

This is a repository copy of *Impact of Conventional and Sustainable Solvents on the Yield, Selectivity, and Recovery of Curcuminoids from Turmeric*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/181654/>

Version: Accepted Version

Article:

Kisanthia, Rattikon, Hunt, Andrew J. orcid.org/0000-0003-3983-8313, Sherwood, James orcid.org/0000-0001-5431-2032 et al. (2 more authors) (2021) Impact of Conventional and Sustainable Solvents on the Yield, Selectivity, and Recovery of Curcuminoids from Turmeric. ACS Sustainable Chemistry and Engineering. 104–114. ISSN 2168-0485

<https://doi.org/10.1021/acssuschemeng.1c04882>

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.

1 **Impact of conventional and sustainable solvents on the yield, selectivity and recovery of**
2 **curcuminoids from turmeric**

3 Rattikon Kisanthia^a, Andrew J. Hunt^b, James Sherwood^c, La-or Somsakeesit^d and Chanokbhorn
4 Phaosiri*^a

5

6

7 ^a Natural Products Research Unit, Center of Excellence for Innovation in Chemistry,
8 Department of Chemistry, Faculty of Science, Khon Kaen University, Khon Kaen,
9 40002, Thailand.

10 ^b Materials Chemistry Research Center (MCRC), Centre of Excellence for Innovation in
11 Chemistry, Department of Chemistry, Faculty of Science, Khon Kaen University, Khon Kaen,
12 40002, Thailand.

13 ^c Green Chemistry Centre of Excellence, Department of Chemistry, University of York,
14 Heslington, YO10 5DD, UK.

15 ^d Department of Chemistry, Faculty of Engineering, Rajamangala University of Technology Isan,
16 Khon Kaen Campus, Khon Kaen, 40000, Thailand.

17

18 *Corresponding to: Dr. Chanokbhorn Phaosiri, Khon Kaen University

19 Email: chapa@kku.ac.th; Phone: 66-897119936; Fax: 66-4320-2373

20 ABSTRACT:

21 Extraction of pharmaceutically important curcuminoid platform molecules has been
22 achieved from turmeric with ultrasound-assisted greener solvent extraction, demonstrating
23 excellent extraction performance and product recovery. Extraction of curcuminoids from turmeric
24 was undertaken with both conventional and potentially bio-based solvents. Sustainable solvents,
25 namely, ethyl acetate, ethanol, Cyrene and deep eutectic solvent (DES-B5, 1:6 ChCl:1,4-
26 butanediol) demonstrated high extraction yields of curcuminoids, including 24.36 ± 3.10 mg/g,
27 25.30 ± 4.58 mg/g, 23.51 ± 2.56 mg/g and 27.40 ± 3.80 mg/g of curcumin, respectively. In contrast,
28 curcumin extracted in lower polarity solvents such as hexane, toluene and tetramethyloxolane
29 (TMO) were low at less than 7.0 mg/g. DES-B5 with 10% water extracted the greatest yield of
30 curcumin (46.70 ± 0.55 mg/g), bisdemethoxycurcumin (46.14 ± 0.82 mg/g) and
31 demethoxycurcumin (10.63 ± 0.35 mg/g), followed by a simple and low energy product recovery
32 method through the addition of water and precipitation. COSMOtherm calculations suggested that
33 extraction efficiency was related to solvent interactions with the cell walls of the biomass, rather
34 than the solubility of the curcuminoids. In addition, application of ultrasound in extraction in
35 combination with DES-B5 enabled the strong destruction of plant matrix that was confirmed by
36 scanning electron microscopy (SEM). The sustainability, efficiency and toxicity of proposed
37 extraction methodologies were evaluated through the CHEM21 green metrics toolkit. The methods
38 utilising ethanol, ethyl acetate and Cyrene in this work demonstrate a significant improvement
39 over those previously published by 3 to 10 times of process mass intensity (PMI) total, while DES-
40 B5 also performed well under the green metric assessment. Because of its high yields, bio-based
41 solvent, low toxicity, being inexpensive and readily available, DES-B5 with 10% water is
42 recommended as the greenest solvent for curcuminoid extraction under ultra-sonication.

43 **KEYWORDS:** Curcuminoids; *Curcuma longa*; Cyrene; Deep eutectic solvents; ultrasound;
44 tetramethyloxolane (TMO).

45

46 **INTRODUCTION**

47 Turmeric (*Curcuma longa*) is an important plant in a family of ginger and can be found
48 across Asia including India, Indonesia and Thailand. It is used widely as an ingredient of food,
49 herbal medicine and in natural dyes.¹ There are three important components in the turmeric
50 rhizome that are collectively known as “curcuminoids”: curcumin (**1**), bisdemethoxycurcumin (**2**)
51 and demethoxycurcumin (**3**), as shown in Figure 1. Curcumin especially is well known as a yellow
52 natural product with bioactive properties including antioxidant, anti-inflammatory, anticancer,
53 anti-diabetic and anti-microbial activities.²⁻⁴ Therefore, it is an attractive platform molecule for the
54 development of new drug candidates by the pharmaceutical industry. Unfortunately, curcuminoids
55 are low-polarity organic compounds that are insoluble in water.⁵ Furthermore, they are thermally
56 liable phenolic compounds that also degrade in strong light.⁶ As such, the use of conventional
57 organic solvents under mild extraction conditions are necessary for further application of
58 curcuminoids. Nowadays, curcuminoids are often extracted with conventional solvents, namely,
59 dichloromethane, methanol or acetone.^{7,8} One possible methodology to enhance safety and
60 efficiency for curcuminoid extraction is an application of green solvents, for instance, bio-based
61 solvents (from starch, wood or vegetable) or environmentally friendly petroleum-based
62 solvents.^{9,10} The utilization of green solvents instead of hazardous or highly hazardous solvents
63 such as benzene, toluene, diethyl ether and halogenated solvents can be less harmful to human
64 health and the environment. Bio-based Cyrene is alternative green solvent with excellent
65 properties, namely, low toxicity, non-mutagenicity and biodegradability.¹¹ It has been successfully

66 utilised instead of conventional dipolar aprotic solvents such as *N,N'*-dimethylformamide (DMF)
67 and *N*-methyl pyrrolidinone (NMP). However, Cyrene offers a higher boiling point (203 °C) and
68 a higher viscosity (1.25 g/mL) that can limit its application in extraction. Tetramethyloxolane
69 (TMO) is non-peroxide forming ether that derives from potentially renewable feedstocks.¹² It is
70 alternative green solvent to toluene, representing similar properties such as melting point (-95 °C),
71 density (0.802 g/mL) and boiling point (112 °C). TMO has the potential to be reaction solvent in
72 esterifications, amidations and Grignard reactions.¹²

73 In addition, deep eutectic solvents (DESs) have been developed as alternative green
74 solvents which are well known as one class of ionic liquids (IL), representing excellent properties
75 such as low price, potential biodegradability, low-toxicity, low-volatility, non-flammability and
76 are stable at high temperature.^{13,14} Generally, DESs are synthesised from two components
77 including a hydrogen bond donor (HBD, *e.g.* ethylene glycol, glycerol and organic acid) and a
78 hydrogen bond acceptor (HBA, *e.g.* quaternary ammonium salts) via hydrogen bond formation. In
79 2020, Altunay *et al.* studied a microextraction of curcumin from spiked foodstuff samples with
80 deep eutectic solvents.¹⁵ The highest percent recovery of curcumin in spiked food samples was
81 observed in DES that was prepared from betaine hydrochloride and glycerol at 1:3 ratio. However,
82 the extraction was studied only in small scale for analytical chemistry. In 2021, Degot *et al.*
83 extracted curcuminoids from turmeric rhizome with ethanol and food additives (triacetin and
84 diacetin).¹⁶ The results showed that the addition of triacetin provided advance extraction power.
85 As a result, the use of combination solvent for curcuminoid extraction can increase the extraction
86 efficiency. Therefore, from the previous reports, the efficient extraction of curcuminoids in greener
87 solvents, namely, Cyrene, TMO and DESs remains of interest.

88 Herein, this work aimed to investigate curcuminoid extraction using conventional and
89 alternative green solvents to develop sustainable methods for natural product extraction.
90 Ultrasound-assisted extraction (UAE) was utilized as an energy efficient green technology to
91 rupture the cell wall of the plant matrix to enhance the extraction process. Furthermore, a series of
92 choline chloride based DESs were synthesised and used in parallel with conventional and other
93 green solvents for curcuminoid extraction. The curcuminoids were evaluated by RP-HPLC
94 analysis. Moreover, computational analysis using Conductor-like Screening Model for Realistic
95 Solvents (COSMO-RS) was conducted with selected solvents to rationalize the experimental
96 results.

97

98 **EXPERIMENTAL SECTION**

99 **Materials**

100 Acetonitrile (HPLC grade, Merck, Germany), DMF, ethanol (HPLC grade), ethyl acetate,
101 hexane, methanol (HPLC grade), glycerol, tetrahydrofuran (THF), toluene and common reagents
102 were obtained from local suppliers. Cyrene from renewable cellulose was supported by Circa
103 company (>99.0%, Australia). 2-Methyltetrahydrofuran (2-MeTHF) (purity >99.5%, bio-
104 renewable source) was purchased from Sigma Aldrich company, USA. TMO was synthesised in
105 the laboratory according to the previous procedure.¹² Choline chloride (ChCl) (purity >98.0%),
106 ethylene glycol (>99.5%) and 1,4-butanediol (99.0%) were obtained from Tokyo Chemical
107 Industry, Japan.

108

109 **Plant material**

110 Turmeric powder from *Curcuma longa* rhizomes was purchased from a local market in
111 Khon Kaen province, Thailand. The dried powder was stored in a sealed dark bag at ambient
112 temperature protected from the light and humid exposure.

113

114 **Preparation of DES**

115 Choline chloride (ChCl) and a hydrogen bond donor (HBD) of either ethylene glycol, 1,4-
116 butanediol or glycerol were combined to prepare DESs via hydrogen bond formation. The choline
117 chloride based DESs were prepared by mixing ChCl and each HBD in 250 mL beaker and heated
118 at 80 °C for 30 minutes with stirring until a homogenous solution was formed. The component
119 ratio of choline chloride based DESs are presented in Table 1. Even though most DESs were
120 synthesised easily at 80 °C, DES-B1 was reheated for five times to form stable DES at room
121 temperature.¹⁷ An attempt to synthesise betaine hydrochloride-based DESs was not successful.

