



This is a repository copy of *Effects of electronic cigarette liquid on monolayer and 3D tissue-engineered models of human gingival mucosa*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/157441/>

Version: Published Version

Article:

Shaikh, Z.N., Alqahtani, A., Almela, T. et al. (3 more authors) (2019) Effects of electronic cigarette liquid on monolayer and 3D tissue-engineered models of human gingival mucosa. *Journal of Advanced Periodontology & Implant Dentistry*, 11 (2). pp. 54-62.

10.15171/japid.2019.010

Reuse

This article is distributed under the terms of the Creative Commons Attribution (CC BY) licence. This licence allows you to distribute, remix, tweak, and build upon the work, even commercially, as long as you credit the authors for the original work. More information and the full terms of the licence here:
<https://creativecommons.org/licenses/>

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Effects of electronic cigarette liquid on monolayer and 3D tissue-engineered models of human gingival mucosa

Zahab N Shaikh¹ • Abdullah Alqahtani¹ • Thafar Almela¹ • Kirsty Franklin¹ • Lobat Tayebi² • Keyvan Moharamzadeh^{1, 2*}

¹School of Clinical Dentistry, University of Sheffield, Claremont Crescent, Sheffield, S10 2TA, UK

²Marquette University School of Dentistry, Milwaukee, WI 53233, USA

*Corresponding Author; E-mail: k.moharamzadeh@sheffield.ac.uk

Received: 24 October 2019; Accepted: 25 December 2019

J Adv Periodontal Implant Dent 2019;11(2):54-62 | doi:10.15171/japid.2019.010

This article is available from <https://japid.tbzmed.ac.ir/>

© 2019 The Authors. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Background. There is limited data available on potential biological effects of E-cigarettes on human oral tissues. The aim of this study was to evaluate the effects of E-cigarette liquid on the proliferation of normal and cancerous monolayer and 3D models of human oral mucosa and oral wound healing after short-term and medium-term exposure.

Methods. Normal human oral fibroblasts (NOF), immortalized OKF6-TERET-2 human oral keratinocytes, and cancerous TR146 keratinocyte monolayer cultures and 3D tissue engineered oral mucosal models were exposed to different concentrations (0.1%, 1%, 5% and 10%) of E-cigarette liquid (12 mg/ml nicotine) for 1 hour daily for three days and for 7 days. Tissue viability was monitored using the PrestoBlue assay. Wounds were also produced in the middle surface of the monolayer systems vertically using a disposable cell scraper. The alterations in the cell morphology and wound healing were visualized using light microscopy and histological examination.

Results. Statistical analysis showed medium-term exposure of TR146 keratinocytes to 5% and 10% E-liquid concentrations significantly increased the viability of the cancer cells compared to the negative control. Short-term exposure of NOFs to 10% E-liquid significantly reduced the cell viability, whereas medium-term exposure to all E-liquid concentrations significantly reduced the NOF cells' viability. OKF6 cells exhibited significantly lower viability following short-term and medium-term exposure to all E-cigarette concentrations compared to the negative control. 3D oral mucosal model containing normal oral fibroblasts and keratinocytes showed significant reduction in tissue viability after exposure to 10% E-liquid, whereas medium-term exposure resulted in significantly lower viability in 5% and 10% concentration groups compared to the negative control. There was a statistically significant difference in wound healing times of both NOF and OKF6 cells after exposure to 1%, 5% and 10% E-cigarette liquid.

Conclusion. Medium-term exposure to high concentrations of the E-cigarette liquid had cytotoxic effects on normal human oral fibroblasts and OKF6 keratinocytes, but a stimulatory cumulative effect on the growth of cancerous TR146 keratinocyte cells as assessed by the PrestoBlue assay and histological evaluation of 3D oral mucosal models. In addition, E-liquid exposure prolonged the wound healing of NOF and OKF6 oral mucosa cells.

Key words: Electronic cigarette, cytotoxicity, keratinocyte, fibroblast, tissue engineering, wound healing.

Introduction

Electronic cigarette (EC) use has rapidly increased since they were introduced. There are significant debates in healthcare and regulatory bodies on this subject, and conflicting advice has been given regarding the appropriate use of ECs.

Data from an international survey showed 15% use in the US, 10% use in the UK, 4% use in Canada, and 2% use in Australia and the number of young people trying the ECs is significantly increasing.¹ The rate of EC use by middle and high school students in the US has tripled from 2011 to 2013. A three-fold increase in EC use was reported from 2010 to 2012 in the Great Britain.²

There is a growing body of evidence that demonstrates ECs are an effective aid to smoking cessation, and they have significant potential to reduce tobacco smoke-related oral diseases.³⁻⁵ However, it is important to balance the potential adverse effects of ECs aerosol with the benefits of reducing tobacco smoking.

Several chemical studies have analyzed the composition of liquids and aerosols from ECs.⁶⁻¹² Several potentially harmful materials have been identified in ECs, including diethylene glycol, propylene glycol, nitrosamines, toluene, acrolein, acetaldehyde, formaldehyde, nickel, lead, aluminum, chromium, cadmium, and silicon. Some ingredients, such as Diacetyl, used in flavored e-liquids are safe to consume orally, though perhaps not to inhale, as it has been associated with bronchiolitis obliterans.

