



UNIVERSITY OF LEEDS

This is a repository copy of *Blood orange juice consumption increases flow-mediated dilation in adults with overweight and obesity: A randomized controlled trial*.

White Rose Research Online URL for this paper:
<https://eprints.whiterose.ac.uk/160668/>

Version: Accepted Version

Article:

Li, L, Lyall, GK orcid.org/0000-0002-5986-4845, Martinez-Blazquez, JA et al. (4 more authors) (2020) Blood orange juice consumption increases flow-mediated dilation in adults with overweight and obesity: A randomized controlled trial. *The Journal of Nutrition*. ISSN 0022-3166

<https://doi.org/10.1093/jn/nxaa158>

Copyright © The Author(s) on behalf of the American Society for Nutrition 2020. This is an author produced version of a paper published in *The Journal of Nutrition*. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Blood orange juice consumption increases flow-mediated dilation in adults with overweight and obesity: A randomized controlled trial

Lu Li^{1,2}, Gemma K. Lyall³, J. Alberto Martinez-Blazquez⁴, Fernando Vallejo⁴, Francisco Tomas-Barberan⁴, Karen M. Birch³, and Christine Boesch²

¹Beijing Advanced Innovation Center for Food Nutrition and Human Health, Beijing Technology and Business University, Beijing 100048, China; ²School of Food Science and Nutrition, University of Leeds, Leeds, United Kingdom; ³School of Biomedical Sciences, University of Leeds, Leeds, United Kingdom; and ⁴CEBAS-CSIC, Quality, Safety and Bioactivity of Plant Foods, Murcia, Spain.

Corresponding authors: C Boesch, School of Food Science and Nutrition, University of Leeds, Leeds, LS2 9JT, United Kingdom. Tel +44 113 3430268. Email: c.bosch@leeds.ac.uk and K Birch, School of Biomedical Sciences, University of Leeds, +44 113 3436713, k.m.birch@leeds.ac.uk

A list of all authors' last names: Li, Lyall, Martinez-Blazquez, Vallejo, Tomas-Barberan, Birch, and Boesch.

Word count: 3972

Number of figures: 3

Number of tables: 2

Supplementary data: none

Running title: Blood orange juice and endothelial function

Abbreviations: ALDH2, aldehyde dehydrogenase; AUC_{peak}, area under the shear rate curve to peak dilation; BOJ, blood orange juice; CD, control drink; cGMP, cyclic guanosine monophosphate; CHD, coronary heart disease; CVD, cardiovascular disease; DBP, diastolic

blood pressure; ET-1, endothelin 1; FMD, flow-mediated dilation; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; IL-6, interleukin 6; LDL, low-density lipoprotein; LPH, lactase phlorizin hydrolase; NO, nitric oxide; RCT, randomized controlled trial; RSD, relative standard deviation; SBP, systolic blood pressure; TNF- α , tumour necrosis factor- α ; UHPLC-QqQ-MS, ultra-high performance liquid chromatography coupled with triple-quadrupole mass spectrometry; VTI, velocity time integral.

Financial support: This work was supported by China Scholarship Council-University of Leeds Scholarship.

Author disclosures: LL, GKL, JAMB, FV, FTB, KMB, and CB, no conflicts of interest.

Clinical Trial Registry number NCT03611114, registered at clinicaltrials.gov (with information on the full protocol); Data described in the manuscript, code book and analytical code will be made available upon request pending (application and approval, payment, other).

1 **Abstract**

2 **Background:** Epidemiological studies have indicated an inverse association between citrus
3 fruit consumption and cardiovascular disease (CVD) risk. There is, however, a paucity of data
4 concerning effects of blood orange juice (BOJ) intake on endothelial function and
5 cardiovascular risk biomarkers.

6 **Objective:** We examined short-term effects of BOJ on endothelial function, blood pressure,
7 lipid profile, and inflammatory markers in healthy participants of European origin who were
8 overweight or obese.

9 **Methods:** In a randomized controlled single-blind crossover trial, 15 men and women (age:
10 28.7 ± 6.5 y; BMI: 28.3 ± 3.1 kg/m²) consumed BOJ or a sugar-matched control drink (CD)
11 (200 mL twice daily) for 2 weeks with a washout period of 1 week. Endothelial function,
12 measured as flow-mediated dilation (FMD) (primary outcome), and the following secondary
13 outcomes blood pressure, anthropometric measures, lipid profile, inflammatory markers,
14 markers of vasodilation and vasoconstriction, and urinary flavanone metabolites were
15 evaluated prior to and at the end of each treatment period following an overnight fast. Changes
16 between treatments over time were assessed using repeated-measures ANOVA.

17 **Results:** The results demonstrate a significant increase in FMD following BOJ consumption
18 (pre: $8.15\% \pm 2.92\%$, post: $10.2\% \pm 3.31\%$, $P=0.002$) compared to CD (pre: $8.11\% \pm 2.52\%$,
19 post: $7.77\% \pm 2.43\%$) (time by treatment interaction: $P=0.001$). Concurrent significant
20 increases in urinary hesperetin-3'-glucuronide and hesperetin-7-glucuronide were observed
21 following BOJ supplementation only (time by treatment interaction: $P \leq 0.01$). Baseline blood
22 pressure, lipid profile, hsCRP, and ET-1 were generally within healthy ranges and unaffected
23 by the intervention.

24 **Conclusions:** A two-week consumption of BOJ exerts favourable effects on endothelial
25 function in healthy women and men who were overweight or obese, which is likely mediated
26 by the combined actions of anthocyanin and flavanone metabolites on mechanisms that
27 contribute to enhancing NO bioavailability. This trial was registered at clinicaltrials.gov as
28 NCT03611114.

29

30 **Keywords:** flavanones, blood orange juice, overweight/obese participants, endothelial

31 function, flow-mediated dilation, shear rate, urinary flavanone metabolites

32

33 **Introduction**

34 Epidemiological studies have suggested that a higher intake of citrus fruit is associated with
35 reduced risk of ischemic stroke (1), lower levels of inflammation and endothelial dysfunction
36 (2). Bioactives in citrus fruits such as hesperidin and naringin have received considerable
37 attention due to *in vitro* and *in vivo* evidence demonstrating anti-atherogenic effects (3, 4).
38 Given the role of inflammation and endothelial dysfunction in the development of
39 atherosclerosis and that endothelial function is a strong prognostic indicator for cardiovascular
40 events (5, 6), the consumption of citrus fruit may have significant impact.

41 However, evidence from randomized controlled trials (RCTs) investigating effects of orange
42 juice on endothelial function and inflammatory markers is conflicting (7-9). A number of factors
43 have been highlighted which may contribute to observed heterogeneity including differences
44 in trial design, food composition, flavonoid source as well as volunteer related (such as age,
45 sex, ethnicity) (10, 11). Indeed, the majority of previous RCTs have involved men or
46 postmenopausal women. Issues associated with the inclusion of premenopausal women
47 mainly relate to fluctuations in either exogenous or endogenous reproductive hormones. The
48 increase in estrogen during the late follicular phase of the menstrual cycle has been seen to
49 markedly increase endothelial function, as measured by flow mediated dilation (FMD) (12, 13).
50 Regulated by an upregulation in nitric oxide (NO), FMD is a direct result of blood flow
51 enhanced shear stress along the endothelium. Thus shear stress as the stimulus for FMD is
52 crucial to the interpretation of FMD data (14) but has rarely been calculated and discussed in
53 the context of polyphenol intervention studies.

54 Genetic variants have been highlighted as important contributors to inter-individual differences
55 (15). Lactase phlorizin hydrolase (LPH), for example, plays a pivotal role in the exclusive
56 hydrolysis of some polyphenol glucosides prior to absorption (16) with low levels occurring in
57 5% of European and 90% of African and Asian adults. Variations in the enzyme aldehyde
58 dehydrogenase (ALDH2), which contributes to nitrate conversion into NO (17), are highly
59 prevalent among Asian populations (18) but relatively uncommon in Caucasians (19).

60 However, details on participant ethnicity are frequently not provided in human intervention
61 studies.

62 In comparison to blond orange juice, blood orange juice (BOJ) has received much less
63 attention yet it is an abundant source for bioactives such as anthocyanins, flavanones,
64 hydroxycinnamic acids (20), and vitamin C (21). With the exception of Buscemi et al. (22) who
65 showed an increase in FMD in men with moderately increased CVD risk after one-week BOJ
66 consumption, none of the other studies were able to demonstrate beneficial changes in CVD
67 biomarkers after chronic exposure of up to 12 weeks (23, 24).

