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1 The effect of mating and the male sex peptide on group behaviour of post-mated female  
2 *Drosophila melanogaster*

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11 **Key words** *Drosophila*, Sleep, Social behaviour, Sex Peptide, Seminal fluid

12

13 **Abstract**

14 Sleep is a highly conserved state in animals, but its regulation and physiological function is poorly  
15 understood. *Drosophila melanogaster* is an excellent model for studying sleep regulation and has been  
16 used to investigate how sex and social interactions can influence wake-sleep profiles. Previously we  
17 have shown that copulation has a profound effect on day time activity and quiescence (siesta sleep) of  
18 individual post-mated females. Here we have studied the effect of mating and the transfer of the 36  
19 amino acid sex peptide in the seminal fluid on the behavior of mated female *Drosophila* populations,  
20 where there will be on-going social interactions. The locomotor activity and sleep patterns of virgin  
21 and post-mated female *D. melanogaster* from three laboratory strains (Oregon-R, Canton-S and  
22 Dahomey) were recorded in social groups of 20 individuals in a 12-12h light-dark cycle. Virgin  
23 female populations from all three fly strains displayed consolidated periods of low activity in between  
24 two sharp peaks of activity, corresponding to lights-on and lights-off. Similar light-correlated peaks  
25 were recorded for the mated female populations, however, the low afternoon activity and siesta seen in

26 virgin populations was abolished after mating in all three strains. In contrast, night activity appeared  
27 unaffected. This post-mating effect was sustained for several days and was dependent on the male SP  
28 acting as a pheromone. Evidence from mixed populations of virgin and mated females suggests that  
29 the siesta of non-mated females is not easily disturbed by the presence of highly active post-mated  
30 females.

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## 35 **Introduction**

36 Sleep, a rapidly reversible resting state accompanied by reduced response to sensory stimuli,  
37 is an enigmatic behaviour that is highly conserved in animals from nematodes to humans [1-  
38 4]. Sleep is critical for animal well-being, however its function and neuronal regulation is  
39 poorly understood [5,3,6]. Two important characteristics of sleep is the regulation by an  
40 internal circadian clock as well as a homeostatic mechanism that can compensate for sleep  
41 disturbance [7]. *Drosophila melanogaster* is proving to be a model par excellence in resolving  
42 the cellular mechanism of the circadian clock and, more recently, for identifying genes  
43 involved in sleep regulation and in unravelling the neuronal networks involved in promoting  
44 wake and sleep states[8,9,4,10] . The importance of neuropeptide signalling in the clock cell  
45 network and in promoting and maintaining sleep in *D. melanogaster* is becoming increasingly  
46 apparent and has highlighted mechanisms by which wake-sleep profiles can be coordinated  
47 with other physiological events, such as feeding and reproduction[11-16].

48 In a previous study we showed that a male 36 amino acid peptide, known as the sex  
49 peptide (SP), can change the wake-sleep pattern of post-mated female *D. melanogaster*. SP is  
50 a well characterised male modulator of female *Drosophila* behaviour serving as a multi-  
51 functional signalling molecule that is passed to the female in the ejaculate [17,18]. The change  
52 in female behaviour by SP is triggered by the silencing of sensory neurons of the female  
53 reproductive tract that communicate with the peptidergic cells of the pars intercerebralis, a  
54 brain neuroendocrine centre homologous to the vertebrate hypothalamus and known to be  
55 involved in the regulation of sleep as well as feeding and reproduction[19,20. SP is synthesised  
56 in the male accessory gland of adult flies as a 55 amino acid preprohormone{Chen, 1988 #702}.  
57 After processing, the mature peptide is secreted into the seminal fluid and on mating is  
58 transferred in the ejaculate to the female where it elicits numerous post-mating behavioural and  
59 physiological responses (PMRs)[21,22], including increased rate of egg-laying, reduced  
60 attractiveness to and rejection of courting males, stimulation of juvenile hormone synthesis  
61 [23-25], yolk accumulation in oocytes [26], increased appetite[27] and altered food preferences  
62 [28-31], elevated rate of excretion [32], loss of day-time sleep [33], enhanced female  
63 aggression [34], release of stored sperm [35] and modulation of the female innate immune  
64 system [36]

65 A notable structural feature of SP is a Trp-rich N-terminal region, which is not required  
66 for biological activity, but is responsible for SP binding to the surface of sperm tails. This

