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1 A dual-mode sensor for colorimetric and fluorescent detection of
2 nitrite in hams based on carbon dots-neutral red system

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14

15 **Abstract**

16 Nitrite residue in hams was detected by a fluorescent and colorimetric sensor
17 based on carbon dots (C-dots) and neutral red (NR). C-dots with green fluorescence
18 was synthesized by a microwave-assisted method. This novel sensor was fabricated
19 by C-dots as donors and NR as acceptors. The presence of nitrite led to decrease of
20 absorbance and increase of fluorescence. Colorimetric and fluorescent methods for
21 nitrite detection were developed with excellent correlation coefficients ($R^2= 0.995$ and
22 0.991) and low limits of detection (196 nM and 0.518 nM). Moreover, nitrite residue
23 in seven types of ham was detected by the colorimetric and fluorescent methods
24 which were verified by a standard method. The results obtained by the proposed
25 method were comparable and agree with that of the Griess-based method (relative
26 errors < 5%). C-dots-NR system as a sensor has a potential application for nitrite
27 detection in hams to monitor its quality and safety.

28 **Keywords:** Carbon dots, Neutral red, Fluorescence resonance energy transfer, Nitrite,
29 Ham

30

31 **1 Introduction**

32 Ham is a meat product highly consumed in the daily diet. Nitrite is a permitted
33 additive in hams to inhibit the growth of spoilage bacteria, and it can improve
34 recognizable colors and flavors of hams (Addiscott & Benjamin, 2004). High intake
35 of nitrite has an adverse impact on human health for nitrite reacting with amines and
36 amides can form a series of carcinogenic N-nitroso compounds (Honikel, 2008; James,
37 Jayashree, & Alice, 2009). In contrast, modest dietary intake of nitrite can mediate
38 physiological effects including vasodilation, modulation of mitochondrial function,
39 and protection from ischemia-reperfusion injury (Lundberg, Weitzberg, Shiva, &
40 Gladwin, 2011). Control of nitrite quantities is necessary to reduce the potential for
41 N-nitroso compound formation and make nitrite have functions of mediating
42 physiological effects. Therefore, accurate measurement of residual unreacted nitrite is
43 required to guarantee quality and safety of hams.

44 Recently, various methods for nitrite detection have been developed including
45 electrochemistry (Gayathri & Balasubramanian, 1999), chromatography (Li, Yu, Jiang,
46 Zhou, & Liu, 2003;) and optical method (Gayathri & Balasubramanian, 1999; Li, Yu,
47 Jiang, Zhou, & Liu, 2003; Zhen, Xue, Chen, & Lin, 2011). Chromatography and
48 Griess-based method were standard methods for nitrite detection set by International
49 Standardization Organization(ISO, 1975), Association of Official Analytical Chemists
50 (AOAC, 1984), Chinese Ministry of Health(CHM, 2016) and the European
51 Committee for Standardization(CEN, 1999). Chromatographic method is sensitive
52 and reliable in the determination of nitrite. However, it is time-consuming with
53 sophisticated operations and high-cost with expensive instrument. The Griess-based
54 colorimetric method has a widespread application with advantages of simplicity and
55 low cost. It is easily affected by colored pigments that may exist in hams due to
56 relying on the color of azo-compounds. Moreover, it is not sensitive to trace-level
57 nitrite. Development of a new method with high sensitivity, simplicity and low cost
58 for nitrite detection is still a research focus.

59 Fluorescence-based methods have attracted increasing interest because of their

60 high sensitivity, simplicity, and ease of operation. In addition, it possesses high
61 selectivity without interference from color components. Carbon dots (C-dots) as
62 inorganic fluorescent nanomaterials has gained considerable attention due to their
63 excellent fluorescence properties, low toxicity, and low cost (Amjadi & Jalili, 2018; Y.
64 Wang & Hu, 2015; Yang. et al., 2011). Fluorescence resonance energy transform
65 (FRET) systems based on C-dots have been successfully developed for bioimaging
66 (Chatzimarkou et al., 2018), biological sensing (Kudr et al., 2017), and chemical
67 sensing (Wu et al., 2017). They have many advantages, such as fast response, low
68 toxicity and high sensitivity(Liu, Zeng, Liu, & Wu, 2012). Therefore, FRET systems
69 based on C-dots have potentials for nitrite detection.

70 A novel FRET system based on C-dots and NR was developed for nitrite
71 detection in hams. C-dots is synthesized by a microwave-assisted method. C-dots-NR
72 system displaying faint fluorescence is fabricated by C-dots as donors and NR as
73 acceptors for the presence of NR can quench fluorescence emission. Colorimetric and
74 fluorescent methods are developed for nitrite detection based on absorbance and
75 fluorescence signals of C-dots-NR system. Nitrite in hams is detected by this
76 colorimetric and fluorescent methods.

77 **2 Materials and methods**

78 **2.1 Materials and instruments**

79 All chemicals were of analytical grade. Polyethylene glycol 200 (PEG 200) was
80 purchased from Sigma-Aldrich (St. Louis, MO, USA). The remaining reagents
81 (sucrose, sodium nitrite (NaNO_2), sodium chloride, sodium bromide, and so on) were
82 purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). All
83 aqueous solutions were prepared by using ultrapure water obtained from Milli-Q filter
84 system (Millipore Co., USA) with a resistivity of $18.2 \text{ M}\Omega \text{ cm}^{-1}$. Seven types of ham
85 at different levels were purchased from the local supermarket.

86 The size and morphology of C-dots were characterized by JEM-2100
87 transmission electron microscopy (TEM) (JEOL Ltd.) at a maximum accelerating

88 voltage of 200 kV. A 5 μ L of C-dots solution was drop-cast on carbon-coated copper
89 grids and subsequently air-dried before TEM analysis. A portable apparatus composed
90 of several commercially devices was used as ultraviolet-visible (UV-Vis)
91 spectroscopy and fluorescence spectroscopy. It includes two light sources (Ocean
92 optics, Halma Company), a USB 2000 spectrometer (Ocean optics, Halma Company),
93 three fiber optics (Ocean optics, Halma Company), a 1-cm pathlength quartz cuvette
94 (Ocean optics, Halma Company), and a tablet PC equipped with Spectrasuite software.
95 Tungsten light or 365 nm laser light source is successively guided into a cuvette
96 carriage through fiber optics for generation of absorbance or fluorescence signals.
97 These signals are gathered by the micro spectrometer and transferred to the tablet PC
98 for analysis (Fig. S1).

