

This is a repository copy of *Supplemental vitamin B-12 enhances the neural response to sensory stimulation in the barrel cortex of healthy rats but does not affect spontaneous neural activity.*

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/141432/>

Version: Accepted Version

---

**Article:**

Kang, Sungmin, Hayashi, Yurie, Bruyns-Haylett, Michael et al. (9 more authors) (2019) Supplemental vitamin B-12 enhances the neural response to sensory stimulation in the barrel cortex of healthy rats but does not affect spontaneous neural activity. *The Journal of Nutrition*. 730 -737. ISSN 0022-3166

<https://doi.org/10.1093/jn/nxz011>

---

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

# Supplemental vitamin B-12 enhances the neural response to sensory stimulation in the barrel cortex of healthy rats but does not affect spontaneous neural activity

Sungmin Kang<sup>1,7</sup>, Yurie Hayashi<sup>1</sup>, Michael Bruyns-Haylett<sup>2</sup>, Daniel H. Baker<sup>3</sup>, Marcia Boura<sup>4</sup>, Xuedan Wang<sup>4</sup>, Kimon-Andreas Karatzas<sup>4,7</sup>, Ines Serra<sup>5</sup>, Angela Bithell<sup>5</sup>, Claire Williams<sup>6</sup>, David T. Field<sup>7</sup>, Ying Zheng<sup>1,7\*</sup>

**\*Corresponding author:** Ying Zheng. Tel: +441183787635. Fax: +441189751994. Email: [ying.zheng@reading.ac.uk](mailto:ying.zheng@reading.ac.uk). Postal address: Biomedical Engineering, School of Biological Sciences, Whiteknights, University of Reading, Reading RG6 7AY, UK

**List of all authors' last names:** Kang, Hayashi, Bruyns-Haylett, Baker, Boura, Wang, Karatzas, Serra, Bithell, Williams, Field, Zheng.

**Word count:** 4120

**Number of figures:** 5

**Number of tables:** 0

**Supplementary data submitted:** 0

**Running title:** Vitamin B-12 supplementation enhances the neural signal

<sup>1</sup>Biomedical Engineering, School of Biological Sciences, Whiteknights, University of Reading, Reading RG6 7AY, UK. <sup>2</sup>Department of Bioengineering, Imperial College, South Kensington Campus, London SW7 2AZ, UK. <sup>3</sup>Department of Psychology and York Biomedical Research Institute, University of York, Heslington, York, YO10 5DD, UK. <sup>4</sup>Food and Nutritional Sciences, School of Chemistry, Food and Pharmacy, Whiteknights, University of Reading, Reading RG6 6AP, UK. <sup>5</sup>Pharmacy, School of Chemistry, Food and Pharmacy, Whiteknights, University of Reading, Reading RG6 7AP, UK. <sup>6</sup>Psychology, School of Psychology and Clinical Language Sciences, Whiteknights, University of Reading, Reading RG6 6AL, UK. <sup>7</sup>Centre for Integrative Neuroscience and Neurodynamics (CINN), University of Reading, Reading RG6 6AL, UK.

Abbreviations: B-12, Vitamin B-12; CON: control; CSD, current source density; EEG, electroencephalography; GABA,  $\gamma$ -aminobutyric acid; LFP, local field potential; MUA, multi-unit activity; PPR, paired-pulse ratio; PSD, power spectral density; SE, standard error.

This work was partly funded by the BBSRC (Grant BB/K010123) and the University of Reading.

Author disclosures: All authors declare no conflicts of interest.

# 1 **Abstract**

2 **Background:** Although vitamin B-12 (B-12) is known to contribute to the structural and  
3 functional development of the brain, it is unclear if B-12 supplementation has any beneficial  
4 effect in healthy populations in terms of enhanced neurological status of the brain or  
5 improved cognitive function.

6 **Objectives:** We investigated the effect of dietary supplementation of B-12 on the cortical  
7 neural activity of well-nourished young adult rats and tested the hypothesis that B-12  
8 supplementation in healthy rats may reduce sensory evoked neural activity due to enhanced  
9 inhibition.

10 **Methods:** Female Lister Hooded rats weighing between 190g to 265g (2 to 4 months old)  
11 were included in the study. The experimental group was fed with B-12 (Cyanocobalamin)  
12 enriched water at a concentration of 1mg/L, and the control (CON) group with tap water for 3  
13 weeks. Animals were then anaesthetised and cortical neural responses to whisker  
14 stimulation were recorded *in vivo* using a multi-channel micro-electrode, from which local  
15 field potentials (LFPs) were extracted.

16 **Results:** Somatosensory evoked LFP was enhanced 25% in the B-12 group ( $4.13 \pm 0.24$ mV)  
17 compared with the CON group ( $3.30 \pm 0.21$ mV) ( $P=0.02$ ). Spontaneous neural activity did not  
18 differ between groups; frequency spectra at each frequency bin of interest did not pass the  
19 cluster-forming threshold at the 5% significance level.

20 **Conclusions:** These findings do not provide evidence supporting the hypothesis of  
21 decreased neural activity due to B-12 supplementation. As the spontaneous neural activity  
22 was unaffected, the increase in somatosensory evoked LFP may be due to enhanced  
23 afferent signal reaching the barrel cortex from the whisker pad, indicating that B-12  
24 supplemented rats may have enhanced sensitivity to sensory stimulation compared to the  
25 CON group. We suggest that this enhancement might be the result of lowered sensory  
26 threshold, although the underlying mechanism has yet to be elucidated.

## 27 **Key words**

28 Local field potential, rat barrel cortex, vitamin B-12, dietary supplementation, sensory  
29 threshold, GABA.

## 30 **1. Introduction**

31 Vitamin B-12 (B-12) is an essential nutrient, vital for the maintenance of blood and nervous  
32 system function. It is a cofactor in the biosynthesis of methionine, a precursor for S-  
33 adenosyl-methionine in the brain. S-adenosyl-methionine is a major methyl donor for  
34 numerous central nervous system methylation reactions involving neurotransmitters, and  
35 plays a crucial role in myelin methylation [1-4].

36 Given the critical biochemical role that B-12 plays in human metabolic processes and in the  
37 synthesis of neurotransmitters, a recent study [5] investigated whether dietary  
38 supplementation with a yeast extract rich in B-12 could alter neural activity produced by  
39 visual patterns in the brains of healthy subjects. Using electroencephalography (EEG), the  
40 researchers observed a reduction in the steady state visual evoked potential for the  
41 intervention group compared to the placebo group, and it was speculated that B-12  
42 supplementation in healthy subjects might lead to increased concentration of the inhibitory  
43 neurotransmitter  $\gamma$ -aminobutyric acid (GABA), which in turn could modulate cortical  
44 excitation and inhibition. We will refer to this as the 'GABA hypothesis'.

