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1 **Combining common genetic variants and non-genetic risk factors to predict risk of**
2 **cutaneous melanoma**

3

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55 **ABSTRACT**

56 Melanoma heritability is among the highest for cancer and single nucleotide
57 polymorphisms (SNPs) contribute to it. To date, only SNPs that reached statistical significance
58 in genome-wide association studies or few candidate SNPs have been included in melanoma risk
59 prediction models. We compared four approaches for building polygenic risk scores (PRS) using
60 12,874 melanoma cases and 23,203 controls from Melanoma Meta-Analysis Consortium as a
61 training set, and newly genotyped 3,102 cases and 2,301 controls from the MelaNostrum
62 consortium for validation. We estimated adjusted odds ratios (ORs) for melanoma risk using
63 traditional melanoma risk factors and the PRS with the largest area under the Receiver Operator
64 Characteristics curve (AUC). We estimated absolute risks combining the PRS and other risk
65 factors, with age- and sex-specific melanoma incidence and competing mortality rates from Italy
66 as an example. The best PRS, including 204 SNPs (AUC= 64.4%; 95% CI=63-65.8%),
67 developed using winner's curse estimate corrections, had a per-quintile OR=1.35 (95% CI=1.30-
68 1.41), corresponding to a 3.33-fold increase comparing the 5th to the 1st PRS quintile. The AUC
69 improvement by adding the PRS was up to 7%, depending on adjusted factors and country. The
70 20-year absolute risk estimates based on the PRS, nevus count and pigmentation characteristics
71 for a 60-year old Italian man ranged from 0.5% to 11.8% (RR=26.34), indicating good
72 separation.

73

74

75 INTRODUCTION

76 The incidence of cutaneous melanoma is increasing in western countries (1-3), with about
77 132,000 new cases worldwide each year. Melanoma is highly curable when detected in its
78 earliest stages, with a 5-year survival rate of 98%. However, notwithstanding improved
79 treatments in recent years (4-6), survival rates decline to 62% and 18% for regional and distant
80 stage disease, respectively (2, 7). Identifying subjects at high risk for melanoma is critical to
81 provide targeted screening and early detection, and numerous melanoma risk prediction models
82 have been built to facilitate this aim (8-20). Previous models mainly included environmental or
83 host risk factors, such as age, family history, sun exposure, sunburns, number of melanocytic
84 nevi, and/or pigmentation characteristics. Several of these risk factors have a strong genetic
85 component and genetic factors are strongly implicated in the etiology of melanoma. Heritability
86 for melanoma has been estimated to be 58%, among the highest for cancer (21). Rare high-risk
87 variants in a few genes, such as *CDKN2A*(22), *CDK4*(23), *BAP1* (24), *TERT* (25), *POT1* (26,
88 27), *ACD* (28) and *PARK2* (29) and variants with intermediate allele frequency (~1-5%),
89 including variants in *MITF* (30), explain ~40% of familial melanoma, but account for a very
90 small proportion of melanoma in the general population.

91 A large proportion of missing heritability is due to common genetic variants(31), which,
92 when combined, may confer substantial risk. Genome-wide association studies (GWAS) of
93 cutaneous melanoma have identified 20 genetic loci associated with melanoma risk to date (32),
94 some of which are near genes related to pigmentation (*ASIP*, *SLC45A2*, *HERC2/OCA2*, *MC1R*,
95 and *TYR*) (33, 34) and/or are associated with nevus count (*TERT*, *PLA2G6*, *CDKN2A-MTAP*,
96 *IRF4*)(32, 35, 36). Building on these findings, a few previous reports of melanoma risk

97 prediction models have combined 11 to 19 SNPs that reached genome-wide significance(37-39)
98 or a few candidate SNPs with biological relevance (38).

99 A considerable proportion of phenotypic variation can be explained by the combination
100 of genetic loci not achieving GWAS significance (40). In this study, we thoroughly explored
101 models that included SNPs selected based on different criteria to build polygenic risk scores that
102 could capture the underlying genetic risk for melanoma. We used the largest meta-analysis of
103 melanoma GWAS data to date from the Melanoma Meta-Analysis Consortium (MMAC) (32) as
104 a training set and validated the performance of the PRS in newly genotyped subjects from
105 Southern Europe, a population typically under-represented in melanoma studies, from the
106 MelaNostrum Consortium. We assessed the association of the PRS with melanoma risk, also
107 adjusting for host/environmental melanoma risk factors. Finally, we built an absolute risk model
108 for melanoma risk by combining relative risks for the PRS and other risk factors using the age-
109 and gender- specific melanoma incidence rates and competing mortality rates from Italy as
110 example. We identified a PRS including 204 SNPs that reached an AUC of 64.4%. The
111 combination of this PRS and the traditional risk factors for melanoma (light hair color, light eye
112 color, high sun sensitivity, large number of nevi as well as older age and male sex) strongly
113 stratified subjects based on melanoma risk.

114 **RESULTS**

115 **Comparison of four models to estimate polygenic risk scores (PRS) using MMAC as a**
116 **training dataset and MelaNostrum as the testing dataset**

117 The characteristics of the MMAC training dataset are reported in Law *et al.* (32). The
118 genotyping testing set from the MelaNostrum Consortium included 5,599 subjects (3,124 cases
119 and 2,475 controls) from Greece, Cyprus, Italy and Spain. Of this set, all the 194 subjects from
120 Cyprus and two additional subjects had no phenotypic covariates and thus were excluded from
121 the analyses including traditional melanoma risk factors. Thus, the MelaNostrum population
122 (Table 1) included 775 melanoma cases and 752 controls from Greece; 1,266 cases and 361
123 controls from Italy; 1,061 cases and 1,188 controls from Spain. Cases included more women
124 than controls, were older, had lighter eye color and hair color, lower skin photo-type, and more
125 nevi. Subjects' characteristics by country and study site are presented in Supplementary Tables
126 1a and 1b.

127
128 The PRS in Model 1, with 17 genome-wide significant SNPs in MMAC (32) plus
129 rs4778138 as proxy for rs7164220, achieved AUC=62.8% (95% CI=61.4%-64.3%) in the testing
130 dataset. In model 2, the best AUC=63.9% (62.5%-65.4%) was achieved with the p -value
131 threshold= 5×10^{-8} and $r^2 = 0.01$ for clumping. This model included 23 SNPs, comprising the 18
132 SNPs included in Model 1 plus five additional SNPs: four on chr.16 in the *MC1R* region, and
133 one on chr.9 in the *CDKN2A/MTAP* region. While keeping the LD clumping criteria at $r^2=0.01$
134 and changing p -value thresholds from 5×10^{-8} up to 10^{-2} (Model 2), the corresponding AUC
135 decreased steadily down to 55.6% (95%CI=54.1-57.1%) for p -value= 10^{-2} (Fig 1). Using LDpred
136 (Model 3), the best AUC was 63.3% (95%CI= 60.8-65.4%). Model 4, correcting for the
137 winner's curse bias and using LD clumping $r^2=0.01$, provided the PRS with the best performance
138 at p -value threshold 10^{-4} . It included 204 SNPs, and had AUC= 64.4% (95% CI=63.0-65.8%)
139 (Figure 1). In the country-specific validation, the AUCs corresponding to the p -value 10^{-4} , were

140 61.3%, 60.9%, and 63.7% (95% CI=61.4-66.0%) for the Greek, Italian and Spanish samples,
141 respectively (Supplementary Table 2). As a sensitivity analysis, we reran the validation
142 excluding all 196 subjects with missing phenotypic covariates to match the population used for
143 the overall analyses and obtained the same 204 SNPs. The 204 SNPs in the PRS with p -
144 value $< 10^{-4}$ are listed in Supplementary Table 3 and the corresponding genotyping data can be
145 found on github at this link: https://github.com/xtmgah/Melanoma_PRS.

