

This is a repository copy of *Tissue inflammation signatures point towards resolution in adhesive capsulitis*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/142206/>

Version: Accepted Version

---

**Article:**

Dakin, SG, Rangan, Amar, Martinez, F et al. (6 more authors) (2019) Tissue inflammation signatures point towards resolution in adhesive capsulitis. *Rheumatology*. pp. 1109-1111. ISSN 1462-0332

<https://doi.org/10.1093/rheumatology/kez007>

---

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

**Tissue inflammation signatures point towards resolution in frozen shoulder**

Journal:	<i>Rheumatology</i>
Manuscript ID	Draft
Manuscript Type:	Letter to the Editor (Case Report)
Date Submitted by the Author:	n/a
Complete List of Authors:	<p>Dakin, Stephanie; University of Oxford, Nuffield Department of Orthopaedics, Rheumatology and Musculoskeletal Sciences</p> <p>Rangan, Amar; University of Oxford Nuffield Department of Orthopaedics Rheumatology and Musculoskeletal Sciences, NDORMS; University of York, Department of Health Sciences</p> <p>Martinez, Fernando; University of Oxford Nuffield Department of Orthopaedics Rheumatology and Musculoskeletal Sciences, NDORMS; University of Surrey, Faculty of Health &amp; Medical Sciences</p> <p>Brealey, Stephen; University of York, Department of Health Sciences</p> <p>Northgraves, Matthew; University of York, Department of Health Sciences</p> <p>Kottam, Lucksy; James Cook University Hospital, Department of Trauma and Orthopaedics</p> <p>Cooper, Cushla; University of Oxford, Nuffield Department of Orthopaedics Rheumatology and Musculoskeletal Science</p> <p>Buckley, Chris; University of Birmingham, Rheumatology Research Group ; University of Oxford Nuffield Department of Orthopaedics Rheumatology and Musculoskeletal Sciences, NDORMS</p> <p>Carr, Andrew J; University of Oxford Nuffield Department of Orthopaedics Rheumatology and Musculoskeletal Sciences, NDORMS</p>
Keywords Please select a minimum FIVE keywords from the list provided. These keywords will be used to select reviewers for this manuscript. The keywords in the main text of your paper do not need to match these words.:	Soft tissue rheumatism < RHEUMATIC DISEASES, Shoulder < REGIONAL RHEUMATISM, Ligaments and tendons < TISSUES, Fibroblast < BASIC & CLINICAL SCIENCES, Inflammation < BASIC & CLINICAL SCIENCES, Macrophages < BASIC & CLINICAL SCIENCES, Histopathology < DIAGNOSTIC METHODS

- 1
- 2
- 3
- 4
- 5
- 6
- 7
- 8
- 9
- 10
- 11
- 12
- 13
- 14
- 15
- 16
- 17
- 18
- 19
- 20
- 21
- 22
- 23
- 24
- 25
- 26
- 27
- 28
- 29
- 30
- 31
- 32
- 33
- 34
- 35
- 36
- 37
- 38
- 39
- 40
- 41
- 42
- 43
- 44
- 45
- 46
- 47
- 48
- 49
- 50
- 51
- 52
- 53
- 54
- 55
- 56
- 57
- 58
- 59
- 60

# Tissue inflammation signatures point towards resolution in frozen shoulder

SG Dakin<sup>1</sup>, A Rangan<sup>1,2</sup>, FO Martinez<sup>3</sup>, S Brealey<sup>2</sup>, M Northgraves<sup>2</sup>,  
L Kottam<sup>4</sup>, C Cooper<sup>1</sup>, CD Buckley<sup>1,5</sup>, AJ Carr<sup>1</sup>

<sup>1</sup> Nuffield Department of Orthopaedics, Rheumatology and Musculoskeletal Sciences University of Oxford, UK

<sup>2</sup> Department of Health Sciences, University of York, UK

<sup>3</sup> Faculty of Health and Medical Sciences, University of Surrey, UK

<sup>4</sup> Department of Trauma & Orthopaedics, The James Cook University Hospital, Middlesbrough, UK

<sup>5</sup> Institute of Inflammation and Ageing, University of Birmingham, UK

**Key message:** Proresolving receptors, macrophage and fibroblast activation point towards a resolving inflammatory milieu in frozen shoulder