Although a few animal studies have failed to show any obvious adverse systemic toxicity associated with inhaled glycerol or propylene glycol,¹³⁻¹⁴ concerns have been raised for humans in terms of respiratory tract irritation,¹⁵ and evidence for potential localized adverse effects of inhaled glycerin is sparse. An animal study has shown evidence of mild epithelial squamous metaplasia following long-term inhalation of aerosolized glycerol in rats.¹⁶

It has been demonstrated that exposure to ECs impairs pulmonary antibacterial and antiviral defenses in a mouse model.¹⁷ In vitro toxicity profiles of electronic and tobacco cigarettes, smokeless tobacco, and nicotine replacement therapy products have been compared in a study using Ames Salmonella mutagenicity testing.¹⁸

The nicotine in the ECs is mainly absorbed through the buccal mucosa and pharyngeal mucosa. Although nicotine has been shown to have wound healing and angiogenic properties,¹⁹ which can make it a potential therapeutic agent, there are some concerns regarding

its ability to promote lung tumor growth through different possible mechanisms, including angiogenesis, cell proliferation, and cell migration.²⁰⁻²¹

There are few representative studies that illustrate the negative effects of ECs on the human oral mucosa. Evidence supports that conventional smoking has a negative influence on a patient's response, as well as their treatment outcome, following non-surgical, surgical, and regenerative periodontal treatment. Smoking also impairs all wound healing phases (particularly the inflammatory and proliferation phases, which lead to delays in wound healing).²²⁻²⁶

Clinical studies of ECs have reported mild harmful effects of vaping on selected cardiovascular²⁷⁻²⁸ and respiratory functional outcomes²⁹⁻³¹ to a considerably less extent when compared with conventional cigarette smoking³². However, it is difficult to assess the prognostic implications of these studies, and there is a need for further research in this area.

Clinical research and survey-type studies to date have shown that the most common patient-reported side effects were symptoms such as xerostomia, throat irritation, and cough.³³⁻³⁶

A five-year multicenter cohort study was conducted to assess the long-term efficacy and safety of ECs,³⁷ and a pilot study investigated the oral mucosa perfusion of intraoral free flaps in EC users and smokers.³⁸

Several toxicological studies have been conducted on ECs, using monolayer cell culture systems.³⁹⁻⁴³ However, there is no study published in the literature comparing the effects of E-cigarette liquid on different normal and cancerous oral mucosa cells, including 3D tissue-engineered models of the human oral mucosa. Therefore, this study aimed to evaluate the biological effects of E-cigarette liquid on both full-thickness 3D oral mucosa models and monolayer cell culture systems utilizing normal oral fibroblasts, OKF6-TERT2 oral keratinocytes, and cancerous TR146 keratinocytes, following short-term (three days) and medium-term (seven days) exposure to flavorless electronic cigarette liquid. Additionally, this study aimed to investigate the influence of E-cigarette liquid on oral mucosa wound healing, using normal oral fibroblasts and OKF6 oral keratinocyte cell cultures.

Methods

Cell Source and Biological Systems

Normal human oral fibroblast cells were obtained from the stocks stored in liquid nitrogen in the laboratories of the School of Clinical Dentistry at the University of Sheffield. These cells had been previously obtained from healthy patients undergoing oral

surgery at Charles Clifford Dental Hospital with their written informed consent under appropriate ethical approval from the UK National Research Ethics Services Committee.

The immortalized OKF6/TERT-2 human oral keratinocyte cell line was kindly provided by Brigham and Women's Hospital, Harvard Institute of Medicine, USA.

The cancerous human TR146 cell line was derived from a neck lymph node metastasis originating from a carcinoma of the oral buccal mucosa. These cells were kindly provided by Cancer Research UK.

Five different biological systems were tested in this study, including:

- Monolayer cultures of normal oral fibroblasts (NOF)
- Monolayer cultures of the immortalized oral keratinocyte cell line (OKF6/TERT-2)
- Monolayer cultures of cancerous TR146 keratinocytes
- Co-cultures of NOF and OKF6/TERT-2 cells
- 3D tissue-engineered oral mucosa models using TR146 keratinocytes and NOFs

Cell Culture

Oral fibroblasts and keratinocytes were cultured in high-glucose Dulbecco's modified Eagle's medium (DMEM) (Sigma, UK), supplemented with 2% L-glutamine (Sigma, UK), 100 IU:100 mg ml⁻¹ Penicillin/Streptomycin (Sigma, UK), and 10% fetal calf serum (FCS) (Biowest Ltd., UK). Six-well plates were utilized for the monolayer cell cultures, inoculating 10,000 cells per well for NOFs, OKF6/TERT2 cells, and TR146 keratinocytes. Additionally, normal oral fibroblasts and keratinocytes were cultured together using six-well plates.

The cultures were maintained in incubators at 5% CO₂ at 37°C. The cells were cultured until 80–100% confluency was achieved.

Tissue-engineered 3D Oral Mucosa Models

Full-thickness 3D tissue-engineered oral mucosa models were manufactured by air/liquid interface culture of TR146 keratinocytes seeded onto fibroblast-populated collagen gels. A solution of 10 × DMEM, 8.5% (v/v) FBS, 2 mM L-glutamine, reconstitution buffer (22 mg mL⁻¹ sodium bicarbonate and 20 mM 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), and 5 mg mL⁻¹ rat tail type I collagen (R & D system, UK) was prepared and neutralized by 1-M sodium hydroxide to pH=7.4 in an ice-cold environment by keeping everything on ice. Normal oral fibroblasts were added to the solution at

a concentration of 500,000 cells/model, and 1 mL of the resultant cell-containing collagen mixture was transferred to cell culture transwell inserts (0.4 μm pore size, Millipore), incubated at 37°C for 2 hours until solidified, and then completely submerged in complete DMEM for 3 days. Subsequently, 1 × 10⁶ keratinocytes were seeded onto each model and kept in submerged culture for three days, after which the oral mucosal models were raised to air/liquid interface and cultured for a further 7 days.

Exposure Protocol

Neutral E-Cigarette Liquid (Vype-UK) with no added flavors, containing medium nicotine level of 12 mg/mL, was used in this experiment. Four different concentrations (0.1%, 1%, 5%, and 10%) of E-cigarette liquid were prepared by diluting the E-liquid with DMEM.

The monolayer cell culture systems and the 3D oral mucosa model's epithelial surfaces were exposed to four different concentrations of E-cigarette liquid (0.1%, 1%, 5%, and 10%), to the negative control (DMEM) and the positive control (70% ethanol). The groups consisted of six samples each (n=6).

