



UNIVERSITY OF LEEDS

This is a repository copy of *Emerging drugs for treating methicillin-resistant Staphylococcus aureus*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/152609/>

Version: Accepted Version

Article:

Bassetti, M, Russo, A, Carnelutti, A et al. (1 more author) (2019) Emerging drugs for treating methicillin-resistant *Staphylococcus aureus*. *Expert Opinion on Emerging Drugs*, 24 (3). pp. 191-204. ISSN 1472-8214

<https://doi.org/10.1080/14728214.2019.1677607>

This article is protected by copyright. This is an author produced version of a journal article published in *Expert Opinion on Emerging Drugs*. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Emerging drugs for treating methicillin-resistant *Staphylococcus aureus*

Matteo Bassetti ^{1,2}, Alessandro Russo ¹, Alessia Carnelutti ¹, Mark Wilcox ³

¹ Infectious Diseases Clinic, Department of Medicine University of Udine and Azienda Sanitaria
Universitaria Integrata di Udine, Udine, Italy

² Department of Health Sciences, University of Genoa, Genoa, Italy

³ Leeds Teaching Hospitals NHS Trust & University of Leeds, Leeds, UK.

Corresponding author:

Matteo Bassett, MD, PhD

Clinica Malattie Infettive, Azienda Sanitaria Universitaria Integrata di Udine, Presidio Ospedaliero
Universitario Santa Maria della Misericordia

Piazzale Santa Maria della Misericordia 15, 33100 Udine, Italy

Phone +39 0432 559353; Fax +39 0432 559360

Email: matteo.bassetti70@gmail.com

Abstract

In clinical practice, methicillin-resistant *Staphylococcus aureus* (MRSA) represents a major threat and has been associated with high rates of inadequate antibiotic treatment and significant increases in morbidity, mortality and overall healthcare costs. The association between the prescription of an inappropriate or delayed antibiotic and impaired clinical outcomes has been widely described, with clinical implications mainly depending on the type of infection and the illness severity.

To face off the threat of MRSA, many new therapeutic options with a peculiar activity against MRSA have been recently developed and approved. New agents are characterized by peculiar issues in terms of spectrum of activity, pharmacokinetics, risk of drug-drug interactions and toxicity, with potential main advantages that should be considered in everyday clinical practice. The most attractive characteristic of new drugs is represented by the broad spectrum of activity against multi-drug resistant pathogens; moreover, new compounds in most cases are characterized by favorable toxicity profiles compared with old drugs currently used in clinical practice.

Some of the new antimicrobials will be also available as oral formulations, with the potential for oral shift even in infections due to resistant pathogens. In particular conditions/populations (e.g. liver failure, renal disease, pregnancy, diabetic, children, elderly) novel antibiotics with reduced toxicity could be an important option, also in discharged patients.

Background

During last decade alarming rates of methicillin-resistance in *S. aureus* have been reported worldwide, with up to 25% of *S.aureus* isolates displaying methicillin-resistance in the majority of countries, including USA, Latin America, Sub-Saharan Africa, Russia, India and China¹. The large SENTRY Antimicrobial Surveillance Program has been monitoring *S.aureus* susceptibility patterns in 45 countries from 1997 to 2016; methicillin-resistant *S.aureus* (MRSA) accounted for 40,3% of isolates overall, with a geographical variability ranging from 26.8% in Europe to 47.0% in North America². In Europe, MRSA accounts for 17% of isolates overall, even with a wide regional variability, ranging from <1% in northern Europe and up to 25% in southern Europe, including Spain, Portugal, Italy and Greece³.

Although the burden of MRSA still belongs to hospital setting⁴⁻⁵, during the last years MRSA has progressively spread also into the community, particularly among outpatients affected by complex clinical problems and having frequent contact with the healthcare system (e.g. residence in a nursing home or long- term-care facility; recent hospitalization during the past 3 months; hemodialysis or intravenous chemotherapy receipt; intravenous therapy, wound care or enteric nutrition at home), thus expanding the at-risk population for the acquisition of infections due to MRSA⁶⁻⁷⁻⁸. Injection drug users represent an additional expanding population for MRSA colonization and infection development, as recently reported⁹.

In clinical practice, MRSA represents a major threat and has been associated with high rates of inadequate antibiotic treatment and significant increases in morbidity, mortality and overall healthcare costs¹⁰⁻¹¹.

A recently published retrospective analysis using data from the National Inpatient Sample from the Agency for Healthcare Research and Quality for the years 2010-2014 found increasing MRSA-related health-care costs during last years, although costs associated with methicillin-susceptible *S.aureus* (MSSA)-related infections have converged with costs of similar MRSA-related hospitalizations.

However, MRSA-related hospitalizations have been associated with a higher adjusted mortality rate compared with MSSA infections¹².

A multicenter, retrospective cohort study including inpatient acute-care episodes in ten European hospitals during the period 2010-2011 tried to estimate the impact of antimicrobial resistance on hospital mortality, excess length of stay (LOS) and overall costs. Per infection, *S. aureus* bloodstream infections had a greater effect on mortality, LOS and costs compared with bloodstream due to Enterobacteriaceae. In this study, however, methicillin resistance did not significantly increase the hazard of death or further prolong the excess LOS compared with bloodstream infections due methicillin-susceptible *S. aureus*¹³.

In clinical practice, *S.aureus* and its methicillin-resistant variant represent a major problem in a wide spectrum of both community and hospital-acquired infections.

According with data from the SENTRY Antimicrobial Surveillance Program, *S.aureus* represents the most frequently encountered pathogen in bacteremia worldwide, accounting for 20,7% of cases, with a significant increase in MRSA prevalence until 2008¹⁴.

In the setting of complicated skin and skin structure infections (cSSTIs) MRSA is a major concern. A recently published study evaluated susceptibility patterns of pathogens isolated from patients with community-acquired cSSTIs and reported MRSA rates varying between 15,8% and 21,4%, with the higher rates in Asia and Pacific regions¹⁵. Moreover, approximately 9% of diabetic patients are colonized by MRSA, and up to 16% of diabetic foot infections are currently due to MRSA¹⁶. Traditionally, MRSA represents a major problem in hospital- and in Intensive Care Unit – acquired pneumonia¹⁷. However, recent data emphasized the emerging role of MRSA also in community acquired pneumonia, particularly among patients presenting with specific risk factors such as age > 65 years, with previous antibiotic use, underlying chronic respiratory disorder or chronic renal failure¹⁸⁻¹⁹

Medical need and existing treatment

The association between the prescription of an inappropriate or delayed antibiotic and impaired clinical outcomes has been widely described, with clinical implications mainly depending on the type of infection and the illness severity²⁰⁻²¹⁻²²⁻²³.

The prompt identification of at-risk population for the development of MRSA infections is crucial to reduce the risk of prescribing an inappropriate empiric antibiotic treatment. However, majority of the recognized risk factors for the acquisition of infections due to MRSA (e.g. older age, hospital-acquired infection, recent antibiotic treatment or hospitalization, chronic underlying diseases) is generic and algorithms with good sensitivity and specificity for the identification of these patients are lacking. MRSA colonization has been strongly associated with an increased risk for the development of infections due to the same pathogen, but nasal swabs are not always performed in routine clinical practice and no recommendations can be made in this regard so far, except for specific clinical settings²⁴⁻²⁵.

In this setting, in areas with high MRSA prevalence, the prescription of an early broad-spectrum empiric therapy followed by a prompt de-escalation upon availability of microbiological data should be strongly recommended in severe infections requiring hospitalization and when risk factors for MRSA are present. Available antimicrobials, however, possess some limitations for the use in clinical practice.

Vancomycin still represents the most used antimicrobial with anti-MRSA activity worldwide²⁶⁻²⁷. However, in a recently published prospective, multicenter, observational study treatment failure and acute kidney injury were reported, respectively, in 18% and 26% of hospitalized adult patients with MRSA bacteremia treated with vancomycin. Moreover, higher vancomycin exposures did not confer a lower treatment failure risk but were associated with more acute kidney injury²⁸. Due to the well-known correlation between plasmatic vancomycin concentrations and the development of nephrotoxicity, Therapeutic Drug Monitoring should be considered, particularly among older patients with pre-existing renal failure, co-administration of nephrotoxic drugs or multiple comorbidities²⁹.

Moreover, vancomycin requires a twice-daily intravenous administration, not allowing the treatment of outpatients.

Daptomycin represents a good alternative to vancomycin for the treatment of the majority of MRSA infections (with the exception of pneumonia due to the inactivation of daptomycin by the pulmonary surfactant) and is widely used in clinical practice due to the rapid bactericidal activity, the good tolerability profile and the once-daily intravenous administration³⁰. The use of high daptomycin doses (8-10 mg/kg) is currently well established and is recommended to improve clinical outcomes while minimizing the risk of resistance selection³¹⁻³². However, due to the need for a once-daily intravenous administration, daptomycin does not represent the ideal option for early-discharge policies and for the treatment of outpatients. Although daptomycin is commonly used as monotherapy in majority of clinical settings, recent data support an investigational use of daptomycin in combinations with new antimicrobials (e.g. caftaroline) for the treatment of MRSA bacteremia, but further studies are needed³³.

Linezolid currently represents a good option for the treatment of MRSA infections, particularly pneumonia and skin and soft tissue infections³⁴⁻³⁵.