45 The primary objective of the current study was to investigate whether dietary  
46 supplementation with B-12 has a significant effect on cortical neural activity. A set of *in vivo*  
47 electrophysiological experiments were conducted to examine the effect of B-12  
48 supplementation on cortical neural activity of healthy rats (2-4 months old) without B-12  
49 deficiency. If B-12 supplementation could lead to increased global GABA concentration in  
50 the brain, the likely effect would be a reduction in both the task-evoked and the spontaneous  
51 neural activity [6]. Thus somatosensory evoked local field potential (LFP) as well as

52 spontaneous neural activity in rats with and without B-12 intervention were collected and  
53 analysed. By examining the temporal dynamics of the evoked LFP, it was possible to assess  
54 how B-12 supplementation may have modulated components of synaptic excitation and  
55 inhibition in the LFP profile [7, 8]. To further investigate possible mechanisms underlying the  
56 observed changes in evoked LFP responses, we used paired-pulse stimulation to compare  
57 the sensory adaptation characteristics of the LFP between the diet groups, as sensory  
58 adaptation has been shown to be related to the intensity of stimulation [9].

## 59 **2. Materials and Methods**

60 All experiments were carried out in accordance with United Kingdom Home Office  
61 regulations (Animals (Scientific Procedures) Act, 1986) and approved by the Research  
62 Ethics Committee at the University of Reading, UK.

### 63 **2.1 Animals and diets**

64 A total of 29 female Lister Hooded rats weighing between 190g to 265g (2 to 4 months old)  
65 were included in the study. The strain, gender and age of the rats were chosen based on our  
66 previous work [8, 10]. This choice allowed us to re-use some of our previous data when  
67 making comparisons between CON and B-12 rats. Rats were housed in a temperature-  
68 controlled room with a 12-h light:dark cycle with *ad libitum* access to food and water, and  
69 were allowed to acclimatise to the animal room conditions and husbandry procedures for 3  
70 days prior to the start of the feeding programme which lasted for 3 weeks.

71 All rats were fed with standard commercial food (Rat and Mouse No.3 Breeding, RM3(E),  
72 801066, Special Diets Services, UK. The proximate composition: Moisture: 10.00%, Crude  
73 Oil: 4.25%. Crude Protein: 22.39%. Crude Fibre: 4.21%. Ash: 7.56%. Nitrogen Free Extract:  
74 51.20%), which has a B-12 (Cyanocobalamin) concentration of 26.78µg/kg, including  
75 17.75µg/kg supplemented B-12 from manufactured sources. Although this is less than the  
76 recommended dietary allowance (RDA) at 50µg/kg diet for rats [11], it is close to the B-12  
77 concentration in standard feeds used in other studies, and is well above the B-12

78 concentration in feeds deficient in B-12 [12, 13]. This was confirmed by the analysis of B-12  
79 concentration in serum samples (see Results).

80 The CON group (n=14) was fed with fresh water, while the B-12 group (n=15) was fed with  
81 B-12 (Cyanocobalamin, Sigma-Aldrich, UK) enriched water. B-12 was added to water  
82 incrementally for the first 3 days of the feeding programme at 25%, 50% and 75% of the final  
83 concentration, which was 40 times the RDA for rats [11]. Assuming the daily intake of food  
84 and water for adult rats to be approximately 5g/100g and 10mL/100g body weight  
85 respectively [14, 15], we estimated the RDA of B-12 to be 0.25µg/100g of rat's body weight.  
86 To provide 100% RDA of B-12 through water, the B-12 concentration would be 0.25µg/10mL  
87 water. Thus the final concentration for the intervention was set at  $40 \times 0.25\mu\text{g}/10\text{mL} =$   
88  $10\mu\text{g}/10\text{mL}$  water, or 1mg/L water. We chose 40 x RDA to be the final concentration in  
89 order to ensure the effectiveness of B-12 supplementation for this study. Dosages much  
90 higher than this have been used in both rodents and humans without evidence of adverse  
91 health effects [16, 17]. Note that fresh water (without B-12) was not supplied to the B-12  
92 group.

## 93 ***2.2 Surgery, neural recording and sample collection***

94 For detailed experimental procedures, the reader is directed to our previous publications [8,  
95 18, 19] for reference. They are briefly reviewed below.

96 Following 3 weeks of supplementation, animals were weighed, anaesthetised and operated  
97 on following our laboratory's standard surgical procedures. Stimulation was in terms of brief  
98 electric current pulses which were applied to the right whisker pad, and the neural activity of  
99 the contralateral barrel cortex was recorded via a 16-channel multi-laminar micro-electrode  
100 inserted perpendicular to the cortical surface of the barrel cortex. The neural signals thus  
101 recorded were typically low-pass filtered below ~500Hz [20] to produce the LFP which  
102 reflected changes in extra-cellular potentials with respect to a reference potential, and was  
103 primarily the weighted sum of post-synaptic activities of the local pyramidal neural

104 population. During whisker stimulation, LFP became more negative as positive currents  
105 flowed from extracellular space into intracellular space to depolarise principal neurons. The  
106 amplitude of the LFP deflection is approximately proportional to the strength of the  
107 stimulation [21], and the LFP deflection during the initial timeframe (1~2ms from the onset of  
108 the deflection) represents solely the excitatory post-synaptic activity of the local pyramidal  
109 neural population [7, 8].

110 A minimum of 100 trials were collected per animal with an inter-trial-interval of at least 5s. All  
111 neural data were sampled at 24.41 kHz. Stimulus intensity of 1.2mA was used for all  
112 animals. After initial analysis which revealed a larger evoked LFP amplitude in the B-12  
113 supplemented group compared with the CON group (see Results), an additional  
114 experimental condition with a stimulus intensity of 1.6mA was added to four rats in the CON  
115 group to investigate if the evoked LFP response under stronger stimulus intensity without B-  
116 12 supplementation could result in similar amplitude increases. The 1.6mA intensity is the  
117 strongest intensity previously tested without causing changes in either blood pressure or  
118 heart rate of rats under the adopted experimental paradigm [21-23]. The LFP data from  
119 these rats were then combined with an existing data set (n=4) collected from previous  
120 experiments conducted in our laboratory using identical experimental protocols and stimulus  
121 intensities [8]. Thus the total number of rats (in the CON group) subjected to both 1.2mA and  
122 1.6mA stimulus intensities was eight.

123 Finally for a subset of subjects (n=9/group), additional paired-pulse stimulation at 1.2mA was  
124 used to investigate if sensory adaptation characteristics could be altered by B-12  
125 supplementation. Stimulus parameters were kept the same for each pulse, while the inter-  
126 pulse-interval was set as 200ms, with the inter-trial-interval set as 10s.

