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1 **CGIAR modeling approaches for resource-constrained scenarios: II. Accelerating crop**
2 **breeding for a changing climate**

3
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40

41

42 **Abstract**

43 Crop improvement efforts aiming at increasing crop production (quantity, quality) and adapting
44 to climate change have been subject of active research over the past years. But, ‘to what extent
45 can breeding gains be achieved under a changing climate, at a pace sufficient to usefully
46 contribute to climate adaptation, mitigation and food security?’. Here, we address this question
47 by critically reviewing how model-based approaches can be used to assist breeding activities,
48 with particular focus on all CGIAR (formerly the Consultative Group on International Agricultural
49 Research but now known simply as CGIAR) breeding programs. Crop modeling can underpin
50 breeding efforts in many different ways, including assessing genotypic adaptability and stability,
51 characterizing and identifying target breeding environments, identifying tradeoffs among traits
52 for such environments, and making predictions of the likely breeding value of the genotypes.
53 Crop modeling science within the CGIAR has contributed to all of these. However, much
54 progress remains to be done if modeling is to effectively contribute to more targeted and
55 impactful breeding programs under changing climates. In a period in which CGIAR breeding
56 programs are undergoing a major modernization process, crop modelers will need to be part of
57 crop improvement teams, with a common understanding of breeding pipelines and model
58 capabilities and limitations, and common data standards and protocols, to ensure they follow
59 and deliver according to clearly defined breeding products. This will, in turn, enable more rapid
60 and better-targeted crop modeling activities, thus directly contributing to accelerated and more
61 impactful breeding efforts.

62 **1. Introduction**

63 Global change projections indicate that many parts of the world will continue to face extreme
64 and erratic climate trends, as a result of rapid population growth, and increasing greenhouse
65 gas (GHG) emissions (IPCC, 2014). Model-based projections indicate greater heat and drought
66 stress during the 21st Century (Teixeira et al., 2013; Gourджи et al., 2013b; Li et al., 2015a),
67 especially (though not only) if no major GHG emission reduction strategies are implemented at
68 scale (Betts et al., 2011; Schleussner et al., 2016; Rogelj et al., 2016). Compounded by other
69 drivers of global change (e.g. population growth, changing economic structures, and changing
70 land use), these changes will reduce crop productivity and increase crop yield variability of
71 many crops (Li et al., 2009; Deryng et al., 2014; Campbell et al., 2016), with major implications
72 on farmer livelihoods (Morton, 2007; Jones and Thornton, 2009) and global food security
73 (Wheeler and von Braun, 2013; Campbell et al., 2016). In light of these projections, crop
74 improvement efforts aiming at increasing crop production (quantity, quality) in a sustainable
75 and efficient way have been subject of active research over the past years.

76

77 Crop breeding programs have contributed to farmers achieving higher yields, food security and
78 income by developing and delivering varieties with higher yield potential, as well as greater
79 resistance to pests and diseases, tolerance to abiotic stresses, and desirable market quality and
80 nutritional characteristics in the public (Evenson and Gollin, 2003; Pfeiffer and McClafferty,
81 2007) and private sectors (Cooper et al., 2014b; Voss-Fels et al., 2019d). Moreover, the use of
82 varieties resistant to heat stress, drought, and possible future pests and diseases can also
83 contribute to climate change adaptation (Takeda and Matsuoka, 2008; Habash et al., 2009;
84 Gourджи et al., 2013a; Gaffney et al., 2015). A key question is, however, ‘to what extent can
85 breeding gains be achieved under changing climates, at a pace sufficient so as to usefully
86 contribute to climate adaptation, mitigation and food security?’. Here, we address this question
87 by reviewing how model-based approaches can assist breeding activities, with particular focus
88 on the CGIAR (formerly the Consultative Group on International Agricultural Research but now
89 known simply as CGIAR). We critically discuss limitations and opportunities in light of the need
90 for greater breeding gains under changing climates. Since the body of published literature

91 (especially in some thematic or geographic areas) is substantial, we concentrate on the most
92 relevant examples, aiming at discussing their strengths and weaknesses, in order to ultimately
93 determine the main crop modeling gaps and strategies for collaboration with researchers, crop
94 improvement teams, farmers, and decision and policymakers. We first review the importance
95 of genotypic adaptation in delivering climate change adaptation outcomes (Sect. 1.1), as well as
96 the challenges in converting potential adaptations into actual genetic improvement (Sect. 1.2).
97 We then review tools and approaches for accelerated breeding (Sect. 2), including modeling of
98 environment- and management- specific yield responses (Sect. 2.1), environmental
99 characterization (Sect. 2.2), ideotype design (Sect. 2.3), the linking of crop modeling and genetic
100 data (Sect. 2.4), and simulation methods for optimizing breeding pipelines (Sect. 2.5). Finally,
101 we discuss limitations in terms of data, models, and approaches (Sect. 3), and conclude by
102 proposing a set of next collaborative research activities that can contribute to maximizing
103 breeding gains under climate change.

104

105 **1.1. The importance of genotypic adaptation under climate change**

106 Modern varieties developed through years of crop improvement have contributed to large
107 increases in crop production in the last 60 years, and they will continue to do so. Evenson and
108 Gollin (2003) reviewed breeding gains during and after the Green Revolution for 11 major food
109 crops, estimating that the contribution of modern varieties to yield growth is in the range 17–
110 50 % in the period 1961–2000. One notable example is the 70 % yield potential increase from
111 the release of the semi-dwarf rice variety IR8 by the International Rice Research Institute (IRRI)
112 in the 1950s and 1960s (Peng et al., 2008). Fischer et al. (2014) indicate a rate of progress in
113 potential yield of 0.5 to 1.08 % per year for wheat, rice, maize and soybean, as a result of crop
114 improvement. Genotypic adaptation to climate change –that is, the process of designing and
115 developing novel crop varieties to enhance productivity and stability under future climates, has
116 the potential to continue delivering productivity gains under changing climates (Rötter et al.,
117 2015; Ramirez-Villegas et al., 2015).

118

119 Evidence of how genotypic adaptation can effectively contribute to climate change adaptation

120 generally arises from two types of studies: those in which models are used to simulate future
121 growing conditions with and without adaptation; and those that quantify the yield benefit of
122 climate-adapted genotypes by means of field experimentation. Model-based studies generally
123 indicate potential for substantial gains in both yield and yield stability. A systematic review of
124 the literature (by searching the keywords 'climate change', 'genotypic adaptation', and
125 'ideotype' in <http://scholar.google.com> in June 2019) identified 19 studies, from which 389
126 individual data points for eight crops were drawn. While some publication bias is expected in
127 the meta-analysis, the identified studies indicate that gains from genotypic adaptation are
128 positive for a number of crops (Fig. 1). For instance, modelling by Semenov and Stratonovitch
129 (2013) suggested that if certain traits could be improved adequately, wheat ideotypes could
130 outperform current cultivars in Europe by up to 65 % under future climates. Similarly,
131 simulations by van Oort and Zwart (2018) showed that favoring varieties with greater thermal
132 time can compensate for climate change-induced yield reductions in African rice systems.
133 Similar findings have been reported for Asian rice (Li and Wassmann, 2010; Mottaleb et al.,
134 2017), groundnut (Singh et al., 2012, 2014b), sorghum (Singh et al., 2014c), pearl millet (Singh
135 et al., 2017), chickpea (Singh et al., 2014a), maize (Tesfaye et al., 2017), and wheat in China
136 (Challinor et al., 2010).

