

This is a repository copy of *Chromoselective Photocatalysis Enables* Stereocomplementary Biocatalytic Pathways.

White Rose Research Online URL for this paper: https://eprints.whiterose.ac.uk/172458/

Version: Published Version

Article:

Schmermund, Luca, Reischauer, Susanne, Bierbaumer, Sarah et al. (9 more authors) (2022) Chromoselective Photocatalysis Enables Stereocomplementary Biocatalytic Pathways. Angewandte Chemie International Edition. ISSN 1433-7851

https://doi.org/10.1002/anie.202100164

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here: https://creativecommons.org/licenses/

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.





Communications



Photobiocatalysis

How to cite: Angew. Chem. Int. Ed. 2021, 60, 6965-6969 International Edition: doi.org/10.1002/anie.202100164 German Edition: doi.org/10.1002/ange.202100164

Chromoselective Photocatalysis Enables Stereocomplementary Biocatalytic Pathways**

Luca Schmermund, Susanne Reischauer, Sarah Bierbaumer, Christoph K. Winkler, Alba Diaz-Rodriguez, Lee J. Edwards, Selin Kara, Tamara Mielke, Jared Cartwright, Gideon Grogan, Bartholomäus Pieber,* and Wolfgang Kroutil*

Abstract: Controlling the selectivity of a chemical reaction with external stimuli is common in thermal processes, but rare in visible-light photocatalysis. Here we show that the redox potential of a carbon nitride photocatalyst (CN-OA-m) can be tuned by changing the irradiation wavelength to generate electron holes with different oxidation potentials. This tuning was the key to realizing photo-chemo-enzymatic cascades that give either the (S)- or the (R)-enantiomer of phenylethanol. In combination with an unspecific peroxygenase from Agrocybe aegerita, green light irradiation of CN-OA-m led to the enantioselective hydroxylation of ethylbenzene to (R)-1-phenylethanol (99% ee). In contrast, blue light irradiation triggered the photocatalytic oxidation of ethylbenzene to acetophenone, which in turn was enantioselectively reduced with an alcohol dehydrogenase from Rhodococcus ruber to form (S)-1-phenylethanol (93 % ee).

Many parameters influence the selectivity of a chemical reaction.^[1] For instance, catalytic reactions can be controlled by varying the catalyst/coordinated ligands, directing groups^[2] or by tuning external parameters (Scheme 1 A). [1a,3] The selectivity of photochemical reactions varies with different wavelengths, [4] but examples that use this for visible-light photocatalysis are rare.[5]

In one example, selective control between either a one- or two-fold substitution of 1,3,5-tribromobenzene with N-methylpyrrole using Rhodamin 6G (Rh-6G) as photocatalyst was demonstrated (Scheme 1B). [5a] This selectivity switch is explained by the chromoselective generation of two photocatalytic species that differ in their reduction potential. Green light irradiation results in a common photoredox cycle and the expected mono-substituted product. In the case of blue light, the Rh-6G radical anion, which is formed after quenching of Rh-6G* with a sacrificial electron donor, can absorb a second photon, resulting in the highly reducing Rh-6G^{-*} species that enables the formation of the di-substituted product.^[5a]

Here we show that electron holes with different oxidation potentials can be generated by using a heterogeneous carbon nitride (CN) catalyst by changing the incident photon energy. The combination of this strategy with two enantioselective biocatalysts^[6] allowed us to selectively produce the (S)- or (R)-enantiomer of a chiral alcohol in photo-chemo-enzymatic reaction sequences (Scheme 1 C).

