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Interaction of Femtosecond Pulsed Lasers with Fe²⁺ and Fe³⁺ Doped Calcium Phosphates for Bone Tissue Engineering

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Abstract: In this work, we aim to investigate the effect of Fe^{2+}/Fe^{3+} doping on the laser sintering of calcium phosphate minerals for the fabrication of bone scaffolds. The laser-matter mechanisms and the biological response are discussed. **OCTS code:** 320.2250, 140.7090.

Introduction: Current trends in tissue engineering (i.e. personalised medicine and near patient manufacturing) favour the use of additive manufacturing techniques for the fabrication of bone scaffolds. Selective laser sintering (SLS) is one of the well-established and widely used methodologies. Many studies have been reported by utilizing the SLS technique for manufacturing of biomaterials; e.g. metal alloys, polymers and bioglass [1]. Of particular interest though, is the investigation of calcium phosphate (CaP) biominerals due to their chemical and structural similarity with the natural bone mineral. Recently, it was reported that doping CaP with Fe²⁺ and Fe³⁺ ions improves energy absorption and allows the localised sintering of the materials without inducing any thermal damage to the surroundings [2, 3]. During irradiation with a femtosecond laser, the Fe-rich phases (e.g. Fe₂O₃ nanoparticles) absorb the laser radiation and act as thermal antennae, dissipating energy to the vicinal mineral phase, thereby triggering the sintering and densification of the surrounding calcium phosphate crystals. Based on this established mechanism of laser radiation absorption, the present work aims to identify an optimum doping concentration of Fe²⁺ and Fe³⁺ ions and investigate the effect of doping on i) the mechanism of laser matter interaction leading to changes in the materials structure; and effect of such changes on ii) the cytobiological adaptability of the laser-radiation transformed materials for ultimate in vivo use.

Materials and methods: In this investigation we synthesised brushite (CaHPO₄·2H₂O) doped with different concentrations of Fe²⁺ and Fe³⁺ ions (0%, 5%, 10%, 20% and 30% mol by substituting Ca²⁺ ions in the phosphate structure). The laser experiments were performed in ambient atmosphere using a Ti:sapphire amplified femtosecond laser system (Coherant-Libra), which emits radiation at a wavelength of 800 nm with 1kHz repetition rate and 100 fs pulse duration. The laser energy was controlled using a polarizer and a half-wave plate. The threshold fluence (F_{th}) of all Fe^{2+,3+}-doped brushite was calculated by using D² method [4]. The pre/post irradiation Fe^{2+,3+}-doped brushite biomaterials were characterised using X-ray diffraction (XRD), Fourier transform infrared spectroscopy (FT-IR) and scanning electron microscopy (SEM) for investigating changes in the crystal structure and phase constitution. The in vitro biocompatibility of laser-irradiated scaffolds was characterised by evaluating the adhesion and proliferation of osteoblast (G-292) cell lines.

Result and discussion: During the synthesis of the minerals some of the iron is incorporated into the crystal lattice while the rest, results in the formation of secondary iron rich phases (e.g. iron phosphate - FePO₄). For doping concentrations up to 10 mol%, brushite is the dominant phase and its original crystal structure remains intact after Fe^{2+}/Fe^{3+} incorporation. At 20% mol. an amorphous iron phosphate phase is formed alongside brushite crystals and for higher iron concentrations (>20%), the amorphous iron phosphate becomes the dominant phase.

Initially, laser irradiation experiments were carried out with different energy densities, in order to identify the ablation threshold (F_{th}) for each material. It was found that the presence of iron results in the shifting of F_{th} to higher energy densities as it increases exponentially with the doping concentration (0.20 J/cm² for un-doped brushite, 0.5 J/cm² for 5% Fe, 1.5 J/cm² for 10% Fe and 5.9 J/cm² for 20% Fe).

In **Figure 1** we compare the surface topology of undoped, 5% Fe-doped and 20% Fe-doped brushite after irradiation of pelletized calcium phosphate materials with constant laser pulse energy (200 μ J), scan speed of 100 mm/s. Using the comparable energy density conditions (1.7 J/cm²), deep microchannels were formed which is a clear indication of ablation. In the case of the 5% Fe^{2+,3+}-doped material, by using identical energy density conditions, partial melting was induced and shallow microchannels were observed. For 20% Fe^{2+,3+}-doping, there is no indication of ablation by forming microchannels, however such highly doped materials readily absorbed energy and cause extensive melting of the crystals. Also due to the localised temperature rise, thermal stresses were developed resulting into the cracking of the surface.



The effect of $Fe^{2+,3+}$ doping on the laser – matter interaction mechanisms is also evident after the phase analysis using XRD of the post irradiated samples. After laser irradiation of 0% Fe doped brushite, the material retained the same crystal structure. However, 5 and 10% doped brushite were transformed into a mixture of monetite (CaHPO₄) and brushite: while for 20% doped brushite, the monetite structure was only observed (**Fig. 2**). The transformation of brushite into monetite normally occurs at ~200 °C during slow heating in air in an oven. Based on the comparative studies of phase transformations in laser irradiated $Fe^{2+,3+}$ -doped calcium phosphate materials, it is evident that the doping concentration Fe^{2+}/Fe^{3+} in the calcium phosphate controls the laser penetration depth and ablation, and also the resulting phase and phase morphologies formed at the irradiated surface.



Fig 2. Comparison of X-ray diffraction patterns of brushite and 5% Fe doped brushite scaffolds after laser irradiation. Osteoblast cell lines (G-292) were used to test the cell viability and proliferation on sintered Fe²⁺brushite materials. For cytotoxicity experiments contact and extract (adenosine triphosphate ATP) method was followed. DNA using picogreen protocol was applied for the proliferation process. It was proven that doping with Fe^{2+/3+} does not induce any toxicological risk, even for high doping up to 20% concentrations. Moreover, cells were attached and proliferated for (1, 3, and 7 days) on the Fe^{2+/3+} containing minerals more extensively than in the case of undoped material (Fig. 3). The results of seeded osteoblast cell lines on the top Fe^{2+/3+} -doped pyro-phosphate microchannels will also be reported by comparing with the cell proliferation data, as exemplified in **Fig. 3**.



Fig 3: a) Proliferation of Fe β -pyrophosphate materials after 7 days b) contact test of 5% Fe β -pyrophosphate presents cells growth around sample.

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