All the samples were exposed to their respective reagents for one hour daily for three continuous days and incubated at 5% CO₂ and 37°C. Following the testing for the short-term exposure, the samples were washed twice with phosphate-buffered saline (PBS) (Sigma, UK), and the medium-term exposure continued for four more days by exposing the samples to the reagents for one hour daily, totaling seven days.

Cell Viability Assay

Following the short-term and medium-term exposures, tissue viability test was carried out using the PrestoBlue assay. The PrestoBlue reagent (Biosource, Camarillo, CA) was added to the samples at a ratio of 9:1 (volume of cells and culture medium: volume of PrestoBlue reagent). The plates were then incubated for 60 minutes at 37°C and 5% CO₂. Following incubation, triplicate 200-μL samples were placed into the wells of a 96-well plate, and the fluorescence intensity of each well was measured at an excitation wavelength of 530 nm and an emission wavelength of 590 nm using a fluorescent plate reader (Infinite 200 PRO TECAN, Switzerland).

Histological and Morphological Assessment

The oral mucosa model samples were first fixed in 10% formalin solution for 24 hours; the samples were then mounted in Optimal Cutting Temperature (OCT) embedding compound, followed by freezing at -20 to -

80°C. Afterward, the samples were sectioned at a thickness of 10–30 μm using a cryostat machine. The sections were then mounted on the histological slides. This was followed by drying the slides for around 30 minutes at room temperature. The slides were then stored in the freezer at -80°C until the processing for hematoxylin and eosin (H&E) staining.

The slides were examined under a light microscope by more than one histopathology expert to assess the changes in the connective tissue and the epithelial layers of the model following the exposure to test agents. Assessment criteria included the continuity and thickness of the epithelium, cell morphology, presence or absence of pyknotic nuclei, presence of a distinct interface between the epithelium and the connective tissue layer.

Wound Healing Assay

Monolayer cultures of normal oral fibroblasts and OKF6/TERT-2 keratinocytes were developed in 6-well tissue culture plates and divided into negative control (DMEM), positive control (70% ethanol), and test groups with various E-Cigarette concentrations (0.1%, 1%, 5%, and 10%) (n=6). The wounds were produced vertically in the middle of the surface of the monolayer systems using a disposable cell scraper. The test groups were exposed to the culture media containing E-cigarette liquid immediately before and continued daily after creating a wound, and then the wounds were monitored until complete healing had occurred. Microscopic images were obtained pre- and post-wound creation daily to assess the healing time in all the groups.

Statistical Analysis

SPSS 20 was used for statistical analysis. The normality of the data was analyzed using the normality test (Shapiro-Wilk test). Means of the samples were compared with ANOVA, followed by multiple comparisons using post hoc Tukey tests to determine the differences between the different groups. The level of significance for all the statistical tests was set at $\alpha=0.05$.

Results

The results of the PrestoBlue assay for normal oral fibroblasts and OKF6/TERT-2 keratinocyte monolayer cell cultures exposed to different concentrations of the E-Liquid for three days and seven days are shown in Figure 1. Short-term exposure to 10% E-liquid solution caused a statistically significant reduction in the viability of NOFs ($P<0.0001$) compared to the negative control group. Following medium-term exposure,

all the E-liquid concentration groups exhibited significantly lower viability compared to the negative control group.

OKF6/TERT-2 keratinocyte monolayers showed significantly lower viability after short-term exposure to all the E-liquid concentrations, whereas medium-term exposure resulted in significantly lower viability in all the groups except for 1% concentration group compared to the negative control group.

The TR146 keratinocyte monolayers did not exhibit any statistically significant difference in the viability between different E-liquid concentration groups compared to the negative control group after short-term exposure. However, medium-term exposure to 5% and 10% E-liquid solution caused a statistically significant increase in the viability of TR146 keratinocytes compared to the negative control group (Figure 2).

Normal oral fibroblast and OKF6/TERT-2 keratinocyte co-culture system showed a significant reduction in cell viability in 10% E-liquid concentration group following short-term exposure. In contrast, medium-

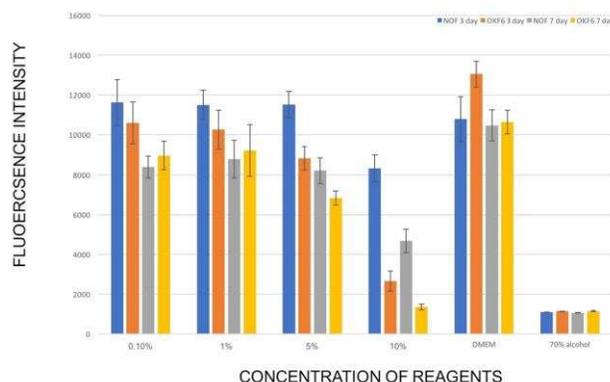


Figure 1. Tissue viability of normal oral fibroblasts and OKF6 keratinocyte monolayer cell cultures exposed to different concentrations of E-liquid as assessed by the PrestoBlue assay.

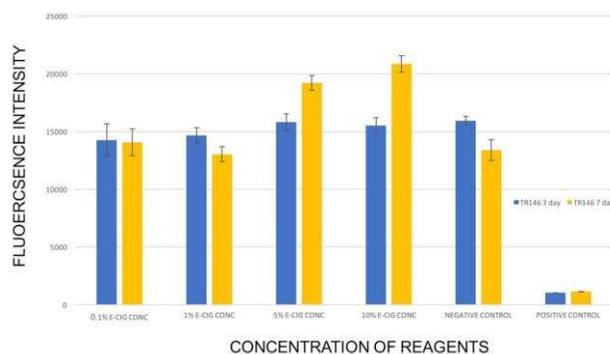


Figure 2. Tissue viability of TR146 keratinocyte monolayer cell cultures exposed to different concentrations of E-liquid as assessed by the PrestoBlue assay.

term exposure resulted in significantly lower viability in 5% and 10% concentration groups ($P < 0.0001$) compared to the negative control group (Figure 3). Histologically, the 3D oral mucosal models utilizing the NOFs and TR146 keratinocytes showed an increase in the thickness of the cancerous epithelial layer in high E-liquid concentration groups compared to the negative control after short-term and medium-term exposure (Figures 4 and 5).