We recently realized that the choice of the wavelength is crucial for high selectivities in metallaphotocatalytic cross couplings using a heterogeneous carbon nitride material, which is made from urea and oxamide in molten salt (CN-OA-m). [5b,c,7] While this can be rationalized by a purely kinetic effect, there is also evidence that a wavelength-controlled generation of excited species with different oxidation potentials might be responsible for this phenomenon. CN-OA-m has a strong absorption up to $\approx 460 \text{ nm}$ and a comparably weaker absorption band up to ≈ 700 nm, which were ascribed as the π - π^* and n- π^* electron transitions, respectively (Figure 1A). $^{[8]}$ The selective induction of the n- π^* electron transition using long wavelengths (525 nm) should result in electron holes with a lower oxidation potential compared to irradiation using blue light (440 nm). The choice of the wavelength should not affect the reduction potential of the electron that is promoted into the valence band. Although

[*] L. Schmermund, S. Bierbaumer, Dr. C. K. Winkler, Prof. W. Kroutil Institute of Chemistry, Department of Organic and Bioorganic Chemistry, University of Graz, NAWI Graz, BioTechMed Graz Heinrichstrasse 28, 8010 Graz (Austria) E-mail: wolfgang.kroutil@uni-graz.at

S. Reischauer, Dr. B. Pieber

Department of Biomolecular Systems, Max Planck Institute of Colloids and Interfaces

Am Mühlenberg1, 14476 Potsdam (Germany) E-mail: Bartholomaeus.Pieber@mpikg.mpg.de

Dr. A. Diaz-Rodriguez, Dr. L. J. Edwards

Chemical Development, Medicinal Science and Technology, Pharma R&D, GlaxoSmithKline Medicines Research Centre Gunnels Wood Road, Stevenage SG1 2NY (UK)

Department of Engineering, Biological and Chemical Engineering, Biocatalysis and Bioprocessing Group, Aarhus University Gustav Wieds Vej 10, 8000 Aarhus (Denmark)

T. Mielke, Dr. J. Cartwright, Prof. G. Grogan Department of Chemistry, University of York Heslington, York, YO10 5DD (UK) Prof. W. Kroutil

Field of Excellence BioHealth-University of Graz 8010 Graz (Austria)

- [**] A previous version of this manuscript has been deposited on a preprint server (https://doi.org/10.26434/chemrxiv.13521527).
- Supporting information and the ORCID identification number(s) for the author(s) of this article can be found under: https://doi.org/10.1002/anie.202100164.
- © 2021 The Authors. Angewandte Chemie International Edition published by Wiley-VCH GmbH. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

6965



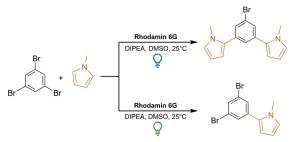




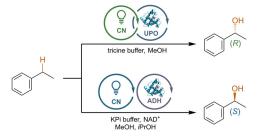


A temperature, pressure, pH, solvent, ligand, catalyst

B Chromoselective access to different reduction potentials



C This work: Chromoselective control of oxidation potentials in photo-chemo-enzymatic cascades



Scheme 1. A) General approaches to control of the outcome of a chemical reaction. B) Chromoselective control in photocatalytic C—H-arylations. ^[5a] C) This study: Chromoselective control of the stereochemical outcome of photo-chemo-enzymatic reactions.

such a behavior was previously suggested, [8] there is, to the best of our knowledge, no report that applies this concept for controlling the selectivity of chemical reactions.

We hypothesized that such a strategy would allow us to induce a photocatalytic reaction of a substrate with green light selectively in the presence of a second compound that is only photo-oxidized when shorter wavelengths are used. The photocatalytic aerobic oxidation of benzylic sp³ C–H bonds, which is feasible with other members of the carbon nitride family and blue light irradiation, [9] served as a model reaction for our initial studies. In a series of experiments, we were indeed able to show that only blue light results in the desired carbonyl products and no reaction occurs at longer wavelengths (Figure 1 B).