Major limitations for the use of linezolid in clinical practice are the potential risk of hematological side effects, mainly thrombocytopenia, which is particularly relevant in patients with plasmatic exposure above 8 mg/liter³⁶; for this reason, Therapeutic Drug Monitoring might probably be useful in majority of patients³⁷. Moreover, due to the inhibition of mono-amino-oxidase enzymes, linezolid is contraindicated in association with drugs with serotonergic activity for the risk of development of serotonergic syndrome³⁸.

To face off the threat of methicillin-resistant *Staphylococcus aureus* (MRSA), many new therapeutic options with a peculiar activity against MRSA have been recently developed and approved. New agents are characterized by peculiar issues in terms of spectrum of activity, pharmacokinetics, risk of drug-drug interactions and toxicity, with potential main advantages that should be considered in everyday clinical practice.

Eravacycline

Eravacycline is a novel fluorocycline in Phase 3 clinical development for cIAI and cUTI. Eravacycline is structurally similar to tigecycline but is not subjected to the mechanisms that are responsible for tetracycline resistance, such as efflux pumps and ribosomal protection protein³⁹. The most attractive characteristic of eravacycline is the broad-spectrum activity against both Gram-positive and Gram-negative resistant pathogens, including MRSA, enterococci (included vancomycin-resistant Enterococci) and Enterobacteriaceae expressing resistance genes from different classes of β -lactamases (particularly ESBL, KPC and OXA), with a 2- to 4-fold greater activity than tigecycline⁴⁰⁻⁴¹. Moreover, eravacycline currently represents the most potent antibiotic against MDR *A.baumannii*, with a fourfold higher activity compared with tigecycline, including strains resistant to sulbactam, imipenem/meropenem, levofloxacin and amikacin/tobramycin⁴². Eravacycline exerts also a potent activity against anaerobic pathogens⁴³. As tigecycline, eravacycline is not effective against *P. aeruginosa*⁴⁴.

Together with the broad-spectrum activity, another attractive characteristic of eravacycline is the availability of both intravenous and oral formulations, making eravacycline a potential option for early oral shift and early discharge in patients with infections due to MDR Gram-negative bacteria⁴⁵. In a recent Phase 3, randomized, double-blind, multicenter study eravacycline was found to be non-inferior compared to ertapenem for the treatment of patients with cIAI (Clinicaltrials.gov, Identifier NCT01844856)⁴⁶. No studies investigating eravacycline efficacy for the treatment of respiratory tract infections are currently ongoing; however, a Phase 1 study conducted in 20 healthy adult volunteers analyzed eravacycline safety and pulmonary concentration after the administration of 1 mg of eravacycline/kg intravenously every 12 h for a total of seven doses over 4 days. Eravacycline was found to achieve 6-fold and a 50-fold higher concentrations in the ELF and alveolar macrophages than in plasma respectively, supporting its potential role for the treatment of respiratory tract infections. Moreover, eravacycline was well tolerated, with no serious adverse events and no treatment

discontinuations⁴⁷. Promising data for the use of eravacycline for the treatment of pneumonia comes from a study by Grossmann et al, showing eravacycline to be as effective as linezolid in a neutropenic MRSA mouse lung infection model⁴⁸. These data make eravacycline an attractive option for the treatment of respiratory tract infections due to resistant pathogens, including MRSA, beta-lactamase producing Enterobacteriaceae and MDR A.baumannii and for oral step-down therapy.

Tedizolid

Tedizolid belongs to the class of oxazolidinones and is currently approved for the treatment of ABSSSIs. Tedizolid is characterized by a potent in vitro activity against Gram-positive pathogens, with a four- to eight-fold greater activity than linezolid; moreover, tedizolid is active against linezolid-nonsusceptible strains⁴⁹.

Tedizolid might represent a promising option for the treatment of MRSA pneumonia because of many advantages over linezolid, including: lower risk of myelotoxicity⁵⁰⁻⁵¹; lower risk of drug – drug interactions with selective serotonin reuptake inhibitors (SSRIs), compounds with serotonergic activity, and adrenergic agents due to its weak and reversible in vitro inhibition of the monoamine oxidase pathway⁵²; high bioavailability (>80%), with in vivo half-life value approximately twofold greater compared with linezolid, allowing once daily administration⁵³.

Moreover, PK/PD studies showed that tedizolid achieves approximately 40-fold higher concentration in ELF relative to free plasma ones, supporting the use of tedizolid in the setting of pneumonia⁵⁴.

The role of tedizolid for the treatment of MRSA respiratory tract infections is only investigational so far. However, promising data supporting the use of tedizolid for the treatment of respiratory infections come from a study conducted in an in vivo murine pneumonia model, showing tedizolid to be as effective as linezolid and more effective than vancomycin for the treatment of MRSA pneumonia⁵⁵.

A Phase 4 study designed to characterize the pharmacokinetics of intravenous and oral tedizolid in patients with cystic fibrosis is currently ongoing (Clinicaltrials.gov, Identifier NCT02444234).

A Phase 3, randomized, double blind study comparing tedizolid (200-mg intravenous once daily for 7

days, or 14 days in bacteremia) versus linezolid (600 mg intravenous every 12 h for 10 days, or 14 days for bacteremia) for the treatment of patients with presumed Gram-positive HAP or VAP is currently recruiting (Clinicaltrials.gov, Identifier NCT02019420).

Telavancin

Telavancin belongs to the class of new lipoglycopeptides and exerts a rapid, concentration-dependent, bactericidal activity against a broad-spectrum of Gram-positive pathogens, including MRSA and *S. pneumoniae*⁵⁶⁻⁵⁷. This drug is characterized by the presence of a lipophilic side chain that attaches to the bacterial membrane showing increased affinity compared with old glycopeptides. Telavancin acts through two different mechanisms of action: inhibition of bacterial wall synthesis (transglycosylation and transpeptidation) and disruption of bacterial membrane function⁵⁸.

PK/PD studies demonstrated that telavancin achieves good concentrations in ELF in healthy volunteers, with a median AUC_{ELF} approximately 75% of the free AUC_{plasma} ⁵⁹.

Non-inferiority of telavancin (10 mg/kg every 24 h) versus vancomycin (1 g every 12 h) for the treatment of HAP has been demonstrated in two Phase 3, randomized, double-blinded studies (ATTAIN studies)⁶⁰. A systematic review and meta-analysis of data coming from ABSSSI and HAP studies on telavancin, however, suggested a higher risk of nephrotoxicity and serious adverse events among telavancin-treated patients compared to vancomycin⁶¹. Particularly, an increased mortality in patients with HAP and moderate-to-severe renal impairment treated with telavancin compared to vancomycin was reported⁶². A post hoc analysis of data from the two Phase 3 ATTAIN trials demonstrated that, in the subset of patients without severe renal impairment or preexisting acute renal failure, clinical and safety outcomes were similar in the telavancin and vancomycin treatment groups⁶³. Telavancin is currently approved by EMA for the treatment of adult patients with HAP (including VAP) only for MRSA known or suspected infections and other alternative treatments are not suitable. Moreover, it is strongly suggested to restrict the use of telavancin only to patients with normal renal function⁶⁴.

Delafloxacin

Delafloxacin belongs to the class of fluoroquinolones and exerts a potent anti-MRSA activity together with a broad-spectrum activity against both Gram-positive (including penicillin-sensitive, penicillin-resistant, and levofloxacin-resistant *S. pneumoniae*, *Streptococcus pyogenes* and Enterococci) and Gram-negative pathogens (*Escherichia coli*, *Klebsiella* spp., *Haemophilus influenzae*, *Moraxella catharralis*, and quinolone-susceptible *P. aeruginosa*)⁶⁵⁻⁶⁶⁻⁶⁷. Moreover, delafloxacin is active against anaerobes and atypical respiratory tract pathogens (e.g., *Legionella*, *Chlamydia*, and *Mycoplasma*)⁶⁸⁻⁶⁹⁻⁷⁰.

Due to the peculiar dual mechanism of DNA target inhibition (DNA gyrase and topoisomerase IV), delafloxacin is characterized by a reduced probability for the selection of resistant *in vitro* mutants⁷¹.

In a neutropenic murine lung infection model delafloxacin demonstrated a high penetration into the lung compartment, as epithelial lining fluid concentrations were substantially higher than plasma ones⁷². The potential role of delafloxacin for the treatment of respiratory tract infection has been evaluated in two Phase 2 studies, with promising results. In a double-blinded, randomized, Phase 2 study, 309 outpatients affected by CAP were treated with once-daily oral administration of delafloxacin at different dosages (100, 200, and 400 mg) for 7 days, with overall clinical and bacteriological cure rates demonstrated in up to 87% of patients. Furthermore, pathogen eradication rates were higher than 90% for *H. influenzae*, *H. parainfluenzae* and other atypical bacteria, and achieved 100% for *S. aureus* and *S. pneumoniae*⁷³. The second study investigated the safety and efficacy of delafloxacin in patients with acute bacterial exacerbation of chronic bronchitis. Four different regimens were tested (100, 200, 400, and 500 mg, given orally every 24 h); clinical response was similar in the four treatment groups, with clinical and microbiological cure rates higher than 70%⁷⁴. Data coming from studies on the use of delafloxacin for the treatment of ABSSSI demonstrate that delafloxacin at the dose of 300 mg every 12h is well tolerated, with diarrhea being the most

common adverse event [111]. Moreover, in healthy volunteers doses up to 900 mg were well tolerated, without any effect on QTc prolongation⁷⁵.

Due to the broad-spectrum activity including MRSA, the availability of an oral formulation, the reduced probability for resistance selection and the good tolerability profile, delafloxacin could represent a promising option for the treatment of respiratory tract infections.

