



UNIVERSITY OF LEEDS

This is a repository copy of *Recent developments in food foams*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/167787/>

Version: Accepted Version

Article:

Murray, BS orcid.org/0000-0002-6493-1547 (2020) Recent developments in food foams. *Current Opinion in Colloid & Interface Science*, 50. 101394. ISSN 1359-0294

<https://doi.org/10.1016/j.cocis.2020.101394>

© 2020, Elsevier. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

Reuse

This article is distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs (CC BY-NC-ND) licence. This licence only allows you to download this work and share it with others as long as you credit the authors, but you can't change the article in any way or use it commercially. More information and the full terms of the licence here: <https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Current Opinion in Colloid & Interface Science

Recent Developments in Food Foams

--Manuscript Draft--

Manuscript Number:	COCIS-D-20-00038R1
Full Title:	Recent Developments in Food Foams
Article Type:	50: Thin Liquid Films and Foams
Short Title:	Food Foams
Keywords:	food; foams; Pickering; particles, biopolymers
Corresponding Author:	Brent S. Murray University of Leeds Leed, W. Yorks UNITED KINGDOM
Corresponding Author's Institution:	University of Leeds
Corresponding Author E-Mail:	b.s.murray@leeds.ac.uk
First Author:	Brent S. Murray
Order of Authors:	Brent S. Murray
Abstract:	<p>The scientific literature between from 2015 onwards (inclusive) with respect to foams and thin films in the context of foods has been reviewed. Proteins are the dominant foaming agents in foods and investigations of the classic, meringue-forming egg white protein still dominate the literature, since the unique properties of this system are still not properly understood. The current drive of many studies is to find suitable replacers of egg proteins, driven by consumer trends for more plant-based alternatives. This has led to investigations of the stabilizing properties of various protein aggregates, 'nanoparticles' and microgel particles as Pickering-type stabilizers of foams (Pickering foams). At the same time, other work has sought to manipulate the surface properties of biopolymer- and non-biopolymer -based particles by chemical means, in order to make the particles adsorb strongly enough. Few, truly novel foam stabilizers have emerged, but two include saponin aggregates and bacteria as particle-type stabilizers.</p>
Author Comments:	<p>Dear Editor, Thank you for the positive response to our review. I note your minor editorial comment and have inserted any missing page numbers or article numbers in the references where these are available. I also thank the Reviewer for their suggestions, which I have addressed as follows, allowing the opportunity to improve the MS further. Below I repeat Reviewer 1's comments/suggestions, followed by my response, in italics for clarity.</p> <p>It is not really clear where the Introduction ends and the material specific discussion starts.</p> <p>In the Introduction I have focused almost exclusively on the generic problems of stabilizing foams, though I have introduced Pickering foams as a special case. At the end of the Introduction I introduced Table 1, a sort of classification summary of the different types of stabilizer as a lead into the following sections, which discuss most of these in turn. However, to make this more clear, I have changed the last sentence of the Introduction to "The main classes and sub-class of foam stabilizer covered in this review are summarized in Table 1 and these will be discussed in the following sections."</p> <p>The Abstract features abbreviations and parentheses, which impairs the optical and reading pleasure. Later on also some sentences are placed parentheses: In my opinion, the parentheses are not needed.</p> <p>There is only one abbreviation in the Abstract, for egg white protein (EWP), which I subsequently used only once. I think this was introduced this mainly to keep the word count down. Nevertheless, I have removed this. There is one set of parentheses, around "Pickering foams". I think this is justified because the term is not entirely widespread yet, unlike Pickering emulsions.</p> <p>On page 7 there is a valid comment on the often-claimed link between foam stability</p>

and interfacial properties. I think the comment is important and should be extended to interfacial shear rheology as well (right now only interfacial dilatational rheo is mentioned).

We agree entirely and so have extended the 4th sentence already there in paragraph 2 of page 7, so that this now reads "Once adsorbed, most proteins are also able to form a thin, viscoelastic, crossed-linked network that can resist further bubble deformation, up to a point, often more reflected in measurements of the interfacial shear rheological properties."

For cellulose (page 13) one could also mention some of the recent published reviews on cellulose at interfaces (<https://doi.org/10.1021/acs.chemmater.7b00531>, <https://doi.org/10.1016/j.cocis.2017.04.001>, <https://doi.org/10.1016/j.cis.2019.102089>). These reviews not entirely focus on foams but might help the reader to understand cellulose interfaces better.

There have been a huge number of recent publications relating to cellulose and various colloidal systems so that it is impossible to know which to select unless they specifically relate to foams, the subject of this review. In addition, there is a lot of confusion between interfacial and network stabilization, which we have discussed. In the light of this, the best of Reviewer 1's recommendations seem to be the 2nd and 3rd (Capron et al., 2017 and Bertsch & Fischer, 2020), which discuss the nature of cellulose surface activity and foams: we have therefore included these two in the revised MS, in the 3rd sentence of the section of cellulose and chitin, which now ends "... or the reason for any apparent natural surface activity of cellulose nanocrystals" with these references inserted.

The first recommendation (De France et al, 2017), relates to aerogels and NOT foams, so we have not included it since the above two cover the required topic adequately.

Page 14: Heading "Combinations of particles and other foaming agents". What do you mean by foaming agents? The foam stabilizing surface-active materials or the gas? In engineering nomenclature the foaming agent would be the gas. Just to avoid confusion.

I was not aware of this engineering nomenclature, so thanks to the Reviewer for bringing this to my attention. I have clarified this by changing the title heading to ""Combinations of particles and other foam stabilizing agents"

Page 16: Section on oleogel. It might be worth to check with two super-recent publications by Mishra et al. on fat foam generation (<https://pubs.acs.org/doi/abs/10.1021/acs.cgd.9b01558>) and the interfacial aggregation of differently treated triglycerides (<https://pubs.acs.org/doi/10.1021/acs.langmuir.0c01195>).

Again this is an area that is expanding rapidly, so that it is hard to keep up, so the recommendations of Reviewer 1 are very welcome. The second (Langmuir paper) actually refers to the work in the first (Crystal Growth and Design) paper, so we have included just the second, adding the sentence: " Some these issues have been nicely discussed in a recent contributions from Mishra et al. [inserted] at the end of the "Oleogels and oleofoams" section.

[Inclusion of these additional reference has obviously activated some reference re-numbering].

Recent Developments in Food Foams

Brent S. Murray^{1}*

b.s.murray@leeds.ac.uk

¹Food Colloids and Bioprocessing Group, School of Food Science and Nutrition, University of Leeds, Leeds, LS2 9JT, UK.

For submission to Current Opinion in Colloid & Interface Science issue Thin Films & Foams

Abstract

The scientific literature between from 2015 onwards (inclusive) with respect to foams and thin films in the context of foods has been reviewed. Proteins are the dominant foaming agents in foods and investigations of the classic, meringue-forming egg white protein (EWP) still dominate the literature, since the unique properties of this system are still not properly understood. The current drive of many studies is to find suitable replacers of EWP, driven by consumer trends for more plant-based alternatives. This has led to investigations of the stabilizing properties of various protein aggregates, 'nanoparticles' and microgel particles as Pickering-type stabilizers of foams (Pickering foams). At the same time, other work has sought to manipulate the surface properties of biopolymer- and non-biopolymer -based particles by chemical means, in order to make the particles adsorb strongly enough. Few, truly novel foam stabilizers have emerged, but two include saponin aggregates and bacteria as particle-type stabilizers.

Keywords: food; foams; Pickering; particles, biopolymers

Introduction

Foam is an important component of many commonly consumed food and drinks (e.g., whipped creams, mousses, soufflés, ice creams, bread, cakes, carbonated beverages) and research into new types of foam and new foam-forming and foam-stabilizing agents continues to grow. Bubbles in consumer products can impart texture, colour, novelty and excitement for the consumer. Additional advantages are that, for air bubbles the bulk material is notoriously cheap, light to transport and is unlikely to result in allergic reactions (for clean air at least) ! If only things were so simple, however, because gas bubbles are one of the most difficult colloidal substances to produce, stabilize and control in large quantities, due to their tendency for relatively rapid creaming, dissolution and coalescence (relative to oil-water emulsions, for example). Consequently, great efforts have been made to design systems with improved foam properties, not least because bubbles represent a zero-calorie filler that might be used to replace other food components and have a beneficial effect on the long-term health and well-being of consumers. Thus, apart from the general case of filling the aqueous matrix of foods with more air, systems have been developed for trapping air bubbles in sugar crystals [1], fats and oils [2]●. In addition, there seems to be increasing awareness of the significance and potential of using foam as a 'processing aid'. Thus bubbles can be deliberately used to fractionate a wide range of more valuable surface active materials, whilst conversely foam may concentrate undesirable chemical species. In addition, foam structure may aid the drying and functional properties of dry powders. On the other hand, foaming is an undesirable phenomenon in other processes and understanding how to enhance the action on particles and surfactants in breaking the thin films between bubbles is also an important area of work. It is worth elaborating at the outset some of the colloidal difficulties referred to above, in order to understand the general direction of recent alternative strategies and formulations in the publications referred to later.

Firstly, in the majority of food foams the gas phase is air, nitrogen or carbon dioxide and the continuous phase is an aqueous solution, so that the discontinuous phase (gas) has reasonable (at least mM) solubility in the continuous phase. This is unlike most emulsions or solid dispersions and means that disproportionation, the equivalent of Ostwald ripening in emulsions and dispersions, is rapid when bubbles approach colloidal dimensions, i.e., ca. 1 μm . Adsorbed layers of even tightly packed low molecular weight surfactants (LMWS), or gel-like films of adsorbed macromolecules, offer little physical barrier to transport of gas molecules across interfaces, so that the only way to halt disproportionation completely is to use solid particles, i.e., Pickering stabilization (Pickering foams) as the stabilizing mechanism [3]. Adsorbed layers of such particles can provide a perfectly elastic barrier with high resistance to compression and desorption of the particles, 'hijacking' shrinkage of bubbles due to the higher internal gas pressure across curved interfaces. (The effect of the Laplace pressure difference across the air-water (A-W) interface is exacerbated by the higher A-W interfacial tension compared to that of the oil-water interface in emulsions). This has led to the search for suitable solid particles that can fulfil this role. So far, the range of *food-grade* solids seem to be limited, so compromises may need to be made in terms of softer particles and/or loose particle aggregates to achieve adequate stability. At the same time, some degree of chemical modification, to impart the desired wetting characteristics to the particles, might need to be employed, although it should be recognized that there is consumer pressure to resist all such modifications. Physical modification, e.g.,

heating/cooling, shearing, sonication and complex formation between different ingredients via physical forces, are much more desirable and acceptable.

Secondly, higher gas-water tensions make bubble disruption under shear much more difficult, i.e., high capillary number (= dynamic viscosity x characteristic velocity/ interfacial tension), so that it is generally hard anyway to achieve the same sort of bubble sizes as in homogenization of oil-water systems, particularly if the viscosity of the continuous phase is increased by inclusion of high dissolved solids or thickening agents. Higher bulk viscosity may slow down bubble creaming, drainage and bubble collisions that normally accelerate coalescence, but high viscosity also slows adsorption of the surface active agent to the bubble surfaces. This is particularly relevant for Pickering foams, where the diffusion coefficient of particles and aggregates is already much lower than that of detergent-like molecules or even small globular proteins. Indeed, modelling work [4,5] has shown that it maybe almost impossible to *form* directly high volume fractions of micron-sized Pickering-stabilized air bubbles in any practical system, since such small bubbles will dissolve away before they achieve adequate particle surface coverage. The alternative route is to form larger bubbles initially then allow them to shrink to smaller size, wherein their particle coverage simultaneously increases if the particles already at the interface are irreversibly adsorbed due to their high desorption energy.

The above issues should be born in mind when considering the various new approaches discussed in the following review, some of which have already been discussed in other summaries elsewhere [6]. It is probably also worth stating at the start that two phenomena should be distinguished, since confusion frequently arises in the literature: foamability versus foam stability. Foamability is the capacity for foam formation: systems that have high foamability form large volumes of foam relative to the starting non-foamed liquid, i.e., high overrun. This does not necessarily mean that these foams will be stable for very long. Foam stability is the capacity of a foam, once formed, to persist. Throughout the following, unless explicitly stated, for brevity we will using the word 'foaming' to imply both foamability and foam stability. Bureiko et al. [7] have considered these issues in comparing the foaming of low molecular *versus* polymeric surfactants, like proteins, plus the competition between them in mixtures. The main classes and sub-class of foam stabilizer covered in this review are summarized in Table 1.

Table 1. 'Index' of the main classes and sub-classes of foam stabilizer discussed in this review.

Main class of stabilizer	Sub-class and corresponding reference numbers
egg white protein (EWP)	reviews 6, 75● general 10, 12, 13, 45, 46, 57, 63● + sugars & polysaccharides 14, 15, 16, 19 + wheat gluten hydrolysates 20, 21
other proteins	sodium caseinate 64● whey protein isolate 17, 102● pea, soy & lentil 18, 47, 48, 49, 59, 62

	wheat gluten & gliadins 20, 21, 22, 36 algal 58; mung bean 60; coconut 53; Quorn® mycoprotein 61 hydrophobin 34●
protein aggregates	β-lactoglobulin 24, 25, 38 lentil 59, 62 soy 49, 94
particles (Pickering)	reviews 28, 29, 30, 33 gluten hydrolysates 20, 21, 22; gliadins 36, 40● zein 39, 40●; hydrophobin 34● chitin 68●; cellulose 69, 70; starch granules 65 saponins 71-75 microgels 31, 32●, 82, CaCO ₃ 83, 84● 'bacteria' 78-79, 81
protein hydrolysates	β-lactoglobulin 51, 56 oat 54● coconut 53, 60 pea 103 EWP 57
covalently cross-linked	α-lactalbumin 52 ovalbumin 13 Na caseinate, OSA-starch & tannic acid 64●
ultrasound-treated proteins	EWP 45, 46 faba bean 47
homogenized proteins	milk 42 pea protein 44
physical complexes & coacervates	whey protein + κ-carageenan 66 whey protein+ alginate 67 lentil protein + guar gum 62 pea protein + OSA-starch 63●
OSA-starch	63●, 64●, 79
fatty alcohols (oleogels/foams)	2●, 97, 98●, 99

Protein-stabilized systems

Egg white protein

Proteins, on their own or combined with other ingredients (see later), still comprise the most widespread foaming agents. It is remarkable that, the proteinaceous culinary foaming agent *par excellence* – egg white, is still the subject of intense investigation. This is connected with our incomplete understanding of the unique stabilizing properties of egg white protein (EWP) and its widespread traditional use in many foamed products, e.g., meringues, mousses, baked goods. At the same time there is strong drive to find plant-based alternatives to this ‘animal-based’ proteinaceous foaming agent, for reasons of environmental sustainability and the relative high proportion of individuals who are allergic to EWP.

The special characteristics of egg albumen stem from the fact that it consists of a mixture of proteins each of which themselves has some less common characteristics: (i) lysozyme, that has a relatively high positive charge at neutral pH, so that it can form electrostatic complexes (aggregates) with the majority of the other (negatively charged) EWP; (ii) ovomucin, a fibrous gelling agent that cannot be denatured up to at least 100 °C that imparts viscoelasticity to the albumen itself; (iii) ovalbumin, the major EWP, a glycoprotein that seems to have a tendency to spontaneous aggregate on adsorption the A-W interface [8],[9]. Daugelaite et al. [10]● have used new, high resolution electrical resistivity measurements as a function of height to investigate free drainage of EWP foams and concluded that the presence of protein aggregates in the Plateau borders has a significant beneficial effect in reducing foam drainage and increasing foam life-times. On the other hand, this tendency for aggregation in the bulk can reduce foamability.

These special features of EWP have lead to a number of treatments to further improve EWP foaming properties, reviewed recently by Gharbi and Labbafi [11]●. A novel treatment is wet ball milling [12], although this seemed to reduce foamability and foam stability due to increased disulfide- and surface hydrophobic-induced aggregation. Similar deliberately induced cross-linking prior to foaming was investigated by Li et al. [13], using hydroxyl radical-induced oxidation of ovalbumin. Moderate oxidation improved the foaming but excessive oxidation reduced it.

In many real products, thickening agents are often added to EWP to try and control the rate of film drainage, particularly when the viscosity of the aqueous phase is already high due the presence of high concentrations of dissolved solids – typically sugars in confectionery products. The higher initial viscosity slows down the incorporation of air to high overrun and inhibits the formation of small enough bubbles. Thickeners, typically polysaccharides, help reduce bubble creaming and drainage whilst the bubble size distribution is reduced to lower sizes by mechanical whipping. They also reduce drainage of the foam product on longer storage times after whipping has ceased, via high values of the low shear rate limiting viscosity or weak gel formation. Understandably, finding the optimum balance of the formulation and processing parameters to give the final desired bubble size distribution, texture, appearance, etc., is not simple and many workers have tried to explain the interaction of all these factors via multi-dimensional response surface studies [14-18]. Ptaszek et al. [19] have shown that a log-normal distribution provides the best description of the bubble size distribution in EWP foamed with pectin and xanthan, confirming the same turbulent eddy-induced Komolgorov mechanism of colloid breakup.

As mentioned above, there is increased interest in substituting EWP with plant-based alternatives. Wouters et al. [20] have shown some improvement in EWP foaming properties by partial substitution with enzymatically hydrolyzed wheat gluten. Gluten hydrolysates had much higher foamability than EWP, whilst EWP showed much higher foam stability than the hydrolysates, but mixtures gave both good foamability and foam stability, suggesting both materials were compatible at the interface. Further measurements of the surface pressure, surface dilatational elasticity (ε) and dilatational viscosity (κ) suggested that the lower molecular weight hydrolysates might dominate the interface with the EWP forming a secondary adsorbed protein layer. Wouters et al. [21] also showed that enzymatically hydrolyzed wheat gluten gives meringues with higher overrun than EWP in a standard recipe (i.e., including sugar) with no notable disadvantages in texture or appearance. More limited gluten hydrolysis seemed to be an advantage due to the formation of more hydrophobic peptides that exhibit higher ε [22].

In many of the studies referred to above and in those that follow, researchers seek to find a link between the interfacial properties of the adsorbed film and bubble stability. The link is not always easy to make. Certainly a high value of ε will help prevent bubble coalescence in the early stages of bubble formation, via the Gibbs- Marangoni mechanism, and many proteins are superior to LMWS in this respect, though the effect is lessened as the kinetics of adsorption increase. Once adsorbed, most proteins are also able to form a thin, viscoelastic, crossed-linked network that can resist further bubble deformation, up to a point. This can aid resistance to spontaneous coalesce but can lead to film fracture under the large deformations imposed by subsequent processing. Discussion of these aspects is not new [9] and beyond the scope of this review, but the recent study of Mezdour et al. [23]● is worthy of mention in this respect. Here the foaming of protein and LMWS solutions were compared under conditions where the equilibrium surface tensions and bulk viscosity were kept constant. Thus the main variables were thought to be the interfacial viscoelasticity and dynamics of surfactant adsorption/desorption. The main conclusion was that higher dynamic dilatational viscoelasticity and more rapid diffusion kinetics resulted in foams with smaller, more stable bubbles. These effects, plus the effect of shear rate in an industrial type mechanical aerators were used to develop a model to predict bubble size.