Figure 6 demonstrates microscopic views of the wound healing at different stages for normal oral fibroblasts and OKF6/TERT-2 keratinocytes. Table 1 presents the mean values and standard deviations of the total time of wound healing for the control and test groups. There was a statistically significant difference in the wound healing time of both NOF and OKF6/TERT-2 monolayer systems exposed to 1%, 5%, and 10% E-liquid solutions compared to those of the negative control group ($P < 0.05$).

Discussion

Three different cell types, including NOFs, OKF6/TERT2 keratinocytes, and cancerous TR146 keratinocytes, were used in this study. Normal oral fibroblasts were selected as they are thought to play a major role in mucosal wound healing; they are also responsible for extracellular matrix synthesis in the connective tissue layer. OKF6/TERT-2 keratinocytes exhibit high reproducibility, avoid batch-to-batch variations; they were selected to represent normal oral epithelial cells. Conversely, squamous cell carcinoma-derived TR146 cells were utilized to assess the effects of E-liquid on oral cancer cells.

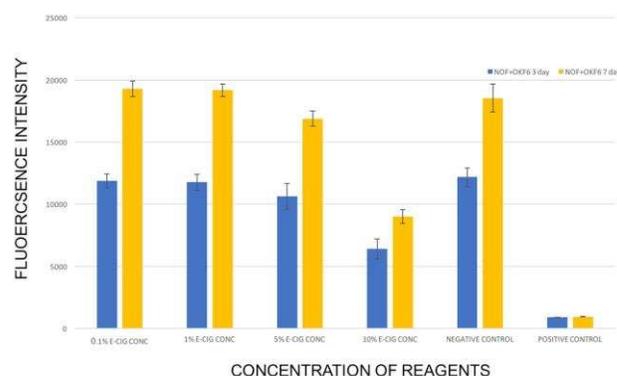


Figure 3. Tissue viability of normal oral fibroblast and OKF6 keratinocyte 3D co-culture systems exposed to different concentrations of E-liquid as assessed by the PrestoBlue assay.

E-liquid with no added flavors was used to eliminate the confounding effects of different additives that are observed with various types of flavoring agents used in E-cigarettes. Different studies have shown that certain flavors, such as menthol, cinnamon, caramel, butterscotch, bubble-gum, and coffee, have more cytotoxic effects on cells compared to some other flavors.³⁹⁻⁴¹

Four different concentrations (0.1%, 1%, 5%, and 10%) of E-liquid solution were prepared to assess the effects of various concentrations of E-liquid on the cells, covering the range from light vapors to heavy e-cigarette users. The duration of exposure included three days and seven days to simulate short-term and medium-term vaping.

In this study, electronic cigarette liquid exhibited an adverse effect on the viability of normal oral

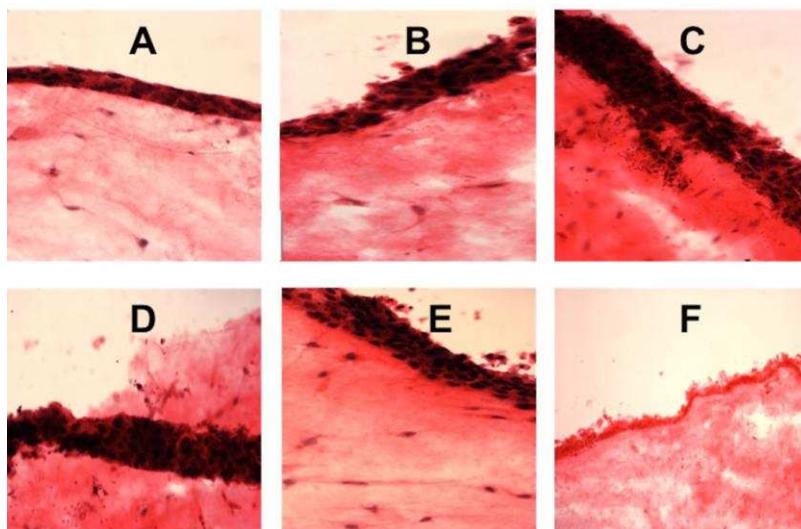


Figure 4. Histological sections of 3D tissue-engineered oral mucosa models after short-term exposure to (A) 0.1% E-liquid; (B) 1% E-liquid; (C) 5% E-liquid; (D) 10% E-liquid; (E) negative control; (F) positive control (H & E staining, original magnification $\times 10$).

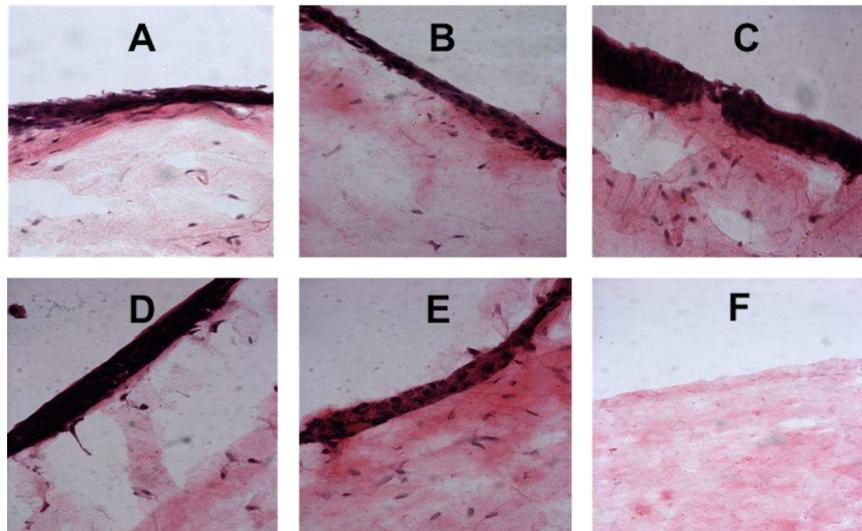


Figure 5. Histological sections of 3D tissue-engineered oral mucosa models after medium-term exposure to (A) 0.1% E-liquid; (B) 1% E-liquid; (C) 5% E-liquid; (D) 10% E-liquid; (E) negative control; (F) positive control (H & E Staining, original magnification $\times 10$).