122

123 **Extraction of curcuminoids from *Curcuma longa***

124 Curcuminoids were extracted from 5 g of turmeric powder in 15 mL of solvent, including
125 hexane, ethyl acetate, DMF, ethanol, toluene, Cyrene, 2-MeTHF and TMO under ultra-sonication
126 (35 kHz, RK 103 H, Bandelin, Germany) for two hours at ambient temperature. Afterward, the
127 crude extracts were filtered using filter paper, and the dried crude extracts were obtained using
128 rotary evaporator (Buchi R-300, Switzerland). The dried crude extracts were transferred, and the
129 volume adjusted in a 25 mL volumetric flask by addition of HPLC grade methanol. Finally,
130 curcuminoids in crude extracts were evaluated using HPLC equipment (Supporting information,
131 Scheme S1). The extractions were carried out in triplicate. In the case of deep eutectic solvents,

132 curcuminoids were also extracted in parallel with the other solvents. However, a centrifugation
133 was applied instead of filtration due to high viscosity of DESs.

$$134 \quad \text{Selectivity of total curcuminoids} = \frac{\text{total curcuminoids (mg)}}{\text{dried crude extract (mg)}} \times 100 \quad (1)$$

$$135 \quad \text{Selectivity of each curcuminoid} = \frac{\text{curcuminoid (mg)}}{\text{total curcuminoids (mg)}} \times 100 \quad (2)$$

136 **HPLC Analysis**

137 Curcuminoids were measured by RP-HPLC following a previous report with some
138 modification.¹⁸ The quantity of curcuminoids was evaluated by Agilent 1200 series HPLC system
139 (Agilent, San Jose, CA, USA). The separation was carried out using acetonitrile (A) and 0.1 M
140 ammonium acetate buffer pH 3.50 (B) as mobile phases with a gradient elution program. The
141 initial composition of mobile phase was 50:50 v/v. The linear gradient was applied to A-B (30:70,
142 v/v) in 8.0 min. Then, the composition of mobile phase was changed to 100% of A in 4.0 min. The
143 C18 column (4.6 × 150 mm × 5 μm, Agilent, USA) was utilized for separation at ambient
144 temperature. The injection volume was 10 μL with flow rate of mobile phase 1 mL/min and the
145 target curcuminoids were detected at 425 nm. The calibration curves of standards of curcumin,
146 bisdemethoxycurcumin and demethoxycurcumin were established from 2-150 ppm in order to
147 identify and determine the quantity of curcuminoid content (Supporting information, Figure S1).

148

149 **Isolation of curcuminoids from DES**

150 The addition of water induced precipitation of curcuminoids in the crude DES extract. The
151 appropriate conditions were studied by varying the amount of water in 1 mL of crude DES extract.
152 The optimised conditions appeared to be three-fold of water. The solid curcuminoid precipitate
153 appeared after the addition of water. Afterward, the solids were filtered, washed with water and

154 dried in an oven at 40 °C. Subsequently, the curcuminoids were kept in a desiccator until a stable
155 mass was observed.

156

157 **Conductor-like Screening Model for Real Solvents (COSMO-RS)**

158 ArgusLab (version 4.0.1, Mark Thompson and Planaria Software LLC, 2004, Seattle, WA,
159 USA) was used to obtain approximate atomic coordinates of compounds. The conformations of
160 the curcuminoids were calculated with COSMOconfX (version 4.0; COSMOlogic GmbH & Co.
161 KG, Leverkusen, Germany, 2015). COSMOthermX (version C30_1705; COSMOlogic GmbH &
162 Co. KG, 2017, TZVP basis set level) was used to provide molecular surface charges and execute
163 the virtual experiments.

164

165 **RESULTS AND DISCUSSION**

166 **Extraction of Curcuminoids from *Curcuma longa***

167 To seek the best solvent for curcuminoid extraction, comparative studies between
168 conventional and green solvents were investigated. Curcuminoids including curcumin,
169 bisdemethoxycurcumin and demethoxycurcumin were extracted from turmeric rhizome powder
170 using conventional solvents (hexane, DMF, toluene, ethyl acetate, ethanol and THF) and
171 alternative green solvents (2-MeTHF, TMO and Cyrene). The selected solvents exhibit a range of
172 properties, as shown in Table 2. Curcuminoids are thermally labile and as such UAE was utilised
173 to enhance the extraction yield and avoid the higher temperatures that may lead to degradation.¹⁹
174 The quantity of extracted curcuminoids were evaluated by RP-HPLC with UV-VIS detection at
175 425 nm. The results demonstrated that the extract was predominantly curcumin and
176 bisdemethoxycurcumin in most solvents, with small amounts of demethoxycurcumin also

177 observed. Curcumin extracted in lower polarity solvents such as hexane, toluene and TMO was
178 less than 7.0 mg/g, whereas in the polar solvents including ethanol, ethyl acetate and Cyrene
179 excellent extraction yields were exhibited of 24.36 ± 3.10 mg/g, 25.30 ± 4.58 mg/g and $23.51 \pm$
180 2.56 mg/g, respectively (Figure 2). These polar solvents also demonstrated excellent extraction
181 yields of bisdemethoxycurcumin and demethoxycurcumin. The highest yield of
182 bisdemethoxycurcumin was 25.57 ± 3.45 mg/g, as observed in ethanol. Ethyl acetate and Cyrene
183 exhibited the similar bisdemethoxycurcumin extraction yields of 23.54 ± 2.64 mg/g and $23.37 \pm$
184 2.44 mg/g, respectively. Ethanol also gave the highest extraction yield of demethoxycurcumin at
185 6.10 ± 0.82 mg/g. The extraction yields of demethoxycurcumin from ethyl acetate and Cyrene
186 were 5.34 ± 0.87 mg/g and 5.97 ± 0.71 mg/g respectively. However, some polar solvents, including
187 THF (15.17 ± 2.08 mg/g), 2-MeTHF (13.80 ± 3.90 mg/g) and DMF (14.43 ± 4.46 mg/g) only led
188 to low to moderate extraction yields of curcumin. These solvents also provided low extraction
189 yields of bisdemethoxycurcumin and demethoxycurcumin.

190 Kumboonma *et al.* investigated the recovery of curcuminoids by the conventional
191 methodology of soaking turmeric rhizome powder in three sequential extractions of
192 dichloromethane at room temperature. The isolated curcumin, bisdemethoxycurcumin and
193 demethoxycurcumin was found to be 33.33 mg/g, 1.66 mg/g and 0.083 mg/g, respectively after
194 column chromatography.²¹ Accordingly, the best three solvents in this current work (ethanol, ethyl
195 acetate and Cyrene) provided a comparable extraction yields of curcumin. Interestingly, the
196 extraction yields of bisdemethoxycurcumin and demethoxycurcumin from the ethanol extract were
197 73 and 15 times greater respectively than that of the dichloromethane extract. Even though
198 excellent extraction yields of curcuminoids were obtained with ethanol, ethyl acetate and Cyrene,
199 these solvents present some minor health and safety hazards. As an alternative, DESs were

200 designed and synthesised from natural sources to further investigate the extraction, due to their
201 attractive properties such as low toxic, reusable, low cost, non-flammable, and biodegradable
202 properties. Therefore, several choline chloride-based DESs were synthesised using ethylene
203 glycol, 1,4-butanediol and glycerol and applied to the curcuminoid extraction from turmeric. The
204 selected chemicals for synthesis of DESs were considered and chosen from their safety, low
205 toxicity and natural source.^{13,38} In addition, ChCl-based DESs increased the flavonoid extraction
206 capacity due to weak acidity from hydroxy groups on flavonoids which related to the structure of
207 curcuminoids.²² Hence, the ChCl-based DESs were selected in this work. The ratio of hydrogen
208 bond acceptor (HBA) and hydrogen bond donor (HBD) was varied from 1:2 to 1:6 HBA:HBD.

209 The method of curcuminoid extraction with DESs was partially modified by application of
210 centrifugation on account of the high viscosity of DESs. The extraction results indicated that both
211 the ethylene glycol and glycerol based DESs tended to decrease the extraction yield when the ratio
212 of HBD was increased. On the other hand, 1,4-butanediol-based DESs exhibited increased
213 extraction yields with the increased ratio of HBD as shown in Figure 3. A poor extraction yield
214 was observed in all glycerol-based DESs, this was likely due to the high viscosity of the solvent
215 hindering the extraction and preventing mass transfer of the target molecules from the plant
216 matrix.²³ Among all DESs, DES-B5 (1 part choline chloride, 6 parts 1,4-butanediol) demonstrated
217 the best yield of curcumin (27.40 ± 3.80 mg/g), bisdemethoxycurcumin (23.24 ± 0.61 mg/g) and
218 demethoxycurcumin (5.22 ± 0.61 mg/g). DES-B5 possessed the lowest viscosity among 1,4-
219 butanediol based series because of the increasing of HBD proportion that allowed mass transfer
220 process.^{23,24}

221

222 **Optimization of the Curcuminoid Extraction Conditions**

223 Due to the highest extraction efficiency, DES-B5 was further selected as the DESs for
224 optimisation of the extraction. The greatest curcuminoid extraction increased with solvent volume
225 until a ratio of 1:15 g/mL. After which, higher quantities of DESs slightly decreased the
226 curcuminoid yield, as shown in Figure 4 (a). Therefore, the ratio with the highest extraction yield
227 (1:15 g/mL) was selected for further optimisation. The water content of the DESs was further
228 investigated with the addition of 0% to 40% v/v. The extraction results indicated that 10% water
229 content afforded advance extraction performance of curcuminoids: curcumin (43.35 ± 1.49 mg/g),
230 bisdemethoxycurcumin (42.23 ± 0.33 mg/g) and demethoxycurcumin (9.62 ± 0.50 mg/g) as shown
231 in Figure 4 (b). A likely explanation for the beneficial effect of adding a small quantity of water
232 (10% v/v) is the reduction in viscosity of the DES, and it might affect to swell of plant matrix and
233 solvent penetration, therefore benefiting mass transfer during the extraction process.²⁵⁻²⁹
234 Moreover, a water content of Cyrene was also studied, however the extraction yield was not
235 enhanced by the addition of water (Figure S3, supporting information).