146

147 **Association between PRS and melanoma risk in the testing dataset considering well** 148 **established melanoma risk factors**

149 Melanoma traditional risk factors were associated with melanoma risk in MelaNostrum
150 data (Supplementary Table 4). The PRS with 204 SNPs was weakly, but significantly, correlated
151 with nevus count and pigmentation variables in MelaNostrum controls overall and in country-
152 specific analyses (Table 2). No correlation was observed with age, sex, and sun exposure. The
153 PRS was significantly associated with melanoma risk in the overall population and in each
154 country separately (Table 3). The OR per PRS quintile was 1.35 (95% CI=1.30-1.41) in the
155 overall population, which corresponds to a 3.3-fold increased melanoma risk comparing the
156 highest vs. the lowest PRS quintile. The ORs per PRS quintile were 1.31 (95%CI: 1.22-1.42) in
157 Greece, 1.32 (95%CI: 1.21-1.43) in Italy, and 1.40 (95%CI: 1.31-1.48) in Spain, corresponding
158 to a 2.98, 3.04 and 3.79-fold risk increase in the highest vs. lowest PRS quintile, respectively.
159 Adjusting for demographic factors did not substantially change the ORs, while adjusting for
160 pigmentation factors and nevus count decreased the per quintile OR of PRS to 1.23 (95%
161 CI=1.13-1.35) in the overall population, and 1.29, 1.23, and 1.26 in Greece, Italy and Spain,
162 respectively. Additionally adjusting for sun exposure-related variables for the Italian and Spanish

163 samples did not affect the results (Table 3). There were no major differences in PRS-melanoma
164 associations by categories of age, sex, nevus count, pigmentation, or tumor characteristics (data
165 not shown).

166 The AUC differences from models without and with PRS varied by country (Table 4).
167 Adding the PRS improved the AUC by 7.3% in Italy and 2.0% in Spain (model with
168 demographic factors); this reflects the age distribution: cases and controls had similar age in the
169 Italian study, while controls were younger than cases in the Spanish study.

170
171 **Absolute risk of developing melanoma in the Italian population**

172
173 Absolute melanoma risk considering competing mortality risk showed substantial risk
174 separation by different risk profiles in the Italian population aged 50, 60 and 70 years; risks
175 ranged from 0.15% [0.16%] to 7.20% [3.66%] at 10 years and from 0.35% [0.29%] to 11.85%
176 [7.10%] at 20 years in men [women] across different combinations of PRS and phenotype risk
177 factors (Figure 2a and 2b and Supplementary Table 5). For example, a 60-year old Italian man in
178 the highest risk category (light eye color, red hair, I-II skin photo-type, 50+ nevi, 5th PRS
179 quintile) had estimated 10-year and 20-year absolute melanoma risks of 5.38% and 11.76%,
180 respectively, compared to 0.21% and 0.48% for a man of the same age in the lowest risk
181 category (dark eye color, brown hair, III-VI skin photo-type, <50 nevi, 1st PRS quintile). Similar
182 patterns were observed for women. The attributable risk of the PRS based on the relative risk
183 estimates from the cases was 0.26 in the Italian population.

184

185 **DISCUSSION**

186 We report on a polygenic risk score for melanoma risk that combines 204 common SNPs
187 and had an AUC of 64.4%. This PRS was obtained using a model that corrected for the winner's
188 curse bias in SNP effect size estimates. Based on the PRS, subjects in the highest quintile had
189 ~2.5-fold risk of melanoma compared to those in the lowest quintile, after adjusting for other
190 major melanoma risk factors. Although not directly comparable, a 2.5 to 3-fold increased risk of
191 melanoma is equivalent or even stronger than the risk of very severe solar damage (10), family
192 history, gender, and many pigmentation and UV-related risk factors (10, 41). This PRS, in
193 combination with pigmentation characteristics and number of nevi, strongly differentiated
194 melanoma risk in the Italian population and thus could be useful towards identifying high-risk
195 subjects who could potentially benefit from increased surveillance.

196 Optimal p -value threshold to select SNPs for disease risk prediction depends on the
197 number of causal SNPs and their effect size distribution, and the sample size of the training data
198 set (40, 42). Accordingly, we thoroughly explored models that included SNPs based on different
199 selection criteria, to build polygenic risk scores that could capture the underlying genetic risk for
200 melanoma. We used a very large training data, to maximize the accuracy of the PRS. The AUC
201 (64.4%) of the best PRS is larger than the PRS-based AUCs for other cancers using the largest
202 GWAS summary data, such as the AUC for lung (56.4%), colorectal (57.4%), pancreatic
203 (58.7%) (43) or breast (61.5%) (44) cancers. It is only slightly smaller than the AUC (65.4%) for
204 prostate cancer (43), which was obtained using a training dataset three-times larger than the one
205 for melanoma. These results are consistent with the heritability estimates across cancers, which
206 are highest for melanoma (58%, 95% CI=43%-73%) and prostate cancer (57%, 95% CI=51%-
207 63%) (21). Absolute risk estimates for melanoma combining PRS and the other melanoma risk
208 factors stratified Italian subjects very well into high and low risk groups, suggesting potential

209 application of PRS in melanoma precision prevention. We used the Italian population because
210 we could obtain age- and sex-specific incidence and mortality rates from cancer registries
211 (AIRTUM) (53, 54), which were not available for Spain and Greece, and we had data on the
212 traditional risk factors for this study population. Moreover, we wanted to investigate the range of
213 estimated absolute risks in a country without routine melanoma screening, where people are not
214 perceived to be at high risk for the disease, and so this model could constitute an important tool
215 for melanoma prevention. Similar calculation can be conducted for other countries using their
216 own age- and gender-specific melanoma incidence and mortality rates. Since the absolute disease
217 risk for short prediction intervals (e.g. 5 years) is proportional to the relative risk multiplied by
218 the age-specific baseline incidence, the PRS effect on absolute risk estimates could be
219 substantially stronger in populations with higher melanoma incidence rates, including Australia
220 and Northern European countries.

221 Several melanoma risk factors have a genetic component, and the PRS, including SNPs at
222 pigmentation- (e.g., *SLC45A2* or *MC1R*) or nevus-associated (e.g., *MTAP*) loci, was weakly
223 correlated with both pigmentation characteristics and nevus count. Overall, the AUC
224 improvement provided by the PRS over traditional risk factors ranged from 0.8% to 1.7%
225 depending on the variables in the models, with some variability also due to the different study
226 designs across the countries. When only age and sex were included in the models, adding the
227 PRS improved the AUC, particularly in the Italian population where cases and controls were
228 matched on age. However, when pigmentation and nevi variables were added, the improvement
229 was reduced overall and for all countries. The impact of the PRS on absolute risk was more
230 noticeable, leading to a doubling of absolute risk for each profile when changing the PRS
231 quintile from the lowest to the highest. This was particularly meaningful for older men, who had

232 the highest melanoma incidence rate in the Italian population. We could not test the effect of
233 PRS in subjects with or without family history of melanoma since few studies collected this
234 information. To avoid oversampling for family history that could bias the PRS effect estimates,
235 we specifically excluded studies that were sampled based on family history.

