



UNIVERSITY OF LEEDS

This is a repository copy of *Relative growth of invasive and indigenous tilapiine cichlid fish in Tanzania*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/156581/>

Version: Accepted Version

---

**Article:**

Bradbeer, SJ orcid.org/0000-0002-6844-6194, Ngatunga, BP, Turner, GF et al. (1 more author) (2020) Relative growth of invasive and indigenous tilapiine cichlid fish in Tanzania. *African Journal of Aquatic Science*, 45 (3). pp. 378-381. ISSN 1608-5914

<https://doi.org/10.2989/16085914.2019.1703169>

---

Copyright © NISC (Pty) Lt. This is an author produced version of an article published in *African Journal of Aquatic Science*. Uploaded in accordance with the publisher's self-archiving policy.

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

# 1 **Relative growth of invasive and indigenous tilapiine cichlid fishes**

## 2 **in Tanzania**

3  
4  
5 SJ Bradbeer<sup>1,2\*</sup>, BP Ngatunga<sup>3,4</sup>, GF Turner<sup>5</sup>, and MJ Genner<sup>1\*</sup>

6  
7 *1 School of Biological Sciences, Life Sciences Building, 24 Tyndall Avenue, University of*  
8 *Bristol, Bristol, BS8 1TQ, United Kingdom*

9 *2 School of Biology, Miall Building, University of Leeds, Leeds, LS2 9JT, United Kingdom.*

10 *3 Tanzania Fisheries Research Institute (TAFIRI), P.O. Box 9750, Dar es Salaam, Tanzania.*

11 *4 Department of Aquatic Sciences and Fisheries, University of Dar es Salaam, P.O. Box 35064,*  
12 *Dar es Salaam, Tanzania.*

13 *5 School of Natural Sciences, Bangor University, Bangor, Gwynedd, LL57 2UW, United*  
14 *Kingdom.*

15  
16 *Corresponding author, email: bssjb@leeds.ac.uk.*

17  
18  
19 Non-native species have been widely distributed across Africa for the enhancement of capture  
20 fisheries, but it can be unclear what benefits in terms of fisheries production the non-native  
21 species bring compared to native species. Here we compared the relative growth rate of  
22 sympatric populations of non-native *Oreochromis niloticus* (Nile tilapia) to native  
23 *Oreochromis jipe* (Jipe tilapia) in three waterbodies in northern Tanzania. Using scale  
24 increments as a proxy for growth, we found that *O. niloticus* had a high growth rate relative to  
25 *O. jipe*, with the highest *O. niloticus* growth rates being observed in Nyumba ya Mungu  
26 reservoir. These results help to explain why *O. niloticus* may be a superior competitor to native  
27 species in some circumstances. However, further introductions of this non-native species  
28 should be undertaken with caution given potential for negative ecological impacts on  
29 threatened indigenous tilapia species.

30  
31  
32 **Keywords:** aquaculture, growth, fisheries, *Oreochromis*, tilapia

33 Non-native invasive species are largely considered to have superior traits relative to their  
34 indigenous counterparts, enabling their establishment and success in invaded ranges.  
35 Characteristics associated with invasion success in fish include fast growth, broad  
36 environmental tolerances and high fecundity (Kolar and Lodge 2002; Moyle and Marchetti  
37 2006). These advantageous traits have been studied alongside environmental characters of the  
38 habitat to both evaluate impacts of non-native species, as well as predict future invasions (Copp  
39 et al. 2009; Marr et al. 2017). In some circumstances, non-native species outcompete  
40 established indigenous species for limited resources, such as food, breeding habitat and shelter  
41 (Bøhn et al. 2008). However, while competition is often inferred based on abundance trends,  
42 or shared patterns of resource use, often there is little evidence of the relative performance of  
43 non-native and native species where they co-occur.