68 The aim of this study was to investigate the effects of short term consumption of BOJ on
69 endothelial function and other CVD risk factors. We chose participants who were overweight
70 or obese and of Caucasian heritage as higher BMI and chronic low-grade inflammation are
71 associated with a higher risk of coronary heart disease (CHD) and impaired endothelial
72 function (25-27).

73

74 **Methods**

75 **Study population**

76 Sixteen healthy men and premenopausal women, aged 20-45 y, were recruited according to
77 the following eligibility criteria: Caucasians (of European origin), generally healthy with
78 absence of any form of CVD, non-smokers, BMI over 25 kg/m², no medications or dietary
79 supplements (vitamins, antioxidants), absence of lactose intolerance. Fifteen participants
80 completed the study as one participant was excluded due to the use of medication during the
81 intervention. The study was approved by Biological Sciences Faculty Research Ethics
82 Committee, University of Leeds (Ethics Reference Number: BIOSCI 15-030), in accordance
83 with ethical principles of the Declaration of Helsinki. Written informed consent from all
84 participants was obtained prior to study commencement.

85

86 **Study design**

87 The study design was a randomized, controlled, single-blind, crossover trial (**Figure 1**). During
88 two 2-week periods, participants were asked to consume sugar-matched 400 mL of BOJ or
89 control drink (CD) (200 mL with breakfast and 200 mL with dinner) daily, with a 1-week
90 washout period between each intervention. Block randomization was conducted to allocate
91 drink sequences to participant codes. After enrolment onto the study, participants visited the
92 vascular laboratory at the University of Leeds on four separate occasions for measurements
93 prior to and following each 2-week period (January-June 2017). Participants were instructed
94 to stay fasted and refrain from exercise for 12 h before measurements in the morning. After
95 the participant was supine and comfortable for 15 min to reach a cardiovascular steady state,
96 blood pressure was measured in triplicate with 2-minute intervals. Endothelial function was
97 evaluated via brachial artery FMD with each measurement performed at the same time of the
98 day and on the same area of the brachial artery, as explained below. Following FMD
99 measurements, venous blood samples from the antecubital vein were collected and
100 participants were asked to provide a spot urine sample. Anthropometric measures were
101 conducted at baseline and following each intervention. Participants were asked to maintain
102 their lifestyle as usual throughout the study, including dietary routines and physical activity
103 level and they were asked to record the time of individual drink consumption on a separate
104 sheet which was returned to the researcher after the 2-week periods. Questionnaires on the
105 habitual intake of citrus fruit/juice and other flavonoid sources were also collected.
106 Researchers who conducted the measurements were blinded and only unblinded post data
107 analysis.

108 Female participants started the intervention on specific days of the menstrual cycle (e.g., day
109 4 of a 28-day menstrual cycle) in order to avoid any measurements during the late follicular
110 phase. For women consuming the oral contraceptive pill, either as combined estrogen and
111 progesterone or progesterone only, all assessments were undertaken during the period of
112 time the pill was being consumed (not in pill breaks).

113

114 **Intervention products**

115 Commercially available blood orange juice (BOJ) (47 kcal/dL) and a low flavonoid control drink
116 (CD) (41 kcal/dL) were obtained from Waitrose, Leeds, UK. The flavanone concentration of
117 the juices was analyzed by HPLC-MS (28). Hesperidin and narirutin were 80.2 ± 2.7 and 9.5
118 ± 0.1 mg/dL for BOJ, and 6.3 ± 0.2 and 1.0 ± 0.1 mg/dL for CD, respectively (both $P < 0.001$).
119 The total anthocyanin concentration, analyzed using pH differential method (29), was $2.40 \pm$
120 0.13 mg/dL for BOJ with no anthocyanins being detectable in CD. Sugar quantification was
121 performed by Dionex ICS-5000 (30) and the total sugar concentration of BOJ and CD
122 calculated at 14.3 ± 0.6 and 14.7 ± 0.7 g/dL, respectively ($P = 0.591$). Test drinks for 2 weeks
123 of consumption were provided to each participant at the start of the intervention and stored in
124 their home refrigerators until consumption.

125

126 **Endothelial function**

127 The local protocol for assessment of brachial artery endothelial function via FMD (31, 32) was
128 in accordance with established guidelines (14, 33) using duplex ultrasonography (Vivid E9
129 with XDclear, GE Healthcare, US). Following 15 min of rest in a supine position in a quiet and
130 temperature-controlled vascular laboratory, the brachial artery was imaged above the
131 antecubital fossa in the longitudinal plane. Resting brachial artery diameter was recorded for
132 20 s at 15 images/s using vascular imaging software (Vascular Imager, Medical Imaging
133 Applications, Iowa, USA). Reactive hyperemia was created by inflating a pneumatic cuff on
134 the forearm for 5 min at 220 mmHg. Post cuff deflation brachial artery diameter and blood flow
135 were recorded for 180 s starting 30 s before cuff deflation. Brachial artery diameter and blood
136 flow were assessed off-line using Brachial Analyzer for Research (version 6, Medical Imaging
137 Applications, Iowa, USA). Peak diameter was calculated from the maximum diameter of the
138 moving averages of 3 consecutive diameters. Absolute FMD (mm; Peak diameter – resting
139 diameter) and relative FMD (%; absolute FMD / resting diameter x 100) were determined.
140 Velocity time integral (VTI) and the area under the shear rate curve (AUC) were calculated as
141 previously described (34). AUC to peak diameter was calculated as an indication of the
142 stimulus for FMD (14). Time to peak diameter was calculated as the time period starting from

143 cuff deflation to peak diameter. Specifically, relative FMD was the primary outcome and the
144 other measures of FMD (resting diameter, AUCpeak, absolute FMD, scaled FMD index, time
145 to peak diameter) were secondary outcomes. To determine reliability of ultrasound
146 measurements, FMD was conducted on 10 healthy participants with each individual examined
147 twice on two consecutive days. Resting brachial artery diameter showed a coefficient of
148 variation (CV) of 0.4% and CV of relative FMD was 6.12%.

149

150 **Plasma cardiovascular risk biomarkers**

151 Serum and EDTA plasma were generated using standard procedures. Urine samples were
152 centrifuged (2000 x g at 4°C, 15 min) and filtered (0.22 µm CA-CN syringe filter). Aliquots of
153 serum, plasma and urine were stored at -80°C prior to analysis. Lipid profile, hsCRP and
154 estradiol were analysed in serum using standardized assays by the Pathology Services (Leeds
155 General Infirmary). Plasma ET-1 and cGMP were measured using commercially available
156 immunoassays (R&D Systems, Abingdon, UK).

157

158 **Urinary metabolite analysis**

159 Major urinary metabolites were analyzed by UHPLC-QqQ-MS as previously described (35).
160 The limit of detection (LOD) was determined as the concentration of analytes with a signal to-
161 noise ratio of at least 3, and the limit of quantification (LOQ) was the lowest standard with a
162 signal-to-noise ratio of at least 10. LOQs were 80 nM for hesperetin, 50 and 80 nM for
163 hesperetin-7-and 3'-glucuronides, respectively, and 40 and 50 nM for hesperetin-7- and 3'-
164 sulfates respectively. LODs were 30 nM for hesperetin, 20 and 30 nM for hesperetin-7- and
165 3'-glucuronides respectively, and 15 and 20 nM for hesperetin-7- and 3'-sulfates respectively.
166 The intra-day repeatability of the UHPLC-QqQ-MS method was assessed from 10 consecutive
167 chromatographic runs using a standard solution with 2.5 µM of every standard in MeOH:0.1%
168 (v/v) formic acid. The inter-day repeatability of the method was assessed by analyzing the
169 same standard solution on 2 consecutive days. The relative standard deviation (RSD) for peak

170 area was in the range of 0.5–4.7% in the intra-day test and 1.3–3.5% in the case of the inter-
171 day test.