Protein aggregates

The tendency for EWP to form aggregates, referred to above, may be one explanation of its excellent foam stabilizing properties, the aggregates providing a thicker and more mechanically resistant film that better resists thin film disruption and provides enhanced steric stabilization. This is providing the aggregates are still surface active and their larger size does not slow down too much their coverage of the A-W interface. In this respect, one might consider EWP as naturally providing something closer to Pickering-type stabilization, although the stabilizing particles are much more complex than traditional Pickering stabilizers. Dombrowski et al. [24,25] have studied the foaming properties of soluble heat-induced aggregates of pure β -lactoglobulin up to sizes of 1 μm . In general, foam stability increased with aggregate size, due to increased protein surface hydrophobicity, and correlated with maxima in the ε and κ . Hu et al. [26] reported similar findings with β -lactoglobulin aggregates of different size and shape - more fibrillar aggregates gave the highest values of

ε and foam stability. The effects of steam pressure and nozzle design on barista-style milk foams as noted by Jimenez-Junca et al. [27] are undoubtedly related to the different extents of milk protein aggregation induced.

Protein-based nanoparticles and microgels

Another way of improving the foaming properties of EWP is to convert the protein into protein microgels [28-30]. For example, Li et al. [31] created submicron (360 nm) EWP microgel (EWPM) particles by homogenization of a thermally cross-linked EWP bulk gel and showed that the EWPM-stabilized aqueous foams via a Pickering-type mechanism. Foamability was less good than EWP-stabilized foams but the EWPM stabilized foams exhibited higher stability to disproportionation and coalescence, in agreement with higher interfacial shear viscosity (η_i) of adsorbed films of EWPM, that were less brittle and liable to fracture. Further work [32]● showed that combinations of EWP and EWPM could achieve the same foamability as EWP alone, whilst still improving stability to bubble shrinkage and coalescence. At the same time, the higher the ratio of EWPM to EWP in the mixtures, the more stable were the foams to freeze-thaw, oven-heating and even microwave heating. The improvement with the mixtures is similar to that obtained with mixtures of gluten hydrolysate and EWP referred to above [20], although here the EWP was not present as microgels but natural aggregates. One might ask what is the difference between a large fractal-type aggregate formed by bottom-up self-assembly of protein molecules and a microgel particle? However, a particle formed by top-down fragmentation of a bulk gel might be expected to retain *internally* the percolating network structure of its parent gel, whilst fractal aggregates do not necessarily have a percolating structure or hold large quantities of solvent. In this review we have reserved the term microgel for those particles that are specifically referred to as such by the authors. The deliberate formation of biopolymer-based particles and aggregates, including microgels, as stabilizing agents of foams (and emulsions) has been reviewed by Dickinson [30]. Microgels in particular have also been reviewed by Murray [29].

The use of protein microgel particles as foam stabilizers exploits the natural superiority of Pickering stabilization, whilst microgels may have other advantages (and disadvantages) compared to traditional solid Pickering stabilizers [28-30,33]. At the same time, other workers have sought to create or exploit other novel protein-based particles as stabilizers that may be considered insoluble in water, or have re-appraised existing proteins as particulate stabilizers. The latter include casein micelles, that may form structures in thin films that significantly curtail drainage [33] and hydrophobin, which is able to form extremely stable small bubbles that can also block drainage of water through the Plateau borders [34]●. The only barrier to use of hydrophobins in foods might be whether or not it represents a significant allergenic potential [35]. Peng et al. [36] created gliadin nanoparticles (diameter \approx 100 – 200 nm) and showed that foaming decreased as the particles became more charged, due to slower adsorption and weaker dynamic dilatational moduli at the A-W interface, whilst effects of varying particle size were negligible. This highlights again the importance of obtaining adequate bubble coverage fast enough with particles

In terms of the minimum bubble surface coverage required for Pickering stabilization, it is well known that particles with a higher aspect ratio than spheres are far more effective. Self-assembly of protein fibrils produced by heating dilute solutions of proteins at low pH have been investigated by various workers and reviewed recently by Mohammadian and

Madadlou [37]●. Thus Peng et al. [38] have investigated β -lactoglobulin fibrils and found that film strength and bubble stability is a maximum close the pI, where the absorbed films have highest dilatational moduli but at the expense of foamability. Raising the pH slightly above the pI and re-introducing some charge to the fibrils improved foamability whilst maintaining good foam stability.

One protein that naturally forms very water-insoluble particles (diameter \approx 100 nm) is the maize storage protein zein and various workers have investigated zein as a natural Pickering stabilizer. Zou et al. [39] found that foams stabilized by zein alone broke down completely in less than 24 h, but if the anionic surfactant sodium dodecyl sulfate (SDS) was added the zein formed fractal clusters of much larger size that give much improved foam stability. Excess SDS reduced stability as adsorption became totally dominated by the LMWS, illustrating another generic difficulty with particle stabilizers. Wouters et al. [40]● also studied zein nanoparticles (size ca. 100 to 150 nm) and also when mixed with wheat gliadin nanoparticles (size ca. 130 to 140 nm), both produced via antisolvent precipitation. In agreement with Zou et al. [39], zein on its own showed very poor foaming, whilst the gliadin on its own was excellent and exhibited much higher surface dilatational moduli. Mixtures were generally worse than the equivalent gliadin concentration alone. This points to the fact that highly hydrophobic and insoluble particles, like the zein, may adsorb strongly, but if there are no cohesive forces between the particles, then the film has little resistance to expansion, as noted earlier for hydrophobins [41].

Modified proteins (physical and chemical)

Physical modification

In this section we will discuss work that has been done to deliberately modify the foaming properties of proteins by physical and chemical means. We have tried to separate out studies of mixtures of proteins with other non-proteinaceous materials, discussed in the following section, although non-covalent complex formation between proteins and other ingredients can still be thought of as a physical modification. We note again that only physical modification is likely to be accepted by regulatory bodies and consumers and also that, much of the work on modified proteins involves plant proteins, in the search for replacements for animal-derived proteins.

Tran et al. [42] have summarized the effects of high-pressure-jet processing on milk proteins in relation to their foaming properties. Apparently this can dissociate casein micelles and denature whey proteins, affecting the bulk viscosity and increasing, in particular, the foamability of whole milk. There is debate about how far the hydrodynamic forces exerted on proteins during their passage through high pressure homogenization can irreversibly affect their conformation [43]●● and therefore how this might affect their ensuing emulsification or foaming properties. It seems obvious that the larger the starting protein aggregates, the more likely it is that such aggregates will be disrupted. Djemaoune et al. [44] have shown that microfluidization only seems to affect the surface activity of pea albumin aggregates when the pH is such that the proteins are in a more compact form. It should always be remembered that the effect of high shear forces on emulsions can sometimes be related to minute contamination with the free-radical inducing transition metal elements in stainless steel.

Ultrasound treatment can also be used to effect physical modifications to proteins, although this can also induce chemical changes due free radical initiation at high power (i.e., sonochemistry). In this respect, there is a similarity with the effects of very high shear (see above), in that both physical and chemical changes might be induced. Sheng et al. [45] and Chen et al. [46] showed significant improvement in EWP foaming properties on sonication, ascribed to formation of aggregates via increased free sulfhydryl content and surface hydrophobicity. Martinez-Velasco et al. [47] used high-intensity ultrasound to increase the solubility and surface activity of faba bean protein isolate which led to improved foam stability but decreased protein digestibility.

Shao, Y. Y., et al. [48] and Wang et al. [49] showed that high quality foaming properties can be obtained by the action of much more simple physical processing, namely heating, of soy protein isolate solutions, again due to opening up of the proteins and exposing more hydrophobic groups. Ossa et al. [50] have shown that pH and temperature treatment are critical in determining the foaming properties of tofu-whey concentrates, an important 'waste' stream of tofu processing. Acidic pH values gave the best foamability and stability, ascribed to the adsorption of more aggregated protein as well as higher concentrations of soluble bean polysaccharides in the aqueous phase.

Chemical modification

Corzo-Martinez et al. [51] studied the effect of limited hydrolysis followed by glycation with galactose (via dry Maillard conjugation), and vice versa, on interfacial and foaming (foamability and foam stability) properties of β -lactoglobulin at pH 7 and pH 5. In particular, the system obtained after glycation followed by limited hydrolysis gave foams with exceptional stability, which might be related to the increase in elastic character and cohesion of the interfacial film, indicated by the increase of the surface dilatational modulus ϵ . Hardly any effect on foaming capacity was observed with the two treatments: hydrolysis followed by glycation, or *vice versa*.

Enzymes, although natural agents themselves, can be used to introduce new covalent cross-links between proteins and create new types of 'unnatural' aggregates. So, although these new aggregates may have improved foaming properties there are doubts about how easily their acceptance might be obtained for use in foods. Thus Dhayal et al. [52] have used transglutaminase or horseradish peroxidase (HRP) to cross-link α -lactalbumin to produce particles with hydrodynamic radii = 20 to 100 nm that gave improved foam stability cf. monomeric α -lactalbumin. However, the stability depended upon which enzyme was used, indicating that the location of the cross-links, i.e., the internal structure of the particles was also important (as might be expected) in terms of surface hydrophobicity, surface area. Kunarayakul et al. [53] have used a glutaminase enzyme to deamidate and improve the foaming properties of coconut protein.

The opposite approach, i.e., cleaving proteins into smaller fragment via enzymes, also continues to be extensively investigated. This has proved particularly useful for plant proteins, which tend to be relatively aggregated and water-insoluble, as demonstrated by Martinez and Pilofof [47] for sunflower protein. As a general rule any enzyme treatment of plant proteins that results in an improvement in aqueous solubility improves foaming properties, but as Bruckner-

Guhmann et al. [54]● showed with oat protein enzymatic hydrolysates, the efficacy depends on the type of enzyme via the different cleavage patterns and subsequent effects on the (mixed) peptide solubility versus pH. One might also expect that with more extensive hydrolysis the smaller peptides become less effective as steric stabilizers, but this may be mitigated by low molecular weight peptide self-assembly. Li, T., et al. [55] have shown synthetically modified dipeptides to be very effective foam stabilizers, either via the peptides self-assembling (via metal ions) into fibre-like networks at the A-W interface or in the continuous phase to form a gel.

Commercial food grade proteins may contain a range of species with different susceptibilities to the same enzyme conditions compared to native proteins. For example, Pein et al. [56] showed that limited hydrolysis of β -lactoglobulin (fractionated from a whey protein isolate) with pepsin or alcalase, increased both foamability and/or foam stability. It was suggested this was due a combination of non-hydrolysed, native β -lactoglobulin plus more surface active peptides derived from denatured β -lactoglobulin. The denatured β -lactoglobulin was presumably a result of the drying of the whey protein isolate, which made it more accessible to the protease enzymes.

The use of food enzymes to modify food proteins still presents regulatory challenges but recently Jiang et al. [57] have applied natural lactic acid fermentation to EWP and showed a modest but significant foamability, presumably due to the opening up of the proteins and exposing more hydrophobic groups. Such 'traditional' enzymatic treatments may be more acceptable.

Other Novel Plant Proteins

Plant-based, non-modified proteins that have received relatively little attention to date include algal proteins [58]. Suitably extracted protein was reportedly superior to a whey protein isolate. In addition to the large amount of work done on soy protein [48,49], other legumes and pulses have been investigated, e.g., lentil protein aggregates [59] and mung bean proteins [60]. Lonchamp et al. [61] have investigated the properties of a currently unexploited bi-product from the Quorn® fermentation process. One fraction, isolated via ultrafiltration, displayed good foamability and foam stability, although the fraction contained a wide range of metabolites and cell fragments, so that is difficult to be sure which were the key components. It is noted that, like most plant proteins, considerable physical processing is required to improve the solubility and functionality of such proteins, whilst further enzymatic and/or chemical modification might improve this further, if permissible.

Complexes

Another way to improve the foaming properties of proteins is to combine them with other ingredients that may improve the protein solubility or moderate its surface hydrophobicity – although the two things are obviously connected. If surface hydrophobicity is too high, proteins will self-aggregate in the aqueous phase rather than adsorb; if surface hydrophobicity is too low then there is little driving force for adsorption. Of course the concept of surface hydrophobicity is of little use if the proteins change their conformation significantly during adsorption, as most do. Complexes between different proteins

or proteins or polysaccharides can arise from electrostatic attraction and hydrogen bonding, whereas hydrophobic bonding may also operate between different proteins or proteins and lipids and surfactants. One can distinguish soluble complexes of dimensions similar to the constituent molecules and much larger coacervates, the latter appearing as colloidal particles in their own right. Here we also ignore any effects of proteins and polysaccharides where the polysaccharide merely acts as an independent thickening agent or induces depletion flocculation of the protein.

Jarpa-Parra et al. [62] demonstrated coacervate formation at mildly acidic pH between lentil proteins and guar gum, xanthan gum and pectin that gave much enhanced foam stability, due to an electrostatically cross-linked gel-like interfacial network. The aggregates also plugged the junctions of the Plateau borders in the same way as noted above with EWP aggregates [10] and with casein micelles [34]. Significantly, the proteins and polysaccharides phase separated segregatively at neutral pH and this led to a loss of foam stability.

Even more complicated foaming mixtures involve proteins and synthetically modified polysaccharides, such as starches modified with octenyl succinic anhydride (OSA-starches) [63,64], the latter also being surface active. Asghari et al. [63]● suggest waxy maize OSA-starch adsorption can exclude pea protein from the interface when the latter is still sufficiently aggregated to prevent enough hydrophobic protein residues accessing the interface. Zhan et al. [64]● studied an even more complex system of an OSA-starch, tannic acid and sodium caseinate, the latter being more surface active than pea protein or EWP. Tannic acid cross-links the caseinate films and makes them more rigid. Measurements of ε over a range of strains suggested that the OSA-starch increased the degree of strain softening on interfacial extension via more isolated adsorbed OSA-starch domains in between protein/tannic acid domains. With EWP, Asghari et al. [63]● suggested OSA co-adsorbs but de-mixing in the interface forces the EWP to form more closely packed regions that increases stability. These workers also compared the effect of the OSA-starches with dry heated (non-OSA treated) rice starch granules. Such dry-heating seems to make the granule surface more hydrophobic in some way [65], though it is not clear if this makes them sufficiently surface active to compete with protein adsorption. The dry-heated granules slowed down the drainage of EWP foams via some mechanism but they were not as effective as the OSA starches.

Wang et al. [66] showed that heating WPI with λ -carageenan in solution under neutral or near neutral pH can induce the formation of soluble complexes with improved functional properties compared to heating WPI then adding the carageenan afterwards. The explanation proposed was that heating the two components together resulted in enhanced electrostatic interactions between them, due to their more open molecular configurations. Xu et al. [67] have investigated similar complexes between WPI and sodium alginate. Foam stability was particularly improved (at low overall protein concentrations) at pH values where surface active soluble electrostatic complexes were formed between the oppositely net charged components at a relatively narrow (2:1 to 1:1) range of weight ratios of WPI to alginate.

All the above studies highlight the complexity of the different effects on foaming properties when there are different adsorbing and non-adsorbing macromolecules and/or particles present, although this is always likely to be the case in real food systems.

Novel, non-protein based systems

Two types of novel non-protein based foaming agents have been mainly investigated: polysaccharides that are modified to make them surface active and non-protein based particle (Pickering) stabilizers. Of the former, OSA-starch has already been mentioned above, though this has largely been used in admixture with other foaming agents, whilst with regard to the latter, polysaccharides are also relevant, in terms of water-insoluble cellulose and chitin particles.

Cellulose & chitin

Tzoumaki et al. [68]● have investigated the foaming of chitin nanocrystals. As with many Pickering foams, the importance of structuring of particles both at the interface and within the thin films between bubbles (affectively forming thin layers of a 'bulk' particle gel) was also considered. Even more work has been done on cellulose as a Pickering foam stabilizer, though debate continues how the cellulose can be made hydrophobic enough to adsorb efficiently enough at the A-W interface. Beatrice et al. [69] required free radical-oxidized nanocellulose in combination with various water-soluble (polysaccharide) thickening agents to produce adequate foaming properties. Another important observation was that, the effect of shear in these systems caused the cellulose nanofibres to flocculate and accumulate at bubble surfaces, though this depended on the attractive interaction between the cellulose and the polysaccharide thickening agents. Such aggregates can potentially act as *defoaming* agents, plus the more aggregated the particles become, the lower the surface area they can cover and so the coarser will be the foam produced. Ahmadzadeh et al. [70] combined chemically modified clay particles to improve the foaming properties of cellulose-based Pickering foams, for foam packing with enhanced physical properties.

Saponins

A relatively new set of Pickering particles that has emerged is based on the natural class of surface active molecules named saponins. Saponins have long been known to be highly surface active and excellent foaming agents in their own right, but Wan et al. [71] have shown that nanofibrils of glycyrrhizic acid can form networks at the A-W and O-W interfaces of oil droplets and bubbles as the system is emulsified and aerated simultaneously, i.e., an emulsion foam, much like whipped dairy cream. The jamming of the emulsion droplets in thin liquid films also enhances foam stability. On cooling the nanofibrils form a three-dimensional net (gel) that prevents coalescence of the droplets and bubbles within. The foam system could be destroyed rapidly on heating because of the melting of the bulk gel. In later work Ma et al. [72] showed that nanofibrils of glycyrrhizic acid alone can form multilayer networks at the A-W interface and also in the bulk aqueous phase, leading to extremely stable foams. Similarly, the foams could be rapidly destabilized by heating, via break-up of the fibrillar networks. This emulsion foam work is related to the findings of Chen et al. [73], who reported that aqueous foams stabilized by Quillaja saponin-coated 'nanodroplets' were far more stable than foams stabilized by the saponin alone, presumably because the droplets created thicker films between adjacent bubbles. The nanodroplets were of sunflower oil, prepared by sonication and with a mean size ca. 150 nm. Santini et al. [74] also investigated Quillaja saponin and mixed saponin-chitosan solutions. The results substantiated the high surface activity of the saponin and showed that it produced the high values of dilatational viscoelasticity. Another effect emphasized was that the highly

charged chitosan molecules + saponin-induced aggregates may prevent thinning of the thin liquid films between bubbles. This therefore prevents coalescence during foam formation, aiding the subsequent saponin adsorption and stabilization of the bubbles. In addition, there is the possibility of electrostatic complex formation of chitosan with adsorbed saponin, increasing electrosteric stabilization of the bubbles. Gonzalez and Sorensen [75]● have recently reviewed work on saponins and investigated the foaming behaviour of those extracted from Soapwort (*Saponaria officinalis*) in the presence of other common food ingredients (salt, sugar, ethanol) and conditions (pH and temperature). In general the foams were resistant to such formulation variations, apart from ethanol.

Whether such systems can overcome the potential toxicity (albeit in large quantities) of saponins remains to be seen, although saponins may have some health benefits in sequestering bile salts and so lowering blood cholesterol [76].

Bacteria-stabilized

It may seem odd to think of bacteria as particulate stabilizers, but it is well known that the high bacterial cell load in fermented products like yoghurt, along with the mucilages they produce, make a significant contribution to the texture and physical stability of such products. Only recently, however, have workers turned to the idea of bacterial cells themselves acting as Pickering-style particles [77,78]. Jiang et al. [79] have taken this a step further in chemically modifying the 'surface' of lactic acid bacteria (using the same octenyl succinic anhydride commonly used for starch modification) to improve their foaming properties. Although a low level of modification maintained cell viability, it is doubtful if such ingredients would be encouraged in real food stuffs. Foaming is a serious problem during anaerobic digestion of food waste and sewage streams He et al. [80], where it is believed that fatty acids generated are the main foaming agent [81]. It remains to be seen if certain bacterial cells themselves contribute to the foam stability by acting as Pickering stabilizers.