137

[Figure 1 near here]

139

140 Experimental studies also provide robust evidence on the benefits of genotypic adaptation,
141 corroborating or extending model-based findings. On-farm maize trials in Africa have shown
142 that drought-tolerant maize can yield between 20–25 % more than current commercial
143 varieties, with no yield penalty in 'good' years (Setimela et al., 2017; Cairns and Prasanna,
144 2018). Suarez Salazar et al. (2018) identified common bean lines adapted to a heat-stress
145 environment in the Colombian Amazon, where commercial bean varieties show low yield.
146 Mondal et al. (2016) estimated genetic yield gains in the range 0.5–0.8% per year when
147 breeding short-cycle heat-adapted wheat varieties in South Asia. Success has also been
148 reported for drought tolerance in maize for the United States corn belt (Cooper et al., 2014a;

149 Messina et al., 2015) and other regions and crops (Sinclair et al., 2020). These studies provide
150 initial evidence that genotypic adaptation can indeed deliver greater yields in stress-prone
151 environments, thus likely contributing to future adaptation outcomes.

152

153 **1.2. Current warming rates will reduce yields unless breeding and seed system efficiency is** 154 **enhanced**

155 The process of breeding a novel cultivar, increasing seed availability and achieving significant
156 adoption often takes more than a decade (Langyintuo et al., 2008; Challinor et al., 2016). This
157 means that temperature increases during the breeding cycle can lead to a systematic (and
158 unintended) yield reduction due to decreases in the duration of the growing cycle (Zheng et al.,
159 2016; Challinor et al., 2016). Researchers confirmed that the challenge is more critical in many
160 subtropical areas with emerging precipitation trends under climate change (Rojas et al., 2019).
161 The breeding of climate-ready crops should, therefore, seek to deliver more productive and
162 resilient crops that keep pace with climate change (Ramirez-Villegas et al., 2015; Challinor et al.,
163 2016). In doing so, it is important to implement a wide range of solutions aiming at making the
164 breeding process more effective and efficient. Anticipatory and predictive tools using crop-
165 climate models (reviewed in Sect. 2 and 3) can enable preemptive breeding and can help
166 enhance and accelerate breeding gains, ultimately ensuring crop improvement contributes
167 effectively to addressing major challenges for agriculture within the context of climate change.

168

169 **2. Tools and approaches for accelerating trait discovery in target environments**

170 For plant breeding, multi-environment trials (METs) are conducted regularly to study Genotype
171 (G) × Environment (E) × Management (M) interactions (G×E×M), assess genotypic adaptability
172 and stability, and make predictions about the breeding value of the genotypes in other
173 environments and years that will allow crop improvement teams to accurately select the
174 parents for the next breeding cycle. This section reviews modeling approaches to assess G×E
175 interactions (Sect. 2.1), characterize target breeding environments (Sect. 2.2), understand ideal
176 plant types for such target environments (Sect. 2.3), predict breeding values (Sect. 2.4), and

177 breeding cycle optimization (Sect. 2.5). We review both process-based eco-physiological models
178 as well as statistical approaches to G×E prediction.

179

180 **2.1. Modeling Genotype × Environment × Management**

181 Accurately predicting G×E responses allows identifying well-adapted genotypes for specific sites
182 or stress situations (Hammer and Broad, 2003; Banterng et al., 2006), or testing ‘virtual’
183 genotypes to inform breeding programs (Cock et al., 1979; Suriharn et al., 2011; Bogard et al.,
184 2020; Hammer et al., 2020). Similarly, predicting management responses allows identification
185 of appropriate levels of fertilization, tillage, irrigation, weed control, amongst others, for a given
186 set of genotypes (Boote et al., 1996; Artacho et al., 2011; Deryng et al., 2011). Accurate
187 prediction of genotype performance across environments and management options is
188 contingent on various elements, including (i) the development of a model with the necessary
189 physiological processes represented at an appropriate level of complexity (Challinor et al.,
190 2009; Boote et al., 2013; Hammer et al., 2019); (ii) the development of a well-constrained
191 parameter set (Iizumi et al., 2009; Angulo et al., 2013); and (iii) high quality environmental (soil,
192 climate) data to drive the model (Lobell, 2013). As discussed below, the CGIAR has made major
193 contributions in these three areas. The use of models for analyzing G × E × M interactions for
194 accelerating breeding is described in Sect. 2.1.2.

195

196 **2.1.1 Model development, parameterization and input data**

197 Model development requires the acquisition of a deep understanding of the biological basis of
198 G×E interactions (i.e. crop physiology), and the translation of such understanding into computer
199 code. Physiological processes of interest, and approaches to model those processes, can vary,
200 sometimes substantially, between contexts. During the early 1980s, progress in process
201 understanding by CGIAR scientists led to the development of three crop models that ably
202 captured G×E×M responses, while also contributing data and knowledge to many other models.
203 Perhaps the earliest crop model developed and used in the CGIAR was the cassava model
204 developed by the International Center for Tropical Agriculture (CIAT) (Cock et al., 1979), upon
205 which various components of the GUMCAS model (Matthews and Hunt, 1994), the CROPSIM-

206 Cassava model (Jones et al., 2003), the model of Gabriel et al. (2014), and the MANIHOT model
207 (CIAT, unpublished), were later developed. The ORYZA1 rice model (Kropff et al., 1993a, 1994),
208 developed at the International Rice Research Institute (IRRI), incorporated many years of eco-
209 physiological research from IRRI and elsewhere. ORYZA1 quickly evolved into ORYZA2000
210 (Bouman et al., 2001), and later into ORYZAv3 (Li et al., 2017). The International Potato Center
211 (CIP) has also led the development of the SOLANUM and the dynamic carbon photosynthesis
212 model (DCPM) models for potato (Condori et al., 2010; Quiroz et al., 2017) and sweet potato
213 (Ramírez et al., 2017). Lastly, CIAT also led the early development of the BEANGRO model,
214 which is currently part of the 'CROPGRO' module within DSSAT (Decision Support System for
215 Agrotechnology Transfer) (White and Izquierdo, 1991; Hoogenboom et al., 1993; White et al.,
216 1995). The International Center for Agricultural Research in the Dry Areas (ICARDA) and the
217 International Crops Research Institute for the Semi-Arid Tropics (ICRISAT) have extensively
218 contributed to the development and improvement of the Simple Simulation Model (SSM)
219 (Soltani and Sinclair, 2012; Sinclair et al., 2020), the CROPGRO for peanut and chickpea, and the
220 CERES model for sorghum and pearl millet, also available in DSSAT.