99 **2.2 Detection of nitrite residue in hams using Griess-based colorimetric method**

100 A Griess-based colorimetric method was used to detect nitrite residue (NO_2^-) in
101 seven types of ham according to GB 5009.33-2016 in China(CHM, 2016) and ISO
102 2918-1975(ISO, 1975). The preprocessing procedures for extraction and isolation of
103 nitrite in hams included: (1) 5 g of homogenized ham was macerated with 14 mL
104 borax solution and 100 mL of distilled water (70°C); (2) the mixture was heated in
105 boiling water for 15 min; (3) 5 mL ferrous potassium cyanide and 5 mL zinc acetate
106 solution were subsequently added; (4) the solution was made up to a final volume of
107 200 mL with water followed by filtration through 0.45 μ m micro-filter. The filtrates
108 were stored at 4°C for analysis.

109 Sodium nitrite standard solution or ham extraction was added into the Griess
110 reagent containing sulfanilic acid solution (0.4 mL, 2 g/L) and N-(1-naphthyl)
111 ethylenediamine dihydrochloride solution (0.2 mL, 1 g/L) incubating for 15 min. The
112 concentration of NaNO_2 was 0, 0.02, 0.04, 0.06, 0.08, 0.10, 0.15, 0.20, 0.25 μ g/mL,
113 respectively. Then absorbance spectra of those mixtures were obtained. A linear
114 calibration curve was established by plotting absorbance values at 538 nm to the
115 corresponding NaNO_2 concentrations (0- 0.25 μ g/mL). The absorbance values of ham

116 extraction at 538 nm were recorded and the nitrite concentration in ham was detected
117 according to equation (1). All measurements were performed in triplicate on different
118 days.

$$119 \quad Q = \frac{(A_{538} - 0.0098) \times 5 \times 200}{6.076 \times 1.371 \times 5} \text{ (mg/kg)} \quad (1)$$

120 Where Q is the nitrite residue in hams, A_{538} represents the absorbance value at 538
121 nm.

122 **2.3 Development of colorimetric and fluorescent methods for nitrite detection**

123 **2.3.1 Synthesis of C-dots**

124 Synthesis of C-dots was according to a microwave-assisted method with minor
125 modifications (Gong et al., 2014). Sucrose solution (1 mL, 30% (w/v)) and
126 concentrated H_2SO_4 (200 μ L) were sequentially added to 6 mL of PEG in a 10 mL
127 glass tube. The mixture was heated in a 900W domestic microwave oven (Midea,
128 China) for 15 seconds. It turned golden yellow indicating the formation of C-dots. The
129 C-dots solution was centrifuged to remove the insoluble substance, and small
130 molecules were removed using a dialysis membrane (molecular weight cut-off =1000)
131 for 24 h. The resultant C-dots was diluted into 100 mL and stored at 4°C for further
132 characterization and utilization.

133 **2.3.2 Optimization of experimental conditions**

134 The concentration of C-dots, concentration of NR, pH and reaction time were
135 optimized to achieve the maximum sensitivity for nitrite detection. Different volumes
136 of C-dots (1, 2, 3 and 4 mL) were analyzed to select the optimal volume of C-dots.
137 C-dots (1 mL) mixing with different amounts of NR (0, 40, 80, 120, 160, 200 and 300
138 μ L) was used for selecting the optimal amount of NR. Different pH values (0.9, 1.0,
139 1.2, 1.7, 2.0, 3.0, 4.0, 5.0, and 6.0) were investigated in presence of 0.10 μ g/mL nitrite,
140 1mL C-dots and 160 μ L NR. The mixture (1 mL C-dots, 160 μ L NR, 100 μ L HCl and
141 0.10 μ g/mL nitrite) was incubated for 5, 10, 15 and 20 min to select the optimal
142 reaction time. All solutions were diluted into 5 mL with water before spectroscopy
143 measurements.

144 **2.3.3 Colorimetric and fluorescent methods for nitrite detection**

145 Colorimetric and fluorescent methods were developed for nitrite detection. NR
146 and of C-dots were mixed to form C-dots-NR system. Then NaNO₂ solution and HCl
147 were added into the C-dots-NR system followed by diluted to 5.0 mL. The final
148 concentration of NaNO₂ were 0, 0.005, 0.01, 0.015, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07,
149 0.08, 0.1, 0.12, 0.015, 0.2, 0.25, and 0.3 µg/mL, respectively. Absorbance and
150 fluorescence spectra were acquired as an average of three independent measurements.
151 Absorbance values and fluorescence intensities were recorded to establish linear
152 calibration curves. The response of C-dots-NR system to other ions was also studied
153 to validate the specificity for NO₂⁻ detection. All experiments were performed in
154 triplicate on different days.