236 The total curcuminoids extracted with each solvent were calculated as the sum of curcumin,
237 bisdemethoxycurcumin and demethoxycurcumin based on HPLC as shown in Table 3. Ethanol,
238 ethyl acetate, Cyrene and DES-B5 (with 10% water) afforded the highest total curcuminoids with
239 56.03 mg/g, 54.17 mg/g, 52.85 mg/g and 55.86 mg/g, respectively. The results indicated that the
240 alternative green solvents, namely, Cyrene and DES-B5 exhibited extraction performance like
241 ethanol and ethyl acetate. The results suggested that the extraction of curcuminoids was not only
242 influenced by polarity and viscosity, but it might be affected by several factors such as penetration
243 of solvent and encapsulation of target molecule from the plant matrix.³⁰

244

245 **Recovery of curcuminoids from Cyrene and DESs Extracts**

246 To isolate the curcuminoids from the crude extracts, vacuum evaporation was applied to
247 most of the conventional and green solvents. However, the high boiling points of Cyrene and DES-
248 B5 make vacuum evaporation an unfeasible method for drying the crude extracts. Cyrene is known
249 to form a geminal diol upon the addition of water, and the intermolecular hydrogen bonding
250 observed in DES is disrupted by the addition of water.³¹⁻³³ Therefore, the addition of water was
251 used to induce precipitation of curcuminoids from the crude Cyrene and DES extracts. The solid
252 curcuminoid precipitate appeared immediately after the addition of water. The precipitation
253 process was left for 1 day to obtain the complete product. This phenomenon of forming a geminal
254 diol reduces curcuminoid solubility and permits isolation of the crude extract without the direct
255 energy consumption required by distillation. Afterward, the solids were filtered, washed with
256 water, and dried in an oven at 40 °C. Subsequently, the curcuminoids were kept in a desiccator
257 until a stable mass was observed. The absence of Cyrene in the dried crude extract was confirmed
258 by proton nuclear magnetic resonance (¹H NMR) spectroscopy. The dried crude extracts of all
259 solvents were represented in Table 3. The results indicated that the maximum dried crude extract
260 was obtained from Cyrene with 124.33 ± 5.13 mg/g. Slightly lower yields were observed in ethyl
261 acetate, ethanol, DES-B5 and DMF, respectively. Lower dried crude yields around 75.0-90.0 mg/g
262 were obtained from TMO, THF and 2-MeTHF, while low-polar solvents, including hexane and
263 toluene, exhibited dry crude extracts lower than 65.0 mg/g. The large mass of the dried crude
264 extracts suggested that non-curcuminoid compounds were also extracted from turmeric powder in
265 all solvents. The non-curcuminoids in the crude extract were presumed to be derived from other
266 organic compounds or pigments found in turmeric.

267

268 **Comparison of curcuminoid selectivity**

269 The selectivity of curcuminoid extraction from DES-B5 (relative to the crude extract) was
270 56%, exhibiting the highest selectivity among all solvents in this work. The selectivity of
271 curcuminoid extraction was high for ethanol, ethyl acetate, Cyrene and 2-MeTHF at 49%, 46%,
272 43% and 41%, respectively. A lower selectivity for curcuminoids (10-39%) was observed from
273 THF, DMF, TMO, toluene and hexane. Most solvents exhibited more than 50% non-curcuminoids
274 in dried crude extracts. A selectivity of each curcuminoid between the three curcuminoids was also
275 calculated as shown in Figure 5. As a result, toluene exhibited the highest selectivity of curcumin
276 at 55%, and DES-B5 provided a higher selectivity towards curcumin than that of most
277 conventional and other green solvents. TMO was highly selective towards bisdemethoxycurcumin.
278 A lone pair of ethereal oxygen of TMO could interact with a proton of phenolic compound via
279 weak hydrogen bond proposed by Byrne *et al.*³⁴ Therefore, TMO with four bulky methyl groups
280 was able to interact with hydroxy group of bisdemethoxycurcumin, which has the lowest steric
281 hindrance of the three curcuminoids. For this reason, TMO specifically interacts with
282 bisdemethoxycurcumin.

283

284 **Exploration of Dissolving Mechanism Using COSMO-RS and Solubility Test**

285 The different performance of solvents in curcuminoid extraction was evaluated by
286 COSMO-RS, calculating the possible interactions of each solvent with the target curcuminoids.
287 Ultimately there is no consistent polarity profile or physical property of the solvents that correlates
288 to extraction performance. The results indicate that the interactions between solvent and the
289 biomass matrix, such as the hydrogen bonding between the structure of the cell wall and DES is
290 critically important.²⁴ A solubility test was conducted to confirm if the curcuminoid extraction
291 yield was strongly influenced by solubility (Table 3) and the major curcuminoid, curcumin, was

292 selected for this test. The results indicated that both THF and DMF had excellent curcumin
293 solubility, at greater than 300 mg/mL. Curcumin solubility was also good for Cyrene and 2-
294 MeTHF at 83.23 mg/mL and 71.79 mg/mL, respectively. A lower curcumin solubility was
295 observed in ethyl acetate and ethanol of 26.49 mg/mL and 7.19 mg/mL, respectively, despite
296 demonstrating high yields of extraction. The experimental results of curcumin solubility were
297 consistent with the predicted solubility of curcumin from COSMO-RS that confirmed DMF and
298 THF as the best solvent for curcumin solubility. These results demonstrate that the solubility of
299 curcuminoids does not correlate with the extraction yield, and the expected increase in solubility
300 with the gradual increase in temperature during ultra-sonication from room temperature to 55 ± 2
301 °C would further diminish any solubility limit on the extraction of curcuminoids from the plant
302 matrix.

303

304 **Destruction and Dissolution of the Plant Matrix in UAE**

305 To investigate the effect of UAE on the plant matrix, a morphology of plant residues was
306 examined by scanning electron microscopy (SEM). The plant residues before and after extraction
307 with Cyrene and DES-B5 were studied as shown in Figure 6. The results indicated that the
308 morphology of turmeric rhizomes cell walls before extraction were intact layers. Post extraction
309 with Cyrene resulted in the partially breakdown plant cell walls leading to the release of target
310 molecule to the solvent. However, in the case of extraction with DES-B5 this effect was further
311 enhanced leading to the rupturing of cell walls. SEM results from Cyrene and DES-B5 were
312 consistent with the extraction yields. This leads to a proposed mechanism for extraction, where the
313 higher extraction performance relates to the strong hydrogen bonding between DES-B5 and
314 biomass leading to the destruction of plant cell walls and dissolution of curcuminoids which is

315 consistent with the literature.³⁵ Several hydroxy groups on plant cell wall (cellulose, and
316 hemicellulose) could form hydrogen bond interactions with DES-B5 as shown in Figure 7.
317 Therefore, target molecules could be released and washed out once hydrogen-bond interactions
318 between cellulose and DES-B5 with application of UAE rupture cell walls.

319

320 **CHEM21 Green Metrics Toolkit Assessment**

321 **Safety and Toxicity of Solvent Extraction**

322 In terms of solvent toxicity, the CHEM21 solvent selection guidelines was applied to
323 suggest the safest solvents for curcuminoid extraction.³⁶ According to their hazards, solvents were
324 categorised as either highly hazardous, hazardous, problematic or preferred. Classification of
325 safety, health and environmental hazards has been made on a 1 (low) to 10 (high) scoring system.
326 Dichloromethane was considered by using CHEM21 solvent selection guide as hazardous. Ethyl
327 acetate and ethanol were categorised as preferred solvents, while Cyrene was classified as
328 problematic due to its high boiling point (Table 4). Despite this, the bio-based Cyrene exhibited
329 lower safety and health issues than that of ethyl acetate and ethanol. Moreover, it represents some
330 excellent properties, including, non-mutagenicity, no acute oral toxicity ($LD_{50} > 2000$ mg/kg, the
331 highest concentration tested), biodegradability (99% in 14 days) and low ecotoxicity.³⁸ The issue
332 of the high boiling point of Cyrene was avoided by precipitation of the extract. Furthermore, the
333 mixture of Cyrene and water offered an advantage as effective solvent extraction in previous
334 work.³⁸ Although isolation of the product was not an energy intensive process, the recovery and
335 reuse of Cyrene after addition of water was challenging.

336 Sustainable DESs were not classified in CHEM21 green metrics toolkits. However, the
337 individual components can be assessed. Ethylene glycol and glycerol are categorised as preferred

338 and problematic solvents respectively, the major difference being their boiling point. The
339 greenness of 1,4-butanediol was calculated according to the CHEM21 toolkit methodology as
340 problematic, despite minimal safety, health and environmental hazards because of its low
341 volatility. Choline chloride is considered safe, biodegradable, and inexpensive, which was
342 frequently used as a favourable HBA in the synthesis of DESs. As the safety and toxicity profiles
343 of individual components were encouraging, and as such it was presumed the toxicity of choline
344 chloride based DESs was also low. Such statements are backed up toxicity measurements with the
345 human HEK-293 cell lines, however, a comprehensive toxicity testing of DESs is needed prior use
346 at scale or application in industry.³⁹

347

348 **Evaluation and Comparison of Solvent Extraction Power Using CHEM21**

349 The CHEM 21 green metrics toolkit was also used to assess an efficiency of curcuminoid
350 extraction processes and was compared with two previous published sustainable
351 methodologies.^{16,40} The green metric selected for the assessment of the extractions was process
352 mass intensity (PMI) (Table 5), the low PMI value referred to an effective method for extraction
353 process. Ethanol extract in this work offered the lowest PMI total with 60.1 g/g, while ethyl acetate
354 and Cyrene exhibited higher PMI with 68.4 g/g and 89.5 g/g, respectively. Although ethyl acetate
355 offered the higher curcuminoid yield than ethanol, the PMI of ethyl acetate was greater than that
356 of ethanol because of solvent density. Degot *et al.* extracted curcuminoids from turmeric powder
357 using a mixture of triacetin (TriA)/Ethanol/H₂O (36/24/40% w/w) and pure ethanol under stirring
358 at room temperature for 1 h, and the quantification of curcuminoids was evaluated by HPLC
359 analysis.¹⁶ Very recently, Huber *et al.* also extracted curcuminoids from turmeric rhizomes with
360 NADES-based surfactant-free microemulsions (NADES-SFME, ChCl + Lactic acid/Ethanol/TriA