Combinations of particles and other foaming agents

The problem of particles adsorbing fast enough to the surface of gas bubbles in order to reach a high enough coverage to stabilize them [4,5] was touched upon in the Introduction. At the same time, there is the difficulty of finding native particles that have the correct wetting characteristics without resorting to chemical modification of their surfaces. Both these difficulties may be overcome via physical means by combining particles with other materials that also adsorb to the bubble surfaces and/or that adsorb to the particle surface and advantageously alter the particle contact angle at the bubble surface. In this way, particles may be made to be more strongly adsorbing or less strongly aggregating in solution and therefore more available for adsorption. Alternatively, combinations of particles with other surface active agents may impart the optimum characteristics to the adsorbed film that give rise to high foamability and foam stability. In this way, Ellis et al. [82] used the unusual cationic surfactant lauric arginate to partially neutralize the charge on k-carrageenan microgel particles so that this adsorbed more strongly but did not aggregate in solution, to improve their foaming characteristics. Similarly Binks et al. [83] used combinations of hydrophilic calcium carbonate particles and the LMWS sodium stearyl-2-lactylate to improve the stability of foams to coalescence and disproportionation. The same technique was applied [84]● to other, more acceptable food surfactants, including β -lactoglobulin, sodium caseinate, sodium dodecanoate and the Quillaja saponins mentioned earlier. However, it was found that the higher molecular weight,

structurally more complex proteins and Quillaja did not increase foam stability, only the LMWS. This is presumably because the latter adsorb 'head-down' on the inorganic particles and give 'clean' hydrophobic patches and the correct hydrophobic-hydrophilic balance for strong adsorption and a high barrier to particle desorption.

Novel applications of foams in foods

Foam mat drying

There has been increasing research into foam mat drying, where material is foamed and moisture removed by application of heat in some way. The higher surface area of the foam can have the advantage of accelerating the drying process, but mitigating against this are the low thermal conductivity of foam and also the increasing viscosity of the aqueous phase if thickening agents are employed or naturally present. Sangamithra et al. [85] have reviewed this technique, which may be particularly useful for drying fruit juices and pulps that are naturally very sticky due to the sugars they contain' but which also contain valuable colours and antioxidants [86-88] that may be easily damaged and lost in spray drying or prolonged heating. Heat may be applied in a number of ways: freezing stable foams then freeze-drying [86,87] or by application of microwave energy [89] to the wet foam, for example. The bubble size distribution, bubble packing and viscoelasticity of the thin liquid films all play a role in determining the efficiency of drying, all of which also depend on the original foam formulation.

Foam fractionation

Foam fractionation exploits the natural tendency of surface active material to accumulate at bubble surfaces and to become trapped in thin film lamellae as the foam drains. The mining industry has exploited this technique for many years in froth flotation, where foaming is used to accumulate and separate out valuable inorganic particulates via their natural tendency to adsorb at the gas-water interface or via introduction of froth flotation agents that deliberately make them do so. However, the technique potentially also has a number of food-related uses.

An interesting case is the work of Ayetigbo et al. [90] who added glycerol monostearate and sodium carboxymethyl cellulose to aid foaming of cassava pulp. In this case the foamed pulp had a significantly lower concentration of toxic cyanogens. In the opposite sense, Larson et al. [91] have shown that foaming concentrates perfluorinated surfactants that may be harmful to the environment. This alters the site-specific impact of such surfactants but potentially might be exploited to remove them from waste streams. Turning this round again, Le Toquin et al. [92]● have shown that foamed disinfectant agents are at least as effective as liquid solutions at the same concentration but generate far less toxic effluent due to lower run off. This has the potential for cleaning in place of food equipment as well as many other scenarios for disinfecting surfaces.

Foam fractionation of surface active material from a high viscosity bulk liquid is clearly more difficult and slow, but Li et al. [93] have developed a novel foam fractionation column with vertical sieve trays. This was applied to recovering yam mucilage from yam starch processing wastewater and resulted in significant enrichment. Li et al. [94] have pointed out that recovery of functionally useful protein from soy processing waste water, via foam fractionation, relies on preventing

irreversible aggregation of the protein. Interestingly, their results suggested that most insoluble aggregates were formed in the *desorption* of protein from the A-W interface, i.e., aggregation was effectively induced via protein adsorption in the first place, similar to EWP. Addition of a defoaming agent based on LMWS therefore reduced the formation of aggregates. Liu et al. [95] used foam fractionation to recover lycopene from the tomato-based processing wastewater via tomato proteins as the collectors. Moreover natural rhamnolipids could be used to enhance foam stabilization via formation of a protein-lycopene complex.

Oleogels and oleofoams

Another way of reducing the overall calorie content of a fatty product is to incorporate gas bubbles directly into the lipid phase, i.e., an oleofoam. This can be achieved by stabilizing the bubbles with oil-dispersible particles, i.e., Pickering stabilization, or via gelation of the oil-phase, i.e., physically trapping the bubbles in a network with sufficient strength to resist movement of the bubbles via gravity or Brownian motion. In the latter case, termed an oleogel [2]●, the network may be provided by fat crystals inherent to the lipid itself, or other oil-insoluble particles introduced into the system. With oleogels there may also be some adsorption of the gelling particles the bubble surfaces, so that it may not always be clear which mechanism is dominant, analogous to water droplet stabilization in margarines via fat crystals. Binks et al. [96] have provided a clear example of pure Pickering stabilization of air-in-oil (A/O) foams via fluorinated inorganic particles (clays and zinc oxide). Stability depended upon both particle wettability and the amount of shear applied, so that some systems broke down or even inverted to O/A systems. If fluorinated particles could be avoided the latter might provide some interesting opportunities for flavour oil release from the dry powders.

Solely Pickering stabilization of A/O dispersions seems to rare - highlighting the difficulty of finding food-grade particles that easily disperse in oils but that are also sufficiently surface active at the A-O interface – much more work has been done on oleogels. Fameau et al. [97] have produced A/O foams (mean bubble diameter < 200 μm) based on an oleogel of fatty alcohols in sunflower oil. The foams are also most indefinitely stable as long as the fatty alcohol crystals do not melt. An interesting idea therefore, was to make the foams light responsive by also incorporating carbon black particles. On application of UV radiation the gel rapidly warmed, the crystals melted and the foam collapsed. Fameau et al. [98]● have reviewed these and other such responsive foams, where the switch from stability to instability can be induced by changes in the bulk or interfacial material on exposure to light, temperature, and magnetic fields. Heymans et al. [99] have produced a similar A/O system but based on monoglyceride crystals.

In all the above oleogel-stabilized foams, successful stabilization depends upon the size, shape and form of the crystals, which in turn depends on the time-temperature history any other physical processing of the system, the chemical composition of the crystals and their solubility in the continuous oil phase. In other words, the fat crystallization kinetics, which is difficult to control and an area that is still not fully understood. In addition, by analogy with aqueous foams, the bulk viscosity of the continuous phase and the effect of the crystallization on this viscosity determines the ease of incorporation of gas. Higher viscosity makes incorporation more difficult (lower foamability), but once incorporated the bubbles will be more stable.

New methods of studying foam formation and stability

As outlined at the outset, foams are relatively unstable compared to many other dispersions in food and this, combined with the relatively large size of bubbles, the tendency for bubbles to cream and pack closely together separated via very thin films, makes them particularly tricky beasts to study. It is therefore of interest to highlight some new methods and techniques that have been developed to improve our understanding of foam formation and destruction.

Many of these new methods centre around the development of microfluidic devices for production of dispersions under controlled and observable conditions, as reviewed recently by Deng et al. [100]●. Labarre and Vigolo [101]●● have developed a method for evaluation of foam stability after a deformation in a microfluidic device. Whether one or more row of bubbles forms in the channels and the size of these bubbles depends on: the gas and liquid pressure drop as bubbles are formed from the flow focussing nozzle; the viscosity of the liquid phase; the surface tension; the ease of bubble deformation at the channel walls. Laporte, M., et al. [102]● applied microfluidics to produce foams stabilized by combinations of whey protein isolate and xanthan gum, showing how on a minute scale this could be used to test and predict the effects of bubble size, bubble volume fraction, continuous phase viscoelasticity and other solution conditions on bulk foam stability. Clarke et al. [103] have studied isolated foam channels created by withdrawing a specific frame geometry from surfactant solutions and using image analysis to monitor film thickness and drainage across the model Plateau borders created. Drainage for aqueous solutions of food grade LMWS, hydroxypropyl methylcellulose and hydrolyzed pea protein were studied. Novel distortions in the border thickness profiles were exhibited with the polymeric surfactants, ascribed to the much higher η_1 produced with such stabilizers. In similar ways, the flow of bulk foam down microchannels can be tested, where wall slip or 'partial coalescence' with the wall are key factors controlling the flow, particularly for bubbles stabilized by particles [104]. Increasing particle coverage creates a more rigid and rough bubble surface that increases the frictional resistance to slip at the microchannel wall. Such effects can also be used to inhibit liquid ingress into narrow channels and capillaries [105].

Zhang et al. [106]●● reported a 'new' type of particle-stabilized foam material, capillary foam, which is stabilized by the synergistic action of particles and a small amount of an immiscible liquid - typically an oil for aqueous-based foams. The stabilization mechanism relies on the wetting of particles by the oil to some extent, which results in them sticking together via capillary bridges. The capillary foam is then formed by aeration of the flocculated dispersion. Alternatively, a particle stabilized foam can be formed first, then the oil added and the system re-agitated to distribute the oil to the particles. In this way, one indirectly tunes the capacity of the particles to stabilize the air-water interface via the particle affinity for the air-oil and oil-water interfaces and the spreading of a thin oil film around the bubbles. This results in foams that are then extremely stable. Oil soluble dyes can be added to make the foam appear highly coloured, or the oil film cured (solidified) to produce highly stiff, load-bearing dry foams. Whilst such systems have been deliberately designed, the effects are also important because of the inadvertent interaction of Pickering foams with oils in their potential usage as enhanced (crude) oil recovery agents and their sealing of capillaries and cracks as discussed above [104,105]. Simulating and visualizing

the flow behaviour of such complex systems in order to understand them better also remains a great challenge, as discussed by Lipsa et al. [107].

Conclusions

Research into novel stabilizers of food foams continues apace, particularly with regard to particulate stabilizers (Pickering foams). Most of these systems still employ proteins as the key stabilizing ingredient, either in the form of particles of some type, or as modifiers of the surface properties of other particulate material. Many of these studies seek to include stable air bubbles as bulking agents and calorie-lowering agents, at the same time seeking to find replacements for traditional stabilizers based on animal proteins (e.g., dairy and egg proteins). Whilst in most food foams the bulk liquid phase is aqueous, interesting work on air-in-oil foams (oleofoams and oleogels) represent additional novel ways of influencing food texture and stability for the benefit of consumers.

References

1. Van Der Vegt A, Bisschop HA: **Sweet particulate fat-containing powder, its preparation and its use.** *Patent WO2012050439A1* 2012.
- 2. Heymans R, Tavernier I, Dewettinck K, Van der Meeren P: **Crystal stabilization of edible oil foams.** *Trends in Food Science & Technology* 2017, **69**:13-24.
A good review of the expanding field of oleofoams and oleogels.
3. Valadbaigi P, Ettelaie R, Kulak AN, Murray BS: **Generation of ultra-stable Pickering microbubbles via polyalkylcyanoacrylates.** *Journal of Colloid and Interface Science* 2019, **536**:618-627.
4. Ettelaie R, Murray B: **Effect of particle adsorption rates on the disproportionation process in pickering stabilised bubbles.** *Journal of Chemical Physics* 2014, **140**.
5. Ettelaie R, Murray BS: **Evolution of bubble size distribution in particle stabilised bubble dispersions: Competition between particle adsorption and dissolution kinetics.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2015, **475**:27-36.
6. Narsimhan G, Xiang N: **Role of Proteins on Formation, Drainage, and Stability of Liquid Food Foams.** In *Annual Review of Food Science and Technology, Vol 9*. Edited by Doyle MP, Klaenhammer TR; 2018:45-63. *Annual Review of Food Science and Technology, vol 9.*
7. Bureiko A, Trybala A, Kovalchuk N, Starov V: **Current applications of foams formed from mixed surfactant-polymer solutions.** *Advances in Colloid and Interface Science* 2015, **222**:670-677.
8. Lechevalier V, Croguennec T, Pezennec S, Guerin-Dubiard C, Pasco M, Nau F: **Evidence for synergy in the denaturation at the air-water interface of ovalbumin, ovotransferrin and lysozyme in ternary mixture.** *Food Chemistry* 2005, **92**:79-87.
9. Murray BS: **Rheological properties of protein films.** *Current Opinion in Colloid & Interface Science* 2011, **16**:27-35.
- 10. Daugelaite D, Guillermic RM, Scanlon MG, Page JH: **Quantifying liquid drainage in egg-white sucrose foams by resistivity measurements.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2016, **489**:241-248.
Based on more accurate measurements of electrical conductivity across a column of foam as function of height, this method could find wide application in many other systems.
- 11. Gharbi N, Labbafi M: **Influence of treatment-induced modification of egg white proteins on foaming properties.** *Food Hydrocolloids* 2019, **90**:72-81.
A useful review of the ongoing modifications to egg white protein.

12. Li M, Li MM, Tan W, Liu XB, Duan X: **Effects of ball-milling treatment on physicochemical and foaming activities of egg ovalbumin.** *Journal of Food Engineering* 2019, **261**:158-164.
 13. Li SG, Huang Y, An FP, Huang Q, Geng F, Ma MH: **Hydroxyl radical-induced early stage oxidation improves the foaming and emulsifying properties of ovalbumin.** *Poultry Science* 2019, **98**:1047-1054.
 14. Sadahira MS, Rodrigues MI, Akhtar M, Murray BS, Netto FM: **Effect of egg white protein-pectin electrostatic interactions in a high sugar content system on foaming and foam rheological properties.** *Food Hydrocolloids* 2016, **58**:1-10.
 15. Sadahira MS, Rodrigues MI, Akhtar M, Murray BS, Netto FM: **Influence of pH on foaming and rheological properties of aerated high sugar system with egg white protein and hydroxypropylmethylcellulose.** *Lwt-Food Science and Technology* 2018, **89**:350-357.
 16. Sadahira MS, Lopes FCR, Rodrigues MI, Yamada AT, Cunha RL, Netto FM: **Effect of pH and interaction between egg white protein and hydroxypropylmethylcellulose in bulk aqueous medium on foaming properties.** *Carbohydrate Polymers* 2015, **125**:26-34.
 17. Laporte M, Della Valle D, Loisel C, Marze S, Riaublanc A, Montillet A: **Rheological properties of food foams produced by SMX static mixers.** *Food Hydrocolloids* 2015, **43**:51-57.
 18. Moll P, Grossmann L, Kutzli I, Weiss J: **Influence of energy density and viscosity on foam stability - A study with pea protein (*Pisum Sativum* L.).** *Journal of Dispersion Science and Technology* 2019.
 19. Ptaszek P, Kabzinski M, Kruk J, Kaczmarczyk K, Zmudzinski D, Liszka-Skoczylas M, Mickowska B, Lukaszewicz M, Banas J: **The effect of pectins and xanthan gum on physicochemical properties of egg white protein foams.** *Journal of Food Engineering* 2015, **144**:129-137.
 20. Wouters AGB, Rombouts I, Fierens E, Brijs K, Blecker C, Delcour JA, Murray BS: **Foaming and air-water interfacial characteristics of solutions containing both gluten hydrolysate and egg white protein.** *Food Hydrocolloids* 2018, **77**:176-186.
 21. Wouters AGB, Rombouts I, Fierens E, Brijs K, Delcour JA: **Enzymatically Hydrolyzed Wheat Gluten as a Foaming Agent in Food: Incorporation in a Meringue Recipe as a Proof-of-Concept.** *Journal of Food Science* 2018, **83**:2119-2126.
 22. Wouters AGB, Rombouts I, Legein M, Fierens E, Brijs K, Blecker C, Delcour JA: **Air-water interfacial properties of enzymatic wheat gluten hydrolyzates determine their foaming behavior.** *Food Hydrocolloids* 2016, **55**:155-162.
 - 23. Mezdour S, Seguineau de Preval E, Granda P, Cuvelier G, Ducept F: **Impact of Interfacial Characteristics on Foam Structure: Study on Model Fluids and at Pilot Scale.** *Oil & Gas Science and Technology-Revue D Ifp Energies Nouvelles* 2017, **72**.
- Not many studies have tried to compare proteins and low molecular weight surfactants, like with like, i.e., where the effects of bulk viscosity and equilibrium tension are equal, but this work does, allowing much clearer interpretation of the differences between them.
24. Dombrowski J, Jöhler F, Warncke M, Kulozik U: **Correlation between bulk characteristics of aggregated beta-lactoglobulin and its surface and foaming properties.** *Food Hydrocolloids* 2016, **61**:318-328.
 25. Dombrowski J, Gschwendtner M, Kulozik U: **Evaluation of structural characteristics determining surface and foaming properties of beta-lactoglobulin aggregates.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2017, **516**:286-295.
 26. Hu J, Yang JX, Xu Y, Zhang K, Nishinari K, Phillips GO, Fang YP: **Comparative study on foaming and emulsifying properties of different beta-lactoglobulin aggregates.** *Food & Function* 2019, **10**:5922-5930.
 27. Jimenez-Junca C, Sher A, Gumy JC, Niranjana K: **Production of milk foams by steam injection: The effects of steam pressure and nozzle design.** *Journal of Food Engineering* 2015, **166**:247-254.
 28. Murray BS: **Pickering Emulsions for Food and Drinks** *Current Opinion in Food Science* 2019, **27**:57-63.
 29. Murray BS: **Microgels at fluid-fluid interfaces for food and drinks.** *Advances in Colloid and Interface Science* 2019, **271**:101990.

30. Dickinson E: **Biopolymer-based particles as stabilizing agents for emulsions and foams.** *Food Hydrocolloids* 2017, **68**:219-231.
31. Li X, Murray BS, Yang YJ, Sarkar A: **Egg white protein microgels as aqueous Pickering foam stabilizers: Bubble stability and interfacial properties.** *Food Hydrocolloids* 2020, **98**.
- 32. Li X, Yang YJ, Murray BS, Sarkar A: **Combination of egg white protein and microgels to stabilize foams: Impact of processing treatments.** *Journal of Food Engineering* 2020, **275**.

Not many other studies of microgel-stabilized foams have subjected the systems to such extreme conditions. These results highlight the great potential of other protein microgels to mimic egg white protein.

33. Dickinson E: **Structuring of colloidal particles at interfaces and the relationship to food emulsion and foam stability.** *Journal of Colloid and Interface Science* 2015, **449**:38-45.
- 34. Dimitrova LM, Petkov PV, Kralchevsky PA, Stoyanov SD, Pelan EG: **Production and characterization of stable foams with fine bubbles from solutions of hydrophobin HFBII and its mixtures with other proteins.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2017, **521**:92-104.

This later work on more applied aspect of hydrophobin highlights again the highly unseal properties of this protein, which acts like a natural Pickering agent for foams, whilst supplementation with other proteins may extend it possible usage.

