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3 **Alemtuzumab-induced halo nevus-like hypopigmentation –**  
4 **New insights into secondary skin autoimmunity in response**  
5 **to an immune cell-depleting antibody**  
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57 The authors of this manuscript declare no conflicts of interest  
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3 Alemtuzumab is an antibody directed against the Cluster of Differentiation (CD)52. It  
4 depletes B and T lymphocytes and is approved for treatment of active relapsing-  
5 remitting multiple sclerosis (RRMS). Secondary autoimmunity is an important risk of  
6 alemtuzumab-treated patients and can affect different organs. 41% of patients develop  
7 autoimmune thyroid disease<sup>2</sup>. Skin autoimmunity under alemtuzumab has but  
8 reported only in the neurological literature<sup>3,4</sup>.  
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11 We describe a 33-year-old male patient who developed hypopigmentation around his  
12 melanocytic nevi with disappearance of the nevi in August 2018. In June 2016 he had  
13 been diagnosed with highly active RRMS and treated with alemtuzumab in September  
14 2016 for the first time. In September 2017 another cycle of alemtuzumab was  
15 administered. The patient's family history for vitiligo was unremarkable and he had  
16 no autoimmune thyroid disease. At time of the occurrence of the hypopigmented spots  
17 the number of his circulating leukocytes in the blood was normal. Upon examination  
18 he showed sharply demarcated hypopigmented spots around his melanocytic nevi  
19 (Fig. 1A). Some of the nevi within the white spots had already disappeared. There  
20 were no other signs of skin hypopigmentation. A biopsy specimen from one of the  
21 halo-like nevi (insert of Fig. 1A) revealed absence of epidermal melanocytes (Fig. 1B)  
22 which was confirmed by immunohistochemistry with an anti-Pan Mel antibody (Fig.  
23 1C). In contrast, melanocytes were detected within the nevus (Fig. 1C).  
24 Immunostaining with an anti-CD3 antibody disclosed a sparsely scattered infiltrate of  
25 T lymphocytes within the upper dermis (Fig. 1D, E). To shed light into the  
26 pathogenesis of the halo nevus-like lesions we wondered whether alemtuzumab might  
27 directly attack epidermal melanocytes. CD52 immunohistochemistry of normal skin  
28 and a melanocytic nevus from a healthy person did not show immunoreactivity in  
29 melanocytes in contrast to lymphoid tissues used as positive control (data not shown).  
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3 Since autoimmune thyroid disease is characterized by circulating anti-thyroid  
4 antibodies, we hypothesized whether anti-melanocyte antibodies may be generated  
5 following alemtuzumab treatment. In patients with non-segmental vitiligo (NSV)  
6 antibodies against tyrosinase (TYR), the key enzyme of melanin synthesis, were also  
7 detected<sup>5</sup>. Therefore, a TnT® T7-Coupled Reticulocyte Lysate System (Promega,  
8 Southampton, UK) was used to produce various [<sup>35</sup>S]-labelled melanocyte antigens *in*  
9 *vitro* from the translation of cDNA in the appropriate plasmid. Serum samples of the  
10 patient before and after alemtuzumab therapy were then analysed for circulating  
11 antibodies against these antigens by radioligand binding assays<sup>5</sup>. An almost 6-fold  
12 increase in anti-TYR antibodies and a ~3-fold increase in antibodies against TYR-  
13 related protein 1 were detected following alemtuzumab (Table 1). In contrast, anti-  
14 thyroid antibodies were not detected throughout the patient's history. In accordance  
15 with this TSH levels were always normal.

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34 Our findings are puzzling as alemtuzumab depletes both B and T cells but still evokes  
35 an autoimmune attack against melanocytes. Recently, three patients with RRMS have  
36 been described who developed NSV 14, 18 and 52 months after initiation of  
37 alemtuzumab<sup>6</sup>. The immune-mediated destruction pathways of melanocytes in classic  
38 halo nevus and NSV may be different<sup>7</sup>. It has been suggested that secondary  
39 autoimmunity induced by alemtuzumab is related to proliferation of chronically  
40 activated, oligoclonal, effector memory CD8<sup>+</sup> T cells<sup>8</sup>. Interestingly, resident and  
41 effector memory T cells have been identified in non-active skin of patients with  
42 NSV<sup>9</sup>. Previous studies showed that alemtuzumab depletes all T cells from the blood  
43 but does not deplete skin-resident memory T cells<sup>10</sup>. Thus, production of anti-  
44 melanocyte-specific antibodies may be secondary to initial destruction of melanocytes  
45 by melanocyte-specific CD8<sup>+</sup> T cells.