236 Since the training data mostly included subjects from Northern Europe, Australia and the
237 US and the validation set included subjects from Southern European countries (MelaNostrum),
238 we evaluated whether the PRS could be useful across different populations. The model
239 performance could be affected by the effect size (i.e., the odds ratio) of the SNPs in the PRS and
240 the variant allele frequency of the genes included in the PRS. We checked whether the effect
241 sizes of each of the 204 SNPs in the best PRS differed between the training set and MelaNostrum
242 subjects (Supplementary Table 6). The large majority of the SNPs had a similar effect size across
243 populations; only three SNPs (rs75286671 at chr.4, rs187989493 at chr.7, and rs139791480 at
244 chr.6) reached a statistically significant difference ($p < 2.45 \times 10^{-4}$). However, as expected, some
245 SNPs in pigmentation-associated loci, such as rs7164220 around *HERC2*, rs250417 around
246 *SCLC45A2*, and rs1805008 around *MC1R*, had different minor allele frequencies between the
247 training set and MelaNostrum (minor allele frequency=0.119 vs. 0.246; 0.03 vs. 0.09; 0.08 vs.
248 0.02, respectively). Thus, the PRS effect estimates can be transferred to other countries of
249 European ancestry, but the ability to discriminate subjects at high or low risk for the disease
250 could vary across different populations.

251 This study has many strengths. For building the PRS, we used the largest melanoma
252 GWAS data to date as a training set, a major determinant of the accuracy of PRS prediction (40).
253 We thoroughly explored different SNP selection criteria and statistical approaches, and chose

254 one with the largest AUC to build the PRS. We genotyped for the first time many subjects from
255 Mediterranean populations, typically under-represented in melanoma studies, for independent
256 validation. We also studied the impact of PRS with and without traditional risk factors for
257 melanoma using various models. Finally, we estimated the absolute risk of melanoma for Italian
258 subjects with different risk profiles and combinations of PRS.

259 Some limitations should also be noted: we lack prospective cohort data for model
260 calibration, which would be ideal for the direct application of the risk prediction model to the
261 public health or clinical setting. However, when we tested the fit of the relative risk model that
262 was the basis of the absolute risk predictions using different approaches as proposed by Song *et*
263 *al.*, (45) none of the tests indicated lack of fit of the model (p-values ranging from 0.08 to 0.78,
264 using 10,000 simulations). Thus, we conclude the relative risk model has adequate fit to the
265 Italian case-control data. An additional limitation is that there was an upward bias for AUC
266 estimate in Models 2, 3 and 4 with a single tuning parameter, because the validation dataset was
267 used for both selecting the tuning parameter and calculating AUC. Such bias is minimal
268 (typically less than 0.15%), as we have shown on simulation studies (43). Moreover, while we
269 conducted imputation for missing data in pigmentation and nevi variables (about 10% and 20%
270 of overall subjects), we had to exclude some traditional risk factors (e.g., family history) from
271 the models because of larger missing data from some studies. Finally, there was heterogeneity
272 among the contributing studies in study design and data collection, e.g., controls in some Spanish
273 and Greek studies were younger than cases, while cases and controls from the Italian studies
274 were matched on age. This discrepancy can explain the differences in the performance of the risk
275 prediction model when including only the demographic variables with the PRS (Table 4).
276 However, we saw no evidence of heterogeneity in SNPs' odds ratios among studies, suggesting

277 that SNP and PRS estimates should be broadly applicable. Moreover, the absolute risk model is
278 not affected by this issue because we only used the Italian studies which were age-matched.

279 Our study suggests that PRS, in combination with traditional melanoma risk factors, may
280 help identify subjects who could benefit from heightened skin examination and sun-avoidance.
281 Prospective analyses of the PRS together with other melanoma risk factors are needed to validate
282 the overall accuracy of risk prediction in Mediterranean and other countries. We expect that risk
283 models combining genetic and non-genetic risk factors will be further improved when larger
284 genetic studies become available in the future.

285 **MATERIALS AND METHODS**

286 **Study population and genotyping**

287 Our PRS was constructed using summary level data from a GWAS meta-analysis from
288 the Melanoma Meta-Analysis Consortium (MMAC) (32), including 11 GWAS from Europe,
289 Australia, and the U.S., totaling 12,874 melanoma cases and 23,203 controls. The details of the
290 study population, genotyping and quality control information are published previously (32).

291 We validated our PRS using independent GWAS data from the MelaNostrum
292 consortium, formed by clinicians and researchers from institutions dedicated to melanoma
293 management in Mediterranean countries. MelaNostrum included cases with histologically-
294 confirmed primary cutaneous melanoma and participants who were melanoma-free at study entry
295 from Italy, Spain, Greece, and Cyprus. Details of the design, data collection, and genotyping
296 methods are presented in the Online Data Supplement. All participants signed an informed
297 consent and the study was reviewed by Institutional Review Boards of the local hospitals and the

298 National Cancer Institute. After quality control, 5,599 subjects (3,124 cases and 2,475 controls)
299 and 707,169 SNPs were used as a validation set for the PRS. Of the 5,599 subjects, 194 subjects
300 from Cyprus and two additional subjects had no phenotypic covariates and thus were excluded
301 from the additional analyses including traditional melanoma risk factors. Thus, the total number
302 of subjects for the overall analyses included 5,403 subjects (3,102 cases, 2,301 controls) from
303 Italy, Spain and Greece. Characteristics of the study population are summarized in Table 1 and
304 Supplementary Tables 1a and 1b.

305

306 **Statistical analyses**

307 Polygenic risk score (PRS) computation

308 We built PRS using four methods based on odds ratios (\widehat{OR}_t) or equivalently $\hat{\beta}_t =$
309 $\log(\widehat{OR}_t)$, and p-values p_t from logistic regression analysis fit to each SNP individually in
310 MMAC (32) (the training data).

311 The first PRS (Model 1) included only $K=18$ SNPs achieving genome-wide significance
312 in MMAC. Note that, for each locus, only the most significant SNPs were selected into the PRS.
313 For each subject i in the validation dataset, the PRS was then calculated as

$$314 \quad PRS_i = \sum_{t=1}^K \hat{\beta}_t g_{it},$$

315 where g_{it} is the genotypic value for SNP t for subject i .

316 The second PRS (Model 2) used different p -value thresholds for SNP inclusion (46).

317 Briefly, we first performed linkage disequilibrium (LD) clumping with PLINK(47) using
318 correlation $r^2 = 0.01$ and window size 5 Mb, guided by the SNP p -values in the training data.

319 Sensitivity analysis was performed using $r^2 = 0.1, 0.2$ and 0.3 . Assuming there are M SNPs after
 320 LD clumping, the PRS for subject i with p -value threshold p is

$$321 \quad PRS_i(p) = \sum_{t=1}^M \hat{\beta}_t g_{it} I(P_t \leq p),$$

322 where $I=1$ if $P_t \leq p$ and $I=0$ otherwise, and the p -value threshold was chosen as $5 \times 10^{-8}, 10^{-7},$
 323 $10^{-6}, 10^{-5}, 10^{-4}, 10^{-3}$ and 10^{-2} . The optimal p -value threshold was the one that maximized the
 324 prediction performance in the validation sample.

