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Drug Discovery for Kinetoplastid Diseases: Future Directions

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ABSTRACT: Kinetoplastid parasites have caused human disease for millennia. Significant progress has been made toward 24 developing new treatments for leishmaniasis (particularly on the Indian subcontinent) and for human African trypanosomiasis 25 (HAT) in Africa. The sustained decrease in the incidence of HAT has made the prospect of elimination a tantalizing reality. 26 Despite the gains, no new chemical or biological entities to treat kinetoplastid diseases have been registered in more than three 27 decades, and more work is needed to discover safe and effective therapies for patients with Chagas disease and leishmaniasis. 28 Advances in tools for drug discovery and novel insights into the biology of the host-parasite interaction may provide 29

- opportunities for accelerated progress. Here, we summarize the output from a gathering of scientists and physicians who met to 30
- discuss the current status and future directions in drug discovery for kinetoplastid diseases. 31

T early a billion people are at risk from the group of vector-32 borne kinetoplastid diseases composed of Chagas 33 34 disease, leishmaniasis, and human African trypanosomiasis 35 (HAT, also known as sleeping sickness). These ancient 36 parasitic illnesses have burdened humans for thousands of 37 years, as evidenced by Trypanosoma DNA sequences found in 38 South American mummies.¹ In the current era, kinetoplastid 39 diseases cause an estimated 30 000 deaths annually and induce 40 crippling morbidities in millions more.

There is reason to be optimistic about trends concerning 41 42 HAT. Public and private partners have jointly tackled the 43 disease in recent decades, with the World Health Organization 44 (WHO) coordinating public health activities and the Drugs for 45 Neglected Diseases Initiative (DNDi) directing global efforts 46 for new therapies. The introduction of nifurtimox-eflornithine 47 therapy in 2009 was a pivotal milestone, which was followed

recently by the demonstrated efficacy of oral fexinidazole² for 48 late-stage disease. (Approval for use is now pending assessment 49 by medicine regulatory agencies.) Fewer than 1500 new cases 50 were reported to WHO in 2017, making disease elimination a 51 tangible goal. 52

Successes in combating HAT are encouraging and contrast 53 with slower progress in containing other kinetoplastid diseases. 54 Most of the current kinetoplastid drugs are repurposed and are 55 often not potent enough to render a sterile cure (i.e., to 56 eliminate all parasites). Safe, effective, short-course practical 57 therapies are urgently needed for Chagas disease and 58 leishmaniasis yet remain elusive. 59

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Figure 1. Clinical manifestations of kinetoplastid diseases and opportunities for intervention.

To understand the present-day challenges and opportunities of related to new medicines for Chagas disease, visceral leishmaniasis (VL), and cutaneous leishmaniasis (CL), the Novartis Institute for Tropical Diseases convened a multidisciplinary group of scientific and medical specialists in parasitology, immunology, and drug discovery in June 2018. The scope of the discussion encompassed unmet medical reeds, the global pipeline of preclinical drug candidates, parasite biology, assays, models, and the potential use of novel immunomodulatory and adjunct therapies to target disease sequelae. This Viewpoint summarizes key workshop information.

72 CURRENT DRUGS: OLD, TOXIC, AND OFTEN 73 INEFFECTIVE

74 Chagas disease, which is endemic in the Americas, is caused by 75 Trypanosoma cruzi. Inoculation typically occurs through 76 infected feces from the triatomine bug, which is scratched or 77 rubbed into the skin or mucosa. Transmission also takes place 78 through blood transfusion and congenital and oral routes. The 79 pathogenesis of Chagas disease is not fully understood, and it is 80 currently impossible to predict which fraction of the patient 81 population (approximately 30%) will develop the serious 82 cardiac or gastrointestinal sequelae that appear years after the 83 initial infection (Figure 1). Sudden death from chronic Chagas 84 cardiomyopathy is an all too common outcome. Two drugs, 85 nitroheterocyclic agents benznidazole and nifurtimox, are used 86 for the treatment of Chagas disease; both were developed 87 decades ago, are contraindicated during pregnancy, and can 88 have serious adverse effects that substantially restrict their use. 89 Benznidazole is the better-tolerated option in adults, although 90 15-30% patients are unable to finish the standard 60-day 91 course, mainly because of skin and nervous system 92 complications. In children, nifurtimox is better tolerated than 93 benznidazole.