A Phase 3 study comparing delafloxacin to moxifloxacin for the treatment of adult patients with CAP (DEFINE-CABP) is currently ongoing (Clinicaltrials.gov, Identifier NCT02679573).

Iclaprim

Iclaprim is related to trimethoprim, as an inhibitor of bacterial dihydrofolate reductase, but is active against Gram-positive bacteria that are resistant to trimethoprim. Notably, iclaprim does not have to be combined with a sulphonamide, which is commonly associated with adverse events. Iclaprim has rapid in vitro bactericidal activity in time-kill studies in human plasma⁷⁶, including against MSSA/MRSA. Using clinical *S. aureus* isolates from US and Europe (2015-16), the MIC₅₀/MIC₉₀ was 0.06/0.06 for MSSA (n=304) and 0.03/0.12 for MRSA (n=314)⁷⁷. The clinical development programme for iclaprim has been protracted, reflecting changes to regulatory guidance, financial constraints by the original company (Arpida) developing the antibiotic, and then the transfer of the development rights to Motif Bio.

Two phase 3 clinical trials of iclaprim (REVIVE-1 and -2)⁷⁸⁻⁷⁹, using a fixed dosage (80 mg IV twice daily for ? days), have recently been completed in ABSSSI; these are in addition to the original phase 3 studies, in which a weight-based dosage was employed. No oral formulation of iclaprim is available. The recent clinical trials showed that iclaprim achieved early clinical response rates comparable with

vancomycin 15 mg/kg IV every 12 hours (79.6% versus 78.8%; treatment difference 0.75%, 95% CI - 3.84 to 5.35%)⁸⁰. Clinical cure rates at test of cure were also very similar (80.4% and 82.5% for patients in the iclaprim and vancomycin groups, respectively; treatment difference -2.04%, 95% CI - 6.44% to 2.36%). *S. aureus* was the most commonly isolated pathogen (n = 595), of which 45.9% were MRSA. The MIC₅₀/MIC₉₀ values for iclaprim and vancomycin for *S. aureus* isolates were 0.12/0.5 mg/L and 1/1 mg/L, respectively.

In a phase II, double-blind, multicenter study in patients with nosocomial pneumonia suspected or confirmed to be caused by Gram-positive pathogens (NCT00543608), subjects were randomized (1:1:1) to iclaprim (0.8 mg/kg IV q12h, n = 23; or 1.2 mg/kg IV q8h, n =24), or vancomycin 1 g IV q12h, n =23) for 7-14 days. The trial was ended early due to financial resource limitations, but the primary endpoint, clinical cure in the intent-to-treat (ITT) population at test of cure (TOC; 7 ± 1 days post treatment) visit, was achieved in 73.9%, 62.5%, and 52.2% of the three treatment groups, respectively. The adverse event profile of both iclaprim dosaging regimens were similar to that of vancomycin.

Motif Bio submitted a New Drug Application (NDA) for iclaprim in 2018 for the treatment of ABSSSI. The FDA has recently stated that it requires an additional clinical trial to be performed, as it has concerns about possible liver toxicity associated with iclaprim. Motif Bio has also stated its desire to develop iclaprim for hospital acquired bacterial pneumonia (HABP), including ventilator associated bacterial pneumonia (VABP), and possibly also for the treatment of *S. aureus* infection in cystic fibrosis, but new clinical trials in these indications have not commenced.

Oritavancin

Oritavancin is a lipoglycopeptide with three described mechanisms of action: inhibition of transglycosylation, inhibition of transpeptidation and disruption of cell membrane integrity. These actions likely explain the faster killing activity of oritavancin against *S. aureus*, including MRSA (at

least 99.9% killing in vitro within 1 hour; i.e. similar to that achieved by daptomycin), compared with vancomycin (similar level of kill, but requiring 24 hours)⁸¹. For clinical isolates recovered in Europe and USA between 2010-16, oritavancin inhibited 99.7-99.8% of *S. aureus* at ≤ 0.12 mg/L (oritavancin MIC_{50/90}, 0.03/0.06 mg/L)⁸². The other key attribute of oritavancin is its very long half life (~ hours), which means that one IV 1.2g dose provides sufficient blood concentrations of antibiotic to exceed the MIC of staphylococci and streptococci over at least 10 days.

Hence, a single iv dose of oritavancin is (FDA/EMA) approved for the treatment of adult patients with cSSSI/ABSSSI caused or suspected to be caused by Gram-positive microorganisms, including MRSA. Approval of oritavancin was based primarily on the two phase 3 (SOLO-I and -II) clinical trials in ABSSSI, where a single dose of oritavancin had comparable efficacy and safety compared with 7-10 days of twice-daily vancomycin⁸³⁻⁸⁴. In a pooled analysis of the results from these trials, there were 1959 patients, of whom 1067 had at least one baseline Gram-positive pathogen and 405 had MRSA⁸⁵. In these MRSA cases, at post-therapy evaluation (days 14–24), clinical success was achieved in 170/204 (83.3) and 169/201 (84.1) treated with oritavancin and vancomycin, respectively. In a real world registry of patients treated with oritavancin, almost 80% of the 112 patients (from 8 sites) had MRSA infections⁸⁶. A positive clinical response was seen in 92.8% of patients, and microbial eradication occurred in 90.0%. Only 4 (3.6%) patients were hospitalized for failure of treatment of the index infection within 28 days following oritavancin administration. Five (4.5%) patients had ≥ 1 possible drug-related AEs, but no drug-related serious AEs were reported. Notably, therefore, single dose treatment with oritavancin can avoid the need for hospital admission or shorten the length of stay.

Ceftaroline/avibactam

Ceftaroline belongs to the new class of fifth generation cephalosporins, and is characterized by a potent activity against MRSA, due to the high binding affinity for the penicillin binding protein (PBP)-2a⁸⁷. Avibactam is a new non-beta-lactam beta-lactamase inhibitor and broadens the spectrum of

activity of ceftaroline, restoring antimicrobial activity against Gram-negative pathogens expressing Ambler class A, C and some class D beta-lactamases⁸⁸. In a Phase 1 study aiming to evaluate safety, tolerability and pharmacokinetics of ceftaroline/avibactam, the compound was found to be safe and was well tolerated at total daily doses of up to 1,800 mg of each compound. Adverse events (AEs), mainly represented by diarrhea, dry mouth and headache, were mild to moderate in severity. Infusion-site reactions were the most common AEs reported after multiple intravenous dosing⁸⁹. A Phase 1 study analyzing pharmacokinetic profiles of ceftaroline and avibactam following intravenous administration of ceftaroline/avibactam in adults with augmented renal clearance has been completed (NCT01624246). A Phase 2 study comparing treatment with ceftaroline/avibactam versus doripenem for the treatment of adult patients with complicated urinary tract infections has recently been completed and results are pending (NCT01281462). Moreover, two studies investigating the potential effect of ceftaroline/ avibactam on QT interval prolongation and on intestinal flora have been completed and results are awaiting (NCT01290900 and NCT01789528).

Due to the broad-spectrum activity, including both MRSA and carbapenemases-producing Enterobacteriaceae, ceftaroline/avibactam might represent an interesting option for the treatment of infections due to MRSA, particularly when a concomitant empiric or targeted treatment against ESBL- or KPC- producing Enterobacteriaceae is required.

Omadacycline

Omadacycline is a semisynthetic antibiotic structurally related to tetracyclines, and has been approved by U.S. FDA on October 2018 for the treatment of acute bacterial skin and skin structure infections and community-acquired bacterial pneumonia. Currently approved schedule consists in a loading dose (200 mg single dose intravenously or 100 mg every 12 hours intravenously) followed by 100 mg every 24 hours intravenously or 300 mg orally once a day for 7-14 days for both ABSSTIs and CAP. For the treatment of ABSSTIs an oral loading dose of 450 mg orally once a day for two days is feasible.

Omadacycline is characterized by a broad-spectrum activity including anaerobes and difficult-to-treat aerobic pathogens, in particular MRSA, vancomycin-resistant *Enterococcus faecium* (VRE), ESBL- and carbapenemase-producing *Enterobacteriaceae*, MDR *Acinetobacter* spp., *Moraxella catharralis* and *Stenotrophomonas maltophilia*⁹⁰⁻⁹¹. Safety and pharmacokinetics of both oral and intravenous formulations have been evaluated in a Phase 1 study including 24 healthy subjects. The absolute bioavailability of the tablets was approximately 34.5% compared with intravenous formulation (a 300-mg dose of the tablet formulation produced a total exposure equivalent to that of a 100-mg intravenous dose), with a consistent inter-subject variability. Overall, omadacycline was well tolerated, with dizziness, nausea and vomiting being the most frequently reported adverse events⁹².

In a recently published, double-blind trial, omadacycline has been found non-inferior to linezolid for the treatment of acute bacterial skin and skin-structure infections, with favorable response rates at 48-72 hours of 84,8% and 85,5%, respectively⁹³. Omadacycline displays also a good penetration into both ELF and alveolar cells, with an overall magnitude of systemic exposure of omadacycline approximately 3-fold higher than that of tigecycline in plasma, ELF and alveolar cells⁹⁴. In a recently published double-blinded, phase III trial, omadacycline was found to be non-inferior compared with moxifloxacin for the treatment of community-acquired bacterial pneumonia in adults, with favorable early clinical response rates at 48-72 hours of 81,1% and 82,7%, respectively⁹⁵. In both studies omadacycline was administered at the dose of 100 mg given intravenously every 12 hours for two doses, then 100 mg given intravenously every 24 hours with an option to transition to oral omadacycline 300 mg every 24 hours after 3 days. A similar safety profile was described for the two compounds, with adverse events, mainly represented by gastrointestinal symptoms, reported in up to 45% of patients⁹⁶⁻⁹⁷.