67 attachment means that SP is transferred with sperm to the sperm storage organs and can be  
68 released over time by proteolytic cleavage at a trypsin-like cleavage site [37,38]. This provides  
69 a mechanism by which the male can extend its influence on female behaviour to several days  
70 by the gradual release of the active SP from the sperm surface. In addition to the sperm-binding  
71 domain, there are two other distinct functional domains; a central region comprising the five 4-  
72 hydroxyproline residues and a modified isoleucine, and a C-terminal section (SP<sup>21-36</sup>)  
73 responsible for receptor binding and initiation of many of the female PMRs [21]. The 4-  
74 hydroxyproline-rich central domain appears to have a role in eliciting an early (24 h after  
75 mating) female immune response to mating [36] [38]. The C-terminal signalling domain (SP<sup>21-</sup>  
76 <sup>36</sup>) is critical for activating the G protein-coupled receptor expressed in sensory neurons of the  
77 female uterus that result in silencing of their neuronal activity. This signalling domain includes  
78 a peptide ring structure with a disulphide bridge between cysteine<sup>24</sup> and cysteine<sup>36</sup> [39] [40],  
79 which is not necessary for receptor activation, but probably protects the peptide from  
80 degradation by seminal fluid peptidases [41].

81         Our observation that the male SP can change the sleep behavior of post-mated females,  
82 added another response to the long list of PMRs triggered by the transfer of this male  
83 pheromone to the female during copulation [33]. When adult *D. melanogaster* are placed in a  
84 light/dark cycle, they display two periods of intense wakeful locomotor activity, one at lights-  
85 on and the other around the time of the light-dark transition. In between these peaks of activity  
86 there are periods of quiescence, or sleep, a behavior that is sexually dimorphic with males  
87 sleeping more than females during the afternoon or siesta period [7,42]. The flies' siesta is a  
88 possible adaptation for survival during hot afternoons which might place individuals at risk  
89 from desiccation [43,44]. Females as well as males should benefit from inactivity during the  
90 siesta, but females differ from males in that they need to balance risks with the demands of  
91 reproduction, which include foraging, to satisfy an increase in appetite and the need for a high-  
92 protein diet to sustain egg production, as well as the seeking of egg laying sites [45]. However,  
93 virgin females are not under the same pressures and therefore appear to reduce exposure to  
94 environmental risks by reducing locomotor activity and increasing levels of day-time  
95 quiescence to levels similar to that seen in males. SP appears to be the molecular switch that  
96 changes the behavior of post-mated females by increasing locomotor activity and reducing  
97 sleep [33]. A similar response to mating by female *D. melanogaster* was also observed in other  
98 studies and in the related fruit fly *D. sukuzii*, which also receives SP in the male seminal fluid  
99 [46,20,47].

100 The circadian timing of locomotor rhythmic activity and the wake-sleep architecture of *D.*  
101 *melanogaster* are influenced by social interactions experienced by couples or flies housed in larger  
102 groups {Levine, 2002 #220;Ganguly-Fitzgerald, 2006 #1010}. For example, Ganguly-Fitzgerald et al.  
103 showed that a 5-day enriched social experience amongst same and mixed sex adults can substantially  
104 increase the amount and quality of sleep of individuals compared to flies that have been deprived of any  
105 social interactions from eclosion [48]. This effect of social interaction on wake-sleep balance resulted  
106 mainly from an increase in day-time sleep and could be reproduced when flies were kept in same-sex  
107 pairs for 3 or more days, resulting in enhanced day-time sleep for male, but not female, *D. melanogaster*.  
108 Recently, population activity monitors have been employed to show that sleep in male and female  
109 populations of 50 flies are regulated by both circadian and homeostatic mechanisms, as reported for  
110 individual flies [49]. This study, however, revealed some sleep differences between populations and  
111 individuals, possibly from olfactory communication between flies within a population. Sexually  
112 dimorphic sleep behavior was also reported with males sleeping more during the day than females,  
113 however, the study did not investigate any impact of mating on sleep on the female population [49].

114 In the present study, we have extended our earlier investigation of the effect of mating on female  
115 sleep from individuals to populations. We now show that mating has a profound effect on the activity  
116 of socially enriched female flies resulting in increased day light locomotor activity and loss of siesta  
117 sleep. Males lacking SP in their ejaculate do not elicit a strong sleep PMR in the female population.  
118 Locomotor data collected from mixed populations of virgin and mated females suggests that the siesta  
119 sleep of virgin flies is robust and is not disturbed by the afternoon excitable activity of the post-mated  
120 population.