221
222 The determination of parameters for crop models, whether they are statistically- or process-
223 based, is crucial to ensure that the model correctly captures genotype behavior across different
224 combinations of climate, soils, and management options. This is especially important for
225 process-based crop models, since the sometimes large number of parameters required means
226 that there can be many more degrees of freedom in the model than can be constrained by the
227 available data (Challinor et al., 2014; Wallach et al., 2016). Progress in model parameterization
228 has been enabled by extensive eco-physiological trials conducted by the CGIAR. Notably, recent
229 progress in characterizing yield changes in response to heat stress for wheat, at least in part,
230 was possible due to data collected in the International Heat Stress Genotype Experiment
231 (IHSGE) carried out by the International Maize and Wheat Improvement Center (CIMMYT)
232 (Asseng et al., 2014). Based on CIMMYT data, too, statistical models by Lobell et al. (2011) and
233 Gourdjji et al. (2013a) assessed maize and wheat genotype responses to temperature,
234 respectively. Li et al. (2015b) used IRRI trial data from Los Baños (Philippines) and Ludhiana

235 (India) to calibrate and evaluate 13 different rice crop models, and Fleisher et al. (2017) used an
236 experiment from Bolivia conducted by the International Potato Center (CIP) as part of the
237 calibration and evaluation dataset for nine potato crop models. The use of remote sensing has
238 also been tested for the estimation of crop model parameters at a low cost (Quiroz et al., 2017).

239
240 Attempts to estimate model parameters from genetic information date to work in the 1990s at
241 CIAT on common bean, where simulations from gene-based estimates of model parameters
242 were generally as accurate as manually estimated parameters (White and Hoogenboom, 1996;
243 Hoogenboom et al., 1997). Similar work in collaboration with CIMMYT, showed that differences
244 in phenology of winter wheat cultivars could be simulated based on genetic information (White
245 et al., 2008). Work also extends into the development of a gene-to-phenotype model for
246 common bean based field trials conducted by CIAT and the University of Florida (UF) (Hwang et
247 al., 2017). Compared to success in linking gene-to-phenotypes achieved by other institutions
248 (Messina et al., 2006; Chenu et al., 2009; Bogard et al., 2020), progress in the CGIAR remains
249 slow.

250
251 The CGIAR has also contributed to the development of key spatially-explicit climate datasets
252 that are used as inputs into crop models. These include WorldClim (Hijmans et al., 2005) and
253 the Climate Change, Agriculture and Food Security (CCAFS)-Climate database (Navarro-Racines
254 et al., 2020), as well as methods to generate daily weather data for crop model simulations
255 (Jones and Thornton, 2000, 2013). Contributions to soil (Jones and Thornton, 2015; Vågen et al.,
256 2016; Piikki et al., 2017) and crop geography (You et al., 2009, 2017; IFPRI, 2019) for crop
257 modeling have also been made in recent years. By contrast, CGIAR work on developing datasets
258 that characterize crop management for crop modeling is limited to specific regional or national
259 studies (see Sect. 2.1.2).

260

261 **2.1.2 Explaining and simulating G×E×M interactions**

262 Using available data and models, CGIAR modelers have studied G×E×M interactions extensively
263 in close coordination with breeding programs. Virtually all centers have done modeling for their

264 mandate crops. Use of models has focused on assessing the stability of economically important
265 traits and predicting the performance of newly developed genotypes evaluated under varying
266 environmental conditions and management practices.

267

268 Statistical approaches have the longest history in plant breeding. These models can be used to
269 study both univariate (Cossa et al., 2004) and multivariate responses (Montesinos-López et al.,
270 2018d). A recent study with on-farm wheat trials (Vargas-Hernández et al., 2018) used a
271 univariate model to assess the combined effects of the wheat lines and their interactions with
272 the farmer-irrigation-year combinations for several traits. For single trait grain yield, the study
273 identified stable wheat lines across all environments, as well as the environments that caused
274 most of the G×E interaction. Multivariate models, though less used, are particularly useful when
275 measurements are available for multiple response variables (i.e. multi-trait), and the objective
276 is to increase our understanding of the complex nature of particular phenomena under field
277 conditions. Many studies have shown that a multivariate approach is better than univariate
278 approaches because it identifies the existing (co)variation between the response variables
279 (Xiong et al., 2014). Moreover, the multivariate analysis also improves accuracy when
280 classifying and identifying superior genetic components (Montesinos-López et al., 2018d). In
281 addition, it increases the precision of genetic correlation parameter estimates between traits,
282 which helps crop improvement teams perform indirect selection. Multivariate models have
283 been implemented using Bayesian analysis (Montesinos-López et al., 2016b) as well as deep
284 machine learning regression (Montesinos-Lopez et al., 2018; Montesinos-López et al., 2018c).
285 Notably, Montesinos-Lopez et al. (2018) report that the performance of multi-trait and multi-
286 environment deep learning (MTDL) is commensurate with that of the Bayesian multi-trait and
287 multi-environment approach. Erzos et al. (2020) and Washburn et al. (2020) review machine-
288 learning approaches in crop improvement.

289

290 Process-based crop models have also been used for assessing G×E×M interactions within the
291 CGIAR. At ICRISAT, crop models are used to investigate whether and how changes in G and M
292 result in positive change in yield across different environments, as a way to prioritize breeding

293 and agronomic intervention decisions, including sowing density (Vadez et al., 2017), irrigation
294 (Vadez et al., 2012), the combination of better-adapted genotypes and irrigation (Soltani et al.,
295 2016), and different traits and combinations of traits (Kholová et al., 2014). ICARDA has
296 employed the Simple Simulation Model (SSM) to characterize the stress scenarios in target
297 regions of focus, as well as to explore plant traits and/or management to be exploited in stress-
298 adapted cultivars for specific target environments (Ghanem et al., 2015; Guiguitant et al.,
299 2017). CIP has used the SOLANUM (Condori et al., 2010) and LINTUL (Spitters, 1988, 1990)
300 models to compare the performance of native and hybrid potato genotypes under extreme
301 climatic conditions (Condori et al., 2010, 2014) and climate change (Quiroz et al., 2018),
302 demonstrating that appropriate choice of germplasm and crop management practices could
303 significantly secure and increase potato production under future climate conditions.

304

305 Similar work has been conducted by IRRI, whereby high yielding and stable genotypes were
306 identified using the ORYZA2000 crop model (Li et al., 2013a). At IRRI, simulations have also
307 been used to simulate potential yield across environments (Kropff et al., 1993b), identify
308 ideotypes for increasing rice yield potential (Kropff et al., 1995; Aggarwal et al., 1997; Dingkuhn
309 et al., 2015, 2016), and to inform national certification processes for the release of crop
310 varieties (Li et al., 2016). At CIMMYT, grid-based global-scale simulations are used to assess the
311 value of certain traits. This modeling capacity was developed in a consortium of UF, CIMMYT,
312 and the International Food Policy Research Institute (IFPRI) that incorporated three crop
313 models, including CERES-wheat, CROPSIM, and NWheat (Gbegbelegbe et al., 2017; Hernandez-
314 Ochoa et al., 2018). At CIAT, crop models have been used to understand drought responses
315 across G and M for rice and beans (Heinemann et al., 2016; Ramirez-Villegas et al., 2018), as
316 well as to assess the value of drought tolerance traits (Heinemann et al., 2019). At IFPRI, a grid-
317 based crop modeling framework was developed and linked with the IMPACT global trade and
318 economic model (Robinson et al., 2015) to simulate the potential impacts of adopting
319 agricultural technologies (e.g. precision agriculture), management practices (e.g. integrated soil
320 fertility management), and breeding target traits (e.g. drought and/or heat tolerance) on global
321 food security and economic implications under climate change scenarios (Rosegrant et al.,

322 2014; Islam et al., 2016). Analyses of climate risk for rice in Africa have also been possible by
323 crop simulation at the Africa Rice Center (van Oort et al., 2015b; a).

