

This is a repository copy of *Elevated platelet count appears to be causally associated with increased risk of lung cancer: A mendelian randomization analysis*.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/155989/

Version: Accepted Version

Article:

Zhu, Y, Wei, Y, Zhang, R et al. (74 more authors) (2019) Elevated platelet count appears to be causally associated with increased risk of lung cancer: A mendelian randomization analysis. Cancer Epidemiology, Biomarkers and Prevention, 28 (5). pp. 935-942. ISSN 1055-9965

https://doi.org/10.1158/1055-9965.epi-18-0356

© 2019 American Association for Cancer Research. This is an author-produced version of a paper subsequently published in Cancer Epidemiology, Biomarkers and Prevention. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



Elevated platelet count appears to be causally associated with increased risk of lung cancer: A Mendelian randomization analysis

Ying Zhu^{1†}, Yongyue Wei^{1,2,3†}, Ruyang Zhang^{1,2,3†}, Xuesi Dong⁴, Sipeng Shen², Yang Zhao^{1,3}, Jianling Bai¹, Demetrios Albanes⁵, Neil E. Caporaso ⁵, Maria Teresa Landi⁵, Bin Zhu⁵, Stephen J. Chanock⁵, Fangyi Gu⁶, Stephen Lam⁷, Ming-Sound Tsao⁸, Frances A. Shepherd⁸, Adonina Tardon⁹, Ana Fernandez-Somoano⁹, Guillermo Fernandez-Tardon⁹, Chu Chen¹⁰, Matthew J. Barnett¹⁰, Jennifer Doherty¹⁰, Stig E. Boiesen¹¹, Mattias Johansson¹², Paul Brennan¹², James D. McKay¹², Robert Carreras-Torres¹², Thomas Muley^{13,14}, Angela Risch^{14,15}, Heunz-Erich Wichmann¹⁶, Heike Bickeböller¹⁷, Albert Rosenberger¹⁷, Gad Rennert¹⁸, Walid Saliba¹⁸, Susanne M. Arnold¹⁹, John K. Field²⁰, Michael P.A. Davies²⁰, Michael W. Marcus²⁰, Xifeng Wu ²¹, Yuanqing Ye²¹, Loic Le Marchand²², Lynne R. Wilkens²², Olle Melander²³, Jonas Manjer²³, Hans Brunnström²⁴, Rayjean J. Hung²⁵, Geoffrey Liu²⁵, Yonathan Brhane²⁵, Linda Kachuri²⁵, Angeline S. Andrew²⁶, Eric J. Duell²⁷, Lambertus A. Kiemeney²⁸, Erik HFM van der Heijden ²⁸, Aage Haugen²⁹, Shanbeh Zienolddiny²⁹, Vidar Skaug²⁹, Kjell Grankvist³⁰, Mikael Johansson³⁰, Penella J. Woll³¹, Angela Cox³¹, Fiona Taylor³¹, Dawn M. Teare³², Philip Lazarus³³, Matthew B. Schabath³⁴, Melinda C. Aldrich³⁵, Richard S. Houlston³⁶, John McLaughlin³⁷, Victoria L. Stevens³⁸, Hongbing Shen³⁹, Zhibin Hu³⁹, Juncheng Dai³⁹, Christopher I. Amos⁴⁰, Younghun Han⁴⁰, Dakai Zhu⁴⁰, Gary E. Goodman⁴¹, Feng Chen^{1,3*}, David C. Christiani^{1,2,3*§}

¹Department of Biostatistics, School of Public Health, Nanjing Medical University, Nanjing, China;

²Department of Environmental Health, Harvard School of Public Health, Boston, Massachusetts, USA;

³China International Cooperation Center (CICC) for Environment and Human Health, Nanjing Medical University, Nanjing, China;

⁴Department of Epidemiology and Biostatistics, School of Public Health, Southeast University, Nanjing, China;

⁵Division of Cancer Epidemiology and Genetics, National Cancer Institute, US National Institutes of Health, Bethesda, Maryland, USA;

⁶ Department of Cancer Prevention and Control , Roswell Park Comprehensive Cancer Center , Buffalo , NY , USA;

⁷British Columbia Cancer Agency, Vancouver, British Columbia, Canada;

⁸University Health Network, Princess Margaret Cancer Centre, Toronto, Ontario,

Canada;

⁹University of Oviedo and CIBERESP, Faculty of Medicine, Oviedo, Spain; ¹⁰Program in Epidemiology, Fred Hutchinson Cancer Research Center, Seattle, Washington, USA;

¹¹Department of Clinical Biochemistry, Herlev and Gentofte Hospital, Copenhagen University Hospital, Copenhagen, Denmark; Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark; Copenhagen General Population Study, Herlev and Gentofte Hospital, Copenhagen, Denmark;

¹²International Agency for Research on Cancer, World Health Organization, Lyon, France;

¹³Thoraxklinik at University Hospital Heidelberg, Heidelberg, Germany;

¹⁴Translational Lung Research Center Heidelberg (TLRC-H), Heidelberg, Germany;

¹⁵German Center for Lung Research (DZL), Heidelberg, Germany; University of Salzburg and Cancer Cluster Salzburg, Salzburg, Austria;

¹⁶ Research Unit of Molecular Epidemiology, Institute of Epidemiology II, Helmholtz Zentrum München, German Research Center for Environmental Health, Neuherberg, Germany;

¹⁷Department of Genetic Epidemiology, University Medical Center, Georg-August-University Göttingen, Germany;

¹⁸Department of Community Medicine and Epidemiology, Clalit National Cancer Control Center at Carmel Medical Center and Technion Faculty of Medicine, Technion-Israel Institute of Technology, Haifa, Israel;

¹⁹Markey Cancer Center, University of Kentucky, Lexington, Kentucky, USA; ²⁰Institute of Translational Medicine, University of Liverpool, Liverpool, UK;

²¹Department of Epidemiology, University of Texas MD Anderson Cancer Center, Houston, Texas, USA;

²²Epidemiology Program, University of Hawaii Cancer Center, Honolulu, Hawaii, USA;

²³Faculty of Medicine, Lund University, Lund, Sweden;

²⁴Department of Pathology, Lund University, Lund, Sweden;

²⁵Lunenfeld-Tanenbaum Research Institute, Sinai Health System, University of Toronto, Toronto, Ontario, Canada;

