1-S phase (to inhibit DNA replication initiation) and G2-M-phase checkpoints (to check MT spindle assembly) (Ju et al., 2020; Petsalaki & Zachos, 2020). Active Chk1 (p-Chk1 Ser345) dissociates from chromatin and phosphorylates and inactivates the Cdc25 family phosphatases (Petsalaki & Zachos, 2020). Chk1 stimulates Wee1 kinase to phosphorylate Cdk1 at Tyr15 (Cdk1-cyclin B1 inhibition). High Chk1-p-Chk1-Wee1-p-Cdc25-p-Cdk1-cyclin B1 activity prevents the onset of mitosis by activating the spindle assembly checkpoint, which persists till DNA damage is not repaired (Petsalaki & Zachos, 2020). A recent study has reported that DNA damage activates MT dynamics and is associated with the spindle checkpoint in the G1 phase arrest (Ma et al., 2021). SLs induce G2/M-G1 phase arrest *via* the proposed Chk1, p-Chk1-Cdc25s-p-Cdk1 (Tyr15)-cyclin B1 pathway. Costunolide upregulates Chk2 (Thr68), p-Cdc25c (Ser216), p-Cdk1 (Tyr15), and cyclin B1 in liver cancer (HA22T/VGH) cells (Liu et al., 2011a). Galiellalactone increases ATM/ATR, p-Chk1 (Ser345), and cyclin B1 protein levels in prostate cancer cells (Garcia et al., 2016). Coronopilin increases cyclin B1 and p-Cdk1 (Tyr15) levels; however, the upregulation of γ H2AX (p-H2AX Ser139) is involved in G1 phase arrest in leukemia (Jurkat and U937) cells (Cotugno et al., 2012).

In the present study, 7HF triggered G1 phase arrest independent of DDR *via* Rb, p-Rb, Chk2, p-Chk2, and γ H2AX signaling. Not surprisingly, Rb and p-Rb (Ser780) expression could not be detected in the 7HF-treated cells, as MDA-MB-468 cells are Rb-null lines (Robinson et al., 2013). However, the deficiency of Rb in breast cancer cells renders them highly sensitive to chemotherapy (DOX and methotrexate) and radiotherapy (Robinson et al., 2013). Reduction in the levels of Chk2, p-Chk2 (Ser516), and phosphorylated H2AX (p-H2AX, γ H2AX) was observed in the 7HF-treated cells at 24 h. 7HF might not induce

DDR via dsDNA breaks in the G1/S/intra S phase checkpoints (*Sharma, Singh & Almasan, 2012; Zannini, Delia & Buscemi, 2014*).

The fibrous nature of the spindle increased in 7HF-treated cells was similar to that observed in 7HF-untreated cells and DOX-treated cells at 24 h (*Figs. 2A–2B*). PTX- and VCR-treated cells exhibited spindle lengthening at the bipolar pole and shrinkage of the radial spindle array, respectively (*Fig. 2B*). 7HF lowered spindle MT stability and α -, β -tubulin polymerization more than PTX (MT stabilizing agent) and VCR (MT destabilizing agent). 7HF bound to a pocket in β -tubulin that differed from the PTX and VCR sites (*Fig. 4A*). Two PTX-tubulin interaction sites have been previously reported (*Field Jessica, Díaz José & Miller John, 2013*). The initial site was located in luminal β -tubulin at position of S7 end (Phe 272), H1 (Asp226), H6-H7 (Leu217, Leu219, His 229, Leu 230, Ala 233), and M-loop (Leu275, Arg278). One of these two is the external pore type I site. An increase in lateral protofilament connections due to PTX and M-loop interaction in the luminal region is critical for tubulin stability. Additional amino acids (Phe214, Thr220, Thr221, and Pro222) occupy the pore type I site (*Field Jessica, Díaz José & Miller John, 2013*). This cavity is formed by the Lys216, Thr218, and Gly271 residues of the β 1 subunit and the Arg77, Pro87, Asp88, and Phe90 residues of the β 2 subunit (*Barasoain et al., 2010*).