155 **2.4 Colorimetric and fluorescent detection of nitrite residue in ham**

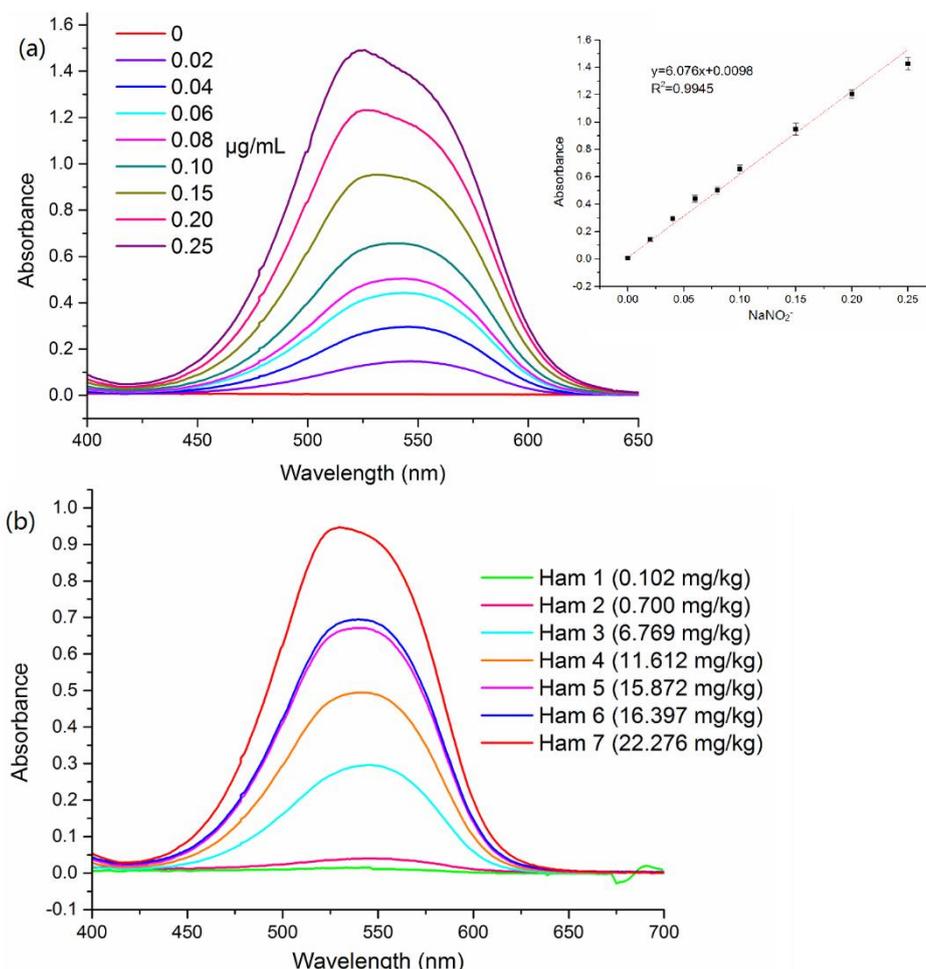
156 Nitrite residue in seven types of ham was detected by the colorimetric and
157 fluorescent methods based on C-dots-NR system according to the Section 2.3.3. The
158 unknown concentration was calculated by putting fluorescent intensity or absorbance
159 value into the corresponding calibration curve. All experiments were performed in
160 triplicate on different days (sample size is 12 for each ham).

161 **3 Results and discussion**

162 **3.1 Nitrite in hams detected by the Griess-based method**

163 Nitrite residue (NO₂⁻) levels in hams detected by Griess-based colorimetric
164 method were between 0.102 and 22.276 mg/kg (Fig. 1). Absorbance values enhanced
165 with increasing concentration of NO₂⁻ (Fig. 1a). Excellent linearity between nitrite
166 concentrations (0-0.25 µg/L) and absorbance values at 538 nm was observed with
167 correlation coefficient of 0.995 (inset in Fig. 1a). The linear equation was $y =$
168 $(6.076 \pm 0.015) x + (0.010 \pm 0.023)$ (x and y represented the NaNO₂ concentration
169 (µg/mL) and absorbance value at 538 nm, respectively). The limit of detection (LOD,
170 3s) was calculated to be 15.8 µg/L (229 nM) according to the International Union of
171 Pure and Applied Chemistry (IUPAC). The average absorbance spectra of Griess

172 reagent in presence of each ham extraction were shown in Fig. 1b. The NO_2^- levels in
 173 seven types of ham were 0.102, 0.700, 6.769, 11.612, 15.872, 16.397, and 22.276
 174 mg/kg by plugging absorbance value at 538 nm into the linear equation (1).



175
 176 Fig.1 (a) absorbance spectra of Griess reagent in presence of NaNO_2 with various
 177 concentrations (inset: A calibration curve between NaNO_2 concentrations and
 178 absorbance values at 538 nm); (b) absorbance spectra of Griess reagent in presence of
 179 ham extractions.

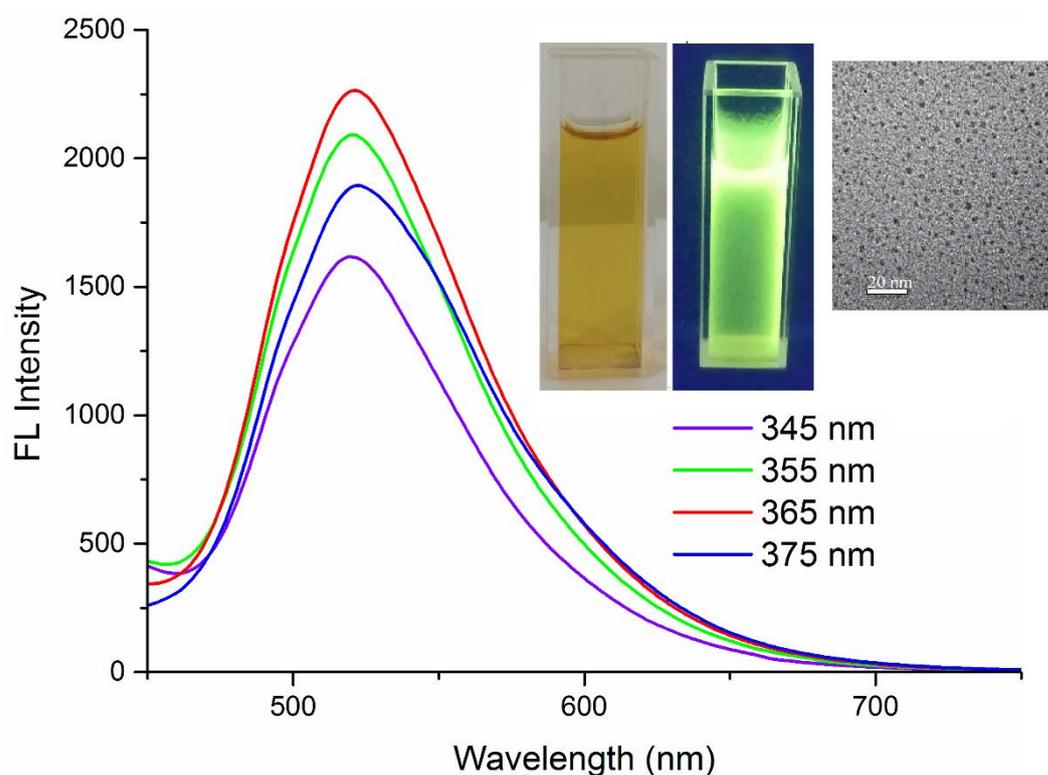
180 3.2 Development of colorimetric and fluorescent methods for nitrite detection

181 3.2.1 Characterization of C-dots

182 C-dots prepared in this paper displayed excellent fluorescence properties and
 183 favorable dispersibility (Fig. 2). It was synthesized using a one-step method
 184 (microwave-assisted method) with sucrose as a carbon source and PEG 200 as a
 185 passivating agent(Yang et al., 2013). C-dots emitted bright green fluorescence after

186 being irradiated by UV light at 365 nm (inset in Fig. 2). The fluorescent intensity at
187 520 nm achieved the highest when the C-dots was irradiated at 365 nm (Fig. 2). TEM
188 image of C-dots displayed that C-dots were well-dispersed with spherical shape (inset
189 in Fig. 2). The average diameter of C-dots was 5.3 nm. Those results are consistent
190 with observation reported previously (Gong et al., 2014).