127 At the end of each experiment, the rat was terminated by cervical dislocation. Blood  
128 collected via cardiac puncture was centrifuged at 3030 x g for 6 minutes at room  
129 temperature, and serum was then collected and stored at -80°C for further analysis. Brains  
130 were extracted, weighed and stored.

131 Note that only 26 (n=13/group) out of the 29 rats in the study provided usable neural data  
132 due to premature death or damage during surgery. Also serum was successfully collected  
133 from 21 (CON, n=10; B12, n=11) rats only. Serum samples were analysed for cobalamin (B-  
134 12) concentration using the Immulite/Immolute 1000 Systems VB Vitamin B12 (Siemens  
135 Healthcare Diagnostics Products Ltd) at the Pathology and Diagnostic Laboratories of the  
136 Royal Veterinary College based in Hertfordshire, UK. The device uses a solid-phase,  
137 competitive chemiluminescent enzyme immunoassay which has an intraassay imprecision  
138 (mean  $\pm$  standard deviation) of  $1308 \pm 77$  (pg/mL). The standard protocol for the quantitative  
139 measurement of B-12 in serum, as detailed in the manufacture's user guide, was followed.

### 140 ***2.3 Data pre-processing and parameter Estimation***

141 Neural recordings from the micro-electrode were first pre-processed using our laboratory's  
142 standard procedure [8]. Briefly, stimulus artifact was removed, data were zero-meant at  
143 baseline and low pass filtered. Inverse Current Source Density (spline iCSD [24]) analysis  
144 was performed to locate the layer IV sink [25] for each data set, and the CSD data were then  
145 used to align both the CSD and the LFP data according to their sink locations across  
146 animals, with the common sink placed 600  $\mu$ m below the pial surface. We used the re-  
147 aligned LFP time series at channel 7, where the sink was located, to represent the evoked  
148 neural activity to whisker pad stimulation, as this channel was located in the cortical layer  
149 which was targeted by thalamocortical afferents, with thalamus acting as a relay to deliver  
150 tactile response to whisker stimulation to the barrel cortex [26-28]. The evoked LFP was  
151 calculated by averaging over 100 trials for each animal. The first negative deflection  
152 observed in the evoked LFP was referred to as N1.

153 In order to compare evoked LFP across groups, the following parameters were extracted  
154 after pre-processing to smooth and align the data: (i) the onset of N1, which was defined as  
155 the time at which N1 exceeded -0.1mV, with the stimulus onset time assigned as zero; (ii)  
156 the initial slope of N1, which was defined as the slope from 2~25% of the N1 peak  
157 amplitude; (iii) the peak amplitude of N1, and (iv) the latency of the N1 peak.



158 For paired-pulse analysis, the amplitude of N1 of the second pulse was also extracted and  
159 the paired-pulse ratio (PPR) was calculated within each trial from

$$160 \quad \text{PPR} = \frac{\text{Amp(N1 of second pulse)}}{\text{Amp(N1 of first pulse)}}$$

## 161 **2.4 Frequency domain analysis**

162 To investigate possible mechanisms giving rise to differences in evoked LFP responses  
163 across diet groups, we checked the anaesthetic levels during the recording period to ensure  
164 that they were not significantly different between groups, as it has been shown that sensory  
165 evoked LFP is sensitive to the level of anaesthesia, and that the anaesthetic level is  
166 reflected in the resting state PSD within the frequency range 1~8Hz [29, 30]. Thus the  
167 resting state PSD below 8Hz was used to compare the anaesthetic depth between groups.  
168 To compute the resting state PSD for each trial, we used the resting state LFP data 0.9~4.9s  
169 post stimulation, down-sampled the data to 10KHz, and calculated PSD in Matlab™ via  
170 Welch's method (Hamming window, 50% overlap).

171 It is also well known that sensory evoked neural activity is closely influenced by spontaneous  
172 activity in the same cortical region [31-33]. One possible explanation for differences in  
173 evoked LFP responses between diet groups could be changes in spontaneous subthreshold  
174 activity and/or spontaneous spiking activity due to B-12 supplementation. Thus we extended  
175 the resting state PSD calculation to include frequencies up to 3000Hz to cover both the  
176 subthreshold (8~500Hz) neural activity and the multi-unit activity (MUA, 500~3000Hz) [20].

## 177 **2.5 Statistical analysis**

178 Throughout the analysis, the significance level  $\alpha$  was set at 0.05. Group analysis was  
179 performed to compare various measurements and parameters extracted from field potential  
180 recordings between the two diet groups using the two-tailed two-sample Student's t-test  
181 under the assumption that the sampling distribution of the mean was normally distributed.  
182 Parameters were presented as mean  $\pm$  standard error (SE). To compare the N1 amplitude in

183 response to two stimulus intensities applied to the same rat, the two-tailed paired-sample  
184 Student's t-test was used.

185 To compare the ratios of brain weight to body weight across the groups, the non-parametric  
186 Wilcoxon rank-sum test was used to test for equal medians, as the ratio of two normally  
187 distributed variables is no longer normally distributed.

188 For comparison of PSDs over the frequency range 1~3000Hz, a non-parametric cluster  
189 correction procedure [34, 35] was used to determine significant clusters across the  
190 frequency range while controlling for multiple comparisons. This involved conducting an  
191 independent two-sample t-test at each frequency bin (width=1 Hz) to compare responses in  
192 the CON and the B-12 groups. Tests that were significant at  $P<0.05$  were aggregated into  
193 clusters across adjacent frequency bins. The summed t-statistic for each cluster was  
194 compared to a null distribution generated by resampling the data from the largest cluster  
195 10,000 times with randomly assigned group labels, and recalculating the summed t-statistic.  
196 Clusters which fell outside of the empirical 95% confidence intervals of the null distribution  
197 were considered significant.

198 Finally, we assumed no significant bias in the weight of the rats between the two groups at  
199 the start of the feeding programme. This was reasonable based on the fact that all rats,  
200 weighing between 175 ~ 224g, were purchased from the same source (Charles River, UK)  
201 on 8 occasions (4 pseudorandom occasions per diet group) across a 20-month period. It  
202 should be noted that rats were only weighed once immediately prior to surgery.

## 203 **3. Results**

### 204 ***3.1 B-12 serum concentration, body and brain weights***

205 The serum cobalamin concentration was 98% greater in the B-12 group compared with the  
206 CON group ( $P<0.01$ ) (**Figure 1A**). The concentration in the CON group was within the  
207 normal range for rats [13, 36, 37], thus confirming that they were not deficient in B-12. There

208 was no significant difference between the final body weight (**Figure 1B**) and brain weight  
209 (**Figure 1C**), and the brain/body weight ratio (**Figure 1D**) between the two diet groups. Thus  
210 our results suggest that, assuming no significant weight difference across diet groups at the  
211 start of the feeding programme, B-12 supplementation did not significantly change body  
212 weight, brain weight, or the ratio between them.