172

173 **Sample size and statistical analysis**

174 To detect a 2.0 unit increase in relative FMD (the primary outcome of the present study),
175 assuming a standard deviation of 2.0 (based on FMD reliability data), with 80% power and at
176 the 5% significance level, a total sample size of 10 participants was required to complete a
177 two-treatment crossover study. Data are presented as means \pm SDs. Statistical analyses were
178 conducted by using Statistical Package for the Social Sciences (SPSS, version 24, IBM
179 Corporation, USA). Data were tested for normality by using Shapiro-Wilk test with normality
180 defined as $P > 0.05$. Differences in study outcomes between treatments were analyzed using
181 a two-factor repeated-measures ANOVA, with treatment and time (baseline and week 2 within
182 each treatment period) as within-subject factors. The main effects of treatment and time as
183 well as the time by treatment interaction were investigated. When significant time, treatment,
184 and/or time by treatment effects were identified, *post hoc* comparisons were carried out by
185 using Bonferroni correction for multiple comparisons. The effect of sex on relative FMD was
186 investigated by using a mixed ANOVA, with treatment and time as within-subject factors and
187 sex as the between-subjects factor, including a time by treatment by sex interaction.
188 Differences in relative FMD adjusted for BMI were analyzed by using a repeated-measures
189 ANOVA, with treatment and time as within-subject factors and BMI as the covariate, including
190 a time by treatment by BMI interaction. Carryover effects on relative FMD were assessed by
191 using a mixed ANOVA, with treatment and time as within-subject factors and treatment
192 sequence as the between-subjects factor. Missing data were not imputed and a complete case
193 analysis was performed. The flavanone concentration in the different drinks was analyzed
194 using an Independent-Samples T-Test. Significance was defined at $P < 0.05$. Correlation
195 analyses were conducted by using Pearson's correlation coefficient.

196

197 **Results**

198 Clinical characteristics of participants at screening are shown in **Table 1**. Among 16 enrolled
199 participants, 15 participants (10 female participants and 5 male participants) completed all
200 arms of the intervention. All 15 participants reported no major changes in diet and lifestyle
201 during the intervention, which was confirmed by unchanged body weight (data not shown).
202 The habitual intake of citrus fruit/juice was generally low (both less than one portion per week)
203 and the total intake of tea and coffee were 2.4 ± 1.1 cups/d (1 cup: 200-250 mL). No carryover
204 effects were observed (P -interaction=0.20). Resting brachial artery diameter did not differ
205 between treatments prior to the intervention (P =1.00; **Table 2**), however a time by treatment
206 interaction (P =0.017) was observed. Compared with baseline, relative FMD markedly
207 increased only following the 2-week consumption of BOJ (**Figure 2A**: time by treatment
208 interaction: P =0.001). Area under the shear rate curve to peak dilation (AUC_{peak}) (**Figure 2B**:
209 time by treatment interaction: P =0.56) and time to peak diameter did not change over time in
210 either treatment (Table 2). To remove the influence of changes in resting artery diameter on
211 FMD, FMD was scaled to resting artery diameter according to Atkinson (36). Analysis of the
212 scaled FMD index also revealed an increase following the 2-week consumption of BOJ only,
213 compared with baseline (P =0.001).

214 Plasma concentrations of estradiol in female participants did not differ during the trial periods
215 (0.33 ± 0.27 and 0.28 ± 0.25 nmol/L for pre and post BOJ consumption, respectively; $0.27 \pm$
216 0.16 and 0.29 ± 0.21 nmol/L for pre and post CD consumption, respectively; time by treatment
217 interaction: P =0.80). Moreover, the effect of the drinks on relative FMD did not differ with sex
218 (the effect of sex: P =0.28; P -interaction=0.70). Although the effect of BMI on relative FMD was
219 not significant (the effect of BMI: P =0.61; P -interaction=0.56), there was a moderate inverse
220 correlation between the BMI of participants and changes in relative FMD following 2-week
221 consumption of BOJ (R =-0.42, P =0.12). Similarly, a moderate inverse correlation between the
222 BMI of participants and changes in relative FMD was also observed following 2-week
223 consumption of CD (R =-0.45, P =0.09).

224 All participants complied with dietary restrictions, confirmed by low (or not detectable) urinary
225 concentrations of hesperetin-3'-glucuronide and hesperetin-7-glucuronide at baseline ($P=0.42$
226 and $P=0.39$, respectively; **Figure 3**). Urinary hesperetin-3'-glucuronide and hesperetin-7-
227 glucuronide both increased following BOJ consumption from 0.17 ± 0.04 to 9.78 ± 2.52 μM
228 ($P=0.007$) and from 0.06 ± 0.02 to 2.71 ± 0.70 μM ($P=0.009$), respectively, but not following the
229 CD (hesperetin-3'-glucuronide from 0.30 ± 0.08 to 0.59 ± 0.15 μM , and hesperetin-7-glucuronide
230 from 0.11 ± 0.03 to 0.16 ± 0.04 μM). Both urinary hesperetin-3'-glucuronide ($R=0.35$, $P=0.007$)
231 and hesperetin-7-glucuronide ($R=0.32$, $P=0.012$) were significantly correlated with relative
232 FMD.

233 Blood pressure (systolic and diastolic), lipids (total cholesterol, triglycerides, and HDL-
234 cholesterol), hsCRP, and endothelin-1 were within healthy ranges (37-40) and not affected by
235 the interventions, except for cGMP with a significant interaction and LDL-cholesterol with a
236 significant treatment effect (Table 2). No significant correlations were observed between these
237 outcomes and relative FMD (data not shown).

238

239 **Discussion**

240 This study demonstrates a significant 2.01% increase in FMD following consumption of
241 anthocyanin-rich blood orange juice as compared to a low flavonoid control drink, with a
242 concurrent significant increase in urinary flavanone metabolites. Importantly, whilst the clinical
243 use of FMD in the calculation of CVD risk has not been recognized, a 1% increase in FMD in
244 large trials has been associated with a range of 8-13% reduction in CVD risk (6, 41).
245 Furthermore, FMD is most successful to monitor effects of interventions, as seen in the current
246 study. The increase in FMD following BOJ intake can be considered as relatively large, given
247 that the chronic effect of flavonoids on FMD is only 0.73 % (ranging from 0.17 to 1.30) as
248 demonstrated by a pooled analysis of flavonoid intervention trials (42). The effects of BOJ on
249 FMD in the present study are comparable to cocoa powder (800 mg cocoa flavonoids/d for 1
250 week) consumed by healthy individuals (43). To our knowledge, the crossover study by
251 Buscemi et al. (22) is the only other currently published study on BOJ consumption (500 mL/d)

252 that can demonstrate a significant increase in endothelial function via FMD alongside
253 decreases in the inflammatory markers CRP, IL-6 and TNF- α after 1 week supplementation in
254 participants with augmented CVD risk. The few other studies on BOJ supplementation were
255 not able to show effects on biomarkers of CVD risk in participants who were overweight
256 (500 mL/d over 28 days) (23) or only a moderate reduction in LDL cholesterol levels in
257 participants with obesity after a 12-week supplementation with 500mL BOJ/d (24). Our current
258 study, using a relatively high volume of 400 mL BOJ per day, did not demonstrate changes in
259 blood pressure, lipid profiles and markers of inflammation, likely due to all values being within
260 a healthy range. Missing data on lipid profiles and markers of inflammation might have limited
261 impact on the interpretation of the results, since the participants were relatively young and
262 healthy, and hence it is likely those outcomes were within a healthy range at baseline and did
263 not change by the intervention. It cannot be excluded that spontaneous changes in the diet
264 were made by the participants to compensate for the energy load in the drinks, as body weight
265 remained unchanged during the study period. Since their habitual diet was low in citrus
266 fruit/juice and not high in tea/coffee, and participants were asked to maintain their diet during
267 the study, the effects of compensatory changes in flavonoid rich foods/drinks were considered
268 to be negligible. This was confirmed by unchanged baseline values of urinary metabolites and
269 endothelial function.

270 To our knowledge, this is the first study to report an impact of polyphenol-rich foods or drink
271 consumption upon resting artery diameter. This indicates a small amount of remodeling of the
272 brachial artery, which may be due to elevated levels of NO following the consumption of BOJ,
273 evidenced by a 29% increase in plasma cGMP. Together with unchanged shear rate stimulus,
274 it suggests a functional improvement in endothelial function probably through enhanced NO
275 bioavailability due to anthocyanin and flavanone metabolites following BOJ consumption and
276 hence for the same shear stimulus the reactivity of the vessel is greater. Indeed, a positive
277 correlation was demonstrated between plasma cGMP and FMD response following a 12-week
278 supplementation with anthocyanins in hypercholesterolemic individuals (44) with cGMP being
279 considered as an indicator for plasma NO levels (45).