The vinca alkaloid interacts with β -tubulin at 177-215 peptides (*Mitra & Sept, 2004; Rai & Wolff, 1996*). Vinorelbine, a vinblastine analogue, also interacts with Tyr224's side chain (*Lobert et al., 2000*). The other side chain of Asp179 moves from nucleotide to drug (*Gigant et al., 2005*). *Table 1* depicts a VCR tubulin interaction site. VCR structure prevented protein binding. Small ligand diffusion into tubulin was proposed here (*Field Jessica, Díaz José & Miller John, 2013*). Unlike PTX/VCR-tubulin interaction, the 7HF-tubulin complex formed hydrophobic and salt bridge contacts with Ile212 and Arg215, close to the external pore type I of MTs (*Field Jessica, Díaz José & Miller John, 2013*). It was found that 7HF gave ΔG_{bind} of -6.77 and -7.12 kcal/mol toward the exterior α - and β -tubulin sites (*Field Jessica, Díaz José & Miller John, 2013*). These numbers were not enough to clearly justify the preference. In addition, 7HF did not interact with any amino acid in luminal areas. Furthermore, 7HF formed H-bonds and hydrophobic interactions with Met301 (near Arg308 in LAU/PEL site) and Ala298, similar to natural MT stabilizing agents laulimalide (LAU) and peloruside A (PEL A) (*Field Jessica, Díaz José & Miller John, 2013*). Chemical bonds were formed between the 7HF lactone ring, ketone, and hydroxyl moieties and amino acid residues (*Table 1* and *Figs. 4B–4C*). These moieties may contribute to a preference for 7HF-tubulin interactions (*Ghantous et al., 2010*).

As ligand- α -tubulin targeting is unknown, we predicted the molecular interaction in the 7HF- α -tubulin complex (*Steinmetz & Prota, 2018*). The 7HF- α -tubulin site differed from that of pironetin. A study showed that Cys316 and Lys352 of α -tubulin covalently bind to pironetin via Michael addition reactions (*Prota et al., 2016; Usui et al., 2004; Yang et al., 2016*). The binding was perfect and perturbed a major loop and helix of α -tubulin, inhibiting MT formation (*Prota et al., 2016*). Furthermore, the covalent bond promoted adducted pironetin formation and shifted the MT equilibrium (*Bañuelos Hernández et al., 2014; Coulup & Georg, 2019; Steinmetz & Prota, 2018*). Unlike the pironetin- α -tubulin interacting site, 7HF (-OH, C-7) used H-bonds at Ile341 and Phe343, as well as hydrophobic

interactions at Lys336 and Phe343, in addition to the interactions with targeted Cys316 and Lys352. As previously stated, the results were insufficient to determine the specificity of 7HF with either α - or β -tubulin because 7HF was predicted to interact with both tubulin sites with close ΔG_{bind} , falling within a range of 1 kcal/mol. Furthermore, the small size of 7HF may be the key for the molecular to enter the tubulin binding pocket. Finally, the specificity of 7HF remained a topic for further investigation.

CONCLUSIONS

To the best of our best knowledge, this is the first report to investigate MT dynamics and its interaction with 7HF in TNBC cells, with the aim of deciphering the mechanism of action of 7HF in TNBC cells. Our findings revealed that 7HF entered the MDA-MB-468 cells, where it disrupted MT dynamics, along with DDR, *via* the induction of G2/M-G1 phase arrest, and upregulation of Chk1, p-Chk1 (Ser345), Bub3, p-Cdk1 (Tyr15), and cyclin B1. *In silico* analysis, 7HF preferably targets β -tubulin over α -tubulin, with the formation of H-bonds and salt bridges between 7HF moieties (lactone, ketone, and hydroxyl groups) and tubulin, which support the formation of the 7HF-MT complex. 7HF uptake prediction follows high passive diffusion and GI absorption routes. Hence, the results of this study will provide information for drug design, development, and therapy for TNBC patients.

ACKNOWLEDGEMENTS

The authors thank the Smythe Lab, School of Biosciences, University of Sheffield, for providing the opportunity to perform the experiments.

ADDITIONAL INFORMATION AND DECLARATIONS

Funding

This work was financed by the Royal Golden Jubilee Ph.D. Program (RGJ-Ph.D. Program), the Thailand Research Fund (TRF)/Thailand Science Research and Innovation (TSRI), and the National Research Council of Thailand (NRCT) (grant number PHD/0091/2558). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Grant Disclosures

The following grant information was disclosed by the authors:

Royal Golden Jubilee Ph.D. Program (RGJ-Ph.D. Program).

Thailand Research Fund (TRF)/Thailand Science Research and Innovation (TSRI).

National Research Council of Thailand (NRCT): PHD/0091/2558.

Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Siriphorn Chimplee conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Carl Smythe conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Varomyalin Tipmanee conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Suchada Sukrong performed the experiments, authored or reviewed drafts of the article, contributed 7HF compound and reagents, and approved the final draft.
- Kanyanatt Kanokwiroon conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.

Data Availability

The following information was supplied regarding data availability:

The raw data are available in the [Supplementary Files](#).

Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.13508#supplemental-information>.

