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Dunneram, Y orcid.org/0000-0002-1012-7350, Greenwood, DC
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Diet, menopause and the risk of ovarian, endometrial and breast cancer

Yashvee Dunneram¹, Darren C. Greenwood², Janet E. Cade¹

1. Nutritional Epidemiology Group, School of Food Science & Nutrition, University of
Leeds, Leeds, UK

2. Division of Epidemiology and Biostatistics, University of Leeds, Leeds, UK

Correspondence to: Yashvee Dunneram; fsyd@leeds.ac.uk

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1 **Abstract**

2 Menopause, the permanent cessation of the menstrual cycle, marks the end of a woman’s
3 reproductive span. In addition to changes in sex hormone levels associated with menopause,
4 its timing is another predictor of future health outcomes such as duration of the presence of
5 vasomotor symptoms (VMS) and the risk of hormone-related cancers. With aging of the
6 population, it is estimated that worldwide 1.2 billion women will be menopausal by the year
7 2030. Previously the effects of reproductive factors (e.g. parity, age at menarche, pregnancy)
8 and socio-demographic factors on intermediate and long-term health outcomes of menopause
9 have been widely documented. However, little is known about whether diet could have an
10 impact on these. Therefore, we review current evidence on the associations of diet with
11 menopause, presence of VMS and the risk of hormone-related cancers such as ovarian,
12 endometrial and breast cancer. Dietary factors could influence the lifespan of the ovaries and
13 sex-hormones levels, hence the timing of natural menopause. Few studies reported an
14 association between diet, in particular soy consumption and a reduced risk of VMS. Sustained
15 estrogen exposure has been associated with a higher risk of hormone-related cancers and thus
16 high fat and meat diets have been linked with an increased risk of these cancers. However, to
17 better understand the mechanistic pathways involved and to make stronger conclusions for
18 these relationships, further studies investigating the associations of dietary intakes and dietary
19 patterns with menopause, presence of VMS and the risk of hormone-related cancers are
20 required.

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34 **Introduction - Menopausal transition and Menopause**

35 Menopause, the last menstrual period, marks the end of reproductive life in women. With aging
36 of the population, it is estimated that worldwide 1.2 billion women will be menopausal by the
37 year 2030⁽¹⁾. While menopause is inevitable, the age at which women reach menopause may
38 vary depending on several factors such as geography and ethnicity. According to a meta-
39 analysis of 36 studies (which included data from 35 countries), the overall mean age of natural
40 menopause was 48.8 years (95% CI 48.3–49.2) with substantial geographic variation. For
41 example, while the mean age of menopause in the United States (49.1y) and Asia (48.8y) were
42 closest to the overall mean, it was higher in Europe (50.5y) and Australia (51.3y) and lower in
43 Africa (48.4y), Latin America (47.2y) and the Middle East (47.4y)^(2, 3).

44 At birth, the human ovaries contain approximately 1,000,000 primordial follicles⁽⁴⁾. This un-
45 replenishable pool of follicles is further reduced to around 100,000 per ovary by the time of
46 menarche. The fate of the remaining follicles is either to develop, reach maturity and then
47 ovulate or degenerate by the process known as atresia⁽⁵⁾. At the perimenopausal transition stage,
48 only about 100 to 1,000 follicles are left in each ovary and exhaustion of the follicle pool is
49 accompanied by permanently elevated levels of pituitary gonadotrophins and the progressive
50 reduction in antimullerian hormone (AMH) which confirms ovarian senescence⁽⁶⁾. The
51 hypoestrogenic changes taking place during the perimenopause (menopausal transition) are a
52 result of the interactions taking place between the hypothalamic-pituitary axis and the
53 reproductive endocrine axis marking this irreversible decline in ovarian responsiveness⁽³⁾.

54 The menopausal transition is the shift from normal reproductive life to the last menstrual period
55 and can last for up to 10–15 years⁽⁷⁾. According to the Staging of Reproductive Aging
56 Workshop⁽⁸⁾, it is divided in two stages: early and late. The early menopausal transition is
57 marked by changes in menstrual cycle length and is characterised by an increase in follicle-
58 stimulating hormone (FSH) level, a decrease in AMH and inhibin B levels, while estrogen level
59 remains stable. The late transition is marked by oligomenorrhea (infrequent periods) and can
60 last for 1–3 years on average. This stage is accompanied by an increase in anovulatory cycles
61 and also major fluctuations in hormonal levels. FSH level remains elevated while there is a
62 consequent decrease in AMH and inhibin levels as well as estrogen level. After the final
63 menstrual period, ovarian ageing is marked by a decrease in antral follicular count, and
64 termination of ovulation and menstruation. In addition, there are further declines in AMH,
65 inhibin and oestradiol levels^(9, 10). Ovarian ageing is also accompanied by loss of
66 responsiveness to FSH and luteinising hormone (LH), hence causing a disruption in the
67 negative feedback mechanism owing to the almost negligible inhibin level and decline in

68 estrogen level. Consequently, the production of gonadotropin-releasing hormone (GnRH) is
69 upregulated, stimulating the release of FSH and LH. Thus, during the initial years after
70 menopause the level of FSH peaks and gradually declines in the last postmenopause stage⁽³⁾.
71 These hormonal fluctuations as a result of the neuroendocrine and reproductive endocrine
72 interactions influence the risk of both intermediate and long term health outcomes associated
73 with menopause⁽³⁾. One of the most common intermediate sequelae of the menopause
74 transition, vasomotor menopausal symptoms (VMS), is defined as either the presence of hot
75 flushes and/or night sweats. VMS is reported by 40-60% perimenopausal women and 8-80%
76 postmenopausal women around the world⁽¹¹⁾. The timing of onset of menopause can influence
77 the length of the menopausal transition and hence the duration for the presence of VMS.
78 Evidence also shows a link between an early onset of menopause and an increased risk of
79 osteoporosis, cardiovascular disease, depression and mortality. On the other hand, a later age
80 at menopause has been associated with a higher prevalence of hormone-related cancers such
81 as breast, endometrial (uterine) and ovarian cancers⁽¹²⁾. Moreover, the presence of VMS has
82 also been associated with an increased risk of cardiovascular disease⁽¹³⁾. Previous studies have
83 demonstrated a link between reproductive factors, sociodemographic factors and the onset of
84 natural menopause, presence of VMS and risk of hormone-related cancer^(14, 15, 16). However, its
85 relationship with diet, a modifiable risk factor, has received less attention and current evidence
86 of association is conflicting. Therefore, the aim of this review is to give an overview of the
87 mechanistic pathway relating diet with age at natural menopause as well as to elucidate the
88 relationship between diet and VMS (an intermediate sequelae of menopause) in addition to the
89 risk of hormone-dependent cancers such as breast, endometrial and ovarian cancers (long-term
90 outcomes of menopause) which are more commonly prevalent in developing countries.

91

92 **Age at natural menopause**

93 Natural menopause refers to cessation of the menstrual cycle without any surgical procedures
94 such as oophorectomy or ovarian failure as a result of chemotherapy or radiotherapy⁽³⁾. A
95 premature menopause is one which is reached before the age of 40 years, an early menopause
96 between 40-45 years and a late menopause is one after the age of 55 years^(17, 18). Depletion of
97 the ovarian reserve and its responsiveness to pituitary gonadotropins governs the lifespan of
98 the ovary and thus influence the onset of the timing of the natural menopause⁽¹⁹⁾. Dietary factors
99 and diet-related disorders can either enhance the lifetime of the ovaries by delaying follicular
100 atresia or by maintaining sex-hormone levels involved in the feedback mechanisms of the
101 menstrual cycle. However, the exact mechanisms still need to be elucidated. The association

102 of age at natural menopause with chronic disease, aging and general health makes it an
103 important subject of clinical and public interest⁽¹²⁾.