280 Large variations in study outcomes have been observed in previous flavonoid
281 supplementation trials, in particular when healthy participants were recruited, making overall
282 interpretation of supplementation effectivity difficult. Our approach was aimed at minimizing
283 potential confounding effects of the female hormone estrogen by scheduling FMD
284 measurements to avoid the late follicular phase. Thereby the reported improvements in
285 endothelial function observed in the present study are likely due to the intervention and not
286 fluctuations in hormone levels. Hence, the present study demonstrates that premenopausal
287 women can be suitable participants for the evaluation of endothelial function under defined
288 experimental conditions.

289 A further factor which may contribute to the conflicting evidence in the literature is shear rate
290 which, if not carefully controlled, might give rise to variations in the resultant FMD values, and
291 be mistaken as a 'functional change' after an intervention. Present improvement in endothelial
292 function following BOJ intake was induced under unchanged shear rate conditions
293 demonstrating a strong and significant correlation between shear rate and FMD. In support of
294 these findings, a significant correlation has only been observed in younger (27 ± 6 y), but not
295 in older adults (58 ± 4 y) (46), which may indicate the loss of endothelial functionality during
296 aging. Likewise, time from cuff deflation to peak diameter was not affected in the present study
297 but is positively associated with increasing age (47). Given that the present participants were
298 young and healthy, the time to reach peak dilation following cuff deflation was relatively quick
299 but consistent with previous research in participants of a similar age (47). Another novel finding
300 of the current study is the differential responses of endothelial function to flavonoid-rich food
301 consumption, depending on the BMI of participants, which to our knowledge, have not been
302 reported in previous studies. In support of this, BMI has been highlighted as a factor impacting
303 the responsiveness of individuals in intervention trials. Azzini et al. (24) reported a
304 lacking/abnormal response of total and LDL cholesterol in participants with obesity compared
305 to lower BMI female participants when given BOJ supplementation.

306 We demonstrate here that the increase in FMD, following a 2-week daily consumption of BOJ,
307 is concurrent with urinary excretion of citrus flavanone metabolites hesperetin-3'- and
308 hesperetin-7-glucuronides. Our results therefore provide compelling evidence that the *in vivo*
309 FMD response is indeed linked to the presence of citrus flavonoids and/or their circulating
310 metabolites. The availability of flavanones from orange juice (as a sum of small intestine and
311 gut microbiota derived compounds) is, despite high inter-individual variation, considered high
312 (48-50). Nevertheless, many orange juice studies are not demonstrating modulation of CVD
313 risk biomarkers or endothelial function. Schär et al. (8) has recently shown that citrus
314 flavonoids from juice in comparison to a hesperidin supplement are much more available to
315 humans. However, in their acute crossover RCT neither orange juice nor hesperidin
316 supplement were able to affect any of the outcome markers such as RH-PAT (reactive
317 hyperemia-peripheral arterial tonometry), or CVD risk biomarkers. Although acute effects were
318 not investigated in the present study, flavanone-rich citrus beverages have been reported to
319 be effective at counteracting the negative impact of a double meal rich in fat on postprandial
320 endothelial function measured by FMD at 7 h post intake (7). In comparison, the
321 measurements in the present study were conducted approximately 12 hours following the final
322 drink, indicating a prolonged effect of the bioactive compounds in BOJ.

323 Anthocyanins, as present in BOJ, are rapidly but poorly absorbed in the small intestine (51);
324 and as a consequence, we were not able to reliably detect anthocyanin metabolites in the
325 urine samples of participants. However, the availability and molecular effects of anthocyanins
326 towards CVD biomarkers have been documented in a number of studies. Indeed, Speciale et
327 al. (52) suggested that anthocyanins prevent stress-induced endothelial dysfunction.
328 Consumption of BOJ for 3 weeks significantly increased plasma antioxidant concentrations
329 (21) and the intake of blackcurrant juice, an abundant source of anthocyanins and other
330 bioactives, for 6 weeks resulted in a significant increase in FMD in healthy adults (53). It has
331 been suggested that the beneficial effects of blood orange may be mediated by the synergistic
332 effects of its different compounds (54).

333 There are several limitations to the present study. First, distinct differences in the colour and
334 taste of the two drinks made double-blinding impossible. However, the researchers who
335 conducted the analyses of the biological samples and FMD data were blinded to which juice
336 the participant was consuming. It is unlikely that the outcomes of the present study (such as
337 endothelial function, lipid profile, hsCRP) were influenced by participants knowing which juice
338 they were consuming. Second, although vitamin C concentration was not matched in the
339 control drink, it is unlikely the observed enhancement in FMD was due to vitamin C presence.
340 Clinical data suggest that doses of vitamin C up to 500 mg do not alter endothelial function,
341 both acutely and chronically (55); and vitamin C concentration in the BOJ ingested in the
342 present study was only 168 mg per day. In addition, given the short half-life of vitamin C
343 (approximately 30 min), it seems unlikely that vitamin C exerted any effect on the markers
344 determined after a 12 h overnight fast. Sex difference analysis was of course hampered by
345 the small sample size. Although Bonferroni corrections were used, there is always a chance
346 of making a Type I error in any study testing multiple secondary outcomes.

347 In conclusion, we observed favourable changes in resting arterial tone and endothelial function
348 in healthy Caucasian men and premenopausal women with overweight or obesity following
349 the consumption of blood orange juice. Further studies are required to better understand the
350 role and potential interactions of individual flavonoids and their metabolites in BOJ and their
351 contribution to reducing CVD risk. The differential effects on FMD according to BMI warrant
352 further confirmation in larger cohorts. In addition, future RCTs on specific participant groups
353 (based on age, sex, ethnicity/genotype, BMI, CVD risk) are needed to investigate effects of
354 polyphenol-rich products following long term consumption.

355

356 **Acknowledgements**

357 We thank Saïd Ibeggazene for his support with FMD practice and Fraser Chadwick for his
358 help with venepuncture.

359 The authors' contributions were as follows: LL, KMB, CB designed the study; LL conducted
360 the human study and statistical analyses; GKL trained LL in FMD and provided venepuncture

361 support; GKL and KMB scaled FMD data and conducted statistical analyses on scaled FMD
362 index; JAMB, FV, FTB analyzed urinary flavanone metabolites; LL drafted the manuscript
363 which was finally edited by GKL, CB and KMB. All authors have read and approved the final
364 manuscript.

References

1. Cassidy A, Bertola M, Chiuve S, Flint A, Forman J, Rimm EB. Habitual intake of anthocyanins and flavanones and risk of cardiovascular disease in men. *Am J Clin Nutr* 2016;104:587-94.
2. Landberg R, Sun Q, Rimm EB, Cassidy A, Scalbert A, Mantzoros CS, Hu FB, van Dam RM. Selected Dietary Flavonoids Are Associated with Markers of Inflammation and Endothelial Dysfunction in U.S. Women. *J Nutr* 2011;141:618-25.
3. Barreca D, Gattuso G, Bellocco E, Calderaro A, Trombetta D, Smeriglio A, Laganà G, Daglia M, Meneghini S, Nabavi SM. Flavanones: citrus phytochemical with health-promoting properties. *BioFactors* 2017;43:495-506.
4. Testai L, Calderone V. Nutraceutical value of citrus flavanones and their implications in cardiovascular disease. *Nutrients* 2017;9:502.
5. Vanhoutte PM, Shimokawa H, Tang EH, Feletou M. Endothelial dysfunction and vascular disease. *Acta Physiol* 2009;196:193-222.
6. Inaba Y, Chen JA, Bergmann SR. Prediction of future cardiovascular outcomes by flow-mediated vasodilatation of brachial artery: a meta-analysis. *Int J Cardiovasc Imaging* 2010;26:631-40.
7. Rendeiro C, Dong H, Saunders C, Harkness L, Blaze M, Hou Y, Belanger RL, Corona G, Lovegrove JA, Spencer JP. Flavanone-rich citrus beverages counteract the transient decline in postprandial endothelial function in humans: a randomised, controlled, double-masked, cross-over intervention study. *Br J Nutr* 2016;116:1999-2010.
8. Schär MY, Curtis PJ, Hazim S, Ostertag LM, Kay CD, Potter JF, Cassidy A. Orange juice-derived flavanone and phenolic metabolites do not acutely affect cardiovascular risk biomarkers: a randomized, placebo-controlled, crossover trial in men at moderate risk of cardiovascular disease. *Am J Clin Nutr* 2015;101:931-8.