REFERENCES

- Adasme MF, Linnemann KL, Bolz SN, Kaiser F, Salentin S, Haupt VJ, Schroeder M. 2021. PLIP 2021: expanding the scope of the protein–ligand interaction profiler to DNA and RNA. *Nucleic Acids Research* 49:W530–W534 DOI 10.1093/nar/gkab294.
- Atta ur R, Choudhary MI, Ata A, Alam M, Farooq A, Perveen S, Shekhani MS, Ahmed N. 1994. Microbial transformations of 7 α -hydroxyfrullanolide. *The Journal of Natural Products* 57:1251–1255 DOI 10.1021/np50111a010.
- Avila J. 1992. Microtubule functions. *Life Science* 50:327–334 DOI 10.1016/0024-3205(92)90433-p.
- Bañuelos Hernández AE, Mendoza-Espinoza JA, Pereda-Miranda R, Cerda-García-Rojas CM. 2014. Studies of (–)-pironetin binding to α -tubulin: conformation, docking, and molecular dynamics. *The Journal of Organic Chemistry* 79:3752–3764 DOI 10.1021/jo500420j.
- Barasoain I, Garcia-Carril AM, Matesanz R, Maccari G, Trigili C, Mori M, Shi JZ, Fang WS, Andreu JM, Botta M, Diaz JF. 2010. Probing the pore drug binding site of microtubules with fluorescent taxanes: evidence of two binding poses. *Chemistry & Biology* 17:243–253 DOI 10.1016/j.chembiol.2010.02.006.

- Bianchini G, De Angelis C, Licata L, Gianni L. 2021.** Treatment landscape of triple-negative breast cancer—expanded options, evolving needs. *Nature Reviews Clinical Oncology* **19**:91–113 DOI [10.1038/s41571-021-00565-2](https://doi.org/10.1038/s41571-021-00565-2).
- Blajeski AL, Phan VA, Kottke TJ, Kaufmann SH. 2002.** G(1) and G(2) cell-cycle arrest following microtubule depolymerization in human breast cancer cells. *Journal Clinical Investigation* **110**:91–99 DOI [10.1172/JCI13275](https://doi.org/10.1172/JCI13275).
- Bocca C, Gabriel L, Bozzo F, Miglietta A. 2004.** A sesquiterpene lactone, costunolide, interacts with microtubule protein and inhibits the growth of MCF-7 cells. *Chemico-biological Interactions* **147**:79–86 DOI [10.1016/j.cbi.2003.10.008](https://doi.org/10.1016/j.cbi.2003.10.008).
- Bosco A, Golsteyn RM. 2017.** Emerging anti-mitotic activities and other bioactivities of sesquiterpene compounds upon human cells. *Molecules* **22**:459 DOI [10.3390/molecules22030459](https://doi.org/10.3390/molecules22030459).
- Bosco A, Molina L, Kerneis SM, Hatzopoulos G, Favez T, Gonczy P, Tantapakul C, Maneerat W, Yeremy B, Williams DE, Andersen RJ, Golsteyn RM. 2021.** Pulchelloid A, a sesquiterpene lactone from the Canadian prairie plant *Gaillardia aristata* inhibits mitosis in human cells. *Molecular Biology Reports* **48**:5459–5471 DOI [10.1007/s11033-021-06554-z](https://doi.org/10.1007/s11033-021-06554-z).
- Caldas LA, Horvath RO, Ferreira-Silva GA, Ferreira MJP, Ionta M, Sartorelli P. 2018.** Calein C, a sesquiterpene lactone isolated from *Calea pinnatifida* (Asteraceae), inhibits mitotic progression and induces apoptosis in MCF-7 cells. *Frontiers in Pharmacology* **9**:1191 DOI [10.3389/fphar.2018.01191](https://doi.org/10.3389/fphar.2018.01191).
- Cecchini MJ, Amiri M, Dick FA. 2012.** Analysis of cell cycle position in mammalian cells. *The Journal of Visualized Experiments* **59**:3941 DOI [10.3791/3491](https://doi.org/10.3791/3491).
- Champoux JJ. 2001.** DNA topoisomerases: structure, function, and mechanism. *Annual Review of Biochemistry* **70**:369–413 DOI [10.1146/annurev.biochem.70.1.369](https://doi.org/10.1146/annurev.biochem.70.1.369).
- Chavez KJ, Garimella SV, Lipkowitz S. 2010.** Triple negative breast cancer cell lines: one tool in the search for better treatment of triple negative breast cancer. *Breast Disease* **32**:35–48 DOI [10.3233/BD-2010-0307](https://doi.org/10.3233/BD-2010-0307).
- Chimlee S, Graidist P, Srisawat T, Sukrong S, Bissanum R, Kanokwiroon K. 2019.** Anti-breast cancer potential of frullanolide from *Grangea maderaspatana* plant by inducing apoptosis. *Oncology Letters* **17**:5283–5291 DOI [10.3892/ol.2019.10209](https://doi.org/10.3892/ol.2019.10209).
- Chimlee S, Roytrakul S, Sukrong S, Srisawat T, Graidist P, Kanokwiroon K. 2022.** Anticancer effects and molecular action of 7- α -hydroxyfrullanolide in G2/M-phase arrest and apoptosis in triple negative breast cancer cells. *Molecules* **27**:407 DOI [10.3390/molecules27020407](https://doi.org/10.3390/molecules27020407).
- Cotugno R, Fortunato R, Santoro A, Gallotta D, Braca A, De Tommasi N, Belisario MA. 2012.** Effect of sesquiterpene lactone coronopilin on leukaemia cell population growth, cell type-specific induction of apoptosis and mitotic catastrophe. *Cell Proliferation* **45**:53–65 DOI [10.1111/j.1365-2184.2011.00796.x](https://doi.org/10.1111/j.1365-2184.2011.00796.x).
- Coulup SK, Georg GI. 2019.** Revisiting microtubule targeting agents: alpha-tubulin and the pironetin binding site as unexplored targets for cancer therapeutics. *Bioorganic & Medicinal Chemistry Letters* **29**:1865–1873 DOI [10.1016/j.bmcl.2019.05.042](https://doi.org/10.1016/j.bmcl.2019.05.042).