Moreover, omadacycline is currently under evaluation for the treatment of urinary tract infections. In a recently published 1b, open label study omadacycline was administered to 31 women with cystitis for 5 days; there groups with ascending doses (group 1: 200 mg intravenously on day 1, then 300 mg orally every 24 h [q24h]; group 2: 300 mg orally every 12 h [q12h] on day 1, then 300 mg orally q24h;

group 3: 450 mg orally q12h on day 1, then 450 mg orally q24h) were evaluated. A good excretion of omadacycline in urine was reported, with favorable clinical outcomes at end of treatment in 94% of cases⁹⁸. A phase-2 study evaluating safety and efficacy of omadacycline compared to levofloxacin for the treatment of acute pyelonephritis is currently recruiting [NCT03757234].

Plazomicin

Plazomicin is a next-generation aminoglycoside that was approved by the FDA in June 2018 for the treatment of cUTIs, including pyelonephritis. Compared with the other aminoglycosides, plazomicin has been structurally modified to prevent inactivation by plasmid-borne aminoglycoside-modifying enzymes, which represent the main resistance mechanism impairing the activity of traditional aminoglycosides. For this reason, plazomicin exerts a potent in vitro bactericidal activity against MDR Enterobacteriaceae, including aminoglycoside-resistant pathogens that encode aminoglycoside-modifying enzymes, and retains activity against most carbapenemase-producing strains, including metallo-beta-lactamase producing isolates⁹⁹⁻¹⁰⁰. Plazomicin was tested against 4,825 clinical isolates collected during 2014 and 2015 in 70 U.S. hospitals as part of the ALERT (Antimicrobial Longitudinal Evaluation and Resistance Trends) program, and was found to be able to inhibit 99.2% of Enterobacteriaceae isolated at ≤ 4 $\mu\text{g/ml}$. Moreover, plazomicin, as well as other aminoglycosides, is effective against *P.aeruginosa* and *A.baumannii*¹⁰¹. Regarding Gram-positive pathogens, plazomicin displays a good activity against staphylococci (both methicillin-susceptible and methicillin-resistant strains), but possess a limited activity against *S.pneumoniae* and Enterococci¹⁰². In vitro synergy between plazomicin and piperacillin/tazobactam or ceftazidime has been reported against MDR Enterobacteriaceae, suggesting a potential role of plazomicin both as monotherapy and as combination therapy for the treatment of serious infections due to this class of pathogens¹⁰³. Moreover, synergy with carbapenems for the treatment of both MDR *A.baumannii* and MRSA has been reported¹⁰⁴⁻¹⁰⁵. Plazomicin at the dose of 15 mg/kg once daily for 5 days was found to be effective in the treatment of adults with cUTIs and acute pyelonephritis (including patients with antibiotic-resistant

Enterobacteriaceae) in a double-blind, Phase 2 study comparing plazomicin with levofloxacin¹⁰⁶. In a recently published Phase 3 plazomicin (15 mg/kg daily) was found non-inferior to meropenem (1 g every 8 hours, with the option to switch to oral levofloxacin after at least four days) for the treatment of cUTIs and acute pyelonephritis caused by Enterobacteriaceae, including multidrug-resistant strains. To note, a higher percentage of patients in the plazomicin group than in the meropenem group were found to have microbiologic eradication, and fewer patients in the plazomicin group than in the meropenem group had microbiologic recurrence (3.7% vs. 8.1%) or clinical relapse (1.6% vs. 7.1%)¹⁰⁷. In a multicenter, randomized, open-label trial tried to evaluate safety and efficacy of plazomicin compared to colistin, both in combination with tigecycline or meropenem for the treatment of serious infections (including HAP, VAP, bloodstream infections, cUTIs and acute pyelonephritis) due to carbapenem-resistant Enterobacteriaceae. Unfortunately, the study was stopped prematurely because of slow enrollment. Overall, 39 patients have been enrolled; among these, 18 were randomized in the plazomicin arm and 21 in colistin arm. Owing the small sample size, no formal hypothesis testing was performed. However, the primary end-point event, represented by death from any cause at 28 days, occurred 4/17 (24%) patients receiving plazomicin and 10/20 (50%) patients receiving colistin, with serious adverse events reported less frequently in plazomicin arm compared to colistin arm¹⁰⁸.

Lefamulin

Lefamulin (formerly known as BC-3781) is the first in class pleuromutilin antibiotic and exhibits a unique mechanism of action through inhibition of protein synthesis by binding to the peptidyl transferase center of the 50S bacterial ribosome, thus preventing the binding of transfer RNA for peptide transfer¹⁰⁹. Lefamulin exerts a potent activity against both Gram-positive pathogens (including MRSA and VRE) and atypical organisms associated with CAP (e.g. *Mycoplasma pneumoniae*, *Legionella pneumophila*, and *Chlamydia pneumoniae*). Additionally, lefamulin retains activity against multidrug-resistant *Neisseria gonorrhoeae* and *Mycoplasma genitalium*¹¹⁰.

Both population pharmacokinetic models and murine models demonstrated a high and rapid penetration into ELF, irrespective of the route of administration (intravenous or oral)¹¹¹⁻¹¹². In a recently published Phase 3, multicenter, randomized, double blind trial, lefamulin at 150 mg intravenously every 12 hours (with the option to switch to oral treatment after 6 doses) was found non inferior to moxifloxacin at 400 mg intravenously every 24 hours for the treatment of community-acquired bacterial pneumonia¹¹³. Due to the peculiar spectrum of activity, lefamulin might represent an interesting option for the treatment of community acquired pneumonia, particularly in patients presenting with risk factors for MRSA infection.

Conclusions

A number of new drugs for the treatment of MRSA infections has been recently approved or are in advanced stage of development (**Table 1**). The most attractive characteristic of new drugs is represented by the broad spectrum of activity against multi-drug resistant pathogens; moreover, new compounds in most cases are characterized by favorable toxicity profiles compared with old drugs currently used in clinical practice. Some of the new antimicrobials will be also available as oral formulations, with the potential for oral shift even in infections due to resistant pathogens.

Expert opinion

The spread of MRSA in the last decades has inevitably changed the therapeutic approach to this infection. Most β -lactams (with the exception of fifth-generation cephalosporins) have become ineffective against proven MRSA, as well as potentially ineffective in the empirical treatment of infections in patients with risk factors for MRSA. Fortunately, several alternatives are now available for suspected or proven MRSA, consisting of the abovementioned fifth-generation cephalosporins combinations and various non- β -lactam antibiotics (e.g., oxazolidinones, glycopeptides, lipopeptides, lipoglycopeptides, delafloxacin). Furthermore, MRSA may remain susceptible to SMX/TMP, tetracyclines, and/or clindamycin.

Since noninferiority was frequently the rule in phase-3 RCT evaluating the efficacy of the anti-MRSA agents mentioned above in the treatment of MRSA infections, other factors may become preeminent when selecting the appropriate antibiotic on a patient-by-patient basis: (i) history of hypersensitivity reactions; (ii) availability of oral formulation for out-patient treatment; (iii) possibility of switch from intravenous to oral therapy and early discharge; (iv) spectrum of activity (e.g., for suspected or proven polymicrobial infections); (v) safety profile of the different therapeutic options in light of the patient's baseline comorbidities and risks for toxicity. Very importantly, with regard to this latter point (toxicity), adequate knowledge of the peculiar safety profile of each drug is essential for guiding monitoring and management of AE, in turn reducing any possible unfavorable impact of toxicity on patients' outcomes.

Data regarding the safety and the efficacy of newer molecules for the treatment of MRSA infections in particular conditions/populations (e.g. liver failure, renal disease, pregnancy, diabetic, children, elderly) are scant, thus dedicated studies are warranted. In our opinion, novel antibiotics with reduced toxicity could be an important option in the elderly (considering the usually non-negligible burden of comorbidities in this population) and in discharged patients. Two examples may be those of well-tolerated fifth-generation cephalosporins in hospitalized patients with impaired renal function, to avoid additional nephrotoxicity due to glycopeptides, and of tedizolid in non-closely monitored discharged patients, by possibly enabling earlier discharge with also a lower risk of thrombocytopenia in comparison with linezolid.

The increasing challenge of antimicrobial dosing for the treatment of MRSA infections in the obese population is also worth mentioning. Notably, the majority of the newly approved molecules does not provide specific indications for dosing in this patient population. In our opinion, drugs with a weight-driven dosage should be preferred. Whenever a β lactam is required, continuous/extended infusions, higher doses, or more frequent dosing should be considered, together with TDM to avoid underexposure. Dose adjustments in obese patients are not recommended for novel, long acting

molecules. However, very limited data have been published to date, and the results of a phase I study evaluating the pharmacokinetics of telavancin in obese subjects are pending.

In diabetic patients, moxifloxacin and delafloxacin could be among the options to be considered for diabetic foot infections, owing to their high penetration in the perinecrotic tissue and the bone, and their good oral bioavailability.