191 C-dots is selected as a fluorescent material to analyze NO_2^- for its unique
192 characteristics (e.g., low cost, simple preparation, excellent dispersibility, favorable
193 biocompatibility, and easy modification). It is synthesized from common carbon
194 sources, such as carbohydrate, natural gas soot and candle soot (Egorova, Tomskaya,
195 An, & Sa, 2017). These raw materials are easily acquired and have no toxicity. The
196 synthesis of C-dots in this paper is fast and simple including: (1) preparation of
197 fluorescent C-dots by heating mixture of sucrose and PEG for 15 seconds in
198 microwave and (2) purification of C-dots by centrifugation and dialysis. Hydroxyl,
199 carbonyl, and carboxylic acid groups on surfaces endow C-dots with excellent
200 water-solubility, favorable biocompatibility and easy modifications (Y. Wang & Hu,
201 2015). All characteristics are conducive to analysis of targets.



202

203 Fig. 2 fluorescent spectra of C-dots excited at 345, 355, 365, and 375 nm (inset (from
204 left to right): image of C-dots under natural light (left); fluorescent image of C-dots
205 under UV light at 365 nm (middle); and TEM image of C-dots (right)).

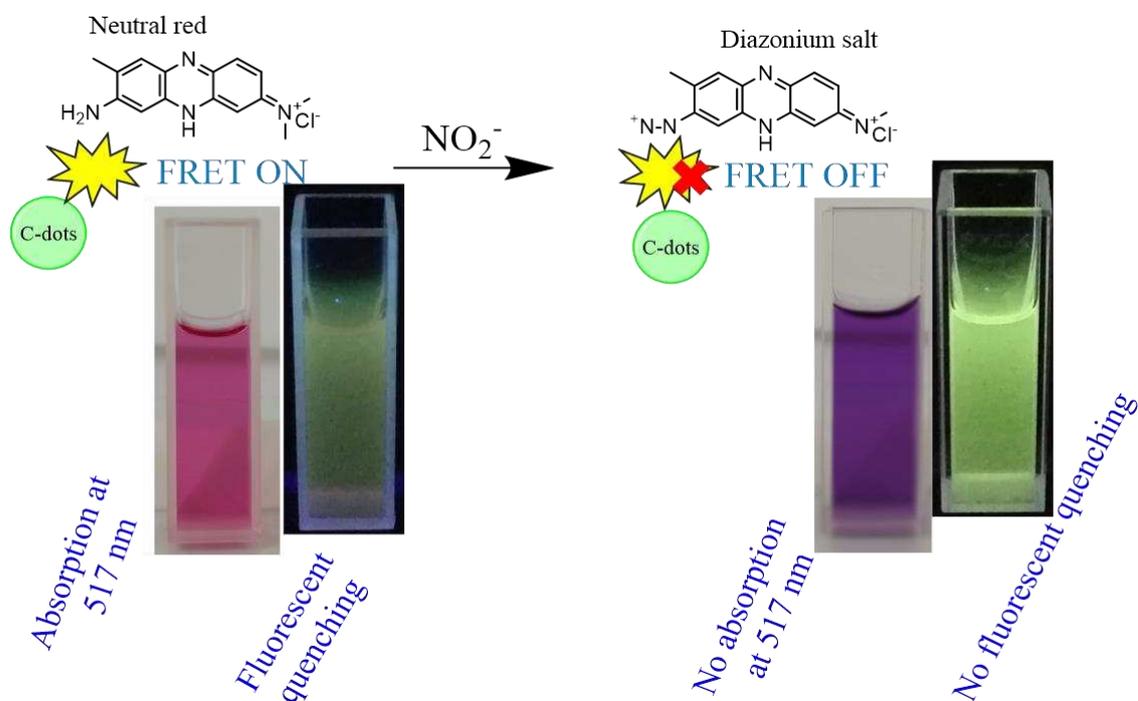
206 **3.2.2 Mechanism of colorimetric and fluorescent detection of nitrite**

207 C-dots-NR system has colorimetric and fluorescent responses to nitrite (Fig. 3).
208 C-dots-NR system showed red color under natural light and faint green fluorescence
209 under UV light of 365 nm. It turned blue in the presence of NO_2^- and the absorbance
210 was decreased. Therefore, C-dots-NR system could be developed as a colorimetric
211 sensor for nitrite detection, where NR acted as the receptor unit and chromogenic
212 agent. Simultaneously, the fluorescent emission of this system recovered by blocking
213 off FRET process with addition of NO_2^- . C-dots-NR also could be designed as a
214 fluorescent sensor for nitrite detection, where NR acted as the receptor unit and
215 C-dots acted as the fluorophore. The nitrite is residual unreacted and
216 free (AbbasAfkhani & AbbasAliMogharnesband, 1994; Li et al., 2003). Some
217 reaction products (nitroso compounds and diazonium salts) were synthesized from
218 free nitrite and amines (or amides). It cannot interact with neutral red and interfere the
219 detection (AbbasAfkhani & AbbasAliMogharnesband, 1994; Gayathri &
220 Balasubramanian, 1999; Han et al., 2014). Therefore, colorimetric and fluorescent
221 detection of nitrite is realized by the C-dots-NR system.

222 Fluorescence and absorbance spectra of NR and diazonium salts displayed that
223 NR had a sensitive response to nitrite (Fig. S2a and b). NR solution with red color
224 (Fig. S2a) has maximal absorbance band at 517 nm (a red line in Fig. S2b). NR can be
225 oxidized in presence of trace amounts of nitrite, forming diazonium salts which
226 exhibit blue color with a maximal absorbance band at 631 nm (a blue line in Fig. S2b).
227 The absorbance of NR at 517 nm was decreased greatly with addition of nitrite. NR
228 has been used to determine nitrite in previous researches (Gayathri &
229 Balasubramanian, 1999).