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## 123 **Materials and methods**

### 124 **Fly strains**

125 Oregon-R were from an established stock maintained in our laboratory for over 20 years. Canton-S and  
126 Dahomey wildtype strain were provided by S. T. Sweeney, University of York, U.K. and T. Chapman,  
127 University of East Anglia, U.K., respectively. SP null mutants (SP<sup>0</sup>) and control wildtype flies (SP<sup>+</sup>)  
128 were generated as described previously using mutant stocks, provided by S. Wigby, University of  
129 Oxford, U.K. and originating from the laboratory of E. Kubli [37,34].

## 130 **Fly culture**

131 Flies were cultured on oatmeal/molasses/yeast/agar medium at 25 °C in 12h:12h light-dark cycle and  
132 were sexed at the pupal stage on the basis of presence/absence of male sex combs.

## 133 **Recording locomotor activity and sleep of fly populations**

134 All experiments were conducted at 25 °C in a 12h:12h light-dark cycle. Unless stated otherwise, female  
135 flies (1 day-old) were mated with males by placing 10 virgin females with 10 virgin males in vials (95  
136 x 25 mm) containing oatmeal/molasses/agar diet for 3 days. Virgin females were kept in groups of 20  
137 for the same length of time under identical conditions. After 3 days, these flies were lightly anaesthetised  
138 using CO<sub>2</sub>, separated by sex and placed in glass vials (95 x 25 mm) containing 6 ml of 2 % (w/v) agar  
139 and 5 % (w/v) sucrose. Vials (95 x 25 mm) were placed in *Drosophila* population activity monitors  
140 (DPM, Trikinetics Inc. Waltham, U.S.A.) that use three arrays of infrared (IR) beams, each set  
141 comprising 15 beams and placed in three positions along the length of the glass vial to detect movement  
142 as the fly walks along the glass tube (Fig. 1a). The apparatus was kept in a vertical position with the  
143 bottom array 1 positioned just above the agar/sucrose, the middle array 2 recorded movement half-way  
144 along the vial and the top array 3 was located close to the cotton plug at the open end of the vial. The  
145 total number of beam breaks was obtained by summing the data for all three sets of IR beams in 5 min  
146 or 30 min time-bins for each sex and strain, and the data analysed using Microsoft Excel. Flies were  
147 allowed to acclimatise for 12 h before data were utilised for analysis. For the purpose of this study,  
148 group sleep was defined as a period of 5 min with no locomotor activity detected by any of the three  
149 arrays of IR beams. Statistical analysis was carried out using GraphPad Prism 7.01.

## 150 **Results**

151 The Trikinetics DPM monitors allow the recording of locomotor activity of adult insects as they break  
152 three sets of IR beams positioned (i) just above the food, (ii) half way along the length of the population  
153 vial and (iii) just below the cotton plug (Fig. 1a). DPMs were used to compare the activity of populations  
154 of virgin and post-mated female *D. melanogaster* from three common laboratory strains (Oregon-R,  
155 Canton-S and Dahomey) in a 12:12h light-dark cycle and constant temperature and humidity. Flies were  
156 placed in monitoring vials (20 females per vial) and were allowed to acclimatise for 12 h before activity  
157 data were collected for analysis. Virgin females of all three strains displayed two prominent peaks of  
158 population activity around lights-on (morning) and lights-off (late afternoon/evening) (Fig.1 b, c and d),  
159 separated by periods of very low activity that lasted for up to 6 h during the middle (afternoon) of the  
160 light period and for up to 9 h during lights-off (night). Mated female populations behaved similarly to

161 their virgin counterparts during night time, but during day-light hours the populations remained very  
162 active during the afternoon, which contrasted with the quiescence of the virgin flies at the same time of  
163 day (Fig. 1 b,c,d). All three arrays of detector beams were repeatedly broken by moving flies showing  
164 that the increased day-time activity of the post-mated population was not restricted to any one position  
165 in the vial, although relatively greater activity was usually detected by array 3 which was furthest away  
166 from the food (Fig. 2e,f).