- Daina A, Michielin O, Zoete V. 2017.** SwissADME: a free web tool to evaluate pharmacokinetics, drug-likeness and medicinal chemistry friendliness of small molecules. *Scientific Reports* 7:42717 DOI [10.1038/srep42717](https://doi.org/10.1038/srep42717).
- De Ruijter TC, Veeck J, De Hoon JP, Van Engeland M, Tjan-Heijnen VC. 2011.** Characteristics of triple-negative breast cancer. *The Journal of Cancer Research and Clinical Oncology* 137:183–192 DOI [10.1007/s00432-010-0957-x](https://doi.org/10.1007/s00432-010-0957-x).
- Duangmano S, Sae-Lim P, Suksamrarn A, Domann FE, Patmasiriwat P. 2012.** Cucurbitacin B inhibits human breast cancer cell proliferation through disruption of microtubule polymerization and nucleophosmin/B23 translocation. *BMC Complementary and Alternative Medicine* 12:185 DOI [10.1186/1472-6882-12-185](https://doi.org/10.1186/1472-6882-12-185).
- Dumontet C, Jordan MA. 2010.** Microtubule-binding agents: a dynamic field of cancer therapeutics. *Nature Reviews Drug Discovery* 9:790–803 DOI [10.1038/nrd3253](https://doi.org/10.1038/nrd3253).
- Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Pineros M, Znaor A, Bray F. 2021.** Cancer statistics for the year 2020: an overview. *International Journal of Cancer* 149:778–789 DOI [10.1002/ijc.33588](https://doi.org/10.1002/ijc.33588).
- Field Jessica J, Díaz José F, Miller John H. 2013.** The binding sites of microtubule-stabilizing agents. *Chemistry & Biology* 20:301–315 DOI [10.1016/j.chembiol.2013.01.014](https://doi.org/10.1016/j.chembiol.2013.01.014).
- Fonseca LC, Dadarkar SS, Lobo AS, Suthar AC, Chauhan VS, Chandrababu S, Sharma SD, Dagia NM, Padigar M. 2010.** 7-hydroxyfrullanolide, a sesquiterpene lactone, inhibits pro-inflammatory cytokine production from immune cells and is orally efficacious in animal models of inflammation. *European Journal of Pharmacology* 644:220–229 DOI [10.1016/j.ejphar.2010.06.052](https://doi.org/10.1016/j.ejphar.2010.06.052).
- Gallagher J, Brian M. 2007.** Microtubule-stabilizing natural products as promising cancer therapeutics. *Current Medicinal Chemistry* 14:2959–2967 DOI [10.2174/092986707782794014](https://doi.org/10.2174/092986707782794014).
- Garcia V, Lara-Chica M, Cantarero I, Sterner O, Calzado MA, Munoz E. 2016.** Galialactone induces cell cycle arrest and apoptosis through the ATM/ATR pathway in prostate cancer cells. *Oncotarget* 7:4490–4506 DOI [10.18632/oncotarget.6606](https://doi.org/10.18632/oncotarget.6606).
- Gascoigne KE, Taylor SS. 2009.** How do anti-mitotic drugs kill cancer cells? *The Journal of Cell Science* 122:2579–2585 DOI [10.1242/jcs.039719](https://doi.org/10.1242/jcs.039719).
- Ghantous A, Gali-Muhtasib H, Vuorela H, Saliba NA, Darwiche N. 2010.** What made sesquiterpene lactones reach cancer clinical trials? *Drug Discovery Today* 15:668–678 DOI [10.1016/j.drudis.2010.06.002](https://doi.org/10.1016/j.drudis.2010.06.002).
- Ghelli Luserna di Rorà A, Cerchione C, Martinelli G, Simonetti G. 2020.** A WEE1 family business: regulation of mitosis, cancer progression, and therapeutic target. *The Journal of Hematology & Oncology* 13:126 DOI [10.1186/s13045-020-00959-2](https://doi.org/10.1186/s13045-020-00959-2).
- Gigant B, Wang C, Ravelli RBG, Roussi F, Steinmetz MO, Curmi PA, Sobel A, Knossow M. 2005.** Structural basis for the regulation of tubulin by vinblastine. *Nature* 435:519–522 DOI [10.1038/nature03566](https://doi.org/10.1038/nature03566).
- Gudimchuk NB, McIntosh JR. 2021.** Regulation of microtubule dynamics, mechanics and function through the growing tip. *Nature Reviews Molecular Cell Biology* 22:777–795 DOI [10.1038/s41580-021-00399-x](https://doi.org/10.1038/s41580-021-00399-x).