35. Yew SM, Chan CL, Ngeow YF, Toh YF, Na SL, Lee KW, Hoh CC, Yee WY, Ng KP, Kuan CS: **Insight into different environmental niches adaptation and allergenicity from the *Cladosporium sphaerospermum* genome, a common human allergy-eliciting Dothideomycetes.** *Scientific Reports* 2016, **6**.
36. Peng DF, Jin WP, Tang C, Lu Y, Wang WQ, Li J, Li B: **Foaming and surface properties of gliadin nanoparticles: Influence of pH and heating temperature.** *Food Hydrocolloids* 2018, **77**:107-116.
- 37. Mohammadian M, Madadlou A: **Technological functionality and biological properties of food protein nanofibrils formed by heating at acidic condition.** *Trends in Food Science & Technology* 2018, **75**:115-128.

A very useful review of this highly topical area, which has implications in many other areas outside foods.

38. Peng DF, Yang JC, Li J, Tang C, Li B: **Foams Stabilized by beta-Lactoglobulin Amyloid Fibrils: Effect of pH.** *Journal of Agricultural and Food Chemistry* 2017, **65**:10658-10665.
39. Zou Y, Wan ZL, Guo J, Wang JM, Yin SW, Yang XQ: **Modulation of the surface properties of protein particles by a surfactant for stabilizing foams.** *Rsc Advances* 2016, **6**:66018-66026.
- 40. Wouters AGB, Schaefer S, Joye IJ, Delcour JA: **Relating the structural, air-water interfacial and foaming properties of wheat (*Triticum aestivum* L.) gliadin and maize (*Zea mays* L.) zein based nanoparticle suspensions.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2019, **567**:249-259.

This paper clearly illustrates that high surface hydrophobicity and solid particle type nature are not necessarily an advantage – proteins particles must unfold and cross-link to some extent to give a coherent film that can stabilize bubbles against coalescence and disproportionation.

41. Wang YW, Bouillon C, Cox A, Dickinson E, Durga K, Murray BS, Xu R: **Interfacial Study of Class II Hydrophobin and Its Mixtures with Milk Proteins: Relationship to Bubble Stability.** *Journal of Agricultural and Food Chemistry* 2013, **61**:1554-1562.
42. Tran M, Roberts R, Felix TL, Harte FM: **Effect of high-pressure-jet processing on the viscosity and foaming properties of pasteurized whole milk.** *Journal of Dairy Science* 2018, **101**:3887-3899.
- 43. Dobson J, Kumar A, Willis LF, Tuma R, Higazi DR, Turner R, Lowe DC, Ashcroft AE, Radford SE, Kapur N, et al.: **Inducing protein aggregation by extensional flow.** *Proceedings of the National Academy of Sciences of the United States of America* 2017, **114**:4673-4678.

Although not directed at foods or even foams, this detailed study of the effect of extensional flow on the conformational change of several physiologically relevant blood proteins shows that, even at relatively low protein concentrations, thermodynamically and kinetically stable proteins can be readily unfolded. This raises many questions about the effects of different stirring conditions on food protein surface activity.

44. Djemaoune Y, Cases E, Saurel R: **The Effect of High-Pressure Microfluidization Treatment on the Foaming Properties of Pea Albumin Aggregates.** *Journal of Food Science* 2019, **84**:2242-2249.
45. Sheng L, Wang YB, Chen JH, Zou J, Wang Q, Ma MH: **Influence of high-intensity ultrasound on foaming and structural properties of egg white.** *Food Research International* 2018, **108**:604-610.
46. Chen YX, Sheng L, Gouda M, Ma MH: **Impact of ultrasound treatment on the foaming and physicochemical properties of egg white during cold storage.** *Lwt-Food Science and Technology* 2019, **113**.
47. Martinez KD, Pilosof AMR: **Foaming behaviour of enzymatically modified sunflower protein in proximity to pl.** *Biointerface Research in Applied Chemistry* 2017, **7**:1883-1886.
48. Shao YY, Lin KH, Kao YJ: **Modification of Foaming Properties of Commercial Soy Protein Isolates and Concentrates by Heat Treatments.** *Journal of Food Quality* 2016, **39**:695-706.
49. Wang MP, Chen XW, Guo J, Yang J, Wang JM, Yang XQ: **Stabilization of foam and emulsion by subcritical water-treated soy protein: Effect of aggregation state.** *Food Hydrocolloids* 2019, **87**:619-628.
50. Ossa JSH, Wagner JR, Palazolo GG: **Influence of chemical composition and structural properties on the surface behavior and foam properties of tofu-whey concentrates in acid medium.** *Food Research International* 2020, **128**.
51. Corzo-Martinez M, Moreno FJ, Villamiel M, Patino JMR, Sanchez CC: **Effect of glycation and limited hydrolysis on interfacial and foaming properties of bovine beta-lactoglobulin.** *Food Hydrocolloids* 2017, **66**:16-26.
52. Dhayal SK, Delahaije R, de Vries RJ, Gruppen H, Wierenga PA: **Enzymatic cross-linking of alpha-lactalbumin to produce nanoparticles with increased foam stability.** *Soft Matter* 2015, **11**:7888-7898.
53. Kunarayakul S, Thaiphant S, Anprung P, Suppavorasatit I: **Optimization of coconut protein deamidation using protein-glutaminase and its effect on solubility, emulsification, and foaming properties of the proteins.** *Food Hydrocolloids* 2018, **79**:197-207.
- 54. Bruckner-Guhmann M, Heiden-Hecht T, Sozer N, Drusch S: **Foaming characteristics of oat protein and modification by partial hydrolysis.** *European Food Research and Technology* 2018, **244**:2095-2106.
- This study illustrates nicely the difficulties of understanding or predicting the effects of enzymatic hydrolysis on protein functional properties, which is dependent on so many external (solution) and intrinsic (state of protein unfolding and or/ aggregation) factors.
55. Li T, Nudelman F, Tavacoli JW, Vass H, Adams DJ, Lips A, Clegg PS: **Long-Lived Foams Stabilized by a Hydrophobic Dipeptide Hydrogel.** *Advanced Materials Interfaces* 2016, **3**.
56. Pein D, Clawin-Radecker I, Lorenzen PC: **Peptic treatment of beta-lactoglobulin improves foaming properties substantially.** *Journal of Food Processing and Preservation* 2018, **42**.
57. Jiang YW, Jia J, Xiong DD, Xu XM, Yang YJ, Liu XB, Duan X: **Effects of short-term fermentation with lactic acid bacteria on egg white: Characterization, rheological and foaming activities.** *Food Hydrocolloids* 2020, **101**.
58. Buchmann L, Bertsch P, Bocker L, Krahenmann U, Fischer P, Mathys A: **Adsorption kinetics and foaming properties of soluble microalgae fractions at the air/water interface.** *Food Hydrocolloids* 2019, **97**.
59. Jarpa-Parra M, Bamdad F, Tian Z, Zeng HB, Temelli F, Chen L: **Impact of pH on molecular structure and surface properties of lentil legumin-like protein and its application as foam stabilizer.** *Colloids and Surfaces B-Biointerfaces* 2015, **132**:45-53.
60. Widjajaseputra AI, Widyastuti TEW: **Potential of coconut milk and mung bean extract combination as foam stabilizer in non-dairy ice cream.** *International Food Research Journal* 2017, **24**:1199-1203.
61. Lonchamp J, Clegg PS, Euston SR: **Foaming, emulsifying and rheological properties of extracts from a co-product of the Quorn fermentation process.** *European Food Research and Technology* 2019, **245**:1825-1839.
62. Jarpa-Parra M, Tian Z, Temelli F, Zeng HB, Chen L: **Understanding the stability mechanisms of lentil legumin-like protein and polysaccharide foams.** *Food Hydrocolloids* 2016, **61**:903-913.

- 63. Asghari AK, Norton I, Mills T, Sadd P, Spyropoulos F: **Interfacial and foaming characterisation of mixed protein-starch particle systems for food-foam applications.** *Food Hydrocolloids* 2016, **53**:311-319.
- 64. Zhan FC, Li J, Shi MQ, Wu D, Li B: **Foaming Properties and Linear and Nonlinear Surface Dilatational Rheology of Sodium Caseinate, Tannin Acid, and Octenyl Succinate Starch Ternary Complex.** *Journal of Agricultural and Food Chemistry* 2019, **67**:2340-2349.

This study and the one above (Asghari et al. [60]) illustrate the potential interfacial incompatibility of the two very different surface active macromolecular species – proteins and hydrophobically modified polysaccharides (in the case OSA-starch) and the different direction in which this can force foam stability.

- 65. Nakagawa M, Tabara A, Ushijima Y, Matsunaga K, Seguchi M: **Hydrophobicity of stored (15, 35 degrees C), or dry-heated (120 degrees C) rice flour and deteriorated breadmaking properties baked with these treated rice flour/fresh gluten flour.** *Bioscience Biotechnology and Biochemistry* 2016, **80**:983-990.
- 66. Wang ZS, Zhang S, Vardhanabhuti B: **Foaming Properties of Whey Protein Isolate and lambda-Carrageenan Mixed Systems.** *Journal of Food Science* 2015, **80**:N1893-N1902.
- 67. Xu Y, Yang N, Yang JX, Hu J, Zhang K, Nishinari K, Phillips GO, Fang YP: **Protein/polysaccharide intramolecular electrostatic complex as superior food-grade foaming agent.** *Food Hydrocolloids* 2020, **101**.
- 68. Tzoumaki MV, Karefyllakis D, Moschakis T, Biliaderis CG, Scholten E: **Aqueous foams stabilized by chitin nanocrystals.** *Soft Matter* 2015, **11**:6245-6253.

Whilst much work has been done recently on cellulose, chitin also represents a much under-utilized source of insoluble food grade particles. This work introduces this topic nicely, focusing on food-grade modifications.

- 69. Beatrice CAG, Rosa-Sibakov N, Lille M, Sozer N, Poutanen K, Ketoja JA: **Structural properties and foaming of plant cell wall polysaccharide dispersions.** *Carbohydrate Polymers* 2017, **173**:508-518.
- 70. Ahmadzadeh S, Nasirpour A, Keramat J, Hamdami N, Behzad T, Desobry S: **Nanoporous cellulose nanocomposite foams as high insulated food packaging materials.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2015, **468**:201-210.
- 71. Wan ZL, Sun YE, Ma LL, Zhou FB, Guo J, Hu SQ, Yang XQ: **Long-Lived and Thermoresponsive Emulsion Foams Stabilized by Self-Assembled Saponin Nanofibrils and Fibrillar Network.** *Langmuir* 2018, **34**:3971-3980.
- 72. Ma LL, Li Q, Du ZY, Su EY, Liu X, Wan ZL, Yang XQ: **A Natural Supramolecular Saponin Hydrogelator for Creation of Ultrastable and Thermostimulable Food-Grade Foams.** *Advanced Materials Interfaces* 2019, **6**.
- 73. Chen XW, Yang DX, Zou Y, Yang XQ: **Stabilization and functionalization of aqueous foams by Quillaja saponin-coated nanodroplets.** *Food Research International* 2017, **99**:679-687.
- 74. Santini E, Jarek E, Ravera F, Liggieri L, Warszynski P, Krzan M: **Surface properties and foamability of saponin and saponin-chitosan systems.** *Colloids and Surfaces B-Biointerfaces* 2019, **181**:198-206.
- 75. Gonzalez PJ, Sorensen PM: **Characterization of saponin foam from *Saponaria officinalis* for food applications.** *Food Hydrocolloids* 2020, **101**. A useful review of the recent work on saponins, saponin fibrils and 'nanocrystals' and their interaction with other food foaming agents.
- 76. Singh B, Singh JP, Singh N, Kaur A: **Saponins in pulses and their health promoting activities: A review.** *Food Chemistry* 2017, **233**:540-549.
- 77. Firoozmand H, Rousseau D: **Tailoring the morphology and rheology of phase-separated biopolymer gels using microbial cells as structure modifiers.** *Food Hydrocolloids* 2014, **42**:204-214.
- 78. Firoozmand H, Rousseau D: **Food-grade bijels based on gelatin-maltodextrin-microbial cell composites.** *Food Hydrocolloids* 2015, **48**:208-212.
- 79. Jiang XY, Falco CY, Dalby KN, Siegmundfeldt H, Arneborg N, Risbo J: **Surface engineered bacteria as Pickering stabilizers for foams and emulsions.** *Food Hydrocolloids* 2019, **89**:224-233.

80. He Q, Li L, Zhao XF, Qu L, Wu D, Peng XY: **Investigation of foaming causes in three mesophilic food waste digesters: reactor performance and microbial analysis.** *Scientific Reports* 2017, **7**.
81. Kong X, Liu JG, Yue XP, Li YA, Wang HT: **Fe-0 inhibits bio-foam generating in anaerobic digestion reactor under conditions of organic shock loading and re-startup.** *Waste Management* 2019, **92**:107-114.
82. Ellis AL, Mills TB, Norton IT, Norton-Welch AB: **The hydrophobic modification of kappa carrageenan microgel particles for the stabilisation of foams.** *Journal of Colloid and Interface Science* 2019, **538**:165-173.
83. Binks BP, Campbell S, Mashinchi S, Piatko MP: **Dispersion Behavior and Aqueous Foams in Mixtures of a Vesicle-Forming Surfactant and Edible Nanoparticles.** *Langmuir* 2015, **31**:2967-2978.
- 84. Binks BP, Muijlwijk K, Koman H, Poortinga AT: **Food-grade Pickering stabilisation of foams by in situ hydrophobisation of calcium carbonate particles.** *Food Hydrocolloids* 2017, **63**:585-592.
- The importance of this study is that it illustrates how low molecular weight surfactants can totally change the surface active properties of particles, much more so than proteins, a fact that is mostly ignored but must be important in many real food formulations.
85. Sangamithra A, Venkatachalam S, John SG, Kuppuswamy K: **FOAM MAT DRYING OF FOOD MATERIALS: A REVIEW.** *Journal of Food Processing and Preservation* 2015, **39**:3165-3174.
86. Darniadi S, Ho P, Murray BS: **Comparison of blueberry powder produced via foam-mat freeze-drying versus spray-drying: evaluation of foam and powder properties.** *Journal of the Science of Food and Agriculture* 2018, **98**:2002-2010.
87. Darniadi S, Ifie I, Ho P, Murray BS: **Evaluation of total monomeric anthocyanin, total phenolic content and individual anthocyanins of foam-mat freeze-dried and spray-dried blueberry powder.** *Journal of Food Measurement and Characterization* 2019, **13**:1599-1606.
88. Chaux-Gutierrez AM, Perez-Monterroza EJ, Telis VRN, Mauro MA: **The Physical and Morphological Characteristics of Mango Powder (*Mangifera indica* L. cv Tommy Atkins) Produced by Foam Mat Drying.** *Food Biophysics* 2017, **12**:69-77.
89. Liu C, Liu CH, Xue HK, Sun Y, Lin Z, Liu HJ, Huo JW, Zheng XZ: **Effect of microwave energy dissipation on drying process of berry puree under microwave foam drying conditions.** *Drying Technology* 2017, **35**:1388-1397.
90. Ayetigbo O, Latif S, Abass A, Muller J: **Preparation, optimization and characterization of foam from white-flesh and yellow-flesh cassava (*Manihot esculenta*) for powder production.** *Food Hydrocolloids* 2019, **97**.
91. Larson ES, Conder JM, Arblaster JA: **Modeling avian exposures to perfluoroalkyl substances in aquatic habitats impacted by historical aqueous film forming foam releases.** *Chemosphere* 2018, **201**:335-341.
- 92. Le Toquin E, Faure S, Orange N, Gas F: **New Biocide Foam Containing Hydrogen Peroxide for the Decontamination of Vertical Surface Contaminated With *Bacillus thuringiensis* Spores.** *Frontiers in Microbiology* 2018, **9**.
- Direct proof of the advantage of such foamed bactericides is surely important in cleaning-in-place in the food industry and not least during Covid-19 pandemic sweeping the world at the time of writing this review.
93. Li HZ, Wu ZL, Liu W, Li ZQ, Hu N, Huang D: **Recovery of yam mucilage from the yam starch processing wastewater by using a novel foam fractionation column.** *Separation and Purification Technology* 2016, **171**:26-33.
94. Li R, Wu ZL, Wang YJ, Ding LL, Liu W: **Prevention of irreversible aggregation of whey soy proteins in their foam fractionation from soy whey wastewater.** *Asia-Pacific Journal of Chemical Engineering* 2016, **11**:673-682.
95. Liu W, Wu ZL, Wang YJ, Li R, Ding LL, Huang D: **Rhamnolipid assisted recovery of lycopene from the tomato-based processing wastewater using foam fractionation.** *Journal of Food Engineering* 2015, **164**:63-69.
96. Binks BP, Johnston SK, Sekine T, Tyowua AT: **Particles at Oil-Air Surfaces: Powdered Oil, Liquid Oil Marbles, and Oil Foam.** *Acs Applied Materials & Interfaces* 2015, **7**:14328-14337.

- 97. Fameau AL, Lam S, Arnould A, Gaillard C, Velev OD, Saint-Jalmes A: **Smart Nonaqueous Foams from Lipid-Based Oleogel**. *Langmuir* 2015, **31**:13501-13510.
- 98. Fameau AL, Carl A, Saint-Jalmes A, von Klitzing R: **Responsive Aqueous Foams**. *Chemphyschem* 2015, **16**:66-75.

This is a good review of these fascinating systems, where a massive change in rheological properties of a low solids system can be induced remotely (i.e., via light or other radiation).

- 99. Heymans R, Tavernier I, Danthine S, Rimaux T, Van der Meeren P, Dewettinck K: **Food-grade monoglyceride oil foams: the effect of tempering on foamability, foam stability and rheological properties**. *Food & Function* 2018, **9**:3143-3154.
- 100. Deng BX, de Ruiter J, Schroen K: **Application of Microfluidics in the Production and Analysis of Food Foams**. *Foods* 2019, **8**.

A beautiful illustration of how microfluidics might potentially allow high throughput screening of new foaming agents on the basis of a solid understanding of the dynamic surface active properties.

- 101. Labarre L, Vigolo D: **Microfluidics approach to investigate foam hysteretic behaviour**. *Microfluidics and Nanofluidics* 2019, **23**.

Another fascinating illustration of how detailed microfluidic design and imaging of bubble formation and flow can be used to understand bubble formation and stability

- 102. Laporte M, Montillet A, Della Valle D, Loisel C, Riaublanc A: **Characteristics of foams produced with viscous shear thinning fluids using microchannels at high throughput**. *Journal of Food Engineering* 2016, **173**:25-33.

Like the work of Deng [97] above, but a more specific example in relation to a systems closer to a real food formulation.

- 103. Clarke C, Spyropoulos F, Norton IT: **Surface rheological measurements of isolated food foam systems**. *Physics of Fluids* 2019, **31**.

- 104. Lv QC, Li ZM, Li BF, Husein M, Shi DS, Zhang C, Zhou TK: **Wall slipping behavior of foam with nanoparticle-armored bubbles and its flow resistance factor in cracks**. *Scientific Reports* 2017, **7**.

- 105. Lv QC, Li ZM, Li BF, Shi DS, Zhang C, Li BL: **Silica nanoparticles as a high-performance filtrate reducer for foam fluid in porous media**. *Journal of Industrial and Engineering Chemistry* 2017, **45**:171-181.