361 35/27.5/37.5% w/w) under the similar condition with Degot *et al.*⁴⁰ PMI calculations of two
362 previously published extraction methods^{16,40} with pure ethanol, a mixture of TriA/Ethanol/H₂O
363 (36/24/40% w/w) and ChCl + Lactic acid/Ethanol/TriA (35/27.5/37.5% w/w), expressed higher
364 total PMI with 757.6 g/g, 327.2 g/g and 505.9 g/g, respectively. Therefore, the methods utilising
365 ethanol, ethyl acetate and Cyrene in this work demonstrate a significant improvement over
366 previously published works. The PMI of the extraction process using DES-B5 was 71.9 g/g and
367 demonstrated a higher value than that of ethanol and ethyl acetate, but lower than that of Cyrene.
368 Moreover, the PMI of the traditional literature method employing dichloromethane for
369 curcuminoid extraction was also estimated based on the quantity of curcuminoids recovered after
370 column chromatography.²¹ The PMI total of dichloromethane was calculated in this work using
371 only the amount of starting material, solvent for extraction and the obtained curcuminoids from
372 the previous work.²¹ PMI total of dichloromethane as 218.1 g/g showed higher number than that
373 of sustainable solvents in this work, namely ethanol, ethyl acetate, Cyrene and DES-B5. Even
374 though the materials used for column chromatography not accounted for, this indirect comparison
375 guided us that the low PMI of sustainable solvents (ethanol, ethyl acetate, Cyrene and DES-B5)
376 offered the potential for substitution the highly hazardous solvent, dichloromethane. These results
377 demonstrate that the proposed method of curcuminoid extraction with application of ultra-
378 sonication significantly improved a lower input mass per gram for curcuminoid extraction.
379 However, there is no universal solvent which is the safest or greenest solvents for all applications,
380 therefore the consideration of solvent remains a compromise between extraction yield,
381 harmfulness and sustainable resource.⁴¹
382 CHEM 21 toolkit demonstrated ethanol as the lowest PMI for curcuminoid extraction under ultra-
383 sonication, therefore overall results suggested DES-B5 as the greenest performance for

384 curcuminoid extraction on account of excellent properties, including, high yields, biodegradability,
385 low toxicity, being inexpensive and readily available.

386

387 **CONCLUSION**

388 Six conventional (hexane, DMF, toluene, ethyl acetate, ethanol and THF) and eighteen
389 alternative green solvents (2-MeTHF, TMO, Cyrene and DESs) were employed to extract
390 curcuminoids from turmeric powder using UAE technique. This is the first report of using Cyrene
391 and TMO for curcuminoid extraction. Greener and bio-based solvents, namely ethanol, ethyl
392 acetate, Cyrene and DES-B5 provided high effective extraction yield amongst those tested. DES-
393 B5 with 10% water content exhibited advance extraction performance. The simple isolation of
394 curcuminoid from the Cyrene and DES-B5 extracts was achieved by addition of water. Thereby
395 avoiding the energy intensive solvent evaporation steps traditionally associated with high boiling
396 point bio-based solvents. COSMO-RS modelling indicated that the extraction efficiency was not
397 influenced directly by the polarity profiles of the solvents. The solubility test suggested that high
398 solubility did not correlate with the efficiency of extraction. SEM images of plant matrix after
399 extraction in DES-B5 indicated significant rupturing of cell walls, which corresponded to high
400 extraction yields. Estimation of the green impact of conventional and green solvents for
401 curcuminoid extraction, was achieved through the application of the CHEM21 guidelines. The best
402 four solvents for curcuminoid extraction exhibited greater safety, low health issues and were more
403 environmental-benign compared to dichloromethane. A PMI total of this proposed method of
404 curcuminoid extraction with application of ultra-sonication offered 3 to 10 times better than
405 previous literature methodologies. Therefore, it significantly improved a lower input mass per
406 gram for curcuminoid extraction. Significantly, DES-B5 with 10% water content was

407 recommended as the greenest performance with several advantages, including high yields, low
408 toxicity, bio-based solvent, being inexpensive, biodegradable and readily available for
409 curcuminoid extraction. This work provides the basis for solvent selection in common natural
410 product extraction and isolation by application of sustainable solvents and green metrics
411 assessment. The bio-based solvent, 2-MeTHF was also used in curcuminoid extraction for the first
412 time. Even though 2-MeTHF is a bio-based solvent with advantages against Cyrene, the extraction
413 yield from 2-MeTHF was only moderate compared to Cyrene. However, it has potential to be used
414 as a sustainable and alternative solvent for natural product extraction which should be further
415 investigated.

416

417 **Supporting information**

418 The identification and quantification of curcuminoids by HPLC; the calibration curves of
419 curcuminoids; the extraction yields, dried crude extract and total curcuminoids in each solvent;
420 impact of water content on Cyrene and extraction yields; investigation of solubility of curcumin;
421 investigation and description of COSMO-RS modelling; SEM images; ^1H NMR spectra of
422 curcuminoid standards.

423

424 **ACKNOWLEDGEMENTS**

425 Financial support from Center of Excellence for Innovation in Chemistry (PERCH-CIC),
426 Ministry of Higher Education, Science, Research and Innovation is gratefully acknowledged. This
427 work was also supported by Materials Chemistry Research Centre (MCRC) and Natural Product
428 Research Unit, Khon Kaen University. Andrew Hunt would like to acknowledge the financial
429 support of the Thailand Research Fund, (Grant number RSA6280031) and Khon Kaen University.

430 We would like to thank Circa Group Pty Ltd. for the provision of Cyrene. COSMO-RS calculations
431 are courtesy of, and authorised for the purposes of, Circa Group Pty Ltd.

432

433 **REFERENCES**

- 434 (1) Kim, D. W.; Lee, S. M.; Woo, H. S.; Park, J. Y.; Ko, B. S.; Heo, J. D.; Ryu, Y. B.; Lee, W. S.
435 Chemical constituents and anti-inflammatory activity of the aerial parts of *Curcuma longa*. *J.*
436 *Funct. Foods* **2016**, *26*, 485–493.
- 437 (2) Hashim, A. F.; Hamed, S. F.; Abdel Hamid, H. A.; Abd-Elsalam, K. A.; Golonka, I.; Musiał,
438 W.; El-Sherbiny, I. M. Antioxidant and antibacterial activities of omega-3 rich oils/curcumin
439 nanoemulsions loaded in chitosan and alginate-based microbeads. *Int. J. Biol. Macromol.*
440 **2019**, *140*, 682–696.
- 441 (3) Ruby, A. J.; Kuttan, G.; Babu, K. D.; Rajasekharan, K. N.; Kuttan, R. Anti-tumor and
442 antioxidant activity of natural curcuminoids. *Cancer Lett.* **1995**, *94*, 79–83.
- 443 (4) Lee, H. H.; Cho, H. Improved anti-cancer effect of curcumin on breast cancer cells by
444 increasing the activity of natural killer cells. *J. Microbiol. Biotechnol.* **2018**, *28*, 874–882.
- 445 (5) Nair, A.; Amalraj, A.; Jacob, J.; Kunnumakkara, A. B.; Gopi, S. Non-Curcuminoids from
446 Turmeric and Their Potential in Cancer Therapy and Anticancer Drug Delivery Formulations.
447 *Biomolecules* **2019**, *9*, 13.
- 448 (6) Alara, O. R.; Abdurahman, N. H.; Ukaegbu, C. I. Soxhlet extraction of phenolic compounds
449 from *Vernonia cinerea* leaves and its antioxidant activity. *J. Appl. Res. Med. Aromat. Plants*
450 **2018**, *11*, 12–17.

- 451 (7) Changtam, C.; Koning, H. P.; Ibrahim, H.; Sajid, M. S.; Gould, M. K.; Suksamrarn, A.
452 Curcuminoid analogs with potent activity against Trypanosoma and Leishmania species. *Eur.*
453 *J. Med. Chem.* **2010**, *45*, 941–956.
- 454 (8) Priyadarsini, K. I.; The chemistry of curcumin: from extraction to therapeutic agent. *Molecules*
455 **2014**, *19*, 20091–20112.
- 456 (9) Climent, M. J.; Corma, A.; Iborra S. Conversion of biomass platform molecules into fuel
457 additives and liquid hydrocarbon fuels. *Green Chem.* **2014**, *16*, 516–547.
- 458 (10) Alonso, D. M.; Bond, J. Q.; Dumesic, J. A. Catalytic conversion of biomass to biofuels. *Green*
459 *Chem.* **2010**, *12*, 1493–1513.
- 460 (11) Sherwood, J.; De bruyn, M.; Constantinou, A.; Moity, L.; McElroy, C. R.; Farmer, T. J.;
461 Duncan, T.; Raverty, W.; Hunt, A. J.; Clark, J. H. Dihydrolevoglucosenone (Cyrene) as a bio-
462 based alternative for dipolar aprotic solvents. *Chem. Commun.* **2014**, *50*, 9650–9652.
- 463 (12) Byrne, F. P.; Forier, B.; Bossaert, G.; Hoebbers, C.; Farmer, T. J.; Clark, J. H.; Hunt, A. J.,
464 2,2,5,5-Tetramethyltetrahydrofuran (TMTHF): a non-polar, non-peroxide forming ether
465 replacement for hazardous hydrocarbon solvents. *Green Chem.* **2017**, *19*, 3671–3678.
- 466 (13) Ruesgas-Ramón, M.; Figueroa-Espinoza, M. C.; Durand, E. Application of Deep Eutectic
467 Solvents (DES) for Phenolic Compounds Extraction: Overview, Challenges, and
468 Opportunities. *J. Agric. Food Chem.* **2017**, *65*, 3591–3601.
- 469 (14) Capello, C.; Fischer, U.; Hungerbühler, K. What is a green solvent? A comprehensive
470 framework for the environmental assessment of solvents. *Green Chem.* **2007**, *9*, 927–934.
- 471 (15) Altunay, N.; Elik, A.; Gükan, R. Preparation and application of alcohol based deep eutectic
472 solvents for extraction of curcumin in food samples prior to its spectrophotometric
473 determination. *Food Chem.* **2020**, *310*, 125933.