104 Metabolic disorders such as diabetes could accelerate reproductive ageing by causing
105 premature ovarian failure through several mechanisms. This has been demonstrated in a
106 study⁽²⁰⁾ including women from 11 Latin American countries. The author reported that diabetic
107 women had an earlier menopause as opposed to non-diabetic women. Similarly, a recent study
108 conducted in the Southern part of India demonstrated that an early menopause was more likely
109 to be reported by diabetic women⁽²¹⁾. This is further supported by a British study which
110 investigated the association between various food groups and the timing of the onset of natural
111 menopause among 914 women in the Women's Cohort Study. It found that a high consumption
112 of refined pasta and rice, high glycaemic index foods, were associated with an earlier onset of
113 natural menopause⁽²²⁾. Furthermore, findings from the prospective Nurses Health Study II
114 demonstrated that a high vitamin D intake was associated with a lower risk of an early onset of
115 menopause⁽²³⁾ which could be due to the fact that a high serum 25-dihydroxyvitamin D
116 concentration could reduce the risk of diabetes as well as metabolic syndrome⁽²⁴⁾. These
117 findings thus indicate that the presence of type II diabetes, a diet-related disease could lead to
118 an earlier onset of menopause.

119 Vegetarianism has also been linked to an earlier age at natural menopause^(22, 25). Vegetarian
120 diets are usually characterised by a high dietary fibre and low fat content, particularly saturated
121 fats. They tend to include more whole grains, vegetable protein sources such as legumes, nuts,
122 and soy protein, and exclude red meat. Dietary fibre may potentially interfere in the
123 enterohepatic circulation of sex hormones, by modifying the metabolic pathway of estrogens,
124 leading to a decrease in estrogen bioavailability^(12, 26). Karelis et al.⁽²⁷⁾ demonstrated that
125 vegetarians had higher levels of sex-hormone binding globulin (SHBG), higher total fibre
126 intake as well as lower levels of free estradiol, free testosterone, dehydroepiandrosterone
127 sulfate and a lower BMI. An intervention study, also reported that a change in fibre intake was
128 significantly and independently associated with a decrease in serum bioavailable estradiol and
129 total estradiol concentrations while no association was found between a change in fat intake
130 and the hormone concentrations⁽²⁸⁾.

131 On the other hand, intakes of green and yellow vegetables as well as fresh legumes have been
132 associated with a delayed onset of menopause^(22, 29). Ovarian ageing is closely associated with
133 increased levels of reactive oxygen species (ROS) which arises mainly due to an imbalance
134 between ROS production and non-enzymatic antioxidant defences⁽³⁰⁾. Oocyte maturation,
135 ovulation, luteolysis and follicle atresia are all affected by ROS⁽³¹⁾. Antioxidant properties of

136 foods have been found to be positively associated with a reduced rate of follicular atresia. A
137 recent *in vivo* study demonstrated a reduced atretic follicle count with use of resveratrol (a
138 polyphenol found in the skin of red grapes and berries)⁽³²⁾. These contradictory findings could
139 be because while few studies looked at the associations with dietary patterns, others considered
140 the associations with individual food items. Moreover, differences in the participants'
141 characteristics and distribution of age at natural menopause could further influence the
142 findings. The confounders used in the analyses and large sample sizes could also explain the
143 differences.

144 High consumptions of meat, fat, and protein have been positively associated with a delayed
145 onset of menopause (Table 1). Cholesterol, the starting product of steroidogenesis can be
146 synthesised by *de novo* synthesis in the endocrine tissue (e.g. granulosa-lutein cells in the
147 ovaries) from acetate, the end-product of fat oxidation⁽³³⁾. Therefore, an excessive dietary fat
148 intake can result in higher serum estradiol levels. In addition, during the menopausal transition
149 significant changes occurs in body composition. For instance, redistribution of body fat takes
150 place such that there is an increase of total and central body fat, and also a redistribution of fat
151 from lower body subcutaneous fat toward the abdominal region. This increase in adipose tissue
152 becomes the main site for estrogen production along with other hormones such as leptin,
153 adiponectin and resistin^(34, 35). Therefore, these endocrine changes taking place during the
154 menopausal transition together with a high fat diet predisposes the woman to a later onset of
155 menopause.

156

157 **Menopause and its associated sequelae**

158 The timing of menopause could determine the duration of the presence of VMS which is mostly
159 prevalent during the perimenopausal years as a consequence of lowered estrogen levels (Figure
160 1). Previous randomised controlled trials have mainly focused on the study of phytoestrogen
161 extracts and their influence on the presence of VMS. However, the study of foods consumed
162 as part of the normal diet in relation to the presence of VMS has received less attention. The
163 decline in estrogen levels during the menopausal transition is postulated to be one of the causes
164 for the presence of VMS. A low estrogen level has been associated with narrowing of the
165 thermoneutral zone between the core body temperatures, resulting in a lowered sweating
166 threshold and hence a higher likelihood to experience hot flushes and night sweats. However,
167 given that around 20% of premenopausal women also report hot flushes suggests that the
168 decline in estrogen levels is not the sole endocrine change causing VMS⁽⁴⁷⁾. Dhanoya et al.⁽⁴⁸⁾

169 demonstrated that both AMH and FSH were associated with the presence of hot flushes while
170 the level of oestradiol was not related with hot flushes.

171 Prolonged exposure to estrogens as a consequence of a delayed menopause increases the risk
172 of hormone dependent cancers such as ovarian, endometrial and breast cancer as demonstrated
173 previously by several epidemiological studies^(49, 50, 51). Other hormones such as progesterone
174 may also be important. These hypotheses have been investigated in earlier published reviews^{(52,}
175 ^{53, 54)}. Other factors such as diet (Figure 1), a modifiable risk factor may also explain the
176 variation in estrogen and other sex hormones levels^(55, 56, 57). Diet-related pathologies may also
177 promote tumorigenesis while some components of the diet may be protective against these
178 cancers. Therefore, the next sections explore the evidence for the hypothesis that diet is a major
179 determinant for the presence of VMS and for the risk of hormone-related cancers.

180

181 **Presence of vasomotor symptoms**

182 VMS such as hot flushes and night sweats are one of the most common symptoms experienced
183 by women during the menopausal transition. The median duration of these symptoms is 4 years
184 but may persist as long as 15 years for some women⁽⁴⁷⁾.

185 Evidence for a link between diet and presence of VMS arises from studies which have
186 previously explored the associations between phytoestrogen extracts or phytoestrogen-rich
187 foods and frequency or severity of VMS. A Cochrane review of 43 randomised controlled trials
188 did not support the beneficial effects of phytoestrogen supplements for the reduction of the
189 frequency or severity of VMS mainly due to the small size of the trials and also the high risk
190 of bias while the same review stated the promising effect of genistein, a phytoestrogen found
191 in soy⁽⁵⁸⁾. A recent review further indicated the beneficial effect of isoflavones against hot
192 flushes⁽⁵⁹⁾.

193 As mentioned previously, women tend to accumulate subcutaneous fat in the abdominal region
194 during the menopausal transition which leads to endocrine changes in terms of higher
195 circulating oestradiol level⁽³⁴⁾. A prospective study of 6040 women demonstrated that a
196 Mediterranean-style diet and a fruit rich diet were both inversely associated with VMS. On the
197 other hand, diets with high fat and sugar contents increased the risk of VMS⁽⁶⁰⁾. This could
198 imply that a healthier diet which prevents obesity could also be protective against VMS. The
199 same study reported that even after adjusting for BMI, the same associations were observed.
200 Therefore, the mechanism involved between diet and presence of VMS still remains unclear.