Word count = 791/800

The authors declare no conflicts of interest exist

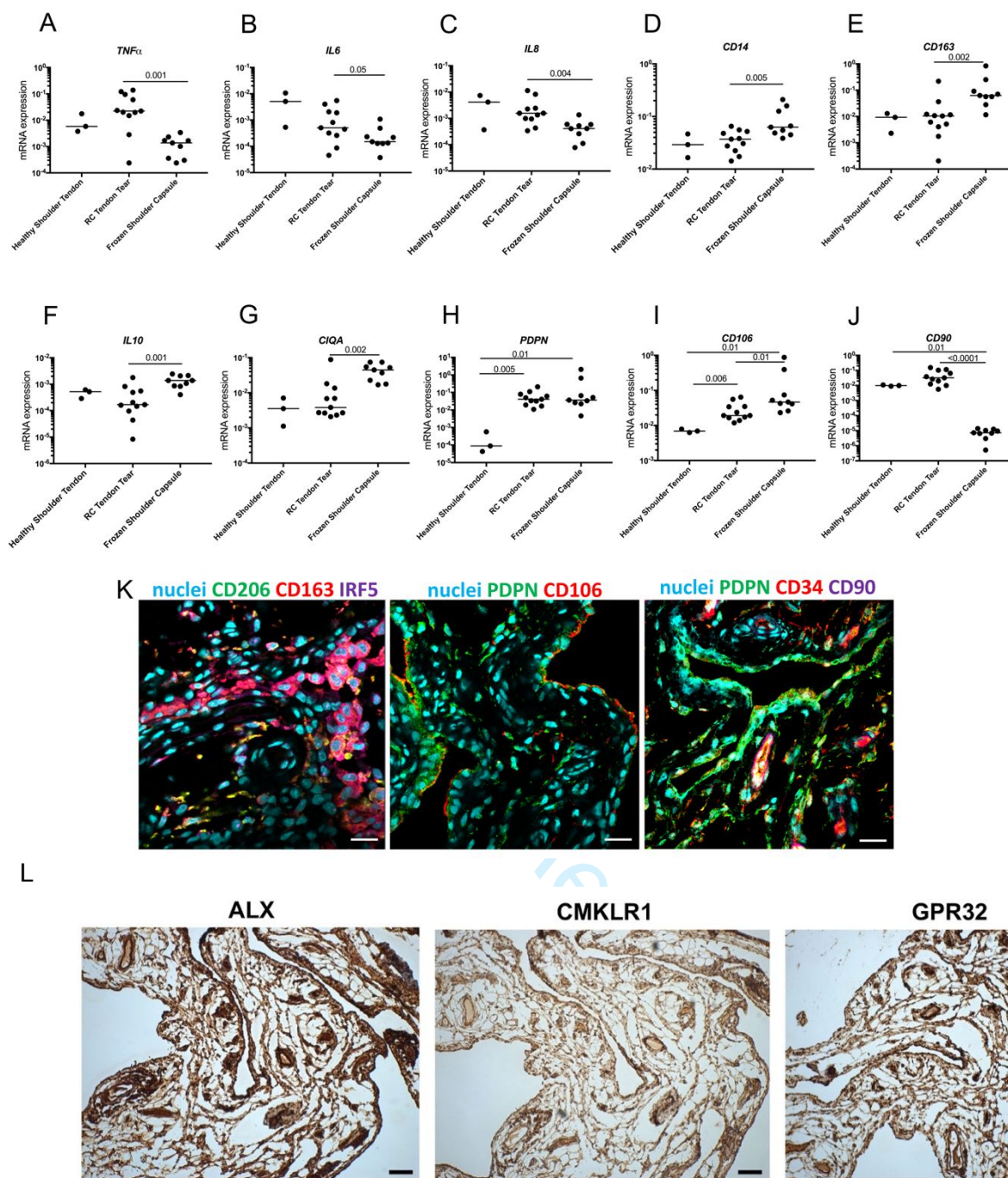
1 SIR, Frozen Shoulder is a remarkable example of a severe, yet self-limiting, inflammatory and fibrotic  
2 condition affecting the shoulder joint capsule. Patients experience pain and restricted shoulder joint motion  
3 for up to 3 years, severely limiting activities and disrupting quality of life [1]. The disease mechanisms are  
4 poorly understood and there are no truly effective therapies for symptomatic patients. The pathological  
5 features of frozen shoulder are reported to include leukocyte and myeloid infiltration, fibroblast accumulation  
6 and increased vascularity [2]. However, the distinct inflammatory pathways and the phenotypes of tissue  
7 resident stromal cells active in disease remain to be identified, and may inform why the condition ultimately  
8 spontaneously resolves. In this case study, we use contrasting manifestations of established shoulder  
9 disease in similarly aged patients to advance understanding of why inflammation is frequently self-limiting in  
10 frozen shoulder but persists in shoulder rotator cuff tendon tears. We therefore investigated inflammation  
11 signatures, characterising the phenotypes of macrophages and fibroblasts in tissue samples from patients  
12 with frozen shoulder, comparing them with tissues from patients with shoulder rotator cuff tendon tears and  
13 with normal rotator cuff tendons. We also investigated if frozen shoulder tissues expressed proresolving  
14 receptors mediating resolution of inflammation.