- 474 (16) Degot, P.; Huber, V.; Hofmann, E.; Hahn, M.; Touraud, D.; Kunz, W. Solubilization and
475 extraction of curcumin from *Curcuma Longa* using green, sustainable, and food-approved
476 surfactant-free microemulsions. *Food Chem.* **2021**, *336*, 127660.
- 477 (17) Krisanti, E. A.; Saputra K.; Arif, M. M.; Mulia, K. Formulation and characterization of betaine-
478 based deep eutectic solvent for extraction phenolic compound from spent coffee grounds. *AIP*
479 *Conference Proceedings* **2019**, *2175*, 020040.
- 480 (18) Amanolahi, F.; Mohammadi, A.; Kazemi Oskuee, R.; Nassirli, H.; Malaekheh-Nikouei, B. A
481 simple, sensitive and rapid isocratic reversed-phase high-performance liquid chromatography
482 method for determination and stability study of curcumin in pharmaceutical samples. *Avicenna*
483 *J. Phytomedicine* **2017**, *7*, 444–453.
- 484 (19) Barbieria, J. B.; Goltza, C.; Cavalheiroa, F. B.; Tocib, A. T.; Mafraa, L. I.; and Mafraa, M. R.
485 Deep eutectic solvents applied in the extraction and stabilization of rosemary (*Rosmarinus*
486 *ofcinalis L.*) phenolic compounds. *Ind. Crops Prod.* **2020**, *144*, 112049.
- 487 (20) Marcus, Y. The properties of organic liquids that are relevant to their use as solvating solvents.
488 *Chem. Soc. Rev.* **1993**, *22*, 409–416.
- 489 (21) Kumboonma, P.; Senawong, T.; Saenglee, S.; Senawong, G.; Somsakeesit, L.; Yenjai, C. and
490 Phaosiri, C. New histone deacetylase inhibitors and anticancer agents from
491 *Curcuma longa*. *Med. Chem. Res.* **2019**, *28*, 1773–1782.
- 492 (22) Wang, M.; Wang, J.; Zhou, Y.; Zhang, M.; Xia, Q.; Bi, W. and Chen, D. D. Y. Ecofriendly
493 Mechanochemical Extraction of Bioactive Compounds from Plants with Deep Eutectic
494 Solvents. *ACS Sustainable Chem. Eng.* **2017**, *5*, 6297–6303.
- 495 (23) Gu Y. and Jérôme, F. Glycerol as a sustainable solvent for green chemistry. *Green Chem.* **2010**,
496 *12*, 1127–1138.

- 497 (24) Cao, D.; Liu, Q.; Jing, W.; Tian, H.; Yan, H.; Bi, W.; Jiang, Y. and Da Yong Chen, D. Insight
498 into the Deep Eutectic Solvent Extraction Mechanism of Flavonoids from Natural Plant. *ACS*
499 *Sustainable Chem. Eng.* 2020, 8, 19169–19177.
- 500 (25) Dai, Y.; Spronsen, J.; Witkamp, G. J.; Verpoorte, R. and Choi, Y. H. Natural deep eutectic
501 solvents as new potential media for green technology. *Anal. Chim. Acta* **2013**, 766, 61–68.
- 502 (26) Cai, C.; Li, F.; Liu, L. and Tan, Z. Deep eutectic solvents used as the green media for the
503 efficient extraction of caffeine from Chinese dark tea. *Sep. Purif. Technol.* **2019**, 227, 115723.
- 504 (27) Tian, H.; Wang, J.; Li, Y.; Bi, W. and Chen, D. D. Y. Recovery of Natural Products from Deep
505 Eutectic Solvents by Mimicking Denaturation. *ACS Sustainable Chem. Eng.* **2019**, 7,
506 9976–9983.
- 507 (28) Dai, Y.; Witkamp, G. J.; Verpoorte, R. and Choi, Y. H. Tailoring properties of natural deep
508 eutectic solvents with water to facilitate their applications. *Food Chem.* **2015**, 187, 14–19.
- 509 (29) Li, J.; Han, Z.; Zou, Y. and Yu, B. Efficient extraction of major catechins in *Camellia sinensis*
510 leaves using green choline chloride-based deep eutectic solvents. *RSC Adv.* **2015**, 5, 93937–
511 93944.
- 512 (30) Ameer, K.; Shahbaz, H. M. and Kwon, J. H. H. Green extraction methods for polyphenols
513 from plant matrices and their byproducts: A review. *Compr. Rev. Food Sci. Food Saf.* **2017**,
514 16, 295–315.
- 515 (31) Zhang, J.; White, G. B.; Ryan, M. D.; Hunt, A. J. and Katz, M. J. Dihydrolevoglucosenone
516 (Cyrene) As a Green Alternative to *N,N*-Dimethylformamide (DMF) in MOF Synthesis. *ACS*
517 *Sustainable Chem. Eng.* **2016**, 4, 7186–7192.

- 518 (32) Bousfield, T. W.; Pearce, K. P. R.; Nyamini, S. B.; Angelis-Dimakis, A. and Camp, J. E.
519 Synthesis of amides from acid chlorides and amines in the bio-based solvent Cyrene™. *Green*
520 *Chem.* 2019, **21**, 3675–3681.
- 521 (33) De bruyn, M.; Budarin, V. L.; Misefari, A.; Shimizu, S.; Fish, H.; Cockett, M.; Hunt, A. J.;
522 Hofstetter, H.; Weckhuysen, B. M.; Clark, J. H. and Macquarrie, D. J. Geminal Diol of
523 Dihydrolevoglucosenone as a Switchable Hydrotrope: A Continuum of Green Nanostructured
524 Solvents. *ACS Sustainable Chem. Eng.* **2019**, *7*, 7878–7883.
- 525 (34) Byrne, F. P.; Hodds, W. M.; Shimizu, S.; Farmer, T. J. and Hunt, A. J. A comparison of the
526 solvation power of the green solvent 2,2,5,5-tetramethyloxolane versus toluene via partition
527 coefficients. *J Clean Prod* **2019**, *240*, 118175.
- 528 (35) Wang, J.; Jing, W.; Tian, H.; Liu, M.; Yan, H.; Bi, W. and Chen, D. D. Y. Investigation of
529 Deep Eutectic Solvent-Based Microwave-Assisted Extraction and Efficient Recovery of
530 Natural Products. *ACS Sustainable Chem. Eng.* **2020**, *8*, 12080–12088.
- 531 (36) Prat, D.; Wells, A.; Hayler, J.; Sneddon, H.; McElroy, C. R.; Abou-Shehadad, S.; Dunn, P. J.
532 CHEM21 selection guide of classical- and less classical-solvents. *Green Chem.* **2016**, *18*, 288–
533 296.
- 534 (37) Yara-Varon, E.; Fabiano-Tixier, A. S.; Balcells, M.; Canela-Garayoa, R.; Antoine Bily, A.
535 and Chemat, F. Is it possible to substitute hexane with green solvents for extraction of
536 carotenoids? A theoretical versus experimental solubility study. *RSC Adv.* **2016**, *6*, 27750
- 537 (38) Milescu, R. A.; Segatto, M. L.; Stahl, A.; McElroy, C. R.; Farmer, T. J.; Clark, J. H.; Zuin,
538 V. G. Sustainable Single-Stage Solid–Liquid Extraction of Hesperidin and Rutin from Agro-
539 Products Using Cyrene. *ACS Sustainable Chem. Eng.* **2020**, *8*, 18245–18257.

- 540 (39) Ahmadi, R.; Hemmateenejad, B.; Safavi, A.; Shojaeifard, Z.; Mohabbati M. and Firuzi, O.
541 Assessment of Cytotoxicity of Choline Chloride-Based Natural Deep Eutectic Solvents
542 Against Human HEK-293 Cells: a QSAR Analysis. *Chemosphere* **2018**, *209*, 831–838.
- 543 (40) Huber, V.; Muller, L.; Degot, P.; Touraud, D.; Kunz, W. NADES-based surfactant-free
544 microemulsions for solubilization and extraction of curcumin from *Curcuma Longa*. *Food*
545 *Chem.* **2021**, *355*, 129624.
- 546 (41) Claux, O.; Santerre, C.; Abert-Vian, M.; Touboul, D.; Vallet, N.; Chemat, F. Alternative and
547 sustainable solvents for green analytical chemistry. *Curr. Opin. Green Sustain. Chem.* **2021**,
548 *31*, 100510.

549

550 **List of Figures and tables**

551 **Table 1** The choline chloride-based DESs.

552 **Table 2** List of conventional and green solvents and their properties.

553 **Table 3** Dried crude extract of curcuminoids and solubility of curcumin from green and
554 conventional solvents.

555 **Table 4** Classification of selected solvents in this work according to CHEM 21 solvent selection
556 guideline.

557 **Table 5** Process Mass Intensity (PMI) of each solvent for curcuminoid extraction from this work
558 and previous work.

559 **Figure 1** Structures of Curcumin (**1**), Bisdemethoxycurcumin (**2**) and Demethoxycurcumin (**3**).

560 **Figure 2** The extraction yields of curcuminoids (mg/g) from conventional and alternative green
561 solvents, under ultrasound-assisted extraction (UAE) at room temperature for 2 hours, and

562 evaluated by RP-HPLC compared to standard.

563 **Figure 3** Extraction yields of curcuminoids (mg/g) from choline chloride-based deep eutectic
564 solvents, by using ultrasound-assisted extraction (UAE) at room temperature for 2 hours, and
565 evaluated by RP-HPLC.

566 **Figure 4 (a)** (a) The extraction yields of curcuminoids (mg/g) from DES-B5 varying solid-liquid
567 ratio (1:6 to 1:21 ratio), by using ultrasound-assisted extraction (UAE) at room temperature for 2
568 hours, and evaluated by RP-HPLC. (b) The extraction yields of curcuminoids (mg/g) from DES-
569 B5 varying water content (0-40% v/v), by using ultrasound-assisted extraction (UAE) at room
570 temperature for 2 hours, and evaluated by RP-HPLC.

571 **Figure 5** Selectivity of each solvent for three curcuminoids: curcumin, bisdemethoxycurcumin
572 and demethoxycurcumin.

573 **Figure 6** SEM images of plant residues before (A, 500X) and after extraction with Cyrene
574 (B, 500X) and DES-B5 (C, D: 500X).

575 **Figure 7** A proposed hydrogen-bond interaction between DES-B5 and plant cell walls (cellulose,
576 hemicellulose and lignin).