### 213 ***3.2 Amplitude of evoked LFP was increased by B-12 supplementation***

214 Along the cortical depth, the B-12 group had a larger LFP response (**Figure 2A**) and a  
215 correspondingly stronger sink/source pair (**Figure 2B**) compared to the CON group. These  
216 were reflected in the brighter blue colour associated with the B-12 group images. Time  
217 series of the evoked LFP responses in the layer IV sink are displayed in **Figure 2C**. The  
218 amplitude of N1 for the B-12 group was 25.2% larger than that of the CON group ( $P=0.02$ )  
219 (**Figure 2D**), while the latency of the N1 peak for the B-12 group was significantly shorter  
220 than the CON group ( $P=0.03$ ) (**Figure 2E**). In addition, the initial slope of N1 for the B-12  
221 group was significantly steeper than the CON group ( $P<0.01$ ) (**Figure 2F**), however the  
222 onset of N1 was not significantly different ( $P=0.39$ ) (**Figure 2G**). Together these  
223 characteristics suggested that the dynamics of the evoked LFP response for the B-12 group  
224 were faster, reflected in the steeper initial slope, and stronger, in terms of the N1 amplitude,  
225 compared to the CON group. However the onset of N1 was not significantly different  
226 between the diet groups, with the important implication that B-12 supplementation for 3  
227 weeks did not significantly change the transmission speed of the afferent neural signal  
228 arriving at the barrel cortex from the whisker pad.

### 229 ***3.3 Sensory adaptation was weakened by B-12 supplementation***

230 Sensory adaptation characteristics of neural responses were investigated using the paired-  
231 pulse stimulus paradigm, results of which are shown in **Figures 3A** and **3B** for CON and B-  
232 12 groups respectively. The PPR for the B-12 group was 21.9% higher than the CON group  
233 ( $P=0.04$ ) (**Figure 3C**), indicating that the second pulse was significantly less adapted for the

234 B-12 group than the CON group. Therefore, despite a higher amplitude of the first evoked  
235 LFP pulse in the B-12 group compared to that of the CON group, the recovery of the second  
236 pulse (200ms apart) was faster in the B-12 group.

### 237 **3.4 Resting state power spectral density (PSD) analysis**

238 There was a clear overlap of PSDs between individuals in the CON group and those in the  
239 B-12 group, indicating no significant difference in either the depth of anaesthesia (**Figure**  
240 **4A**), or the subthreshold and MUA neural activity (**Figure 4B**) between the two diet groups.  
241 The nonparametric cluster correction analysis showed that, across all frequency bins, the  
242 maximum absolute t-statistic was 1.84, less than the critical t-value of 2.06 (n=13/group,  
243 degree of freedom=24) for significance at the 5% level, further confirming that there was no  
244 significant difference in PSDs between the two diet groups in the frequency range  
245 1~3000Hz.

### 246 **3.5 Effect of stimulus intensity**

247 For rats without B-12 supplementation, the evoked LFP amplitude to the 1.6mA stimulation  
248 was 13.9% higher than that to the 1.2mA stimulation ( $P<0.01$ ) (**Figure 5A**). For comparison,  
249 we re-plotted the LFP responses of CON and B-12 groups to the 1.2mA stimulation (**Figure**  
250 **5B**). As stated previously in Section 3.2, the N1 amplitude for the B-12 group was 25.2%  
251 higher than that of the CON group.

## 252 **4. Discussion**

253 To the best of our knowledge, this is the first study to show that healthy rats supplemented  
254 with B-12 demonstrate an increase in sensory evoked synaptic activity in the somatosensory  
255 cortex. We discuss here possible mechanisms underlying the observed phenomena and  
256 their implications for future research.

### 257 **4.1 B-12 supplementation and the myelin sheath**

258 It is well-known that B-12 plays a crucial role in myelin methylation. Recent research further  
259 suggests that the myelin sheath is more than an inert insulating membrane structure [38-40].  
260 A study on rat somatic sensorimotor system has shown that the structure of myelin sheath in  
261 the spinal cord underwent changes throughout the aging process [41]. Furthermore  
262 myelination properties have been shown to be regulated by neuronal activity and the  
263 environment [42, 43]. It is therefore plausible that in young adult rats, as used in our study,  
264 myelination properties such as myelin sheath length and/or thickness could be altered within  
265 a 3-week period, and that the increased neural response described here could be the result  
266 of strengthened myelination of neurons in B-12 rats. However, the onset of the N1 deflection  
267 in our data across the diet groups did not differ significantly (Figure 2G), suggesting that the  
268 neuron conduction velocity was not changed by the supplementation. However, we also  
269 recognise that there is a minimum difference of onset that could be detected in our  
270 measurement at a 5% level of significance. This can be estimated using the two-sample t-  
271 statistic:

$$272 \quad \frac{\bar{x}_1 - \bar{x}_2}{\sqrt{s_1^2/n_1 + s_2^2/n_2}} = t_{n_1+n_2-2, 0.05}$$

273 With the standard errors of onset shown in Figure 2G (CON: SE=0.08ms; B-12: SE=0.06ms;  
274 n=13/group), we found this difference to be 0.21ms. In other words, if the onset difference  
275 between the two groups was 0.21ms or less, we would not be able to detect it at a 5% level  
276 of significance.

#### 277 **4.2 B-12 supplementation and the ‘GABA hypothesis’**

278 We are not aware of any study providing evidence linking dietary supplementation of B-12 to  
279 changes in GABA in the brain. However using intracerebroventricular infusion, Ikeda et al  
280 [44] investigated the effect of B-12 on circadian pace-making in rodents and found that B-12  
281 infusion significantly increased the content of GABA in the suprachiasmatic nucleus of the  
282 hypothalamus, while the content of the excitatory neurotransmitter glutamate in the same  
283 region was significantly decreased. The authors speculated that B-12 may modulate the

284 metabolism of GABA and glutamate by facilitating glutamic acid decarboxylase activity. In  
285 addition, a recent human study investigated the effect of dietary intervention of a yeast  
286 extract substance on steady state visual evoked potentials (VEPs), and found reduced  
287 neural responses in the diet group compared to the placebo group [5]. As the yeast  
288 substance was richer in B-12 in comparison to the placebo substance, the researchers  
289 suggested that the observed reduction could be the result of increased GABA concentration  
290 in the brain due to dietary supplementation of B-12.

291 Direct comparison of our study to the above human study is not possible, not least because  
292 the stimulus paradigms used in the two studies were very different. However if dietary  
293 supplementation with B-12 increased the global GABA concentration in the brain, the likely  
294 effect on the spontaneous as well as task-evoked neural activity would be a reduction in both  
295 [6]. PSD analysis of our data during resting state showed no significant difference between  
296 the two groups over the frequency range 1~3000Hz (Figure 4). In addition, the evoked LFP  
297 for the B-12 group showed significantly increased amplitude and faster temporal dynamics.  
298 Both of these observations could be taken as evidence against the GABA hypothesis.