²⁶Department of Epidemiology, Geisel School of Medicine, Hanover, New Hampshire, USA;

²⁷Unit of Nutrition and Cancer, Catalan Institute of Oncology (ICO-IDIBELL), Barcelona, Spain; ²⁸Radboud University Medical Center, Radboud Institute for Health Sciences, Nijmegen, the Netherlands;

²⁹National Institute of Occupational Health, Oslo, Norway;

³⁰Department of Medical Biosciences, Umeå University, Umeå, Sweden;

³¹Department of Oncology and Metabolism, University of Sheffield, Sheffield, UK;

³²School of Health and Related Research, University Of Sheffield, England, UK;

³³Department of Pharmaceutical Sciences, College of Pharmacy, Washington State University, Spokane, Washington, USA;

³⁴Department of Cancer Epidemiology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, Florida, USA;

³⁵Department of Thoracic Surgery, Division of Epidemiology, Vanderbilt University Medical Center;

³⁶The Institute of Cancer Research, London, England;

³⁷Public Health Ontario, Canada;

³⁸American Cancer Society, Inc., Atlanta, Georgia, USA;

³⁹Department of Epidemiology and Biostatistics, Jiangsu Key Lab of Cancer

Biomarkers, Prevention and Treatment, Collaborative Innovation Center for Cancer

Personalized Medicine, School of Public Health, Nanjing Medical University, Nanjing, P.R. China;

⁴⁰Biomedical Data Science, Geisel School of Medicine at Dartmouth, Hanover New Hampshire, USA;

⁴¹Swedish Medical Group, Seattle, WA, USA.

[†]Authors contributed equally to this work. [§]Senior author.

Running title: Relationship between platelets and risk of lung cancer

Key words: Lung cancer, Mendelian randomization, Platelet count, Instrumental variable.

Funding

This study was supported by the National Institute of Health (NIH) (CA092824 and CA209414 to D.C. Christiani), National Natural Science Foundation of China (81530088 and 81473070 to F. Chen, and 81373102 to Y. Zhao), State's Key Project of Research and Development Program (2016YFE0204900 to F. Chen), Key Project

of Natural Science Foundation of Jiangsu, China (14JA31002 to F. Chen), A Project Funded by the Priority Academic Program Development of Jiangsu Higher Education Institutions (PAPD), and Outstanding Young Teachers Training Program of Nanjing Medical University (to Y.Y. Wei). Transdisciplinary Research for Cancer in Lung (TRICL) and Oncoarray funding sources are detailed following. Sponsors had no role in design of the study, collection and analysis of data, or preparation of the manuscript. Transdisciplinary Research for Cancer in Lung (TRICL) of the International Lung Cancer Consortium (ILCCO) was supported by grants U19-CA148127 and CA148127S1. ILCCO data harmonization was supported by the Cancer Care Ontario Research Chair of Population Studies (to R.J. Hung) and the Lunenfeld-Tanenbaum Research Institute, Sinai Health System. The CAPUA study was supported by FIS-FEDER/Spain grants FIS-01/310, FIS-PI03-0365, and FIS-07-BI060604; FICYT/Asturias grants FICYT PB02-67 and FICYT IB09-133; and the University Institute of Oncology (IUOPA) of the University of Oviedo and the Ciber de Epidemiologiay Salud Pública (CIBERESP), Spain. Work performed in the CARET study was supported by the National Institute of Health (NIH)/National Cancer Institute (NCI) UM1 CA167462 (PI: G.E. Goodman), NIH UO1-CA6367307 (PIs: Omen, G.E. Goodman), NIH R01 CA111703 (PI: C. Chen), and NIH 5R01 CA151989-01A1 (PI: J. Doherty). The Liverpool Lung project was supported by the Roy Castle Lung Cancer Foundation. The Harvard Lung Cancer Study was supported by the NIH/NCI grants CA092824, CA090578, and CA074386. The Multiethnic Cohort Study was partially supported by NIH grants CA164973, CA033619, CA63464, and CA148127. Work performed in the MSH-PMH study was supported by the Canadian Cancer Society Research Institute (020214), Ontario Institute of Cancer and Cancer Care Ontario Chair Award (to R.J. Hung and G. Liu), and Alan Brown Chair and Lusi Wong Programs at Princess Margaret Hospital Foundation. NJLCS was funded by the State Key Program of National Natural Science of China (81230067), National Key Basic Research Program (2011CB503805), and Major Program of the National Natural Science Foundation of China (81390543). The Norway study was supported by the Norwegian Cancer Society, Norwegian Research Council. The Shanghai Cohort Study (SCS) was supported by NIH

4

R01 CA144034 (PI: J.M. Yuan) and UM1 CA182876 (PI: J.M. Yuan). The Singapore Chinese Health Study (SCHS) was supported by NIH R01 CA144034 (PI: J.M. Yuan) and UM1 CA182876 (PI: J.M. Yuan). Work in the TLC study has been supported in part the James & Esther King Biomedical Research Program (09KN-15), NIH Specialized Programs of Research Excellence (SPORE) (P50 CA119997), and a Cancer Center Support Grant (CCSG) at the H. Lee Moffitt Cancer Center and Research Institute, an NCI designated Comprehensive Cancer Center (P30-CA76292). The Vanderbilt Lung Cancer Study - BioVU dataset used for the described analyses was obtained from Vanderbilt University Medical Center's BioVU, which is supported by institutional funding, the 1S10RR025141-01 instrumentation award, and by the Vanderbilt Nature Genetics: doi:10.1038/ng.3892. The Clinical and Translational Science Awards (CTSA) grant UL1TR000445 was from the National Center for Advancing Translational Sciences (NCATS)/NIH. M.C. Aldrich was supported by NIH/NCI K07CA172294 (PI: M.C. Aldrich), and Dr. Bush was supported by National Human Genome Research Institute (NHGRI)/NIH U01HG004798 (PI: Crawford). The Copenhagen General Population Study (CGPS) was supported by the Chief Physician Johan Boserup and Lise Boserup Fund, the Danish Medical Research Council, and Herlev Hospital. The NELCS study was supported by grant P20RR018787 from the National Center for Research Resources (NCRR), a component of the NIH. The Kentucky Lung Cancer Research Initiative was supported by the Department of Defense [Congressionally Directed Medical Research Program, US Army Medical Research and Materiel Command Program] under award 10153006 (W81XWH-11-1-0781). This research was also supported by unrestricted infrastructure funds from the UK Center for Clinical and Translational Science, NIH grant UL1TR000117, and Markey Cancer Center NCI Cancer Center Support Grant (P30 CA177558) Shared Resource Facilities: Cancer Research Informatics, Biospecimen and Tissue Procurement, and Biostatistics and Bioinformatics. The M.D. Anderson Cancer Center study was supported in part NIH grants P50 CA070907 and R01 CA176568 (to X.F. Wu), Cancer Prevention & Research Institute of Texas RP130502 (to X.F. Wu), and University of Texas MD Anderson Cancer Center institutional support for the Center for Translational and Public Health