201

202

203 **Ovarian cancer**

204 Women of reproductive age undergo cyclical cellular changes in their genital tract during the
205 menstrual cycle⁽⁶¹⁾. During each cycle several follicles containing an ovum undergo a
206 maturation and selection process where ordinarily one of them is selected and released from
207 the ovary during ovulation on or around the 14th day of the cycle⁽⁶²⁾. The menstrual cycle is
208 under the influence of various hormones namely gonadotrophin releasing hormone, luteinizing
209 hormone, follicle stimulating hormone, estrogen, and progesterone⁽⁶³⁾. During ovulation the
210 surface of the ovary ruptures to release the ovum, following which the cells on the surface of
211 the ovary, known as the epithelial cells, proliferate to close the breach under the influence of
212 estrogen. Improper proliferation of those cells can result in formation of cysts or even cancers
213 like surface epithelial tumours which are a sub group among the diverse types of ovarian
214 tumours⁽⁶⁴⁾.

215 Estrogen and progesterone are steroid hormones synthesised from cholesterol⁽³³⁾; individuals
216 having a high fat diet provide the substrate for excessive estrogen synthesis which stimulates
217 cell proliferation in the female genital tract. Diets high in animal protein also contains xeno-
218 estrogens which have carcinogenic potential⁽⁶⁵⁾. Leptin, another hormone secreted by the
219 adipose tissue under the influence of factors like high lipid levels in blood, has several effects
220 on the body like producing a feeling of satiety, as well as stimulating the release of GnRH
221 which in turn stimulates release of LH and FSH⁽⁶⁶⁾. High levels of LH may result in the
222 immature release of the ovum and high levels of estrogen secondary to high circulating
223 cholesterol levels in the body (as a result of high saturated fat and energy intake). Consequently,
224 this may result in improper re-epithelialisation of the ovaries. Chronic stimulation of ovaries
225 in this way may predispose to development of abnormal growths which subsequently can
226 undergo malignant transformation. Therefore, diets high in energy, fats, or animal protein may
227 promote development of ovarian cancer.

228 Omega-3 fatty acid, a polyunsaturated fatty acid (PUFA) can be obtained through dietary
229 sources (flaxseeds, walnuts, canola oil, and oily fish) only. The n-3 family of PUFAs comprises
230 alpha-linolenic acid (ALA), eicosapentaenoic acid (EPA), and docosahexaenoic acid (DHA).
231 According to *in vivo* studies, EPA and DHA have been found to be precursors for anti-
232 inflammatory lipid mediators⁽⁶⁷⁾. Estrogen has proliferative effects on estrogen-sensitive
233 tissues and thus could be involved in the pathogenesis of some hormone-dependent cancers
234 such as ovarian cancer. Dietary n-3 PUFAs deter the promotion and progression stages of
235 carcinogenesis through several mechanistic pathways. One of the mechanisms involves

236 changes in estrogen metabolism which could result in reduced estrogen-stimulated cell growth
237 (68, 69).

238 Prostaglandin E2 (PGE2), an arachidonic acid (an n-6 PUFA found in meat and fish)-derived
239 eicosanoid stimulates the activity of aromatase P450, which converts 19-carbon steroids to
240 estrogens while on the other hand, PGE3 (derived from EPA metabolism), does not activate
241 aromatase P450. Hence, an increased intake of EPA, which leads to increased production of
242 PGE3 and decreased production of PGE2, is expected to decrease estrogen production and thus
243 reduce estrogen-stimulated cell growth⁽⁶⁸⁾. In addition, n-3 PUFAs can influence the regulation
244 of two transcription factors; sterol regulatory element binding protein-1c (SREBP-1c) and
245 peroxisome proliferator activated receptor alpha (PPAR alpha). SREBP-1c is involved in
246 inducing a set of lipogenic enzymes in liver and n-3 PUFAs can potentially inhibit the
247 expression and processing of SREBP-1c and thus inhibits the de novo lipogenesis of fatty acids,
248 making it an important consideration for the carcinogenesis. Merritt *et al.* ⁽⁷⁰⁾ demonstrated that
249 a higher intake of omega-3 may be protective for ovarian cancer, while a greater consumption
250 of trans-fat was associated with an increased risk of ovarian cancer. However, the clinical
251 effects of n-3 PUFAs does not solely rely on its concentration alone, but most importantly on
252 the ratio of n-3 PUFAs to n-6 PUFAs in the cells⁽⁷¹⁾. For instance, in a study using a knockout
253 mouse model it was demonstrated that a dietary ratio of omega-6/omega-3 PUFA lower than 5
254 was effective in suppressing tumour growth, and prolonging animal lifespan⁽⁷²⁾. Thus, a high
255 intake of n-3 PUFAs relative to that of n-6 PUFAs may decrease endogenous estrogen
256 production and reduce the risk of ovarian cancer.

257 Along with hormonal control, diet can also interfere at the level of fatty acid (FA) and
258 cholesterol biosynthesis and eventually affect sex steroid metabolism and thus risk of ovarian
259 cancer⁽⁷³⁾. For instance, it has been found that feeding previously fasted animals a diet high in
260 carbohydrate and low in fat content causes a dramatic induction of enzymes such as fatty acid
261 synthase (FAS) and mitochondrial glycerol-3-phosphate acyltransferase (GPAT) which are
262 involved in FA and triacylglycerol (TAG) synthesis. FAS and GPAT are the two critical
263 enzymes involved in FA and TAG biosynthesis. FAS catalyses the synthesis of long-chain fatty
264 acids, primarily palmitate, using acetyl-CoA and malonyl-CoA as substrates and NADPH as
265 the reducing equivalent while GPAT catalyses the first committed as well as the rate-limiting
266 step in TAG and phospholipid biosynthesis⁽⁷⁴⁾.

267 Dietary variations are responsible for fluctuations in nutrient intake which can result in changes
268 in circulating glucose, which in turn signal the secretion of hormones. For example, ingestion
269 of a high-carbohydrate diet leads to a high circulating insulin level which consequently induces

270 enzymes involved in FA and TAG synthesis, thus providing FA for membrane phospholipid
271 biosynthesis in cancer cells. On the other hand, during a state of fasting or starvation, glucagon
272 level is elevated which suppresses activities of enzymes involved in FA and TAG biosynthesis
273 by increasing the intracellular cAMP level^(73, 74, 75). Moreover, *in vivo* studies have
274 demonstrated that high carbohydrate and low fat diets lead to higher rates of lipogenesis than
275 diets rich in fat and low in carbohydrates. The type of carbohydrate also affects lipogenesis
276 such that diets with fructose as the primary source of carbohydrate cause higher rates of FA
277 synthesis and higher activities of the lipogenic enzymes than diets containing equivalent
278 amounts of glucose⁽⁷⁶⁾.

279

280 **Endometrial cancer**

281 As mentioned previously, the cells in the endometrium undergo cyclical cellular changes
282 during the menstrual cycle. Hormones like estrogen have a mitogenic effect on the cells of the
283 endometrium^(77, 78). Excessive exposure to estrogen either exogenous or endogenous secondary
284 to high fat diet may cause increased proliferation of the endometrial cells. Cells proliferating
285 at a faster rate are more prone to errors during DNA replication and the mutated cells can
286 subsequently undergo malignant transformation, most commonly adenocarcinomas.