325 The third PRS (Model 3) was constructed using LDpred (48). LDpred includes all
 326 analyzed SNPs while re-estimating the effect size β_t as the posterior mean by conditioning on
 327 the marginal effect size estimates for all SNPs and LD information in a local region. Compared
 328 to the other models that require LD clumping, LDpred may have better performance when
 329 multiple SNPs in a local region are independently associated with the phenotype.

330 Finally, the fourth method (Model 4) is similar to Model 2 but corrects the effect size
 331 estimation for winner's curse, i.e. the fact that effect estimates for SNP selected based on having
 332 small p -values are upwardly biased. We recently demonstrated that correcting for this bias can
 333 improve the predictive performance of PRS (43). Following this approach, we bias-corrected the
 334 SNP specific estimates $\hat{\beta}_t$, to obtain

$$335 \quad \hat{\beta}_t^{wcc}(p) = \text{sign}(\hat{\beta}_t) \left(|\hat{\beta}_t| - \lambda(p) \right) I(|\hat{\beta}_t| > \lambda(p)),$$

336 where $\lambda(p)$ depends on the p -value threshold p : $\lambda(p) = \Phi^{-1} \left(1 - \frac{p}{2} \right) \hat{\sigma}_t$. Here, $\Phi()$ is the
 337 probability distribution function for a standard normal distribution.

338 The rs4778138 SNP was reported as significant in MMAC but was not imputed well in
339 MelaNostrum; thus, we included rs7164220 (LD $R^2=0.6$ with rs4778138) in all models even if it
340 did not achieve the required significance level.

341 We evaluated the prediction performance of the four PRS scores in the MelaNostrum
342 GWAS (the testing data) by calculating the area under the Receiving Operator Characteristics
343 (ROC) curve (AUC) using the R package “pROC” (49) with bootstrap confidence intervals.

344

345 Contribution of PRS on melanoma risk prediction

346 We assessed the association of the PRS with the best predictive performance (coded in
347 quintiles) with melanoma risk, alone and with additional risk factors, and evaluated its
348 performance in risk prediction in the MelaNostrum data.

349 We imputed traditional risk factors, allowing for interactions with case-status. The
350 variables were assumed to be categorical and included: age at diagnosis for cases or at study
351 enrollment for controls, eye color (dark, medium, light), hair color (black, dark
352 brown/light/reddish brown, blond, red), intermittent sun exposure (none/some, high), sunlamp
353 use (yes, no), actinic keratosis (yes, no), chronic sun exposure (yes, no), skin type (I-II, III-VI),
354 sunscreen use (yes, no). We did not impute missing family history and did not use this
355 information in the model. The imputation was conducted using IVEware (50), and we analyzed
356 the M=5 imputed datasets, accounting for the random imputation in the variance computation
357 using PROC MIANALYZE (Inc. SI. SAS 9.3. Cary, NC2011) (51). The largest amount of
358 missingness was seen for sunscreen use (57.76%, excluded from the model); eye and hair color

359 had $\leq 15\%$ missing data. We observed no substantial differences in our findings when we
360 excluded individuals with missing values in a sensitivity analysis (data not shown).

361 ORs and 95% confidence intervals (CIs) for association were calculated using logistic
362 regression models (PROC Logistic, SAS 9.3). PRS quintiles were coded as an ordinal variable.
363 We used data harmonized across the different studies and countries to adjust the PRS models.
364 Specifically, models were: 1) not adjusted; 2) adjusted for demographic factors only (age, sex,
365 country of residence: Greece, Italy, Spain); 3) adjusted for demographic factors, pigmentation
366 variables (eye color, hair color, skin phototype) and nevus count. Models adjusted for linear
367 combinations of pigmentation characteristics obtained using factor analysis (52) yielded similar
368 estimates and are thus not shown. We included an age \times study site interaction term in the models
369 to accommodate different age distributions across studies. We computed two-sided p -values
370 using Wald tests; $p < 0.05$ was considered statistically significant.

371 We also stratified all analyses by country of residence. We further adjusted Italian models
372 for chronic sun exposure, intermittent sun exposure and history of sunburns, and Spanish models
373 for chronic sun exposure, chronic sun damage, acute sun damage and history of sunburns.

374 Contributions of PRS to prediction performance were evaluated by the difference of AUC
375 between models with and without PRS, computed based on cross-validation, overall and by
376 country.

377 Projecting probabilities (absolute risk) of developing melanoma in Italian subjects

378 The absolute risk $r^*(a,b)$ of melanoma in the age interval (a,b) is the probability of
379 developing melanoma during that interval, given that one is alive and free of previous melanoma
380 at age a ,

381
$$r^*(a,b) = \int_a^b \lambda_m(t,x) \exp(-\int_a^t \lambda_m(u,x) + \lambda_D(u,x) du) dt. \quad (1)$$

382 The melanoma hazard rate λ_m was modeled as $\lambda_m(a,x) = (1-AR(x)) \exp(\beta x) \lambda^*(a)$ as the
383 product of one minus the age- and sex-specific attributable risk for all the risk factors in the
384 model, the relative risk, $\exp(\beta x)$, that includes covariates x , and age and sex specific incidence
385 rates from ITACAN, <http://itacan.ispo.toscana.it>, pooling data from 38 Italian cancer registries in
386 2009. For details see Pfeiffer and Gail (53), Chapter 5. The competing deaths hazard λ_D was
387 estimated by subtracting 5-year age and sex-specific mortality rates for melanoma from 5 year-
388 age and sex specific all-cause mortality rates from ITACAN.