44

45 One indicator of the relative fitness of sympatric species is growth. In fish, growth can be  
46 measured using a range of methods including quantifying the deposition of calcified layers on  
47 otoliths, vertebrae and scales (Cheung et al. 2007; Martin et al. 2012). Higher growth rates are  
48 considered advantageous as they enable individuals to reach reproductive age quicker, with  
49 less time spent at the more vulnerable juvenile life stage (Sutherland 1996). Furthermore in  
50 female fish, body size is directly related to egg output potential and therefore larger body sizes  
51 can enhance reproductive output (Barneche et al. 2018). Large body size may also pose an  
52 advantage for males in competition for spawning territories. Taken together, this evidence  
53 suggests that comparisons of growth rates of sympatric species with similar life history  
54 strategies can indicate relative competitive performance (Chifamba and Videler 2014).

55

56 *Oreochromis niloticus* (Nile tilapia (Linnaeus 1758)) is native to northern Africa, including the  
57 Nile and Niger river systems (Trewavas 1983). In Tanzania, the species is naturally distributed  
58 only in the Lake Tanganyika catchment (Shechonge et al. 2019a), but over recent decades the  
59 species has been widely distributed across the country (Shechonge et al. 2019b). Such  
60 introductions have been both deliberate to promote capture fisheries, and accidental following  
61 escapes from aquaculture facilities. Where *O. niloticus* is present in Tanzania, it typically co-  
62 occurs with indigenous tilapiine species (Bradbeer et al. 2019; Shechonge et al. 2019b).  
63 However, the fundamental ecological characteristics of populations of *O. niloticus* relative to  
64 those of native species in sympatric environments are largely unknown, including fisheries-  
65 related traits such as growth rates.

66

67 Here, we report a study comparing the relative growth of non-native *O. niloticus* to native  
68 *Oreochromis jipe* (Jipe tilapia (Lowe 1955)), a large bodied species endemic to the Pangani  
69 catchment that partially supports multiple artisanal fisheries in the region (Shechonge et al.  
70 2019b). When first described, this taxon was believed to represent a complex of three closely-  
71 related morphologically similar species, with *O. jipe* and *O. girigan* occupying different niches  
72 within Lake Jipe and *O. pangani* occupying the main Pangani river (Lowe 1955). These  
73 populations have not been studied in depth since and have not generally been distinguished as  
74 separate taxa by subsequent workers. Instead, they are now treated as a single species (Seegers  
75 et al. 2003; Fricke et al. 2019), and we followed this approach by assigning all studies  
76 populations to *O. jipe*. However, further research may support original species-level  
77 designation of Lowe (1955). We sampled fishers catches from three locations: Lake Kumba  
78 (4.806°S, 38.621°E, altitude 367m), Nyumba ya Mungu reservoir (3.612°S, 37.459°E, altitude  
79 519m) and the Pangani Falls reservoir (5.347°S, 38.645°E, altitude 191m) in August 2015  
80 (Figure 1). Lake Kumba is a natural lake with a surface area of 0.5km<sup>2</sup>, and a maximum depth  
81 of 7 metres. The Nyumba-ya-Mungu reservoir was formed when the Pangani river was  
82 dammed in 1965, and has a maximum surface area of 180km<sup>2</sup> and a maximum depth of  
83 approximately 45 metres (Petr et al. 1975; Bailey 1996). The Pangani Falls reservoir was  
84 formed when the Pangani river was dammed in 1994, and has a surface area of 0.5km<sup>2</sup> and a  
85 maximum depth of 10 metres (Anderson et al. 2006).

86 All sampled fishes were identified to species, individually labelled, and stored in 70% ethanol  
87 (Table 1). To assess growth rates, we followed the scale measurement method of Martin et al.  
88 (2012) that has been validated in experimental trials as a technique for quantifying recent  
89 growth of tilapiine cichlids. For each specimen, three scales were removed from the area  
90 superior to the lateral line and posterior to the head. Scales were then placed onto a microscope  
91 slide, treated with glycerol and covered with a glass coverslip. Images with a superimposed  
92 scale bar were taken using a M205C microscope (Leica, Wetzlar, Germany). Image files were  
93 loaded into tpsDIG 2.2 (Rohlf 2015) and from each scale, five measurements were recorded,  
94 namely the scale total width (longest distance across the scale; Figure 2a) and four separate  
95 “increment size” measurements of the distance between the first and fifth circuli on primary  
96 radii viewed from the anterior field of the scale (Figure 2b). From these measurements we  
97 calculated a mean scale width, and the mean increment size of the individual. Scale total width  
98 was employed as a covariate of increment size, alongside the factor variables species and  
99 sampling site, in an analysis of covariance in R version 3.6.0 (R Core Team 2019). Size-