Carbon nitrides are used to catalyse the formation of O_2 and H_2 via water oxidation^[10] and the production of hydrogen peroxide from oxygen and alcohols, which requires the reduction of O_2 .^[11] Hydrogen peroxide can then be used as stoichiometric oxidant in the enantioselective hydroxylation of ethylbenzene derivatives catalysed by the unspecific peroxygenase (UPO)^[12] from *A. aegerita*^[13] (*Aae*UPO) acting as chiral catalyst.^[14]

We hypothesized that a chromoselective activation of CN-OA-m with green light enables the selective formation of

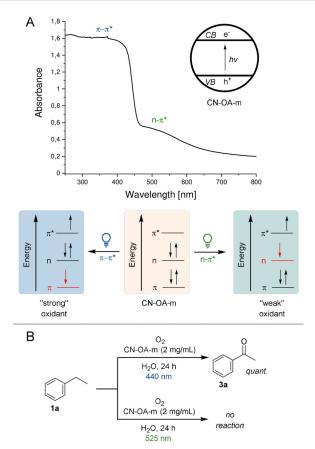


Figure 1. Chromoselective generation of excited CN-OA-m species with different oxidation potentials. A) Switching between $\pi-\pi^*$ and $n-\pi^*$ electron transitions using different wavelengths. B) The oxidation of ethylbenzene **1 a** to acetophenone **3 a** is only possible using blue light.

H₂O₂ in the presence of ethylbenzene (1) and the *Aae*UPO, which in turn catalyses the asymmetric hydroxylation of 1 (Figure 2). Performing the reaction in tricine buffer using 528 nm LEDs indeed resulted in a high selectivity towards (*R*)-1-phenylethanol formation [(*R*)-2a, up to 3.8 mM, 98% *ee*] with low amounts (3%) of acetophenone (3a). When the same reaction was carried out using shorter wavelengths, 3a became the main product, thus supporting our hypothesis. Ketone (3a) formation was also the preferred reaction in the presence of blue light in phosphate buffer. It is worth to note, that the type of buffer had a significant influence on the outcome on the reaction, whereby the molecular reason needs to be clarified.

It was previously shown that UPOs are deactivated in the presence of blue light, a photocatalyst and O₂ due to the generation of reactive oxygen species (ROS) that harm the enzyme. ^[15] Consequently, one might expect that green light might be less harmful to the UPO and lead to higher conversions in comparison to blue light. To investigate this aspect, UPO and CN-OA-m were incubated for one hour in the presence of oxygen and green or blue light, before **1a** was added (Figure S48). The mixture incubated at longer wavelengths indeed led to a higher conversion for the asymmetric hydroxylation after addition of **1a**.



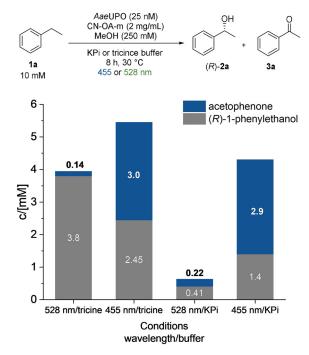


Figure 2. Influence of different wavelengths and buffers on the photochemo-enzymatic hydroxylation of ethylbenzene; reaction conditions: AaeUPO (25 nM), ethylbenzene (10 mM), CN-OA-m (2 mg mL⁻¹), MeOH (250 mM), KPi (100 mM, pH 7.5) or tricine (100 mM, pH 7.5), 455 nm (1440 μ mol photons m⁻² s⁻¹) or 528 nm (1330 μ mol photons m⁻² s⁻¹), 30 °C, 8 h.

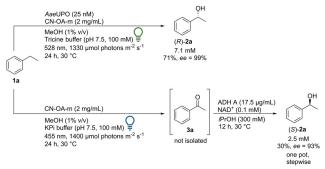
Scheme 2. Substrate scope of AaeUPO using H2O2 generated by CN-OA-m under green light irradiation. Absolute configurations were determined by reference material except otherwise stated. [a] Based on external calibration curves of 2i. [b] (R)-Enantiomer determined by measurement of the specific rotation (20 °C, c = 1.00, CHCl₃) and comparison to literature.

The milder conditions subsequently allowed an extension of the substrate scope for AaeUPO (Scheme 2). Nine additional substrates were converted with high stereoselectivity (> 98 % ee) to the corresponding alcohols with concentrations of 1.0-6.0 mM. None of these ethylbenzene derivatives has been transformed with AaeUPO using an in situ H₂O₂ generation system before.