Genomics. The deCODE study of smoking and nicotine dependence was funded in part by grant R01- DA017932 from the National Institutes on Drug Abuse (NIDA). The Lodz center study was partially funded by Nofer Institute of Occupational Medicine under task NIOM 10.13: Predictors of mortality from non-small cell lung cancer - field study. Genetic sharing analysis was funded by NIH grant CA194393. The ResoLuCENT study (Resource for the Study of Lung Cancer Epidemiology in North Trent) is funded by the Sheffield Hospitals Charity, Sheffield Experimental Cancer Medicine Centre and Weston Park Hospital Cancer Charity. F.Taylor was supported by a Cancer Research UK/Yorkshire Cancer Research Clinical Fellowship. B. Zhu's work was supported by the Intramural Research Program of the Division of Cancer Epidemiology and Genetics, NCI. The Environment And Genetics in Lung cancer Etiology (EAGLE) study (PI: M.T. Landi) was supported by the Intramural Research Program of NIH, NCI, Division of Cancer Epidemiology and Genetics. F.Y. Gu was also supported by Roswell Park Cancer Institute and Cancer Center Supporting Grant P30CA016056. L.L. Marchand was supported by a program project grant from the NCI, NIH: P01 CA168530 and grant U01 CA164973. The Toronto study (PI: J. McLaughlin) was supported by Canadian Cancer Society Research Institute (020214). The Canadian Urban Environmental Health Research Consortium is funded by the Canadian Institutes for Health Research (J. McLaughlin). S.J. Chanock was supported by the Intramural Research Program of the National Institutes of Health NCI's Division of Cancer Epidemiology, and the American Cancer Society. This work was also supported by Cancer Research UK (C1298/A8362 to R.S. Houlston, C1298/A8780 and C1298/A8362 to J. McLaughlin and C18281/A19169 to R. Carreras-Torres).

*Send correspondence to

Dr. Feng Chen, SPH Building Room 412, 101 Longmian Avenue, Nanjing, Jiangsu 211166, China; e-mail: fengchen@njmu.edu.cn
Dr. David C. Christiani, Building I Room 1401, 665 Huntington Avenue, Boston, MA 02115, USA; e-mail: <u>dchris@hsph.harvard.edu</u>

Competing interests

The authors below are claiming a Conflict of Interest. Geoffrey Liu was on the Speaker's Bureau and received honoraria from Pfizer, AstraZeneca, Takeda, Roche, Novartis, BMS, Merck. Erik H.F.M. van der Heijden recieved commercial research support from Philips Medical Systems, and Astra Zeneca Oncology, and was on the Speaker's Bureau and received honoraria from Pentax Medical. The other authors declare no potential conflicts of interest.

Manuscript word count: 2000 words Tables: 2 Figures: 3

Abbreviations: SNPs, single nucleotide polymorphisms; NSCLC, non-small cell lung cancer; OR, odds ratio; PLT, platelet count; MR, Mendelian Randomization; IV, instrumental variable; GWAS, genome-wide association studies; SMR, summary data-based Mendelian randomization; AC, adenocarcinoma; SqCC, squamous cell carcinoma; SCLC, small cell lung cancer; IVW, inverse-variance weighted.

Abstract

Background: Platelets are a critical element in coagulation and inflammation, and activated platelets are linked to cancer risk through diverse mechanisms. However, a causal relationship between platelets and risk of lung cancer remains unclear. **Methods:** We performed single and combined multiple instrumental variable Mendelian Randomization (MR) analysis by an inverse-weighted (IVW) method, in addition to a series of sensitivity analyses. Summary data for associations between single nucleotide polymorphisms (SNPs) and platelet count is from a recent publication including 48,666 Caucasian Europeans and International Lung Cancer Consortium and Transdisciplinary Research in Cancer of the Lung data consisting of 29,266 cases and 56,450 controls analyze associations between candidate single nucleotide polymorphisms and lung cancer risk.

Results: Multiple instrumental variable analysis incorporating six SNPs showed a 62% increased risk of overall NSCLC (OR, 1.62; 95%Cl, 1.15–2.27; P = 0.005) and 200% increased risk for small cell lung cancer (OR, 3.00; 95%Cl, 1.27–7.06; P = 0.01), respectively. Results showed only a trending association with NSCLC histological subtypes, which may be due to insufficient sample size and/or weak effect size. A series of sensitivity analysis retained these findings.

Conclusion: Our findings suggest a causal relationship between elevated platelet count and increased risk of lung cancer and provide evidence of possible anti-platelet interventions for lung cancer prevention.

Impact: Our findings suggest a causal relationship of increased platelet count and risk of lung cancer, which also provide a better understanding of lung cancer etiology and potential evidence for anti-platelet interventions for lung cancer prevention.

Introduction

Lung cancer, a highly invasive, rapidly metastasizing cancer, has been the leading cause of cancer deaths worldwide for decades, accounting for more than one million deaths each year [1]. Smoking is a major risk factor for lung cancer and accounts for about 80% of male and 50% of female lung cancer cases [2]. In addition, environmental–occupational exposures [3, 4], lifestyle, and genetic variants [5] have been broadly explored as risks/predisposing factors for lung cancer. However, aspects of lung cancer risk remain largely unexplained and thus warrant further study.