9. Morand C, Dubray C, Milenkovic D, Lioger D, Martin JF, Scalbert A, Mazur A. Hesperidin contributes to the vascular protective effects of orange juice: a randomized crossover study in healthy volunteers. *Am J Clin Nutr* 2011;93:73-80.
10. Rees A, Dodd G, Spencer J. The Effects of Flavonoids on Cardiovascular Health: A Review of Human Intervention Trials and Implications for Cerebrovascular Function. *Nutrients* 2018;10:1852.
11. Cassidy A, Minihane A-M. The role of metabolism (and the microbiome) in defining the clinical efficacy of dietary flavonoids. *Am J Clin Nutr* 2016;105:10-22.
12. Adkisson EJ, Casey DP, Beck DT, Gurovich AN, Martin JS, Braith RW. Central, peripheral and resistance arterial reactivity: fluctuates during the phases of the menstrual cycle. *Exp Biol Med* 2010;235:111-8.
13. Williams MR, Westerman RA, Kingwell BA, Paige J, Blombery PA, Sudhir K, Komesaroff PA. Variations in endothelial function and arterial compliance during the menstrual cycle. *J Clin Endocrinol Metab* 2001;86:5389-95.
14. Thijssen DH, Black MA, Pyke KE, Padilla J, Atkinson G, Harris RA, Parker B, Widlansky ME, Tschakovsky ME, Green DJ. Assessment of flow-mediated dilation in humans: a methodological and physiological guideline. *Am J Physiol Heart Circ Physiol* 2011;300:H2-H12.
15. Manach C, Milenkovic D, Wiele T, Rodriguez-Mateos A, Roos B, Garcia-Conesa MT, Landberg R, Gibney ER, Heinonen M, Tomás-Barberán F. Addressing the inter-individual variation in response to consumption of plant food bioactives: Towards a better understanding of their role in healthy aging and cardiometabolic risk reduction. *Mol Nutr Food Res* 2017;61:1600557.
16. Day AJ, Cañada FJ, Díaz JC, Kroon PA, Mclauchlan R, Faulds CB, Plumb GW, Morgan MR, Williamson G. Dietary flavonoid and isoflavone glycosides are hydrolysed by the lactase site of lactase phlorizin hydrolase. *FEBS Lett* 2000;468:166-70.
17. Lundberg JO, Weitzberg E, Gladwin MT. The nitrate–nitrite–nitric oxide pathway in physiology and therapeutics. *Nat Rev Drug Discov* 2008;7:156-67.

18. Chang JS, Hsiao J-R, Chen C-H. ALDH2 polymorphism and alcohol-related cancers in Asians: a public health perspective. *J Biomed Sci* 2017;24:19.
19. Brennan P, Lewis S, Hashibe M, Bell DA, Boffetta P, Bouchardy C, Caporaso N, Chen C, Coutelle C, Diehl SR. Pooled analysis of alcohol dehydrogenase genotypes and head and neck cancer: a HuGE review. *Am J Epidemiol* 2004;159:1-16.
20. Rapisarda P, Carollo G, Fallico B, Tomaselli F, Maccarone E. Hydroxycinnamic acids as markers of Italian blood orange juices. *J Agric Food Chem* 1998;46:464-70.
21. Riso P, Visioli F, Gardana C, Grande S, Brusamolino A, Galvano F, Galvano G, Porrini M. Effects of Blood Orange Juice Intake on Antioxidant Bioavailability and on Different Markers Related to Oxidative Stress. *J Agric Food Chem* 2005;53:941-7.
22. Buscemi S, Rosafio G, Arcoleo G, Mattina A, Canino B, Montana M, Verga S, Rini G. Effects of red orange juice intake on endothelial function and inflammatory markers in adult subjects with increased cardiovascular risk. *Am J Clin Nutr* 2012;95:1089-95.
23. Hollands WJ, Armah CN, Doleman JF, Perez-Moral N, Winterbone MS, Kroon PA. 4-Week consumption of anthocyanin-rich blood orange juice does not affect LDL-cholesterol or other biomarkers of CVD risk and glycaemia compared with standard orange juice: a randomised controlled trial. *Br J Nutr* 2018;119:415-21.
24. Azzini E, Venneria E, Ciarapica D, Foddai M, Intorre F, Zaccaria M, Maiani F, Palomba L, Barnaba L, Tubili C. Effect of Red Orange Juice Consumption on Body Composition and Nutritional Status in Overweight/Obese Female: A Pilot Study. *Oxid Med Cell Longev* 2017;2017.
25. Willett WC, Manson JE, Stampfer MJ, Colditz GA, Rosner B, Speizer FE, Hennekens CH. Weight, weight change, and coronary heart disease in women: risk within the 'normal' weight range. *JAMA* 1995;273:461-5.
26. Woo KS, Chook P, Chung WY, Sung RY, Qiao M, Leung SS, Lam CW, Metreweli C, Celermajer DS. Effects of diet and exercise on obesity-related vascular dysfunction in children. *Circulation* 2004;109:1981-6.

27. Perticone F, Ceravolo R, Candigliota M, Ventura G, Iacopino S, Sinopoli F, Mattioli PL. Obesity and body fat distribution induce endothelial dysfunction by oxidative stress. *Diabetes* 2001;50:159-65.
28. Sweidan AMA. Bioavailability of citrus flavanones and their effect on cardiovascular health biomarkers. University of Leeds, 2015.
29. Lee J, Durst RW, Wrolstad RE. Determination of total monomeric anthocyanin pigment content of fruit juices, beverages, natural colorants, and wines by the pH differential method: collaborative study. *J AOAC Int* 2005;88:1269-78.
30. Øbro J, Harholt J, Scheller HV, Orfila C. Rhamnogalacturonan I in *Solanum tuberosum* tubers contains complex arabinogalactan structures. *Phytochemistry* 2004;65:1429-38.
31. Harris E, Rakobowchuk M, Birch KM. Interval exercise increases angiogenic cell function in postmenopausal women. *BMJ Open Sport Exerc Med* 2017;3:e000248.
32. Harris E, Rakobowchuk M, Birch KM. Sprint interval and sprint continuous training increases circulating CD34+ cells and cardio-respiratory fitness in young healthy women. *PloS One* 2014;9:e108720.
33. Harris RA, Nishiyama SK, Wray DW, Richardson RS. Ultrasound assessment of flow-mediated dilation. *Hypertension* 2010;55:1075-85.
34. Rakobowchuk M, Harris E, Taylor A, Baliga V, Cubbon RM, Rossiter HB, Birch KM. Heavy and moderate interval exercise training alters low-flow-mediated constriction but does not increase circulating progenitor cells in healthy humans. *Exp Physiol* 2012;97:375-85.
35. Rangel-Huerta OD, Aguilera CM, Martin MV, Soto MJ, Rico MC, Vallejo F, Tomas-Barberan F, Perez-de-la-Cruz AJ, Gil A, Mesa MD. Normal or High Polyphenol Concentration in Orange Juice Affects Antioxidant Activity, Blood Pressure, and Body Weight in Obese or Overweight Adults. *J Nutr* 2015;145:1808-16.
36. Atkinson G. Shear rate normalization is not essential for removing the dependency of flow-mediated dilation on baseline artery diameter: past research revisited. *Physiol Meas* 2014;35(9):1825.

37. Williams B, Mancia G, Spiering W, Agabiti Rosei E, Azizi M, Burnier M, Clement DL, Coca A, De Simone G, Dominiczak A. 2018 ESC/ESH Guidelines for the management of arterial hypertension: The Task Force for the management of arterial hypertension of the European Society of Cardiology (ESC) and the European Society of Hypertension (ESH). *Eur Heart J* 2018;39:3021-104.
38. Program NCE. Expert Panel on Detection, Evaluation, and treatment of high blood cholesterol in Adults (Adult treatment Panel III). Third report of the National cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and treatment of high blood cholesterol in Adults (Adult treatment Panel III) final report. *Circulation* 2002;106:3143-421.
39. Pearson TA, Mensah GA, Alexander RW, Anderson JL, Cannon III RO, Criqui M, Fadl YY, Fortmann SP, Hong Y, Myers GL. Markers of inflammation and cardiovascular disease: application to clinical and public health practice: a statement for healthcare professionals from the Centers for Disease Control and Prevention and the American Heart Association. *Circulation* 2003;107:499-511.
40. Abdel-Sayed S, Nussberger J, Aubert J-F, Gohlke P, Brunner HR, Brakch N. Measurement of plasma endothelin-1 in experimental hypertension and in healthy subjects*. *Am J Hypertens* 2003;16:515-21.
41. Ras RT, Streppel MT, Draijer R, Zock PL. Flow-mediated dilation and cardiovascular risk prediction: a systematic review with meta-analysis. *Int J Cardiol* 2013;168:344-51.
42. Kay CD, Hooper L, Kroon PA, Rimm EB, Cassidy A. Relative impact of flavonoid composition, dose and structure on vascular function: A systematic review of randomised controlled trials of flavonoid-rich food products. *Mol Nutr Food Res* 2012;56:1605-16.
43. Grassi D, Desideri G, Necozone S, Di Giosia P, Barnabei R, Allegaert L, Bernaert H, Ferri C. Cocoa consumption dose-dependently improves flow-mediated dilation and arterial stiffness decreasing blood pressure in healthy individuals. *J Hypertens* 2015;33:294-303.