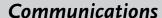
Ethylbenzenes bearing a methyl substituent in the orthoor meta-position were hydroxylated with 99 % regioselectivity at the ethyl group to give the desired chiral alcohols (R)-2b,c. This ability to distinguish between a methyl and an ethyl group has not been reported before. A possible explanation for this selectivity might be a preferred formation of the secondary intermediate radical over the primary radical. Acetophenone substituted with ethyl in the *para*-position (1i) allowed one to access a bi-functionalised chiral hydroxyketone 2i, which is otherwise difficult to make. The same is true for 2j.

Recycling experiments further showed that CN-OA-m can be reused by centrifugation and one washing step with water. CN-OA-m was reused three times after drying at room temperature (Figure S49-S51). Transferring the photochemo-enzymatic hydroxylation from a total volume of 1 mL in 1.5 mL glass vials successfully to a larger scale (7 mL volume, 10 mL tubes) in another photoreactor (provided by GlaxoSmithKline, S5), [16] showed the robustness and reproducibility of the approach. The hydroxylation of 1a worked equally well giving up to 7.5 mM of (R)-2a.

Recently, photo-chemo-biocatalytic cascades reported combining a photoredox oxidation of ethylbenzene with an enzymatic reduction.^[17] In a related approach a photochemo-biocatalytic cascade that yields the corresponding (S)enantiomers was set up by taking advantage of the chromoselective activation of CN-OA-m (Scheme 3). The blue-light mediated oxidation of 1a to 3a proceeded smoothly in KPi buffer. The resulting ketone (3a) was stereoselectively reduced using an alcohol dehydrogenase (ADH-A) from Rhodococcus ruber in the presence of NAD⁺ as cofactor.^[18] The optimized two-step one-pot procedure led to 2.5 mM (S)-2a with an ee of 93%. The lower ee obtained in the photochemo-enzymatic cascade compared to previous reports by ADH-A (ee 99%), [19] can be explained by the formation of a small amount of rac-1-phenylethanol during the photocatalytic reaction under blue light irradiation (Table S3). This cascade represents a stereocomplementary pathway compared to the pathway with AaeUPO using the same photocatalyst. Interestingly, it was noticed that MeOH was not required for the reaction to hydroxylate ethylbenzene with AaeUPO. Without MeOH the same concentration of



Scheme 3. Light-driven enantioselective oxyfunctionalizations of 1a by using chromoselective CN-OA-m and AaeUPO or ADH-A.







product was detected. Thus, the reaction is possible without a sacrificial electron donor like MeOH or formate, which is in contrast to some examples reported in literature. [14a,20] For practical reasons, MeOH was still used since it simplified the preparation of stock solutions of the hydrophobic substrates. To test whether the cascade can also be transferred to other substrates, *para*- and *ortho*-bromo-substituted ethylbenzene (1g, 1h) were investigated: Using the blue-light pathway, (S)-2g was obtained with an ee of >99% (1 mM) and (S)-2h with an ee of 94% (1.4 mM) (Figure S32 and S37).

To the best of our knowledge this is the first example in which it was possible to utilize the same photocatalyst to either oxidize an organic substrate or to provide in situ formed H₂O₂ without photocatalytic oxidation of the substrate, all controlled only by the choice of the wavelength.

In summary, we showed that electron holes with different oxidation potentials can be generated using a carbon nitride material by simply changing the photon energy. In the presence of blue light this enables the oxidation of ethylbenzene to acetophenone in an aqueous solution. Using green light, the organic substrate does not react and only H₂O₂ is formed. This was the key for designing chromoselective photo-chemo-enzymatic cascade reactions. Selective hydrogen peroxide generation enabled the hydroxylation of ethylbenzene to give (R)-1-phenylethanol (R)-2a using an UPO, whereas the photocatalytic oxidation to acetophenone was coupled with an enantioselective reduction to (S)-1-phenylethanol (S)-2a by an ADH. Additionally, low-energy photons (green light) increased the stability of UPO compared to blue light, which permitted the expansion of the substrate scope of this enzyme. Controlling the outcome of a photocatalytic reaction merely through the choice of wavelength employed presents exciting new options in reaction design and could be an important new tool for controlling reactivity and stereoselection in organic synthesis.