fibroblasts and OKF6/TERT-2 keratinocytes. These results are consistent with a previous study that tested different types of E-cigarette liquids on the human periodontal ligament fibroblasts and showed a reduction in the viability of cells in the samples that were exposed to E-liquids.³⁹

Conversely, a dose-dependent stimulatory effect of E-liquid on the growth of cancerous TR146 cells was observed in our study, exhibiting increased viability and proliferation of TR146 cells with increasing concentrations of E-liquid. Similarly, histological evaluation of the 3D oral mucosa models showed an increase in the thickness of the cancerous epithelial layer exposed to high concentrations of E-liquid. This is the first study reporting the use of a full-thickness

3D tissue-engineered oral mucosal model for the biological evaluation of electronic cigarettes on cancerous oral tissues. These findings have not been reported previously and might indicate tumor-promoting effects of the ingredients of the E-liquid tested in this study.

Previous studies have raised some concerns regarding the potential effects of nicotine on promoting tumors in the lungs through various possible mechanisms, such as cell migration, proliferation and angiogenesis.²⁰⁻²¹

The influence of nicotine on dysplastic oral keratinocyte cell line and precancerous lesions of the mouse tongue has been investigated previously, showing an inhibitory effect on apoptosis and a

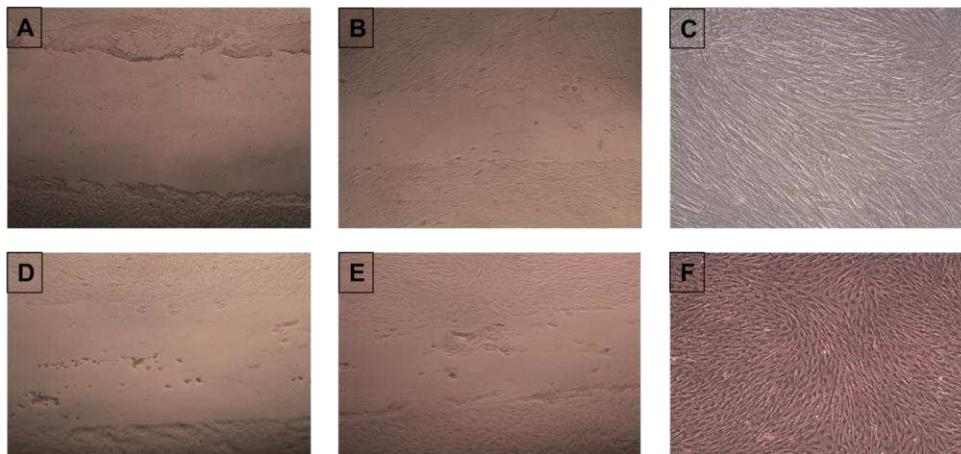


Figure 6. Microscopic views of the wound healing assay showing (A) the center of the wound on day 1 of fibroblast culture; (B) central wound on day 3 of fibroblast culture; (C) completely healed fibroblast cultures; (D) the center of the wound on day 1 of keratinocyte culture; (E) central wound on day 3 of keratinocyte culture; and (F) completely healed keratinocyte cultures.

Table 1. Mean values and standard deviations of the total time of wound healing for the control and test groups

Cells	Groups	Mean (days)	SD
NOF	0.1% E-liquid	6.17	0.75
	1% E-liquid	7.33	0.51
	5% E-liquid	9.167	0.75
	10% E-liquid	12.33	1.03
	Control (DMEM)	5.17	0.41
OKF6/ TERT2	0.1% E-liquid	6.00	0.63
	1% E-liquid	6.67	0.52
	5% E-liquid	7.67	0.52
	10% E-liquid	10.50	0.84
	Control (DMEM)	5.05	0.63

stimulatory effect on the growth of oral precancerous lesions.⁴⁴ Similarly, Chernyavsky et al⁴² assessed the tumor-promoting effects of nicotine on oral and lung cancer cells. Nicotine exhibited resistance to apoptosis, increasing the counts of both lung and oral cancer cells.

Some other studies have used monolayer cell culture systems to assess the cytotoxicity of electronic cigarettes.³⁹⁻⁴³ Different types of cell lines have been exposed to either electronic cigarette aerosols or E-liquid solutions. These studies have also confirmed the adverse effects of E-cigarettes, with some cell lines being more sensitive than the others.

Recently, studies have been conducted to assess the effects of electronic cigarettes on oral mucosal cells. In a study by Yu et al,⁴³ electronic cigarette exposure reduced cell viability along with high levels of apoptosis and necrosis, with alterations in augmented DNA strands, in normal epithelial and squamous cell carcinoma (head and neck) cell lines. However, Guttenplan et al⁴⁴ observed a stimulatory effect of E-cigarette exposure on the proliferation of human oral leukoplakia cells. Conversely, Willershausen et al³⁷ reported a reduction in cell proliferation of periodontal ligament fibroblasts after exposure to various E-liquids.