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3 In summary, alemtuzumab-induced skin autoimmunity is a condition a dermatologist  
4 should be aware of. Our report underscores the complex pathogenesis of immune-  
5 mediated destruction of epidermal melanocytes that may follow even upon depletion  
6 of both T and B cells.  
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19 The patient in this manuscript has given written informed consent to the publication of  
20 his case details.  
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**Figure 1**

(A) Clinical appearance of the posterior trunk of the patient. The insert depicts the halo-like nevus from which the biopsy was taken; (B) Haematoxylin & eosin staining and (C) Pan Mel immunohistochemistry of the lesion (as shown in insert of Fig 1A). Note the absence of epidermal melanocytes; (D, E) CD3 immunostaining of the lesion. Note scattered T cells adjacent to the dermal melanocytes (D) and along the dermal-epidermal junction devoid of melanocytes (E).

For Peer Review

**Table 1:** Presence of circulating anti-melanocyte antibodies in the patient's serum before (Sept/2016) and after initiation of alemtuzumab (Aug/2018 and Feb/2019).

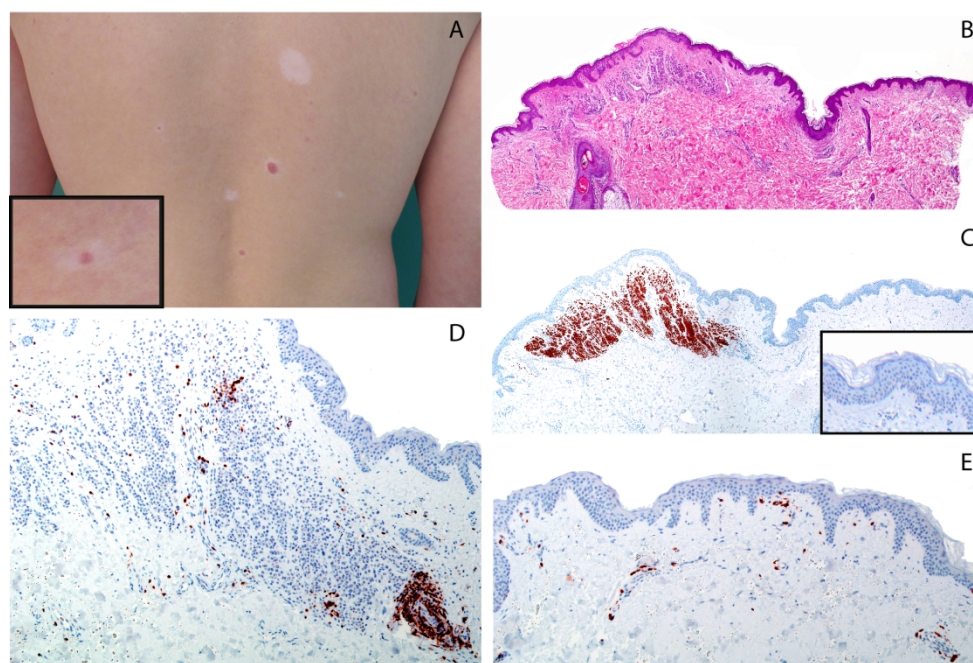
Antigen used in radioligand binding assay <sup>1</sup>	Before treatment antibody index <sup>2</sup>	After <sup>2</sup> treatment antibody index <sup>2</sup>	After <sup>2</sup> treatment antibody index <sup>2</sup>	Upper limit of normal for radioligand binding assay <sup>3</sup>
<b>LMNA</b>	1.23	1.09	1.15	1.59
<b>TYR</b>	<b>2.23</b>	<b>12.89</b>	<b>13.12</b>	1.72
<b>TYRP1</b>	1.25	<b>3.04</b>	<b>3.89</b>	1.57
<b>DCT</b>	0.95	0.99	1.03	1.36
<b>PMEL</b>	1.14	1.26	1.19	1.45
<b>TH</b>	1.09	1.12	1.15	1.84
<b>MCHR1</b>	0.95	0.94	0.98	1.48

<sup>1</sup>Radioligand binding assays (RLBA) were used to test for antibodies against LMNA, laminA, TYR, tyrosinase; TYRP1, TYR-related protein 1; DCT, dopachrome tautomerase; PMEL, premelanosome protein; TH, tyrosine hydroxylase; and MCHR1, melanin-concentrating hormone receptor 1.

<sup>2</sup>An antibody index for the patient's serum was calculated for each RLBA as: counts per min (cpm) immunoprecipitated by serum/mean cpm immunoprecipitated by 20 healthy control sera. Positive antibody indices are in bold.

<sup>3</sup>The upper limit of normal for the RLBA was calculated using the mean antibody index + 3 SD of the population of 20 healthy control sera.





(A) Clinical appearance of the posterior trunk of the patient. The insert depicts the halo-like nevus from which the biopsy was taken; (B) Haematoxylin & eosin staining and (C) Pan Mel immunohistochemistry of the lesion (as shown in insert of Fig 1A). Note the absence of epidermal melanocytes; (D, E) CD3 immunostaining of the lesion. Note scattered T cells adjacent to the dermal melanocytes (D) and along the dermal-epidermal junction devoid of melanocytes (E).

196x136mm (300 x 300 DPI)