230 NR quenched fluorescence of C-dots effectively via FRET process based on NR

231 as acceptors and C-dots as donors (Fig. S2c). Fluorescent intensity of C-dots at 520
 232 nm was decreased with addition of NR. This observation was in agreement with these
 233 fluorescent images (inset in Fig. S2c). Nitrite, diazonium salts and nitrosative species
 234 had no impact on C-dots for the fluorescence spectra remained unchanged in the
 235 presence of nitrite, diazonium salts (reaction products of nitrite and neutral red) and
 236 N-Nitrosodimethylamine (Fig. S2c). As shown in Fig. S2b, the UV-vis absorbance
 237 peak of NR (red line) and fluorescence emission peak of C-dots (green line) were at
 238 517 and 520 nm, respectively. The absorbance spectrum of NR almost exactly
 239 overlapped with the emission spectrum of C-dots. The principle of fluorescence
 240 resonance energy transfer (FRET) is that absorption spectrum of acceptor should
 241 overlap with emission spectrum of donor and the distance between them must be
 242 linked in close proximity (Tang, Du, & Su, 2013). Therefore, an efficient FRET system
 243 could be established with NR as acceptors and C-dots as donors because of their
 244 overlaps and close distance. The FRET efficiency was estimated by $E=1-I/I_0$, where I
 245 and I_0 were fluorescent intensity of C-dots in presence of NR and C-dots alone (no
 246 NR), respectively. The quenching efficiency exceeded 77% by adding 160 μL of NR.



247

248 Fig. 3 illustration of the mechanism for nitrite detection using C-dots-NR system

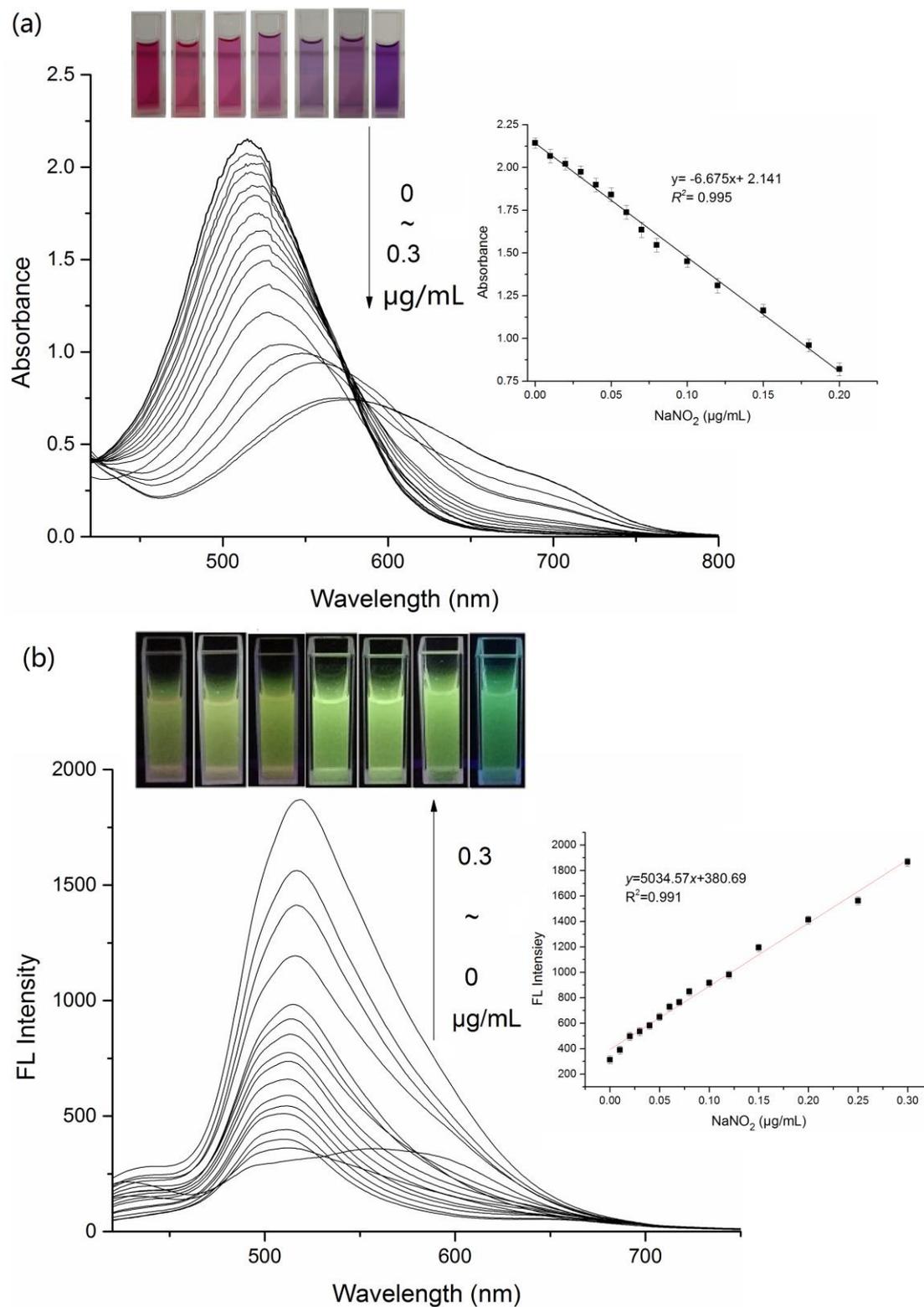
249 3.2.3 Colorimetric and fluorescent detection of nitrite

250 C-dots(1 mL), NR (160 μ L), HCl (100 μ L) and reaction time (10 minutes) were
251 selected as the optimal reaction parameters (Fig. S3). One milliliter of C-dots had
252 strong fluorescent emission (Fig. S3a) and their fluorescent emission was almost
253 quenched by adding 160 μ L of NR (Fig. S3b). The ratio of fluorescent intensity (F/F_0)
254 reached the maximum at pH value of 1.2 indicating that the reaction was the fastest
255 (Fig. S3c). The fluorescence ratio (F/F_0) remained stable after 10 min, indicating that
256 the reaction was almost completed (Fig. S3d).