In the next five years, we expect to witness a continuous refining of therapeutic algorithm for maximizing the cost-effectiveness of the treatment of MRSA infections, in which, considering the similar efficacy of novel treatment, potential toxicity will play a critical role in establishing the best available therapy for each specific patient, together with consideration regarding the possibility of avoiding hospitalization or allowing switch from intravenous to oral therapy and early discharge.

Against this backdrop, it will be

interesting to define also the specific place in therapy of other novel agents that could be available in the future, by accurately considering and weighing their safety data from currently ongoing RCT for MRSA infections.

Disclosures

Outside the submitted work, MB has participated in advisory boards and/or received speaker honoraria from Achaogen, Angelini, Astellas, Bayer, Basilea, Biomerieux, Cidara, Gilead, Menarini, MSD, Nabriva, Paratek, Pfizer, Roche, Melinta, Shionogi, Tetrphase, VenatoRx and Vifor and has received study grants from Angelini, Basilea, Astellas, Shionogi, Cidara, Melinta, Gilead, Pfizer and MSD.

REFERENCES

¹ https://cddep.org/sites/default/files/swa_2015_final.pdf

² Diekema DJ, Pfaller MA, Shortridge D, Zervos M, Jones RN. Twenty-Year Trends in Antimicrobial Susceptibilities Among *Staphylococcus aureus* From the SENTRY Antimicrobial Surveillance Program. *Open Forum Infect Dis.* 2019 Mar 15;6(Suppl 1):S47-S53. doi: 10.1093/ofid/ofy270. eCollection 2019 Mar. Erratum in: *Open Forum Infect Dis.* 2019 May 20;6(5):ofz202

³ <https://ecdc.europa.eu/sites/portal/files/documents/EARS-Net-report-2017-update-jan-2019.pdf>

⁴ Sader HS, Mendes RE, Jones RN, Flamm RK. Antimicrobial susceptibility patterns of community- and hospital-acquired methicillin-resistant *Staphylococcus aureus* from United States Hospitals: results from the AWARE Ceftaroline Surveillance Program (2012-2014). *Diagn Microbiol Infect Dis.* 2016;86:76-9.

⁵ Bassetti M, Righi E, Peghin M, et al. Is first-line antimicrobial therapy still adequate to treat MRSA in the ICU? A report from a highly endemic country. *Crit Care.* 2016;20:246

⁶ Nillius D, von Müller L, Wagenpfeil S, Klein R, Herrmann M. Methicillin-Resistant *Staphylococcus aureus* in Saarland, Germany: The Long-Term Care Facility Study. *PLoS One.* 2016;11:e0153030

⁷ European Centre for Disease Prevention and Control (ECDC) network: HEALTHCARE-ASSOCIATED INFECTIONS. Available at:

http://ecdc.europa.eu/en/activities/surveillance/HAI/Documents/2008_HAI_%20special_chapter.pdf

(Last accessed 05 June 2019).

⁸ Friedman ND, Kaye KS, Stout JE et al. Health care--associated bloodstream infections in adults: a reason to change the accepted definition of community-acquired infections. *Ann Intern Med* 2002;137:791-7

⁹ Packer S, Pichon B, Thompson S, Neale J, Njoroge J, Kwiatkowska RM, Oliver I, Telfer M, Doumith M, Buunaaisie C, Heinsbroek E, Hopewell-Kelly N, Desai M, Hope V, Williams OM, Kearns A, Hickman M, Gobin M. Clonal expansion of community-associated methicillin-resistant *Staphylococcus aureus* (MRSA) in people who inject drugs (PWID): prevalence, risk factors and molecular epidemiology, Bristol, United Kingdom, 2012 to 2017. *Euro Surveill*. 2019 Mar;24(13). doi: 10.2807/1560-7917

¹⁰ Ruhe JJ, Smith N, Bradsher RW, Menon A. Community-onset methicillin-resistant *Staphylococcus aureus* skin and soft-tissue infections: impact of antimicrobial therapy on outcome. *Clin Infect Dis* 2007;44:777-84

¹¹ Eagye KJ, Kim A, Laohavaleeson S et al. Surgical site infections: does inadequate antibiotic therapy affect patient outcomes? *Surg Infect (Larchmt)* 2009;10:323-31

¹² Klein EY, Jiang W, Mojica N, Tseng KK, McNeill R, Cosgrove SE, Perl TM. National Costs Associated With Methicillin-Susceptible and Methicillin-Resistant *Staphylococcus aureus* Hospitalizations in the United States, 2010-2014. *Clin Infect Dis*. 2019 Jan 1;68(1):22-28. doi: 10.1093/cid/ciy399

¹³ Stewardson AJ, Allignol A, Beyersmann J, Graves N, Schumacher M, Meyer R, Tacconelli E, De Angelis G, Farina C, Pezzoli F, Bertrand X, Gbaguidi-Haore H, Edgeworth J, Tosas O, Martinez JA,

Ayala-Blanco MP, Pan A, Zoncada A, Marwick CA, Nathwani D, Seifert H, Hos N, Hagel S, Pletz M, Harbarth S; TIMBER Study Group. The health and economic burden of bloodstream infections caused by antimicrobial-susceptible and non-susceptible Enterobacteriaceae and *Staphylococcus aureus* in European hospitals, 2010 and 2011: a multicenter retrospective cohort study. *Euro Surveill.* 2016 Aug 18;21(33). doi: 10.2807/1560-7917.ES.2016.21.33.30319

¹⁴ Diekema DJ, Hsueh PR, Mendes RE, Pfaller MA, Rolston KV, Sader HS, Jones RN. The Microbiology of Bloodstream Infection: 20-Year Trends from the SENTRY Antimicrobial Surveillance Program. *Antimicrob Agents Chemother.* 2019 Apr 22. pii: AAC.00355-19. doi: 10.1128/AAC.00355-19. [Epub ahead of print]

¹⁵ Sader HS, Streit JM, Carvalhaes CG, Huband MD, Pfaller MA. Frequency and antimicrobial susceptibility of bacterial isolates from patients hospitalized with community-acquired skin and skin-structure infection in Europe, Asia and Latin America. *J Glob Antimicrob Resist.* 2018 Nov 17;17:103-108. doi: 10.1016/j.jgar.2018.11.013. [Epub ahead of print]

¹⁶ Stacey HJ, Clements CS, Welburn SC, Jones JD. The prevalence of methicillin-resistant *Staphylococcus aureus* among diabetic patients: a meta-analysis. *Acta Diabetol.* 2019 Apr 6. doi: 10.1007/s00592-019-01301-0. [Epub ahead of print]

¹⁷ Bassetti M, Righi E, Vena A, Graziano E, Russo A, Peghin M. Risk stratification and treatment of ICU-acquired pneumonia caused by multidrug resistant/extensively drug-resistant/pandrug-resistant bacteria. *Curr Opin Crit Care.* 2018 Oct;24(5):385-393. doi: 10.1097/MCC.0000000000000534

-
- ¹⁸ Cillóniz C, Dominedò C, Nicolini A, Torres A. PES Pathogens in Severe Community-Acquired Pneumonia. *Microorganisms*. 2019 Feb 12;7(2)
- ¹⁹ Torres A, Chalmers JD, Dela Cruz CS, Dominedò C, Kollef M, Martin-Loeches I, Niederman M, Wunderink RG. Challenges in severe community-acquired pneumonia: a point-of-view review. *Intensive Care Med*. 2019 Feb;45(2):159-171. doi: 10.1007/s00134-019-05519-y.
- ²⁰ Berger A, Oster G, Edelsberg J, Huang X, Weber DJ. Initial treatment failure in patients with complicated skin and skin structure infections. *Surg Infect (Larchmt)*. 2013;14:304-12
- ²¹ Ostermann H, Blasi F, Medina J, Pascual E, McBride K, Garau J; REACH study group. Resource use in patients hospitalized with complicated skin and soft tissue infections in Europe and analysis of vulnerable groups: the REACH study. *J Med Econ*. 2014;17:719-29
- ²² Kumar A, Ellis P, Arabi Y, Roberts D, Light B, Parrillo JE, Dodek P, Wood G, Kumar A, Simon D, Peters C, Ahsan M, Chateau D; Cooperative Antimicrobial Therapy of Septic Shock Database Research Group. Initiation of inappropriate antimicrobial therapy results in a fivefold reduction of survival in human septic shock. *Chest* 2009;136:1237-48
- ²³ Vallés J, Rello J, Ochagavía A, Garnacho J, Alcalá MA. Community-acquired bloodstream infection in critically ill adult patients: impact of shock and inappropriate antibiotic therapy on survival. *Chest* 2003;123:1615-24

²⁴ Butler-Laporte G, Cheng MP, McDonald EG, Lee TC. Screening swabs surpass traditional risk factors as predictors of MRSA bacteremia. *BMC Infect Dis.* 2018 Jun 11;18(1):270. doi: 10.1186/s12879-018-3182-x.

²⁵ Gunderson CG, Holleck JL, Chang JJ, Merchant N, Lin S, Gupta S. Diagnostic accuracy of methicillin-resistant *Staphylococcus aureus* nasal colonization to predict methicillin-resistant *S aureus* soft tissue infections. *Am J Infect Control.* 2016;44:1176-1177

²⁶ Liu C, Bayer A, Cosgrove SE, Daum RS, Fridkin SK, Gorwitz RJ, Kaplan SL, Karchmer AW, Levine DP, Murray BE, J Rybak M, Talan DA, Chambers HF; Infectious Diseases Society of America. Clinical practice guidelines by the infectious diseases society of america for the treatment of methicillin-resistant *Staphylococcus aureus* infections in adults and children. *Clin Infect Dis.* 2011 Feb 1;52(3):e18-55. doi: 10.1093/cid/ciq146.