167 To assess the influence of the male SP on the behaviour of mated female populations,  
168 Oregon-R females were mated with SP<sup>0</sup> males that do not make SP, but otherwise have  
169 normal seminal fluid. The resulting PMR was compared with that of populations of females  
170 mated with genetically matched control flies (SP<sup>+</sup>) and virgin females [37] [34]. The high  
171 afternoon activity of the mated female population recorded previously was reproduced when  
172 SP<sup>+</sup> males producing normal levels of SP were used for insemination, but not when mated  
173 with SP null (SP<sup>0</sup>) males (Fig. 2a). The high level of SP-induced afternoon activity of the  
174 female population progressively declined with time until at around 7 days it reached the same  
175 level of day light activity recorded for the virgin female population (Fig. 2b). Night activity  
176 also declined steadily over this period, but there was no apparent difference in the population  
177 activity between virgin females and females mated with either SP<sup>+</sup> or SP<sup>0</sup> males (Fig. 2c),  
178 emphasising that this SP-induced PMR only occurred during the afternoon period.

179 The locomotor activity data was transformed to provide a measure of sleep, defined as 5 min  
180 time bins in which no movement in the entire population was detected by any of the three sets of IR  
181 arrays (Fig. 3). For the first 6 days of the experiment, the afternoon siesta, a characteristic of the virgin  
182 female population, was essentially abolished for females mated with control SP<sup>+</sup> males. In contrast, the  
183 female population mated to males lacking SP (SP<sup>0</sup>) did sleep during the afternoon, although the total  
184 sleep was not as great as that experienced by the virgin female population. The relatively low sleep value  
185 for day 1 probably reflects poor acclimatisation after transfer to the DAM vials. Mating with males  
186 expressing SP, but not with SP null males, also appeared to trigger a reduction in night-time sleep,  
187 however, only by around 25%.

188 To investigate possible day light social interactions between virgin and mated females, the  
189 locomotor activity of a mixed population comprising 10 mated and 10 virgin females was compared  
190 with a population of 20 virgins (20) and a population of 10 mated females for 2 days in the standard  
191 12:12h light-dark cycle. The expected peaks of morning and evening activity were observed for all three  
192 populations as well as the mating-induced rise in afternoon activity in the mated fly population compared



193 to the virgin population (Fig. 4a). Increasing the population of flies from 10 to 20 by mixing mated and  
194 virgin females increased the activity levels at lights-on and lights-off, but not during the afternoon siesta  
195 period. To emphasise this point and provide statistical support, the day light activity for the three female  
196 populations was split into four 3 h periods. Period 1 covers the morning, period 2 and 3 is the midday  
197 afternoon and period 4 is the late afternoon/evening. As expected, mixing 10 mated and 10 virgin  
198 females raised activity levels during peak periods 1 and 4 when compared to the levels recorded for 10  
199 mated females. In contrast, mixing populations of mated females (10) with virgin females (10) made no  
200 significant difference to the activity levels during the siesta periods 2 and 3 compared with 10 mated  
201 females, suggesting that the virgin flies remain quiescent despite the elevated activity of the co-housed  
202 10 mated females (Fig.4b).

## 203 **Discussion**

204 Previous studies have shown that socially enriched individuals sleep more compared to flies that are  
205 socially deprived [48,50]. These studies focused on the social experience prior to the monitoring of  
206 sleep and wakeful activity in individual flies and therefore differed from the present study and that of  
207 Liu et al. [49], who studied social behaviour in populations where flies experience ongoing interactions  
208 with other members of the community. Liu et al. used the LAM25H Trikinetics activity equipment  
209 which allowed monitoring of the activity/sleep behaviour of populations of 50 adult *D. melanogaster*  
210 [49]. This system although using vials of the same dimension as those used in the present study, differed  
211 significantly from our population monitors (DPM) in that the LAM25H has just one set of IR beams and  
212 detectors positioned to detect moving flies in the central axial region of a horizontal population vial.  
213 DPMs have 3 sets of IR beams/detectors positioned not only to detect flies crossing the middle of the  
214 vials, but also to detect flies moving close to the food surface and at the cotton plug interface. The study  
215 of Liu et al. showed that sleep/wake behaviour of same-sex populations was under both circadian and  
216 homeostatic control and like individual flies was sexually dimorphic [49]. Some differences between  
217 the behaviour of individuals and populations were however reported (e.g. more rapid synchronisation of  
218 sleep onset in populations) and these were likely the result of social interactions mediated by multiple  
219 sensory stimuli including visual, tactile and olfactory [49]. Interestingly, when females were placed  
220 with males in a mixed-sex population (female to male ratio of 2:1) total day time sleep was much lower  
221 than that recorded for both single sex populations, suggesting that sexual encounters were stimulating  
222 activity of both sexes.