577

578

579

580

581

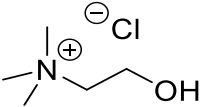
582

583

584

585

586 **Table 1** The choline chloride-based DESs.

| Solvents | Component | | Molar ratio |
|----------|---|-----------------|-------------|
| | HBA | HBD | |
| DES-E1 | Choline chloride | Ethylene glycol | 1:2 |
| DES-E2 |  | Ethylene glycol | 1:3 |
| DES-E3 | | Ethylene glycol | 1:4 |
| DES-E4 | | Ethylene glycol | 1:5 |
| DES-E5 | | Ethylene glycol | 1:6 |
| DES-B1 | | 1,4-Butanediol | 1:2 |
| DES-B2 | 1,4-Butanediol | 1:3 | |
| DES-B3 | 1,4-Butanediol | 1:4 | |
| DES-B4 | 1,4-Butanediol | 1:5 | |
| DES-B5 | 1,4-Butanediol | 1:6 | |
| DES-G1 | Glycerol | 1:2 | |
| DES-G2 | Glycerol | 1:3 | |
| DES-G3 | Glycerol | 1:4 | |
| DES-G4 | Glycerol | 1:5 | |
| DES-G5 | Glycerol | 1:6 | |

587

588

589

590

591

592 **Table 2** List of conventional and green solvents and their properties.

| Solvents | Kamlet-Taft parameters | | | | Boiling point (°C) | Flash point (°C) | Density (g/mL) | M _w (g/mol) |
|---------------|------------------------|--------------------|---------------------|-----------|--------------------------|------------------------|---------------------|---------------------------|
| | α | β | π^* | $E_T(30)$ | | | | |
| Hexane | 0.00 ²⁰ | 0.00 ²⁰ | -0.04 ²⁰ | 31.0 | 69 ¹² | -26 | 0.661 ¹² | 86.2 ¹² |
| Toluene | 0.00 ²⁰ | 0.11 ²⁰ | 0.54 ²⁰ | 33.9 | 111 ¹² | 4 | 0.867 ¹² | 92.1 ¹² |
| DMF | 0.00 ²⁰ | 0.69 ²⁰ | 0.88 ²⁰ | 43.8 | 153 | 58 | 0.944 | 73.09 |
| THF | 0.00 ²⁰ | 0.55 ²⁰ | 0.58 ²⁰ | 37.4 | 66 ¹² | -17 | 0.883 ¹² | 72.11 ¹² |
| Ethyl acetate | 0.00 ²⁰ | 0.45 ²⁰ | 0.55 ²⁰ | 38.1 | 77 | -3 | 0.902 | 88.1 |
| Ethanol | 0.86 ²⁰ | 0.75 ²⁰ | 0.54 ²⁰ | 55.4 | 78 | 14 | 0.789 | 46.1 |
| 2-MeTHF | 0.00 ²⁰ | 0.45 ²⁰ | 0.53 ¹² | 36.5 | 78 ¹² | -10 | 0.854 ¹² | 86.1 ¹² |
| TMO | 0.00 ¹² | 0.77 ¹² | 0.35 ¹² | - | 112 ¹² | 4 | 0.802 ¹² | 128.25 ¹² |
| Cyrene | 0.00 ¹¹ | 0.61 ¹¹ | 0.93 ¹¹ | - | 203 ¹¹ | 108 | 1.25 ¹¹ | 128.13 |

593

594

595

596

597

598

599

600

601

602

603

604 **Table 3** Dried crude extract of curcuminoids and solubility of curcumin from green and
 605 conventional solvents.

| Solvents | Dried crude extract (mg/g) | Total curcuminoids (mg/g) | Solubility of curcumin (mg/mL) |
|-------------------------------|-------------------------------|------------------------------|-----------------------------------|
| Hexane | 25.63 ± 7.42 | 2.66 | 0.003 ± 0.00 |
| Toluene | 52.69 ± 1.64 | 8.22 | 1.75 ± 0.11 |
| DMF | 104.70 ± 12.40 | 32.30 | >300 |
| THF | 84.73 ± 26.24 | 33.04 | >300 |
| Ethyl acetate | 117.01 ± 9.18 | 54.17 | 26.49 ± 0.42 |
| Ethanol | 114.32 ± 3.87 | 56.03 | 7.19 ± 0.28 |
| 2-MeTHF | 76.79 ± 20.17 | 31.77 | 71.79 ± 5.49 |
| TMO | 88.85 ± 8.73 | 21.74 | 2.27 ± 0.07 |
| Cyrene | 124.33 ± 5.13 | 52.85 | 83.23 ± 2.29 |
| DES-B5 (10% H ₂ O) | 100.06 ± 1.57 | 55.86 | 6.55 ± 0.24 |

606

607

608

609

610

611

612

613

614

615 **Table 4** Classification of selected solvents in this work according to CHEM 21 solvent selection
 616 guideline.³⁶

| Solvents | Resource | BP (°C) | FP (°C) | Safety score | Health score | Env. score | Ranking by default | Ranking after discussion |
|-----------------|---------------------------|------------|------------|-----------------|-----------------|---------------|-----------------------|-----------------------------|
| Ethanol | Cereal crop ³⁷ | 78 | 14 | 4 | 3 | 3 | recommended | recommended |
| Ethyl acetate | Cereal crop ³⁷ | 77 | -3 | 5 | 3 | 3 | recommended | recommended |
| Cyrene | Wood | 203 | 108 | 1 | 2 | 7 | problematic | problematic |
| TMO | Biomass ¹² | 112 | 4 | 1 | 5 | 5 | problematic | problematic |
| Dichloromethane | Petroleum | 40 | n.a. | 1 | 7 | 7 | hazardous | hazardous |

| | | | |
|-------------|-------------|-----------|------------------|
| recommended | problematic | hazardous | Highly hazardous |
|-------------|-------------|-----------|------------------|

617

618 *A solvent or chemical was not classified by CHEM 21 toolkit., n.a.: not available.

619

620

621

622

623

624

625

626

627

628

629 **Table 5** Process Mass Intensity (PMI) of each solvent for curcuminoid extraction from this work
 630 and previous work.

| Solvents | Turmeric powder (g) | Weight of solvent (g) | Curcuminoids (g) | PMI total | PMI solvent |
|---|---------------------|-----------------------|----------------------|-----------|-------------|
| Ethyl acetate ^a | 5 | 13.53 | 0.2709 ^b | 68.4 | 50.0 |
| Ethanol ^a | 5 | 11.84 | 0.2802 ^b | 60.1 | 42.2 |
| Cyrene ^a | 5 | 18.75 | 0.2643 ^b | 89.5 | 71.0 |
| DES-B5 (10% H ₂ O) ^a | 5 | 15.13 | 0.2793 ^b | 71.9 | 54.1 |
| TriA/Ethanol/H ₂ O (36/24/40) ¹⁶ | 4 | 16.00 | 0.0611 ^b | 327.2 | 261.8 |
| Ethanol ¹⁶ | 4 | 16.00 | 0.0264 ^b | 757.6 | 606.1 |
| ChCl + Lac/Ethanol/TriA (35/27.5/37.5) ⁴⁰ | 2 | 16.00 | 0.0356 ^b | 505.9 | 449.7 |
| Dichloromethane ²¹ | 600 | 3990 | 21.0438 ^c | 218.1 | 189.6 |

631

632 ^aThis work. ^bA quantity of curcumin based on HPLC. ^cAn amount of curcumin after isolation.

633

634

635

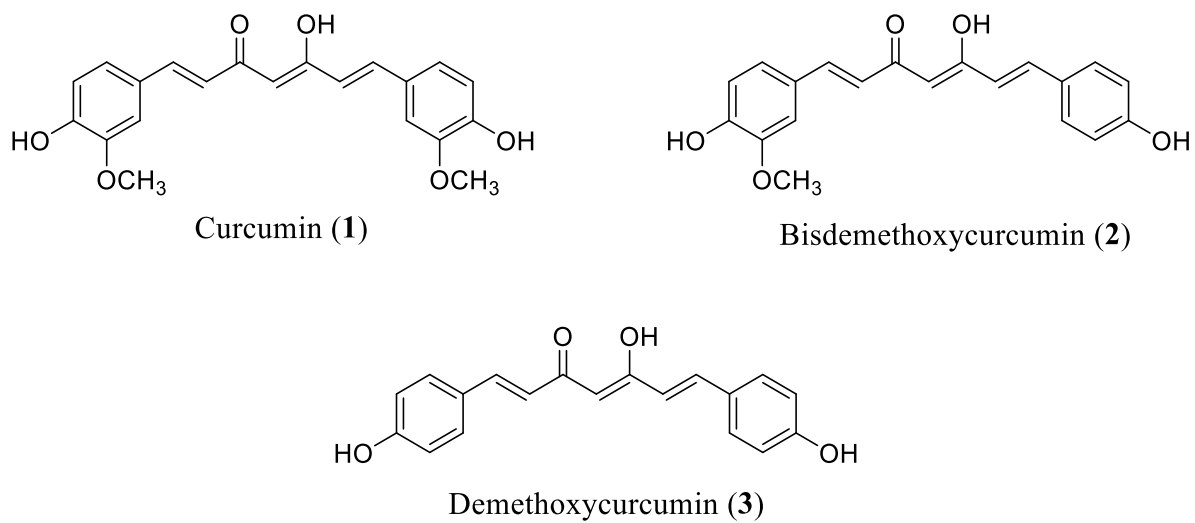
636

637

638

639

640



641

642 **Figure 1** Structures of Curcumin (1), Bisdemethoxycurcumin (2) and Demethoxycurcumin (3).