100 standardised increment size (hereafter termed “relative growth”) was compared using marginal  
101 means and pairwise *post-hoc* Tukey’s tests in the emmeans package (Lenth et al. 2018).

102

103 We first observed a positive dependence of scale total width on increment size ( $F_{1,142} = 138.53$ ,  
104  $P < 0.001$ ), and after accounting for this covariation we interpret differences in increment size  
105 among populations as differences in growth rates. We observed an overall difference in growth  
106 rates among tilapia species from the different water bodies ( $F_{2,142} = 57.55$ ,  $P < 0.001$ ), and we  
107 observed that overall *O. niloticus* had a greater growth rate than *O. jipe* ( $F_{1,142} = 30.49$ ,  $P <$   
108  $0.001$ ). However, the extent of the differences in growth rates between the two species varied  
109 among locations ( $F_{2,142} = 12.72$ ,  $P < 0.001$ ; Figure 3). The clearest difference between the  
110 species was at Nyumba ya Mungu, where *O. niloticus* grew significantly faster than *O. jipe* ( $t$   
111  $= -7.303$ ,  $P < 0.001$ ). However, there were no significant growth differences between the  
112 species at either Lake Kumba ( $t = -0.946$ ,  $P = 0.346$ ) or the Pangani falls reservoir ( $t = -1.427$ ,  
113  $P = 0.156$ ). When comparing growth rates of *O. niloticus* between the water bodies, we found  
114 fish at Nyumba ya Mungu grew faster than those at Pangani falls ( $t = -4.710$ ,  $P < 0.001$ ) and  
115 Lake Kumba ( $t = -11.629$ ,  $P < 0.001$ ), while fish at Pangani falls also grew significantly faster  
116 than Lake Kumba ( $t = 5.625$ ,  $P < 0.001$ ). Similarly we found that *O. jipe* grew significantly  
117 faster at Nyumba ya Mungu than Lake Kumba ( $t = -2.876$ ,  $P = 0.013$ ), but there were no  
118 significant differences in *O. jipe* growth rates between the Pangani Falls and either Nyumba ya  
119 Mungu ( $t = 0.245$ ,  $P = 0.967$ ) or Lake Kumba ( $t = 2.364$ ,  $P = 0.051$ ).

120

121 The key results of this study are that *O. niloticus* had higher growth relative to the indigenous  
122 *O. jipe*, but also that extent of differences varied among locations. Such differences may have  
123 multiple explanations. Since *Oreochromis* can respond rapidly to selection on body size traits  
124 (Hulata et al. 1986), and recent work has identified significant genetic differences in neutral  
125 markers among the three sampled *O. niloticus* populations (Shechonge et al. 2019a), then  
126 genetic variation may underpin growth differences among the populations of both species. This  
127 may reflect historic selection from aquaculture prior to being introduced, or perhaps fisheries-  
128 induced evolution (Heino et al. 2015). Alternatively, the different sampled environments may  
129 differentially favour the species, with conditions within the Nyumba ya Mungu reservoir  
130 particularly well suited to the growth of *O. niloticus* relative to *O. jipe* present. It is unknown  
131 to what extent these species use different niches within each of the sampled environments. To  
132 fully understand the underlying reasons for the differences in growth rates between and within  
133 species would require more detailed study of growth rates in common-garden conditions, in

134 addition to an improved understanding of the relative differences among populations in habitat,  
135 diet and levels of fisheries exploitation.