389 The attributable risk of the PRS was estimated using the Bruzzi formula (54).

390

391

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406

407 **CONFLICTS OF INTEREST**

408 The authors declare no conflicts of interest

409

410 **REFERENCES**

- 411 1 Christenson, L.J., Borrowman, T.A., Vachon, C.M., Tollefson, M.M., Otley, C.C.,
412 Weaver, A.L. and Roenigk, R.K. (2005) Incidence of basal cell and squamous cell carcinomas in
413 a population younger than 40 years. *Jama*, **294**, 681-690.
- 414 2 Cancer Facts & Figures. (2017). American Cancer Society, pp. 24-25.
- 415 3 Weinstock, M.A. (2001) Epidemiology, etiology, and control of melanoma. *Medicine and*
416 *health, Rhode Island*, **84**, 234-236.
- 417 4 Long, G.V., Hauschild, A., Santinami, M., Atkinson, V., Mandala, M., Chiarion-Sileni,
418 V., Larkin, J., Nyakas, M., Dutriaux, C., Haydon, A. *et al.* (2017) Adjuvant Dabrafenib plus
419 Trametinib in Stage III BRAF-Mutated Melanoma. *The New England journal of medicine*, **377**,
420 1813-1823.
- 421 5 Weber, J., Mandala, M., Del Vecchio, M., Gogas, H.J., Arance, A.M., Cowey, C.L.,
422 Dalle, S., Schenker, M., Chiarion-Sileni, V., Marquez-Rodas, I. *et al.* (2017) Adjuvant
423 Nivolumab versus Ipilimumab in Resected Stage III or IV Melanoma. *The New England journal*
424 *of medicine*, **377**, 1824-1835.
- 425 6 Wolchok, J.D., Chiarion-Sileni, V., Gonzalez, R., Rutkowski, P., Grob, J.J., Cowey, C.L.,
426 Lao, C.D., Wagstaff, J., Schadendorf, D., Ferrucci, P.F. *et al.* (2017) Overall Survival with
427 Combined Nivolumab and Ipilimumab in Advanced Melanoma. *The New England journal of*
428 *medicine*, **377**, 1345-1356.
- 429 7 Gershenwald, J.E., Scolyer, R.A., Hess, K.R., Sondak, V.K., Long, G.V., Ross, M.I.,
430 Lazar, A.J., Faries, M.B., Kirkwood, J.M., McArthur, G.A. *et al.* (2017) Melanoma staging:
431 Evidence-based changes in the American Joint Committee on Cancer eighth edition cancer
432 staging manual. *CA: a cancer journal for clinicians*, **67**, 472-492.
- 433 8 Cho, E., Rosner, B.A., Feskanich, D. and Colditz, G.A. (2005) Risk factors and
434 individual probabilities of melanoma for whites. *Journal of clinical oncology : official journal of*
435 *the American Society of Clinical Oncology*, **23**, 2669-2675.
- 436 9 Davies, J.R., Chang, Y.M., Bishop, D.T., Armstrong, B.K., Bataille, V., Bergman, W.,
437 Berwick, M., Bracci, P.M., Elwood, J.M., Ernstoff, M.S. *et al.* (2015) Development and
438 validation of a melanoma risk score based on pooled data from 16 case-control studies. *Cancer*
439 *epidemiology, biomarkers & prevention : a publication of the American Association for Cancer*
440 *Research, cosponsored by the American Society of Preventive Oncology*, **24**, 817-824.
- 441 10 Fears, T.R., Guerry, D.t., Pfeiffer, R.M., Sagebiel, R.W., Elder, D.E., Halpern, A., Holly,
442 E.A., Hartge, P. and Tucker, M.A. (2006) Identifying individuals at high risk of melanoma: a
443 practical predictor of absolute risk. *Journal of clinical oncology : official journal of the*
444 *American Society of Clinical Oncology*, **24**, 3590-3596.
- 445 11 Fortes, C., Mastroeni, S., Bakos, L., Antonelli, G., Alessandroni, L., Pilla, M.A., Alotto,
446 M., Zappala, A., Manoorannparampill, T., Bonamigo, R. *et al.* (2010) Identifying individuals at
447 high risk of melanoma: a simple tool. *European journal of cancer prevention : the official*
448 *journal of the European Cancer Prevention Organisation (ECP)*, **19**, 393-400.
- 449 12 Guthrie, S., Ramrath, K., Dyal-Smith, D., Landthaler, M. and Stolz, W. (2012)
450 Development of a targeted risk-group model for skin cancer screening based on more than
451 100,000 total skin examinations. *Journal of the European Academy of Dermatology and*
452 *Venereology : JEADV*, **26**, 86-94.

453 13 MacKie, R.M., Freudenberger, T. and Aitchison, T.C. (1989) Personal risk-factor chart
454 for cutaneous melanoma. *Lancet (London, England)*, **2**, 487-490.

455 14 Mar, V., Wolfe, R. and Kelly, J.W. (2011) Predicting melanoma risk for the Australian
456 population. *The Australasian journal of dermatology*, **52**, 109-116.

457 15 Molinaro, A.M., Ferrucci, L.M., Cartmel, B., Loftfield, E., Leffell, D.J., Bale, A.E. and
458 Mayne, S.T. (2015) Indoor tanning and the MC1R genotype: risk prediction for basal cell
459 carcinoma risk in young people. *American journal of epidemiology*, **181**, 908-916.

460 16 Olsen, C.M., Neale, R.E., Green, A.C., Webb, P.M., The, Q.S., The Epigene, S. and
461 Whiteman, D.C. (2015) Independent validation of six melanoma risk prediction models. *The*
462 *Journal of investigative dermatology*, **135**, 1377-1384.

463 17 Quereux, G., Moyse, D., Lequeux, Y., Jumbou, O., Brocard, A., Antonioli, D., Dreno, B.
464 and Nguyen, J.M. (2011) Development of an individual score for melanoma risk. *European*
465 *journal of cancer prevention : the official journal of the European Cancer Prevention*
466 *Organisation (ECP)*, **20**, 217-224.

467 18 Schuchter, L., Schultz, D.J., Synnestvedt, M., Trock, B.J., Guerry, D., Elder, D.E.,
468 Elenitsas, R., Clark, W.H. and Halpern, A.C. (1996) A prognostic model for predicting 10-year
469 survival in patients with primary melanoma. The Pigmented Lesion Group. *Annals of internal*
470 *medicine*, **125**, 369-375.

471 19 Sneyd, M.J., Cameron, C. and Cox, B. (2014) Individual risk of cutaneous melanoma in
472 New Zealand: developing a clinical prediction aid. *BMC cancer*, **14**, 359.

473 20 Williams, L.H., Shors, A.R., Barlow, W.E., Solomon, C. and White, E. (2011)
474 Identifying Persons at Highest Risk of Melanoma Using Self-Assessed Risk Factors. *Journal of*
475 *clinical & experimental dermatology research*, **2**.

476 21 Mucci, L.A., Hjelmberg, J.B., Harris, J.R., Czene, K., Havelick, D.J., Scheike, T., Graff,
477 R.E., Holst, K., Moller, S., Unger, R.H. *et al.* (2016) Familial Risk and Heritability of Cancer
478 Among Twins in Nordic Countries. *Jama*, **315**, 68-76.

479 22 Goldstein, A.M., Chan, M., Harland, M., Hayward, N.K., Demenais, F., Bishop, D.T.,
480 Azizi, E., Bergman, W., Bianchi-Scarra, G., Bruno, W. *et al.* (2007) Features associated with
481 germline CDKN2A mutations: a GenoMEL study of melanoma-prone families from three
482 continents. *Journal of medical genetics*, **44**, 99-106.

483 23 Zuo, L., Weger, J., Yang, Q., Goldstein, A.M., Tucker, M.A., Walker, G.J., Hayward, N.
484 and Dracopoli, N.C. (1996) Germline mutations in the p16INK4a binding domain of CDK4 in
485 familial melanoma. *Nature genetics*, **12**, 97-99.

486 24 Wiesner, T., Obenaus, A.C., Murali, R., Fried, I., Griewank, K.G., Ulz, P.,
487 Windpassinger, C., Wackernagel, W., Loy, S., Wolf, I. *et al.* (2011) Germline mutations in
488 BAP1 predispose to melanocytic tumors. *Nature genetics*, **43**, 1018-1021.

489 25 Horn, S., Figl, A., Rachakonda, P.S., Fischer, C., Sucker, A., Gast, A., Kadel, S., Moll, I.,
490 Nagore, E., Hemminki, K. *et al.* (2013) TERT promoter mutations in familial and sporadic
491 melanoma. *Science (New York, N.Y.)*, **339**, 959-961.

492 26 Robles-Espinoza, C.D., Harland, M., Ramsay, A.J., Aoude, L.G., Quesada, V., Ding, Z.,
493 Pooley, K.A., Pritchard, A.L., Tiffen, J.C., Petljak, M. *et al.* (2014) POT1 loss-of-function
494 variants predispose to familial melanoma. *Nature genetics*, **46**, 478-481.