287 Endometrial cancer is a hormone-driven cancer, with approximately 80% of endometrial
288 cancers potentially arising due to either an excess of estrogen or a lack of progesterone. In the
289 normal endometrium, the proliferative effects of estrogen are normally countered by
290 progesterone, but the absence of progesterone allows estrogen to induce oncogenesis, an effect
291 that is amplified in situations of excess estrogen (Figure 1). One of the major emerging causes
292 of the estrogen/progesterone imbalance is obesity which is known to influence hormonal
293 balance and level of growth factors^(79, 80). Evidence shows a positive link between increased
294 dietary fat intake and obesity, thus associating fat intake to an increased risk of endometrial
295 cancer⁽⁸¹⁾. Central obesity, characterised by high abdominal fatness is commonly observed
296 among women during the menopausal years and is responsible for the increase in circulating
297 free fatty acids and consequently promotes an increase in insulin resistance⁽⁸²⁾. In addition,
298 long-term consumption of high glycaemic index diet is another risk factor for obesity and
299 insulin resistance and is also hypothesised to be involved in the pathogenesis of endometrial
300 cancer⁽⁸³⁾. Hyperinsulinaemia increases the risk of endometrial cancers mainly by the binding
301 of insulin to insulin receptors on endometrial cells to stimulate the growth of endometrial
302 stromal cells as well as through other pathways⁽⁷⁷⁾.

303

304 **Breast cancer**

305 The pathogenesis of breast cancer is intricate and multifactorial. The aetiology of breast cancer
306 could include mutation in the BRCA1 gene, a family history of breast cancer or mutagens
307 which can lead to DNA damage. It can also involve a similar hormonal pathogenesis as ovarian
308 cancer⁽⁸⁴⁾. Importantly, estrogen influences growth, differentiation and functioning of the
309 breast tissue (Figure 1). Aromatase, an enzyme found in the adipose tissues helps convert
310 circulating cholesterol to oestradiol⁽⁸⁵⁾. Due to the higher proportion of fat cells in breasts of
311 older women, their level of oestradiol in the breast tissues particularly post menopause is likely
312 to be higher than the plasma circulating level. The high oestradiol level in the breast tissues
313 can trigger differential effects on the estrogen receptor expression which are found in those
314 tissues, thus influencing the behaviour of cancer cells⁽⁸⁶⁾. Stromal cells in the breast tissues can
315 also support metastatic activity as they do not only control growth of normal breast epithelial
316 cells but also that of neoplastic epithelial cells by secreting growth factors in response to the
317 levels of endogenous hormones⁽⁸⁷⁾.

318 High cholesterol level, as a result of a high fat diet has also been stated as a risk factor for breast
319 cancer among women during the late peri-menopausal and post-menopausal state⁽⁸⁸⁾.
320 According to studies in mice^(89, 90), oxysterol 27-hydroxycholesterol (27-HC), a metabolite of
321 cholesterol synthesis has been identified in the pathogenesis of breast cancer. 27-HC could
322 stimulate the growth of breast cancer cell lines by binding and activating estrogen receptors
323 (ER) in a similar way as oestradiol. There is also evidence that postmenopausal women
324 experience an increase in their cholesterol level and thus its metabolite 27-HC which could
325 help explain the increase in breast cancer risk among obese and hypercholesterolaemic
326 women⁽⁹¹⁾. However, according to a recent EPIC-Heidelberg Cohort study publication
327 including 530 incident cases of breast cancer, a high level of 27-HC was associated with a
328 reduced risk of breast cancer among postmenopausal women and no association was found
329 among premenopausal women⁽⁹²⁾.

330 Moreover, a fat-rich diet is positively correlated with insulin resistance⁽⁹³⁾. Insulin resistance,
331 a major factor in the pathogenesis of premenopausal breast cancer, is also involved in the
332 aetiology of postmenopausal breast cancer. Insulin can bind to insulin receptors found on the
333 epithelial cells of the breast. This insulin signalling can contribute to cancer through mitogenic
334 activity mediated by the phosphatidylinositol-3 kinase and mitogen-activated protein
335 kinase/Akt signalling pathways⁽⁹⁴⁾. Insulin also has anti-apoptotic characteristics and thus
336 promotes tumour invasive activity. Insulin resistance is also accompanied by high levels of
337 proinflammatory cytokines and leptin as well as a decreased level of adiponectin which

338 concomitantly lead to both ER-positive and ER-negative breast cancer⁽⁹⁵⁾. Moreover, insulin
339 resistance is associated with an increased estrogen level as a result of enhanced aromatase
340 activity and decreased production of SHBG^(96, 97). This mechanistic pathway has been
341 supported by an Italian-nested case-control study which demonstrated that both pre- and
342 post-menopausal women with hyperglycaemia had an increased risk of breast cancer⁽⁹⁸⁾.

343

344 In addition to the high circulating level of estrogen as a result of obesity, the associated high
345 levels of inflammatory markers, insulin-like growth factors and adipokines from the visceral
346 fat also increases the risk of breast cancer among postmenopausal women⁽⁹⁹⁾. While high
347 circulating estrogen level among premenopausal women can be a risk factor for breast
348 cancer⁽¹⁰⁰⁾, some studies have demonstrated that obesity can be protective among
349 premenopausal women. Obesity can lead to irregular ovarian cycles and hence lower
350 circulating estrogen levels. As demonstrated by a meta-analysis of prospective studies, waist
351 circumference was associated with ER-positive and progesterone receptor positive breast
352 cancers in postmenopausal women while in premenopausal women waist circumference was
353 positively associated with ER-negative breast cancer⁽¹⁰¹⁾. This would suggest a lower
354 likelihood of a hormonal pathogenesis for breast cancer among premenopausal women.
355 Chronic inflammation, abnormally high levels of insulin-like growth factor (IGF) and insulin
356 resistance have been linked to premenopausal breast cancer⁽¹⁰²⁾.

357

358 **Other protective effect of diet against the risk of hormone-dependent cancers**

359 Vitamins like B₆, B₁₂ and folate are required for normal DNA repair mechanisms and proper
360 DNA replication. Folate receptor alpha expression is correlated with stage and grade of ovarian
361 cancer, suggesting this pathway may be relevant to ovarian carcinogenesis and progression⁽¹⁰³⁾.
362 Ascorbic acid, vitamin E and other trace elements like selenium having antioxidant properties
363 help to protect from free radical injury and maintain normal cellular function. Vitamin C is
364 recognised for its beneficial effect in cancer chemoprevention mainly as it has the potential to
365 stimulate immune function, impede nitrosamine formation, minimise DNA damage and block
366 the metabolic activation of carcinogens⁽¹⁰⁴⁾. Vitamin A helps to control epithelisation of tissues
367 and also has antioxidant properties to help protect from DNA damage⁽¹⁰⁵⁾. Although these
368 current theories support the plausible role of these micronutrients in hormone-dependent
369 cancer, prospective studies as well as a recent pooled analysis of cohort studies and meta-
370 analyses reported no association between dietary vitamins A, C or E and the risk of ovarian

371 and endometrial cancers^(104, 106, 107, 108). In addition, the WCRF/AICR⁽⁸²⁾ reported inconclusive
372 association between nutrients such as vitamin A, C, E as well as folate and the incidence of
373 ovarian, endometrial, breast cancers.

374 Moreover, a recent meta-analysis of cohort and case-control studies suggested that vitamin D
375 intake was protective against premenopausal breast cancer⁽¹⁰⁹⁾. A large cohort study including
376 68,567 postmenopausal women further demonstrated that women with a high intake of
377 calcium, and vitamin D had a reduced risk of postmenopausal breast cancer⁽¹¹⁰⁾. Experimental
378 studies have also suggested that vitamin D intake could reduce the stimulatory effect of
379 androgen in human ovarian cancer cell lines and also reduce obesity induced endometrial
380 cancer^(111, 112). However, systematic reviews concluded that the evidence to support the
381 association between vitamin D intake and endometrial and ovarian cancers are not consistent
382 and strong, thus calling for further prospective studies. One of the limitations was that since
383 most of the studies included in this systematic review were case-control studies, diet was thus
384 measured only at 1-time period and was very prone to misreporting due to recall bias, therefore
385 not accounting for diet change over time and vitamin D production through the skin^(113, 114).