- 106. Zhang Y, Allen MC, Zhao RY, Deheyn DD, Behrens SH, Meredith JC: **Capillary Foams: Stabilization and Functionalization of Porous Liquids and Solids**. *Langmuir* 2015, **31**:2669-2676.

Lovely explanation of this new type of foam system, with either an aqueous or non-aqueous based continuous phase.

- 107. Lipsa DR, Roberts RC, Laramée RS: **FoamVis, A Visualization System for Foam Research: Design and Implementation**. *Computers* 2015, **4**:39-60.

Recent Developments in Food Foams

Brent S. Murray^{1}*

b.s.murray@leeds.ac.uk

¹Food Colloids and Bioprocessing Group, School of Food Science and Nutrition, University of Leeds, Leeds, LS2 9JT, UK.

For submission to Current Opinion in Colloid & Interface Science issue Thin Films & Foams

Abstract

The scientific literature between from 2015 onwards (inclusive) with respect to foams and thin films in the context of foods has been reviewed. Proteins are the dominant foaming agents in foods and investigations of the classic, meringue-forming egg white protein still dominate the literature, since the unique properties of this system are still not properly understood. The current drive of many studies is to find suitable replacers of egg proteins, driven by consumer trends for more plant-based alternatives. This has led to investigations of the stabilizing properties of various protein aggregates, 'nanoparticles' and microgel particles as Pickering-type stabilizers of foams (Pickering foams). At the same time, other work has sought to manipulate the surface properties of biopolymer- and non-biopolymer -based particles by chemical means, in order to make the particles adsorb strongly enough. Few, truly novel foam stabilizers have emerged, but two include saponin aggregates and bacteria as particle-type stabilizers.

Keywords: food; foams; Pickering; particles, biopolymers

Introduction

Foam is an important component of many commonly consumed food and drinks (e.g., whipped creams, mousses, soufflés, ice creams, bread, cakes, carbonated beverages) and research into new types of foam and new foam-forming and foam-stabilizing agents continues to grow. Bubbles in consumer products can impart texture, colour, novelty and excitement for the consumer. Additional advantages are that, for air bubbles the bulk material is notoriously cheap, light to transport and is unlikely to result in allergic reactions (for clean air at least)! If only things were so simple, however, because gas bubbles are one of the most difficult colloidal substances to produce, stabilize and control in large quantities, due to their tendency for relatively rapid creaming, dissolution and coalescence (relative to oil-water emulsions, for example). Consequently, great efforts have been made to design systems with improved foam properties, not least because bubbles represent a zero-calorie filler that might be used to replace other food components and have a beneficial effect on the long-term health and well-being of consumers. Thus, apart from the general case of filling the aqueous matrix of foods with more air, systems have been developed for trapping air bubbles in sugar crystals [1], fats and oils [2]●. In addition, there seems to be increasing awareness of the significance and potential of using foam as a 'processing aid'. Thus bubbles can be deliberately used to fractionate a wide range of more valuable surface active materials, whilst conversely foam may concentrate undesirable chemical species. In addition, foam structure may aid the drying and functional properties of dry powders. On the other hand, foaming is an undesirable phenomenon in other processes and understanding how to enhance the action on particles and surfactants in breaking the thin films between bubbles is also an important area of work. It is worth elaborating at the outset some of the colloidal difficulties referred to above, in order to understand the general direction of recent alternative strategies and formulations in the publications referred to later.

Firstly, in the majority of food foams the gas phase is air, nitrogen or carbon dioxide and the continuous phase is an aqueous solution, so that the discontinuous phase (gas) has reasonable (at least mM) solubility in the continuous phase. This is unlike most emulsions or solid dispersions and means that disproportionation, the equivalent of Ostwald ripening in emulsions and dispersions, is rapid when bubbles approach colloidal dimensions, i.e., ca. 1 μm . Adsorbed layers of even tightly packed low molecular weight surfactants (LMWS), or gel-like films of adsorbed macromolecules, offer little physical barrier to transport of gas molecules across interfaces, so that the only way to halt disproportionation completely is to use solid particles, i.e., Pickering stabilization (Pickering foams) as the stabilizing mechanism [3]. Adsorbed layers of such particles can provide a perfectly elastic barrier with high resistance to compression and desorption of the particles, 'hijacking' shrinkage of bubbles due to the higher internal gas pressure across curved interfaces. (The effect of the Laplace pressure difference across the air-water (A-W) interface is exacerbated by the higher A-W interfacial tension compared to that of the oil-water interface in emulsions). This has led to the search for suitable solid particles that can fulfil this role. So far, the range of *food-grade* solids seem to be limited, so compromises may need to be made in terms of softer particles and/or loose particle aggregates to achieve adequate stability. At the same time, some degree of chemical modification, to impart the desired wetting characteristics to the particles, might need to be employed, although it should be recognized that there is consumer pressure to resist all such modifications. Physical modification, e.g.,

heating/cooling, shearing, sonication and complex formation between different ingredients via physical forces, are much more desirable and acceptable.

Secondly, higher gas-water tensions make bubble disruption under shear much more difficult, i.e., high capillary number (= dynamic viscosity x characteristic velocity/ interfacial tension), so that it is generally hard anyway to achieve the same sort of bubble sizes as in homogenization of oil-water systems, particularly if the viscosity of the continuous phase is increased by inclusion of high dissolved solids or thickening agents. Higher bulk viscosity may slow down bubble creaming, drainage and bubble collisions that normally accelerate coalescence, but high viscosity also slows adsorption of the surface active agent to the bubble surfaces. This is particularly relevant for Pickering foams, where the diffusion coefficient of particles and aggregates is already much lower than that of detergent-like molecules or even small globular proteins. Indeed, modelling work [4,5] has shown that it maybe almost impossible to *form* directly high volume fractions of micron-sized Pickering-stabilized air bubbles in any practical system, since such small bubbles will dissolve away before they achieve adequate particle surface coverage. The alternative route is to form larger bubbles initially then allow them to shrink to smaller size, wherein their particle coverage simultaneously increases if the particles already at the interface are irreversibly adsorbed due to their high desorption energy.

The above issues should be born in mind when considering the various new approaches discussed in the following review, some of which have already been discussed in other summaries elsewhere [6]. It is probably also worth stating at the start that two phenomena should be distinguished, since confusion frequently arises in the literature: foamability versus foam stability. Foamability is the capacity for foam formation: systems that have high foamability form large volumes of foam relative to the starting non-foamed liquid, i.e., high overrun. This does not necessarily mean that these foams will be stable for very long. Foam stability is the capacity of a foam, once formed, to persist. Throughout the following, unless explicitly stated, for brevity we will using the word 'foaming' to imply both foamability and foam stability. Bureiko et al. [7] have considered these issues in comparing the foaming of low molecular *versus* polymeric surfactants, like proteins, plus the competition between them in mixtures. The main classes and sub-class of foam stabilizer covered in this review are summarized in Table 1 and these will be discussed in the following sections.

Table 1. 'Index' of the main classes and sub-classes of foam stabilizer discussed in this review.

Main class of stabilizer	Sub-class and corresponding reference numbers
egg white protein (EWP)	reviews 6, 77● general 10, 12, 13, 45, 46, 57, 63● + sugars & polysaccharides 14, 15, 16, 19 + wheat gluten hydrolysates 20, 21
other proteins	sodium caseinate 64● whey protein isolate 17, 105● pea, soy & lentil 18, 47, 48, 49, 59, 62

	wheat gluten & gliadins 20, 21, 22, 36 algal 58; mung bean 60; coconut 53; Quorn® mycoprotein 61 hydrophobin 34●
protein aggregates	β-lactoglobulin 24, 25, 38 lentil 59, 62 soy 49, 96
particles (Pickering)	reviews 28, 29, 30, 33 gluten hydrolysates 20, 21, 22; gliadins 36, 40● zein 39, 40●; hydrophobin 34● chitin 68●; cellulose 71, 72; starch granules 65 saponins 73-77 microgels 31, 32●, 84, CaCO ₃ 85, 86● 'bacteria' 80-81, 83
protein hydrolysates	β-lactoglobulin 51, 56 oat 54● coconut 53, 60 pea 106 EWP 57
covalently cross-linked	α-lactalbumin 52 ovalbumin 13 Na caseinate, OSA-starch & tannic acid 64●
ultrasound-treated proteins	EWP 45, 46 faba bean 47
homogenized proteins	milk 42 pea protein 44
physical complexes & coacervates	whey protein + κ-carageenan 66 whey protein+ alginate 67 lentil protein + guar gum 62 pea protein + OSA-starch 63●
OSA-starch	63●, 64●, 81
fatty alcohols (oleogels/foams)	2●, 99, 100●, 101

Protein-stabilized systems

Egg white protein

Proteins, on their own or combined with other ingredients (see later), still comprise the most widespread foaming agents. It is remarkable that, the proteinaceous culinary foaming agent *par excellence* – egg white, is still the subject of intense investigation. This is connected with our incomplete understanding of the unique stabilizing properties of egg white protein (EWP) and its widespread traditional use in many foamed products, e.g., meringues, mousses, baked goods. At the same time there is strong drive to find plant-based alternatives to this ‘animal-based’ proteinaceous foaming agent, for reasons of environmental sustainability and the relative high proportion of individuals who are allergic to EWP.

The special characteristics of egg albumen stem from the fact that it consists of a mixture of proteins each of which themselves has some less common characteristics: (i) lysozyme, that has a relatively high positive charge at neutral pH, so that it can form electrostatic complexes (aggregates) with the majority of the other (negatively charged) EWP; (ii) ovomucin, a fibrous gelling agent that cannot be denatured up to at least 100 °C that imparts viscoelasticity to the albumen itself; (iii) ovalbumin, the major EWP, a glycoprotein that seems to have a tendency to spontaneous aggregate on adsorption the A-W interface [8],[9]. Daugelaite et al. [10]● have used new, high resolution electrical resistivity measurements as a function of height to investigate free drainage of EWP foams and concluded that the presence of protein aggregates in the Plateau borders has a significant beneficial effect in reducing foam drainage and increasing foam life-times. On the other hand, this tendency for aggregation in the bulk can reduce foamability.

These special features of EWP have lead to a number of treatments to further improve EWP foaming properties, reviewed recently by Gharbi and Labbafi [11]●. A novel treatment is wet ball milling [12], although this seemed to reduce foamability and foam stability due to increased disulfide- and surface hydrophobic-induced aggregation. Similar deliberately induced cross-linking prior to foaming was investigated by Li et al. [13], using hydroxyl radical-induced oxidation of ovalbumin. Moderate oxidation improved the foaming but excessive oxidation reduced it.

In many real products, thickening agents are often added to EWP to try and control the rate of film drainage, particularly when the viscosity of the aqueous phase is already high due the presence of high concentrations of dissolved solids – typically sugars in confectionery products. The higher initial viscosity slows down the incorporation of air to high overrun and inhibits the formation of small enough bubbles. Thickeners, typically polysaccharides, help reduce bubble creaming and drainage whilst the bubble size distribution is reduced to lower sizes by mechanical whipping. They also reduce drainage of the foam product on longer storage times after whipping has ceased, via high values of the low shear rate limiting viscosity or weak gel formation. Understandably, finding the optimum balance of the formulation and processing parameters to give the final desired bubble size distribution, texture, appearance, etc., is not simple and many workers have tried to explain the interaction of all these factors via multi-dimensional response surface studies [14-18]. Ptaszek et al. [19] have shown that a log-normal distribution provides the best description of the bubble size distribution in EWP foamed with pectin and xanthan, confirming the same turbulent eddy-induced Komolgorov mechanism of colloid breakup.

As mentioned above, there is increased interest in substituting EWP with plant-based alternatives. Wouters et al. [20] have shown some improvement in EWP foaming properties by partial substitution with enzymatically hydrolyzed wheat gluten. Gluten hydrolysates had much higher foamability than EWP, whilst EWP showed much higher foam stability than the hydrolysates, but mixtures gave both good foamability and foam stability, suggesting both materials were compatible at the interface. Further measurements of the surface pressure, surface dilatational elasticity (ε) and dilatational viscosity (κ) suggested that the lower molecular weight hydrolysates might dominate the interface with the EWP forming a secondary adsorbed protein layer. Wouters et al. [21] also showed that enzymatically hydrolyzed wheat gluten gives meringues with higher overrun than EWP in a standard recipe (i.e., including sugar) with no notable disadvantages in texture or appearance. More limited gluten hydrolysis seemed to be an advantage due to the formation of more hydrophobic peptides that exhibit higher ε [22].

In many of the studies referred to above and in those that follow, researchers seek to find a link between the interfacial properties of the adsorbed film and bubble stability. The link is not always easy to make. Certainly a high value of ε will help prevent bubble coalescence in the early stages of bubble formation, via the Gibbs-Marangoni mechanism, and many proteins are superior to LMWS in this respect, though the effect is lessened as the kinetics of adsorption increase. Once adsorbed, most proteins are also able to form a thin, viscoelastic, crossed-linked network that can resist further bubble deformation, up to a point, often more reflected in measurements of the interfacial *shear* rheological properties. This can aid resistance to spontaneous coalesce but can lead to film fracture under the large deformations imposed by subsequent processing. Discussion of these aspects is not new [9] and beyond the scope of this review, but the recent study of Mezdour et al. [23]● is worthy of mention in this respect. Here the foaming of protein and LMWS solutions were compared under conditions where the equilibrium surface tensions and bulk viscosity were kept constant. Thus the main variables were thought to be the interfacial viscoelasticity and dynamics of surfactant adsorption/desorption. The main conclusion was that higher dynamic dilatational viscoelasticity and more rapid diffusion kinetics resulted in foams with smaller, more stable bubbles. These effects, plus the effect of shear rate in an industrial type mechanical aerators were used to develop a model to predict bubble size.

Protein aggregates

The tendency for EWP to form aggregates, referred to above, may be one explanation of its excellent foam stabilizing properties, the aggregates providing a thicker and more mechanically resistant film that better resists thin film disruption and provides enhanced steric stabilization. This is provided the aggregates are still surface active and their larger size does not slow down too much their coverage of the A-W interface. In this respect, one might consider EWP as naturally providing something closer to Pickering-type stabilization, although the stabilizing particles are much more complex than traditional Pickering stabilizers. Dombrowski et al. [24,25] have studied the foaming properties of soluble heat-induced aggregates of pure β -lactoglobulin up to sizes of 1 μm . In general, foam stability increased with aggregate size, due to increased protein surface hydrophobicity, and correlated with maxima in the ε and κ . Hu et al. [26] reported similar findings with β -lactoglobulin aggregates of different size and shape - more fibrillar aggregates gave the highest values of

ε and foam stability. The effects of steam pressure and nozzle design on barista-style milk foams as noted by Jimenez-Junca et al. [27] are undoubtedly related to the different extents of milk protein aggregation induced.

Protein-based nanoparticles and microgels

Another way of improving the foaming properties of EWP is to convert the protein into protein microgels [28-30]. For example, Li et al. [31] created submicron (360 nm) EWP microgel (EWPM) particles by homogenization of a thermally cross-linked EWP bulk gel and showed that the EWPM-stabilized aqueous foams via a Pickering-type mechanism. Foamability was less good than EWP-stabilized foams but the EWPM stabilized foams exhibited higher stability to disproportionation and coalescence, in agreement with higher interfacial shear viscosity (η_i) of adsorbed films of EWPM, that were less brittle and liable to fracture. Further work [32]● showed that combinations of EWP and EWPM could achieve the same foamability as EWP alone, whilst still improving stability to bubble shrinkage and coalescence. At the same time, the higher the ratio of EWPM to EWP in the mixtures, the more stable were the foams to freeze-thaw, oven-heating and even microwave heating. The improvement with the mixtures is similar to that obtained with mixtures of gluten hydrolysate and EWP referred to above [20], although here the EWP was not present as microgels but natural aggregates. One might ask what is the difference between a large fractal-type aggregate formed by bottom-up self-assembly of protein molecules and a microgel particle? However, a particle formed by top-down fragmentation of a bulk gel might be expected to retain *internally* the percolating network structure of its parent gel, whilst fractal aggregates do not necessarily have a percolating structure or hold large quantities of solvent. In this review we have reserved the term microgel for those particles that are specifically referred to as such by the authors. The deliberate formation of biopolymer-based particles and aggregates, including microgels, as stabilizing agents of foams (and emulsions) has been reviewed by Dickinson [30]. Microgels in particular have also been reviewed by Murray [29].

The use of protein microgel particles as foam stabilizers exploits the natural superiority of Pickering stabilization, whilst microgels may have other advantages (and disadvantages) compared to traditional solid Pickering stabilizers [28-30,33]. At the same time, other workers have sought to create or exploit other novel protein-based particles as stabilizers that may be considered insoluble in water, or have re-appraised existing proteins as particulate stabilizers. The latter include casein micelles, that may form structures in thin films that significantly curtail drainage [33] and hydrophobin, which is able to form extremely stable small bubbles that can also block drainage of water through the Plateau borders [34]●. The only barrier to use of hydrophobins in foods might be whether or not it represents a significant allergenic potential [35]. Peng et al. [36] created gliadin nanoparticles (diameter \approx 100 – 200 nm) and showed that foaming decreased as the particles became more charged, due to slower adsorption and weaker dynamic dilatational moduli at the A-W interface, whilst effects of varying particle size were negligible. This highlights again the importance of obtaining adequate bubble coverage fast enough with particles

In terms of the minimum bubble surface coverage required for Pickering stabilization, it is well known that particles with a higher aspect ratio than spheres are far more effective. Self-assembly of protein fibrils produced by heating dilute solutions of proteins at low pH have been investigated by various workers and reviewed recently by Mohammadian and

Madadlou [37]●. Thus Peng et al. [38] have investigated β -lactoglobulin fibrils and found that film strength and bubble stability is a maximum close the pI, where the absorbed films have highest dilatational moduli but at the expense of foamability. Raising the pH slightly above the pI and re-introducing some charge to the fibrils improved foamability whilst maintaining good foam stability.

One protein that naturally forms very water-insoluble particles (diameter \approx 100 nm) is the maize storage protein zein and various workers have investigated zein as a natural Pickering stabilizer. Zou et al. [39] found that foams stabilized by zein alone broke down completely in less than 24 h, but if the anionic surfactant sodium dodecyl sulfate (SDS) was added the zein formed fractal clusters of much larger size that give much improved foam stability. Excess SDS reduced stability as adsorption became totally dominated by the LMWS, illustrating another generic difficulty with particle stabilizers. Wouters et al. [40]● also studied zein nanoparticles (size ca. 100 to 150 nm) and also when mixed with wheat gliadin nanoparticles (size ca. 130 to 140 nm), both produced via antisolvent precipitation. In agreement with Zou et al. [39], zein on its own showed very poor foaming, whilst the gliadin on its own was excellent and exhibited much higher surface dilatational moduli. Mixtures were generally worse than the equivalent gliadin concentration alone. This points to the fact that highly hydrophobic and insoluble particles, like the zein, may adsorb strongly, but if there are no cohesive forces between the particles, then the film has little resistance to expansion, as noted earlier for hydrophobins [41].

Modified proteins (physical and chemical)

Physical modification

In this section we will discuss work that has been done to deliberately modify the foaming properties of proteins by physical and chemical means. We have tried to separate out studies of mixtures of proteins with other non-proteinaceous materials, discussed in the following section, although non-covalent complex formation between proteins and other ingredients can still be thought of as a physical modification. We note again that only physical modification is likely to be accepted by regulatory bodies and consumers and also that, much of the work on modified proteins involves plant proteins, in the search for replacements for animal-derived proteins.