Acknowledgements

This project received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-Curie grant agreement No 764920. S.B. acknowledges the Austrian Science Fund (FWF) for funding within the project CATALOX (DOC 46-B21). The University of Graz and the Field of Excellence BioHealth are acknowledged for financial support. We thank GlaxoSmithKline for providing the Photochemistry LED Illuminator (Pacer Components Ltd. Pangbourne, UK). S.R. and B.P. acknowledge the Max-Planck Society and the German Chemical Industry Fund (Liebig Fellowship, Fonds der Chemischen Industrie, FCI) for generous financial support. B.P. thanks the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) under Germany's Excellence Strategy-EXC 2008-390540038—UniSysCat for financial support. T.M. was funded by a studentship awarded by the industrial affiliates of the Centre of Excellence for Biocatalysis, Biotransformations and Biomanufacture (CoEBio3). We thank Prof. Klaus Zangger and Bernd Werner for recording of NMR spectra.

Conflict of interest

The authors declare no conflict of interest.

Keywords: carbon nitrides \cdot C-H activation \cdot chromoselectivity \cdot photobiocatalysis \cdot unspecific peroxygenases

- [1] a) T. Gaich, E. Winterfeldt, Directed Selectivity in Organic Synthesis: A Practical Guide, 1st ed., Wiley-VCH, Weinheim, 2014; b) G. Statham, Synthese 2017, 194, 4815-4838.
- [2] a) I. Fleming, A. Barbero, D. Walter, *Chem. Rev.* 1997, 97, 2063–2192; b) C. Sambiagio, D. Schönbauer, R. Blieck, T. Dao-Huy, G. Pototschnig, P. Schaaf, T. Wiesinger, M. F. Zia, J. Wencel-Delord, T. Besset, B. U. W. Maes, M. Schnürch, *Chem. Soc. Rev.* 2018, 47, 6603–6743; c) G. Rousseau, B. Breit, *Angew. Chem. Int. Ed.* 2011, 50, 2450–2494; *Angew. Chem.* 2011, 123, 2498–2543; d) Z. Huang, G. Dong, *Acc. Chem. Res.* 2017, 50, 465–471.
- [3] J. Jurczak, *Physica B* + C **1986**, 139-140, 709-716.
- [4] S. Protti, D. Ravelli, M. Fagnoni, *Photochem. Photobiol. Sci.* 2019, 18, 2094–2101.
- [5] a) I. Ghosh, B. König, Angew. Chem. Int. Ed. 2016, 55, 7676–7679; Angew. Chem. 2016, 128, 7806–7810; b) S. Gisbertz, S. Reischauer, B. Pieber, Nat. Catal. 2020, 3, 611–620; c) C. Cavedon, E. T. Sletten, A. Madani, O. Niemeyer, P. H. Seeberger, B. Pieber, Org. Lett. 2021, 23, 514–518; d) A. M. Martínez-Gualda, R. Cano, L. Marzo, R. Pérez-Ruiz, J. Luis-Barrera, R. Mas-Ballesté, A. Fraile, V. A. de la Peña O'Shea, J. Alemán, Nat. Commun. 2019, 10, 2634.