386

387 Flavonoids, a group of heterogeneous polyphenols, have multiple health benefits. The main
388 sources of flavonoids include fruits, vegetables, tea, and wine⁽¹¹⁵⁾. Flavonoids reportedly have
389 several properties which contribute to the various health benefits including antioxidant, anti-
390 mutagenic, and anti-proliferative properties. Among them, isoflavones and some flavones,
391 flavanones, and flavanols also have estrogenic or anti-estrogenic activity, which makes these
392 compounds of particular interest for modulation of reproductive cancer risks⁽¹¹⁶⁾. According to
393 a large prospective cohort study including 171,940 US women, 723 of whom developed
394 ovarian cancer over a period of 16–22y of follow-up, demonstrated inverse associations
395 between flavonol and flavanone intakes and ovarian cancer risk⁽¹¹⁷⁾. Further supporting the
396 chemoprotective role of the flavonol in ovarian cancer risk, two *in vitro* studies demonstrated
397 that kaempferol induces apoptosis in ovarian cancer cells by regulating pro-apoptotic and anti-
398 apoptotic protein expressions and by preventing angiogenesis in ovarian cancer cells^(118, 119).
399 Furthermore, a meta-analysis of six cohort and six case-control studies demonstrated that
400 intakes of flavonols and flavones are protective against breast cancer, especially among
401 postmenopausal women⁽¹²⁰⁾, thus supporting the chemo-preventive role of fruits and vegetables
402 in hormone-related cancers.

403

404

405 **Recommendations and future research based on the current evidence**

406 In summary, evidence shows that diets predisposing to obesity and insulin resistance are the
407 main drivers of sex hormone fluctuations among both pre- and post-menopausal women.
408 Fluctuations in estrogen levels have been associated with the timing of the onset of natural
409 menopause, the presence of VMS, and longer-term sequelae such as ovarian, endometrial and
410 breast cancer. Studies have demonstrated that both the consumption of more balanced diets,
411 rich in fibre, fruits and vegetables (and, by contrast, those less healthy containing processed
412 meats and rich in fat) can alter circulating levels of estrogen and other sex hormones. Diet could
413 consequently influence the timing of natural menopause and hence affect its associated
414 sequelae. However, further evidence around the hypothesis that diet might influence timing of
415 menopause and presence of VMS are required in observational trials and use of metabolomics
416 may be valuable in revealing mechanistic pathways. Additional observational studies may also
417 clarify the association between diet and hormone-related cancers.

418

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426

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428 Y.D. drafted the manuscript. J.E.C and D.C.G critically revised the manuscript for important
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References

1. Hill K (1996) The demography of menopause. *Maturitas* **23**, 113-127.
2. Schoenaker D, Jackson CA, Rowlands JV *et al.* (2014) Socioeconomic position, lifestyle factors and age at natural menopause: a systematic review and meta-analyses of studies across six continents. *Int J Epidemiol* **43**, 1542-1562.
3. Davis SR, Lambrinoudaki I, Lumsden M *et al.* (2015) Menopause. *Nat Rev Dis Primers* **1**, 15004. doi: 10.1038/nrdp.2015.4.
4. Edmonds DK, Dewhurst JS (2006) *Dewhurst's textbook of obstetrics and gynaecology*. vol. 7th. Malden, Mass: Blackwell Pub.
5. Sherwood L (1993) *Human physiology: from cells to systems*. vol. 2nd. St. Paul: West Publishing Company.
6. O'Connor KA, Holman DJ, Wood JW (2001) Menstrual cycle variability and the perimenopause. *Am J Hum Bio* **13**, 465-478.
7. Morrison JH, Brinton RD, Schmidt PJ *et al.* (2006) Estrogen, menopause, and the aging brain: how basic neuroscience can inform hormone therapy in women. *J Neurosci* **26**, 10332-10348.
8. Harlow SD, Gass M, Hall JE *et al.* (2012) Executive summary of the Stages of Reproductive Aging Workshop + 10: addressing the unfinished agenda of staging reproductive aging. *J Clin Endocrinol Metab* **97**, 1159-1168.
9. Burger HG, Hale GE, Dennerstein L *et al.* (2008) Cycle and hormone changes during perimenopause: the key role of ovarian function. *Menopause* **15**, 603-612.
10. Su HI, Freeman EW (2009) Hormone changes associated with the menopausal transition. *Minerva Ginecol* **61**, 483-489.
11. Freeman EW, Sherif K (2007) Prevalence of hot flushes and night sweats around the world: a systematic review. *Climacteric* **10**, 197-214.
12. Gold EB (2011) The timing of the age at which natural menopause occurs. *Obstet Gynecol Clin North Am* **38**, 425-440.
13. Pines A (2011) Vasomotor symptoms and cardiovascular disease risk. *Climacteric* **14**, 535-536.

14. Kaczmarek M (2007) The timing of natural menopause in Poland and associated factors. *Maturitas* **57**, 139-153.
15. Kato I, Toniolo P, Akhmedkhanov A *et al.* (1998) Prospective Study of Factors Influencing the Onset of Natural Menopause. *J Clin Epidemiol* **51**, 1271-1276.
16. Lawlor DA, Ebrahim S, Smith GD (2003) The association of socio-economic position across the life course and age at menopause: the British Women's Heart and Health Study. *BJOG* **110**, 1078-1087.
17. Okeke TC, Anyaehie UB, Ezenyeaku CC (2013) Premature Menopause. *Ann Med Health Sci Res* **3**, 90-95.
18. Faubion SS, Kuhle CL, Shuster LT *et al.* (2015) Long-term health consequences of premature or early menopause and considerations for management. *Climacteric* **18**, 483-491.
19. Sherman S (2005) Defining the menopausal transition. *Am J Med* **118**, 3-7.
20. Monterrosa-Castro A, Blümel JE, Portela-Buelvas K *et al.* (2013) Type II diabetes mellitus and menopause: a multinational study. *Climacteric* **16**, 663-672.
21. Sekhar TVDS, Medarametla S, Rahman A *et al.* (2015) Early Menopause in Type 2 Diabetes – A Study from a South Indian Tertiary Care Centre. *J Clin Diagn Res* **9**, OC08-OC10.
22. Dunneram Y, Greenwood DC, Burley VJ *et al.* (2018) Dietary intake and age at natural menopause: results from the UK Women's Cohort Study. *J Epidemiol Community Health* **72**, 733-740.
23. Purdue-Smithe AC, Whitcomb BW, Szegda KL *et al.* (2017) Vitamin D and calcium intake and risk of early menopause. *Am J Clin Nutr* **105**, 1493-1501.
24. Parker J, Hashmi O, Dutton D *et al.* (2010) Levels of vitamin D and cardiometabolic disorders: systematic review and meta-analysis. *Maturitas* **65**, 225-236.
25. Biela U (2002) [Determinants of the age at natural menopause]. *Przegl Lek* **59**, 165-169.
26. Bagga D, Ashley JM, Geffrey SP *et al.* (1995) Effects of a very low fat, high fiber diet on serum hormones and menstrual function. Implications for breast cancer prevention. *Cancer* **76**, 2491-2496.