In both acute and chronic T. cruzi infection, treatment 94 reduces the parasite load and can yield clearance from blood 95 using available assays (e.g., PCR). Even so, in some cases, 96 parasites presumably may persist intracellularly, and it is 97 unclear in adults how the reductions in the parasite load 98 modulate the severity of chronic disease in the absence of 99 complete parasite clearance. Current drugs are inadequate 100 because they fail too often and are not dependable. However, 101 WHO recommends off-label use of benznidazole for the 102 treatment of chronically infected patients, even though its 103 efficacy in later stages of the disease is debatable. A large study 104 of patients with chronic Chagas cardiomyopathy (the 105 BENEFIT trial) demonstrated that benznidazole treatment 106 reduced the parasite burden but did not significantly reduce 107 disease progression. However, most had New York Heart 108 Association class I (largely asymptomatic) heart disease, which 109 may have confounded the findings.³ Some nonrandomized, 110 unblinded studies using benznidazole in indeterminate patients 111 without heart failure showed reduced disease progression, 112 emphasizing the need for controlled randomized studies for 113 indeterminate patients.4 114

Similarly, the approved therapeutic arsenal for leishmaniasis 115 has important limitations. Leishmaniasis is distributed across 116 the tropics and subtropics, though a majority of VL cases are 117 reported in only seven countries: Brazil, Ethiopia, Kenya, 118 Somalia, Sudan, South Sudan, and India. Patients succumb to 119 VL gradually over a period of 2 years. Anorexia and 120 pancytopenia give rise to wasting and increased susceptibility 121 to bacterial superinfections. VL is fatal without treatment, and 122 even those who undergo therapy remain at risk of a disfiguring 123 dermal form of relapsing disease called postkala-azar dermal 124 leishmaniasis (PKDL) that may also contribute to continued 125 disease transmission (Figure 1). CL has a higher global burden 126 than VL, with the greatest prevalence in Africa, the 127 Mediterranean, and South America. It does not cause systemic 128 129 morbidities or death but can result in grievous disfiguration 130 and stigma. Drugs targeting *Leishmania* parasites have generally 131 been repurposed from other indications. Antimonials, 132 amphotericin B, paromomycin sulfate, and miltefosine have 133 variable efficacy against the more than 20 *Leishmania* species 134 that cause disease. While WHO-recommended treatment 135 regimens for VL on the Indian subcontinent include liposomal 136 amphotericin B or oral miltefosine, these medicines are poorly 137 effective in patients in other global regions. Treatment courses 138 are generally long, require hospitalization, and have significant 139 toxicities that mandate frequent monitoring. Differences in the 140 treatment protocol by region, high costs, and low availability of 141 some drugs understandably stretch the limits of under-142 resourced health systems in countries where these diseases 143 are endemic.

144THE ANTIPARASITIC PIPELINE IS FILLING, BUT145THERE ARE GAPS

146 Antiparasitics are the cornerstone of therapy for kinetoplastid 147 diseases. It is well accepted that the clinical event cascades in 148 Chagas disease and leishmaniasis are induced by the presence 149 of parasites, and evidence suggests that eliminating parasites as 150 early as possible after infection could mitigate severe disease. 151 Unfortunately, the current preclinical pipeline for Chagas 152 disease treatments is meager. Only three classes of compounds 153 have been shown to achieve high cure rates in stringent mouse 154 models of infection: nitroimidazoles (e.g., fexinidazole, 155 currently in phase II), oxaboroles (e.g., DNDi-6148, active 156 against both leishmaniasis and Chagas), and proteasome 157 inhibitors (e.g., GNF6702⁵). The future is brighter in drug 158 discovery for leishmaniasis, where there are at least six 159 candidates in preclinical or clinical phases that have five 160 distinct mechanisms of action.⁶

Proposed target product profiles for new drugs are listed in Box 1. For Chagas disease, medicines should achieve cures that reason beyond the acute stage must be simple to administer and safe because patients in the indeterminate phase typically reason beyond the acute stage must be simple to administer response to a safe because patients in the indeterminate phase typically response to a safe because a relapse-free cure with no or minor response a verse a relapse-free cure with no or minor response to that of current drugs and improved safety response to that of current drugs and improved safety response to the step forward.