223 The present study focused on the behaviour of female-only populations of *D. melanogaster* and  
224 the impact of mating status on the sleep/activity states of different laboratory strains (Oregon-R, Canton-  
225 S and Dahomey). This study allowed comparison with our previous published work describing the role

226 of SP in abolishing the siesta sleep of individual virgin females [33]. The stimulating social environment  
227 of a population might be expected to increase the level of activity and reduce quiescence in the  
228 population, especially during the afternoon when sleep is less intense. Our data shows that for all three  
229 strains of *D. melanogaster*, virgin females display synchronised activity and sustained quiescence during  
230 both night and the afternoon siesta period despite the obvious potential for disruptive social interactions  
231 with other individuals in the population. Higher activity levels were noted for the Dahomey strain  
232 compared to the other two. The reason for this difference is not clear, however it has been previously  
233 reported that strains and even sub-strains can have markedly different locomotor behaviour [51,20].

234 We have previously shown that mating results in increased in locomotor activity and concomitant  
235 loss of sleep during the afternoon for individual females and that the male SP is the principal molecule  
236 responsible for switching female behaviour [33]. The same mating-induced loss of a siesta has now  
237 been reproduced in female populations for three strains of *D. melanogaster*. This change in population  
238 behaviour, at least for Oregon-R, is SP-dependent and persists for up to 1 week, presumably because of  
239 the previously reported slow-release of the peptide from stored sperm in the female [38]. A similar  
240 persistence of the post-mating response was observed in the earlier study of individually housed post-  
241 mated females [33]. Mixing of equal numbers of mated flies with high afternoon activity with virgin  
242 females experiencing a siesta period resulted in significant increase in the morning and evening peaks  
243 of activity because of the larger population size. This increase was not 100%, probably because the  
244 relationship between number of beam breaks and population size is not linear due to the greater chance  
245 of multiple flies breaking beams in the same time bin as the number of individuals in the vial increases.  
246 In contrast, doubling the population size did not increase the afternoon activity of the mixed population  
247 at all, suggesting that the siesta of virgin females is robust and not easily disrupted by the presence of  
248 the more excited post-mated females in the population.

249 In summary, this study has established that the SP-induced loss of the afternoon siesta by female  
250 *D. melanogaster* that was first observed in individual flies, can be replicated in populations experiencing  
251 ongoing social interactions and that the change of behaviour mated females is independent of fly strain.  
252 Furthermore, the siesta of virgin females appears to be maintained in mixed populations comprising  
253 equal number of the resting virgins and the very active post-mated females.

254

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396 **Figure legends**

397 Fig.1. Mating elevates afternoon locomotor activity in populations of female *D.*

398 *melanogaster*. The activity of virgin (---) and mated (—) populations was monitored over  
399 48 h using DAM population monitors that record the movement of flies breaking three arrays  
400 of IR beams positioned as indicated in **a, b, c, d** Activity of populations (20 females) of  
401 Oregon-R, Canton-S and Dahomey strains, respectively. Populations were maintained in a  
402 12:12h light-dark cycle indicated by the open (lights-on, day light) and solid black bars  
403 (lights-off, night). The activity is the sum of the beam breaks for all three arrays of IR beams  
404 recorded in 30 min time bins. **e,f** Beam breaks/30 min recorded by each of the three arrays for  
405 **e** virgin Dahomey population and **f** mated Dahomey population. These activities were  
406 summed to generate the data plotted in **d**.

407 Fig. 2 Sex peptide (SP) of the male seminal fluid is necessary for elevating the afternoon  
408 activity in populations of post-mated *D. melanogaster* females. **a** Population activity of virgin  
409 Oregon-R females (●●●) and females mated to either SP null (SP<sup>0</sup>, - - -) or control (SP<sup>+</sup>,  
410 —) males kept in a 12:12 h light-dark cycle indicated by the open (day light) and solid black  
411 bars (night). **b** Data from **a** plotted as total population activity in the 12 h of day light. **c** Data  
412 from **a** plotted as total population activity in the 12 h of night.