The lung was recently noted to play a major role in platelet biogenesis and act as an ideal bioreactor for production of mature platelets from megakaryocytes, which account for ~50% of total platelet production [6]. Platelets are an important element in coagulation and inflammation, and diverse mechanisms link activated platelets to cancer progression [7, 8]. It has been identified that several variants in those chromosomal regions associated with platelet count (PLT) have associations with myocardial infraction, autoimmune and hematologic disorders. Tumor-educated blood platelets (TEPs) have emerged as promising biomarker sources for noninvasive detection of cancer, and it was demonstrated to discriminates patients with NSCLC from healthy individuals and patients with various non-cancerous inflammatory conditions [9, 10]. Indeed, high platelet count (PLT) is associated with increased mortality in a variety of cancers, including malignant mesothelioma [11], gynecological malignancies [12], and breast cancer [13]. In addition, platelet-tolymphocyte ratio and mean platelet volume also add value in early diagnosis of lung cancer [14] and prognosis prediction [15, 16]. These findings, taken together, indicate that disordered platelet production may be connected to lung carcinogenesis. However, due to potential unmeasured confounders in observational studies, the association between PLT and lung cancer risk remains unclear.

Mendelian Randomization (MR) is based on the principle that an individual's genotype is randomized at conception[17] and utilizes genetic variants as instrumental variables (IV) for the association between phenotypic exposures and outcomes to eliminate bias due to unmeasured confounders. Genetic variants used as instrumental variables should meet the following assumptions: (1) genetic variants are associated with exposure, (2) genetic variants affect outcome only via the

exposure, and (3) genetic variants are not associated with any confounders of the exposure–outcome association.[18] By finding a genetic marker that satisfies instrumental variable assumptions, Mendelian randomization analysis has been broadly used to estimate unconfounded associations between exposure and outcome [19], such as the effect of higher adult height on escalated cancer risk [20-24].

In this study, we performed summary data-based Mendelian randomization (SMR) [25] analysis which is the extension of two sample Mendelian randomization, using curated platelet count-related SNPs as instrumental variables to evaluate the association between platelet count and lung cancer risk by using summary statistics from recent large scale genome-wide association studies (GWAS).

Materials and Methods

Data source and study population

Mendelian Randomization analysis was conducted to estimate the effect of platelet count (X) on risk of lung cancer (Y) using genetic variants (G) as instrumental variables.[26] According to the MR analysis diagram described in Figure 1, we used coefficients of genetic variants on platelet count (b_{XG}) and their standard errors (SE_{XG}) from the recently published study of Gieger et al., which pooled 23 studies and included approximately 48,666 individuals of European descent [27].

The 54 genetic variants were identified that were associated with PLT (Table S1). One of the key assumptions underlying Mendelian Randomization is that the genetic variants (SNPs) used as instrumental variables are only related to the outcome of interest through the exposure variable under study. No pleiotropic pathways should exist from platelet-related SNPs to lung cancers through intermediates other than platelet count. Thus, six genetic variants (rs17030845, rs6141, rs3792366, rs210134, rs708382, and rs6065) where further selected as qualified instrumental variables that have prior functional knowledge supporting their association with platelets and no apparent link to cancer through intermediates other than platelets. By the way, the SNP rs6141 in THPO narrowly misses the level required for nominal significance ($P<5\times10^{-8}$) with $P=6.18\times10^{-8}$ in Europeans, but shows genome-wide significance in Japanese [28] . Therefore, it is still included serving as instrument variable for platelet count.

Coefficients (β_{YG}) and corresponding standard errors (SE_{YG}) of the association

between genetic variants and lung cancer risk were obtained from meta-analysis of existing Oncoarray and TRICL GWAS studies, which were detailed previously [29]. Briefly, overall non-small cell lung cancer (NSCLC) samples were composed from Oncoarray and TRICL GWASs, including 29,266 cases and 56,450 controls, and subgroup analyses were performed for 11,273 adenocarcinoma (AC), 7,426 squamous cell carcinoma (SqCC), and 2,664 small cell lung cancer (SCLC) cases (Table S2).

Mendelian Randomization (MR) analysis

Mendelian Randomization analysis with multiple instrumental variables was performed using an inverse-variance weighted (IVW) method combining the effect of genetic variants by weighted score. This score was used as an instrumental variable to estimate the effect of PLT on lung cancer risk [26]:

$$\hat{b}_{YX_{IVW}} = \frac{\sum_{i=1}^{N} \left(\frac{b_{XG_{i}} b_{YG_{i}}}{SE_{YG_{i}}^{2}} \right)}{\sum_{i=1}^{N} \left(\frac{b_{XG_{i}}}{SE_{YG_{i}}} \right)^{2}}, SE_{YX_{IVW}} = \sqrt{\frac{1}{\sum_{i=1}^{N} \left(\frac{b_{XG_{i}}}{SE_{YG_{i}}} \right)^{2}}}$$
(1)

Additionally, penalized IVW, robust IVW, MR-Egger, penalized MR-Egger, and robust MR-Egger methods were used for sensitivity analyses to evaluate robustness of the findings [30]. Step forward modeling was used to add an optimal instrumental variable each time from the left 48 SNPs, adding to the 6 curated SNPs for multiple instrumental variable analysis, until there was no improvement of statistical significance (*P*-value) for the test of causal effect. The modeling process was terminated when no added SNP increased -log10 (*P*-value) by 20% or 10%. Besides, MR analysis with a single IV (one SNP at a time) was performed as supplementary. Effect of PLT on Lung cancer risk [b_{YX} in log odds ratio (OR) scale] and its standard error (SE_{YX}) were estimated as follows [31]:

$$\hat{b}_{YX} = \frac{b_{YG}}{b_{XG}}, SE_{YX} = \frac{SE_{YG}}{b_{XG}}$$
 (2)

All analyses were performed using R Software Version 3.3.1 (The R Foundation). All tests were two-sided, and $P \le 0.05$ was considered statistically significant unless stated otherwise.

Results

Among 48,666 Europeans, 54 SNPs were quantitatively associated with platelet count with $P \le 5 \times 10^{-8}$ (Table S1) [27]. Associations of those 54 SNPs with risk of lung cancer were analyzed among 29,266 cases and 56,450 controls from OncoArray and previous GWAS studies. Demographics and study descriptions were detailed previously [29] and are briefly listed in Table S2 as well. Summarized association results of SNPs and lung cancer risk are listed in Table S3. According to instrumental variable assumptions that had evidence only related to platelets, six SNPs which are relatively independent and situated in different chromosomes were selected for MR analysis (Table 1), and 48 SNPs were excluded (Table S4).