643

644

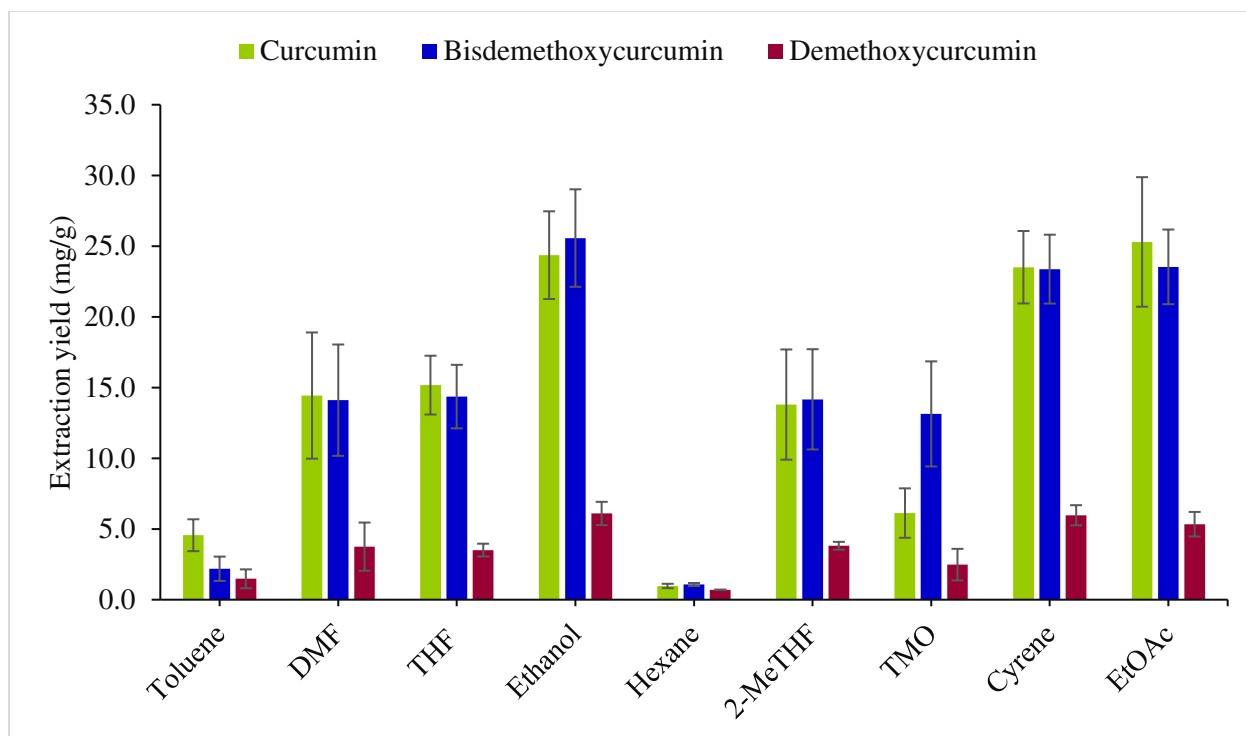
645

646

647

648

649



650

651 **Figure 2** The extraction yields of curcuminoids (mg/g) from conventional and alternative green
652 solvents, under ultrasound-assisted extraction (UAE) at room temperature for 2 hours, and
653 evaluated by RP-HPLC compared to standard.

654

655

656

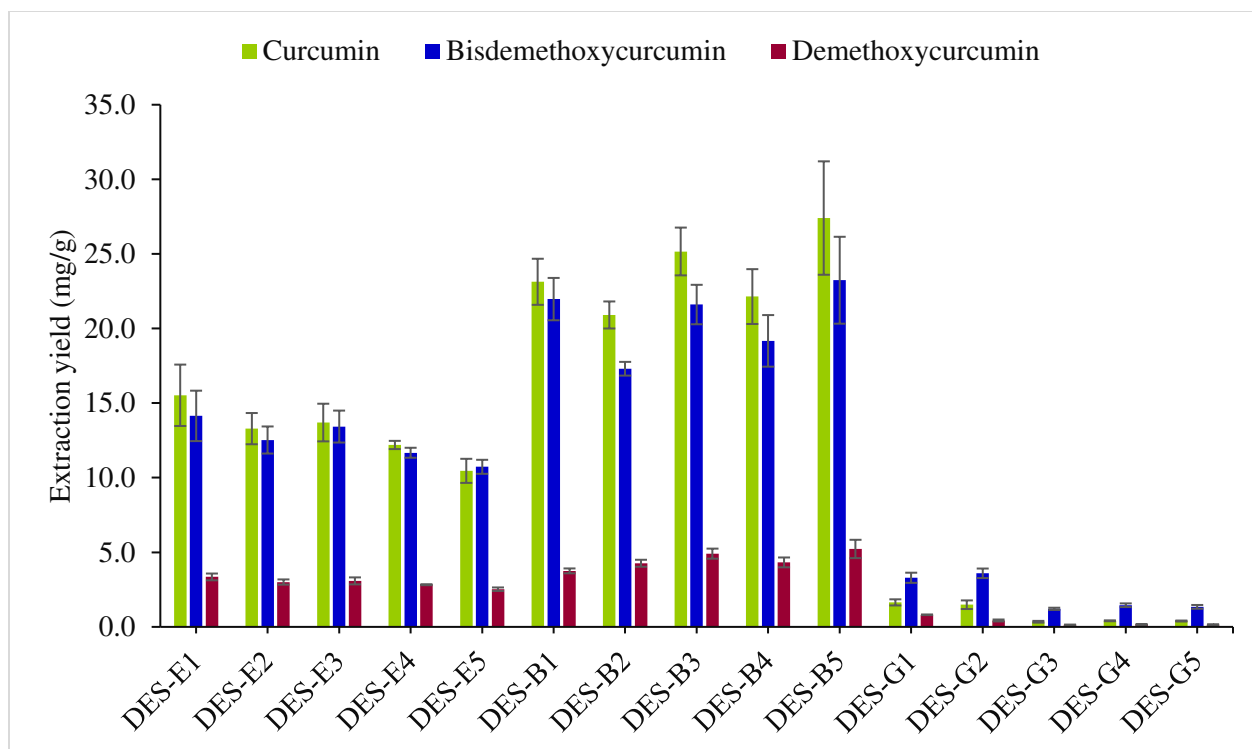
657

658

659

660

661



662

663 **Figure 3** Extraction yields of curcuminoids (mg/g) from choline chloride-based deep eutectic
664 solvents, by using ultrasound-assisted extraction (UAE) at room temperature for 2 hours, and
665 evaluated by RP-HPLC.

666

667

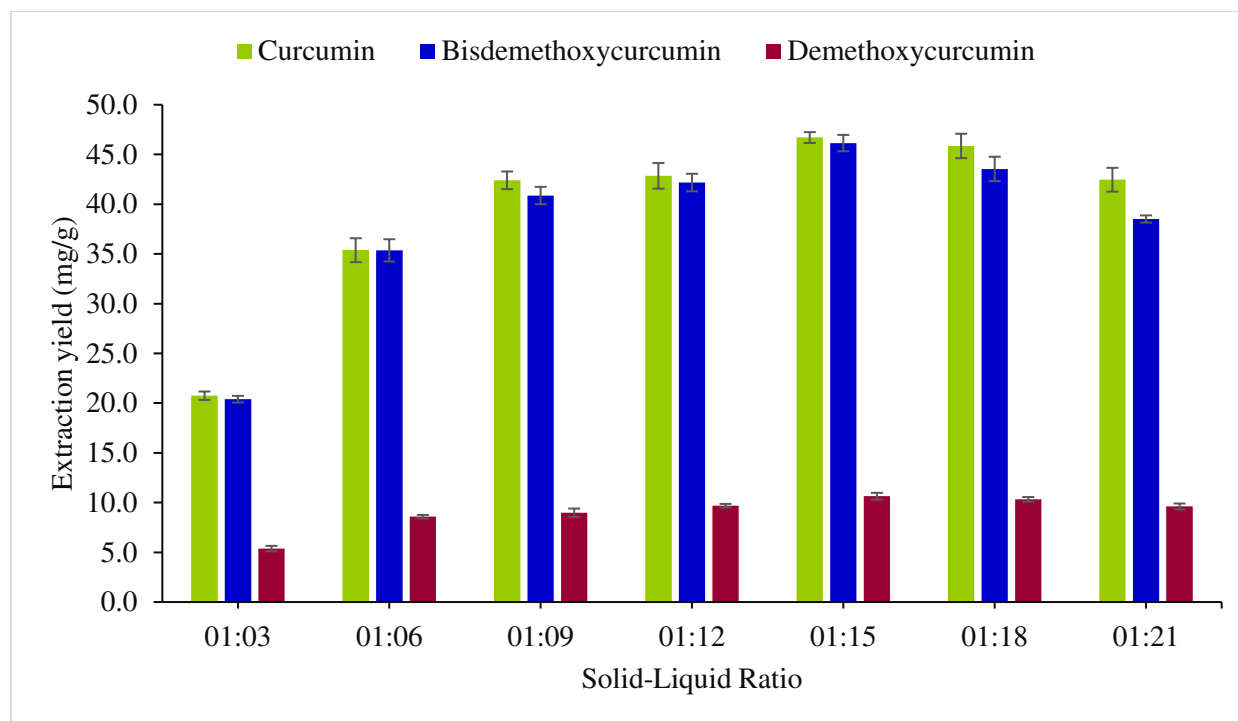
668

669

670

671

672



673

674

675 **Figure 4 (a)** The extraction yields of curcuminoids (mg/g) from DES-B5 varying solid-liquid ratio
676 (1:6 to 1:21 ratio), by using ultrasound-assisted extraction (UAE) at room temperature for 2 hours,
677 and evaluated by RP-HPLC.

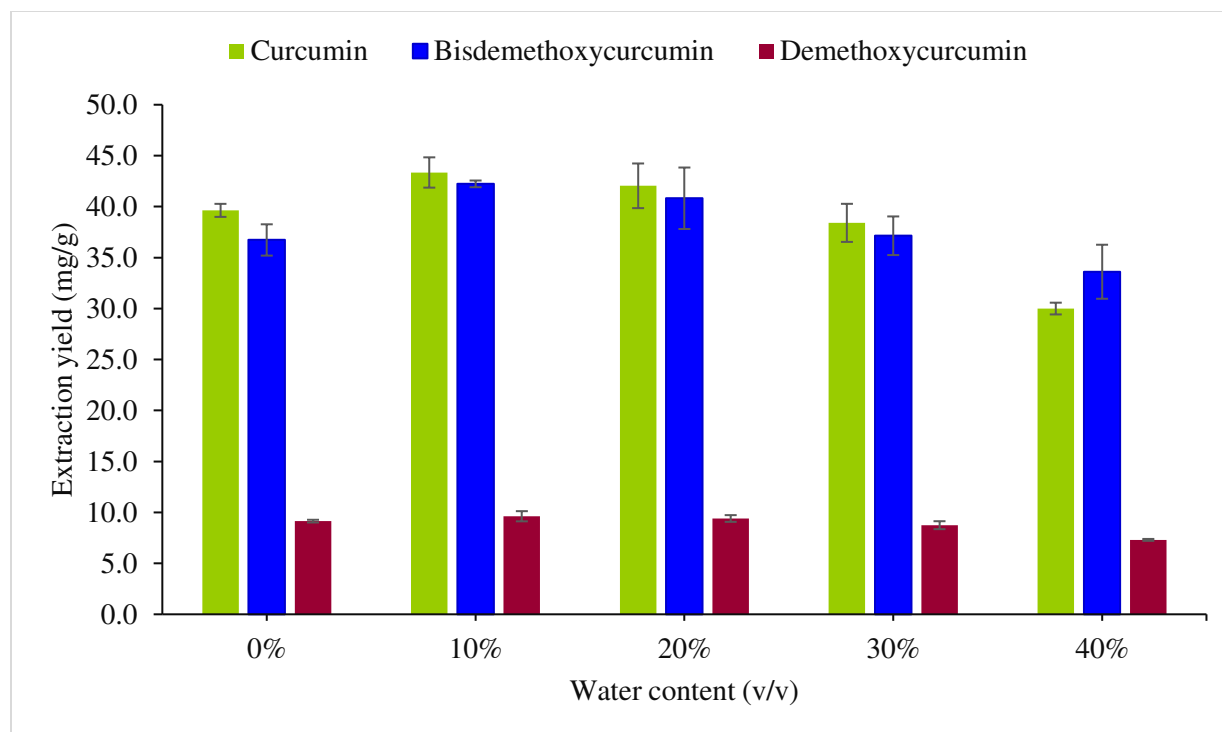
678

679

680

681

682



683

684

685 **Figure 4 (b)** The extraction yields of curcuminoids (mg/g) from DES-B5 varying water content
686 (0-40% v/v), by using ultrasound-assisted extraction (UAE) at room temperature for 2 hours, and
687 evaluated by RP-HPLC.