136

137 Although our analysis of scale increments suggest higher growth for *O. niloticus* than *O. jipe*,  
138 to compare fisheries productivity, other relevant phenotypic characters need to be assessed  
139 including maximum length, age of maturity and food conversion rate. Higher individual growth  
140 rate need not always translate into greater rate of total fish biomass production, which is likely  
141 to be more relevant for small-scale fishery yields. Whether the differences we observed will  
142 have relevance for ecological interactions between the species is also unclear. It is possible that  
143 a faster growth rate may be advantageous for the non-native *O. niloticus* when competing with  
144 *O. jipe* for limited resources, including food, breeding space or shelter from predators. This is  
145 potentially of concern given the Critically Endangered IUCN red list status of the *O. jipe*, linked  
146 to its narrow geographic range and overall decreasing population trajectory (Bayona and  
147 Hanssens 2006). In Lake Kariba, *O. niloticus* has been shown to possess faster growth rate than  
148 indigenous *Oreochromis mortimeri* (Trevawas, 1966; Chifamba and Videler 2014). This,  
149 coupled with evidence of a rapid population expansion of *O. niloticus* matching a decline in *O.*  
150 *mortimeri* from the late 1990s onwards (Chifamba 2006), and evidence of overlapping resource  
151 use patterns (Mhlanga 2000), suggestss strong potential for *O. niloticus* to outcompete  
152 indigenous species. Equivalent monitoring of the abundance changes, resource use patterns and  
153 detailed analyses of life history parameters of both native and non-native tilapia populations in  
154 invaded habitats are needed to understand the full effects of introduced tilapia species across  
155 East Africa.

156

157 Acknowledgements - The work was funded by Royal Society-Leverhulme Trust Africa Awards  
158 AA100023 and AA130107. We thank the Tanzania Commission for Science and Technology  
159 (COSTECH) for research permits, and staff of the Tanzania Fisheries Research Institute for  
160 contributions to fieldwork.

161

## 162 **References**

163

164 Anderson R, Wanseth F, Cueller M, Von Mitzlaff U. 2006. Pangani falls re-development  
165 project in Tanzania. Swedish International Development Cooperation Agency,  
166 Stockholm.

167 Bailey RG. 1996. Changes in fish and fisheries ecology of a large man-made lake in

168 Tanzania, 1965-94. *Fisheries Management and Ecology* 3: 251-260.

169 Barneche DR, Robertson DR, White CR, Marshall DJ. 2018. Fish reproductive-energy output  
170 increases disproportionately with body size. *Science* 360: 642-645.

171 Bayona JDR, Hanssens M. 2006. *Oreochromis jipe*. The IUCN Red List of Threatened  
172 Species 2006: e.T60628A12388450. <https://bit.ly/2Kag28O>. Downloaded on 01 August  
173 2019.

174 Bøhn T, Amundsen PA, Sparrow A. 2008. Competitive exclusion after invasion? *Biological*  
175 *Invasions* 10: 359-368.

176 Bradbeer SJ, Harrington J, Watson H, Warriach A, Shechonge A, Smith A, Tamatamah R,  
177 Ngatunga BP, Turner GF, Genner MJ. 2019. Limited hybridization between introduced  
178 and Critically Endangered indigenous tilapia fishes in northern Tanzania. *Hydrobiologia*,  
179 832: 257-268.

180 Cheung CHY, Chaillé PM, Randall DJ, Gray JS, Au DWT. 2007. The use of scale increments  
181 as a means of indicating fish growth and growth impairments. *Aquaculture* 266: 102-111.

182 Chifamba PC, Videler JJ. 2014. Growth rates of alien *Oreochromis niloticus* and indigenous  
183 *Oreochromis mortimeri* in Lake Kariba, Zimbabwe. *African Journal of Aquatic Science*  
184 39: 167-176.

185 Chifamba PC. 2006. Spatial and historical changes of indigenous *O. mortimeri* following  
186 introduction of exotic *O. niloticus* in Lake Kariba. In: Odada EO, Olago DO, Ochala W,  
187 Ntiba N, Wandiga S, Gichuki N, Oyieke H (eds), 11th World Lakes Conference, Nairobi,  
188 Kenya, 31 October–4th November 2005, Proceedings Vol. II. Nairobi: Ministry of Water  
189 and Irrigation, Kenya, and International Lake Environment Committee. pp 500–504.