In multiple IV analysis combining all six relatively independent SNPs situated in different chromosomes, a significant association between PLT and overall NSCLC risk is revealed, showing that each 100×10⁹/L increment of PLT was associated with a 62% increase in NSCLC risk (95%Cl, 1.15–2.27; P = 0.005) (Figure 2A and Figure 3A). In addition, five different methods of sensitivity analysis, including penalized IVW, robust IVW, MR-Egger, penalized MR-Egger, and robust MR-Egger, retained this association (Table 2). In NSCLC subtype analysis, it failed to detected significant associations between PLT and the risk of lung Adenocarcinoma (AC) (OR, 1.51; 95%Cl, 0.92–2.48; P = 0.11) (Figure 2B and Figure 3B) and squamous cells carcinomas (SqCC) (OR, 1.59; 95%Cl, 0.86–2.92; P = 0.14) (Figure 2C and Figure 3C). On the other hand, it is suggested that PLT is significantly associated with the risk of small cell lung cancer (SCLC) (OR, 3.00; 95%Cl, 1.27–7.06; P = 0.01) (Figure 2D and Figure 3D). The results of single IV are presented in supplementary (Table S5). No correction was conducted for them because a single weak instrument will have lower power to reject the null hypothesis [32].

We also performed a step forward modeling strategy to include more instrumental SNPs in the multiple instrumental variable model. Including more SNPs as instrumental variables yielded similar, yet more significant, causal estimates (Table S6 and Figure S1).

Discussion

This Mendelian randomization study suggests that each 100×10⁹/L increment in platelets results in a 62% increased risk of non-small cell lung cancer and, notably, a 200% increased risk of small cell lung cancer. However, this study failed to show evidence of a relationship between PLT and risk of AC and SqCC, probably resulting from insufficient sample size. As comparing with SCLC, the effect size of PLT on AC and SqCC are weaker, larger sample size is needed [33].

Platelets have been studied for decades as an important regulator of inflammation and thrombosis [34], which are broadly interrelated with human carcinogenesis [13]. Platelets are also recognized as a stimulator of proangiogenic factors [13] and a major source of vascular endothelial growth factor (VEGF) [35], platelet-derived growth factor (PDGF) [36, 37], and basic fibroblast growth factor (bFGF) [37], which act as promoters of tumor growth in lung [38-44]. New evidence suggests that platelets are relevant to defensive, physiological immune responses of the lungs and to inflammatory lung diseases [45]. Thus, higher platelet count has a potential biological connection to increased risk of lung cancer. Interestingly, p-selectin, an important adhesion molecule expressed on the surface of activated platelets, is more highly expressed in lung adenocarcinomas and squamous cell carcinomas than in healthy populations [46]. These results indicate a considerable role of platelets in lung carcinogenesis.

Intriguingly, a recent study indicates that cancer cells depend on platelets to avoid anoikis and succeed in metastasis [47]. Platelets induce resistance to anoikis in vitro and are critical for metastasis in vivo by activating RhoA-MYPT1-PP1-mediated YAP1 dephosphorylation and promoting its nuclear translocation to inhibit apoptosis. However, the unknown underlying mechanism warrants future well-designed functional experiments to clarify the role of platelets in these cellular processes.

In addition, anti-platelet agents, such as purinergic antagonists, are used clinically because they affect inflammatory pathways [48]. Recent publications demonstrate that platelets suppress T-cell responses against tumors through production and activation of immunosuppressive factors. These results suggest the use of a combination of immunotherapy and platelet inhibitors, such as aspirin [49, 50] and clopidogrel, as a therapeutic strategy against cancer [51, 52]. Therefore, it is possible that anti-platelet therapy could reduce lung cancer risk.

However, we acknowledge some limitations in our study. First, some associations between genetic instrumental variables and phenotype (platelet count) were insufficient and thus may result in a "weak instrument" phenomenon [53]. Second, in some scenarios, inconsistent results were observed between inverse-variance weighted and MR-Egger (or regular and penalized/robust) models. This phenomenon indicates that genetic variants probably have horizontal pleiotropy, and thus MR assumptions are likely violated [54]. Moreover, there is heterogeneity across results incorporating different SNP sets as instrumental variables, which indicates that the instrumental variable should be curated carefully before Mendelian randomization analysis. In this study, all platelet count-related SNPs were curated, and six were retained to better satisfy MR assumptions. Third, a linear association was assumed between PLT and lung cancer risk. However, the shape could be non-linear and thus warrants further study incorporating individual-level data. Fourth, we only evaluated platelet count as a potential causal factor, whereas platelet function plays a comparable causal role in this pathway. More detailed platelet information should be measured in future studies, including immature platelet fractions and function. In addition, we assumed that study populations used for the genetic instrument for platelet count and for risk of lung cancer were representative of the same general Caucasian population, which may not be true. Therefore, additional functional studies are needed to further evaluate the mechanisms that underlie associations between platelets and lung cancer risk.

Nonetheless, our findings do suggest a role of platelet count in risk of lung cancer. The results provide a better understanding of lung cancer etiology and evidence for a possible role of anti-platelet interventions in lung cancer prevention.

Acknowledgements

We thank the participants and staff for their important contributions to this study.