688

689

690

691

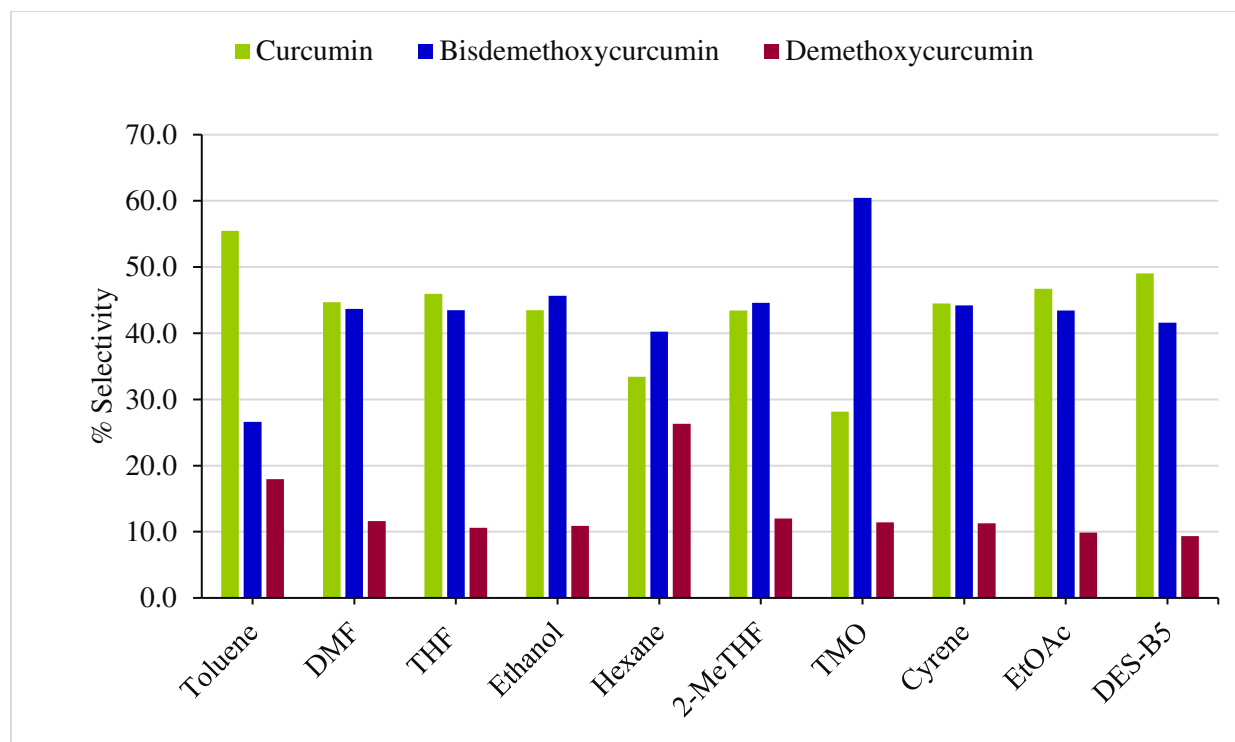
692

693

694

695

696



697

698

699 **Figure 5** Selectivity of each solvent for three curcuminoids: curcumin, bisdemethoxycurcumin

700 and demethoxycurcumin.

701

702

703

704

705

706

707

708

709

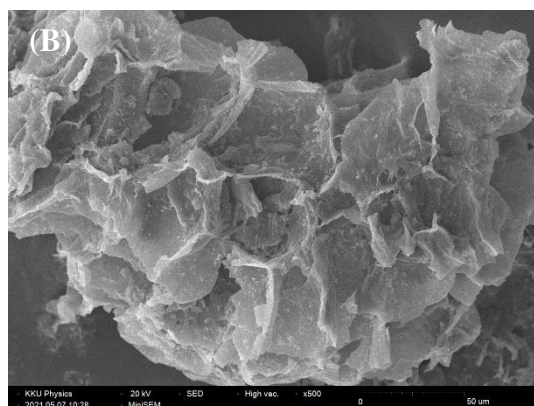
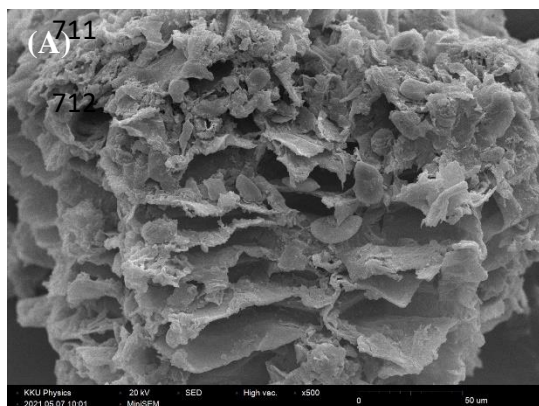
710

713

714

715

716

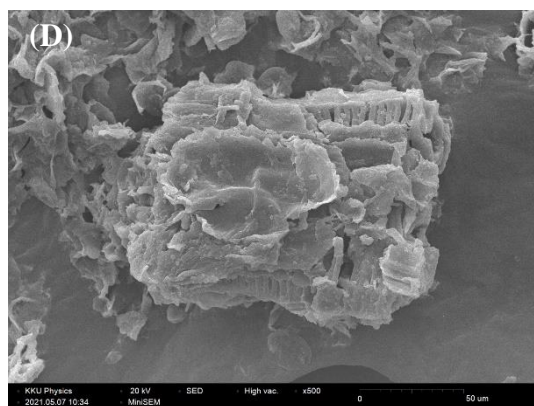
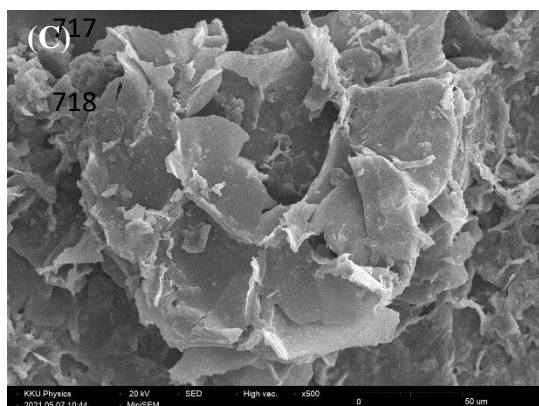


719

720

721

722



723

724 **Figure 6** SEM images of plant residues before (A, 500X) and after extraction with Cyrene

725 (B, 500X) and DES-B5 (C, D: 500X).

726

727

728

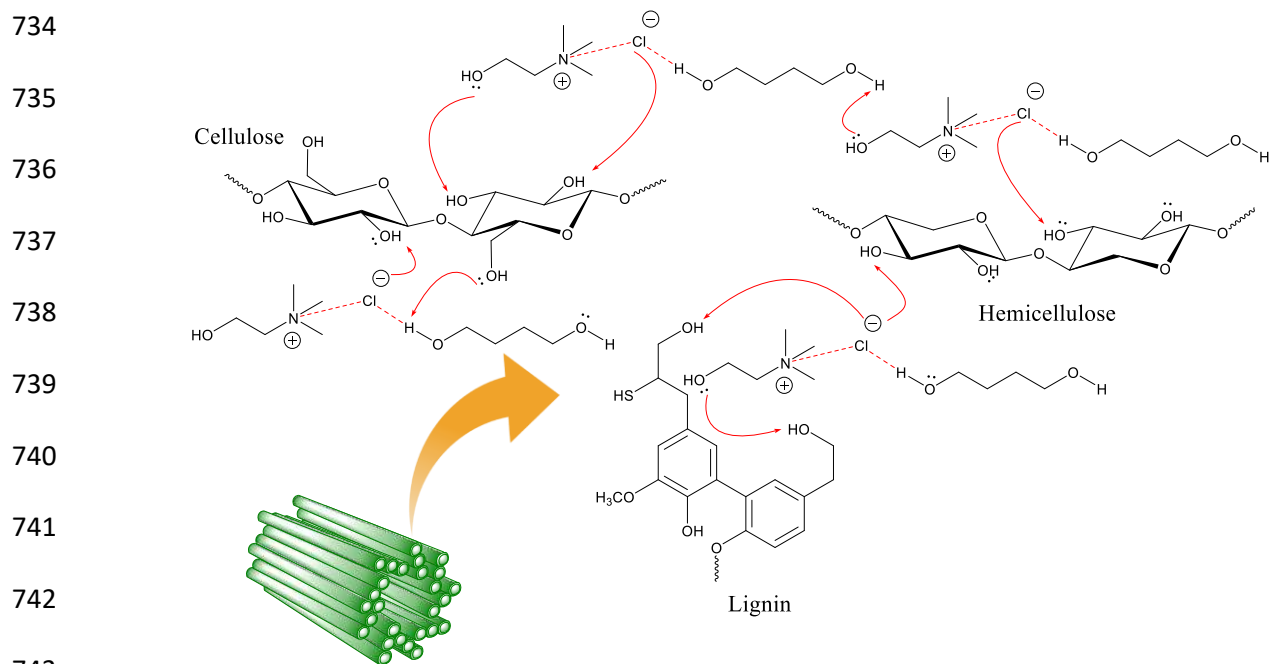
729

730

731

732

733



745 **Figure 7** A proposed hydrogen-bond interaction between DES-B5 and plant cell walls (cellulose,
746 hemicellulose and lignin).

747

748

749

750

751

752

753

754

755

756

757

758

759 **For Table of Contents Use Only**

760

761

762

763

764

765

766



767 Sustainable and effective extraction of curcuminoids from *Curcuma longa* with several alternative

768 green solvents using ultrasound-assisted extraction