Tran et al. [42] have summarized the effects of high-pressure-jet processing on milk proteins in relation to their foaming properties. Apparently this can dissociate casein micelles and denature whey proteins, affecting the bulk viscosity and increasing, in particular, the foamability of whole milk. There is debate about how far the hydrodynamic forces exerted on proteins during their passage through high pressure homogenization can irreversibly affect their conformation [43]●● and therefore how this might affect their ensuing emulsification or foaming properties. It seems obvious that the larger the starting protein aggregates, the more likely it is that such aggregates will be disrupted. Djemaoune et al. [44] have shown that microfluidization only seems to affect the surface activity of pea albumin aggregates when the pH is such that the proteins are in a more compact form. It should always be remembered that the effect of high shear forces on emulsions can sometimes be related to minute contamination with the free-radical inducing transition metal elements in stainless steel.

Ultrasound treatment can also be used to effect physical modifications to proteins, although this can also induce chemical changes due free radical initiation at high power (i.e., sonochemistry). In this respect, there is a similarity with the effects of very high shear (see above), in that both physical and chemical changes might be induced. Sheng et al. [45] and Chen et al. [46] showed significant improvement in EWP foaming properties on sonication, ascribed to formation of aggregates via increased free sulfhydryl content and surface hydrophobicity. Martinez-Velasco et al. [47] used high-intensity ultrasound to increase the solubility and surface activity of faba bean protein isolate which led to improved foam stability but decreased protein digestibility.

Shao, Y. Y., et al. [48] and Wang et al. [49] showed that high quality foaming properties can be obtained by the action of much more simple physical processing, namely heating, of soy protein isolate solutions, again due to opening up of the proteins and exposing more hydrophobic groups. Ossa et al. [50] have shown that pH and temperature treatment are critical in determining the foaming properties of tofu-whey concentrates, an important 'waste' stream of tofu processing. Acidic pH values gave the best foamability and stability, ascribed to the adsorption of more aggregated protein as well as higher concentrations of soluble bean polysaccharides in the aqueous phase.

Chemical modification

Corzo-Martinez et al. [51] studied the effect of limited hydrolysis followed by glycation with galactose (via dry Maillard conjugation), and vice versa, on interfacial and foaming (foamability and foam stability) properties of β -lactoglobulin at pH 7 and pH 5. In particular, the system obtained after glycation followed by limited hydrolysis gave foams with exceptional stability, which might be related to the increase in elastic character and cohesion of the interfacial film, indicated by the increase of the surface dilatational modulus ϵ . Hardly any effect on foaming capacity was observed with the two treatments: hydrolysis followed by glycation, or *vice versa*.

Enzymes, although natural agents themselves, can be used to introduce new covalent cross-links between proteins and create new types of 'unnatural' aggregates. So, although these new aggregates may have improved foaming properties there are doubts about how easily their acceptance might be obtained for use in foods. Thus Dhayal et al. [52] have used transglutaminase or horseradish peroxidase (HRP) to cross-link α -lactalbumin to produce particles with hydrodynamic radii = 20 to 100 nm that gave improved foam stability cf. monomeric α -lactalbumin. However, the stability depended upon which enzyme was used, indicating that the location of the cross-links, i.e., the internal structure of the particles was also important (as might be expected) in terms of surface hydrophobicity, surface area. Kunarayakul et al. [53] have used a glutaminase enzyme to deamidate and improve the foaming properties of coconut protein.

The opposite approach, i.e., cleaving proteins into smaller fragment via enzymes, also continues to be extensively investigated. This has proved particularly useful for plant proteins, which tend to be relatively aggregated and water-insoluble, as demonstrated by Martinez and Pilofof [47] for sunflower protein. As a general rule any enzyme treatment of plant proteins that results in an improvement in aqueous solubility improves foaming properties, but as Bruckner-

Guhmann et al. [54]● showed with oat protein enzymatic hydrolysates, the efficacy depends on the type of enzyme via the different the cleavage patterns and subsequent effects on the (mixed) peptide solubility versus pH. One might also expect that with more extensive hydrolysis the smaller peptides become less effective as steric stabilizers, but this may be mitigated by low molecular weight peptide self-assembly. Li, T., et al. [55] have shown synthetically modified dipeptides to be very effective foam stabilizers, either via the peptides self-assembling (via metal ions) into fibre-like networks at the A-W interface or in the continuous phase to form a gel.

Commercial food grade proteins may contain a range of species with different susceptibilities to the same enzyme conditions compared to native proteins. For example, Pein et al. [56] showed that limited hydrolysis of β -lactoglobulin (fractionated from a whey protein isolate) with pepsin or alcalase, increased both foamability and/or foam stability. It was suggested this was due a combination of non-hydrolysed, native β -lactoglobulin plus more surface active peptides derived from denatured β -lactoglobulin. The denatured β -lactoglobulin was presumably a result of the drying of the whey protein isolate, which made it more accessible to the protease enzymes.

The use of food enzymes to modify food proteins still presents regulatory challenges but recently Jiang et al. [57] have applied natural lactic acid fermentation to EWP and showed a modest but significant foamability, presumably due to the opening up of the proteins and exposing more hydrophobic groups. Such 'traditional' enzymatic treatments may be more acceptable.

Other Novel Plant Proteins

Plant-based, non-modified proteins that have received relatively little attention to date include algal proteins [58]. Suitably extracted protein was reportedly superior to a whey protein isolate. In addition to the large amount of work done on soy protein [48,49], other legumes and pulses have been investigated, e.g., lentil protein aggregates [59] and mung bean proteins [60]. Lonchamp et al. [61] have investigated the properties of a currently unexploited bi-product from the Quorn® fermentation process. One fraction, isolated via ultrafiltration, displayed good foamability and foam stability, although the fraction contained a wide range of metabolites and cell fragments, so that is difficult to be sure which were the key components. It is noted that, like most plant proteins, considerable physical processing is required to improve the solubility and functionality of such proteins, whilst further enzymatic and/or chemical modification might improve this further, if permissible.

Complexes

Another way to improve the foaming properties of proteins is to combine them with other ingredients that may improve the protein solubility or moderate it's surface hydrophobicity – although the two things are obviously connected. If surface hydrophobicity is too high, proteins will self-aggregate in the aqueous phase rather than adsorb; if surface hydrophobicity is too low then there is little driving force for adsorption. Of course the concept of surface hydrophobicity is of little use if the proteins change their conformation significantly during adsorption, as most do. Complexes between different proteins

or proteins or polysaccharides can arise from electrostatic attraction and hydrogen bonding, whereas hydrophobic bonding may also operate between different proteins or proteins and lipids and surfactants. One can distinguish soluble complexes of dimensions similar to the constituent molecules and much larger coacervates, the latter appearing as colloidal particles in their own right. Here we also ignore any effects of proteins and polysaccharides where the polysaccharide merely acts as an independent thickening agent or induces depletion flocculation of the protein.

Jarpa-Parra et al. [62] demonstrated coacervate formation at mildly acidic pH between lentil proteins and guar gum, xanthan gum and pectin that gave much enhanced foam stability, due to an electrostatically cross-linked gel-like interfacial network. The aggregates also plugged the junctions of the Plateau borders in the same way as noted above with EWP aggregates [10] and with casein micelles [34]. Significantly, the proteins and polysaccharides phase separated segregatively at neutral pH and this led to a loss of foam stability.

Even more complicated foaming mixtures involve proteins and synthetically modified polysaccharides, such as starches modified with octenyl succinic anhydride (OSA-starches) [63,64], the latter also being surface active. Asghari et al. [63]● suggest waxy maize OSA-starch adsorption can exclude pea protein from the interface when the latter is still sufficiently aggregated to prevent enough hydrophobic protein residues accessing the interface. Zhan et al. [64]● studied an even more complex system of an OSA-starch, tannic acid and sodium caseinate, the latter being more surface active than pea protein or EWP. Tannic acid cross-links the caseinate films and makes them more rigid. Measurements of ε over a range of strains suggested that the OSA-starch increased the degree of strain softening on interfacial extension via more isolated adsorbed OSA-starch domains in between protein/tannic acid domains. With EWP, Asghari et al. [63]● suggested OSA co-adsorbs but de-mixing in the interface forces the EWP to form more closely packed regions that increases stability. These workers also compared the effect of the OSA-starches with dry heated (non-OSA treated) rice starch granules. Such dry-heating seems to make the granule surface more hydrophobic in some way [65], though it is not clear if this makes them sufficiently surface active to compete with protein adsorption. The dry-heated granules slowed down the drainage of EWP foams via some mechanism but they were not as effective as the OSA starches.

Wang et al. [66] showed that heating WPI with λ -carageenan in solution under neutral or near neutral pH can induce the formation of soluble complexes with improved functional properties compared to heating WPI then adding the carageenan afterwards. The explanation proposed was that heating the two components together resulted in enhanced electrostatic interactions between them, due to their more open molecular configurations. Xu et al. [67] have investigated similar complexes between WPI and sodium alginate. Foam stability was particularly improved (at low overall protein concentrations) at pH values where surface active soluble electrostatic complexes were formed between the oppositely net charged components at a relatively narrow (2:1 to 1:1) range of weight ratios of WPI to alginate.

All the above studies highlight the complexity of the different effects on foaming properties when there are different adsorbing and non-adsorbing macromolecules and/or particles present, although this is always likely to be the case in real food systems.

Novel, non-protein based systems

Two types of novel non-protein based foaming agents have been mainly investigated: polysaccharides that are modified to make them surface active and non-protein based particle (Pickering) stabilizers. Of the former, OSA-starch has already been mentioned above, though this has largely been used in admixture with other foaming agents, whilst with regard to the latter, polysaccharides are also relevant, in terms of water-insoluble cellulose and chitin particles.

Cellulose & chitin

Tzoumaki et al. [68]● have investigated the foaming of chitin nanocrystals. As with many Pickering foams, the importance of structuring of particles both at the interface and within the thin films between bubbles (affectively forming thin layers of a 'bulk' particle gel) was also considered. Even more work has been done on cellulose as a Pickering foam stabilizer, though debate continues how the cellulose can be made hydrophobic enough to adsorb efficiently enough at the A-W interface, or the reason for any apparent natural surface activity of cellulose nanocrystals.[69,70] Beatrice et al. [71] required free radical-oxidized nanocellulose in combination with various water-soluble (polysaccharide) thickening agents to produce adequate foaming properties. Another important observation was that, the effect of shear in these systems caused the cellulose nanofibres to flocculate and accumulate at bubble surfaces, though this depended on the attractive interaction between the cellulose and the polysaccharide thickening agents. Such aggregates can potentially act as *defoaming* agents, plus the more aggregated the particles become, the lower the surface area they can cover and so the coarser will be the foam produced. Ahmadzadeh et al. [72] combined chemically modified clay particles to improve the foaming properties of cellulose-based Pickering foams, for foam packing with enhanced physical properties.

Saponins

A relatively new set of Pickering particles that has emerged is based on the natural class of surface active molecules named saponins. Saponins have long been known to be highly surface active and excellent foaming agents in their own right, but Wan et al. [73] have shown that nanofibrils of glycyrrhizic acid can form networks at the A-W and O-W interfaces of oil droplets and bubbles as the system is emulsified and aerated simultaneously, i.e., an emulsion foam, much like whipped dairy cream. The jamming of the emulsion droplets in thin liquid films also enhances foam stability. On cooling the nanofibrils form a three-dimensional net (gel) that prevents coalescence of the droplets and bubbles within. The foam system could be destroyed rapidly on heating because of the melting of the bulk gel. In later work Ma et al. [74] showed that nanofibrils of glycyrrhizic acid alone can form multilayer networks at the A-W interface and also in the bulk aqueous phase, leading to extremely stable foams. Similarly, the foams could be rapidly destabilized by heating, via break-up of the fibrillar networks. This emulsion foam work is related to the findings of Chen et al. [75], who reported that aqueous foams stabilized by Quillaja saponin-coated 'nanodroplets' were far more stable than foams stabilized by the saponin alone, presumably because the droplets created thicker films between adjacent bubbles. The nanodroplets were of sunflower oil, prepared by sonication and with a mean size ca. 150 nm. Santini et al. [76] also investigated Quillaja saponin and mixed saponin-chitosan solutions. The results substantiated the high surface activity of the saponin and showed that it produced the high values of dilatational viscoelasticity. Another effect emphasized was that the highly

charged chitosan molecules + saponin-induced aggregates may prevent thinning of the thin liquid films between bubbles. This therefore prevents coalescence during foam formation, aiding the subsequent saponin adsorption and stabilization of the bubbles. In addition, there is the possibility of electrostatic complex formation of chitosan with adsorbed saponin, increasing electrosteric stabilization of the bubbles. Gonzalez and Sorensen [77]● have recently reviewed work on saponins and investigated the foaming behaviour of those extracted from Soapwort (*Saponaria officinalis*) in the presence of other common food ingredients (salt, sugar, ethanol) and conditions (pH and temperature). In general the foams were resistant to such formulation variations, apart from ethanol.

Whether such systems can overcome the potential toxicity (albeit in large quantities) of saponins remains to be seen, although saponins may have some health benefits in sequestering bile salts and so lowering blood cholesterol [78].

Bacteria-stabilized

It may seem odd to think of bacteria as particulate stabilizers, but it is well known that the high bacterial cell load in fermented products like yoghurt, along with the mucilages they produce, make a significant contribution to the texture and physical stability of such products. Only recently, however, have workers turned to the idea of bacterial cells themselves acting as Pickering-style particles [79,80]. Jiang et al. [81] have taken this a step further in chemically modifying the 'surface' of lactic acid bacteria (using the same octenyl succinic anhydride commonly used for starch modification) to improve their foaming properties. Although a low level of modification maintained cell viability, it is doubtful if such ingredients would be encouraged in real food stuffs. Foaming is a serious problem during anaerobic digestion of food waste and sewage streams He et al. [82], where it is believed that fatty acids generated are the main foaming agent [83]. It remains to be seen if certain bacterial cells themselves contribute to the foam stability by acting as Pickering stabilizers.

Combinations of particles and other foam stabilizing agents

The problem of particles adsorbing fast enough to the surface of gas bubbles in order to reach a high enough coverage to stabilize them [4,5] was touched upon in the Introduction. At the same time, there is the difficulty of finding native particles that have the correct wetting characteristics without resorting to chemical modification of their surfaces. Both these difficulties may be overcome via physical means by combining particles with other materials that also adsorb to the bubble surfaces and/or that adsorb to the particle surface and advantageously alter the particle contact angle at the bubble surface. In this way, particles may be made to be more strongly adsorbing or less strongly aggregating in solution and therefore more available for adsorption. Alternatively, combinations of particles with other surface active agents may impart the optimum characteristics to the adsorbed film that give rise to high foamability and foam stability. In this way, Ellis et al. [84] used the unusual cationic surfactant lauric arginate to partially neutralize the charge on k-carageenan microgel particles so that this adsorbed more strongly but did not aggregate in solution, to improve their foaming characteristics. Similarly Binks et al. [85] used combinations of hydrophilic calcium carbonate particles and the LMWS sodium stearyl-2-lactylate to improve the stability of foams to coalescence and disproportionation. The same technique was applied [86]● to other, more acceptable food surfactants, including β -lactoglobulin, sodium caseinate, sodium dodecanoate and the Quillaja saponins mentioned earlier. However, it was found that the higher molecular weight,

structurally more complex proteins and Quillaja did not increase foam stability, only the LMWS. This is presumably because the latter adsorb 'head-down' on the inorganic particles and give 'clean' hydrophobic patches and the correct hydrophobic-hydrophilic balance for strong adsorption and a high barrier to particle desorption.

Novel applications of foams in foods

Foam mat drying

There has been increasing research into foam mat drying, where material is foamed and moisture removed by application of heat in some way. The higher surface area of the foam can have the advantage of accelerating the drying process, but mitigating against this are the low thermal conductivity of foam and also the increasing viscosity of the aqueous phase if thickening agents are employed or naturally present. Sangamithra et al. [87] have reviewed this technique, which may be particularly useful for drying fruit juices and pulps that are naturally very sticky due to the sugars they contain' but which also contain valuable colours and antioxidants [88-90] that may be easily damaged and lost in spray drying or prolonged heating. Heat may be applied in a number of ways: freezing stable foams then freeze-drying [88,89] or by application of microwave energy [91] to the wet foam, for example. The bubble size distribution, bubble packing and viscoelasticity of the thin liquid films all play a role in determining the efficiency of drying, all of which also depend on the original foam formulation.

Foam fractionation

Foam fractionation exploits the natural tendency of surface active material to accumulate at bubble surfaces and to become trapped in thin film lamellae as the foam drains. The mining industry has exploited this technique for many years in froth flotation, where foaming is used to accumulate and separate out valuable inorganic particulates via their natural tendency to adsorb at the gas-water interface or via introduction of froth flotation agents that deliberately make them do so. However, the technique potentially also has a number of food-related uses.

An interesting case is the work of Ayetigbo et al. [92] who added glycerol monostearate and sodium carboxymethyl cellulose to aid foaming of cassava pulp. In this case the foamed pulp had a significantly lower concentration of toxic cyanogens. In the opposite sense, Larson et al. [93] have shown that foaming concentrates perfluorinated surfactants that may be harmful to the environment. This alters the site-specific impact of such surfactants but potentially might be exploited to remove them from waste streams. Turning this round again, Le Toquin et al. [94]● have shown that foamed disinfectant agents are at least as effective as liquid solutions at the same concentration but generate far less toxic effluent due to lower run off. This has the potential for cleaning in place of food equipment as well as many other scenarios for disinfecting surfaces.

Foam fractionation of surface active material from a high viscosity bulk liquid is clearly more difficult and slow, but Li et al. [95] have developed a novel foam fractionation column with vertical sieve trays. This was applied to recovering yam mucilage from yam starch processing wastewater and resulted in significant enrichment. Li et al. [96] have pointed out that recovery of functionally useful protein from soy processing waste water, via foam fractionation, relies on preventing

irreversible aggregation of the protein. Interestingly, their results suggested that most insoluble aggregates were formed in the *desorption* of protein from the A-W interface, i.e., aggregation was effectively induced via protein adsorption in the first place, similar to EWP. Addition of a defoaming agent based on LMWS therefore reduced the formation of aggregates. Liu et al. [97] used foam fractionation to recover lycopene from the tomato-based processing wastewater via tomato proteins as the collectors. Moreover natural rhamnolipids could be used to enhance foam stabilization via formation of a protein-lycopene complex.

Oleogels and oleofoams

Another way of reducing the overall calorie content of a fatty product is to incorporate gas bubbles directly into the lipid phase, i.e., an oleofoam. This can be achieved by stabilizing the bubbles with oil-dispersible particles, i.e., Pickering stabilization, or via gelation of the oil-phase, i.e., physically trapping the bubbles in a network with sufficient strength to resist movement of the bubbles via gravity or Brownian motion. In the latter case, termed an oleogel [2]●, the network may be provided by fat crystals inherent to the lipid itself, or other oil-insoluble particles introduced into the system. With oleogels there may also be some adsorption of the gelling particles the bubble surfaces, so that it may not always be clear which mechanism is dominant, analogous to water droplet stabilization in margarines via fat crystals. Binks et al. [98] have provided a clear example of pure Pickering stabilization of air-in-oil (A/O) foams via fluorinated inorganic particles (clays and zinc oxide). Stability depended upon both particle wettability and the amount of shear applied, so that some systems broke down or even inverted to O/A systems. If fluorinated particles could be avoided the latter might provide some interesting opportunities for flavour oil release from the dry powders.