44. Zhu Y, Xia M, Yang Y, Liu F, Li Z, Hao Y, Mi M, Jin T, Ling W. Purified anthocyanin supplementation improves endothelial function via NO-cGMP activation in hypercholesterolemic individuals. *Clin Chem* 2011;57:1524-33.
45. Minamino T, Kitakaze M, Sato H, Asanuma H, Funaya H, Koretsune Y, Hori M. Plasma levels of nitrite/nitrate and platelet cGMP levels are decreased in patients with atrial fibrillation. *Arterioscler Thromb Vasc Biol* 1997;17:3191-5.
46. Thijssen DH, Bullens LM, van Bommel MM, Dawson EA, Hopkins N, Tinken TM, Black MA, Hopman MT, Cable NT, Green DJ. Does arterial shear explain the magnitude of flow-mediated dilation?: a comparison between young and older humans. *Am J Physiol Heart Circ Physiol* 2009;296:H57-H64.
47. Black MA, Cable NT, Thijssen DH, Green DJ. Importance of measuring the time course of flow-mediated dilatation in humans. *Hypertension* 2008;51:203-10.
48. Pereira-Caro G, Borges G, Van Der Hoof J, Clifford MN, Del Rio D, Lean ME, Roberts SA, Kellerhals MB, Crozier A. Orange juice (poly) phenols are highly bioavailable in humans. *Am J Clin Nutr* 2014;100:1378-84.
49. Brett GM, Hollands W, Needs PW, Teucher B, Dainty JR, Davis BD, Brodbelt JS, Kroon PA. Absorption, metabolism and excretion of flavanones from single portions of orange fruit and juice and effects of anthropometric variables and contraceptive pill use on flavanone excretion. *Br J Nutr* 2008;101:664-75.
50. Vallejo F, Larrosa M, Escudero E, Zafrilla MP, Cerdá B, Boza J, García-Conesa MT, Espín JC, Tomás-Barberán FA. Concentration and Solubility of Flavanones in Orange Beverages Affect Their Bioavailability in Humans. *J Agric Food Chem* 2010;58:6516-24.
51. Vitaglione P, Donnarumma G, Napolitano A, Galvano F, Gallo A, Scafì L, Fogliano V. Protocatechuic acid is the major human metabolite of cyanidin-glucosides. *J Nutr* 2007;137:2043-8.
52. Speciale A, Cimino F, Saija A, Canali R, Virgili F. Bioavailability and molecular activities of anthocyanins as modulators of endothelial function. *Genes Nutr* 2014;9:404.

53. Khan F, Ray S, Craigie AM, Kennedy G, Hill A, Barton KL, Broughton J, Belch JJF. Lowering of oxidative stress improves endothelial function in healthy subjects with habitually low intake of fruit and vegetables: A randomized controlled trial of antioxidant- and polyphenol-rich blackcurrant juice. *Free Radic Biol Med* 2014;72:232-7.
54. Grosso G, Galvano F, Mistretta A, Marventano S, Nolfo F, Calabrese G, Buscemi S, Drago F, Veronesi U, Scuderi A. Red orange: experimental models and epidemiological evidence of its benefits on human health. *Oxid Med Cell Longev* 2013;2013.
55. Ashor AW, Siervo M, Lara J, Oggioni C, Afshar S, Mathers JC. Effect of vitamin C and vitamin E supplementation on endothelial function: a systematic review and meta-analysis of randomised controlled trials. *Br J Nutr* 2015;113:1182-94.

Tables

TABLE 1

Clinical characteristics of study participants at baseline ¹

	Values
Age (y)	28.7 ± 6.5 (20-45)
BMI (kg/m ²)	28.3 ± 3.1 (25.5-36.5)
SBP (mmHg)	110.0 ± 12.9 (91.0-128.7)
DBP (mmHg)	71.9 ± 9.5 (59.3-92.3)
Total cholesterol (mmol/L)	4.7 ± 0.6 (4.0-5.8)
HDL-cholesterol (mmol/L)	1.5 ± 0.5 (0.8-2.1)
LDL-cholesterol (mmol/L)	2.6 ± 0.4 (2.1-3.2)
Triglycerides (mmol/L)	1.3 ± 0.6 (0.5-2.8)
hsCRP (mg/L)	0.9 ± 0.9 (0.2-3.4)

¹Values are mean ± SD (range), n=15 except for lipids and hsCRP, n=10. DBP, diastolic blood pressure; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; LDL, low-density lipoprotein; SBP, systolic blood pressure.

Table 2

Endothelial function, blood pressure, and circulating inflammatory markers, lipids, endothelin 1, and cGMP in healthy adults with overweight and obesity at baseline and following 2-week consumption of blood orange juice or control drink in a random sequence ¹

	Blood orange juice		Control drink		<i>P</i> -treatment ²	<i>P</i> -time ³	<i>P</i> -interaction ⁴
	Basal	2 wk	Basal	2 wk			
Resting diameter (mm)	3.62 ± 0.56	3.64 ± 0.54	3.63 ± 0.57	3.62 ± 0.56	0.77	0.36	0.017
AUC _{peak} (1000 a.u.)	55.4 ± 19.1	57.3 ± 22.3	57.4 ± 27.4	56.6 ± 21.0	0.25	0.91	0.56
Absolute FMD (mm)	0.29 ± 0.08 ^b	0.36 ± 0.09 ^a	0.29 ± 0.07 ^b	0.27 ± 0.07 ^b	0.001	0.06	0.001
Relative FMD (%)	8.15 ± 2.92 ^b	10.2 ± 3.31 ^a	8.11 ± 2.52 ^b	7.77 ± 2.43 ^b	0.001	0.06	0.001
Scaled FMD index	1.23±0.03 ^b	1.25±0.03 ^a	1.23±0.02 ^b	1.22±0.02 ^b	0.001	0.06	<0.001
Time to peak diameter (s)	47.6 ± 13.6	47.5 ± 14.8	47.5 ± 17.1	46.5 ± 17.7	0.88	0.82	0.83
SBP (mmHg)	108 ± 12	108 ± 11	108 ± 11	108 ± 11	0.97	0.70	0.97
DBP (mmHg)	71 ± 8	69 ± 7	72 ± 9	70 ± 8	0.61	0.06	0.87
hsCRP (mg/L)	0.59 ± 0.29	0.58 ± 0.35	0.87 ± 0.82	1.14 ± 1.47	0.15	0.64	0.63
Total cholesterol (mmol/L)	4.63 ± 0.60	4.68 ± 0.56	4.55 ± 0.57	4.54 ± 0.77	0.24	0.82	0.81
Triglycerides (mmol/L)	1.23 ± 0.77	1.28 ± 0.61	1.38 ± 0.68	1.28 ± 0.61	0.58	0.37	0.57
LDL-cholesterol (mmol/L)	2.62 ± 0.39	2.60 ± 0.23	2.39 ± 0.46	2.46 ± 0.46	0.005	0.78	0.74
HDL-cholesterol (mmol/L)	1.53 ± 0.49	1.57 ± 0.57	1.59 ± 0.57	1.50 ± 0.49	0.82	0.61	0.27
Endothelin 1 (pg/mL)	1.08 ± 0.18	1.05 ± 0.27	1.15 ± 0.22	1.09 ± 0.29	0.51	0.40	0.83
cGMP (pmol/mL)	61.2 ± 19.0	78.9 ± 29.1	72.4 ± 15.0	65.9 ± 16.5	0.84	0.37	0.043

¹Values are mean \pm SD, n=15 except for lipids, endothelin 1, cGMP and hsCRP, n=10. Values with different superscript letters differ ($P<0.05$).