413 Fig. 3 The siesta sleep of a virgin female population is abolished after mating with males  
414 expressing SP, but not when mated with males lacking seminal fluid SP. Data from Fig.2 was  
415 transformed to minutes of sleep/12 h, calculated from the number of 5 min periods of zero  
416 beam breaks recorded by any of the three sets of arrays of the DAM population monitors. **a**  
417 amount of day light (siesta) sleep and **b** night sleep experienced by virgin females and  
418 females mated to either SP null (SP<sup>0</sup>) males or control (SP<sup>+</sup>) males over 7 days.

419 Fig.4 The effect of mixing mated and virgin female populations on locomotor activity. **a** The  
420 population activity expressed as total beam breaks/h for 10 mated females (---), 20 virgin  
421 females (●●●) and a mixed population of 10 virgin and 10 mated females (—). The plotted  
422 activities are the means of data collected over 48 h from 4 separate experiments using flies of  
423 the Dahomey strain. The data points and error bars have been omitted for clarity. Open and  
424 solid black bars indicate the light and dark periods, respectively. **b** Activity data from the first

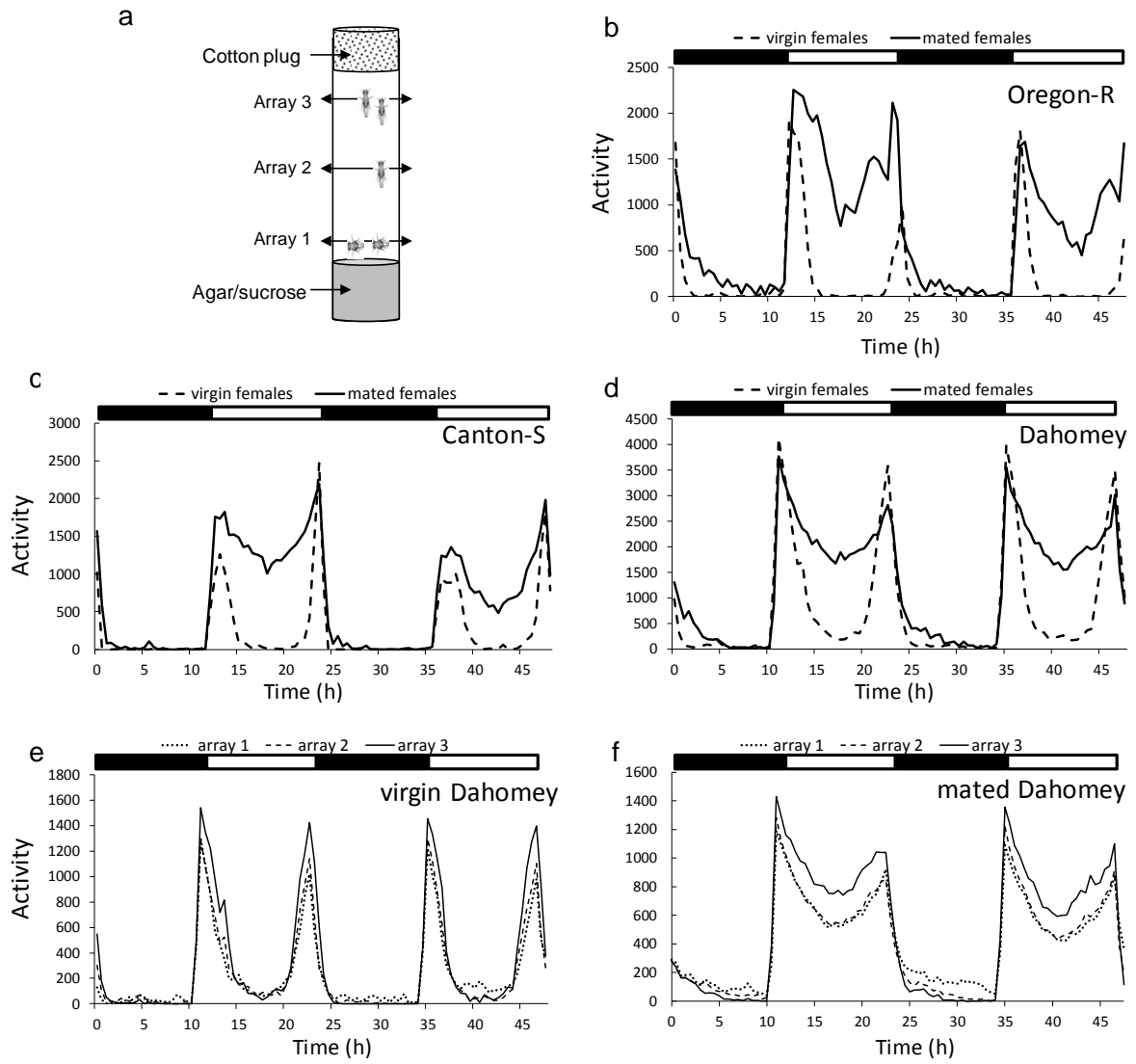
425 day in **a** were summed into four 3 h day light periods (period 1, 13-15 h; period 2, 16-18 h;  
426 period 3, 19-21 h; period 4, 22-24 h) and expressed as the mean  $\pm$  s.e.m. (n = 4). Statistical  
427 analysis was conducted using student's t test and one-way ANOVA (GraphPad Prism 7.01).  
428 ns, P > 0.05; \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001. Similar results were obtained when the  
429 second 24 h of data was analysed in the same way (plots not shown).