Solely Pickering stabilization of A/O dispersions seems to rare - highlighting the difficulty of finding food-grade particles that easily disperse in oils but that are also sufficiently surface active at the A-O interface – much more work has been done on oleogels. Fameau et al. [99] have produced A/O foams (mean bubble diameter < 200 μm) based on an oleogel of fatty alcohols in sunflower oil. The foams are also most indefinitely stable as long as the fatty alcohol crystals do not melt. An interesting idea therefore, was to make the foams light responsive by also incorporating carbon black particles. On application of UV radiation the gel rapidly warmed, the crystals melted and the foam collapsed. Fameau et al. [100]● have reviewed these and other such responsive foams, where the switch from stability to instability can be induced by changes in the bulk or interfacial material on exposure to light, temperature, and magnetic fields. Heymans et al. [101] have produced a similar A/O system but based on monoglyceride crystals.

In all the above oleogel-stabilized foams, successful stabilization depends upon the size, shape and form of the crystals, which in turn depends on the time-temperature history any other physical processing of the system, the chemical composition of the crystals and their solubility in the continuous oil phase. In other words, the fat crystallization kinetics, which is difficult to control and an area that is still not fully understood. In addition, by analogy with aqueous foams, the bulk viscosity of the continuous phase and the effect of the crystallization on this viscosity determines the ease of incorporation of gas. Higher viscosity makes incorporation more difficult (lower foamability), but once incorporated the

bubbles will be more stable. Some these issues have been nicely discussed in a recent contributions from Mishra et al.[102]

New methods of studying foam formation and stability

As outlined at the outset, foams are relatively unstable compared to many other dispersions in food and this, combined with the relatively large size of bubbles, the tendency for bubbles to cream and pack closely together separated via very thin films, makes them particularly tricky beasts to study. It is therefore of interest to highlight some new methods and techniques that have been developed to improve our understanding of foam formation and destruction.

Many of these new methods centre around the development of microfluidic devices for production of dispersions under controlled and observable conditions, as reviewed recently by Deng et al. [103]●. Labarre and Vigolo [104]●● have developed a method for evaluation of foam stability after a deformation in a microfluidic device. Whether one or more row of bubbles forms in the channels and the size of these bubbles depends on: the gas and liquid pressure drop as bubbles are formed from the flow focussing nozzle; the viscosity of the liquid phase; the surface tension; the ease of bubble deformation at the channel walls. Laporte, M., et al. [105]● applied microfluidics to produce foams stabilized by combinations of whey protein isolate and xanthan gum, showing how on a minute scale this could be used to test and predict the effects of bubble size, bubble volume fraction, continuous phase viscoelasticity and other solution conditions on bulk foam stability. Clarke et al. [106] have studied isolated foam channels created by withdrawing a specific frame geometry from surfactant solutions and using image analysis to monitor film thickness and drainage across the model Plateau borders created. Drainage for aqueous solutions of food grade LMWS, hydroxypropyl methylcellulose and hydrolyzed pea protein were studied. Novel distortions in the border thickness profiles were exhibited with the polymeric surfactants, ascribed to the much higher γ , produced with such stabilizers. In similar ways, the flow of bulk foam down microchannels can be tested, where wall slip or 'partial coalescence' with the wall are key factors controlling the flow, particularly for bubbles stabilized by particles [107]. Increasing particle coverage creates a more rigid and rough bubble surface that increases the frictional resistance to slip at the microchannel wall. Such effects can also be used to inhibit liquid ingress into narrow channels and capillaries [108].

Zhang et al. [109]●● reported a 'new' type of particle-stabilized foam material, capillary foam, which is stabilized by the synergistic action of particles and a small amount of an immiscible liquid - typically an oil for aqueous-based foams. The stabilization mechanism relies on the wetting of particles by the oil to some extent, which results in them sticking together via capillary bridges. The capillary foam is then formed by aeration of the flocculated dispersion. Alternatively, a particle stabilized foam can be formed first, then the oil added and the system re-agitated to distribute the oil to the particles. In this way, one indirectly tunes the capacity of the particles to stabilize the air-water interface via the particle affinity for the air-oil and oil-water interfaces and the spreading of a thin oil film around the bubbles. This results in foams that are then extremely stable. Oil soluble dyes can be added to make the foam appear highly coloured, or the oil film cured (solidified) to produce highly stiff, load-bearing dry foams. Whilst such systems have been deliberately designed, the effects are also important because of the inadvertent interaction of Pickering foams with oils in their potential usage as enhanced (crude)

oil recovery agents and their sealing of capillaries and cracks as discussed above [107,108]. Simulating and visualizing the flow behaviour of such complex systems in order to understand them better also remains a great challenge, as discussed by Lipsa et al. [110].

Conclusions

Research into novel stabilizers of food foams continues apace, particularly with regard to particulate stabilizers (Pickering foams). Most of these systems still employ proteins as the key stabilizing ingredient, either in the form of particles of some type, or as modifiers of the surface properties of other particulate material. Many of these studies seek to include stable air bubbles as bulking agents and calorie-lowering agents, at the same time seeking to find replacements for traditional stabilizers based on animal proteins (e.g., dairy and egg proteins). Whilst in most food foams the bulk liquid phase is aqueous, interesting work on air-in-oil foams (oleofoams and oleogels) represent additional novel ways of influencing food texture and stability for the benefit of consumers.

References

1. Van Der Vegt A, Bisschop HA: **Sweet particulate fat-containing powder, its preparation and its use.** *Patent WO2012050439A1* 2012.
- 2. Heymans R, Tavernier I, Dewettinck K, Van der Meeren P: **Crystal stabilization of edible oil foams.** *Trends in Food Science & Technology* 2017, **69**:13-24.
A good review of the expanding field of oleofoams and oleogels.
3. Valadbaigi P, Ettelaie R, Kulak AN, Murray BS: **Generation of ultra-stable Pickering microbubbles via polyalkylcyanoacrylates.** *Journal of Colloid and Interface Science* 2019, **536**:618-627.
4. Ettelaie R, Murray B: **Effect of particle adsorption rates on the disproportionation process in pickering stabilised bubbles.** *Journal of Chemical Physics* 2014, **140**:204713.
5. Ettelaie R, Murray BS: **Evolution of bubble size distribution in particle stabilised bubble dispersions: Competition between particle adsorption and dissolution kinetics.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2015, **475**:27-36.
6. Narsimhan G, Xiang N: **Role of Proteins on Formation, Drainage, and Stability of Liquid Food Foams.** In *Annual Review of Food Science and Technology, Vol 9.* Edited by Doyle MP, Klaenhammer TR; 2018:45-63. *Annual Review of Food Science and Technology, vol 9.*
7. Bureiko A, Trybala A, Kovalchuk N, Starov V: **Current applications of foams formed from mixed surfactant-polymer solutions.** *Advances in Colloid and Interface Science* 2015, **222**:670-677.
8. Lechevalier V, Croguennec T, Pezennec S, Guerin-Dubiard C, Pasco M, Nau F: **Evidence for synergy in the denaturation at the air-water interface of ovalbumin, ovotransferrin and lysozyme in ternary mixture.** *Food Chemistry* 2005, **92**:79-87.
9. Murray BS: **Rheological properties of protein films.** *Current Opinion in Colloid & Interface Science* 2011, **16**:27-35.
- 10. Daugelaite D, Guillermic RM, Scanlon MG, Page JH: **Quantifying liquid drainage in egg-white sucrose foams by resistivity measurements.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2016, **489**:241-248.
Based on more accurate measurements of electrical conductivity across a column of foam as function of height, this method could find wide application in many other systems.
- 11. Gharbi N, Labbafi M: **Influence of treatment-induced modification of egg white proteins on foaming properties.** *Food Hydrocolloids* 2019, **90**:72-81.
A useful review of the ongoing modifications to egg white protein.
12. Li M, Li MM, Tan W, Liu XB, Duan X: **Effects of ball-milling treatment on physicochemical and foaming activities of egg ovalbumin.** *Journal of Food Engineering* 2019, **261**:158-164.

13. Li SG, Huang Y, An FP, Huang Q, Geng F, Ma MH: **Hydroxyl radical-induced early stage oxidation improves the foaming and emulsifying properties of ovalbumin.** *Poultry Science* 2019, **98**:1047-1054.
 14. Sadahira MS, Rodrigues MI, Akhtar M, Murray BS, Netto FM: **Effect of egg white protein-pectin electrostatic interactions in a high sugar content system on foaming and foam rheological properties.** *Food Hydrocolloids* 2016, **58**:1-10.
 15. Sadahira MS, Rodrigues MI, Akhtar M, Murray BS, Netto FM: **Influence of pH on foaming and rheological properties of aerated high sugar system with egg white protein and hydroxypropylmethylcellulose.** *Lwt-Food Science and Technology* 2018, **89**:350-357.
 16. Sadahira MS, Lopes FCR, Rodrigues MI, Yamada AT, Cunha RL, Netto FM: **Effect of pH and interaction between egg white protein and hydroxypropylmethylcellulose in bulk aqueous medium on foaming properties.** *Carbohydrate Polymers* 2015, **125**:26-34.
 17. Laporte M, Della Valle D, Loisel C, Marze S, Riaublanc A, Montillet A: **Rheological properties of food foams produced by SMX static mixers.** *Food Hydrocolloids* 2015, **43**:51-57.
 18. Moll P, Grossmann L, Kutzli I, Weiss J: **Influence of energy density and viscosity on foam stability - A study with pea protein (*Pisum Sativum* L.).** *Journal of Dispersion Science and Technology* 2019.
 19. Ptaszek P, Kabzinski M, Kruk J, Kaczmarczyk K, Zmudzinski D, Liszka-Skoczylas M, Mickowska B, Lukasiewicz M, Banas J: **The effect of pectins and xanthan gum on physicochemical properties of egg white protein foams.** *Journal of Food Engineering* 2015, **144**:129-137.
 20. Wouters AGB, Rombouts I, Fierens E, Brijs K, Blecker C, Delcour JA, Murray BS: **Foaming and air-water interfacial characteristics of solutions containing both gluten hydrolysate and egg white protein.** *Food Hydrocolloids* 2018, **77**:176-186.
 21. Wouters AGB, Rombouts I, Fierens E, Brijs K, Delcour JA: **Enzymatically hydrolyzed wheat gluten as a foaming agent in food: incorporation in a meringue recipe as a proof-of-concept.** *Journal of Food Science* 2018, **83**:2119-2126.
 22. Wouters AGB, Rombouts I, Legein M, Fierens E, Brijs K, Blecker C, Delcour JA: **Air-water interfacial properties of enzymatic wheat gluten hydrolyzates determine their foaming behavior.** *Food Hydrocolloids* 2016, **55**:155-162.
 - 23. Mezdour S, Segueineau de Preval E, Granda P, Cuvelier G, Ducept F: **Impact of interfacial characteristics on foam structure: study on model fluids and at pilot scale.** *Oil & Gas Science and Technology- Revue D Ifp Energies Nouvelles* 2017, **72(2)**:13.
- Not many studies have tried to compare proteins and low molecular weight surfactants, like with like, i.e., where the effects of bulk viscosity and equilibrium tension are equal, but this work does, allowing much clearer interpretation of the differences between them.
24. Dombrowski J, Johler F, Warncke M, Kulozik U: **Correlation between bulk characteristics of aggregated beta-lactoglobulin and its surface and foaming properties.** *Food Hydrocolloids* 2016, **61**:318-328.
 25. Dombrowski J, Gschwendtner M, Kulozik U: **Evaluation of structural characteristics determining surface and foaming properties of beta-lactoglobulin aggregates.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2017, **516**:286-295.
 26. Hu J, Yang JX, Xu Y, Zhang K, Nishinari K, Phillips GO, Fang YP: **Comparative study on foaming and emulsifying properties of different beta-lactoglobulin aggregates.** *Food & Function* 2019, **10**:5922-5930.
 27. Jimenez-Junca C, Sher A, Gumy JC, Niranjana K: **Production of milk foams by steam injection: The effects of steam pressure and nozzle design.** *Journal of Food Engineering* 2015, **166**:247-254.
 28. Murray BS: **Pickering Emulsions for Food and Drinks** *Current Opinion in Food Science* 2019, **27**:57-63.
 29. Murray BS: **Microgels at fluid-fluid interfaces for food and drinks.** *Advances in Colloid and Interface Science* 2019, **271**:101990.
 30. Dickinson E: **Biopolymer-based particles as stabilizing agents for emulsions and foams.** *Food Hydrocolloids* 2017, **68**:219-231.
 31. Li X, Murray BS, Yang YJ, Sarkar A: **Egg white protein microgels as aqueous Pickering foam stabilizers: Bubble stability and interfacial properties.** *Food Hydrocolloids* 2020, **98**:105292.

- 32. Li X, Yang YJ, Murray BS, Sarkar A: **Combination of egg white protein and microgels to stabilize foams: Impact of processing treatments.** *Journal of Food Engineering* 2020, **275**.

Not many other studies of microgel-stabilized foams have subjected the systems to such extreme conditions. These results highlight the great potential of other protein microgels to mimic egg white protein.

- 33. Dickinson E: **Structuring of colloidal particles at interfaces and the relationship to food emulsion and foam stability.** *Journal of Colloid and Interface Science* 2015, **449**:38-45.

- 34. Dimitrova LM, Petkov PV, Kralchevsky PA, Stoyanov SD, Pelan EG: **Production and characterization of stable foams with fine bubbles from solutions of hydrophobin HFBII and its mixtures with other proteins.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2017, **521**:92-104.

This later work on more applied aspect of hydrophobin highlights again the highly unseal properties of this protein, which acts like a natural Pickering agent for foams, whilst supplementation with other proteins may extend its possible usage.

- 35. Yew SM, Chan CL, Ngeow YF, Toh YF, Na SL, Lee KW, Hoh CC, Yee WY, Ng KP, Kuan CS: **Insight into different environmental niches adaptation and allergenicity from the *Cladosporium sphaerospermum* genome, a common human allergy-eliciting Dothideomycetes.** *Scientific Reports* 2016, **6**:27008.

- 36. Peng DF, Jin WP, Tang C, Lu Y, Wang WQ, Li J, Li B: **Foaming and surface properties of gliadin nanoparticles: Influence of pH and heating temperature.** *Food Hydrocolloids* 2018, **77**:107-116.

- 37. Mohammadian M, Madadlou A: **Technological functionality and biological properties of food protein nanofibrils formed by heating at acidic condition.** *Trends in Food Science & Technology* 2018, **75**:115-128.

A very useful review of this highly topical area, which has implications in many other areas outside foods.

- 38. Peng DF, Yang JC, Li J, Tang C, Li B: **Foams Stabilized by beta-Lactoglobulin Amyloid Fibrils: Effect of pH.** *Journal of Agricultural and Food Chemistry* 2017, **65**:10658-10665.

- 39. Zou Y, Wan ZL, Guo J, Wang JM, Yin SW, Yang XQ: **Modulation of the surface properties of protein particles by a surfactant for stabilizing foams.** *Rsc Advances* 2016, **6**:66018-66026.

- 40. Wouters AGB, Schaefer S, Joye IJ, Delcour JA: **Relating the structural, air-water interfacial and foaming properties of wheat (*Triticum aestivum* L.) gliadin and maize (*Zea mays* L.) zein based nanoparticle suspensions.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2019, **567**:249-259.

This paper clearly illustrates that high surface hydrophobicity and solid particle type nature are not necessarily an advantage – proteins particles must unfold and cross-link to some extent to give a coherent film that can stabilize bubbles against coalescence and disproportionation.

- 41. Wang YW, Bouillon C, Cox A, Dickinson E, Durga K, Murray BS, Xu R: **Interfacial study of class II hydrophobin and its mixtures with milk proteins: relationship to bubble stability.** *Journal of Agricultural and Food Chemistry* 2013, **61**:1554-1562.

- 42. Tran M, Roberts R, Felix TL, Harte FM: **Effect of high-pressure-jet processing on the viscosity and foaming properties of pasteurized whole milk.** *Journal of Dairy Science* 2018, **101**:3887-3899.

- 43. Dobson J, Kumar A, Willis LF, Tuma R, Higazi DR, Turner R, Lowe DC, Ashcroft AE, Radford SE, Kapur N, et al.: **Inducing protein aggregation by extensional flow.** *Proceedings of the National Academy of Sciences of the United States of America* 2017, **114**:4673-4678.

Although not directed at foods or even foams, this detailed study of the effect of extensional flow on the conformational change of several physiologically relevant blood proteins shows that, even at relatively low protein concentrations, thermodynamically and kinetically stable proteins can be readily unfolded. This raises many questions about the effects of different stirring conditions on food protein surface activity.

- 44. Djemaoune Y, Cases E, Saurel R: **The effect of high-pressure microfluidization treatment on the foaming properties of pea albumin aggregates.** *Journal of Food Science* 2019, **84**:2242-2249.

- 45. Sheng L, Wang YB, Chen JH, Zou J, Wang Q, Ma MH: **Influence of high-intensity ultrasound on foaming and structural properties of egg white.** *Food Research International* 2018, **108**:604-610.

- 46. Chen YX, Sheng L, Gouda M, Ma MH: **Impact of ultrasound treatment on the foaming and physicochemical properties of egg white during cold storage.** *Lwt-Food Science and Technology* 2019, **113**:108303.