257 Under these optimal assay conditions, C-dots-NR system was applied
258 successfully for colorimetric detection of NO_2^- (Fig. 4a). The absorbance spectrum of
259 C-dots-NR system was centered at 517 nm, which displayed in red color. The color
260 changed from red to blue with addition of NO_2^- , which can be recognized by the
261 naked eye (inset in Fig. 4a). Absorbance values at 517 nm decreased gradually and a
262 new absorbance peak at 575 nm raised with wavelength shift of 58 nm. There was a
263 good linear correlation ($R^2= 0.995$) between absorbance intensity and NaNO_2
264 concentration in the range of 0- 0.2 $\mu\text{g/mL}$ (0- 2.9 μM) (inset in Fig. 4a). The
265 associated linear regression equation was $y= (-6.367\pm 0.092) x+ (2.141\pm 0.039)$ (x and
266 y represented the NaNO_2 concentration ($\mu\text{g/mL}$) and absorbance value at 517 nm,
267 respectively). The limit of detection for NaNO_2 was approximately 13.5 $\mu\text{g/L}$ (196
268 nM) at a 3:1 signal-to-noise (S/N) ratio, which was lower than the maximum
269 concentration limit of nitrite in drinking water (about 1 μM) (Keeton, 2017).

270 Simultaneously, fluorescent detection of NO_2^- was performed successfully using
271 C-dots-NR system (Fig. 4b). Fluorescence intensities of C-dots-NR system at 520 nm
272 enhanced gradually with increasing NO_2^- concentrations (Fig. 4b). The fluorescence
273 recovered with addition of nitrite which was attributed to decrease of NR and
274 reduction of FRET efficiency (inset in Fig. 4b). There was an excellent linear
275 correlation (0.991) between fluorescent intensities at 520 nm and NaNO_2
276 concentrations in the range of 0- 0.3 $\mu\text{g/mL}$ (0- 4.34 μM) (inset in Fig. 4b). The

277 associated linear regression equation was $y = (5034.570 \pm 2.597) x + (380.690 \pm 1.743)$ (x
278 and y represented the NaNO_2 concentration ($\mu\text{g/mL}$) and absorbance value at 520 nm,
279 respectively). Limit of detection for NaNO_2 detection was calculated to be 35.75 ng/L
280 (0.518 nM), which was lower than most methods reported to nitrite detection (Table 1)
281 (Abbas & Mostafa, 2000; Büldt & Karst, 1999; Jedličková, Paluch, & Alušík, 2002;
282 Li et al., 2003; Losada, Armada, García, Casado, & Alonso, 2017; Zheng, Liang, Li,
283 Zhang, & Qiu, 2016).



284

285 Fig. 4 (a) absorbance spectra of C-dots-NR system after addition of NaNO₂ with
 286 different concentrations (0, 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.1, 0.12,
 287 0.15, 0.18, 0.2, 0.25, 0.3 μg/mL) (inset: photographs of C-dots-NR system with
 288 different concentrations of NaNO₂ and a calibration curve established by absorbance

289 values at 517 nm and NaNO₂ concentrations (0, 0.01, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07,
290 0.08, 0.1, 0.12, 0.15, 0.18, 0.2 µg/mL)); (b) fluorescence spectra of C-dots-NR system
291 with addition of NaNO₂ with various concentrations (0, 0.005, 0.01, 0.015, 0.02, 0.03,
292 0.04, 0.05, 0.06, 0.07, 0.08, 0.1, 0.12, 0.015, 0.2, 0.25, 0.3 µg/mL) (inset: photographs
293 of C-dots-NR system with addition of NaNO₂ under UV light at 365 nm and a
294 calibration curve established by fluorescence intensities at 520 nm and NaNO₂
295 concentrations(0, 0.005, 0.01, 0.015, 0.02, 0.03, 0.04, 0.05, 0.06, 0.07, 0.08, 0.1, 0.12,
296 0.015, 0.2, 0.25, 0.3 µg/mL)).

297 The detectable concentration range based on fluorescence signal (0.072- 4.34 µM)
298 was wider than that of absorbance signal (0.145- 2.9 µM); the LOD of fluorescence
299 signal (0.518 nM) was much lower than that of absorbance signal (196 nM); the
300 relationship coefficient of absorbance signal ($R^2= 0.995$) was higher than that of
301 fluorescence signal ($R^2= 0.991$). Both absorbance and fluorescent signals were
302 successfully used for nitrite detection and they had their own unique characteristics.
303 Therefore, signal fusion of fluorescence and absorbance was performed in this study.
304 Binary regression equation y (µg/mL) $= (0.188 \pm 0.024) - (0.098 \pm 0.006) x_1 +$
305 $(6.180 \pm 0.037) \times 10^{-5} x_2$ ($R^2= 0.995$, x_1 and x_2 were the absorbance value at 517 nm and
306 the fluorescence intensity at 520 nm, respectively, y was the NaNO₂ concentration
307 (µg/mL)) was obtained by combining absorbance and fluorescence signal. The
308 correlation coefficient based on dual signals was higher than that based on
309 colorimetric signals and was equal to that based on fluorescent signals.

310 Cu nanoclusters(Zheng et al., 2016), Au nanoclusters(H. Liu, Yang, Abdel-Halim,
311 & Zhu, 2013), CdSe quantum dots(X. Liu, Guo, Cheng, & Ju, 2009; B. Wang, Huang,
312 Ma, Shi, & Cai, 2014) and upconversion nanoparticles (UCNPs) (Han et al., 2014)
313 have been successfully used for determination of NO₂⁻. It is simple and sensitive to
314 detect nitrite based on these nanosensors. However, CdSe quantum dots are
315 synthesized from semiconductors containing heavy metals (Cd) and their applications
316 are subject to their toxicity and potential hazard to the environment (Guo et al., 2013;

317 Zhen et al., 2011). Cu nanoclusters and Au nanoclusters have low water-solubility and
 318 are harmful to the environment as their preparations are based on heavy metals (Cu,
 319 Au). UCNPs doped with certain rare-earth ions need complex procedures to convert
 320 hydrophobic form to hydrophilic variants for making them water-soluble. In addition,
 321 the rare earth elements are scarce (Han et al., 2014). Compared with other methods,
 322 the proposed method exhibits superiority in nitrite detection.