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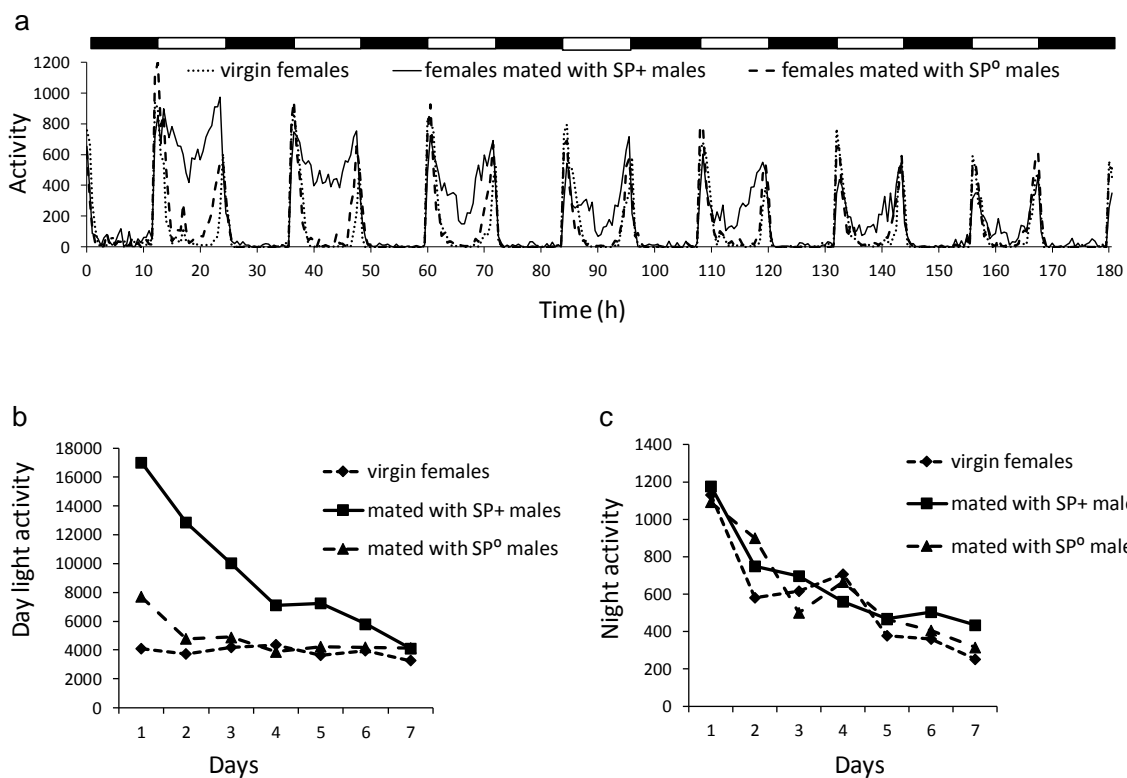
Fig.1



