



UNIVERSITY OF LEEDS

This is a repository copy of *On Measuring the Specific Surface Area of inhalation-grade lactose powders*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/160915/>

Version: Accepted Version

Proceedings Paper:

Styliari, ID, Nguyen, TTH, Gajjar, P et al. (6 more authors) (2020) On Measuring the Specific Surface Area of inhalation-grade lactose powders. In: Proceedings of the Respiratory Drug Delivery 2020 conference (3). Respiratory Drug Delivery 2020, 26 Apr - 30 Jun 2020, Online conference. Respiratory Drug Delivery , pp. 551-554. ISBN 9781942911487

Copyright© 2020 by Virginia Commonwealth University. Reproduced with permission from Respiratory Drug Delivery 2020, Virginia Commonwealth University and RDD Online.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

On Measuring the Specific Surface Area of inhalation-grade lactose powders

Ioanna Danai Styliari¹, Thai T.H. Nguyen², Parmesh Gajjar³, Benjamin Tordoff⁴, Timothy L. Burnett³, Philip J. Withers³, Robert Hammond², Kevin Roberts² and Darragh Murnane²

¹ School of Life and Medical Sciences, University of Hertfordshire, Hatfield, AL10 9AB, UK.

² Centre for the Digital Design of Drug Products, School of Chemical and Process Engineering, University of Leeds, Leeds, UK

³ Henry Moseley X-ray Imaging Facility, School of Materials, The University of Manchester, Manchester, M13 9PL, UK

⁴ Carl Zeiss Microscopy GmbH, Carl-Zeiss-Straße 22, 73447 Oberkochen, Germany

KEYWORDS: Specific surface area (SSA), inverse gas chromatography (iGC), nitrogen adsorption, lactose, X-Ray computed tomography (XCT)

INTRODUCTION

Measuring and monitoring the specific surface area (SSA) of powders during storage or after processing is a valuable metric that has been associated with changes in the performance of inhaled powder formulations, as it depends on both particle size distribution (PSD), the porosity and the surface roughness of the powders [1,2]. The value is also needed in surface energy experiments using inverse gas chromatography (iGC). Typically, gas-adsorption techniques use inert gases including Nitrogen (N₂) and Krypton to measure SSA; the adsorption isotherm is analysed and if it is of type II or IV, then the Brunauer-Emmett-Teller (BET) theory can be applied to extract the SSA_{BET} [3,4]. However, the widely used N₂ adsorption method is challenging for powders with low SSA, such as inhalation grade α -lactose monohydrate. More recently, iGC has been employed as an alternative method, using n-alkanes (heptane-C₇, octane-C₈) as probe molecules [5–7]. Advances in X-Ray Computed Tomography (XCT) have allowed the 3D imaging of inhalation grade lactose powders, and useful

metrics, such as PSD to be extracted [8,9]. The SSA_{BET} determined by N_2 adsorption and iGC-C₈ has been compared for inhalation grade lactose and XCT has been assessed as a complimentary technique.

METHODS

Inhalation grade α -lactose monohydrate, namely Lactohale 100 (sieved), Lactohale 200 (milled), Lactohale 206 (milled with fines removed) and Lactohale 20X (milled with fines removed) were kindly supplied by DFE Pharma (GmbH). PSD measurements were achieved by laser diffraction, as previously reported [8]. For the iGC SSA_{BET} measurements, the Surface Energy Analyzer (iGC-SEA, Surface Measurement Systems Ltd, UK) was used. Approximately 1.5 g of lactose was packed into silanised iGC glass columns (Analytical Columns, Croydon, UK). Prior to any measurements, the columns were conditioned using helium carrier gas at 10 sccm (standard cubic centimeters per minute) for 2 h at 30 °C and 0% RH. Methane gas was injected at the start and the end of the experiments for the dead volume calculation. SSA was calculated via the BET theory (SSA_{BET}), based on the C₈ adsorption isotherm data [10]. N_2 adsorption isotherms were obtained via a TriStar 3000 (Micromeritics Instrument Corp. USA). Around 500- 1400 mg of lactose sample were filled into 3/8" flat bottom cells with filler rods and conditioned under a helium purge at 25 °C for 8-12h. N_2 isotherms were measured at -196 °C. The BET analysis was based on the linear region of the nitrogen adsorption isotherm (from $p/p^\circ = 0.06-0.2$). Measurements were done in duplicate with the exception of LH20X which was measured just once.

LH100 was prepared for XCT scanning by spooning powder into a separate Kapton tube sample mount as described in [9]. A Zeiss Xradia Versa 520-DCT instrument (Carl Zeiss Microscopy) was used for this experiment. Scan settings and image

analysis methodology have been described elsewhere [8]. The SSA was calculated as the ratio of the total surface area within the analyzed sample to the mass of the analyzed sample, with the mass calculated as the product of the sample volume and density of 1.54 g/cm³.

RESULTS AND DISCUSSION

SSA depends on the surface roughness, porosity and the PSD of the particles, which is influenced by the inclusion of milling or classification steps in the manufacturing process. Table 1 shows the SSA results based on the techniques. By measuring, both iGC-C₈ and N₂ adsorption, it was possible to distinguish between the different grades of lactose (milled>milled with fines removed>sieved). Better agreement between the iGC-C₈ and the N₂ was observed for the milled lactose, (LH200), however, a significant difference was observed for the sieved lactose (LH100). This is due to the detection limit of the technique, as it is challenging to measure SSA_{BET} values less than 0.1 m²/g with N₂. LH100 SSA_{BET-N₂} was 0.064 m²/g with poor correlation in the measured partial pressure range (R² values were 0.984 and 0.959). Krypton adsorption could be employed but due to the experimental setup, there is the potential risk of change in structural form [6].