References

- Siegel R, Ma J Fau Zou Z, Zou Z Fau Jemal A, Jemal A. Cancer statistics, 2014. CA Cancer J Clin 2014;64(1):9-29. doi: 10.3322/caac.21208:1542-4863.
- [2] Jemal A, Bray F Fau Center MM, Center Mm Fau Ferlay J, Ferlay J Fau -Ward E, Ward E Fau - Forman D, Forman D. Global cancer statistics. CA Cancer J Clin 2011;65(2):87-108. doi: 10.3322/caac.21262:1542-4863.
- [3] Katanoda K, Sobue T Fau Satoh H, Satoh H Fau Tajima K, Tajima K Fau -Suzuki T, Suzuki T Fau - Nakatsuka H, Nakatsuka H Fau - Takezaki T, et al. An association between long-term exposure to ambient air pollution and mortality from lung cancer and respiratory diseases in Japan. J Epidemiol 2011;21(2):132-43(1349-9092 (Electronic)).
- [4] Cohen BL. Testing a BEIR-VI suggestion for explaining the lung cancer vs. radon relationship for U.S. counties. Health Phys 2000;78(5):522-7(0017-9078 (Print)).
- [5] Kligerman S, White C. Epidemiology of lung cancer in women: risk factors, survival, and screening. AJR Am J Roentgenol 2011;196(2):287-95. doi: 10.2214/AJR.10.5412(1546-3141 (Electronic)).
- [6] Lefrancais E, Ortiz-Munoz G, Caudrillier A, Mallavia B, Liu F, Sayah DM, et al. The lung is a site of platelet biogenesis and a reservoir for haematopoietic progenitors. Nature 2017;544(7648):105-109. doi: 10.1038/nature21706(1476-4687 (Electronic)).
- [7] Gasic Gj Fau Gasic TB, Gasic Tb Fau Stewart CC, Stewart CC.
 Antimetastatic effects associated with platelet reduction. Proc Natl Acad Sci U S A 1968;;61(1):46-52(0027-8424 (Print)).
- [8] Ji Y, Sheng L Fau Du X, Du X Fau Qiu G, Qiu G Fau Su D, Su D. Elevated platelet count is a strong predictor of poor prognosis in stage I nonsmall cell lung cancer patients. Platelets 2015;26(2):138-42. doi: 10.3109/09537104.2014.888547(1369-1635 (Electronic)).
- [9] Best MG, Sol N, In 't Veld S, Vancura A, Muller M, Niemeijer AN, et al. Swarm Intelligence-Enhanced Detection of Non-Small-Cell Lung Cancer Using Tumor-Educated Platelets. Cancer cell 2017;32(2):238-52.e9.
- [10] Joosse SA, Pantel K. Tumor-Educated Platelets as Liquid Biopsy in Cancer Patients. Cancer cell 2015;28(5):552-4.
- [11] Tural Onur S, Sokucu SN, Dalar L, Iliaz S, Kara K, Buyukkale S, et al. Are neutrophil/lymphocyte ratio and platelet/lymphocyte ratio reliable parameters as prognostic indicators in malignant mesothelioma? Ther Clin Risk Manag

2016;12:651-6. doi: 10.2147/TCRM.S104077(1176-6336 (Print)).

- [12] Menczer J. Preoperative elevated platelet count and thrombocytosis in gynecologic malignancies. Archives of gynecology and obstetrics 2016;295(1):9-15. doi: 10.1007/s00404-016-4212-9(1432-0711 (Electronic)).
- [13] Franco ATA-Ohoo, Corken A, Ware JA-Ohoo. Platelets at the interface of thrombosis, inflammation, and cancer. Blood 2015;126(5):582-8. doi: 10.1182/blood-2014-08-531582(1528-0020 (Electronic)).
- [14] Nikolic I Fau Kukulj S, Kukulj S Fau Samarzija M, Samarzija M Fau Jelec V, Jelec V Fau Zarak M, Zarak M, Orehovec B Fau Taradi I, et al. Neutrophil-to-lymphocyte and platelet-to-lymphocyte ratio help identify patients with lung cancer, but do not differentiate between lung cancer subtypes. Croat Med J 2016;57(3):287-92(1332-8166 (Electronic)).
- [15] Omar M, Tanriverdi O, Cokmert S, Oktay E, Yersal O, Pilanci KN, et al. Role of increased mean platelet volume (MPV) and decreased MPV/platelet count ratio as poor prognostic factors in lung cancer. LID - 10.1111/crj.12605 [doi]. Clin Respir J 2016; doi: 10.1111/crj.12605(1752-699X (Electronic)).
- [16] Oncel M, Kiyici A Fau Oncel M, Oncel M Fau Sunam GS, Sunam Gs Fau -Sahin E, Sahin E Fau - Adam B, Adam B. Evaluation of Platelet Indices in Lung Cancer Patients. Asian Pac J Cancer Prev 2015;16(17):7599-602(2476-762X (Electronic)).
- [17] Palmer TM, Sterne Ja Fau Harbord RM, Harbord Rm Fau Lawlor DA, Lawlor Da Fau - Sheehan NA, Sheehan Na Fau - Meng S, Meng S Fau -Granell R, et al. Instrumental variable estimation of causal risk ratios and causal odds ratios in Mendelian randomization analyses. Am J Epidemiol 2010;173(12):1392-403. doi: 10.1093/aje/kwr026(1476-6256 (Electronic)).
- [18] VanderWeele TJ, Tchetgen Tchetgen Ej Fau Cornelis M, Cornelis M Fau -Kraft P, Kraft P. Methodological challenges in mendelian randomization.
 Epidemiology 2014;25(3):427-35. doi: 10.1097/EDE.000000000000081(1531-5487 (Electronic)).
- [19] Didelez V, Sheehan N. Mendelian randomization as an instrumental variable approach to causal inference. Stat Methods Med Res 2007;16(4):309-30(0962-2802 (Print)).
- [20] Thrift AP, Risch HA, Onstad L, Shaheen NJ, Casson AG, Bernstein L, et al. Risk of esophageal adenocarcinoma decreases with height, based on consortium analysis and confirmed by Mendelian randomization. Clin Gastroenterol Hepatol 2014;12(10):1667-76.e1. doi: 10.1016/j.cgh.2014.01.039(1542-7714 (Electronic)).

- [21] Thrift AP, Gong J, Peters U, Chang-Claude J, Rudolph A, Slattery ML, et al. Mendelian randomization study of height and risk of colorectal cancer. Int J Epidemiol 2015;44(2):662-72. doi: 10.1093/ije/dyv082(1464-3685 (Electronic)).
- [22] Nuesch E, Dale C, Palmer TM, White J, Keating BJ, van Iperen EP, et al. Adult height, coronary heart disease and stroke: a multi-locus Mendelian randomization meta-analysis. Int J Epidemiol 2015;45(6):1927-1937. doi: 10.1093/ije/dyv074(1464-3685 (Electronic)).
- [23] Khankari NK, Shu XO, Wen W, Kraft P, Lindstrom S, Peters U, et al. Association between Adult Height and Risk of Colorectal, Lung, and Prostate Cancer: Results from Meta-analyses of Prospective Studies and Mendelian Randomization Analyses. PLoS Med 2016;13(9):e1002118. doi: 10.1371/journal.pmed.1002118(1549-1676 (Electronic)).
- [24] Davies NM, Gaunt TR, Lewis SJ, Holly J, Donovan JL, Hamdy FC, et al. The effects of height and BMI on prostate cancer incidence and mortality: a Mendelian randomization study in 20,848 cases and 20,214 controls from the PRACTICAL consortium. Cancer Causes Control 2015;26(11):1603-16. doi: 10.1007/s10552-015-0654-9(1573-7225 (Electronic)).
- [25] Zhu Z, Zhang F, Hu H, Bakshi A, Robinson MR, Powell JE, et al. Integration of summary data from GWAS and eQTL studies predicts complex trait gene targets. 2016;48(5):481-7.
- [26] Burgess S, Butterworth A Fau Thompson SG, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. Genet Epidemiol 2013;37(7):658-65. doi: 10.1002/gepi.21758(1098-2272 (Electronic)).
- [27] Gieger C, Radhakrishnan A Fau Cvejic A, Cvejic A Fau Tang W, Tang W Fau - Porcu E, Porcu E Fau - Pistis G, Pistis G Fau - Serbanovic-Canic J, et al. New gene functions in megakaryopoiesis and platelet formation. Nature 2011;480(7376):201-8. doi: 10.1038/nature10659(1476-4687 (Electronic)).
- [28] Kamatani Y, Matsuda K, Okada Y, Kubo M, Hosono N, Daigo Y, et al. Genome-wide association study of hematological and biochemical traits in a Japanese population. Nature genetics 2010;42(3):210-5.
- [29] McKay JD, Hung RJ, Han Y, Zong X, Carreras-Torres R, Christiani DC, et al. Large-scale association analysis identifies new lung cancer susceptibility loci and heterogeneity in genetic susceptibility across histological subtypes. Nature genetics 2017.
- [30] Bowden J, Del Greco MF, Minelli C, Davey Smith G, Sheehan N, Thompson