Sundar et al⁴⁵ utilized human periodontal ligament fibroblasts and a 3D gingival epithelium-only tissue model to assess the effects of electronic cigarettes, reporting an increase in pro-inflammatory cytokines and an elevated oxidative/carbonyl stress resulting in the production of high levels of cyclooxygenase-2 and prostaglandin E2. Although the 3D split-thickness tissue model used in this study was more clinically relevant than monolayer cell culture systems, it lacked the connective tissue component.

A study by Sancilio et al⁴⁶ raised concerns regarding E-cigarette's role in the pathogenesis of oral diseases. Human gingival fibroblasts exposed to E-Liquids showed decreased production of collagen I and increased levels of lactate dehydrogenase.

There is a lack of research on the effects of E-cigarettes on human oral mucosa wound healing. It has been indicated that exposure to E-cigarettes reduces the viability of cells and compromises cell migration.⁴⁶ Therefore, this study aimed to investigate the effects of E-cigarette liquids with different concentrations on oral mucosa wound healing. Two wound healing models were used in this study based on normal oral fibroblasts to represent connective tissue wound healing and OKF6/TERT2 keratinocytes to simulate epithelial wound healing. The results of this in vitro assay were consistent with the results of the cytotoxicity tests and indicated that the E-liquid tested in this study might have potential dose-dependent adverse effects on oral mucosa wound healing, prolonging the healing times both in the epithelial and the connective tissue layers. In a recent study,⁵⁰ human gingival fibroblasts were exposed to three different groups (cigarette smoke condensate, nicotine-free or nicotine-rich electronic cigarette vapor condensates), and the results showed that both cigarette smoke and electronic cigarette vapors affected the proliferation and migration of fibroblasts. Additionally, the cell scratch test revealed delayed wound healing, consistent with the findings of this study.

There were certain limitations in our study; firstly, this was an in-vitro study, and the results may not be extrapolated to an in-vivo situation. Secondly, only E-liquids were tested in this study rather than the E-cigarette aerosols. Ideally, the cytotoxic effects of E-cigarettes should be assessed in both liquid and vapor form, as the E-cigarette vapors come into direct contact with oral mucosa.

Hence, further research is required to overcome these limitations by assessing E-cigarettes of different flavors and various concentrations of nicotine. The liquid, as well as the vapor form of E-cigarettes, can be tested and compared with the conventional cigarette smoke. Furthermore, evaluating the long-term effects of E-cigarettes would further add to our understanding of the biological effects of E-cigarettes on human oral tissues.

Conclusion

This in vitro study revealed that short-term and medium-term exposure to the electronic cigarette liquid had dose-dependent cytotoxic effects on normal human oral fibroblasts and OKF6/TERT-2 oral keratinocytes. However, E-liquid exposure had a cumulative stimulatory effect on the growth of cancerous TR146 keratinocytes using both monolayer and 3D cell culture systems.

The full-thickness 3D tissue-engineered human oral

mucosal model has the potential to be used as a clinically relevant biological test system for the evaluation of electronic cigarettes.

In addition, E-liquid exposure prolonged the wound healing of both normal oral fibroblasts and OKF6/TERT-2 epithelial cells.

Competing Interests

The authors declare no conflict(s) of interest related to the publication of this work.

Authors' Contributions

Conceptualization, Z.S., A.A., T.A., K.F., L.T. and K.M.; Data curation, Z.S. and A.A.; Investigation, Z.S., A.A., T.A., K.F., L.T. and K.M.; Methodology, Z.S., A.A., T.A. and K.F.; Supervision, L.T. and K.M.; Validation, Z.S., and A.A.; Writing—original draft, Z.S. and A.A.; Writing—review and editing, T.A., K.F., L.T. and K.M.

Acknowledgments

The authors would like to acknowledge Cancer Research UK for kindly providing the TR146 cell line and Brigham and Women's Hospital, Harvard Institute of Medicine, for providing the immortalized OKF6/TERT-2 human oral keratinocyte cell line.

Funding

This research received no external funding.

Ethics Approval

This study has been approved by National Research Ethics Service, NRES Committee London—Hampstead. Research Ethics Committee (REC) Reference number: 15/LO/0116; date of approval: 21/01/2015.