299 GABA concentration in rat brain can be measured using techniques such as Gas  
300 Chromatography-Mass Spectrometry, immunohistochemistry and magnetic resonance  
301 spectroscopy. We plan to conduct some of these tests for our future studies.

#### 302 ***4.3 B-12 enhanced LFP may implicate enhanced sensitivity to sensory stimulation***

303 Figure 2F showed that the initial slope of N1 for the B-12 group is steeper than that of the  
304 CON group. Based on our previous study [8], this suggests faster excitatory post-synaptic  
305 activity for the B-12 group. On the other hand we didn't observe significant difference in the  
306 resting state neural activity between the groups. The scenario is analogous to the barrel  
307 cortex responding to whisker stimulation with two levels of intensity, the stimulus with  
308 stronger intensity will evoke a higher LFP response amplitude than that evoked by the lower  
309 intensity stimulus, while the resting state LFP will be unaffected by stimulus strength [8, 21,

23]. In other words, the enhanced LFP response for the B-12 group could be due to enhanced thalamo-cortical afferent signal, suggesting that B-12 supplementation in well-nourished rats may have enhanced the sensitivity of neurons to sensory stimulation in the lemniscal pathway linking peripheral nerves in the whisker pad to neurons in the thalamus. This is further supported by our results on sensory adaptation. The mechanism underlying sensory adaptation and stimulus strength was studied in detail by Ganmor et al [9] who demonstrated that stronger whisker stimulation produced weaker sensory adaptation in the somatosensory cortex of rodent. They pinpointed the source of this weaker adaptation to neurons in the brainstem trigeminal complex and argued that such coding strategy may be used to discriminate stimulus intensities during adaptation in order to counterbalance the effect of short-term synaptic depression in the thalamus and subsequently in the cortex. Based on their work, our observed weaker sensory adaptation in the B-12 group could be due to neurons in the brainstem responding more strongly to the same stimulus compared to the CON group, with faster recovery and subsequently less adaptation to the second stimulus. Further experiments will be needed to confirm this by using a wider range of stimulus intensities with neural recordings from the thalamus and the brainstem of both diet groups.

#### 327 ***4.4 B-12 supplementation and sensory threshold***

328 Changes in sensory evoked potentials have been linked to changes in sensory threshold. 329 Lund et al [45] found that, post-surgery, sensory threshold to cutaneous electrical stimulation 330 was increased, while the peak-to-peak amplitudes of somatosensory evoked potentials were 331 decreased significantly. A more recent study on pain-threshold and aggressiveness found 332 that individuals who more often behave aggressively had a higher pain threshold, and 333 aggressiveness was negatively correlated to the amplitude of pain-related evoked potentials 334 [46]. In auditory research, evoked potentials have been found to correlate with auditory 335 signal detection, specifically the amplitude of auditory evoked potentials associated with 336 correctly detected signals were found to be much higher than those corresponding to falsely

337 reported signals, undetected signals or correctly reported non-signals [47]. Furthermore,  
338 sensory thresholds can be lowered by training. Using the human visual system, Skrandies et  
339 al [48] demonstrated that sensory threshold decreased during repeated presentation of  
340 visual hyper-acuity stimuli. This was accompanied by significantly larger amplitude of VEPs.  
341 Interestingly, they also observed significantly shorter peak latency in VEPs post training,  
342 which agrees with the shorter N1 peak latency for the B-12 group (Figure 2E).

343 Based on this body of literature, we suggest that B-12 supplementation may have the effect  
344 of lowering the sensory threshold, thus enhancing the sensitivity of neurons to sensory  
345 stimulation.

#### 346 ***4.5 Future work***

347 The conclusion of the study is limited by several shortcomings. One is that the concentration  
348 of GABA in the brain was not measured, thus we were unable to confirm if the sensory  
349 evoked LFP difference between the diet groups was related to the different levels of GABA  
350 concentration. The other is that we did not measure B-12 concentration in the brain,  
351 although its concentrations in serum samples were obtained. Furthermore, cognitive  
352 correlates of B-12 supplementation were not assessed. We plan to incorporate these  
353 measurements in future studies to further elucidate the role that B-12 may play in shaping  
354 the neurological and cognitive functions of the brain.

355 Dietary supplementation of B12 is inexpensive and non-toxic. If it can be demonstrated to  
356 slow down age-related cognitive decline through increased responsiveness to sensory  
357 stimulation, it will have significant impact on the well-being of older people, and generate  
358 considerable economic as well as public health benefits.

#### 359 **Acknowledgement**

360 We thank Prof. Simon Andrews, Dr. Stephen Elmore, Dr. Peter Harris, Ms. Amanpreet Kaur,  
361 Mr. Andrew Cripps and the BRU at the University of Reading, and Ms Sue Rodway at the



362 Royal Veterinary College based in Hertfordshire, UK for their assistance during the project.  
363 The authors' contributions to the manuscripts are as follows: D.T.F., M.B-H., Y.Z. and D.H.B.  
364 conceived the project. Y.Z., M.B-H. and C.W. designed the research. S.K., Y.H., M.B-H. and  
365 Y.Z. collected data. M.B., X.W., K.K., I.S. and A.B. provided essential materials. Y.Z., S.K.,  
366 M.B-H., D.H.B. analysed data. Y.Z. wrote the manuscript. D.T.F., C.W., A.B., M.B-H., D.H.B.  
367 and I.S. contributed to the editing of the manuscript. Y.Z. had primary responsibility for the  
368 final content. All authors have read and approved the final manuscript.

369

## References

1. Young SN, Shalchi M: **The effect of methionine and S-adenosylmethionine on S-adenosylmethionine levels in the rat brain.** *J Psychiatry Neurosci* 2005, **30**:44-48.
2. Gröber U, Kisters K, Schmidt J: **Neuroenhancement with Vitamin B12—Underestimated Neurological Significance.** *Nutrients* 2013, **5**:5031-5045.
3. Kennedy DO: **B Vitamins and the Brain: Mechanisms, Dose and Efficacy—A Review.** *Nutrients* 2016, **8**:68.
4. Porter K, Hoey L, Hughes FC, Ward M, McNulty H: **Causes, Consequences and Public Health Implications of Low B-Vitamin Status in Ageing.** *Nutrients* 2016, **8**.
5. Smith A, K., Wade A, R. , Penkman K, E. H. , Baker D, H. : **Dietary modulation of cortical excitation and inhibition.** *J Psychopharmacol (Oxf)* 2017, **31**:632-637.
6. Duncan NW, Wiebking C, Northoff G: **Associations of regional GABA and glutamate with intrinsic and extrinsic neural activity in humans—A review of multimodal imaging studies.** *Neurosci Biobehav Rev* 2014, **47**:36-52.
7. Zheng Y, Luo JJ, Harris S, Kennerley A, Berwick J, Billings SA, Mayhew J: **Balanced excitation and inhibition: Model based analysis of local field potentials.** *Neuroimage* 2012, **63**:81-94.