324

325 **2.2. Environmental characterization for setting breeding priorities**

326 The existence of significant G×E×M interactions can slow plant breeding progress for broad
327 adaptation and/or for adaptation to specific conditions within a region (Löffler et al., 2005;
328 Chenu et al., 2011). The Target Population of Environments (TPE) approach aims at addressing
329 G×E×M through model-based environmental characterization (Lacaze and Roumet, 2004;
330 Chenu, 2015). In the TPE approach, process-based crop models are used to assess and detect
331 stress patterns and their impacts. This, in turn, allows prioritizing stress types by their intensity
332 and frequency across geographies, as well as identifying sites where selection for given stresses
333 is likely to be more successful. Thus, TPEs offer a concrete way to aid breeding programs
334 through effectively setting trait and geographic priorities. The TPE approach has been used with
335 success by wheat breeding programs in Australia (Chenu et al., 2011, 2013; Lobell et al., 2015),
336 and more recently has been applied to maize in Europe (Harrison et al., 2014).

337

338 Compared with the applications described in Sect. 2.1, CGIAR's work on TPEs for breeding
339 programs is less in terms of number of crops covered and published studies (Fig. 2A, B). CGIAR's
340 collaborative efforts include studies addressing drought for rice (Heinemann et al., 2015;
341 Ramirez-Villegas et al., 2018) and beans (Heinemann et al., 2016, 2017) under current and
342 future climate in Brazil. Significant breeding progress has resulted from these studies, including
343 improvements in drought phenotyping in a drought-prone environment which allows
344 controlling the timing, intensity, and duration of drought, reducing the uncertainty associated
345 with climate variability trials in the main season (Martinez et al., 2014).

346

347 [Figure 2 near here]

348

349 For rainfed beans, EMBRAPA (*Empresa Brasileira de Pesquisa Agropecuaria*) initiated a drought
350 tolerance breeding program following the results of Heinemann et al. (2016, 2017), though its

351 implementation was halted due to the Brazilian economic crisis. For post-rainy sorghum in
352 India, Kholová et al. (2013) report five main types of stress conditions requiring different
353 breeding/agronomic approaches (Fig. 2B). A related larger-scale method, referred to as Mega-
354 Environments, has been used by CIMMYT to target breeding since the 1990s (Rajaram et al.,
355 1994; van Ginkel et al., 2002; Cairns et al., 2013). Though less mainstream in the CGIAR in terms
356 of centers and traits, TPE and Mega-Environment work has the potential to help better-target
357 breeding programs across scales (see Fig. 2C). Similar TPE analysis has also been done for
358 chickpea in India (Hajjarpoor et al., 2018).

359

360 The first step across breeding programs should be to map what stresses exist currently (in
361 recent decades) to then analyze changes in stress patterns under future climates (Harrison et
362 al., 2014; Lobell et al., 2015). Using TPE results to stratify the target geographic area of the
363 breeding program, in combination with socio-economic (e.g. farmer preference) studies,
364 breeding programs can then decide which products are most relevant and impactful. Contrary
365 to the private sector (Cooper et al., 2014a; Voss-Fels et al., 2019a; c), however, to the best of
366 our knowledge, the extent to which this is currently done in a systematic way across the CGIAR
367 system is very limited. Yet, taking into account the TPE definition as part of the definition of the
368 breeding products will allow modelers to impact breeders, while allowing breeders to discuss
369 model results from the start of the breeding process.

370

371 **2.3. Design of ideotypes for future target environments**

372 With a clear understanding of the target stresses for breeding, a key use of process-based crop
373 models is to determine which traits can maximize yield in each target environment. When
374 applied to a range of traits simultaneously, this then becomes a process referred to as ideotype
375 design (Donald, 1968; Rasmusson, 1987). Ideotypes can be developed for current as well as for
376 future climates via a variety of methods ranging from iterative testing changes in model
377 parameters (Suriharn et al., 2011; Dingkuhn et al., 2015), optimization to maximize mean yield
378 and minimize yield variability (Semenov and Stratonovitch, 2013; Hammer et al., 2020), or by
379 developing gene- or trait × gene-specific components into the crop models (White and

380 Hoogenboom, 1996; Hoogenboom et al., 2004; Messina et al., 2006). Modeling in this case is
381 based on traits that have previously indicated promise for example in boosting yield potential
382 (Reynolds et al., 2012), adaptation to heat stress (Cossani and Reynolds, 2012), amongst others.
383 Table 1 summarizes all existing studies in which models have been used to design ideotypes
384 within the CGIAR; that is, conducted by CGIAR scientists on CGIAR mandate crops and
385 geographic areas.

386

387 [Table 1 near here]

388

389 The 12 studies listed in Table 1, published in a span of 9 years, indicate that CGIAR Centers are
390 very active in this area of work. The list, while not extensive in terms of crops or countries (i.e.
391 covers five crops across five countries), offers valuable insights as to the methods used and the
392 potential value of these analyses for breeding programs. Foremost, we note that all studies use
393 systematic parameter modifications to create ideotypes, which suggests opportunities to
394 explore optimization methods as well as more direct gene-to-phenotype modeling (e.g. van
395 Eeuwijk et al., 2019). Additionally, the similarity in the ideotypes proposed for different studies
396 (e.g. chickpea, sorghum and groundnut) suggests the need for refinement in the traits assessed
397 through discussion with crop improvement teams, or through the use of more detailed eco-
398 physiological models (Rebolledo et al., 2015; Dingkuhn et al., 2016). Such similarity could also
399 suggest that the models may fail to capture cropping system dynamics realistically when
400 subjected to these parameter modifications. Furthermore, little connection is seen in most
401 studies between the parameter variations proposed and existing ideotypes for these crops,
402 except for the study of Mottaleb et al. (2017). Additionally, there is a need to ensure that
403 parameter modifications, especially when several traits are simulated simultaneously, are done
404 within realistic biological bounds (Koornneef and Stam, 2001).

405

406 Finally, we note that moving from a set of prescribed changes in model parameter values (as
407 reported in the studies listed in Table 1) to a range of phenotypic screens that can be feasibly
408 measured and selected for in breeding trials is not a trivial process. Most notably, it requires

409 delivering information on (i) the available genetic diversity, (ii) heritability, and (iii) high-
410 throughput phenotyping methods for the trait in question. Future research within and outside
411 the CGIAR will need to capitalize on existing well-calibrated models, results from environmental
412 characterization, methods to connect eco-physiological models with genetic data (see Sect.
413 2.4), in better connection with existing ideotypes and crop improvement teams and their
414 knowledge, needs and priorities.