27. Karelis AD, Fex A, Filion ME *et al.* (2010) Comparison of sex hormonal and metabolic profiles between omnivores and vegetarians in pre- and post-menopausal women. *Br J Nutr* **104**, 222-226.
28. Rock CL, Flatt SW, Thomson CA *et al.* (2004) Effects of a High-Fiber, Low-Fat Diet Intervention on Serum Concentrations of Reproductive Steroid Hormones in Women With a History of Breast Cancer. *J Clin Oncol* **22**, 2379-2387.
29. Nagata C, Takatsuka N, Kawakami N *et al.* (2000) Association of diet with the onset of menopause in Japanese women. *Am J Epidemiol* **152**, 863-867.
30. Agarwal A, Gupta S, Sharma RK (2005) Role of oxidative stress in female reproduction. *Reprod Biol Endocrino* **3**, 28-28.
31. Ruder EH, Hartman TJ, Blumberg J *et al.* (2008) Oxidative stress and antioxidants: exposure and impact on female fertility. *Hum Reprod Update* **14**, 345-357.
32. Ozcan P, Ficicioglu C, Yildirim OK *et al.* (2015) Protective effect of resveratrol against oxidative damage to ovarian reserve in female Sprague-Dawley rats. *Reprod Biomed Online* **31**, 404-410.
33. Berg J, Tymoczko J, Stryer L (2002) Important Derivatives of Cholesterol Include Bile Salts and Steroid Hormones. In *Biochemistry*, 5th ed. New York: W H Freeman.
34. Zsakai A, Karkus Z, Utczas K *et al.* (2016) Body fatness and endogenous sex hormones in the menopausal transition. *Maturitas* **87**, 18-26.
35. Ho SC, Wu S, Chan SG *et al.* (2010) Menopausal transition and changes of body composition: a prospective study in Chinese perimenopausal women. *Int J Obes (Lond)* **34**, 1265-1274.
36. Torgerson DJ, Avenell A, Russell IT *et al.* (1994) Factors associated with onset of menopause in women aged 45–49. *Maturitas* **19**, 83-92.
37. Torgerson DJ, Thomas RE, Campbell MK *et al.* (1997) Alcohol consumption and age of maternal menopause are associated with menopause onset. *Maturitas* **26**, 21-25.
38. Nagata C, Takatsuka N, Inaba S *et al.* (1998) Association of diet and other lifestyle with onset of menopause in Japanese women. *Maturitas* **29**, 105-113.
39. Nagel G, Altenburg HP, Nieters A *et al.* (2005) Reproductive and dietary determinants of the age at menopause in EPIC-Heidelberg. *Maturitas* **52**, 337-347.

40. Martin LJ, Greenberg CV, Kriukov V *et al.* (2006) Intervention with a low-fat, high-carbohydrate diet does not influence the timing of menopause. *Am J Clin Nutr* **84**, 920-928.
41. Dorjgochoo T, Kallianpur A, Gao YT *et al.* (2008) Dietary and lifestyle predictors of age at natural menopause and reproductive span in the Shanghai Women's Health Study. *Menopause* **15**, 924-933.
42. Nagata C, Wada K, Nakamura K *et al.* (2012) Associations of physical activity and diet with the onset of menopause in Japanese women. *Menopause* **19**, 75-81.
43. Carwile JL, Willett WC, Michels KB (2013) Consumption of Low-Fat Dairy Products May Delay Natural Menopause. *J Nutr* **143**, 1642-1650.
44. Boutot ME, Purdue-Smithe A, Whitcomb BW *et al.* (2018) Dietary Protein Intake and Early Menopause in the Nurses' Health Study II. *Am J Epidemiol* **187**, 270-277.
45. Wang M, Gong WW, Hu RY *et al.* (2018) Age at natural menopause and associated factors in adult women: Findings from the China Kadoorie Biobank study in Zhejiang rural area. *PLoS One* **13**, e0195658.
46. Purdue-Smithe AC, Whitcomb BW, Manson JE *et al.* (2018) A Prospective Study of Dairy Food Intake and Early Menopause. *Am J Epidemiol*, doi: 10.1093/aje/kwy212.
47. O'Neill S, Eden J (2017) The pathophysiology of menopausal symptoms. *Obstet Gynaecol Reprod Med* **27**, 303-310.
48. Dhanoya T, Sievert LL, Muttukrishna S *et al.* (2016) Hot flushes and reproductive hormone levels during the menopausal transition. *Maturitas* **89**, 43-51.
49. Collaborative Group on Hormonal Factors in Breast Cancer (2012) Menarche, menopause, and breast cancer risk: individual participant meta-analysis, including 118 964 women with breast cancer from 117 epidemiological studies. *The Lancet Oncology* **13**, 1141-1151.
50. Tavani A, Ricci E, La Vecchia C *et al.* (2000) Influence of menstrual and reproductive factors on ovarian cancer risk in women with and without family history of breast or ovarian cancer. *Int J Epidemiol* **29**, 799-802.
51. Xu WH, Xiang YB, Ruan ZX *et al.* (2004) Menstrual and reproductive factors and endometrial cancer risk: Results from a population-based case-control study in urban Shanghai. *Int J Cancer* **108**, 613-619.

52. Travis RC, Key TJ (2003) Oestrogen exposure and breast cancer risk. *Breast Cancer Res* **5**, 239-247.
53. Clemons M, Goss P (2001) Estrogen and the risk of breast cancer. *N Engl J Med* **344**, 276-285.
54. Mungenast F, Thalhammer T (2014) Estrogen biosynthesis and action in ovarian cancer. *Front Endocrinol (Lausanne)* **5**, 192.
55. Wu AH, Pike MC, Stram DO (1999) Meta-analysis: Dietary Fat Intake, Serum Estrogen Levels, and the Risk of Breast Cancer. *J Natl Cancer Ins* **91**, 529-534.
56. Nagata C, Nagao Y, Shibuya C *et al.* (2005) Fat Intake Is Associated with Serum Estrogen and Androgen Concentrations in Postmenopausal Japanese Women. *J Nutr* **135**, 2862-2865.
57. Gaskins AJ, Mumford SL, Zhang C *et al.* (2009) Effect of daily fiber intake on reproductive function: the BioCycle Study. *Am J Clin Nutr* **90**, 1061-1069.
58. Lethaby A, Marjoribanks J, Kronenberg F *et al.* (2013) Phytoestrogens for menopausal vasomotor symptoms. *The Cochrane database of systematic reviews*, Cd001395.
59. Schmidt M, Arjomand-Wolkart K, Birkhauser MH *et al.* (2016) Consensus: soy isoflavones as a first-line approach to the treatment of menopausal vasomotor complaints. *Cochrane Database Syst Rev* **32**, 427-430.
60. Herber-Gast GC, Mishra GD (2013) Fruit, Mediterranean-style, and high-fat and -sugar diets are associated with the risk of night sweats and hot flushes in midlife: results from a prospective cohort study. *Am J Clin Nutr* **97**, 1092-1099.
61. Begum S, S A (2012) Study of immune profile during different phases of menstrual cycle. *IJBMR* **3**, 1407-1409.
62. Nair AR, Taylor HS (2010) The Mechanism of Menstruation. In *Amenorrhea: A Case-Based, Clinical Guide*, pp. 21-34 [NF Santoro and G Neal-Perry, editors]. New York: Humana Press.
63. Beshay VE, Carr BR (2013) Hypothalamic-Pituitary-Ovarian Axis and Control of the Menstrual Cycle. In *Clinical Reproductive Medicine and Surgery: A Practical Guide*, 2 ed., pp. 31-42 [T Falcone and WW Hurd, editors]. New York: Springer.