495 27 Shi, J., Yang, X.R., Ballew, B., Rotunno, M., Calista, D., Fargnoli, M.C., Ghiorzo, P.,
496 Bressac-de Paillerets, B., Nagore, E., Avril, M.F. *et al.* (2014) Rare missense variants in POT1
497 predispose to familial cutaneous malignant melanoma. *Nature genetics*, **46**, 482-486.

498 28 Aoude, L.G., Pritchard, A.L., Robles-Espinoza, C.D., Wadt, K., Harland, M., Choi, J.,
499 Gartside, M., Quesada, V., Johansson, P., Palmer, J.M. *et al.* (2015) Nonsense mutations in the
500 shelterin complex genes ACD and TERF2IP in familial melanoma. *JNCI Journal of the National*
501 *Cancer Institute*, **107**.

502 29 Hu, H.H., Kannengiesser, C., Lesage, S., Andre, J., Mourah, S., Michel, L., Descamps,
503 V., Basset-Seguin, N., Bagot, M., Bensussan, A. *et al.* (2016) PARKIN Inactivation Links
504 Parkinson's Disease to Melanoma. *JNCI Journal of the National Cancer Institute*, **108**.

505 30 Yokoyama, S., Woods, S.L., Boyle, G.M., Aoude, L.G., MacGregor, S., Zismann, V.,
506 Gartside, M., Cust, A.E., Haq, R., Harland, M. *et al.* (2011) A novel recurrent mutation in MITF
507 predisposes to familial and sporadic melanoma. *Nature*, **480**, 99-103.

508 31 Lu, Y., Ek, W.E., Whiteman, D., Vaughan, T.L., Spurdle, A.B., Easton, D.F., Pharoah,
509 P.D., Thompson, D.J., Dunning, A.M., Hayward, N.K. *et al.* (2014) Most common 'sporadic'
510 cancers have a significant germline genetic component. *Hum. Mol. Genet.*, **23**, 6112-6118.

511 32 Law, M.H., Bishop, D.T., Lee, J.E., Brossard, M., Martin, N.G., Moses, E.K., Song, F.,
512 Barrett, J.H., Kumar, R., Easton, D.F. *et al.* (2015) Genome-wide meta-analysis identifies five
513 new susceptibility loci for cutaneous malignant melanoma. *Nature genetics*, **47**, 987-995.

514 33 Kenny, E.E., Timpson, N.J., Sikora, M., Yee, M.C., Moreno-Estrada, A., Eng, C.,
515 Huntsman, S., Burchard, E.G., Stoneking, M., Bustamante, C.D. *et al.* (2012) Melanesian blond
516 hair is caused by an amino acid change in TYRP1. *Science (New York, N.Y.)*, **336**, 554.

517 34 Liu, F., Visser, M., Duffy, D.L., Hysi, P.G., Jacobs, L.C., Lao, O., Zhong, K., Walsh, S.,
518 Chaitanya, L., Wollstein, A. *et al.* (2015) Genetics of skin color variation in Europeans: genome-
519 wide association studies with functional follow-up. *Human genetics*, **134**, 823-835.

520 35 Duffy, D.L., Iles, M.M., Glass, D., Zhu, G., Barrett, J.H., Hoim, V., Zhao, Z.Z., Sturm,
521 R.A., Soranzo, N., Hammond, C. *et al.* (2010) IRF4 variants have age-specific effects on nevus
522 count and predispose to melanoma. *American journal of human genetics*, **87**, 6-16.

523 36 Falchi, M., Bataille, V., Hayward, N.K., Duffy, D.L., Bishop, J.A., Pastinen, T., Cervino,
524 A., Zhao, Z.Z., Deloukas, P., Soranzo, N. *et al.* (2009) Genome-wide association study identifies
525 variants at 9p21 and 22q13 associated with development of cutaneous nevi. *Nature genetics*, **41**,
526 915-919.

527 37 Fang, S., Han, J., Zhang, M., Wang, L.E., Wei, Q., Amos, C.I. and Lee, J.E. (2013) Joint
528 effect of multiple common SNPs predicts melanoma susceptibility. *PloS one*, **8**, e85642.

529 38 Stefanaki, I., Panagiotou, O.A., Kodela, E., Gogas, H., Kypreou, K.P., Chatzinasiou, F.,
530 Nikolaou, V., Plaka, M., Kalfa, I., Antoniou, C. *et al.* (2013) Replication and predictive value of
531 SNPs associated with melanoma and pigmentation traits in a Southern European case-control
532 study. *PloS one*, **8**, e55712.

533 39 Kypreou, K.P., Stefanaki, I., Antonopoulou, K., Karagianni, F., Ntritsos, G., Zaras, A.,
534 Nikolaou, V., Kalfa, I., Chasapi, V., Polydorou, D. *et al.* (2016) Prediction of Melanoma Risk in
535 a Southern European Population Based on a Weighted Genetic Risk Score. *The Journal of*
536 *investigative dermatology*, **136**, 690-695.

537 40 Dudbridge, F. (2013) Power and predictive accuracy of polygenic risk scores. *PLoS*
538 *genetics*, **9**, e1003348.

539 41 Ford, D., Bliss, J.M., Swerdlow, A.J., Armstrong, B.K., Franceschi, S., Green, A., Holly,
540 E.A., Mack, T., MacKie, R.M., Osterlind, A. *et al.* (1995) Risk of cutaneous melanoma
541 associated with a family history of the disease. The International Melanoma Analysis Group
542 (IMAGE). *International journal of cancer*, **62**, 377-381.

543 42 Chatterjee, N., Wheeler, B., Sampson, J., Hartge, P., Chanock, S.J. and Park, J.H. (2013)
544 Projecting the performance of risk prediction based on polygenic analyses of genome-wide
545 association studies. *Nature genetics*, **45**, 400-405, 405e401-403.

546 43 Shi, J., Park, J.H., Duan, J., Berndt, S.T., Moy, W., Yu, K., Song, L., Wheeler, W., Hua,
547 X., Silverman, D. *et al.* (2016) Winner's Curse Correction and Variable Thresholding Improve
548 Performance of Polygenic Risk Modeling Based on Genome-Wide Association Study Summary-
549 Level Data. *PLoS genetics*, **12**, e1006493.

550 44 Mavaddat, N., Pharoah, P.D.P., Michailidou, K., Tyrer, J., Brook, M.N., Bolla, M.K.,
551 Wang, Q., Dennis, J., Dunning, A.M., Shah, M. *et al.* (2015) Prediction of Breast Cancer Risk
552 Based on Profiling With Common Genetic Variants. *JNCI Journal of the National Cancer
553 Institute*, **107**.

554 45 Song, M., Kraft, P., Joshi, A.D., Barrdahl, M. and Chatterjee, N. (2015) Testing
555 calibration of risk models at extremes of disease risk. *Biostatistics (Oxford, England)*, **16**, 143-
556 154.

557 46 Purcell, S.M., Wray, N.R., Stone, J.L., Visscher, P.M., O'Donovan, M.C., Sullivan, P.F.
558 and Sklar, P. (2009) Common polygenic variation contributes to risk of schizophrenia and
559 bipolar disorder. *Nature*, **460**, 748-752.

560 47 Purcell, S., Neale, B., Todd-Brown, K., Thomas, L., Ferreira, M.A., Bender, D., Maller,
561 J., Sklar, P., de Bakker, P.I., Daly, M.J. *et al.* (2007) PLINK: a tool set for whole-genome
562 association and population-based linkage analyses. *American journal of human genetics*, **81**,
563 559-575.