15 The frozen shoulder cohort consisted of 12 female and 4 male patients aged between 43-72 undergoing  
16 arthroscopic capsular release surgery as part of the NIHR-HTA programme funded UK FROST study [3].  
17 Frozen shoulder patient tissues were compared with those from similarly aged patients with torn  
18 supraspinatus tendons undergoing surgical debridement and repair (n=11). Healthy supraspinatus tendons  
19 were collected from patients undergoing shoulder stabilisation surgery (n=3). Tissues were collected under  
20 research ethics from the Oxford Musculoskeletal Biobank (09/H0606/11) and NRES Committee, Newcastle  
21 and North Tyneside (14/NE/1176). Full informed consent according to the Declaration of Helsinki was  
22 obtained from all patients. Collected tissues were processed for RNA isolation and histology. RT-qPCR and  
23 immunohistochemistry were performed using previously published protocols [4] to identify activation markers  
24 for macrophages and fibroblasts and proresolving receptors in collected tissues.

25 Inflammation signatures differed between tissues collected from frozen shoulder compared to tendon tear  
26 patients. Frozen shoulder tissues showed reduced expression of NF $\kappa$ B response genes including *TNF-alpha*,  
27 *IL6* and *IL8* compared to tissues from tendon tear patients (Figure 1A-C, p=0.001, 0.05 and 0.004  
28 respectively). Frozen shoulder tissues showed increased *CD14*, *CD163*, *IL10* and *C1QA* mRNA expression  
29 compared to torn tendons (Figure 1D-G, p=0.005, 0.002, 0.001 and 0.002 respectively). Fibroblast activation  
30 markers Podoplanin (*PDPN*) and *CD106* (VCAM-1) were highly expressed in frozen shoulder and torn  
31 tendons compared to healthy tendons (Figure 1 H-I). However, the fibroblast activation marker *CD90* was  
32 significantly reduced in frozen shoulder compared to healthy and diseased tendon tissues (Figure 1J p=0.01  
33 and p<0.0001 respectively). Immunostaining supported increased CD163, PDPN and CD106 and reduced  
34 CD90 expression in tissue sections from frozen shoulder patients (Figure 1K). Proresolving receptors  
35 mediating resolution of inflammation including ALX, CMKLR and GPR32 were highly expressed in frozen  
36 shoulder tissues (Figure 1L).

37 Investigating common shoulder diseases in similarly aged patients presents a unique opportunity to  
38 understand why inflammation ultimately resolves in frozen shoulder but persists in tendon tears. We identify  
39 tissues from patients with frozen shoulder differentially express markers of macrophage and fibroblast  
40 activation compared to those from patients with shoulder rotator cuff tendon tears. Frozen shoulder tissues  
41 showed reduced NF $\kappa$ B response genes and increased *IL10* compared to tendon tears, suggestive of a  
42 resolving inflammatory milieu. In support of this, increased CD163 suggests macrophages in frozen shoulder  
43 tissues exhibit a glucocorticoid receptor activation signature, associated with dampening inflammation and  
44 tissue repair [5]. Fibroblast activation markers PDPN and CD106 were highly expressed in both conditions,  
45 however *CD90* was significantly reduced in frozen shoulder compared to tendon tears. CD90 (Thy1) is  
46 expressed by pathogenic synovial fibroblasts from Rheumatoid Arthritis patients with a pro-inflammatory and  
47 invasive phenotype [6, 7]. The current study suggests the phenotypes of fibroblast subsets populating  
48 diseased shoulder tissues differ between self-limiting and persistent inflammation. CD90 therefore represents  
49 an important pathogenic marker and possible molecular checkpoint regulating persistent stromal mediated  
50 inflammation in common soft tissue disease of the joint. The identification of proresolving receptors ALX,  
51 CMKLR and GPR32 suggests proresolving pathways mediating resolution of inflammation are active in  
52 frozen shoulder. These proresolving proteins were highly expressed in frozen shoulder compared to our  
53 previous study on patients with established shoulder tendon tears [4]. Collectively, these findings provide  
54 novel insight into the disease mechanisms underpinning self-limiting inflammation in frozen shoulder,  
55 identifying proresolving receptors, macrophage and fibroblast activation signatures that point towards a  
56 resolving inflammatory milieu. Improved understanding of the biological mechanisms governing successful  
57 resolution of inflammation will inform the development of new therapeutic strategies targeting stromal  
58 mediated inflammation. These therapies are required to accelerate disease resolution in symptomatic frozen  
59 shoulder patients and in other common soft tissue diseases of the joint.  
60