323 Table 1 the results of NO₂⁻ detection reported by other methods

Technique	Linear range (μM)	LOD(nM)	Reference
Neutral red (colorimetry)	0.87- 4.35	176.1	(Li, Yu, Jiang, Zhou, & Liu, 2003)
Griess reagent (colorimetry)	0.22- 48	20	(Abbas & Mostafa, 2000)
HPLC-UV-vis ^a (chromatography)	0- 20	100	(Jedličková, Paluch, & Alušík, 2002)
HPLC-FL ^b (chromatography)	0.215- 7.17	60	(Büldt & Karst, 1999)
Amperometry (electrochemistry)	0.005- 10	0.02	(Losada, Armada, García, Casado, & Alonso, 2017)
Cu nanoclusters (fluorometry)	0.0125- 125	3.6	(Zheng, Liang, Li, Zhang, & Qiu, 2016)
C-dots-NR system (colorimetry and fluorometry)	0.145- 4.34 (colorimetry) 0.072- 4.34 (fluorometry)	196 0.518	This work

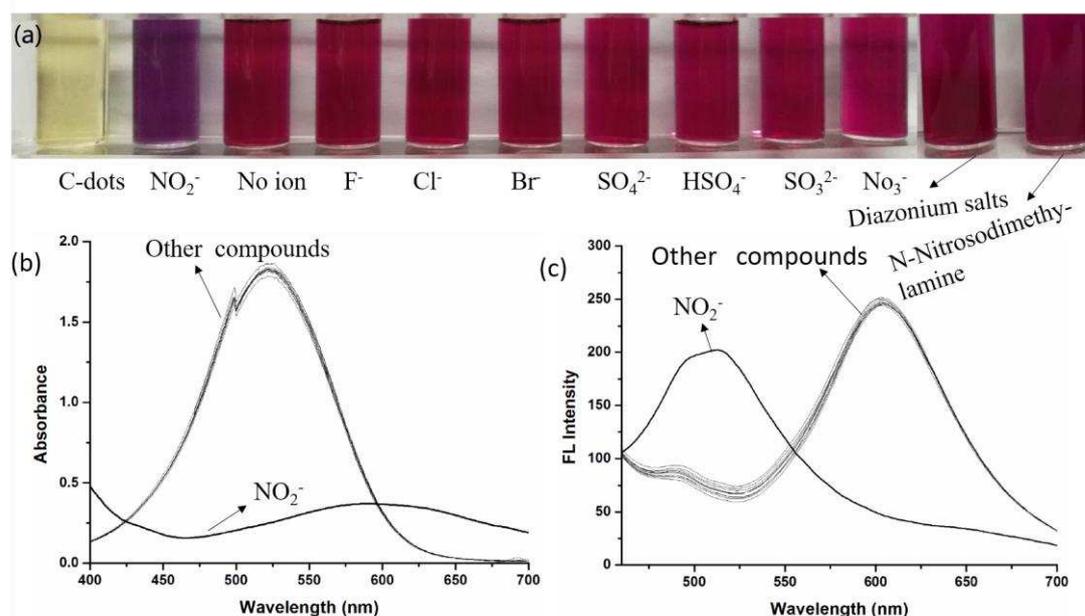
324 ^a High performance liquid chromatography with UV-vis absorbance detection

325 ^b High performance liquid chromatography with fluorescence detection

326 3.2.4 Selectivity of C-dots-NR system for nitrite detection

327 The C-dots-NR system as a colorimetric and fluorescent sensor displayed superb
 328 selectivity in NO₂⁻ detection (Fig. 5). The presence of NO₂⁻ led to significant color
 329 change (from red to blue), absorbance reduction at 517 nm (Fig. 5a and b). The
 330 fluorescence emission of 520 nm recovered by adding NO₂⁻ (Fig. 5c). Potentially
 331 competing compounds (F⁻, Cl⁻, Br⁻, HSO₄⁻, SO₄²⁻, SO₃²⁻, NO₃⁻, dianoium salts
 332 (reaction products of nitrite and neutral red), and N-Nitrosodimethylamine) were

333 added into the C-dots-NR system under identical conditions. Absorbance at 517 nm
 334 and fluorescent intensity at 520 nm showed no change indicating that those
 335 compounds did not interfere with nitrite detection (Fig. 5b and c). The high selectivity
 336 of NO_2^- detection using this sensor may be attributed to the unique reaction between
 337 NO_2^- and NR.



338
 339 Fig. 5 images (a), absorbance spectra (b), and fluorescent spectra (c) of C-dots-NR
 340 system in presence of NO_2^- , F^- , Cl^- , Br^- , HSO_4^- , SO_4^{2-} , SO_3^{2-} , NO_3^- , diazonium salts
 341 and N-Nitrosodimethylamine.

342 3.3 Colorimetric and fluorescent detection of nitrite in hams

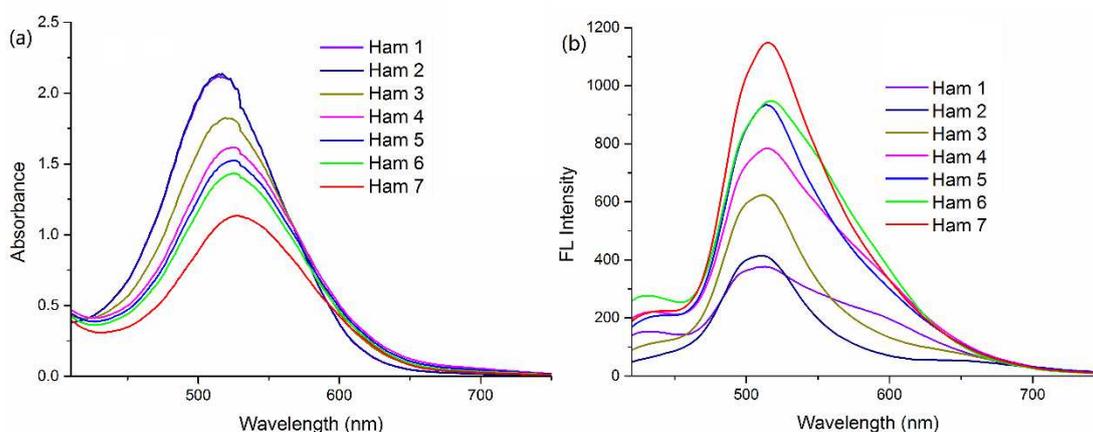
343 Nitrite in the seven types of ham was detected by the proposed method (Fig. 6)
 344 and the performance was evaluated by the Griess-based colorimetric method (Table 2).
 345 Absorbance spectra of C-dots-NR system in presence of ham extractions were shown
 346 in Fig. 6a. Based on the absorbance values at 517 nm (A) and linear correlation ($Q=$
 347 $(2.141- A)/6.675$), the concentrations of NO_2^- in hams were 0.098, 0.677, 7.059,
 348 12.020, 16.205, 17.189 and 22.969 mg/kg, respectively. Relative errors were between
 349 2.098% and 4.830% compared with results detected by the Griess-based method. The
 350 results demonstrate that the NO_2^- levels predicted by absorbance signal are in
 351 accordance with that detected by the standard method. Fluorescent spectra of