²⁷ Stevens DL, Bisno AL, Chambers HF et al. Infectious Diseases Society of America. Practice guidelines for the diagnosis and management of skin and soft tissue infections: 2014 update by the Infectious Diseases Society of America. *Clin Infect Dis* 2014;59:e10-52. Erratum in: *Clin Infect Dis* 2015;60:1448

²⁸ Lodise TP, Rosenkranz SL, Finnemeyer M, Evans S, Sims M, Zervos MJ, Creech CB, Patel PC, Keefer M, Riska P, Silveira FP, Scheetz M, Wunderink RG, Rodriguez M, Schrank J, Bleasdale SC, Schultz S, Barron M, Stapleton A, Wray D, Chambers H, Fowler V, Holland TL; Antibacterial Resistance Leadership Group. The Emperor's New Clothes: Prospective Observational Evaluation of the Association between Initial Vancomycin Exposure and Failure Rates among Adult Hospitalized

Patients with MRSA Bloodstream Infections (PROVIDE). *Clin Infect Dis*. 2019 Jun 3. pii:vciz460. doi: 10.1093/cid/ciz460. [Epub ahead of print]

²⁹ Ye ZK, Li C, Zhai SD. Guidelines for therapeutic drug monitoring of vancomycin: a systematic review. *PLoS One* 2014;9:e99044

³⁰ Gonzalez-Ruiz A, Seaton RA, Hamed K. Daptomycin: an evidence-based review of its role in the treatment of Gram-positive infections. *Infect Drug Resist*. 2016;9:47-58 // Seaton RA, Gonzalez-Ruiz A, Cleveland KO, Couch KA, Pathan R, Hamed K. Real-world daptomycin use across wide geographical regions: results from a pooled analysis of CORE and EU-CORE. *Ann Clin Microbiol Antimicrob*. 2016;15:18

³¹ Timbrook TT, Caffrey AR, Luther MK, Lopes V, LaPlante KL. Association of Higher Daptomycin Dose (7 mg/kg or Greater) with Improved Survival in Patients with Methicillin-Resistant *Staphylococcus aureus* Bacteremia. *Pharmacotherapy*. 2018 Feb;38(2):189-196. doi: 10.1002/phar.2070. Epub 2018 Jan 8. PubMed PMID: 29235661

³² Gonzalez-Ruiz A, Seaton RA, Hamed K. Daptomycin: an evidence-based review of its role in the treatment of Gram-positive infections. *Infect Drug Resist*. 2016 Apr 15;9:47-58. doi: 10.2147/IDR.S99046.

³³ Geriak M, Haddad F, Rizvi K, Rose W, Kullar R, LaPlante K, Yu M, Vasina L, Ouellette K, Zervos M, Nizet V, Sakoulas G. Clinical Data on Daptomycin plus Ceftaroline versus Standard of Care Monotherapy in the Treatment of Methicillin-Resistant *Staphylococcus aureus* Bacteremia. *Antimicrob Agents Chemother*. 2019 Apr 25;63(5). pii: e02483-18. doi: 10.1128/AAC.02483-18

³⁴ Torres A, Niederman MS, Chastre J, Ewig S, Fernandez-Vandellos P, Hanberger H, Kollef M, Li Bassi G, Luna CM, Martin-Loeches I, Paiva JA, Read RC, Rigau D, François Timsit J, Welte T, Wunderink R. Summary of the international clinical guidelines for the management of hospital-acquired and ventilator-acquired pneumonia. *ERJ Open Res.* 2018 Jun 26;4(2). pii: 00028-2018. doi: 10.1183/23120541.00028-2018.

³⁵ Bassetti M, Baguneid M, Bouza E, et al. European perspective and update on the management of complicated skin and soft tissue infections due to methicillin-resistant *Staphylococcus aureus* after more than 10 years of experience with linezolid. *Clin Microbiol Infect.* 2014;20 Suppl 4:3-18

³⁶ Pea F, Viale P, Cojutti P, Del Pin B, Zamparini E, Furlanut M. Therapeutic drug monitoring may improve safety outcomes of long-term treatment with linezolid in adult patients. *J Antimicrob Chemother.* 2012 Aug;67(8):2034-42. doi: 10.1093/jac/dks153.

³⁷ Pea F, Cojutti PG, Baraldo M. A 10-Year Experience of Therapeutic Drug Monitoring (TDM) of Linezolid in a Hospital-wide Population of Patients Receiving Conventional Dosing: Is there Enough Evidence for Suggesting TDM in the Majority of Patients? *Basic Clin Pharmacol Toxicol.* 2017 Oct;121(4):303-308. doi: 10.1111/bcpt.12797.

³⁸ Go AC, Golightly LK, Barber GR, Barron MA. Linezolid interaction with serotonin reuptake inhibitors: report of two cases and incidence assessment. *Drug Metabol Drug Interact* 2010;25:41-7

³⁹ Clark RB, Hunt DK, He M, et al. Fluorocyclines. 2. Optimization of the C-9 side-chain for antibacterial activity and oral efficacy. *J Med Chem* 2012; 55:606 – 622.

-
- ⁴⁰ Abdallah M, Olafisoye O, Cortes C, et al. Activity of eravacycline against Enterobacteriaceae and *Acinetobacter baumannii*, including multidrug-resistant isolates, from New York City. *Antimicrob Agents Chemother* 2015; 59:1802 – 1805.
- ⁴¹ Zhanel GG, Baxter MR, Adam HJ, et al. In vitro activity of eravacycline against 2213 Gram-negative and 2424 Gram-positive bacterial pathogens isolated in Canadian hospital laboratories: CANWARD surveillance study 2014-2015. *Diagn Microbiol Infect Dis*. 2017
- ⁴² Seifert H, Stefanik D, Sutcliffe JA, et al. In-vitro activity of the novel fluorocycline eravacycline against carbapenem non-susceptible *Acinetobacter baumannii*. *Int J Antimicrob Agents*. 2018;51:62-64
- ⁴³ Snyderman DR, McDermott LA, Jacobus NV, et al. Evaluation of the in vitro activity of eravacycline against a broad spectrum of recent clinical anaerobic isolates. *Antimicrob Agents Chemother*. 2018.
- ⁴⁴ Sutcliffe JA, O'Brien W, Fyfe C, et al. Antibacterial activity of eravacycline (TP-434), a novel fluorocycline, against hospital and community pathogens. *Antimicrob Agents Chemother* 2013; 57:5548 – 5558.
- ⁴⁵ Bassetti M, Righi E. Eravacycline for the treatment of intra-abdominal infections. *Expert Opin Investig Drugs* 2014;23:1575 – 1584
- ⁴⁶ Solomkin J, Evans D, Slepavicius A, Lee P, et al. Assessing the Efficacy and Safety of Eravacycline vs Ertapenem in Complicated Intra-abdominal Infections in the Investigating Gram-Negative Infections Treated With Eravacycline (IGNITE 1) Trial: A Randomized Clinical Trial. *JAMA Surg*. 2017;152:224-232.

-
- ⁴⁷ Connors KP, Housman ST, Pope JS, et al. Phase I, open-label, safety and & pharmacokinetic study to assess bronchopulmonary disposition of intravenous eravacycline in healthy men and women. *Antimicrob Agents Chemother* 2014; 58:2113 – 2118
- ⁴⁸ Grossman TH, Murphy TM, Slee AM, et al. Eravacycline (TP-434) is efficacious in animal models of infection. *Antimicrob Agents Chemother* 2015; 59:2567 – 2571.
- ⁴⁹ Li S, Guo Y, Zhao C, et al. In vitro activities of tedizolid compared with other antibiotics against Gram-positive pathogens associated with hospital-acquired pneumonia, skin and soft tissue infection and bloodstream infection collected from 26 hospitals in China. *J Med Microbiol.* 2016;65:1215-1224.
- ⁵⁰ Lodise TP, Fang E, Minassian SL, Prokocimer PG. Platelet profile in patients with acute bacterial skin and skin structure infections receiving tedizolid or linezolid: findings from the Phase 3 ESTABLISH clinical trials. *Antimicrob Agents Chemother* 2014; 58:7198 – 7204.
- ⁵¹ Shorr AF, Lodise TP, Corey GR, et al. Analysis of the Phase 3 ESTABLISH trials of tedizolid versus linezolid in acute bacterial skin and skin structure infections. *Antimicrob Agents Chemother* 2015; 59:864 – 871
- ⁵² Shaw KJ, Barbachyn MR. The oxazolidinones: past, present, and future. *Ann NY Acad Sci* 2011; 1241:48 – 70
- ⁵³ Flanagan S, Passarell J, Lu Q. Tedizolid population pharmacokinetics, exposure response, and target attainment. *Antimicrob Agents Chemother* 2014; 58:6462 – 6470
- ⁵⁴ Lodise TP, Drusano GL. Use of pharmacokinetic/pharmacodynamic systems analyses to inform dose selection of tedizolid phosphate. *Clin Infect Dis* 2014; 58(Suppl 1):S28–S34