415

416 **2.4. Assisting varietal selection through linking crop models and genetic information**

417 A more recent area of work aims at directly linking crop model and genetic information with the
418 aim of addressing two different, but related, questions (i) what is the phenotypic response of a
419 set of genotypes for which the genetics are known, but on which no phenotyping has been
420 conducted?; and (ii) what is the phenotypic response of a set of genotypes (with known
421 genetics) in a location where environmental (soil, climate) characteristics are known, but no
422 phenotyping has been conducted? As the methods to be used may depend on the crop and
423 geographic areas of interest (e.g. due to differences in data availability, targets, and breeding
424 methods), several potential avenues need to be explored to address these questions (Asseng et
425 al., 2019a). These are discussed below.

426

427 **2.4.1 Link environmental information into genomic selection models**

428 Genomic selection (GS) that leverages genome-wide molecular marker information to select
429 individuals based on their predicted genetic merit (Meuwissen et al., 2001) is a promising tool
430 for accelerating crop genetic gains in the face of climate change. In a recent paper, Zhang et al.
431 (2017) reported genetic gains of 0.225 ton ha⁻¹ per cycle (or 0.100 ton ha⁻¹ year⁻¹) from rapid
432 cycling genomic selection for four recombination cycles in a multi-parental CIMMYT tropical
433 maize population (Fig. 3). However, in spite of these early findings and the fact that GS has
434 revolutionized animal breeding by increasing the accuracy of selections and reducing cycle time
435 and cost (Hayes et al., 2013; Hickey et al., 2017), its implementation in CGIAR crop breeding
436 programs is still limited (focusing primarily on the major cereals), in part due to costs associated
437 with routine evaluation and relatively low prediction accuracy due to G×E.

438

439

[Figure 3 near here]

440

441 CGIAR has done extensive research to evaluate the genomic predictabilities of several traits
442 including phenology, grain yield and its components, disease resistance, quality and
443 micronutrients (Grenier et al., 2015; Crossa et al., 2016a; Juliana et al., 2017a, 2018; Sukumaran
444 et al., 2018). The accuracy of forward predictions for grain yield (using a previous nursery/year
445 to predict the next nursery/year) is, however, low and highly influenced by the environment
446 (Juliana et al., 2018), thereby highlighting the importance of incorporating environmental data
447 in genomic prediction models for grain yield (van Eeuwijk et al., 2019; Bhandari et al., 2019).
448 Several novel methods and statistical models for modeling genomic relationships, pedigree
449 relationships, environmental data and genomic \times environment (G \times E) interactions have been
450 developed and evaluated in the CGIAR. These methods (see Table 2) vary in the type of
451 information they use as input, the way they assess G \times E interactions, and their prediction
452 purpose and accuracy. Notably, studies comparing the predictive abilities of some of these
453 approaches have also been conducted (Pérez-Rodríguez et al., 2012; Juliana et al., 2017b;
454 Montesinos-López et al., 2018a).

455

456

[Table 2 near here]

457

458 While GS models are promising tools to accelerate breeding gains, further research is needed to
459 understand how they fit in different stages of the breeding cycle, their comparative advantage
460 over conventional breeding, their integration with rapid cycling technologies such as speed
461 breeding (Voss-Fels et al., 2019b), and the type of approach used to integrate crop and genomic
462 models (Messina et al., 2018; Voss-Fels et al., 2019a; van Eeuwijk et al., 2019). Experience for
463 hybrid maize breeding in the private sector, however, offers evidence of the potential of GS for
464 enhancing breeding gains (Cooper et al., 2014b, 2020).

465

466 **2.4.2 Models that capture trait-trait relationships**

467 Crop models aimed at capturing trait-trait relationships, developed with sufficient simplicity to
468 be understandable, yet with enough mechanistic detail to be robust, can also help crop
469 improvement teams in the selection process. CGIAR involvement and leadership in this area is
470 very limited. These models can be useful in situations where a trait is too difficult to screen but
471 is clearly predictable on the basis of other, more easily measurable, traits. Fundamental
472 changes in the structure of current crop models would, however, be required for this approach
473 to be implemented. That is, models should be sufficiently generic to be applicable across
474 genotypes with limited or no calibration requirements (Soltani and Sinclair, 2012; Kholová et al.,
475 2014; Holzworth et al., 2014). More flexibility in the types of model inputs may also be required
476 when dynamic changes in certain plant traits are used as predictors. For instance, prediction
477 modeling for genotype values can use correlated physiological traits measured using high-
478 throughput phenotyping (HTP) platforms. This, in turn, facilitates indirect selection for grain
479 yield in early-generations. Integration of HTP data for canopy reflectance and vegetation indices
480 in genomic and pedigree-relationship based prediction models has proven to increase
481 prediction accuracies in several studies (Rutkoski et al., 2016; Sun et al., 2017, 2019; Juliana et
482 al., 2019).

483

484 **2.4.3 Gene-based crop simulation models**

485 Another way to couple crop models and genetic data is to develop models or model
486 components that, from the start, use genetic and environmental information and are based on
487 empirical relationships from available agronomic trial and marker data. This can be achieved
488 through a highly dynamic approach (Hwang et al., 2017), at the expense of increasing
489 uncertainty, or through prediction of crop state variables at coarser time scales, at the expense
490 of mechanistic detail. Dynamic approaches that link genetic information with crop simulation
491 models have proved successful for crop development variables (i.e. phenology) (White and
492 Hoogenboom, 1996; Yin et al., 2004; White et al., 2008) as well as for more complex traits
493 (Chenu et al., 2009; Bertin et al., 2010). The current level of direct engagement and leadership
494 by the CGIAR in this line of work is very limited, likely due to a combination of CGIAR center-
495 specific focus, funding sources for modelers, and limited uptake and applicability of these

496 models in CGIAR breeding programs.

497

498 **2.5. Optimization of breeding methods through genetic modeling and simulation**

499 In addition to modeling of cropping systems and trait-specific responses, simulation work also
500 extends to the design of breeding pipelines. While not directly related to crop modeling, we
501 include this area of work in our review as constitutes part of the simulation tools available to
502 crop improvement teams. These computer tools are capable of simulating the performance of a
503 breeding strategy. For instance, QuLine, can simulate the selection of inbred lines, which means
504 most major food cereals in the world, plus basically all leguminous crops (Wang et al., 2003,
505 2005; Wang and Pfeiffer, 2007). QuLine has been used to compare and optimize conventional
506 selection strategies (Wang et al., 2003, 2009; Li et al., 2013b), to predict cross performance
507 using known gene information (Wang et al., 2005), and optimize marker assisted selection to
508 pyramid multiple genes (Wang et al., 2007).