- Han JS, Vitre B, Fachinetti D, Cleveland DW. 2014.** Bimodal activation of BubR1 by Bub3 sustains mitotic checkpoint signaling. *Proceedings of the National Academy of Sciences of the United States of America* **111**:E4185–E4193 DOI [10.1073/pnas.1416277111](https://doi.org/10.1073/pnas.1416277111).
- Humphrey W, Dalke A, Schulten K. 1996.** VMD: Visual molecular dynamics. *The Journal of Molecular Graphics* **14**:33–38 DOI [10.1016/0263-7855\(96\)00018-5](https://doi.org/10.1016/0263-7855(96)00018-5).
- Jackman M. 2011.** G2/M transition. *Encyclopedia of cancer* 1482–1484.
- Ju J-Q, Li X-H, Pan M-H, Xu Y, Sun M-H, Xu Y, Sun S-C. 2020.** CHK1 monitors spindle assembly checkpoint and DNA damage repair during the first cleavage of mouse early embryos. *Cell Proliferation* **53**:e12895 DOI [10.1111/cpr.12895](https://doi.org/10.1111/cpr.12895).
- Kellogg EH, Hejab NMA, Howes S, Northcote P, Miller JH, Diaz JF, Downing KH, Nogales E. 2017.** Insights into the distinct mechanisms of action of taxane and non-taxane microtubule stabilizers from cryo-EM structures. *The Journal of Molecular Biology* **429**:633–646 DOI [10.1016/j.jmb.2017.01.001](https://doi.org/10.1016/j.jmb.2017.01.001).
- Kousholt AN, Menzel T, Sorensen CS. 2012.** Pathways for genome integrity in G2 phase of the cell cycle. *Biomolecules* **2**:579–607 DOI [10.3390/biom2040579](https://doi.org/10.3390/biom2040579).
- Lehmann BD, Pietenpol JA. 2014.** Identification and use of biomarkers in treatment strategies for triple-negative breast cancer subtypes. *The Journal of Pathology* **232**:142–150 DOI [10.1002/path.4280](https://doi.org/10.1002/path.4280).
- Liao GS, Apaya MK, Shyur LF. 2013.** Herbal medicine and acupuncture for breast cancer palliative care and adjuvant therapy. *Evidence-based Complementary Alternative Medicine* **2013**:437948 DOI [10.1155/2013/437948](https://doi.org/10.1155/2013/437948).
- Lindqvist A, Rodriguez-Bravo V, Medema RH. 2009.** The decision to enter mitosis: feedback and redundancy in the mitotic entry network. *The Journal of Cell Biology* **185**:193–202 DOI [10.1083/jcb.200812045](https://doi.org/10.1083/jcb.200812045).
- Lipinski CA, Lombardo F, Dominy BW, Feeney PJ. 2001.** Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Advanced Drug Delivery Reviews* **46**:3–26 DOI [10.1016/s0169-409x\(00\)00129-0](https://doi.org/10.1016/s0169-409x(00)00129-0).
- Liu CY, Chang HS, Chen IS, Chen CJ, Hsu ML, Fu SL, Chen YJ. 2011a.** Costunolide causes mitotic arrest and enhances radiosensitivity in human hepatocellular carcinoma cells. *Radiation Oncology* **6**:56 DOI [10.1186/1748-717X-6-56](https://doi.org/10.1186/1748-717X-6-56).
- Liu Y, Chen XQ, Liang HX, Zhang FX, Zhang B, Jin J, Chen YL, Cheng YX, Zhou GB. 2011b.** Small compound 6-O-angeloylplenolin induces mitotic arrest and exhibits therapeutic potentials in multiple myeloma. *PLOS ONE* **6**:e21930 DOI [10.1371/journal.pone.0021930](https://doi.org/10.1371/journal.pone.0021930).
- Liu ST, Zhang H. 2016.** The mitotic checkpoint complex (MCC): looking back and forth after 15 years. *AIMS Molecular Science* **3**:597–634 DOI [10.3934/molsci.2016.4.597](https://doi.org/10.3934/molsci.2016.4.597).
- Lobert S, Fahy J, Hill BT, Duflos A, Etievant C, Correia JJ. 2000.** Vinca alkaloid-induced tubulin spiral formation correlates with cytotoxicity in the leukemic L1210 cell line. *Biochemistry* **39**:12053–12062 DOI [10.1021/bi001038r](https://doi.org/10.1021/bi001038r).