64. Lengyel E (2010) Ovarian cancer development and metastasis. *Am J Pathol* **177**, 1053-1064.
65. Fucic A, Gamulin M, Ferencic Z *et al.* (2012) Environmental exposure to xenoestrogens and oestrogen related cancers: Reproductive system, breast, lung, kidney, pancreas, and brain. *Environ Health* **11**, S8-S8.
66. Ogura K, Irahara M, Kiyokawa M *et al.* (2001) Effects of leptin on secretion of LH and FSH from primary cultured female rat pituitary cells. *Eur J Endocrinol* **144**, 653-658.
67. Azrad M, Turgeon C, Demark-Wahnefried W (2013) Current evidence linking polyunsaturated Fatty acids with cancer risk and progression. *Front Oncol* **3**, 224. doi: 10.3389/fonc.2013.00224.
68. Larsson SC, Kumlin M, Ingelman-Sundberg M *et al.* (2004) Dietary long-chain n-3 fatty acids for the prevention of cancer: a review of potential mechanisms. *Am J Clin Nutr* **79**, 935-945.
69. Calder PC (2013) Omega-3 polyunsaturated fatty acids and inflammatory processes: nutrition or pharmacology? *Br J Clin Pharmacol* **75**, 645-662.
70. Merritt MA, Cramer DW, Missmer SA *et al.* (2014) Dietary fat intake and risk of epithelial ovarian cancer by tumour histology. *Br J Cancer* **110**, 1392-1401.
71. Hu F, Zhang Y, Song Y (2013) *Lipid metabolism, metabolic syndrome, and cancer*: INTECH Open Access Publisher.
72. Berquin IM, Min Y, Wu R *et al.* (2007) Modulation of prostate cancer genetic risk by omega-3 and omega-6 fatty acids. *J Clin Invest* **117**, 1866-1875.
73. Wong RHF, Sul HS (2010) Insulin signaling in fatty acid and fat synthesis: a transcriptional perspective. *Curr Opin Pharmacol* **10**, 684-691.
74. Sul HS, Wang D (1998) Nutritional and hormonal regulation of enzymes in fat synthesis: Studies of fatty acid synthase and mitochondrial glycerol-3-phosphate acyltransferase gene transcription. *Annu Rev Nutr* **18**, 331-351.
75. Currie E, Schulze A, Zechner R *et al.* (2013) Cellular Fatty Acid Metabolism and Cancer. *Cell Metab* **18**, 153-161.

76. Hillgartner F, Salati LM, Goodridge AG (1995) Physiological And Molecular Mechanisms Involved In Nutritional Regulation Of Fatty-Acid Synthesis. *Physiol Rev* **75**, 47-76.
77. Kaaks R, Lukanova A, Kurzer MS (2002) Obesity, endogenous hormones, and endometrial cancer risk: A synthetic review. *Cancer Epidemiol Biomarkers Prev* **11**, 1531-1543.
78. Losordo DW, Isner JM (2001) Estrogen and Angiogenesis: A Review. *Arterioscler Thromb Vasc Biol* **21**, 6-12.
79. Xu WH, Matthews CE, Xiang YB *et al.* (2005) Effect of adiposity and fat distribution on endometrial cancer risk in Shanghai women. *Am J Epidemiol* **161**, 939-947.
80. Carlson MJ, Thiel KW, Yang S *et al.* (2012) Catch it before it kills: progesterone, obesity, and the prevention of endometrial cancer. *Discov Med* **14**, 215-222.
81. Goodman MT, Wilkens LR, Hankin JH *et al.* (1997) Association of soy and fiber consumption with the risk of endometrial cancer. *Am J Epidemiol* **146**, 294-306.
82. World Cancer Research Fund/American Institute for Cancer Research (2007) *Food, Nutrition, Physical Activity, and the Prevention of Cancer: a Global Perspective*. Available at dietandcancerreport.org
83. Mulholland HG, Murray LJ, Cardwell CR *et al.* (2008) Dietary glycaemic index, glycaemic load and endometrial and ovarian cancer risk: a systematic review and meta-analysis. *Br J Cancer* **99**, 434-441.
84. Yoneda A, Lendorf ME, Couchman JR *et al.* (2012) Breast and Ovarian Cancers: A Survey and Possible Roles for the Cell Surface Heparan Sulfate Proteoglycans. *J Histochem Cytochem* **60**, 9-21.
85. Cleary MP, Grossmann ME (2009) Obesity and Breast Cancer: The Estrogen Connection. *Endocrinology* **150**, 2537-2542.
86. Malara NM, Leotta A, Sidoti A *et al.* (2006) Ageing, hormonal behaviour and cyclin D1 in ductal breast carcinomas. *Breast* **15**, 81-89.
87. Bussard KM, Mutkus L, Stumpf K *et al.* (2016) Tumor-associated stromal cells as key contributors to the tumor microenvironment. *Breast Cancer Res* **18**, 84.

88. Llaverias G, Danilo C, Mercier I *et al.* (2011) Role of Cholesterol in the Development and Progression of Breast Cancer. *Am J Pathol* **178**, 402-412.
89. Wu Q, Ishikawa T, Sirianni R *et al.* (2013) 27-Hydroxycholesterol Promotes Cell-autonomous ER-positive Breast Cancer Growth. *Cell Rep* **5**, 637-645.
90. Baek AE, Yu Y-RA, He S *et al.* (2017) The cholesterol metabolite 27 hydroxycholesterol facilitates breast cancer metastasis through its actions on immune cells. *Nat Commun* **8**, 864.
91. Derby CA, Crawford SL, Pasternak RC *et al.* (2009) Lipid Changes During the Menopause Transition in Relation to Age and Weight The Study of Women's Health Across the Nation. *Am J Epidemiol* **169**, 1352-1361.
92. Lu D-L, Le Cornet C, Sookthai D *et al.* (2018) Circulating 27-Hydroxycholesterol and Breast Cancer Risk: Results From the EPIC-Heidelberg Cohort. *J Natl Cancer Inst* doi: 10.1093/jnci/djy115.
93. De Souza CT, Araujo EP, Bordin S *et al.* (2005) Consumption of a fat-rich diet activates a proinflammatory response and induces insulin resistance in the hypothalamus. *Endocrinology* **146**, 4192-4199.
94. Orgel E, Mittelman SD (2013) The Links Between Insulin Resistance, Diabetes, and Cancer. *Curr Diab Rep* **13**, 213-222.
95. Rose DP, Gracheck PJ, Vona-Davis L (2015) The Interactions of Obesity, Inflammation and Insulin Resistance in Breast Cancer. *Cancers* **7**, 2147-2168.
96. Mukhopadhyay KD, Liu Z, Bandyopadhyay A *et al.* (2015) Aromatase Expression Increases the Survival and Malignancy of Estrogen Receptor Positive Breast Cancer Cells. *PLoS One* **10**, e0121136.
97. Daka B, Rosen T, Jansson PA *et al.* (2013) Inverse association between serum insulin and sex hormone-binding globulin in a population survey in Sweden. *Endocr Connect* **2**, 18-22.
98. Sieri S, Muti P, Claudia A *et al.* (2012) Prospective study on the role of glucose metabolism in breast cancer occurrence. *Int J Cancer* **130**, 921-929.
99. Howe LR, Subbaramaiah K, Hudis CA *et al.* (2013) Molecular pathways: adipose inflammation as a mediator of obesity-associated cancer. *Clin Cancer Res* **19**, 6074-6083.