509

510 **3. Limitations of existing approaches and future work**

511 The use of crop models to accelerate breeding under changing climates is a complex and rapidly
512 evolving area of work, especially with regards to linking crop models and genetic data. At the
513 same time, with the availability and affordability of high throughput phenotyping and
514 genotyping technologies, most breeding programs are undergoing major transformations in the
515 way they operate, most notably through the incorporation of genomic selection and modeling.
516 The CGIAR is no exception to these transformations, as shown by the establishment of the
517 Excellence in Breeding Platform and the Crops to End Hunger Initiative¹, and the existing
518 research on genomic selection (Sect. 2.4.1). Under these initiatives, breeding programs are
519 expected to become more focused and impactful, with clearly set product profiles that clearly
520 outline geographic, farmer and consumer, as well as trait priorities. Hence, it is in the context of
521 these transformations that crop modeling needs to operate, in an effective, flexible and agile

¹ The CGIAR Crops to End Hunger Initiative (CtEH Initiative) seeks to improve and modernize CGIAR crop breeding programs, moving toward using improved breeding approaches. See document of the 8th CGIAR System Council meeting here <https://storage.googleapis.com/cgiarorg/2019/04/SC8-08-CtEH-Module.pdf>.

522 way, to provide crop improvement teams with tools and information that can help them make
523 informed decisions.

524

525 An emerging result from the review of Sect. 2 is that there is no common protocol or approach
526 in the CGIAR to inform breeding programs. This is in part due to the diversity of methods and
527 approaches used, but also due to the lack of collaboration platforms for crop modelers, as well
528 as between modelers and crop improvement teams. In addition, varying degrees of leadership
529 by the CGIAR and coordination between CGIAR Centers also exists with respect to the
530 integration of modeling into breeding programs. As a result, crop modeling activities have little
531 perceived and actual impact on breeding decisions and the breeding process itself. We highlight
532 four suggestions for targeted joint work across the modeling and breeding communities.

533 **(i) Actively take part in the transformation of the breeding programs.** Many CGIAR modeling
534 studies, especially those focused on ideotype design, fail to capture the range of traits
535 relevant in crop improvement, the range of model outputs and spatial and temporal scales
536 that would be useful to them, and the parts of the breeding process that need to be
537 informed (see Sect. 2.3). As breeding programs become more modern (Voss-Fels et al.,
538 2019c), crop modelers need to be an active part of crop improvement, ensuring crop
539 improvement teams are truly multidisciplinary, including crop physiology and modeling,
540 quantitative genetics, genomic prediction and breeding. Given its potential to enhance
541 breeding gains (Messina et al., 2018; Voss-Fels et al., 2019a), a critical part of this endeavor
542 will be the integration of crop modeling with genomic selection (Sect. 2.4.1), and the use of
543 crop models to map and stratify stress variation and response in the target breeding area
544 (Sect. 2.2). As has been demonstrated by some private sector breeding programs (e.g.
545 Cooper et al. 2014), if these tools are integrated to enable the definition and
546 implementation of breeding products, the impact of the breeding programs can be
547 maximized.

548 **(ii) Move towards simpler models that ably simulate key traits and their responses across**
549 **environments and management conditions.** In the last decade, most model improvements
550 have been relatively slow (compared to the rate of knowledge generation), limited by data

551 availability, typically incremental (i.e. without thinking out of the box), and focused on a
552 small range of crops (Challinor et al., 2014; Maiorano et al., 2017). At the same time,
553 because crop models are increasingly being used beyond their original design purposes,
554 they have also tended to become overly complex. Furthermore, as summarized by Rotter et
555 al. (2011) and Challinor et al. (2018), major limitations exist in process-based crop models
556 regarding the processes they consider, the accuracy and precision with which they do so,
557 and the true significance of their parameters. New models need to be designed that
558 specifically incorporate those traits that are of importance to CGIAR breeding programs and
559 crops, as well as their response to key stresses and their interactions, considering the
560 proper balance between parsimony, and biological relevance (Hammer et al., 2019).
561 Leapfrog changes in crop modeling frameworks, such as those proposed by Droutsas et al.
562 (2019) and Soltani and Sinclair (2011, 2012), offer promise in creating models that can be
563 more effectively and rapidly improved to support the breeding process (e.g. by adding new
564 processes and/or traits, or by connecting them with genetic or phenotypic data) [e.g.
565 Messina et al. (2015)]. A documented portfolio of models will allow selection of best-bet
566 models on a case-by-case basis.

567 **(iii) Modernize data storage and interoperability.** Collaboration across researchers in crop
568 modeling in global or regional projects, including the Agricultural Model Inter-comparison
569 Project (AgMIP), has helped the crop modeling community to identify high-value datasets
570 (Asseng et al., 2015; Raymundo et al., 2018), resulting in improved models with greater
571 applicability for breeding under future climates, for example for heat stress response on
572 wheat (Asseng et al., 2013, 2014, 2019b), or CO₂ response on maize (Durand et al., 2018).
573 As breeding programs become more data-driven (e.g. through the application of genomic
574 selection), joint efforts between the modeling and breeding communities will help develop
575 and deploy common standards and inter-connected data storage, translation, transfer, and
576 use platforms that enable the seamless integration of crop modeling into breeding
577 methods.

578 **(iv) Fully take advantage of phenotyping and breeding data for modeling key traits.** Lack of
579 appropriate documentation and benchmarking and extensive model evaluation across

580 target breeding environments implies that the range of model capabilities is generally
581 poorly understood within the modeling community itself (Ramirez-Villegas et al., 2015;
582 Challinor et al., 2018), and even less so by the breeding community. Testing models against
583 experimental data will generate closer links between crop model parameter sets and
584 specific crop varieties, and enable faster and more targeted model development and
585 improvement.

586

587 **4. Conclusions**

588 We have reviewed the use of crop models in support of accelerated breeding, with a particular
589 focus on the CGIAR. Crop modeling can support breeding efforts in many ways, including
590 assessing genotypic adaptability and stability, characterizing and identifying target breeding
591 environments, identifying traits and/or eco-physiological characteristics that maximize yield for
592 such environments, and making predictions about the breeding value of the genotypes. Crop
593 modeling science, especially within the CGIAR, has contributed to all of these, with clear
594 strengths around knowledge generation on eco-physiology, the translation of such knowledge
595 into crop model development and evaluation, and the assessment of G×E×M interactions.
596 However, much progress remains to be made if crop modeling is to effectively contribute to the
597 accelerated breeding rates required to adapt to climate change (see Sect. 1.2).

598

599 In a decade in which major CGIAR system breeding program transformations are expected, crop
600 modelers will need to be part of crop improvement teams, with a common understanding of
601 breeding pipelines and model capabilities and limitations, and common data and protocols,
602 ensuring they follow and deliver according to common and clearly defined breeding products.
603 Doing so will imply more rapid and better targeted crop model improvement activities, and
604 ‘thinking out of the model box’ to create novel approaches that capitalize on the availability of
605 genetic data, thus ultimately allowing the use of the knowledge embedded in current models to
606 effectively address breeding program questions. Standard tests of crop model skill, whilst
607 requiring perhaps a little courage on the part of modelers, will ultimately be of great service to
608 the modelling and breeding communities, as well as those who use the results of their work.