564 48 Vilhjalmsjon, B.J., Yang, J., Finucane, H.K., Gusev, A., Lindstrom, S., Ripke, S.,
565 Genovese, G., Loh, P.R., Bhatia, G., Do, R. *et al.* (2015) Modeling Linkage Disequilibrium
566 Increases Accuracy of Polygenic Risk Scores. *American journal of human genetics*, **97**, 576-592.

567 49 Robin, X., Turck, N., Hainard, A., Tiberti, N., Lisacek, F., Sanchez, J.C. and Muller, M.
568 (2011) pROC: an open-source package for R and S+ to analyze and compare ROC curves. *BMC
569 bioinformatics*, **12**, 77.

570 50 Raghunathan TES, P.W., Hoewyk, J.V. (2002) IVEWARE: Imputation and variance
571 estimation software. University of Michigan.

572 51 Little, R.J. and Rubin, D.B. (2000) Causal effects in clinical and epidemiological studies
573 via potential outcomes: concepts and analytical approaches. *Annual review of public health*, **21**,
574 121-145.

575 52 Bartholomew, D. (1987) *Latent variable models and factor analysis*. Griffin, London.

576 53 Ruth M. Pfeiffer, Mitchell H. Gail. (2017) *Absolute Risk: Methods and Applications in
577 Clinical Management and Public Health*. Chapman and Hall/CRC Reference.

578 54 Bruzzi, P., Green, S.B., Byar, D.P., Brinton, L.A. and Schairer, C. (1985) Estimating the
579 population attributable risk for multiple risk factors using case-control data. *American journal of
580 epidemiology*, **122**, 904-914.

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582

583 **FIGURE LEGENDS**

584 **Figure 1.** Area under the curve (AUC) and 95% confidence intervals (CI) for three different
585 models. Model 2 (see Methods section for details) used LD clumping $r^2=0.01$, and different p-
586 value thresholds for SNP inclusion. Model 3 was constructed using LDpred (47). Model 4 is
587 similar to Model 2 but corrects the effect size estimation for winner's curse (43). Model 1 is not
588 represented in the Figure; it has AUC=62.8%.

589
590 **Figure 2.** 10- and 20-year absolute risk of melanoma for Italian men (Fig. 2a) and women (Fig.
591 2b), by age and risk profile. The absolute risk was estimated in a model that includes the PRS
592 and other established risk factors, using age- and sex- specific incidence rates of melanoma as
593 well as death rates of other causes from the Italian population. Corresponding risks are also
594 shown in Supplementary Table 5.

595

596 **SUPPLEMENTARY MATERIAL**

597 **Supplementary Figure 1.** Quantile-quantile plot for the genome wide association study of
598 melanoma in the MelaNostrum consortium

599 **Supplementary Table 1a.** Characteristics of 5403 subjects in the MelaNostrum Consortium
600 (validation set), by country. **Supplementary Table 1b.** Characteristics of subjects in the
601 MelaNostrum Consortium (validation set), by Italian sites

602 **Supplementary Table 2.** Performance of risk prediction model (area under the curve) based on
603 continuous polygenic risk score by p-value thresholds and study site

604 **Supplementary Table 3.** 204 SNPs in the polygenic risk score using the Winner's Curse
605 Correction mode

606 **Supplementary Table 4.** Association between traditional risk factors and melanoma risk in the
607 Mediterranean population using imputed data

608 **Supplementary Table 5.** 10- and 20-year absolute risk of melanoma for Italian men and women,
609 by age and risk profile.

610 **Supplementary Table 6.** Comparing the effect size and minor allele frequency of the 204 SNPs
611 in the PRS between the Melanoma Meta-Analysis Consortium and MelaNostrum Consortium
612 data

613 **ONLINE DATA SUPPLEMENTS:** Study populations in MelaNostrum consortium and
614 supplementary methods

615

616

Table 1. Characteristics of the MelaNostrum study population (n=5403)

		Case		Control	
		N=3102	%	N=2301	%
Study site	Greece	775	25.0	752	32.7
	Italy	1266	40.8	361	15.7
	Spain	1061	34.2	1188	51.6
Sex ^a	Male	1453	46.8	1241	53.9
	Female	1649	53.2	1060	46.1
Age ^b	≤29	241	7.8	544	23.6
	30-39	494	15.9	576	25.0
	40-49	652	21.0	528	22.9
	50-59	636	20.5	319	13.9
	60-77	870	28.0	242	10.5
	≥78	143	4.6	26	1.1
	Missing	66	2.1	66	2.9
Family history of melanoma	No	1919	61.9	853	37.1
	Yes	227	7.3	159	6.9
	Missing	956	30.8	1289	56.0
Eye color ^a	Dark	1198	38.6	1262	54.8
	Medium	1065	34.3	644	28.0
	Light	575	18.5	241	10.5
	Missing	264	8.5	154	6.7
Hair color ^a	Black	323	10.4	342	14.9
	Dark brown/light/reddish brown	1874	60.4	1607	69.8
	Blond	486	15.7	147	6.4
	Red	126	4.1	37	1.6
	Missing	293	9.4	168	7.3
Skin phototype ^a	III-VI	1521	49.0	1250	54.3
	I-II	1349	43.5	779	33.9
	Missing	232	7.5	272	11.8
Nevi ^a	≤50	816	26.3	1143	49.7
	>50	1702	54.9	631	27.4
	Missing	584	18.8	527	22.9
Acute sun damage ^c	No	465	43.8	782	65.8
	Yes	521	49.1	334	28.1
	Missing	75	7.1	72	6.1
Chronic sun damage ^c	No	822	77.5	1089	91.7
	Yes	180	17.0	44	3.7
	Missing	59	5.6	55	4.6
Sunburns ^c	No	604	26.0	238	10.2

	Yes	1458	62.7	1123	48.2
	Missing	265	11.4	188	8.1
Intermittent sun exposure ^c	No/some	740	58.5	248	68.7
	High	429	33.9	83	23.0
	Missing	97	7.7	30	8.3
Chronic sun exposure ^c	No	1288	55.4	1108	71.5
	Yes	483	20.8	298	19.2
	Missing	556	23.9	143	9.2
Melanoma body site	Head/neck	347	11.2		
	Trunk	1254	40.4		
	Upper limbs	383	12.3		
	Lower limbs	703	22.7		
	Hands/feet	154	5.0		
	Unknown	212	6.8		
	Missing	49	1.6		
Melanoma type	SSM	1733	55.9		
	NM	365	11.8		
	LM	162	5.2		
	Acral	88	2.8		
	Mucosal	2	0.1		
	Undetermined	298	9.6		
	Missing	454	14.6		
Multiple melanoma	No	2564	82.7		
	Yes	342	11.0		
	Missing	196	6.3		
Thickness according to Breslow (mm)	<1.00	1060	34.2		
	1.01-2.00	440	14.2		
	2.01-4.00	335	10.8		
	>4.00	215	6.9		
	Undetermined	176	5.7		
	Missing	876	28.2		

619 ^aVariables included in all analyses

620 ^bAge at diagnosis for cases and age at study enrollment for controls

621 ^cDue to high missing rates in some studies, these variables were only evaluated, and therefore
622 presented here, in subgroups of studies: acute and chronic sun damage are included in the
623 Spanish study; intermittent sun exposure is included in the Italian study; sunburn and chronic sun
624 exposure are included in both the Spanish and Italian studies.