352 C-dots-NR system in presence of these ham extractions were presented in Fig. 6b.
353 Based on fluorescent intensities at 520 nm (F) and the linear correlation ($Q = (F -$
354 $380.69) / 5034.57$), the concentrations of NO_2^- were 0.099, 0.676, 6.714, 11.628,
355 15.899, 16.429 and 22.190 mg/kg, respectively. Relative errors (0.138-3.429%)
356 indicate that the NO_2^- concentration levels predicted by fluorescent signal are in
357 agreement with that detected by Griess-based method perfectly. The nitrite content in
358 hams were 0.087, 0.850, 6.921, 11.709, 15.788, 16.599 and 22.276 mg/kg based on
359 absorbance values and fluorescence intensities combined with binary regression
360 equation ($Q = 0.188 - 0.098 A + 6.180E-5 F$), respectively. Relative errors
361 (0.000-21.429%) are not as good as those obtained by colorimetric or fluorescent
362 method.

363 Compared with these results of nitrite detection in seven types of ham, the
364 fluorescent detection results were more excellent than colorimetric detection results
365 and dual-signal detection results. It may be ascribed to that pigments in hams have a
366 negative impact on colorimetric detection results. All results, especially the
367 fluorescent detection results indicate that C-dots-NR system can be utilized to realize
368 simple and sensitive detection of nitrite in hams.

369 The difference between this proposed method and Griess-based method was
370 investigated by using Student's two-tailed t test. The developed colorimetric,
371 fluorescent and dual-signal methods are comparable to the standard method since the
372 differences in nitrite concentration between them are fairly negligible (P value (0.939,
373 0.980 and 0.987) > 0.05). In addition, the LOD of this method (196 nM for
374 colorimetric detection and 0.518 nM for fluorescent detection) is much lower than that
375 of Griess-based method (229 nM). This method yields excellent quantification results
376 ($R^2 = 0.995$ and 0.991) as accurate as Griess-based method ($R^2 = 0.994$). Those results
377 demonstrate that most of common compounds (e.g., ions and reaction products of
378 nitrite) do not interfere with the measurement of nitrite with the use of the proposed
379 method (Li et al., 2003).

380 The other standard methodology for nitrite detection is ion chromatography
381 (CHM, 2016; ISO, 1975). The detection time (about 30 min) by using this method to
382 measure a set of 7 samples is less than ion chromatography (about 3 hours). This
383 method has an advantage of requiring simpler sample preparation because less
384 centrifugation and filtration, as well as no cleanup steps (Siu & Henshall, 1998).
385 Generally, only 3 steps are needed to measure nitrite after sample preparation for this
386 method: (1) mixing nitrite extraction and C-dots-NR system; (2) acquiring spectra; (3)
387 calculating nitrite concentration. This method is much faster and simpler than ion
388 chromatography for nitrite detection in hams(Q.-H. Wang et al., 2017).



389
390 Fig. 6 absorbance (a) and fluorescent (b) spectra of C-dots-NR system with addition
391 of ham extractions.

392 Table 2 detection results of nitrite in hams based on Griess reagent and C-dots-NR
 393 system

Sample	Griess	C-dots-NR system					
	reagent	absorbance		fluorescence		signal fusion	
	C(NO ₂ ⁻) mg/kg)	C(NO ₂ ⁻) mg/kg)	Error (%)	C(NO ₂ ⁻) mg/kg)	Ratio (%)	C(NO ₂ ⁻) mg/kg)	Error (%)
Ham 1	0.102	0.098	3.922	0.099	2.914	0.087	14.706
Ham 2	0.700	0.677	3.286	0.676	3.429	0.850	21.429
Ham 3	6.769	7.059	4.284	6.714	0.813	6.921	2.245
Ham 4	11.612	12.020	3.514	11.628	0.138	11.709	0.835
Ham 5	15.872	16.205	2.098	15.899	0.170	15.788	0.529
Ham 6	16.397	17.189	4.830	16.429	0.195	16.599	1.232
Ham 7	22.276	22.969	3.110	22.190	0.386	22.276	0.000

394 **4 Conclusion**

395 A novel FRET system based on C-dots and NR has been applied successfully
 396 for colorimetric and fluorescent detection of nitrite residue in hams. Nitrite
 397 concentrations had excellent linear relation with absorbance values at 517 nm ($R^2=$
 398 0.995) and fluorescent intensities at 520 nm ($R^2=0.991$). Limit of detection for
 399 nitrite using colorimetric and fluorescent method was 169 nM and 0.517 nM,
 400 respectively. Furthermore, the performance based on this proposed method was
 401 accordance with that of the Griess-based method for detection of nitrite in hams
 402 (relative errors < 5%). The dual-mode optical sensor has several advantages in
 403 sensitive detection, convenient instrumentation and simple operation over
 404 conventional methods. In summary, the developed C-dots-NR system is a potential
 405 colorimetric and fluorescent sensor for simple, selective and sensitive detection of
 406 nitrite in hams.

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417 **Compliance with Ethical Standards**

418 **Conflict of Interest** The authors declare that they have no conflict of interest.

419 **Ethical Approval** This article does not contain any studies with human participants or
420 animals performed by any of the authors.

421 **Informed Consent** Informed consent is not applicable for the nature of this study.

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