References

- Chapman SL, Wu LT. E-cigarette prevalence and correlates of use among adolescents versus adults: a review and comparison. *Journal of psychiatric research*. 2014 Jul 1; 54:43-54.
- Dockrell M, Morrison R, Bauld L, McNeill A. E-cigarettes: prevalence and attitudes in Great Britain. *Nicotine & Tobacco Research*. 2013 Oct 1;15(10):1737-44.
- Bullen C, McRobbie H, Thornley S, Glover M, Lin R, Laugesen M. Effect of an electronic nicotine delivery device (e cigarette) on desire to smoke and withdrawal, user preferences and nicotine delivery: randomised cross-over trial. *Tobacco control*. 2010 Apr 1;19(2):98-103.
- Bullen C, Howe C, Laugesen M, McRobbie H, Parag V, Williman J, Walker N. Electronic cigarettes for smoking cessation: a randomised controlled trial. *The Lancet*. 2013 Nov 16;382(9905):1629-37.
- Caponnetto P, Auditore R, Russo C, Cappello G, Polosa R. Impact of an electronic cigarette on smoking reduction and cessation in schizophrenic smokers: a prospective 12-month pilot study. *International journal of environmental research and public health*. 2013 Feb;10(2):446-61.
- Hadwiger ME, Trehy ML, Ye W, Moore T, Allgire J, Westerberger B. Identification of amino-tadalafil and rimonabant in electronic cigarette products using high pressure liquid chromatography with diode array and tandem mass spectrometric detection. *Journal of chromatography A*. 2010 Nov 26;1217(48):7547-55.
- Pellegrino RM, Tinghino B, Mangiaracina G, Marani A, Vitali M, Protano C, Osborn JF, Cattaruzza MS. Electronic cigarettes: an evaluation of exposure to chemicals and fine particulate matter (PM). *Ann Ig*. 2012 Jul;24(4):279-88.
- Kim, H.J. and Shin, H.S., 2013. Determination of tobacco-specific nitrosamines in replacement liquids of electronic cigarettes by liquid chromatography–tandem mass spectrometry. *Journal of Chromatography A*, 1291, pp.48-55.
- Etter JF, Zäther E, Svensson S. Analysis of refill liquids for electronic cigarettes. *Addiction*. 2013 Sep;108(9):1671-9.
- Goniewicz ML, Knysak J, Gawron M, Kosmider L, Sobczak A, Kurek J, Prokopowicz A, Jablonska-Czapla M, Rosik-Dulewska C, Havel C, Jacob P. Levels of selected carcinogens and toxicants in vapour from electronic cigarettes. *Tobacco control*. 2014 Mar 1;23(2):133-9.
- Jensen RP, Luo W, Pankow JF, Strongin RM, Peyton DH. Hidden formaldehyde in e-cigarette aerosols. *New England Journal of Medicine*. 2015 Jan 22;372(4):392-4.
- Williams M, Villarreal A, Bozhilov K, Lin S, Talbot P. Metal and silicate particles including nanoparticles are present in electronic cigarette cartomizer fluid and aerosol. *PloS one*. 2013 Mar 20;8(3):e57987.
- Robertson OH, Loosli CG, Puck TT, Wise H, Lemon HM, Lester W. Tests for the chronic toxicity of propylene glycol and triethylene glycol on monkeys and rats by vapor inhalation and oral administration. *Journal of Pharmacology and Experimental Therapeutics*. 1947 Sep 1;91(1):52-76.
- Werley MS, McDonald P, Lilly P, Kirkpatrick D, Wallery J, Byron P, Venitz J. Non-clinical safety and pharmacokinetic evaluations of propylene glycol aerosol in Sprague-Dawley rats and Beagle dogs. *Toxicology*. 2011 Sep 5;287(1-3):76-90.
- Wieslander G, Norbäck D, Lindgren T. Experimental exposure to propylene glycol mist in aviation emergency training: acute ocular and respiratory effects. *Occupational and Environmental Medicine*. 2001 Oct 1;58(10):649-55.
- Renne RA, Wehner AP, Greenspan BJ, Deford HS, Ragan HA, Westerberg RB, Buschbom RL, Burger GT, Hayes AW, Suber RL, Mosberg AT. 2-Week and 13-week inhalation studies of aerosolized glycerol in rats. *Inhalation toxicology*. 1992 Jan 1;4(2):95-111.
- Sussan TE, Gajghate S, Thimmulappa RK, Ma J, Kim JH, Sudini K, Consolini N, Cormier SA, Lomnicki S, Hasan F, Pekosz A. Exposure to electronic cigarettes impairs pulmonary anti-bacterial and anti-viral defenses in a mouse model. *PloS one*. 2015 Feb 4;10(2):e0116861.
- Misra M, Leverette R, Cooper B, Bennett M, Brown S. Comparative in vitro toxicity profile of electronic and tobacco cigarettes, smokeless tobacco and nicotine replacement therapy products: e-liquids, extracts and collected aerosols. *International journal of environmental research and public health*. 2014 Nov;11(11):11325-47.
- Jacobi J, Jang JJ, Sundram U, Dayoub H, Fajardo LF, Cooke JP. Nicotine accelerates angiogenesis and wound healing in genetically diabetic mice. *The American journal of pathology*. 2002 Jul 1;161(1):97-104.
- Schaal C, Chellappan SP. Nicotine-mediated cell proliferation and tumor progression in smoking-related cancers. *Molecular Cancer Research*. 2014 Jan 1;12(1):14-23.
- Murray RP, Connett JE, Zapawa LM. Does nicotine replacement therapy cause cancer? Evidence from the Lung Health Study. *Nicotine & Tobacco Research*. 2009 Jul 1;11(9):1076-