625

626

627 **Table 2. Correlation of polygenic risk score^a and phenotypes in the MelaNostrum control**
628 **population, overall and by country of residence**
629

Phenotype	Corr	P	N
Overall			
Sex: 0=male 1=female	-0.01	0.55	2301
Age ^b	0.04	0.07	2235
Nevus count: 1= ≤50; 2= >50	0.13	<0.0001	1774
Eye color: 0=dark, 1=medium, 2=light	0.09	<0.0001	2147
Hair color: 1=black,2=dark brown/light/reddish brown, 3=blond,4=red	0.14	<0.0001	2144
Skin phototype: 0=III-VI; 1=I-II	0.15	<0.0001	2029
Greece			
Sex: 0=male 1=female	-0.05	0.14	752
Age ^b	0.003	0.94	692
Nevus count: 1= ≤50; 2= >50	0.16	0.006	313
Eye color: 0=dark, 1=medium, 2=light	0.1	0.02	634
Hair color: 1=black,2=dark brown/light/reddish brown, 3=blond,4=red	0.18	<0.0001	636
Skin phototype: 0=III-VI; 1=I-II	0.17	<0.0001	623
Italy			
Sex: 0=male 1=female	-0.02	0.74	361
Age ^b	-0.04	0.41	358
Nevus count: 1= ≤50; 2= >50	0.07	0.22	304
Eye color: 0=dark, 1=medium, 2=light	0.09	0.10	354
Hair color: 1=black,2=dark brown/light/reddish brown, 3=blond,4=red	0.14	0.008	345
Skin phototype: 0=III-IV; 1=I-II	0.23	<0.0001	355
Sunburns: 0=no; 1=yes	0.04	0.53	249
Intermittent sun exposure: 0=none/some; 1=high	-0.04	0.53	331
Chronic sun exposure: 0=no; 1=yes	-0.04	0.50	249
Spain			
Sex: 0=male 1=female	0.01	0.83	1188
Age ^b	0.02	0.50	1185
Nevus count: 1= ≤50; 2= >50	0.07	0.02	1157
Eye color: 0=dark, 1=medium, 2=light	0.03	0.26	1159
Hair color: 1=black,2=dark brown/light/reddish brown, 3=blond,4=red	0.11	0.0001	1152
Skin phototype: 0=III-VI; 1=I-II	0.11	0.0002	1095
Acute sun damage: 0=no; 1=yes	-0.01	0.81	1116
Chronic sun damage (actinic keratoses): 0=no; 1=yes	0.05	0.06	1133
Sunburns: 0=no; 1=yes	-0.007	0.81	1112
Chronic sun exposure: 0=no; 1=yes	-0.001	0.95	1157

630 ^aContinuous score based on the best winner's curse model
631 ^bAge at diagnosis for cases and age at study enrollment for controls
632

633 **Table 3. Odds Ratios (OR) between PRS and melanoma risk, adjusting for different**
634 **melanoma risk factors**
635

	OR _{per} quintile	L95	U95	P	OR _{5th vs.1st} quintile
Overall					
PRS	1.35	1.30	1.41	<0.0001	3.33
PRS+Demographics ^a	1.35	1.29	1.41	<0.0001	3.30
PRS+Demographics + pigmentation ^b + nevi	1.23	1.13	1.35	<0.0001	2.32
Greece					
PRS	1.31	1.22	1.42	<0.0001	2.98
PRS+Demographics ^a	1.33	1.23	1.44	<0.0001	3.11
PRS+Demographics + pigmentation ^b + nevi	1.29	1.19	1.40	<0.0001	2.76
Italy					
PRS	1.32	1.21	1.43	<0.0001	3.04
PRS+Demographics ^a	1.32	1.21	1.44	<0.0001	3.02
PRS+Demographics + pigmentation ^b + nevi	1.23	1.13	1.35	0.0003	2.32
PRS+Fully adjusted ^c	1.23	1.12	1.35	<0.0001	2.29
Spain					
PRS	1.40	1.31	1.48	<0.0001	3.79
PRS+Demographics ^a	1.38	1.29	1.48	<0.0001	3.63
PRS+Demographics + pigmentation ^b + nevi	1.26	1.16	1.37	<0.0001	2.55
PRS+Fully adjusted ^d	1.27	1.17	1.38	<0.0001	2.62

636 PRS: Polygenic risk score

637 ^a Demographic includes age, sex, and country (for overall population)

638 ^b Pigmentation includes eye color, hair color and skin phototype

639 ^c Full model in the Italian population additionally adjusted for chronic sun exposure, intermittent
640 sun exposure, and history of sunburns

641 ^d Full model in the Spanish population additionally adjusted for chronic sun exposure, chronic
642 sun damage, acute sun damage and history of sunburns

643

644 **Table 4. Performance of risk prediction model with and without polygenic risk score**
645

Traditional covariates in models	Area Under the Curve (95% CI)			P-difference
	Model without PRS	Model with PRS	AUC difference	
Overall				
Demographic ^a	76.5% (75.2%-77.8%)	78.2% (77.0%-79.4%)	1.7% (1.1%-2.2%)	<0.0001
Demographic+pigmentation ^b +nevi	80.1% (78.9%-81.3%)	81.0% (79.8%-82.2%)	0.8% (0.5%-1.2%)	<0.0001
Greece				
Demographic ^a	67.9% (65.2%-70.7%)	70.7% (68.0%-73.4%)	2.7% (1.3%-4.1%)	0.0002
Demographic + pigmentation ^b + nevi	69.8% (67.1%-72.5%)	71.7% (69.1%-74.4%)	1.9% (0.7%-3.1%)	0.003
Italy				
Demographic ^a	53.9% (50.6%-57.2%)	61.2% (57.8%-64.5%)	7.3% (3.4%-11.2%)	0.0001
Demographic + pigmentation ^b + nevi	64.8% (61.6%-68.1%)	66.6% (63.4%-69.8%)	1.7% (0.6%-3.0%)	0.04
Fully adjusted ^c	67.0% (63.7%-70.3%)	68.5% (65.4%-71.7%)	1.4% (-0.1%-2.9%)	0.07
Spain				
Demographic ^a	78.6% (76.7%-80.5%)	80.6% (78.8%-82.4%)	2.0% (1.2%-2.8%)	<0.0001
Demographic + pigmentation ^b + nevi	87.7% (86.3%-89.3%)	88.3% (86.8%-89.7%)	0.5% (0.1%-0.8%)	0.005
Fully adjusted ^d	88.7% (87.3%-90.1%)	89.1% (87.6%-90.5%)	0.4% (0.1%-0.7%)	0.005

646 PRS: Polygenic risk score

647 ^a Demographic includes age, sex, and country (for overall population)

648 ^b Pigmentation includes eye color, hair color and skin phototype

649 ^c Full model in the Italian population additionally adjusted for chronic sun exposure, intermittent sun exposure, and history of sunburns

651 ^d Full model in the Spanish population additionally adjusted for chronic sun exposure, chronic sun damage, acute sun damage and history of sunburns

652
653

654 **ABBREVIATIONS**

655 SNPs: single nucleotide polymorphisms

656 PRS: polygenic risk scores

657 OR: odds ratio

658 CI: confidence intervals

659 AUC: Operator Characteristics curve

660 GWAS: Genome-wide association study

661 LD: linkage disequilibrium