47. Martinez KD, Pilosof AMR: **Foaming behaviour of enzymatically modified sunflower protein in proximity to pl.** *Biointerface Research in Applied Chemistry* 2017, **7**:1883-1886.
48. Shao YY, Lin KH, Kao YJ: **Modification of Foaming Properties of Commercial Soy Protein Isolates and Concentrates by Heat Treatments.** *Journal of Food Quality* 2016, **39**:695-706.
49. Wang MP, Chen XW, Guo J, Yang J, Wang JM, Yang XQ: **Stabilization of foam and emulsion by subcritical water-treated soy protein: Effect of aggregation state.** *Food Hydrocolloids* 2019, **87**:619-628.
50. Ossa JSH, Wagner JR, Palazolo GG: **Influence of chemical composition and structural properties on the surface behavior and foam properties of tofu-whey concentrates in acid medium.** *Food Research International* 2020, **128**:108772.
51. Corzo-Martinez M, Moreno FJ, Villamiel M, Patino JMR, Sanchez CC: **Effect of glycation and limited hydrolysis on interfacial and foaming properties of bovine beta-lactoglobulin.** *Food Hydrocolloids* 2017, **66**:16-26.
52. Dhayal SK, Delahaije R, de Vries RJ, Gruppen H, Wierenga PA: **Enzymatic cross-linking of alpha-lactalbumin to produce nanoparticles with increased foam stability.** *Soft Matter* 2015, **11**:7888-7898.
53. Kunarayakul S, Thaiphant S, Anprung P, Suppavorasatit I: **Optimization of coconut protein deamidation using protein-glutaminase and its effect on solubility, emulsification, and foaming properties of the proteins.** *Food Hydrocolloids* 2018, **79**:197-207.
- 54. Bruckner-Guhmann M, Heiden-Hecht T, Sozer N, Drusch S: **Foaming characteristics of oat protein and modification by partial hydrolysis.** *European Food Research and Technology* 2018, **244**:2095-2106.
This study illustrates nicely the difficulties of understanding or predicting the effects of enzymatic hydrolysis on protein functional properties, which is dependent on so many external (solution) and intrinsic (state of protein unfolding and or/aggregation) factors.
55. Li T, Nudelman F, Tavacoli JW, Vass H, Adams DJ, Lips A, Clegg PS: **Long-lived foams stabilized by a hydrophobic dipeptide hydrogel.** *Advanced Materials Interfaces* 2016, **3**:1500601.
56. Pein D, Clawin-Radecker I, Lorenzen PC: **Peptic treatment of beta-lactoglobulin improves foaming properties substantially.** *Journal of Food Processing and Preservation* 2018, **42**:e13543.
57. Jiang YW, Jia J, Xiong DD, Xu XM, Yang YJ, Liu XB, Duan X: **Effects of short-term fermentation with lactic acid bacteria on egg white: Characterization, rheological and foaming activities.** *Food Hydrocolloids* 2020, **101**:105507.
58. Buchmann L, Bertsch P, Bocker L, Krahenmann U, Fischer P, Mathys A: **Adsorption kinetics and foaming properties of soluble microalgae fractions at the air/water interface.** *Food Hydrocolloids* 2019, **97**.
59. Jarpa-Parra M, Bamdad F, Tian Z, Zeng HB, Temelli F, Chen L: **Impact of pH on molecular structure and surface properties of lentil legumin-like protein and its application as foam stabilizer.** *Colloids and Surfaces B-Biointerfaces* 2015, **132**:45-53.
60. Widjajaseputra AI, Widyastuti TEW: **Potential of coconut milk and mung bean extract combination as foam stabilizer in non-dairy ice cream.** *International Food Research Journal* 2017, **24**:1199-1203.
61. Lonchamp J, Clegg PS, Euston SR: **Foaming, emulsifying and rheological properties of extracts from a co-product of the Quorn fermentation process.** *European Food Research and Technology* 2019, **245**:1825-1839.
62. Jarpa-Parra M, Tian Z, Temelli F, Zeng HB, Chen L: **Understanding the stability mechanisms of lentil legumin-like protein and polysaccharide foams.** *Food Hydrocolloids* 2016, **61**:903-913.
- 63. Asghari AK, Norton I, Mills T, Sadd P, Spyropoulos F: **Interfacial and foaming characterisation of mixed protein-starch particle systems for food-foam applications.** *Food Hydrocolloids* 2016, **53**:311-319.
- 64. Zhan FC, Li J, Shi MQ, Wu D, Li B: **Foaming properties and linear and nonlinear surface dilatational rheology of sodium caseinate, tannin acid, and octenyl succinate starch ternary complex.** *Journal of Agricultural and Food Chemistry* 2019, **67**:2340-2349.
This study and the one above (Asghari et al. [63]) illustrate the potential interfacial incompatibility or the two very different surface active macromolecular species – proteins and hydrophobically modified polysaccharides (in the case OSA-starch) and the different direction in which this can force foam stability.
65. Nakagawa M, Tabara A, Ushijima Y, Matsunaga K, Seguchi M: **Hydrophobicity of stored (15, 35 degrees C), or dry-heated (120 degrees C) rice flour and deteriorated breadmaking properties baked with**

these treated rice flour/fresh gluten flour. *Bioscience Biotechnology and Biochemistry* 2016, **80**:983-990.

66. Wang ZS, Zhang S, Vardhanabhuti B: **Foaming Properties of Whey Protein Isolate and lambda-Carrageenan Mixed Systems.** *Journal of Food Science* 2015, **80**:N1893-N1902.
67. Xu Y, Yang N, Yang JX, Hu J, Zhang K, Nishinari K, Phillips GO, Fang YP: **Protein/polysaccharide intramolecular electrostatic complex as superior food-grade foaming agent.** *Food Hydrocolloids* 2020, **101**:105474.
- 68. Tzoumaki MV, Karefyllakis D, Moschakis T, Biliaderis CG, Scholten E: **Aqueous foams stabilized by chitin nanocrystals.** *Soft Matter* 2015, **11**:6245-6253.
- Whilst much work has been done recently on cellulose, chitin also represents a much under-utilized source of insoluble food grade particles. This work introduces this topic nicely, focusing on food-grade modifications.
69. Capron I, Rojas OJ, Bordes R: **Behavior of nanocelluloses at interfaces.** *Current Opinion in Colloid & Interface Science* 2017, **29**:83-95.
70. Bertsch P, Fischer P: **Adsorption and interfacial structure of nanocelluloses at fluid interfaces.** *Advances in Colloid and Interface Science* 2020, **276**:102089.
71. Beatrice CAG, Rosa-Sibakov N, Lille M, Sozer N, Poutanen K, Ketoja JA: **Structural properties and foaming of plant cell wall polysaccharide dispersions.** *Carbohydrate Polymers* 2017, **173**:508-518.
72. Ahmadzadeh S, Nasirpour A, Keramat J, Hamdami N, Behzad T, Desobry S: **Nanoporous cellulose nanocomposite foams as high insulated food packaging materials.** *Colloids and Surfaces a-Physicochemical and Engineering Aspects* 2015, **468**:201-210.
73. Wan ZL, Sun YE, Ma LL, Zhou FB, Guo J, Hu SQ, Yang XQ: **Long-Lived and Thermoresponsive Emulsion Foams Stabilized by Self-Assembled Saponin Nanofibrils and Fibrillar Network.** *Langmuir* 2018, **34**:3971-3980.
74. Ma LL, Li Q, Du ZY, Su EY, Liu X, Wan ZL, Yang XQ: **A Natural Supramolecular Saponin Hydrogelator for Creation of Ultrastable and Thermostimulable Food-Grade Foams.** *Advanced Materials Interfaces* 2019, **6**:1900417.
75. Chen XW, Yang DX, Zou Y, Yang XQ: **Stabilization and functionalization of aqueous foams by Quillaja saponin-coated nanodroplets.** *Food Research International* 2017, **99**:679-687.
76. Santini E, Jarek E, Ravera F, Liggieri L, Warszynski P, Krzan M: **Surface properties and foamability of saponin and saponin-chitosan systems.** *Colloids and Surfaces B-Biointerfaces* 2019, **181**:198-206.
- 77. Gonzalez PJ, Sorensen PM: **Characterization of saponin foam from Saponaria officinalis for food applications.** *Food Hydrocolloids* 2020, **101**:105541.

A useful review of the recent work on saponins, saponin fibrils and 'nanocrystals' and their interaction with other food foaming agents.

78. Singh B, Singh JP, Singh N, Kaur A: **Saponins in pulses and their health promoting activities: A review.** *Food Chemistry* 2017, **233**:540-549.
79. Firoozmand H, Rousseau D: **Tailoring the morphology and rheology of phase-separated biopolymer gels using microbial cells as structure modifiers.** *Food Hydrocolloids* 2014, **42**:204-214.
80. Firoozmand H, Rousseau D: **Food-grade bijels based on gelatin-maltodextrin-microbial cell composites.** *Food Hydrocolloids* 2015, **48**:208-212.
81. Jiang XY, Falco CY, Dalby KN, Siegumfeldt H, Arneborg N, Risbo J: **Surface engineered bacteria as Pickering stabilizers for foams and emulsions.** *Food Hydrocolloids* 2019, **89**:224-233.
82. He Q, Li L, Zhao XF, Qu L, Wu D, Peng XY: **Investigation of foaming causes in three mesophilic food waste digesters: reactor performance and microbial analysis.** *Scientific Reports* 2017, **7**:13701.
83. Kong X, Liu JG, Yue XP, Li YA, Wang HT: **Fe-0 inhibits bio-foam generating in anaerobic digestion reactor under conditions of organic shock loading and re-startup.** *Waste Management* 2019, **92**:107-114.
84. Ellis AL, Mills TB, Norton IT, Norton-Welch AB: **The hydrophobic modification of kappa carrageenan microgel particles for the stabilisation of foams.** *Journal of Colloid and Interface Science* 2019, **538**:165-173.
85. Binks BP, Campbell S, Mashinchi S, Piatko MP: **Dispersion Behavior and Aqueous Foams in Mixtures of a Vesicle-Forming Surfactant and Edible Nanoparticles.** *Langmuir* 2015, **31**:2967-2978.

- 86. Binks BP, Muijlwijk K, Koman H, Poortinga AT: **Food-grade Pickering stabilisation of foams by in situ hydrophobisation of calcium carbonate particles.** *Food Hydrocolloids* 2017, **63**:585-592.

The importance of this study is that it illustrates how low molecular weight surfactants can totally change the surface active properties of particles, much more so than proteins, a fact that is mostly ignored but must be important in many real food formulations.

- 87. Sangamithra A, Venkatachalam S, John SG, Kuppuswamy K: **Foam mat drying of food materials: a review.** *Journal of Food Processing and Preservation* 2015, **39**:3165-3174.
- 88. Darniadi S, Ho P, Murray BS: **Comparison of blueberry powder produced via foam-mat freeze-drying versus spray-drying: evaluation of foam and powder properties.** *Journal of the Science of Food and Agriculture* 2018, **98**:2002-2010.
- 89. Darniadi S, Ifie I, Ho P, Murray BS: **Evaluation of total monomeric anthocyanin, total phenolic content and individual anthocyanins of foam-mat freeze-dried and spray-dried blueberry powder.** *Journal of Food Measurement and Characterization* 2019, **13**:1599-1606.
- 90. Chaux-Gutierrez AM, Perez-Monterroza EJ, Telis VRN, Mauro MA: **The Physical and Morphological Characteristics of Mango Powder (*Mangifera indica* L. cv Tommy Atkins) Produced by Foam Mat Drying.** *Food Biophysics* 2017, **12**:69-77.
- 91. Liu C, Liu CH, Xue HK, Sun Y, Lin Z, Liu HJ, Huo JW, Zheng XZ: **Effect of microwave energy dissipation on drying process of berry puree under microwave foam drying conditions.** *Drying Technology* 2017, **35**:1388-1397.
- 92. Ayetigbo O, Latif S, Abass A, Muller J: **Preparation, optimization and characterization of foam from white-flesh and yellow-flesh cassava (*Manihot esculenta*) for powder production.** *Food Hydrocolloids* 2019, **97**:105205.
- 93. Larson ES, Conder JM, Arblaster JA: **Modeling avian exposures to perfluoroalkyl substances in aquatic habitats impacted by historical aqueous film forming foam releases.** *Chemosphere* 2018, **201**:335-341.
- 94. Le Toquin E, Faure S, Orange N, Gas F: **New Biocide Foam Containing Hydrogen Peroxide for the Decontamination of Vertical Surface Contaminated With *Bacillus thuringiensis* Spores.** *Frontiers in Microbiology* 2018, **9**:2295.

Direct proof of the advantage of such foamed bactericides is surely important in cleaning-in-place in the food industry and not least during Covid-19 pandemic sweeping the world at the time of writing this review.

- 95. Li HZ, Wu ZL, Liu W, Li ZQ, Hu N, Huang D: **Recovery of yam mucilage from the yam starch processing wastewater by using a novel foam fractionation column.** *Separation and Purification Technology* 2016, **171**:26-33.
- 96. Li R, Wu ZL, Wang YJ, Ding LL, Liu W: **Prevention of irreversible aggregation of whey soy proteins in their foam fractionation from soy whey wastewater.** *Asia-Pacific Journal of Chemical Engineering* 2016, **11**:673-682.
- 97. Liu W, Wu ZL, Wang YJ, Li R, Ding LL, Huang D: **Rhamnolipid assisted recovery of lycopene from the tomato-based processing wastewater using foam fractionation.** *Journal of Food Engineering* 2015, **164**:63-69.
- 98. Binks BP, Johnston SK, Sekine T, Tyowua AT: **Particles at Oil-Air Surfaces: Powdered Oil, Liquid Oil Marbles, and Oil Foam.** *Acs Applied Materials & Interfaces* 2015, **7**:14328-14337.
- 99. Fameau AL, Lam S, Arnould A, Gaillard C, Velev OD, Saint-Jalmes A: **Smart Nonaqueous Foams from Lipid-Based Oleogel.** *Langmuir* 2015, **31**:13501-13510.
- 100. Fameau AL, Carl A, Saint-Jalmes A, von Klitzing R: **Responsive Aqueous Foams.** *Chemphyschem* 2015, **16**:66-75.

This is a good review of these fascinating systems, where a massive change in rheological properties of a low solids system can be induced remotely (i.e., via light or other radiation).

- 101. Heymans R, Tavernier I, Danthine S, Rimaux T, Van der Meeren P, Dewettinck K: **Food-grade monoglyceride oil foams: the effect of tempering on foamability, foam stability and rheological properties.** *Food & Function* 2018, **9**:3143-3154.

102. Mishra K, Bergfreund J, Bertsch P, Fischer P, Windhab EJ: **Crystallization-Induced Network Formation of Tri- and Monopalmitin at the Middle-Chain Triglyceride Oil/Air Interface.** *Langmuir* 2020, **36**:7566-7572.

● 103. Deng BX, de Ruiter J, Schroen K: **Application of Microfluidics in the Production and Analysis of Food Foams.** *Foods* 2019, **8**:476.

A beautiful illustration of how microfluidics might potentially allow high throughput screening of new foaming agents on the basis of a solid understanding of the dynamic surface active properties.

●● 104. Labarre L, Vigolo D: **Microfluidics approach to investigate foam hysteretic behaviour.** *Microfluidics and Nanofluidics* 2019, **23**:129.

Another fascinating illustration of how detailed microfluidic design and imaging of bubble formation and flow can be used to understand bubble formation and stability.

● 105. Laporte M, Montillet A, Della Valle D, Loisel C, Riaublanc A: **Characteristics of foams produced with viscous shear thinning fluids using microchannels at high throughput.** *Journal of Food Engineering* 2016, **173**:25-33.

Like the work of Deng [103] above, but a more specific example in relation to a systems closer to a real food formulation.

106. Clarke C, Spyropoulos F, Norton IT: **Surface rheological measurements of isolated food foam systems.** *Physics of Fluids* 2019, **31**:092002.

107. Lv QC, Li ZM, Li BF, Husein M, Shi DS, Zhang C, Zhou TK: **Wall slipping behavior of foam with nanoparticle-armored bubbles and its flow resistance factor in cracks.** *Scientific Reports* 2017, **7**:5063.

108. Lv QC, Li ZM, Li BF, Shi DS, Zhang C, Li BL: **Silica nanoparticles as a high-performance filtrate reducer for foam fluid in porous media.** *Journal of Industrial and Engineering Chemistry* 2017, **45**:171-181.

●● 109. Zhang Y, Allen MC, Zhao RY, Deheyn DD, Behrens SH, Meredith JC: **Capillary Foams: Stabilization and Functionalization of Porous Liquids and Solids.** *Langmuir* 2015, **31**:2669-2676.

Lovely explanation of this new type of foam system, with either an aqueous or non-aqueous based continuous phase.

110. Lipsa DR, Roberts RC, Laramée RS: **FoamVis, A Visualization System for Foam Research: Design and Implementation.** *Computers* 2015, **4**:39-60.

Conflict of Interest Statement

Regarding the manuscript: "Recent Developments in Food Foams", authored by myself, Brent S. Murray
I declare there are no conflicts of interest.

Dear Editor, Thank you for the positive response to our review. I note your minor editorial comment and have inserted any missing page numbers or article numbers in the references where these are available. I also thank the Reviewer for their suggestions, which I have addressed as follows, allowing the opportunity to improve the MS further. Below I repeat Reviewer 1's comments/suggestions, followed by my response, in italics for clarity.

It is not really clear where the Introduction ends and the material specific discussion starts.

In the Introduction I have focused almost exclusively on the generic problems of stabilizing foams, though I have introduced Pickering foams as a special case. At the end of the Introduction I introduced Table 1, a sort of classification summary of the different types of stabilizer as a lead into the following sections, which discuss most of these in turn. However, to make this more clear, I have changed the last sentence of the Introduction to "The main classes and sub-class of foam stabilizer covered in this review are summarized in Table 1 and these will be discussed in the following sections."

The Abstract features abbreviations and parentheses, which impairs the optical and reading pleasure. Later on also some sentences are placed parentheses: In my opinion, the parentheses are not needed.

There is only one abbreviation in the Abstract, for egg white protein (EWP), which I subsequently used only once. I think this was introduced this mainly to keep the word count down. Nevertheless, I have removed this. There is one set of parentheses, around "Pickering foams". I think this is justified because the term is not entirely widespread yet, unlike Pickering emulsions.

On page 7 there is a valid comment on the often-claimed link between foam stability and interfacial properties. I think the comment is important and should be extended to interfacial shear rheology as well (right now only interfacial dilatational rheo is mentioned).

We agree entirely and so have extended the 4th sentence already there in paragraph 2 of page 7, so that this now reads "Once adsorbed, most proteins are also able to form a thin, viscoelastic, crossed-linked network that can resist further bubble deformation, up to a point, often more reflected in measurements of the interfacial shear rheological properties."

For cellulose (page 13) one could also mention some of the recent published reviews on cellulose at interfaces

(<https://doi.org/10.1021/acs.chemmater.7b00531>, <https://doi.org/10.1016/j.cocis.2017.04.001>,

<https://doi.org/10.1016/j.cis.2019.102089>). These reviews not entirely focus on foams but might help the reader to understand cellulose interfaces better.

There have been a huge number of recent publications relating to cellulose and various colloidal systems so that it is impossible to know which to select unless they specifically relate to foams, the subject of this review. In addition, there is a lot of confusion between interfacial and network stabilization, which we have discussed. In the light of this, the best of Reviewer 1's recommendations seem to be the 2nd and 3rd (Capron et al., 2017 and Bertsch & Fischer, 2020), which discuss the nature of cellulose surface activity and foams: we have therefore included these two in the revised MS, in the 3rd sentence of the section of cellulose and chitin, which now ends ..." or the reason for any apparent natural surface activity of cellulose nanocrystals" with these references inserted.

The first recommendation (De France et al, 2017), relates to aerogels and NOT foams, so we have not included it since the above two cover the required topic adequately.

Page 14: Heading "Combinations of particles and other foaming agents". What do you mean by foaming agents? The foam stabilizing surface-active materials or the gas? In engineering nomenclature the foaming agent would be the gas. Just to avoid confusion.

I was not aware of this engineering nomenclature, so thanks to the Reviewer for bringing this to my attention. I have clarified this by changing the title heading to ""Combinations of particles and other foam stabilizing agents"

Page 16: Section on oleogel. It might be worth to check with two super-recent publications by Mishra et al. on fat foam generation (<https://pubs.acs.org/doi/abs/10.1021/acs.cgd.9b01558>) and the interfacial aggregation of differently treated triglycerides (<https://pubs.acs.org/doi/10.1021/acs.langmuir.0c01195>).

Again this is an area that is expanding rapidly, so that it is hard to keep up, so the recommendations of Reviewer 1 are very welcome. The second (Langmuir paper) actually refers to the work in the first (Crystal Growth and Design) paper, so we have included just the second, adding the sentence: "Some these issues have been nicely discussed in a recent contributions from Mishra et al. [inserted] at the end of the "Oleogels and oleofoams" section.

[Inclusion of these additional reference has obviously activated some reference re-numbering].

Declaration of Interest Statement

Regarding the manuscript: "Recent Developments in Food Foams", authored by myself, Brent S. Murray
I declare there are no conflicts of interest.