J. A framework for the investigation of pleiotropy in two-sample summary data Mendelian randomization. Stat Med 2017;36(11):1783-802.

- [31] Burgess S, Small DS, Thompson SG. A review of instrumental variable estimators for Mendelian randomization. Stat Methods Med Res 2016;pii: 0962280215597579(1477-0334 (Electronic)).
- [32] Davies NM, von Hinke Kessler Scholder S, Farbmacher H, Burgess S, Windmeijer F, Smith GD. The many weak instruments problem and Mendelian randomization. Stat Med 2015;34(3):454-68.
- [33] Burgess S. Sample size and power calculations in Mendelian randomization with a single instrumental variable and a binary outcome. Int J Epidemiol 2014;43(3):922-9.
- [34] Thomas MR, Storey RF. The role of platelets in inflammation. Thromb Haemost 2015;114(3):449-58. doi: 10.1160/TH14-12-1067(0340-6245 (Print)).
- [35] Verheul HM, Hoekman K Fau Luykx-de Bakker S, Luykx-de Bakker S Fau -Eekman CA, Eekman Ca Fau - Folman CC, Folman Cc Fau - Broxterman HJ, Broxterman Hj Fau - Pinedo HM, et al. Platelet: transporter of vascular endothelial growth factor. Clin Cancer Res 1997;3(12 Pt 1):2187-90(1078-0432 (Print)).
- [36] Pinedo HM, Verheul Hm Fau D'Amato RJ, D'Amato Rj Fau Folkman J,
 Folkman J. Involvement of platelets in tumour angiogenesis? Lancet
 1998;352(9142):1775-7(0140-6736 (Print)).
- [37] Cross MJ, Claesson-Welsh L. FGF and VEGF function in angiogenesis: signalling pathways, biological responses and therapeutic inhibition. Trends Pharmacol Sci 2010;22(4):201-7(0165-6147 (Print)).
- [38] Ferrara N, Gerber Hp Fau LeCouter J, LeCouter J. The biology of VEGF and its receptors. Nat Med 2003;9(6):669-76(1078-8956 (Print)).
- [39] Xiao XY, Lang XP. Correlation Between MMP-7 and bFGF Expressions in Non-small Cell Lung Cancer Tissue and Clinicopathologic Features. Cell Biochem Biophys 2015;73(2):427-432. doi: 10.1007/s12013-015-0656y(1559-0283 (Electronic)).
- [40] Otaka Y, Rokudai S, Kaira K, Fujieda M, Horikoshi I, Iwakawa-Kawabata R, et al. STXBP4 Drives Tumor Growth and Is Associated with Poor Prognosis through PDGF Receptor Signaling in Lung Squamous Cell Carcinoma. Clin Cancer Res 2017;doi: 10.1158/1078-0432.CCR-16-1815(1078-0432 (Print)).
- [41] Naykoo NA, Dil A, Rasool R, Shah S, Ahangar AG, Bhat IA, et al. Single nucleotide polymorphisms, haplotype association and tumour expression of the vascular endothelial growth factor (VEGF) gene with lung carcinoma.

Gene 2017;608:95-102. doi: 10.1016/j.gene.2017.01.007(1879-0038 (Electronic)).

- [42] Jahanban-Esfahlan R, Seidi K, Monfaredan A, Shafie-Irannejad V, Abbasi MM, Karimian A, et al. The herbal medicine Melissa officinalis extract effects on gene expression of p53, Bcl-2, Her2, VEGF-A and hTERT in human lung, breast and prostate cancer cell lines. (1879-0038 (Electronic)).
- [43] Hu M, Hu Y, He J, Li B. Prognostic Value of Basic Fibroblast Growth Factor (bFGF) in Lung Cancer: A Systematic Review with Meta-Analysis. PLoS One 2016;11(1):e0147374. doi: 10.1371/journal.pone.0147374(1932-6203 (Electronic)).
- [44] Dadrich M, Nicolay NH, Flechsig P, Bickelhaupt S, Hoeltgen L, Roeder F, et al. Combined inhibition of TGFbeta and PDGF signaling attenuates radiationinduced pulmonary fibrosis. Oncoimmunology 2016;5(5):e1123366. doi: 10.1080/2162402X(2162-4011 (Print)).
- [45] Middleton EA, Weyrich AS, Zimmerman GA. Platelets in Pulmonary Immune Responses and Inflammatory Lung Diseases. Physiological reviews 2016;96(4):1211-59.
- [46] Gong L, Cai Y, Zhou X, Yang H. Activated platelets interact with lung cancer cells through P-selectin glycoprotein ligand-1. Pathology oncology research : POR 2012;18(4):989-96.
- [47] Haemmerle M, Taylor ML, Gutschner T, Pradeep S, Cho MS, Sheng J, et al. Platelets reduce anoikis and promote metastasis by activating YAP1 signaling. 2017;8(1):310.
- [48] Pitchford SC. Novel uses for anti-platelet agents as anti-inflammatory drugs.Br J Pharmacol 2007;152(7):987-1002(0007-1188 (Print)).
- [49] Cao Y, Nishihara R, Wu K, Wang M, Ogino S, Willett WC, et al. Populationwide Impact of Long-term Use of Aspirin and the Risk for Cancer. JAMA Oncol 2016;2(6):762-9.
- [50] Oh SW, Myung SK, Park JY, Lee CM, Kwon HT. Aspirin use and risk for lung cancer: a meta-analysis. Annals of Oncology 2011;22(11):2456-65.
- [51] Bordon Y. Tumour immunology: Platelets a new target in cancer immunotherapy? Nature reviews Immunology 2017;17(6):348. doi: 10.1038/nri.2017.61(6):348.
- [52] Saleh Rachidi AM, Brian Riesenberg, Bill X. Wu, Michelle H. Nelson, Caroline Wallace, Chrystal M. Paulos, Mark P. Rubinstein, Elizabeth Garrett-Mayer, Mirko Hennig, Daniel W. Bearden, Yi Yang, Bei Liu and Zihai Li. Platelets subvert T cell immunity against cancer via GARP-TGFβ axis. Science