- [6] C. K. Winkler, J. H. Schrittwieser, W. Kroutil, ACS Cent. Sci. 2021, 7, 55—71.
- [7] B. Pieber, J. A. Malik, C. Cavedon, S. Gisbertz, A. Savateev, D. Cruz, T. Heil, G. Zhang, P. H. Seeberger, *Angew. Chem. Int. Ed.* 2019, 58, 9575–9580; *Angew. Chem.* 2019, 131, 9676–9681.
- [8] G. Zhang, G. Li, Z.-A. Lan, L. Lin, A. Savateev, T. Heil, S. Zafeiratos, X. Wang, M. Antonietti, Angew. Chem. Int. Ed. 2017, 56, 13445 13449; Angew. Chem. 2017, 129, 13630 13634.
- [9] a) P. Geng, Y. Tang, G. Pan, W. Wang, J. Hu, Y. Cai, Green Chem. 2019, 21, 6116-6122; b) W. Zhang, A. Bariotaki, I. Smonou, F. Hollmann, Green Chem. 2017, 19, 2096-2100.
- [10] W. J. Ong, L. L. Tan, Y. H. Ng, S. T. Yong, S. P. Chai, Chem. Rev. 2016, 116, 7159 – 7329.
- [11] a) Y. Shiraishi, S. Kanazawa, Y. Kofuji, H. Sakamoto, S. Ichikawa, S. Tanaka, T. Hirai, Angew. Chem. Int. Ed. 2014, 53, 13454–13459; Angew. Chem. 2014, 126, 13672–13677; b) Y. Shiraishi, S. Kanazawa, Y. Sugano, D. Tsukamoto, H. Sakamoto, S. Ichikawa, T. Hirai, ACS Catal. 2014, 4, 774–780; c) Z. Wei, M. Liu, Z. Zhang, W. Yao, H. Tan, Y. Zhu, Energy Environ. Sci. 2018, 11, 2581–2589; d) X. Zeng, Y. Liu, Y. Kang, Q. Li, Y. Xia, Y. Zhu, H. Hou, M. H. Uddin, T. R. Gengenbach, D. Xia, C. Sun, D. T. McCarthy, A. Deletic, J. Yu, X. Zhang, ACS Catal. 2020, 10, 3697–3706.
- [12] a) Y. Wang, D. Lan, R. Durrani, F. Hollmann, Curr. Opin. Chem. Biol. 2017, 37, 1–9; b) M. Faiza, S. Huang, D. Lan, Y. Wang, BMC Evol. Biol. 2019, 19, 76; c) M. Hofrichter, R. Ullrich, Curr. Opin. Chem. Biol. 2014, 19, 116–125; d) M. Hobisch, D. Holtmann, P. Gomez de Santos, M. Alcalde, F. Hollmann, S. Kara, Biotechnol. Adv. 2020, 0, 107615.
- [13] a) P. Molina-Espeja, E. Garcia-Ruiz, D. Gonzalez-Perez, R. Ullrich, M. Hofrichter, M. Alcalde, *Appl. Environ. Microbiol.* 2014, 80, 3496–3507; b) P. Molina-Espeja, S. Ma, D. M. Mate, R. Ludwig, M. Alcalde, *Enzyme Microb. Technol.* 2015, 73–74, 29–33.
- [14] a) W. Zhang, B. O. Burek, E. Fernandez-Fueyo, M. Alcalde, J. Z. Bloh, F. Hollmann, *Angew. Chem. Int. Ed.* 2017, *56*, 15451–15455; *Angew. Chem.* 2017, *129*, 15654–15658; b) L. Schmermund, V. Jurkaš, F. F. Özgen, G. D. Barone, H. C. Büchsen-