AUC_{peak}, area under the shear rate curve to peak dilation; cGMP, cyclic guanosine monophosphate; DBP, diastolic blood pressure; FMD, flow-mediated dilation; HDL, high-density lipoprotein; hsCRP, high-sensitivity C-reactive protein; LDL, low-density lipoprotein; SBP, systolic blood pressure.

²Indicates the main effect of treatment, $P<0.05$.

³Indicates the main effect of time, $P<0.05$.

⁴Indicates the time by treatment interaction, $P<0.05$.

Legends for figures

Figure 1

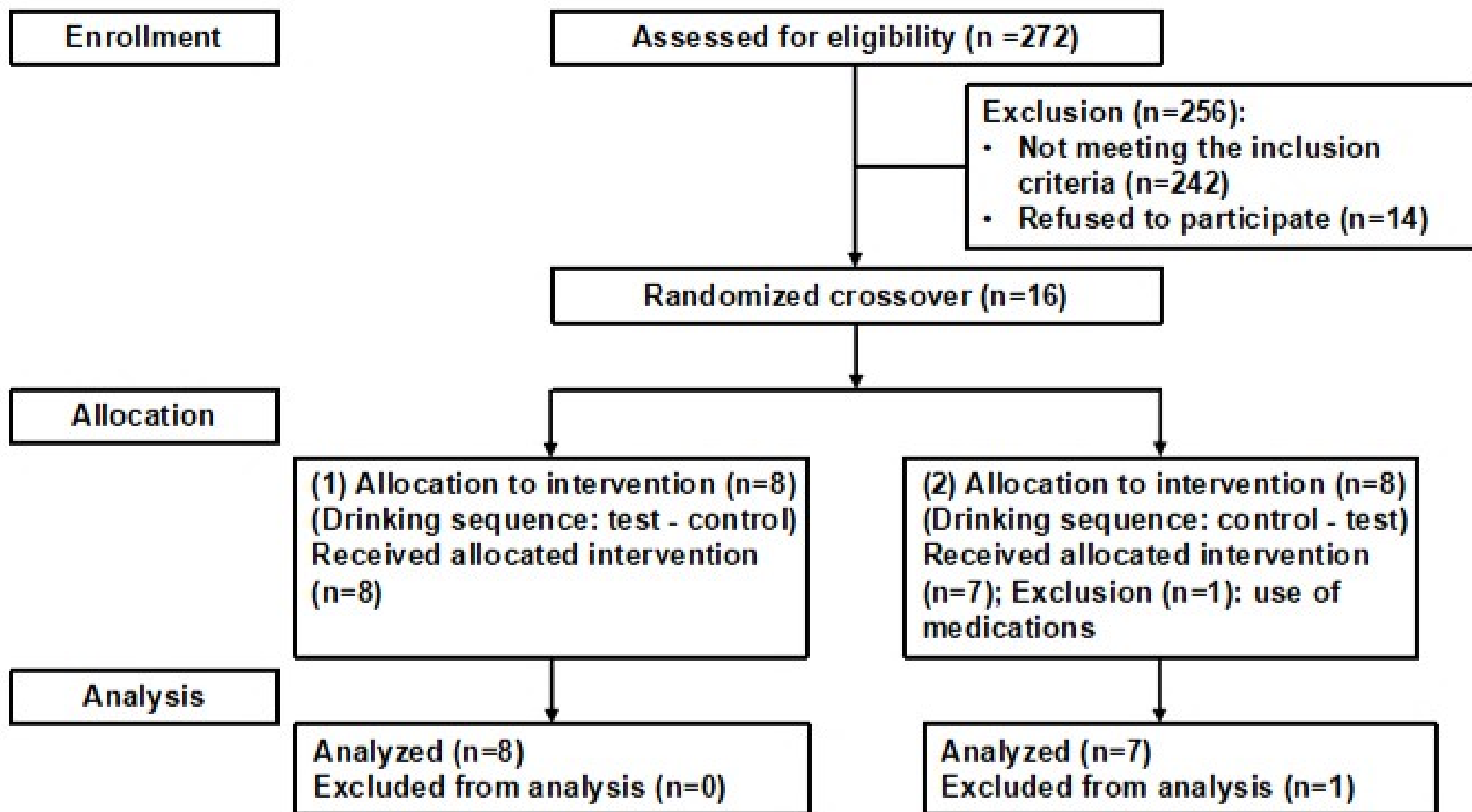
Participant flow diagram. Simple randomization was used to determine the different groups (starting with test drink or control drink). Block randomization was conducted to randomly allocate participants into groups to ensure an equal number in each group.

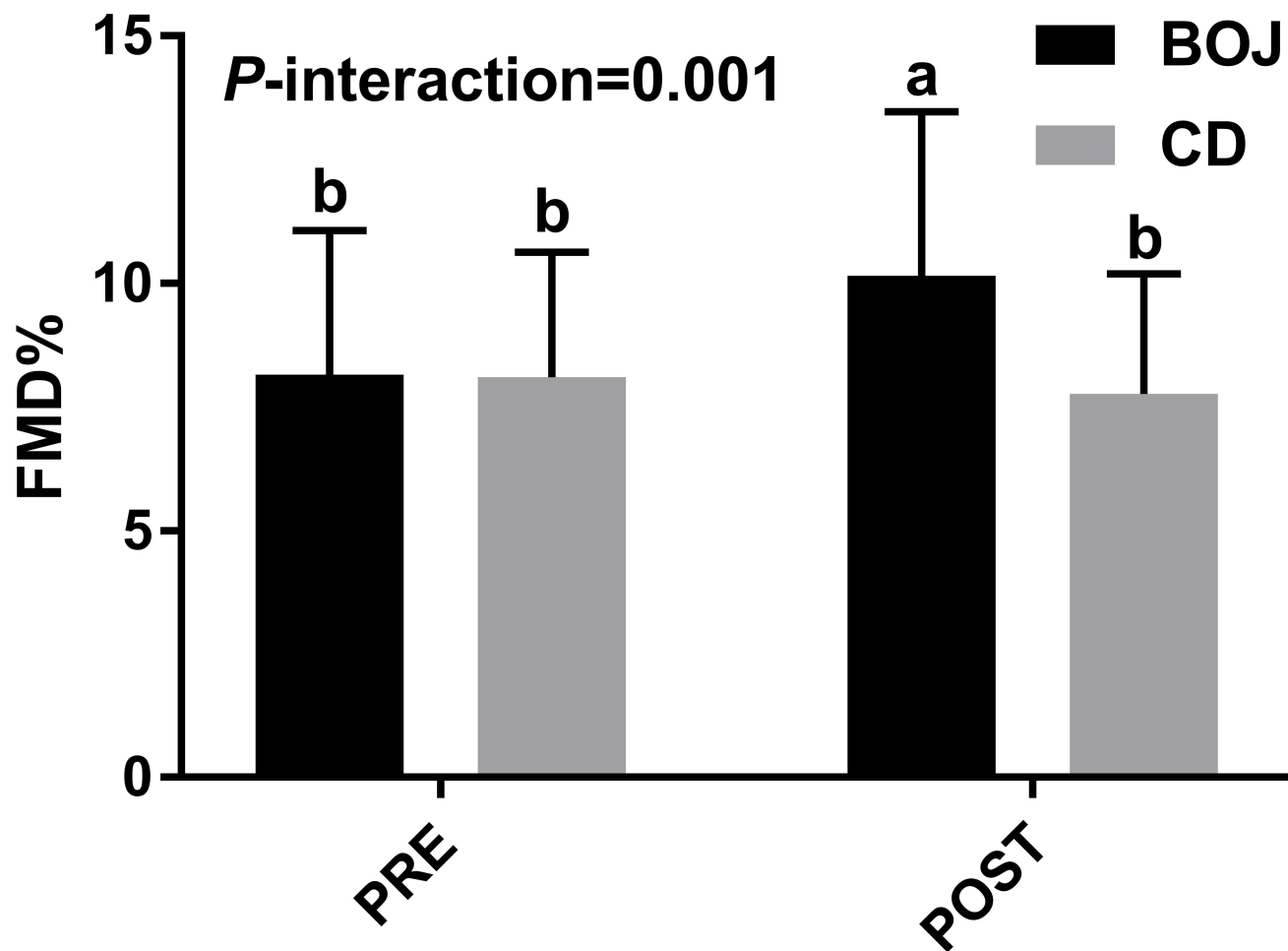
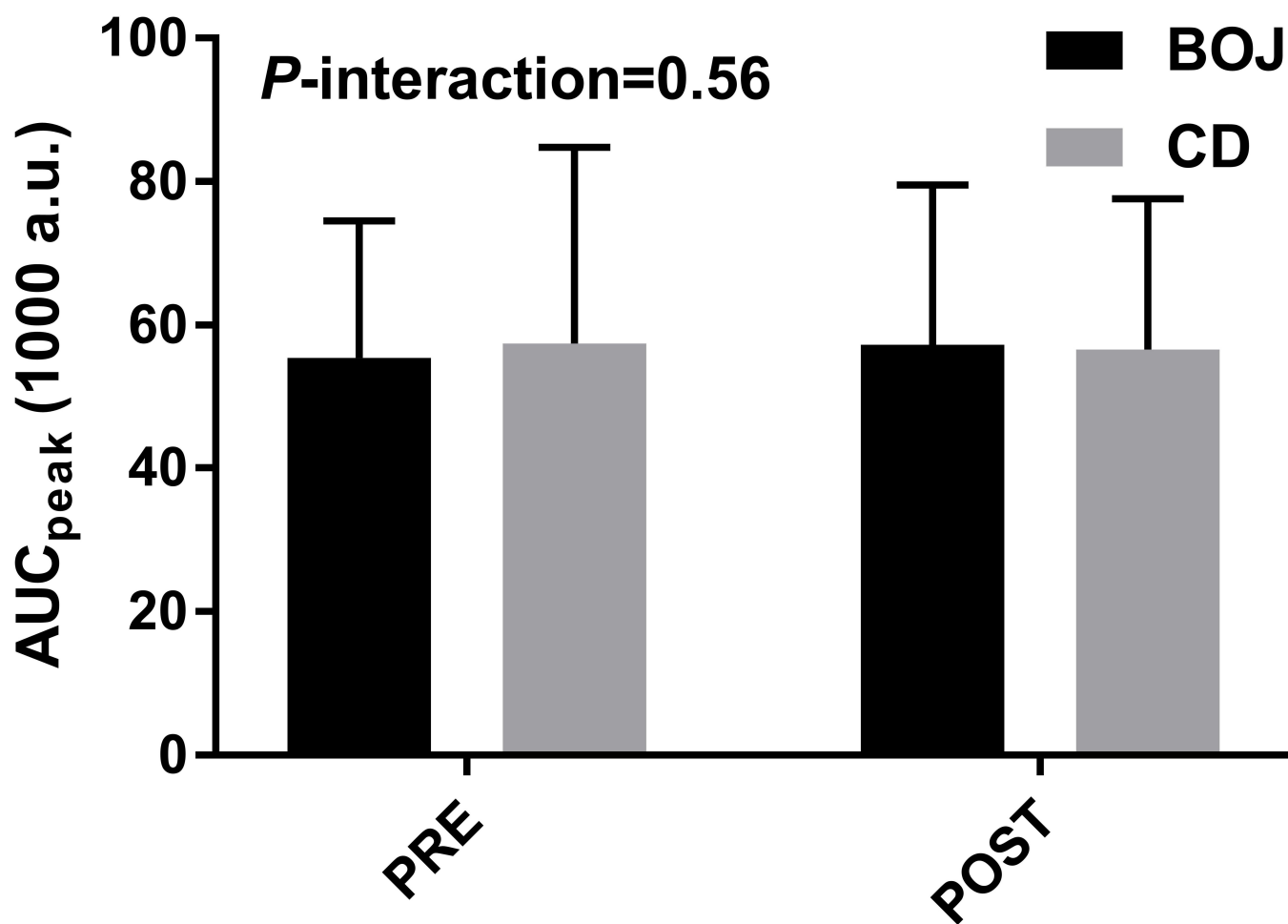
Figure 2