- Lottersberger F, Karssemeijer RA, Dimitrova N, De Lange T. 2015.** 53BP1 and the LINC complex promote microtubule-dependent DSB mobility and DNA repair. *Cell* **163**:880–893 DOI [10.1016/j.cell.2015.09.057](https://doi.org/10.1016/j.cell.2015.09.057).
- Ma G, Chong L, Li Z, Cheung AH, Tattersall MH. 2009.** Anticancer activities of sesquiterpene lactones from *Cyathocline purpurea* in vitro. *Cancer Chemotherapy and Pharmacology* **64**:143–152 DOI [10.1007/s00280-008-0863-y](https://doi.org/10.1007/s00280-008-0863-y).
- Ma S, Rong Z, Liu C, Qin X, Zhang X, Chen Q. 2021.** DNA damage promotes microtubule dynamics through a DNA-PK-AKT axis for enhanced repair. *The Journal of Cell Biology* **220**:e201911025 DOI [10.1083/jcb.201911025](https://doi.org/10.1083/jcb.201911025).
- Manchado E, Guillamot M, Malumbres M. 2012.** Killing cells by targeting mitosis. *Cell Death and Differentiation* **19**:369–377 DOI [10.1038/cdd.2011.197](https://doi.org/10.1038/cdd.2011.197).
- Matthews HK, Bertoli C, De Bruin RAM. 2021.** Cell cycle control in cancer. *Nature Reviews Molecular Cell Biology* **23**:74–88 DOI [10.1038/s41580-021-00404-3](https://doi.org/10.1038/s41580-021-00404-3).
- Mitra A, Sept D. 2004.** Localization of the antimitotic peptide and depsipeptide binding site on beta-tubulin. *Biochemistry* **43**:13955–13962 DOI [10.1021/bi0487387](https://doi.org/10.1021/bi0487387).
- Molina L, Williams DE, Andersen RJ, Golsteyn RM. 2021.** Isolation of a natural product with anti-mitotic activity from a toxic Canadian prairie plant. *Heliyon* **7**:e07131 DOI [10.1016/j.heliyon.2021.e07131](https://doi.org/10.1016/j.heliyon.2021.e07131).
- Mollinedo F, Gajate C. 2003.** Microtubules, microtubule-interfering agents and apoptosis. *Apoptosis* **8**:413–450 DOI [10.1023/a:1025513106330](https://doi.org/10.1023/a:1025513106330).
- Morris GM, Huey R, Lindstrom W, Sanner MF, Belew RK, Goodsell DS, Olson AJ. 2009.** AutoDock4 and AutoDockTools4: automated docking with selective receptor flexibility. *The Journal of Computational Chemistry* **30**:2785–2791 DOI [10.1002/jcc.21256](https://doi.org/10.1002/jcc.21256).
- Mukhtar E, Adhami VM, Mukhtar H. 2014.** Targeting microtubules by natural agents for cancer therapy. *Molecular Cancer Therapeutics* **13**:275–284 DOI [10.1158/1535-7163.MCT-13-0791](https://doi.org/10.1158/1535-7163.MCT-13-0791).
- Nahata A, Saxena A, Suri N, Saxena AK, Dixit VK. 2013.** *Sphaeranthus indicus* induces apoptosis through mitochondrial-dependent pathway in HL-60 cells and exerts cytotoxic potential on several human cancer cell lines. *Integrative Cancer Therapies* **12**:236–247 DOI [10.1177/1534735412451997](https://doi.org/10.1177/1534735412451997).
- Nunez Abad M, Calabuig-Farinas S, Lobo de Mena M, Jose Godes Sanz de Bremond M, Garcia Gonzalez C, Torres Martinez S, Garcia-Garcia JA, Iranzo Gonzalez-Cruz V, Camps Herrero C. 2021.** Update on systemic treatment in early triple negative breast cancer. *Therapeutic Advances in Medical Oncology* **13**:1758835920986749 DOI [10.1177/1758835920986749](https://doi.org/10.1177/1758835920986749).
- Pandey P, Singh D, Hasanain M, Ashraf R, Maheshwari M, Choyal K, Singh A, Datta D, Kumar B, Sarkar J. 2019.** 7-hydroxyfrullanolide, isolated from *Sphaeranthus indicus*, inhibits colorectal cancer cell growth by p53-dependent and -independent mechanism. *Carcinogenesis* **40**:791–804 DOI [10.1093/carcin/bgy176](https://doi.org/10.1093/carcin/bgy176).
- Parker AL, Kavallaris M, McCarroll JA. 2014.** Microtubules and their role in cellular stress in cancer. *Frontiers in Oncology* **4**:153 DOI [10.3389/fonc.2014.00153](https://doi.org/10.3389/fonc.2014.00153).