Immunology 2017;DOI: 10.1126/sciimmunol.aai7911.

- [53] Burgess S, Thompson SG. Bias in causal estimates from Mendelian randomization studies with weak instruments. Stat Med 2011;30(11):1312-23. doi: 10.1002/sim.4197(1097-0258 (Electronic)).
- [54] Bowden J, Burgess S, Smith GD. Difficulties in Testing the Instrument Strength Independent of Direct Effect Assumption in Mendelian Randomization. JAMA cardiology 2017.

SNP	Chr:Position (hg19)	Gene	Reference allele	Effect allele	EAF (%)	Function	b (95% CI)	Р
rs17030845	2: 43687879	THADA	С	Т	09.65	intron	-3.58 (-4.67, -2.49)	1.27×10 ⁻¹⁰
rs6141	3: 184090266	THPO	Т	С	47.39	3' UTR	–2.47 (–3.36, –1.57)	6.18×10 [–]
rs3792366	3: 122839876	PDIA5	А	G	38.68	intron	2.153 (1.44, 2.87)	3.60×10 ^{−9}
rs210134	6: 33540209	BAK1	G	А	29.29	500bp downstream	-4.96 (-5.73, -4.18)	7.11×10 ⁻³⁶
rs708382	17: 42442344	FAM171A2-ITGA2B	Т	С	39.66	2kb upstream	-2.44 (-3.28, -1.59)	1.51×10 ⁻⁸
rs6065	17: 4836381	GP1BA	С	Т	08.53	missense	4.19 (2.96, 5.43)	2.92×10 ⁻¹¹

Table 1. SNPs of specific platelet-related genes

EAF, effect allele frequency; UTR, untranslated region

SNP	Overall NSCLC		AC		SqCC		SCLC	
SINF	OR (95%CI)	Р	OR (95%CI)	Р	OR (95%CI)	Р	OR (95%CI)	Р
IVW	1.62 (1.15, 2.27)	0.005	1.51 (0.92, 2.48)	0.11	1.59 (0.86, 2.92)	0.14	3.00 (1.27, 7.06)	0.01
Penalized IVW	1.62 (1.15, 2.27)	0.005	1.51 (0.92, 2.48)	0.11	1.59 (0.86, 2.92)	0.14	3.00 (1.27, 7.06)	0.01
Robust IVW	1.63 (1.26, 2.11)	<0.001	1.51 (0.90, 2.53)	0.12	1.54 (0.93, 2.56)	0.09	3.30 (1.52, 7.15)	0.003
MR-Egger	3.25 (1.16, 9.11)	0.03	6.06 (1.45, 25.27)	0.01	1.75 (0.22, 13.84)	0.59	3.29 (0.24, 45.11)	0.37
Penalized MR-Egger	3.25 (1.16, 9.11)	0.03	6.06 (1.45, 25.27)	0.01	1.75 (0.22, 13.84)	0.59	3.29 (0.24, 45.11)	0.37
Robust MR-Egger	3.23 (1.80, 5.78)	<0.001	5.88 (2.74, 12.61)	<0.001	1.70 (0.48, 6.08)	0.41	3.56 (1.25, 10.14)	0.02

Table 2. Association between platelet count and risk of lung cancer using multiple IV analysis

OR, odds ratio of platelet count (PLT) on lung cancer risk per 100×10⁹/L increment of PLT; NSCLC, non-small cell lung cancer; AC, adenocarcinoma; SqCC, squamous cell carcinoma; SCLC, small-cell carcinoma; IVW, inverse-variance weighted.

Figure Legends.

Figure 1. Diagram of Mendelian randomization (MR) analysis. MR aims to estimate the unbiased causal relationship between PLT and lung cancer risk by incorporating genetic variants as instrumental variables (IVs). Dashed line represents the association between instrumental variable (SNP) and outcome (risk of lung cancer), denoted using b_{YG} in log(odds ratio) scale and its standard error (SE_{YG}), which were obtained from GWAS. Estimates of quantitative trait loci relationship between SNP and phenotype (platelet count) were obtained from a recently published article, which were described by b_{XG} and SE_{XG} . Lung cancer risk was assessed for non-small cell lung cancer (NSCLC), adenocarcinoma (AC), squamous cell carcinoma (SqCC), and small cell carcinoma (SCLC).

Figure 2. Causal associations between platelet count and lung cancer risk. Forest plots of causal associations between platelet count (PLT) and risk of lung cancer using Mendelian randomization (MR) analysis incorporating different genetic variants as instrumental variables (IVs). Associations of PLT with risk of (A) non-small cell lung cancer (NSCLC), (B) adenocarcinoma (AC), (C) squamous cell carcinoma (SqCC), and (D) small-cell lung cancer (SCLC) were analyzed based on single IV or multiple IVs using inverse-variance weighted (IVW) analysis.

Figure 3. Assocations between SNPs and lung cancer risk. Scatter plots displaying estimates of the association between each SNP and risk of lung cancer against quantitative relationship of each SNP on platelet count (PLT) for (A) non-small cell lung cancer (NSCLC), (B) adenocarcinoma (AC), (C) squamous cell carcinoma (SqCC), and (D) small cell lung cancer (SCLC). Slope of the blue dashed line through the plot represents inverse-variance weighted (IVW) regression estimate for the causal effect of PLT on lung cancer risk.