Figure 1



**Figure 1. Activation of macrophages and fibroblasts and the presence of proresolving receptors point towards a resolving inflammatory milieu in tissues from patients with frozen shoulder.** Tissues were collected from healthy patients undergoing shoulder stabilisation surgery (n=3, healthy supraspinatus tendon), patients undergoing surgery to repair a supraspinatus rotator cuff (RC) tendon tear (n=11) or arthroscopic capsular release surgery for Frozen Shoulder (n=9). mRNA expression was determined for NF $\kappa$ B response genes (A-C), myeloid activation (D-E), anti-inflammatory cytokine (F), complement activation (G) and fibroblast activation markers (H-J). Statistically significant differences were calculated using pairwise Mann-Whitney U tests. Gene expression is normalized to  $\beta$ -actin; bars represent median values. (K) Representative immunofluorescence images of sections of Frozen Shoulder tissues stained for markers of macrophage (CD206, CD163, IRF5) and fibroblast activation (PDPN, CD106, CD90). Cyan represents POPO-1 nuclear counterstain. Scale bar, 20 $\mu$ m. (L) Representative images of immunostaining (brown) for proresolving receptors in sections of frozen shoulder tissues. Proresolving receptors ALX, CMKLR1 and GPR32 are highly expressed in frozen shoulder. Nuclear counterstain is haematoxylin. Scale bar, 100 $\mu$ m.

## Acknowledgements

SGD is funded by an Oxford-UCB Prize Fellowship in Biomedical Sciences. Research at Oxford University is supported by the NIHR Oxford Biomedical Research Centre. The views expressed are those of the authors and not necessarily the NHS or the Department of Health. This research was also funded by the NIHR HTA Programme (project number 13/26/01). The views expressed are those of the authors and not necessarily those of the NHS, the NIHR or the Department of Health and Social Care. A research grant from the British Elbow and Shoulder Society funded the transport costs of the tissue in the nested study. The UK FROST team acknowledges the support of the NIHR Clinical Research Network. We thank the trial participants who agreed to providing tissue samples and the participating hospitals who helped collect patient tissue samples.

## References

- 1 Rangan A, Hanchard N, McDaid C. What is the most effective treatment for frozen shoulder? *BMJ* 2016;354:i4162.
- 2 Hand GC, Athanasou NA, Matthews T, Carr AJ. The pathology of frozen shoulder. *J Bone Joint Surg Br* 2007;89(7):928-32.
- 3 Brealey S, Armstrong AL, Brooksbank A, et al. United Kingdom Frozen Shoulder Trial (UK FROST), multi-centre, randomised, 12 month, parallel group, superiority study to compare the clinical and cost-effectiveness of Early Structured Physiotherapy versus manipulation under anaesthesia versus arthroscopic capsular release for patients referred to secondary care with a primary frozen shoulder: study protocol for a randomised controlled trial. *Trials* 2017;18(1):614.
- 4 Dakin SG, Martinez FO, Yapp C, et al. Inflammation activation and resolution in human tendon disease. *Science translational medicine* 2015;7(311):311ra173.
- 5 Murray PJ, Allen JE, Biswas SK, et al. Macrophage Activation and Polarization: Nomenclature and Experimental Guidelines. *Immunity* 2014;41(1):14-20.
- 6 Mizoguchi F, Slowikowski K, Wei K, et al. Functionally distinct disease-associated fibroblast subsets in rheumatoid arthritis. *Nature communications* 2018;9(1):789.
- 7 Dakin SG, Coles M, Sherlock J, Powrie F, Carr AJ, Buckley CD. Pathogenic stromal cells as therapeutic targets in joint inflammation. *Nature reviews. Rheumatology In Press*.