- Petsalaki E, Zachos G. 2020.** DNA damage response proteins regulating mitotic cell division: double agents preserving genome stability. *The FEBS Journal* **287**:1700–1721 DOI [10.1111/febs.15240](https://doi.org/10.1111/febs.15240).
- Piccolo M, Misso G, Ferraro MG, Riccardi C, Capuozzo A, Zarone MR, Maione F, Trifuoggi M, Stiuso P, D’Errico G, Caraglia M, Paduano L, Montesarchio D, Irace C, Santamaria R. 2019.** Exploring cellular uptake, accumulation and mechanism of action of a cationic Ru-based nanosystem in human preclinical models of breast cancer. *Scientific Reports* **9**:7006 DOI [10.1038/s41598-019-43411-3](https://doi.org/10.1038/s41598-019-43411-3).
- Prota AE, Setter J, Waight AB, Bargsten K, Murga J, Diaz JF, Steinmetz MO. 2016.** Pironetin binds covalently to alpha Cys316 and perturbs a major loop and helix of alpha-tubulin to inhibit microtubule formation. *The Journal of Molecular Biology* **428**:2981–2988 DOI [10.1016/j.jmb.2016.06.023](https://doi.org/10.1016/j.jmb.2016.06.023).
- Rahman STA, Elbashir AA, El-Mukhtar M, Ibrahim MM. 2016.** Application of spectrophotometric methods for the determination of thiamine (VB1) in pharmaceutical formulations using 7-chloro-4-nitrobenzoxadiazole (NBD-Cl). *The Journal of Analytical and Pharmaceutical Research* **2**:00018 DOI [10.15406/japlr.2016.02.00018](https://doi.org/10.15406/japlr.2016.02.00018).
- Rai SS, Wolff J. 1996.** Localization of the vinblastine-binding site on beta-tubulin. *The Journal of Biological Chemistry* **271**:14707–14711 DOI [10.1074/jbc.271.25.14707](https://doi.org/10.1074/jbc.271.25.14707).
- Robinson TJ, Liu JC, Vizeacoumar F, Sun T, Maclean N, Egan SE, Schimmer AD, Datti A, Zacksenhaus E. 2013.** RB1 status in triple negative breast cancer cells dictates response to radiation treatment and selective therapeutic drugs. *PLOS ONE* **8**:e78641 DOI [10.1371/journal.pone.0078641](https://doi.org/10.1371/journal.pone.0078641).
- Ruangrungsi N, Kasiwong S, Likhitwitayawuid K, Lange GL, Decicco CP. 1989.** Constituents of *Grangea maderaspatana*. a new eudesmanolide. *The Journal of Natural Products* **52**:130–134 DOI [10.1021/np50061a016](https://doi.org/10.1021/np50061a016).
- Saechan C, Nguyen UH, Wang Z, Sugimoto S, Yamano Y, Matsunami K, Otsuka H, Phan GM, Pham VH, Tipmanee V, Kaewsrichan J. 2021.** Potency of bisresorcinol from *Heliciopsis terminalis* on skin aging: in vitro bioactivities and molecular interactions. *PeerJ* **9**:e11618 DOI [10.7717/peerj.11618](https://doi.org/10.7717/peerj.11618).
- Sato M, Toda T. 2004.** Reconstruction of microtubules: entry into interphase. *Developmental Cell* **6**:456–458 DOI [10.1016/S1534-5807\(04\)00101-7](https://doi.org/10.1016/S1534-5807(04)00101-7).
- Schmidt M, Rohe A, Platzer C, Najjar A, Erdmann F, Sippl W. 2017.** Regulation of G2/M transition by inhibition of WEE1 and PKMYT1 kinases. *Molecules* **22**:2045 DOI [10.3390/molecules22122045](https://doi.org/10.3390/molecules22122045).
- Sharma P. 2016.** Biology and management of patients with triple-negative breast cancer. *Oncologist* **21**:1050–1062 DOI [10.1634/theoncologist.2016-0067](https://doi.org/10.1634/theoncologist.2016-0067).
- Sharma P. 2018.** Update on the treatment of early-stage triple-negative breast cancer. *Current Treatment Options in Oncology* **19**:22 DOI [10.1007/s11864-018-0539-8](https://doi.org/10.1007/s11864-018-0539-8).
- Sharma A, Singh K, Almasan A. 2012.** Histone H2AX phosphorylation: a marker for DNA damage. *Methods in Molecular Biology* **920**:613–626 DOI [10.1007/978-1-61779-998-3_40](https://doi.org/10.1007/978-1-61779-998-3_40).