8. Bruyns-Haylett M, Luo J, Kennerley AJ, Harris S, Boorman L, Milne E, Vautrelle N, Hayashi Y, Whalley BJ, Jones M, et al: **The neurogenesis of P1 and N1: A concurrent EEG/LFP study.** *Neuroimage* 2017, **146**:575-588.
9. Ganmor E, Katz Y, Lampl I: **Intensity-Dependent Adaptation of Cortical and Thalamic Neurons Is Controlled by Brainstem Circuits of the Sensory Pathway.** *Neuron* 2010, **66**:273-286.
10. Mayhew J, Zheng Y, Hou YQ, Vuksanovic B, Berwick J, Askew S, Coffey P: **Spectroscopic analysis of changes in remitted illumination: the response to increased neural activity in brain.** *Neuroimage* 1999, **10**:304-326.
11. Council NR: *Nutrient Requirements of Laboratory Animals, : Fourth Revised Edition, 1995.* Washington, DC: The National Academies Press; 1995.
12. Jaffé WG: **Requirements of Rats for Vitamin B12 During Growth, Reproduction and Lactation.** *The Journal of Nutrition* 1956, **59**:135-146.
13. Birn H, Nexø E, Christensen EI, Nielsen R: **Diversity in rat tissue accumulation of vitamin B12 supports a distinct role for the kidney in vitamin B12 homeostasis.** *Nephrology Dialysis Transplantation* 2003, **18**:1095-1100.
14. Hubert M-F, Laroque P, Gillet J-P, Keenan KP: **The Effects of Diet, ad Libitum Feeding, and Moderate and Severe Dietary Restriction on Body Weight, Survival, Clinical Pathology Parameters, and Cause of Death in Control Sprague-Dawley Rats.** *Toxicol Sci* 2000, **58**:195-207.
15. Beale KEL, Murphy KG, Harrison EK, Kerton AJ, Ghatei MA, Bloom SR, Smith KL: **Accurate Measurement of Body Weight and Food Intake in Environmentally Enriched Male Wistar Rats.** *Obesity* 2011, **19**:1715-1721.
16. Richardson LR, Brock R: **Studies of Reproduction in Rats Using Large Doses of Vitamin B12 and Highly Purified Soybean Proteins.** *The Journal of Nutrition* 1956, **58**:135-145.

17. Eussen SM, de Groot LM, Clarke R, et al.: **Oral cyanocobalamin supplementation in older people with vitamin b12 deficiency: A dose-finding trial.** *Arch Intern Med* 2005, **165**:1167-1172.
18. Boorman L, Harris S, Bruyns-Haylett M, Kennerley A, Zheng Y, Martin C, Jones M, Redgrave P, Berwick J: **Long-Latency Reductions in Gamma Power Predict Hemodynamic Changes That Underlie the Negative BOLD Signal.** *The Journal of Neuroscience* 2015, **35**:4641-4656.
19. Kang S, Bruyns-Haylett M, Hayashi Y, Zheng Y: **Concurrent Recording of Co-localized Electroencephalography and Local Field Potential in Rodent.** *Journal of Visualized Experiments : JoVE* 2017:56447.
20. Einevoll GT, Kayser C, Logothetis NK, Panzeri S: **Modelling and analysis of local field potentials for studying the function of cortical circuits.** *Nat Rev Neurosci* 2013, **14**:770-785.
21. Jones M, Hewson-Stoate N, Martindale J, Redgrave P, Mayhew J: **Nonlinear coupling of neural activity and CBF in rodent barrel cortex.** *Neuroimage* 2004, **22**:956-965.
22. Mayhew J, Johnston D, Berwick J, Jones M, Coffey P, Zheng Y: **Spectroscopic analysis of neural activity in brain: Increased oxygen consumption following activation of barrel cortex.** *Neuroimage* 2000, **12**:664-675.
23. Hewson-Stoate N, Jones M, Martindale J, Berwick J, Mayhew J: **Further nonlinearities in neurovascular coupling in rodent barrel cortex.** *Neuroimage* 2005, **24**:565-574.
24. Pettersen KH, Devor A, Ulbert I, Dale AM, Einevoll GT: **Current-source density estimation based on inversion of electrostatic forward solution: Effects of finite extent of neuronal activity and conductivity discontinuities.** *J Neurosci Methods* 2006, **154**:116-133.

25. Mitzdorf U: **Current source-density method and application in cat cerebral cortex: investigation of evoked potentials and EEG phenomena.** *Physiol Rev* 1985, **65**:37-100.
26. Feldmeyer D: **Excitatory neuronal connectivity in the barrel cortex.** *Frontiers in neuroanatomy* 2012, **6**:24-24.
27. Constantinople CM, Bruno RM: **Deep cortical layers are activated directly by thalamus.** *Science (New York, NY)* 2013, **340**:1591-1594.
28. Mease RA, Krieger P, Groh A: **Cortical control of adaptation and sensory relay mode in the thalamus.** *Proceedings of the National Academy of Sciences* 2014, **111**:6798-6803.
29. Friedberg MH, Lee SM, Ebner FF: **Modulation of Receptive Field Properties of Thalamic Somatosensory Neurons by the Depth of Anesthesia.** *J Neurophysiol* 1999, **81**:2243-2252.
30. Devonshire IM, Grandy TH, Dommett EJ, Greenfield SA: **Effects of urethane anaesthesia on sensory processing in the rat barrel cortex revealed by combined optical imaging and electrophysiology.** *Eur J Neurosci* 2010, **32**:786-797.
31. Petersen CCH, Hahn TTG, Mehta M, Grinvald A, Sakmann B: **Interaction of sensory responses with spontaneous depolarization in layer 2/3 barrel cortex.** *Proc Natl Acad Sci U S A* 2003, **100**:13638-13643.
32. Schölvinck ML, Friston KJ, Rees G: **The influence of spontaneous activity on stimulus processing in primary visual cortex.** *Neuroimage* 2012, **59**:2700-2708.
33. He BJ: **Spontaneous and Task-Evoked Brain Activity Negatively Interact.** *The Journal of Neuroscience* 2013, **33**:4672-4682.
34. Maris E, Oostenveld R: **Nonparametric statistical testing of EEG- and MEG-data.** *J Neurosci Methods* 2007, **164**:177-190.
35. Baker DH: **Decoding eye-of-origin outside of awareness.** *Neuroimage* 2017, **147**:89-96.