190 Copp GH, Vilizzi L, Mumford J, Fenwick GV, Godard MJ, Gozlan RE. 2009. Calibration of  
191 FISK, an invasiveness screening tool for nonnative freshwater fishes. *Risk Analysis: An*  
192 *International Journal* 29: 457-467.

193 Fricke R, Eschmeyer, WN Van der Laan R. (eds) 2019. Eschmeyer's Catalog of Fishes:  
194 Genera, Species, References. Electronic version accessed 15 Aug 2019.

195 Heino M, Pauli BD, Dieckmann U. 2015. Fisheries-induced evolution. *Annual Review of*  
196 *Ecology, Evolution and Systematics* 46: 461-480.

197 Hulata G, Wohlfarth GW, Halevy A. 1986. Mass selection for growth rate in the Nile tilapia  
198 (*Oreochromis niloticus*). *Aquaculture* 57: 177-184.

199 Kolar CS, Lodge DM. 2002. Ecological predictions and risk assessment for alien fishes in  
200 North America. *Science* 298: 1233-1236.

201 Lenth R 2019. emmeans: estimated marginal means, aka least-squares means. *R Package*

202           version 1.4. <http://CRAN.R-project.org/package=emmeans>

203 Marr SM, Ellender BR, Woodford DJ, Alexander ME, Wasserman RJ, Ivey P, Zengeya T,  
204           Weyl OL. 2017. Evaluating invasion risk for freshwater fishes in South Africa. *Bothalia-*  
205           *African Biodiversity and Conservation* 47: 1-10.

206 Martin, CH. 2012. Weak disruptive selection and incomplete phenotypic divergence in two  
207           classic examples of sympatric speciation: Cameroon Crater lake cichlids. *The American*  
208           *Naturalist* 180: 90-109.

209 Mhlanga L. 2000. The diet of five cichlid fish species from Lake Kariba, Zimbabwe.  
210           *Transactions of the Zimbabwe Scientific Association* 74: 16-21.

211 Moyle PB, Marchetti MP. 2006. Predicting invasion success: freshwater fishes in California as  
212           a model. *BioScience* 56: 515-524.

213 Petr T 1975. Limnology and fisheries of the Nyumba ya Mungu, a man-made lake in Tanzania.  
214           *African Journal of Tropical Hydrobiology and Fisheries* 4: 39-50.

215 Rohlf FJ. 2015. tpsDig, digitize landmarks and outlines, version 2.2. Department of Ecology  
216           and Evolution, State University of New York at Stony Brook

217 Seegers L, de Vos LDG, Okeyo DO. 2003. Annotated checklist of the freshwater fishes of  
218           Kenya (excluding the lacustrine haplochromines from Lake Victoria). *Journal of East*  
219           *African Natural History* 92: 11-47.

220 Shechonge A, Ngatunga BP, Tamatamah R, Bradbeer SJ, Sweke E, Smith A, Turner G.F.,  
221           Genner MJ. 2019a. Population genetics of Nile tilapia confirms the Lake Tanganyika  
222           population as a unique genetic resource. *Environmental Biology of Fishes* 102: 1107-  
223           1117

224 Shechonge A, Ngatunga BP, Bradbeer SJ, Day JJ, Freer JJ, Ford AGP, Kihedu J, Richmond T,  
225           Mzighani S, Smith AM, Sweke EA, Tamatamah R, Tyers AM, Turner GF, Genner MJ.  
226           2019b. Widespread colonisation of Tanzania catchments by introduced *Oreochromis*  
227           tilapia fishes: the legacy from decades of deliberate introduction. *Hydrobiologia* 832:  
228           235-253.

229 Sutherland WJ. 1996. From individual behaviour to population ecology. Oxford University  
230           Press.

231 Trewavas E. 1983. Tilapiine fishes of the genera *Sarotherodon*, *Oreochromis* and *Danakilia*.  
232           British Mus. Nat. Hist., London, UK.