- Steinbruck L, Pereira G, Efferth T. 2010.** Effects of artesunate on cytokinesis and G(2)/M cell cycle progression of tumour cells and budding yeast. *Cancer Genomics Proteomics* 7:337–346.
- Steinmetz MO, Prota AE. 2018.** Microtubule-targeting agents: strategies to hijack the cytoskeleton. *Trends in Cell Biology* 28:776–792 DOI 10.1016/j.tcb.2018.05.001.
- Sudakin V, Chan GK, Yen TJ. 2001.** Checkpoint inhibition of the APC/C in HeLa cells is mediated by a complex of BUBR1, BUB3, CDC20, and MAD2. *The Journal of Cell Biology* 154:925–936 DOI 10.1083/jcb.200102093.
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, Bray F. 2021.** Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: A Cancer Journal for Clinicians* 71:209–249 DOI 10.3322/caac.21660.
- Syred HM, Welburn J, Rappsilber J, Ohkura H. 2013.** Cell cycle regulation of microtubule interactomes: multi-layered regulation is critical for the inter-phase/mitosis transition. *Molecular and Cellular Proteomics* 12:3135–3147 DOI 10.1074/mcp.M113.028563.
- Tischer J, Gergely F. 2019.** Anti-mitotic therapies in cancer. *The Journal of Cell Biology* 218:10–11 DOI 10.1083/jcb.201808077.
- Uppatanpreecha P. 2009.** Topoisomerase I inhibitory activity from Thai medicinal plants in yeast cell-based assay. M.Sc. Thesis, Chulalongkorn University.
- Usui T, Watanabe H, Nakayama H, Tada Y, Kanoh N, Kondoh M, Asao T, Takio K, Watanabe H, Nishikawa K, Kitahara T, Osada H. 2004.** The anticancer natural product pironetin selectively targets Lys352 of alpha-tubulin. *Chemistry & Biology* 11:799–806 DOI 10.1016/j.chembiol.2004.03.028.
- Wang C, Kar S, Lai X, Cai W, Arfuso F, Sethi G, Lobie PE, Goh BC, Lim LHK, Hartman M, Chan CW, Lee SC, Tan SH, Kumar AP. 2018.** Triple negative breast cancer in Asia: an insider's view. *Cancer Treatment Reviews* 62:29–38 DOI 10.1016/j.ctrv.2017.10.014.
- Whipple RA, Vitolo MI, Boggs AE, Charpentier MS, Thompson K, Martin SS. 2013.** Parthenolide and costunolide reduce microtentacles and tumor cell attachment by selectively targeting detyrosinated tubulin independent from NF-kappa B inhibition. *Breast Cancer Research* 15:R83 DOI 10.1186/bcr3477.
- Yang NJ, Hinner MJ. 2015.** Getting across the cell membrane: an overview for small molecules, peptides, and proteins. *Methods in Molecular Biology* 1266:29–53 DOI 10.1007/978-1-4939-2272-7_3.
- Yang J, Wang Y, Wang T, Jiang J, Botting CH, Liu H, Chen Q, Yang J, Naismith JH, Zhu X, Chen L. 2016.** Pironetin reacts covalently with cysteine-316 of α -tubulin to destabilize microtubule. *Nature Communications* 7:12103 DOI 10.1038/ncomms12103.
- Yin L, Duan J-J, Bian X-W, Yu S-C. 2020.** Triple-negative breast cancer molecular subtyping and treatment progress. *Breast Cancer Research* 22:61 DOI 10.1186/s13058-020-01296-5.
- Zannini L, Delia D, Buscemi G. 2014.** CHK2 kinase in the DNA damage response and beyond. *The Journal of Molecular Cell Biology* 6:442–457 DOI 10.1093/jmcb/mju045.

Zhu H. 2012. Cell proliferation assay by flow cytometry (BrdU and PI staining). *Bioprotocol* 2:e198 DOI [10.21769/BioProtoc.198](https://doi.org/10.21769/BioProtoc.198).