In order to assess SSA in an indirect way, further analysis of the XCT results was employed. To the best of the authors knowledge it is the first time that an indirect SSA evaluation based on a 3D imaging technique is presented. SSA_{XCT} for LH100 was 0.0854 m²/g, comparable with the other two techniques. However, due to the image resolution of the instrument used, surface roughness and porosity cannot be sufficiently estimated. This could potentially be addressed with the microstructural

information obtained from higher resolution XCT systems like nanoCT which can achieve voxel resolutions of 16 nm [11].

Table 1 – Characteristics, PSD and SSA values of inhalation grade lactose via N₂ adsorption, iGC and XCT. Errors correspond to duplicate measurements for the N₂ adsorption experiments and for triplicate measurements in one column for iGC.

Sample		LH100	LH200	LH206	LH20X
Grade		Sieved	Milled	Milled (fines removed)	Milled (fines removed)
LD PSD [μm] (2.0 bar)	D ₁₀	55	13	31	60
	D ₅₀	131	77	81	102
	D ₉₀	216	143	160	151
SSA [m ² /g]	N ₂	0.06 ± 0.02	0.25 ± 0.03	0.14 ± 0.03	0.13
	iGC-C ₈	0.18 ± 0.002	0.29 ± 0.003	0.22 ± 0.001	0.24 ± 0.001
	XCT	0.09	-	-	-

CONCLUSIONS

A comparison between the standard N₂-based adsorption and the C₈-based iGC for the measurement of the SSA of inhalation grade powders showed a good agreement between the two techniques for coarse lactose. For sieved lactose that exhibits an SSA value close to the detection limit of the instruments (0.1 m²/g), XCT was used as an alternative indirect measurement. The XCT value was in good agreement with the two other techniques suggesting that in the future, XCT imaging with built in porosity and roughness parameters will be able to provide SSA values.

ACKNOWLEDGEMENTS

This work is an output of the INFORM2020 Consortium (EPSRC grant EP/N025075/1). We thank Prof. Dr. Regina Scherließ for the N₂ adsorption data of LH100. We are grateful to consortium partner DFE Pharma for the supply of materials, and 3M,

AstraZeneca, GlaxoSmithKline, Malvern Panalytical and Zeiss Microscopy for their membership of the INFORM 2020 Consortium. Beamtime was provided by the Henry Moseley X-ray Imaging Facility (HMXIF), established through EPSRC Grant Nos. EP/F007906/1, EP/I02249X/1, and EP/F028431/1. HMXIF is one part of the Henry Royce Institute for Advanced Materials, established by EPSRC Grants EP/R00661X/1, EP/P025498/1, and EP/P025021/1.

REFERENCES

1. Watling CP, Elliott JA, Scruton C, Cameron RE: Surface modification of lactose inhalation blends by moisture. *Int J Pharm* 2010, 391: 29–37.
2. Depasquale R, Lee SL, Saluja B, Shur J, Price R: The Influence of Secondary Processing on the Structural Relaxation Dynamics of Fluticasone Propionate. *AAPS PharmSciTech* 2015, 16: 589–600.
3. ISO [International Organization for Standardization]: Determination of the specific surface area of solids by gas adsorption — BET method. 2010.
4. Brunauer S, Emmett PH, Teller E: Adsorption of Gases in Multimolecular Layers. *J Am Chem Soc* 1938, 60: 309–19.
5. Duralliu A, Matejtschuk P, Williams DR: Measuring the specific surface area (SSA) of freeze-dried biologics using inverse gas chromatography. *Eur J Pharm Biopharm Elsevier*; 2019, 142: 216–21.
6. Sharif S, Dimemmo LM, Thommes M, Hubert M, Sarsfield BA: A simplified approach to determine effective surface area and porosity of low bulk density active pharmaceutical ingredients in early development. *Adv Powder Technol The Society of Powder Technology Japan*; 2015, 26: 337–48.
7. Jaffari S, Forbes B, Collins E, Khoo J, Martin GP, Murnane D: Evidence for the existence of powder sub-populations in micronized materials: Aerodynamic size-fractions of aerosolized powders possess distinct physicochemical properties. *Pharm Res* 2014, 31: 3251–64.
8. Gajjar P, Styliari ID, Nguyen TTH, Carr J, Chen X, Elliott JA, et al.: 3D Characterisation of Dry Powder Inhaler Formulations: Developing X-ray Micro Computed Tomography Approaches. *Int J Pharm* 2020, In press.
9. Gajjar P, Styliari ID, Burnett TL: Multiscale Tomography : Probing The Nano- , Micro- , And Meso-Scale Resolution Of Inhalation Powder Structure Multiscale Tomography : Probing the Nano- , Micro- , and Meso-scale Resolution of Inhalation Powder Structure. *RDD Eur* 2019 2019, 1: 155–68.
10. Ramachandran V, Murnane D, Hammond RB, Pickering J, Roberts KJ, Soufian M, et al.: Formulation pre-screening of inhalation powders using computational atom-atom systematic search method. *Mol Pharm* 2015, 12: 18–33.

11. Withers PJ: X-ray nanotomography. *Mater Today* 2007, 10: 26–34.