Communications



schütz, C. K. Winkler, S. Schmidt, R. Kourist, W. Kroutil, ACS Catal. 2019, 9, 4115-4144; c) B. O. Burek, S. Bormann, F. Hollmann, J. Z. Bloh, D. Holtmann, Green Chem. 2019, 21, 3232-3249; d) S. J. P. Willot, E. Fernández-Fueyo, F. Tieves, M. Pesic, M. Alcalde, I. W. C. E. Arends, C. B. Park, F. Hollmann, ACS Catal. 2019, 9, 890-894; e) F. Tieves, S. J.-P. Willot, M. M. C. H. van Schie, M. C. R. Rauch, S. H. H. Younes, W. Zhang, J. Dong, P. Gomez de Santos, J. M. Robbins, B. Bommarius, M. Alcalde, A. S. Bommarius, F. Hollmann, Angew. Chem. Int. Ed. 2019, 58, 7873-7877; Angew. Chem. 2019, 131, 7955-7959; f) Y. Ni, E. Fernández-Fueyo, A. G. Baraibar, R. Ullrich, M. Hofrichter, H. Yanase, M. Alcalde, W. J. H. van Berkel, F. Hollmann, Angew. Chem. Int. Ed. 2016, 55, 798-801; Angew. Chem. 2016, 128, 809-812; g) D. S. Choi, Y. Ni, E. Fernández-Fueyo, M. Lee, F. Hollmann, C. B. Park, ACS Catal. 2017, 7, 1563-1567; h) E. Churakova, M. Kluge, R. Ullrich, I. Arends, M. Hofrichter, F. Hollmann, Angew. Chem. Int. Ed. **2011**, 50, 10716–10719; Angew. Chem. **2011**, 123, 10904–10907; i) M. Hobisch, M. M. C. H. van Schie, J. Kim, K. R. Andersen, M. Alcalde, R. Kourist, C.B. Park, F. Hollmann, S. Kara, ChemCatChem 2020, 12, 4009-4013; j) J. Yoon, J. Kim, F. Tieves, W. Zhang, M. Alcalde, F. Hollmann, C. B. Park, ACS Catal. 2020, 10, 5236-5242; k) W. Zhang, E. Fernandez-Fueyo, Y. Ni, M. van Schie, J. Gacs, R. Renirie, R. Wever, F. G. Mutti, D. Rother, M. Alcalde, F. Hollmann, Nat. Catal. 2018, 1, 55-62.

[15] a) S. Bormann, A. Gomez Baraibar, Y. Ni, D. Holtmann, F. Hollmann, Catal. Sci. Technol. 2015, 5, 2038-2052; b) B. O. Burek, S. R. de Boer, F. Tieves, W. Zhang, M. van Schie, S.

- Bormann, M. Alcalde, D. Holtmann, F. Hollmann, D. W. Bahnemann, J. Z. Bloh, ChemCatChem 2019, 11, 3093-3100.
- [16] a) H. E. Bonfield, J. D. Williams, W. X. Ooi, S. G. Leach, W. J. Kerr, L. J. Edwards, ChemPhotoChem 2018, 2, 938-944; b) H. E. Bonfield, K. Mercer, A. Diaz-Rodriguez, G. C. Cook, B. S. J. McKay, P. Slade, G. M. Taylor, W. X. Ooi, J. D. Williams, J. P. M. Roberts, J. A. Murphy, L. Schmermund, W. Kroutil, T. Mielke, J. Cartwright, G. Grogan, L. J. Edwards, ChemPhoto-Chem 2020, 4, 45-51.
- [17] a) R. C. Betori, C. M. May, K. A. Scheidt, Angew. Chem. Int. Ed. **2019**, 58, 16490 – 16494; Angew. Chem. **2019**, 131, 16642 – 16646; b) Y. Peng, D. Li, J. Fan, W. Xu, J. Xu, H. Yu, X. Lin, Q. Wu, Eur. J. Org. Chem. 2020, 821 – 825.
- [18] K. Edegger, C. C. Gruber, T. M. Poessl, S. R. Wallner, I. Lavandera, K. Faber, F. Niehaus, J. Eck, R. Oehrlein, A. Hafner, W. Kroutil, Chem. Commun. 2006, 2402-2404.
- [19] W. Stampfer, B. Kosjek, C. Moitzi, W. Kroutil, K. Faber, Angew. Chem. Int. Ed. 2002, 41, 1014-1017; Angew. Chem. 2002, 114, 1056 - 1059.
- [20] M. M. C. H. van Schie, W. Zhang, F. Tieves, D. S. Choi, C. B. Park, B. O. Burek, J. Z. Bloh, I. W. C. E. Arends, C. E. Paul, M. Alcalde, F. Hollmann, ACS Catal. 2019, 9, 7409-7417.

Manuscript received: January 5, 2021 Accepted manuscript online: February 2, 2021 Version of record online: February 26, 2021