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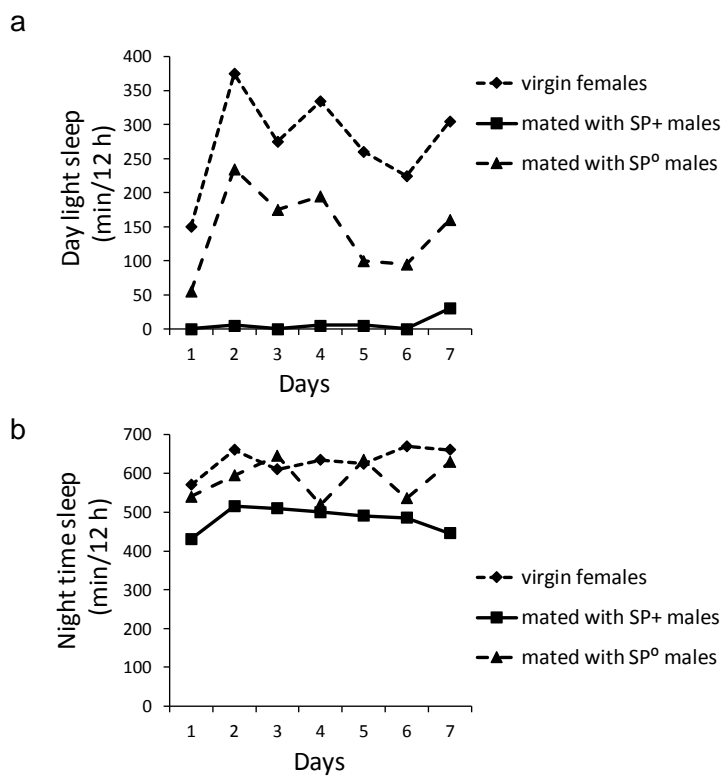


Fig. 2



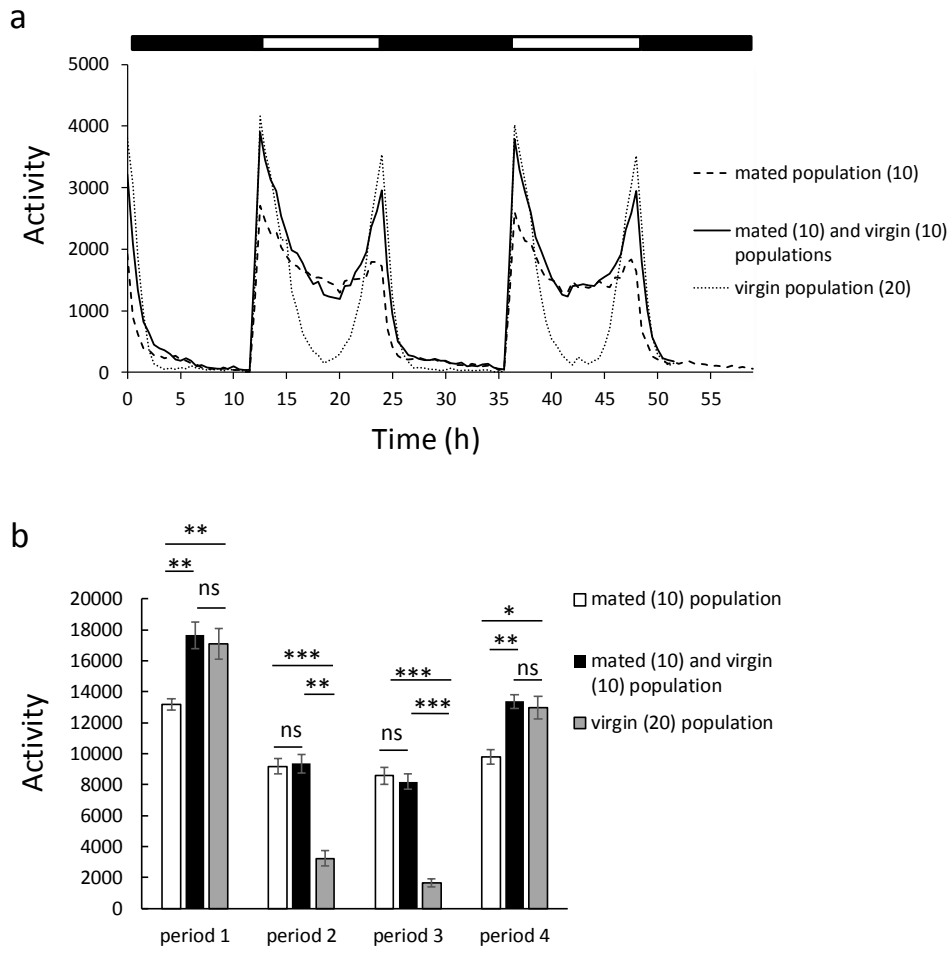
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Fig. 3



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Fig. 4



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