609

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623

624 **Conflict of interest**

625 The authors declare no conflict of interest

626

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1257 **Figure captions**

1258

1259 **Figure 1** Average simulated future potential benefits from genotypic adaptation (including ideotype
1260 design) as derived from 19 modelling studies for wheat (n=15 simulations), sorghum (n=4), pearl millet
1261 (n=48), groundnut (n=12), chickpea (n=48), rice (n=159), maize (n=19), and barley (n=48). The number of
1262 data points used to compute means and error bars follows the number of studies, and the number of
1263 sites, varieties, and scenarios reported in each study. The height of the bar shows the mean of all
1264 reported simulations for each crop, and error bars extend 5–95 % of the data.

1265

1266 **Figure 2** Three major CGIAR examples of environmental characterization to support breeding. (A)
1267 Drought stress patterns for rice in central Brazil (Ramirez-Villegas et al., 2018); (B) drought stress
1268 patterns for post-rainy sorghum in India (Kholová et al., 2013); and (C) map of maize breeding mega-
1269 environments from CIMMYT (Cairns et al., 2013). Panels A and B are redrawn from the original studies,
1270 and data from C was provided by CIMMYT.

1271

1272 **Figure 3** Response to rapid GS cycling for grain yield from the rapid cycling recombination genomic
1273 selection for four cycles (C1, C2, C3, and C4). Colored dots indicate means of the checks (red) and of the
1274 entries (blue). Figure taken from Zhang et al. (2017).

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Table 1 Key CGIAR model-based ideotype design studies

Crop	Region	Model	Proposed ideotype / trait change	Reference
Chickpea	South Asia and East Africa	CROPGRO (DSSAT)	<ul style="list-style-type: none"> – Increased maximum leaf photosynthesis rate, partitioning of daily growth to pods and seed-filling duration. – Drought and heat tolerance: greater rooting density, water extraction capacity, and lower sensitivity for seed-set, individual seed growth, and partitioning (depending on location) 	Singh et al. (2014a)
Peanut	India	CROPGRO (DSSAT)	Longer maturity	Singh et al. (2012)
Peanut	India and West Africa	CROPGRO (DSSAT)	<ul style="list-style-type: none"> – Increased crop maturity, leaf photosynthesis, partitioning to seeds, and seed filling duration – Greater heat and drought (root traits) tolerance 	Singh et al. (2014b)
Peanut	India	GLAM	Increasing maximum photosynthetic rates, total assimilate partitioned to seeds, and, where enough soil moisture is available, also maximum transpiration rates	Ramirez-Villegas and Challinor (2016)
Lentil	East Africa	SSM	<ul style="list-style-type: none"> – Shorter cycle of lentil – Limited transpiration rates under high vapor pressure deficit 	Ghanem et al. (2015)
Lentil	South Asia	SSM	<ul style="list-style-type: none"> – Shorter cycle of lentil – Limited transpiration rates under high vapor pressure deficit 	Guiguitant et al. (2017)
Sorghum	India and West Africa	CERES-Sorghum	<ul style="list-style-type: none"> – Increased crop maturity, radiation use efficiency, relative leaf size and partitioning of assimilates to the panicle. – Greater heat (lower sensitivity of reproductive processes) and drought (root traits) tolerance 	Singh et al. (2014c)
Sorghum	India	APSIM	Limited transpiration rates under high vapor pressure deficit, especially combined with enhanced water extraction capacity at the root level. Smaller canopy size, later plant vigor or increased leaf appearance rate.	Kholová et al. (2014)
Pearl millet	India and West Africa	CERES-Pearl millet	<ul style="list-style-type: none"> – Increased crop duration and yield potential traits (photosynthesis, partitioning) – Drought and heat tolerance in arid and semi-arid hot tropical climates. 	Singh et al. (2017)
Rice	Africa	ORYZA2000	Greater crop duration and increased maximum photosynthetic rate at high temperatures	van Oort and Zwart (2018)
Rice	South Asia	ORYZA2000	Deeper roots (from 45 to 50 cm) to reduce plant sensitivity to drought. Drought onset occurs 3 weeks after transplanting.	Mottaleb et al. (2017)
Rice	Philippines	ORYZA2000	Greater duration and tolerance to extreme temperatures	Li and Wassman (2010)

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1281 **Table 2** List of statistical approaches that incorporate environmental information into genomic

1282 prediction models

	Method description	Reference(s)
i	Prediction model integrating pedigree based additive genetic covariances between relatives and G×E interactions	Crossa et al. (2006)
ii	Multi-environment prediction framework for modeling G×E interactions using pedigree and genomic information	Burgueño et al. (2012)
iii	Reaction norm model for incorporating the main and interaction effects of high-dimensional markers and environmental covariates	Jarquín et al. (2014)
iv	Threshold models incorporating G×E and additive × additive × environment (G×G×E) interactions for predicting ordinal categorical traits	Montesinos-Lopez et al. (2015)
v	Bayesian mixed-negative binomial genomic regression model for count data that integrates G×E interactions	Montesinos-Lopez et al. (2016a)
vi	Marker × environment interaction (Mk×E) genomic model for predicting non-phenotyped individuals and identifying genomic regions associated with yield stability and environmental specificity	Crossa et al. (2016b)
vii	Models integrating genomic, pedigree and environmental covariates for predicting grain yield in different agro-ecological zones	Saint Pierre et al. (2016)
viii	G×E interaction kernel regression models using nonlinear Gaussian kernels for modelling marker main effects and marker-specific interaction effects	Cuevas et al. (2017)
ix	Single-step approach incorporating genomic, pedigree and G×E interaction information for predicting wheat lines in South Asia	Perez-Rodriguez et al. (2017)
x	Pedigree-based reaction norm model incorporating G×E interactions for multi-environment trial data	Sukumaran et al. (2017),
xi	Bayesian approach and a recommender systems approach for predicting multiple traits evaluated in multiple environments	Montesinos-Lopez et al. (2016b, 2018b)
xii	G×E interaction model in durum wheat evaluated using three cross-validation (CV) schemes for predicting incomplete field trials (CV2), new lines (CV1), and lines in untested environments (CV0)	Sukumaran et al. (2018); Roorkiwal et al. (2018)