36. Sivakumar B, Nath N, Nath MC: **Effect of Various High Protein Diets on Vitamin B12 Status in Rats.** *The Journal Of Vitaminology* 1969, **15**:151-154.
37. Nix WA, Zirwes R, Bangert V, Kaiser RP, Schilling M, Hostalek U, Obeid R: **Vitamin B status in patients with type 2 diabetes mellitus with and without incipient nephropathy.** *Diabetes Res Clin Pract* 2015, **107**:157-165.
38. Simons M, Nave K-A: **Oligodendrocytes: Myelination and Axonal Support.** *Cold Spring Harbor perspectives in biology*, **8**:a020479-a020479.
39. Young KM, Psachoulia K, Tripathi RB, Dunn S-J, Cossell L, Attwell D, Tohyama K, Richardson WD: **Oligodendrocyte dynamics in the healthy adult CNS: evidence for myelin remodeling.** *Neuron* 2013, **77**:873-885.
40. Fields RD: **White matter in learning, cognition and psychiatric disorders.** *Trends Neurosci* 2008, **31**:361-370.
41. Xie F, Liang P, Fu H, Zhang J, & , Chen J: **Effects of normal aging on myelin sheath ultrastructures in the somatic sensorimotor system of rats.** *Molecular Medicine Reports* 2014, **10**:459-466.
42. Michalski J-P, Kothary R: **Oligodendrocytes in a Nutshell.** *Frontiers in cellular neuroscience* 2015, **9**:340-340.
43. Gibson EM, Purger D, Mount CW, Goldstein AK, Lin GL, Wood LS, Inema I, Miller SE, Bieri G, Zuchero JB, et al: **Neuronal activity promotes oligodendrogenesis and adaptive myelination in the mammalian brain.** *Science (New York, NY)* 2014, **344**:1252304-1252304.
44. Ikeda M, Azuma S, Inoue S: **Vitamin B12 enhances GABA content but reduces glutamate content in the rat suprachiasmatic nucleus.** *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 1997, **273**:R359-R363.
45. Lund C, Hansen OB, Kehlet H: **Effect of surgery on sensory threshold and somatosensory evoked potentials after skin stimulation.** *Br J Anaesth* 1990, **65**:173-176.

46. Ring C, Kavussanu M, Willoughby A: **Pain thresholds, pain-induced frontal alpha activity and pain-related evoked potentials are associated with antisocial behavior and aggressiveness in athletes.** *Psychology of Sport and Exercise* 2016, **22**:303-311.
47. Hillyard SA, Squires KC, Bauer JW, Lindsay PH: **Evoked Potential Correlates of Auditory Signal Detection.** *Science* 1971, **172**:1357-1360.
48. Skrandies W, Lang G, Jedynek A: **Sensory thresholds and neurophysiological correlates of human perceptual learning.** *Spat Vis* 1996, **9**:475-489.

## Figure legends

**FIGURE 1.** Serum cobalamin concentration (A), final body (B) and brain weights (C), and the brain/body weight ratio (D) in adult female rats that did not or did consume B-12 for 3 weeks. Values are means  $\pm$  SEs. For serum samples, CON, n=10; B-12, n=11. For body and brain weights, CON, n=14; B-12, n=15. Asterisks indicate different from CON:  $**P < 0.01$ . B-12, vitamin B12; CON, control.

**FIGURE 2.** Neural responses and associated parameters in adult female rats that did not or did consume B-12 for 3 weeks. (A) Mean LFPs displayed as images for the two diet groups. Cortical depth is along the vertical axis, with the top of the image being 200 $\mu$ m below pia mater. Time is along the horizontal axis, with the black triangle indicating stimulus onset. (B) Similar to (A) but CSD of the two diet groups. (C) Mean evoked LFP time series of the two diet groups. Shadows indicate SE. Stimulus onset is at t=0. (D) The amplitude, (E) the latency, (F) the initial slope, and (G) the onset of N1. Values are means  $\pm$  SEs, n=13/group. Asterisks indicate different from CON:  $*P < 0.05$ ;  $**P < 0.01$ . B-12, vitamin B12; CON, control; CSD, current source density; LFP, local field potential.

**FIGURE 3.** Mean LFP responses to paired-pulse stimulation in adult female rats that did not (A) and did (B) consume B-12 for 3 weeks. Shadows indicate SE. Stimulus onsets are at t=0 and t=200ms respectively. (C) PPR of the two diet groups. Values are means  $\pm$  SEs, n=9/group. Asterisks indicate different from CON:  $*P < 0.05$ . B-12, vitamin B12; CON, control; LFP, local field potential; PPR, paired-pulse ratio.

**FIGURE 4.** Mean resting state PSD in the frequency range 1~8Hz (A), and 8~3000Hz (B) of adult female rats that did not or did consume B-12 for 3 weeks. Individual subject's PSD are also displayed, n=13/group. B-12, vitamin B12; CON, control; PSD, power spectral density.

**FIGURE 5.** (A) Mean LFP responses to whisker stimulation at intensities 1.2mA (n=13) and 1.6mA (n=8) respectively for adult female rats that did not consume B-12. (B) Mean LFP

responses of the two diet groups at the same stimulus intensity of 1.2mA, n=13/group. Error bars indicate SEs. B-12, vitamin B12; CON, control; LFP, local field potential.