-
- ⁵⁵ Tessier PR, Keel RA, Hagihara M, et al. Comparative in vivo efficacies of epithelial lining fluid exposures of tedizolid, linezolid, and vancomycin for methicillin-resistant *Staphylococcus aureus* in a mouse pneumonia model. *Antimicrob Agents Chemother* 2012; 56:2342 – 2346
- ⁵⁶ Smith JR, Barber KE, Hallesy J, et al. Telavancin demonstrates activity against methicillin-resistant *Staphylococcus aureus* isolates with reduced susceptibility to vancomycin, daptomycin, and linezolid in broth microdilution MIC and one-compartment pharmacokinetic/pharmacodynamic models. *Antimicrob Agents Chemother* 2015; 59:5529 – 5534
- ⁵⁷ Pfaller MA, Mendes RE, Sader HS, Jones RN. Telavancin activity against Gram-positive bacteria isolated from respiratory tract specimens of patients with nosocomial pneumonia. *J Antimicrob Chemother* 2010; 65:2396 – 2404
- ⁵⁸ Zhanel GG, Calic D, Schweizer F, et al. New lipoglycopeptides: a comparative review of dalbavancin, oritavancin and telavancin. *Drugs* 2010; 70:859 – 886; Erratum in: *Drugs* 2011;71:526.
- ⁵⁹ Lodise TP Jr, Gotfried M, Barriere S, Drusano GL. Telavancin penetration into human epithelial lining fluid determined by population pharmacokinetic modeling and Monte Carlo simulation. *Antimicrob Agents Chemother* 2008; 52:2300 – 2304
- ⁶⁰ Rubinstein E, Lalani T, Corey GR, et al., ATTAIN Study Group. Telavancin versus vancomycin for hospital-acquired pneumonia due to gram-positive pathogens. *Clin Infect Dis* 2011; 52:31 – 40.
- ⁶¹ Polyzos KA, Mavros MN, Vardakas KZ, et al. Efficacy and safety of telavancin in clinical trials: a systematic review and meta-analysis. *PLoS One* 2012; 7:e41870.

⁶² Barriere SL. The ATTAIN trials: efficacy and safety of telavancin compared with vancomycin for the treatment of hospital-acquired and ventilator-associated bacterial pneumonia. *Future Microbiol* 2014; 9:281 – 289.

⁶³ Torres A, Rubinstein E, Corey GR, et al. Analysis of Phase 3 telavancin nosocomial pneumonia data excluding patients with severe renal impairment and acute renal failure. *J Antimicrob Chemother* 2014; 69:1119 – 1126.

⁶⁴ Masterton R, Cornaglia G, Courvalin P, et al. The clinical positioning of telavancin in Europe. *Int J Antimicrob Agents* 2015; 45:213 – 220

⁶⁵ Almer LS, Hoffrage JB, Keller EL, et al. In vitro and bactericidal activities of ABT-492, a novel fluoroquinolone, against Gram-positive and Gram-negative organisms. *Antimicrob Agents Chemother* 2004; 48:2771– 2777.

⁶⁶ Gunderson SM, Hayes RA, Quinn JP, Danziger LH. In vitro pharmacodynamic activities of ABT-492, a novel quinolone, compared to those of levofloxacin against *Streptococcus pneumoniae*, *Haemophilus influenzae*, and *Moraxella catarrhalis*. *Antimicrob Agents Chemother* 2004; 48:203 – 208

⁶⁷ Pfaller MA, Sader HS, Rhomberg PR, Flamm RK. In Vitro Activity of Delafloxacin against Contemporary Bacterial Pathogens from the United States and Europe, 2014. *Antimicrob Agents Chemother*. 2017;61. Erratum in: *Antimicrob Agents Chemother*. 2018 Jan 25;62(2)

⁶⁸ Goldstein EJ, Citron DM, Merriam CV, et al. In vitro activities of ABT-492, a new fluoroquinolone, against 155 aerobic and 171 anaerobic pathogens isolated from antral sinus puClinicalTrials.Gov,

IdentifierNCTure specimens from patients with sinusitis. *Antimicrob Agents Chemother* 2003; 47:3008–3011.

⁶⁹ Hammerschlag MR, Roblin PM. The in vitro activity of a new fluoro-quinolone, ABT-492, against recent clinical isolates of *Chlamydia pneumoniae*. *J Antimicrob Chemother* 2004; 54:281–282.

⁷⁰ Waites KB, Crabb DM, Duffy LB. Comparative in vitro susceptibilities and bactericidal activities of investigational fluoroquinolone ABT-492 and other antimicrobial agents against human mycoplasmas and ureaplasmas. *Antimicrob Agents Chemother* 2003; 47:3973–3975.

⁷¹ Remy JM, Tow-Keogh CA, McConnell TS, et al. Activity of delafloxacin against methicillin-resistant *Staphylococcus aureus*: resistance selection and characterization. *J Antimicrob Chemother* 2012; 67:2814–2820

⁷² Thabit AK, Crandon JL, Nicolau DP. Pharmacodynamic and pharmacokinetic profiling of delafloxacin in a murine lung model against community-acquired respiratory tract pathogens. *Int J Antimicrob Agents*. 2016;48:535-541

⁷³ Longcor J, Hopkins S, Wickler M, Laurence L. A Phase 2 study of the safety and efficacy of oral delafloxacin (DLX) in community acquired pneumonia (CAP). Presented at ID Week 2012; San Diego, California, USA.

⁷⁴ O’Riordan W, Mehra P, Manos P, et al. A randomized Phase 2 study comparing two doses of delafloxacin with tigecycline in adults with complicated skin and skin-structure infections. *Int J Infect Dis* 2015; 30:67–73

⁷⁵ Litwin JS, Benedict MS, Thorn MD, et al. A thorough QT study to evaluate the effects of therapeutic and supratherapeutic doses of delafloxacin on cardiac repolarization. *Antimicrob Agents Chemother* 2015; 59:3469 – 3473

⁷⁶ Ader HS, Fritsche TR, Jones RN. Potency and bactericidal activity of iclaprim against recent clinical gram-positive isolates. *Antimicrob Agents Chemother* 2009;53:2171–5.

⁷⁷ Huang DB, Magnet S, De Angelis S, Holland TL, File TM Jr, Dryden M, Corey GR, Torres A, Wilcox MH. Surveillance of iclaprim activity: in vitro susceptibility of Gram-positive skin infection pathogens collected from 2015 to 2016 from North America and Europe. *Diagn Microbiol Infect Dis*. 2019 Feb;93(2):154-158.

⁷⁸ Huang DB, O'Riordan W, Overcash JS, Heller B. A phase 3, Randomized, double-blind, multicenter study to Evaluate the safety and efficacy of intravenous Iclaprim versus Vancomycin for the treatment of acute bacterial skin and skin structure infections suspected or confirmed to be due to Gram-positive pathogens: REVIVE-1. *Clin Infect Dis* 2018b;66:1222–9.

⁷⁹ Holland TL, O'Riordan W, McManus A, Shin E, Borghei A, File Jr TM, Wilcox MH, Torres A, Dryden M, Lodise T, Oguri T, Corey GR, McLeroth P, Shukla R, Huang DB. Phase 3, Randomized, Double-Blind, Multicenter Study To Evaluate the Safety and Efficacy of Intravenous Iclaprim versus Vancomycin for Treatment of Acute Bacterial Skin and Skin Structure Infections Suspected or Confirmed To Be Due to Gram-Positive Pathogens (REVIVE-2 Study). *Antimicrob Agents Chemother* 2018;26(62):e02580-17.

⁸⁰ Huang DB, Corey GR, Holland TL, Lodise T, O'Riordan W, Wilcox MH, File TM Jr, Dryden M, Balsler B, Desplats E, Torres A. Pooled analysis of the phase 3 REVIVE trials: randomised, double-blind studies to evaluate the safety and efficacy of iclaprim versus vancomycin for treatment of acute bacterial skin and skin-structure infections. *Int J Antimicrob Agents*. 2018 Aug;52(2):233-240.

⁸¹ Sweeney D, Shinabarger DL, Arhin FF, Belley A, Moeck G, Pillar CM. Comparative in vitro activity of oritavancin and other agents against methicillin-susceptible and methicillin-resistant *Staphylococcus aureus*. *Diagn Microbiol Infect Dis*. 2017 Feb;87(2):121-128.

⁸² Mendes RE, Sader HS, Castanheira M, Flamm RK. Distribution of main Gram-positive pathogens causing bloodstream infections in United States and European hospitals during the SENTRY Antimicrobial Surveillance Program (2010-2016): concomitant analysis of oritavancin in vitro activity. *J Chemother*. 2018 Sep;30(5):280-289.

⁸³ Corey GR, Kabler H, Mehra P, Gupta S, Overcash JS, Porwal A, et al. Single-dose oritavancin in the treatment of acute bacterial skin infections. *N Engl J Med* 2014;370:2180–90.

⁸⁴ Corey GR, Good S, Jiang H, Moeck G, Wikler M, Green S, et al. Single-dose oritavancin versus 7–10 days of vancomycin in the treatment of Gram-positive acute bacterial skin and skin structure infections: the SOLO II noninferiority study. *Clin Infect Dis* 2015;60:254–62.

⁸⁵ Corey GR, Arhin FF, Wikler MA, Sahm DF, Kreiswirth BN, Mediavilla JR, Good S, Fiset C, Jiang H, Moeck G, Kabler H, Green S, O'Riordan W; SOLO I, SOLO II Investigators. Pooled analysis of single-dose oritavancin in the treatment of acute bacterial skin and skin-structure infections caused by

Gram-positive pathogens, including a large patient subset with methicillin-resistant *Staphylococcus aureus*. *Int J Antimicrob Agents*. 2016 Nov;48(5):528-534.