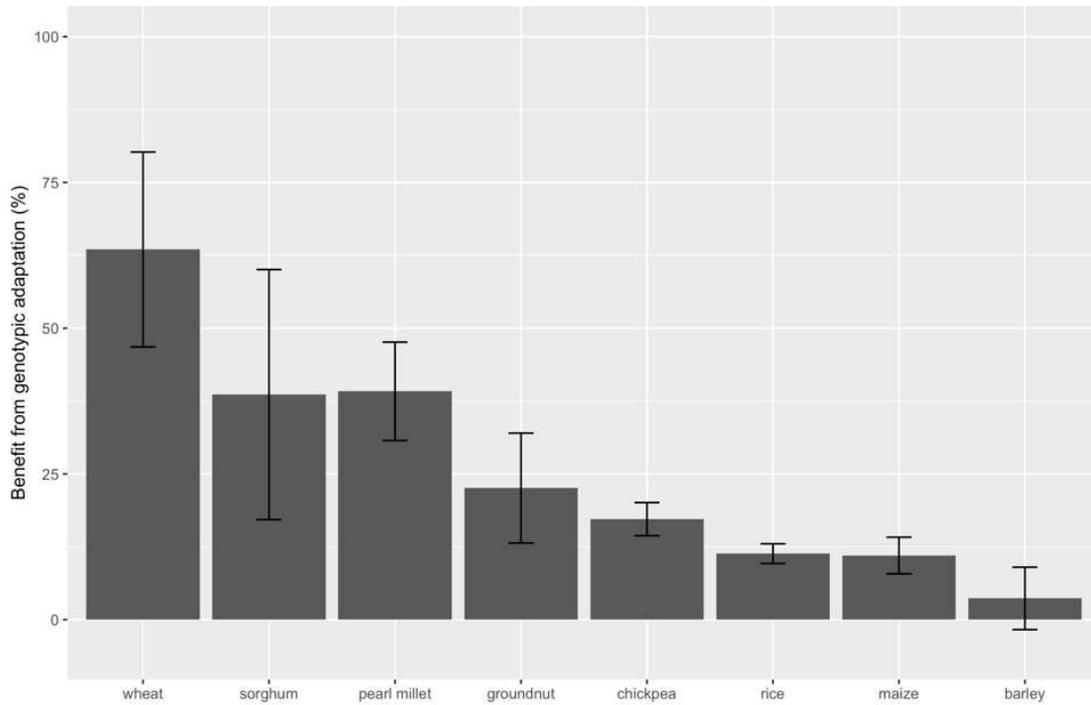
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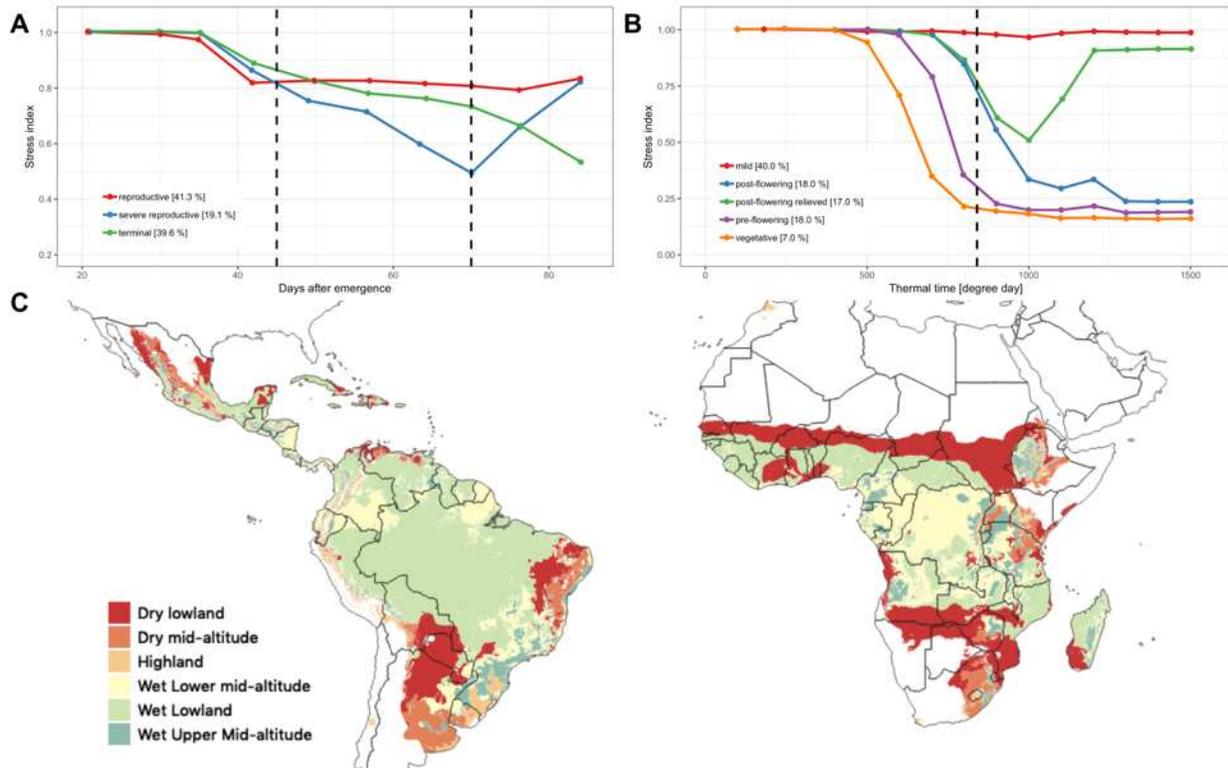
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Figure 1 Average simulated future potential benefits from genotypic adaptation (including ideotype design) as derived from 19 modelling studies for wheat (n=15 simulations), sorghum (n=4), pearl millet (n=48), groundnut (n=12), chickpea (n=48), rice (n=159), maize (n=19), and barley (n=48). The number of data points used to compute means and error bars follows the number of studies, and the number of sites, varieties, and scenarios reported in each study. The height of the bar shows the mean of all reported simulations for each crop, and error bars extend 5–95 % of the data.

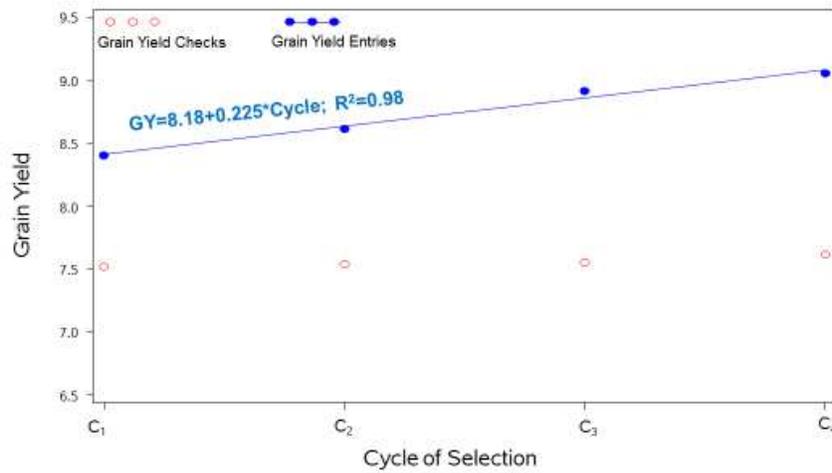
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Figure 2 Three major CGIAR examples of environmental characterization to support breeding. (A) Drought stress patterns for rice in central Brazil (Ramirez-Villegas et al., 2018); (B) drought stress patterns for post-rainy sorghum in India (Kholová et al., 2013); and (C) map of maize breeding mega-environments from CIMMYT (Cairns et al., 2013). Panels A and B are redrawn from the original studies, and data from C was provided by CIMMYT.

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Figure 3 Response to rapid GS cycling for grain yield from the rapid cycling recombination genomic selection for four cycles (C₁, C₂, C₃, and C₄). Colored dots indicate means of the checks (red) and of the entries (blue). Figure taken from Zhang et al. (2017).