- 82.
21. Broughton G, Janis J, Attinger CE. The basic science of wound healing. *Plast Reconstr Surg* 2006. p. 12S-34S.
 22. Campos AC, Groth AK, Branco AB. Assessment and nutritional aspects of wound healing. *Curr Opin Clin Nutr Metab Care*. 2008;11(3):281-8.
 23. Mosser DM, Edwards JP. Exploring the full spectrum of macrophage activation. *Nat Rev Immunol*. 2008;8(12):958-69.
 24. Gosain A, DiPietro LA. Aging and Wound Healing. *World J Surg*. 2004;28(3):321-6.
 25. Guo S, DiPietro L. Critical Review in Oral Biology & Medicine: Factors Affecting Wound Healing. Houston: American Association for Dental Research/American Academy of Implant Dentistry; 2010. p. 219-29.
 26. Farsalinos K, Tsiapras D, Kyrzopoulos S, Voudris V. Acute and Chronic Effects of Smoking on Myocardial Function in Healthy Heavy Smokers: A Study of Doppler Flow, Doppler Tissue Velocity, and Two-Dimensional Speckle Tracking Echocardiography. *Echocardiography*. 2013 Mar;30(3):285-92.
 27. Van Staden SR, Groenewald M, Engelbrecht R, Becker PJ, Hazelhurst LT. Carboxyhaemoglobin levels, health and lifestyle perceptions in smokers converting from tobacco cigarettes to electronic cigarettes. *South African medical journal*. 2013;103(11):864-8.
Vardavas CI, Anagnostopoulos N, Kougias M, Evangelopoulou V, Connolly GN, Behrakis PK. Short-term pulmonary effects of using an electronic cigarette: impact on respiratory flow resistance, impedance, and exhaled nitric oxide. *Chest*. 2012 Jun 1;141(6):1400-6.
 28. Schober W, Szendrei K, Matzen W, Osiander-Fuchs H, Heitmann D, Schettgen T, Jörres RA, Fromme H. Use of electronic cigarettes (e-cigarettes) impairs indoor air quality and increases FeNO levels of e-cigarette consumers. *International journal of hygiene and environmental health*. 2014 Jul 1;217(6):628-37.
 29. Flouris AD, Chorti MS, Pouliantiti KP, Jamurtas AZ, Kostikas K, Tzatzarakis MN, Wallace Hayes A, Tsatsakis AM, Koutedakis Y. Acute impact of active and passive electronic cigarette smoking on serum cotinine and lung function. *Inhalation toxicology*. 2013 Feb 1;25(2):91-101.
 30. Farsalinos KE, Polosa R. Safety evaluation and risk assessment of electronic cigarettes as tobacco cigarette substitutes: a systematic review. *Therapeutic advances in drug safety*. 2014 Apr;5(2):67-86.
 31. Hua M, Alfi M, Talbot P. Health-related effects reported by electronic cigarette users in online forums. *Journal of medical Internet research*. 2013 Apr;15(4).
 32. Polosa R, Morjaria JB, Caponnetto P, Campagna D, Russo C, Alamo A, Amaradio M, Fisichella A. Effectiveness and tolerability of electronic cigarette in real-life: a 24-month prospective observational study. *Internal and emergency medicine*. 2014 Aug 1;9(5):537-46.
Dawkins L, Turner J, Roberts A, Soar K. 'Vaping' profiles and preferences: an online survey of electronic cigarette users. *Addiction*. 2013 Jun;108(6):1115-25.
 33. Etter JF, Bullen C. Electronic cigarette: users profile, utilization, satisfaction and perceived efficacy. *Addiction*. 2011 Nov;106(11):2017-28.
 34. Manzoli L, La Vecchia C, Flacco ME, Capasso L, Simonetti V, Boccia S, Di Baldassarre A, Villari P, Mezzetti A, Cicolini G. Multicentric cohort study on the long-term efficacy and safety of electronic cigarettes: study design and methodology. *BMC public health*. 2013 Dec;13(1):883.
 35. Reuther WJ, Brennan PA. Is nicotine still the bad guy? Summary of the effects of smoking on patients with head and neck cancer in the postoperative period and the uses of nicotine replacement therapy in these patients. *British Journal of Oral and Maxillofacial Surgery*. 2014 Feb 1;52(2):102-5.
 36. Willershausen I, Wolf T, Weyer V, Sader R, Ghanaati S, Willershausen B. Influence of E-smoking liquids on human periodontal ligament fibroblasts. *Head & face medicine*. 2014 Dec;10(1):39.
 37. Bahl V, Lin S, Xu N, Davis B, Wang YH, Talbot P. Comparison of electronic cigarette refill fluid cytotoxicity using embryonic and adult models. *Reproductive toxicology*. 2012 Dec 1;34(4):529-37.
 38. Romagna G, Alliffranchini E, Bocchietto E, Todeschi S, Esposito M, Farsalinos KE. Cytotoxicity evaluation of electronic cigarette vapor extract on cultured mammalian fibroblasts (ClearStream-LIFE): comparison with tobacco cigarette smoke extract. *Inhalation toxicology*. 2013 May 1;25(6):354-61.
 39. Farsalinos K, Romagna G, Alliffranchini E, Ripamonti E, Bocchietto E, Todeschi S, Tsiapras D, Kyrzopoulos S, Voudris V. Comparison of the cytotoxic potential of cigarette smoke and electronic cigarette vapour extract on cultured myocardial cells. *International journal of environmental research and public health*. 2013 Oct;10(10):5146-62.
 40. Behar RZ, Davis B, Wang Y, Bahl V, Lin S, Talbot P. Identification of toxicants in cinnamon-flavored electronic cigarette refill fluids. *Toxicology in vitro*. 2014 Mar 1;28(2):198-208.
 41. Wang C, Niu W, Chen H, Shi N, He D, Zhang M, Ge L, Tian Z, Qi M, Chen T, Tang X. Nicotine suppresses apoptosis by regulating $\alpha 7nAChR/Prx1$ axis in oral precancerous lesions. *Oncotarget*. 2017 Sep 26;8(43):75065.
 42. Chernyavsky AI, Shchepotin IB, Grando SA. Mechanisms of growth-promoting and tumor-protecting effects of epithelial nicotinic acetylcholine receptors. *International immunopharmacology*. 2015 Nov 1;29(1):36-44.
 43. Yu V, Rahimy M, Korrapati A, Xuan Y, Zou AE, Krishnan AR, Tsui T, Aguilar JA, Advani S, Alexander LE, Brumund KT. Electronic cigarettes induce DNA strand breaks and cell death independently of nicotine in cell lines. *Oral oncology*. 2016 Jan 1;52:58-65.
 44. Guttenplan JB, Sun YW, Chen KM, Kosinska W, El-Bayoumy K. Effects of electronic cigarettes (E-cigs) and hookah on the metabolic activation of the tobacco carcinogen, benzo (a) pyrene (BaP), and cellular proliferation in human oral leukoplakia cells. *Proceedings of AACR 107th Annual Meeting 2016*. DOI: 10.1158/1538-7445.AM2016-4063.
 45. Sundar IK, Javed F, Romanos GE, Rahman I. E-cigarettes and flavorings induce inflammatory and pro-senescence responses in oral epithelial cells and periodontal fibroblasts. *Oncotarget*. 2016 Nov 22;7(47):77196.
 46. Sancilio, S., Gallorini, M., Cataldi, A., Sancillo, L., Rana, R.A. and di Giacomo, V., 2017. Modifications in human oral fibroblast ultrastructure, collagen production, and lysosomal compartment in response to electronic cigarette fluids. *Journal of periodontology*, 88(7), 673-680.
 47. Alanazi, H., Park, H.J., Chakir, J., Semlali, A. and Rouabhia, M., 2018. Comparative study of the effects of cigarette smoke and electronic cigarettes on human gingival fibroblast proliferation, migration and apoptosis. *Food and chemical toxicology*, 118, 390-398.