100. Key TJ, Appleby PN, Reeves GK *et al.* (2013) Sex hormones and risk of breast cancer in premenopausal women: a collaborative reanalysis of individual participant data from seven prospective studies. *Lancet Oncol* **14**, 1009-1019.
101. Chen G-C, Chen S-J, Zhang R *et al.* (2016) Central obesity and risks of pre- and postmenopausal breast cancer: a dose–response meta-analysis of prospective studies. *Obes Rev* **17**, 1167-1177.
102. Luque RM, López-Sánchez LM, Villa-Osaba A *et al.* (2017) Breast cancer is associated to impaired glucose/insulin homeostasis in premenopausal obese/overweight patients. *Oncotarget* **8**, 81462-81474.
103. Harris HR, Cramer DW, Vitonis AF *et al.* (2012) Folate, vitamin B 6, vitamin B 12, methionine and alcohol intake in relation to ovarian cancer risk. *Int J Cancer* **131**, E518-E529.
104. Bandera EV, Gifkins DM, Moore DF *et al.* (2009) Antioxidant Vitamins and the Risk of Endometrial Cancer: A Dose–Response Meta-Analysis. *Cancer Causes Control* **20**, 699-711.
105. Donaldson MS (2004) Nutrition and cancer: a review of the evidence for an anti-cancer diet. *Nutr J* **3**, 19.
106. Navarro Silvera SA, Jain M, Howe GR *et al.* (2006) Carotenoid, vitamin A, vitamin C, and vitamin E intake and risk of ovarian cancer: A prospective cohort study. *Cancer Epidemiol Biomarkers Prev* **15**, 395-397.
107. Thomson CA, Neuhouser ML, Shikany JM *et al.* (2008) The Role of Antioxidants and Vitamin A in Ovarian Cancer: Results From the Women's Health Initiative. *Nutr Cancer* **60**, 710-719.
108. Koushik A, Wang M, Anderson KE *et al.* (2015) Intake of vitamins A, C, and E and folate and the risk of ovarian cancer in a pooled analysis of 10 cohort studies. *Cancer Causes Control* **26**, 1315-1327.
109. Estébanez N, Gómez-Acebo I, Palazuelos C *et al.* (2018) Vitamin D exposure and Risk of Breast Cancer: a meta-analysis. *Sci Rep* **8**, 9039.
110. McCullough ML, Rodriguez C, Diver WR *et al.* (2005) Dairy, Calcium, and Vitamin D Intake and Postmenopausal Breast Cancer Risk in the Cancer Prevention Study II Nutrition Cohort. *Cancer Epidemiol Biomarkers Prev* **14**, 2898-2904.

111. Ahonen MH, Zhuang YH, Aine R *et al.* (2000) Androgen receptor and vitamin D receptor in human ovarian cancer: Growth stimulation and inhibition by ligands. *Int J Cancer* **86**, 40-46.
112. Yu W, Cline M, Maxwell LG *et al.* (2010) Dietary vitamin D exposure prevents obesity-induced increase in endometrial cancer in Pten^{+/-} mice. *Cancer Prev Res (Phila)* **3**, 1246-1258.
113. Cook LS (2010) A systematic literature review of vitamin D and ovarian cancer. *Am J Obstet Gynecol* **203**, 70.e71-70.e78.
114. McCullough ML, Bandera EV, Moore DF *et al.* (2008) Vitamin D and calcium intake in relation to risk of endometrial cancer: A systematic review of the literature. *Prev Med* **46**, 298-302.
115. Romagnolo DF, Selmin OI (2012) Flavonoids and Cancer Prevention: A Review of the Evidence. *J Nutr Gerontol Geriatr* **31**, 206.
116. Rossi M, La Vecchia C (2014) Flavonoids and the risk of ovarian cancer. *Am J Clin Nutr* **100**, 1217-1219.
117. Cassidy A, Huang TY, Rice MS *et al.* (2014) Intake of dietary flavonoids and risk of epithelial ovarian cancer. *Am J Clin Nutr* **100**, 1344-1351.
118. Luo H, Rankin GO, Li Z *et al.* (2011) Kaempferol induces apoptosis in ovarian cancer cells through activating p53 in the intrinsic pathway. *Food Chem* **128**, 513-519.
119. Luo H, Rankin GO, Juliano N *et al.* (2012) Kaempferol inhibits VEGF expression and in vitro angiogenesis through a novel ERK-NF kappa B-cMyc-p21 pathway. *Food Chem* **130**, 321-328.
120. Hui C, Qi X, Qianyong Z *et al.* (2013) Flavonoids, Flavonoid Subclasses and Breast Cancer Risk: A Meta-Analysis of Epidemiologic Studies. *PLoS ONE* **8**, e54318. doi: 10.1371/journal.pone.0054318.

Table 1. Evidence for the associations between diet and onset of menopause

Author, year	Study Design, sample size	Intervention/exposure	Findings	
			Early	Late
Torgerson et al., 1994 ⁽³⁶⁾	Cross-sectional, 2,074	Meat, alcohol	-	Meat Alcohol
Torgerson et al., 1997 ⁽³⁷⁾	Prospective, 1,227	Meat, alcohol	-	Alcohol
Nagata et al., 1998 ⁽³⁸⁾	Cross-sectional, 3,704	Total energy; macronutrients; cholesterol; calcium; crude fibre; vitamins A, C, D, and E; carotene; soy product; retinol; coffee; alcohol	Soy products Coffee	Fat Cholesterol
Nagata et al., 2000 ⁽²⁹⁾	Prospective, 1,130	Energy, macronutrients, animal protein/fat, vegetable protein/fat, fat from fish, cholesterol, calcium, crude fibre, vitamin A, retinol, vitamin C, vitamin E, green and yellow vegetables, other vegetables, soy products	-	Green & yellow vegetable
Nagel et al., 2005 ⁽³⁹⁾	Prospective, 5,568	Macronutrients, alcohol, meat, dairy products, fish, vegetables, fruit, cereal products, fibre, soy products, sweets, added animal fat, added vegetable fat	Carbohydrate Vegetable Fibre Cereal products	Total fat Protein Meat
Martin et al., 2006 ⁽⁴⁰⁾	Randomised clinical trial, 2,611	Low-fat high-carbohydrate diet	-	-
Dorjgochoo et al., 2008 ⁽⁴¹⁾	Prospective, 33,054	Energy, macronutrients, vegetables, fruit, red meat, saturated fat, total soy, total fibre, tea, alcohol	-	Energy Fruits Protein Carbohydrate
Nagata et al., 2012 ⁽⁴²⁾	Prospective, 3,115	Energy, total fat, SFAs, PUFAs, MUFAs, long omega-3 FAs, dietary fibre, soy isoflavones, alcohol	Polyunsaturated fat	-
Carwile et al., 2013 ⁽⁴³⁾	Prospective, 46,059	High-fat dairy, total low-fat dairy, skim milk, whole milk, dairy fat, dairy protein, calcium, vitamin D, lactose	-	Low fat dairy Skim milk Vitamin D from dairy sources
Purdue-Smithe et al., 2017 ⁽²³⁾	Prospective, 116,430	Vitamin D, calcium intake from dairy and non-dairy sources		Calcium from dairy sources

Boutot et al., 2017 ⁽⁴⁴⁾	Prospective, 85,682	Vegetable protein, animal protein, total protein, all meat, red meat, processed meat, chicken/turkey, seafood, eggs, soy/tofu, beans/lentils, peanuts, peas/lima beans, other nuts, peanut butter, pasta, dark bread, cold cereal	-	Vegetable protein Pasta Dark bread Cold cereal
Wang et al., 2018 ⁽⁴⁵⁾	Cross-sectional, 17,076	Meat, seafood, fresh eggs, soybean products, fresh fruits, dairy products, vitamins, minerals	Seafood Fresh eggs Fresh fruits Vitamins	Meat
Dunneram et al., 2018 ⁽²²⁾	Prospective, 35,375	Wholegrain products, refined grain products, low-fibre breakfast cereals, high-fibre breakfast cereals, plain potatoes, potatoes with added fat, refined pasta and rice, wholegrain pasta and rice, low-fat dairy products, high-fat dairy products, butter and hard margarine, margarine, low-fat spreads, high-fat dressing, low-fat dressing, soya bean products, textured vegetable protein, pulses, eggs/egg dishes, fish & fish dishes, oily fish, shellfish, red meat, processed meat, poultry, offal, vegetables, fruits, dried fruits, other foods groups, tea, coffee, soft drinks, wines, spirits, beer and cider, port/sherry/liqueurs	Refined pasta and rice	Oily fish Fresh legumes Vitamin B6 Zinc
Purdue-Smithe et al., 2018 ⁽⁴⁶⁾	Prospective, 116,429	Low-fat dairy foods, high-fat dairy foods, total dairy	-	Total dairy Low-fat dairy foods

Figure 1: Potential mechanistic pathways through which diet can influence circulating estrogen levels and women's reproductive health