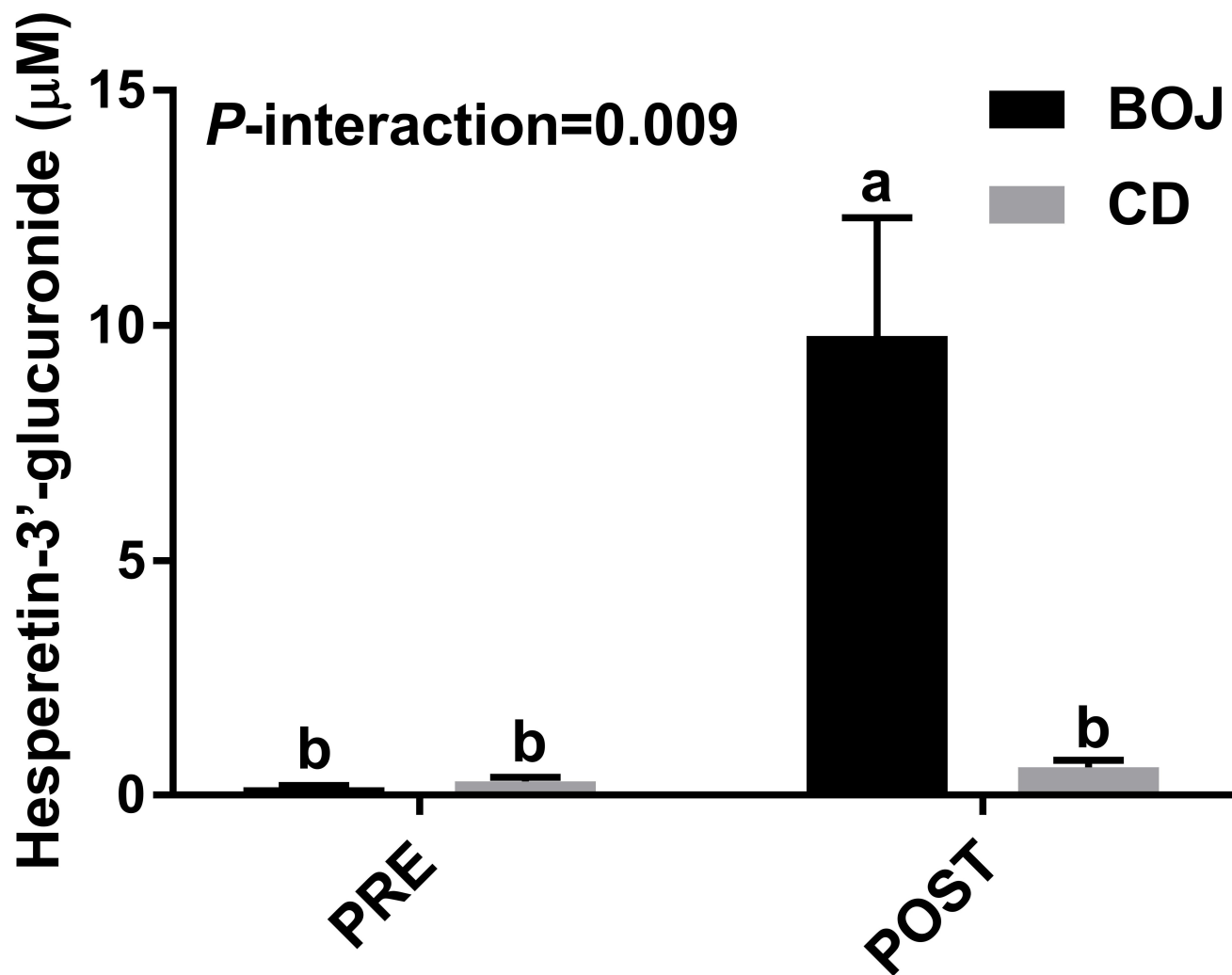
Endothelial function in healthy adults with overweight and obesity prior to and following 2-week consumption of blood orange juice or control drink (2 x 200 mL/d) in randomized order. (A) Relative FMD, (B) AUC_{peak}. Data are mean ± SD, n=15. Labeled means without a common letter differ, $P < 0.05$. AUC_{peak}, area under the shear rate curve to peak dilation; BOJ, blood orange juice; CD, control drink; FMD, flow-mediated dilation.

Figure 3

Urinary flavanone metabolites hesperetin-3'-glucuronide (A) and hesperetin-7-glucuronide (B) in healthy adults with overweight and obesity prior to and following consumption of blood orange juice or control drink (2 x 200 mL/d) in randomized order. Data are mean ± SD, n=15. Labeled means without a common letter differ, $P < 0.05$. BOJ, blood orange juice; CD, control drink.



A**B**

A**B**