⁸⁶ Redell M, Moeck G, Lucasti C, Durso S, Kennedy C, Fusaro K, Loutit J, Dudley M. A Real-world Patient Registry for Oritavancin Demonstrates Efficacy and Safety Consistent With the Phase 3 SOLO Program. *Open Forum Infect Dis*. 2018 Mar 19;5(6):ofy051.

⁸⁷ Saravolatz LD, Stein GE, Johnson LB. Ceftaroline: a novel cephalosporin with activity against methicillin-resistant *Staphylococcus aureus*. *Clin Infect Dis* 2011;52:1156-63

⁸⁸ Nichols WW, Newell P, Critchley IA, Riccobene T, Das S. Avibactam Pharmacokinetic/Pharmacodynamic Targets. *Antimicrob Agents Chemother*. 2018 May 25;62(6). pii: e02446-17. doi: 10.1128/AAC.02446-17

⁸⁹ Riccobene TA, Su SF, Rank D (2013) Single- and Multiple-Dose Study To Determine the Safety, Tolerability, and Pharmacokinetics of Ceftaroline Fosamil in Combination with Avibactam in Healthy Subjects *Antimicrob Agents Chemother*; 57: 1496–1504

⁹⁰ Pfaller MA, Huband MD, Shortridge D, Flamm RK (2018) Surveillance of Omadacycline Activity Tested against Clinical Isolates from the United States and Europe as Part of the 2016 SENTRY Antimicrobial Surveillance Program. *Antimicrob Agents Chemother*;62

⁹¹ Huband MD, Pfaller MA, Shortridge D, Flamm RK. Surveillance of omadacycline activity tested against clinical isolates from the United States and Europe: Results from the SENTRY Antimicrobial Surveillance Programme, 2017. *J Glob Antimicrob Resist*. 2019 Feb 27. pii: S2213-7165(19)30057-8.

⁹² Sun H, Ting L, Machineni S, et al (2016) Randomized, Open-Label Study of the Pharmacokinetics and Safety of Oral and Intravenous Administration of Omadacycline to Healthy Subjects. *Antimicrob Agents Chemother*;60:7431-7435

⁹³ O'Riordan W, Green S, Overcash JS, Puljiz I, Metallidis S, Gardovskis J, Garrity-Ryan L, Das AF, Tzanis E, Eckburg PB, Manley A, Villano SA, Steenbergen JN, Loh E. Omadacycline for Acute Bacterial Skin and Skin-Structure Infections. *N Engl J Med*. 2019 Feb 7;380(6):528-538

⁹⁴ Gotfried MH, Horn K, Garrity-Ryan L, et al (2017) Comparison of Omadacycline and Tigecycline Pharmacokinetics in the Plasma, Epithelial Lining Fluid, and Alveolar Cells of Healthy Adult Subjects. *Antimicrob Agents Chemother*;61

⁹⁵ Stets R, Popescu M, Gonong JR, Mitha I, Nseir W, Madej A, Kirsch C, Das AF, Garrity-Ryan L, Steenbergen JN, Manley A, Eckburg PB, Tzanis E, McGovern PC, Loh E. Omadacycline for Community-Acquired Bacterial Pneumonia. *N Engl J Med*. 2019 Feb 7;380(6):517-527

⁹⁶ O'Riordan W, Green S, Overcash JS, Puljiz I, Metallidis S, Gardovskis J, Garrity-Ryan L, Das AF, Tzanis E, Eckburg PB, Manley A, Villano SA, Steenbergen JN, Loh E. Omadacycline for Acute Bacterial Skin and Skin-Structure Infections. *N Engl J Med*. 2019 Feb 7;380(6):528-538

⁹⁷ Stets R, Popescu M, Gonong JR, Mitha I, Nseir W, Madej A, Kirsch C, Das AF, Garrity-Ryan L, Steenbergen JN, Manley A, Eckburg PB, Tzanis E, McGovern PC, Loh E. Omadacycline for Community-Acquired Bacterial Pneumonia. *N Engl J Med*. 2019 Feb 7;380(6):517-527

-
- ⁹⁸ Overcash JS, Bhiwandi P, Garrity-Ryan L, Steenbergen J, Bai S, Chitra S, Manley A, Tzanis E. Pharmacokinetics, Safety, and Clinical Outcomes of Omadacycline in Women with Cystitis: Results from a Phase 1b Study. *Antimicrob Agents Chemother.* 2019 Apr 25;63(5)
- ⁹⁹ Serio AW, Keepers T, Krause KM. Plazomicin Is Active Against Metallo- β -Lactamase-Producing Enterobacteriaceae. *Open Forum Infect Dis.* 2019 Mar 12;6(4):ofz123
- ¹⁰⁰ Galani I, Nafplioti K, Adamou P, Karaiskos I, Giamarellou H, Souli M; Study Collaborators. Nationwide epidemiology of carbapenem resistant *Klebsiella pneumoniae* isolates from Greek hospitals, with regards to plazomicin and aminoglycoside resistance. *BMC Infect Dis.* 2019 Feb 15;19(1):167
- ¹⁰¹ Castanheira M, Davis AP, Mendes RE, Serio AW, Krause KM, Flamm RK (2018) In Vitro Activity of Plazomicin against Gram-Negative and Gram-Positive Isolates Collected from U.S. Hospitals and Comparative Activities of Aminoglycosides against Carbapenem-Resistant Enterobacteriaceae and Isolates Carrying Carbapenemase Genes. *Antimicrob Agents Chemother*;62]
- ¹⁰² Walkty A, Karlowsky JA, Baxter MR, Adam HJ, Zhanel GG. In Vitro Activity of Plazomicin against Gram-Negative and Gram-Positive Bacterial Pathogens Isolated from Patients in Canadian Hospitals from 2013 to 2017 as Part of the CANWARD Surveillance Study. *Antimicrob Agents Chemother.* 2018 Dec 21;63(1)
- ¹⁰³ Thwaites M, Hall D, Stoneburner A, et al (2018) Activity of plazomicin in combination with other antibiotics against multidrug-resistant Enterobacteriaceae. *Diagn Microbiol Infect Dis*

¹⁰⁴ García-Salguero C, Rodríguez-Avial I, Picazo JJ, Culebras E (2015) Can Plazomicin Alone or in Combination Be a Therapeutic Option against Carbapenem-Resistant *Acinetobacter baumannii*? *Antimicrob Agents Chemother*;59:5959-66

¹⁰⁵ López Díaz MC, Ríos E, Rodríguez-Avial I, Simaluiza RJ, Picazo JJ, Culebras E. In-vitro activity of several antimicrobial agents against methicillin-resistant *Staphylococcus aureus* (MRSA) isolates expressing aminoglycoside-modifying enzymes: potency of plazomicin alone and in combination with other agents. *Int J Antimicrob Agents*. 2017 Aug;50(2):191-196

¹⁰⁶ Connolly LE, Riddle V, Cebrik D, Armstrong ES, Miller LG (2018) A Multicenter, Randomized, Double-Blind, Phase 2 Study of the Efficacy and Safety of Plazomicin Compared with Levofloxacin in the Treatment of Complicated Urinary Tract Infection and Acute Pyelonephritis. *Antimicrob Agents Chemother*;62

¹⁰⁷ Wagenlehner FME, Cloutier DJ, Komirenko AS, Cebrik DS, Krause KM, Keepers TR, Connolly LE, Miller LG, Friedland I, Dwyer JP; EPIC Study Group. Once-Daily Plazomicin for Complicated Urinary Tract Infections. *N Engl J Med*. 2019 Feb 21;380(8):729-740.

¹⁰⁸ McKinnell JA, Dwyer JP, Talbot GH, Connolly LE, Friedland I, Smith A, Jubb AM, Serio AW, Krause KM, Daikos GL; CARE Study Group. Plazomicin for Infections Caused by Carbapenem-Resistant Enterobacteriaceae. *N Engl J Med*. 2019 Feb 21;380(8):791-793

¹⁰⁹ Veve MP, Wagner JL (2018) Lefamulin: Review of a Promising Novel Pleuromutilin Antibiotic. *Pharmacotherapy*

¹¹⁰ Paukner S, Gelone SP, Arends SJR, Flamm RK, Sader HS. Antibacterial Activity of Lefamulin against Pathogens Most Commonly Causing Community-Acquired Bacterial Pneumonia: SENTRY Antimicrobial Surveillance Program (2015-2016). *Antimicrob Agents Chemother.* 2019 Mar 27;63(4)

¹¹¹ Zhang L, Wicha WW, Bhavnani SM, Rubino CM. Prediction of lefamulin epithelial lining fluid penetration after intravenous and oral administration using Phase 1 data and population pharmacokinetics methods. *J Antimicrob Chemother.* 2019 Apr 1;74(Supplement_3):iii27-iii34

¹¹² Wicha WW, Strickmann DB, Paukner S. Pharmacokinetics/pharmacodynamics of lefamulin in a neutropenic murine pneumonia model with *Staphylococcus aureus* and *Streptococcus pneumoniae*. *J Antimicrob Chemother.* 2019 Apr 1;74(Supplement_3):iii11-iii18

¹¹³ File TM Jr, Goldberg L, Das A, Sweeney C, Saviski J, Gelone SP, Seltzer E, Paukner S, Wicha WW, Talbot GH, Gasink LB. Efficacy and Safety of IV-to-Oral Lefamulin, a Pleuromutilin Antibiotic, for Treatment of Community-Acquired Bacterial Pneumonia: The Phase 3 LEAP 1 Trial. *Clin Infect Dis.* 2019 Feb 4