Figure 1

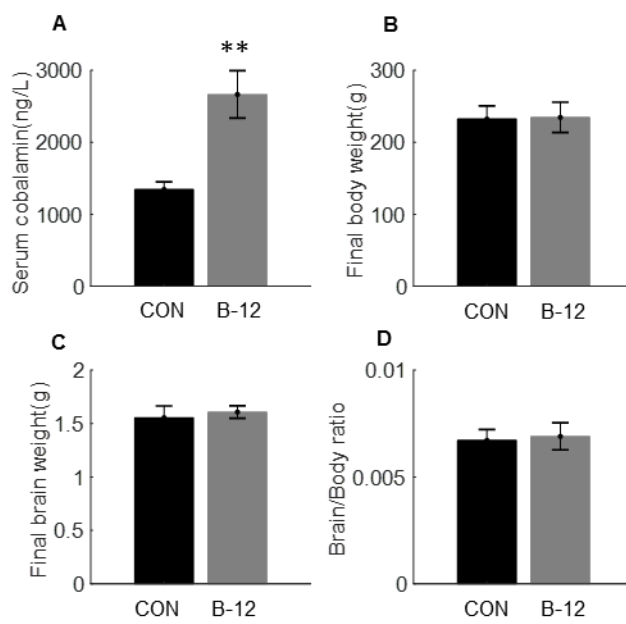




Figure 2

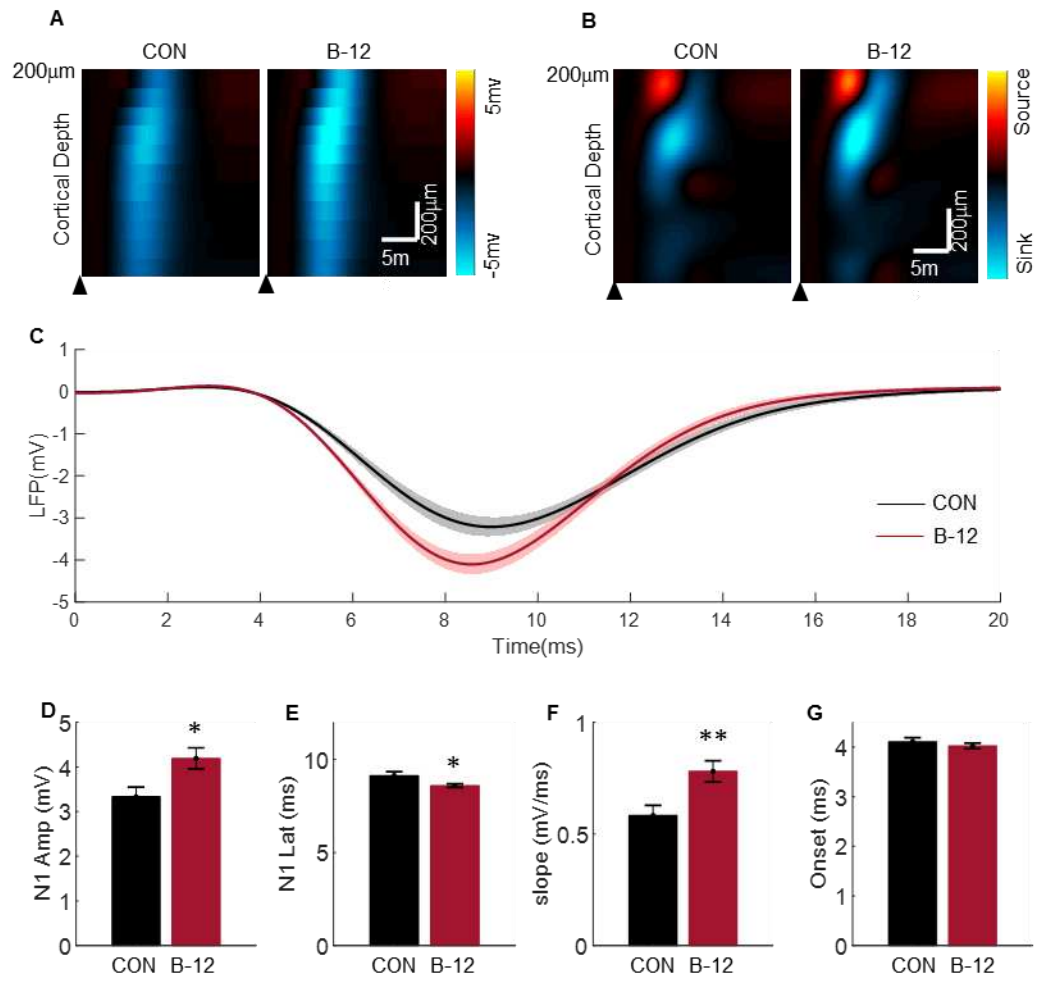


Figure 3

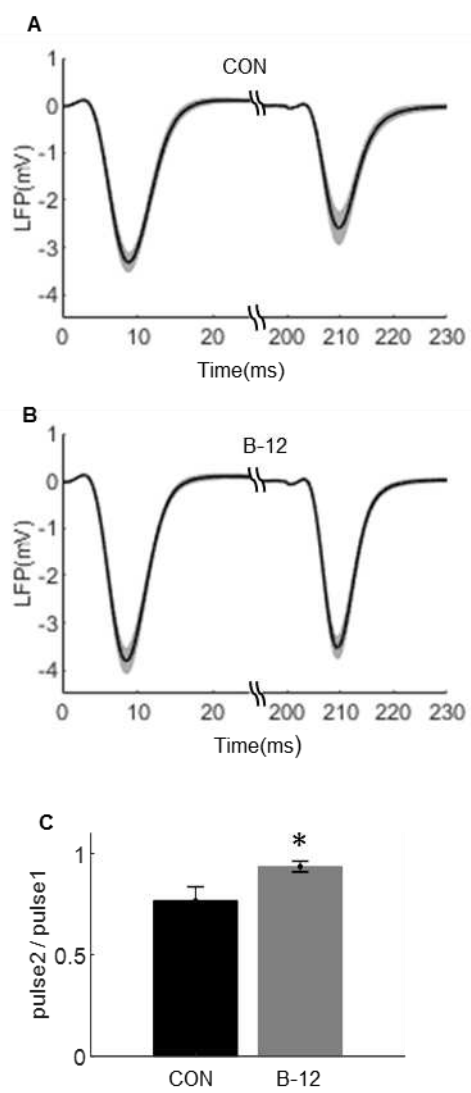


Figure 4

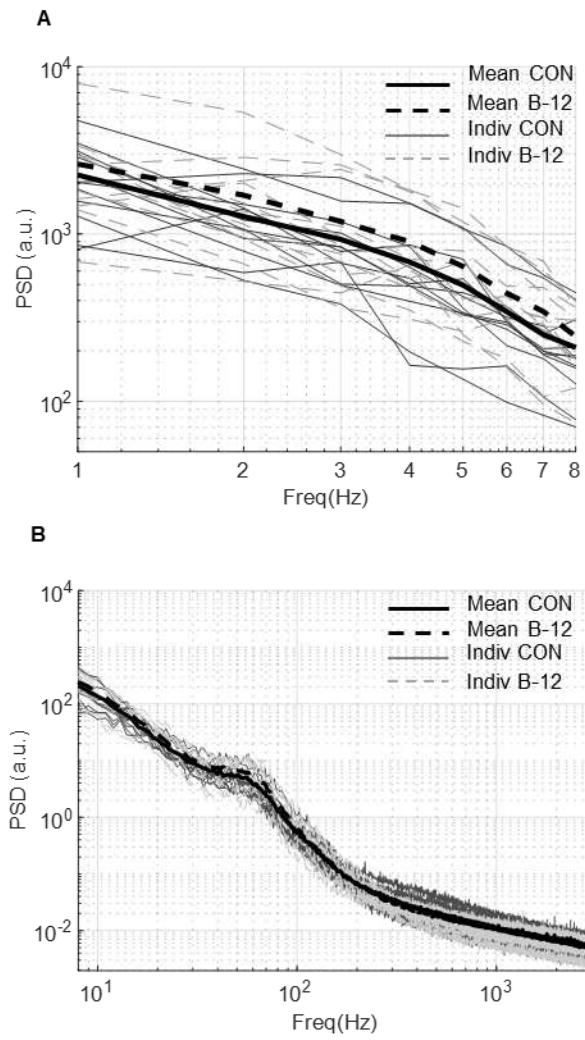


Figure 5