Whether a sterile cure (i.e., the elimination of all parasites) is row essential is a topic of debate. Some parasitologists advocate row strongly that a sterile cure must be achieved in Chagas disease row the reproliferation of parasites and enduring row pathogenicity. A sterile cure may not be critical for VL and row L. Reducing the parasite load in these infections could be row sufficient if the host immune system can complete the job of row parasite control or clearance. A sterile cure is more likely needed for PKDL (which appears to result from latent parasites) and for VL in individuals with HIV coinfection or row the immunodeficiency syndromes. A condition known as leishmaniasis recidivans in CL may also result from the recrudescence of latent parasites that survive therapy.

185 NEW INSIGHTS INTO DISEASE BIOLOGY WILL 186 INDICATE THE NEED FOR NEW MODELS

187 Given how little we know about the biology of *T. cruzi* and 188 *Leishmania* species and the lack of validated drug targets, it is

Box 1. Proposed Target Product Profile (TPP) for Chagas Disease and Leishmaniasis

Proposed TPP for Chagas

Eliminates all parasites, including in blood and tissue Active against all distinct typing units (DTUs)

Oral, safe, and well tolerated for use at all ages and during pregnancy and lactation with no monitoring required

Simple treatment regimen, amenable for use in a setting of weak health systems/infrastructure, accessible and affordable

Potency and safety not affected by pharmacogenomic factors

Can be used repeatedly (e.g., in the case of reinfections)

No significant drug-drug interaction

Low probability of resistance

Shelf life >2 years under tropical conditions

Proposed TPP for Leishmaniasis

Effective against all VL and CL parasites from varying geographic regions

Potency and safety not affected by pharmacogenomic factors

Potency/efficacy, >95% parasite clearance for VL, 99.9% parasite clearance from periphery, 99% from seclusion sites for CL

Short treatment regimen (as short as 1 week for both VL and CL, 14 day maximum for VL, 21 day maximum for CL)

Amenable for use in a setting of weak health systems/ infrastructure, accessible and affordable treatment regimen

Oral, safe, and well tolerated for use at all ages and during

pregnancy with no monitoring required Effective in immune-deficient individuals (e.g., HIV-VL)

and against PKDL

Avoids risk of resistance

not surprising that most current pipeline compounds 189 originated from phenotypic screens. Assays are available to 190 test the growth inhibition of amastigotes for *T. cruzi* 191 (intracellular) and *Leishmania* (intracellular and extracellular), 192 and these are compatible with high-throughput screening. It 193 may be important to evaluate the antiparasitic effect of 194 compounds by using intracellular parasites grown in diseaserelevant tissues. Cidality, time-to-kill kinetics, and washout 196 assays may be used to further enhance the confidence of hits 197 and to assist prioritization.

Highly sensitive in vivo imaging with bioluminescent T. cruzi 199 that enables the monitoring of the mouse parasite burden in 200 real time has highlighted how the parasite load varies by tissue 201 type over time. Furthermore, this model has predictive power. 202 It demonstrated the limited efficacy of posaconazole, a Chagas 203 disease drug candidate that had previously shown potency in 204 animal models but has failed to consistently eliminate 205 parasitemia in patients. By comparison, benznidazole was 206 shown to be efficacious in both mice and humans.⁸ Similarly, 207 novel murine and hamster models for VL and CL using 208 bioluminescent parasites have improved our understanding of 209 disease progression. New chemical entities should be tested in 210 mouse models with specific questions in mind, such as how the 211 treatment duration and curative exposures could translate from 212 mice to humans. 213

An important unknown for Chagas disease is the role played 214 by amastigotes that spontaneously adopt a "persister" 215 phenotype. These nonreplicative and phenotypically drug- 216 resistant forms of the parasite are later able to differentiate to 217 218 trypomastigotes and reinfect new host cells.⁸ Future work is 219 needed to understand how the development of persistent 220 forms is triggered, if they are metabolically active, whether they 221 can be forced out of dormancy, and what their role is in disease 222 progression. In the meantime, screening against persistent 223 parasites to find novel inhibitors would be beneficial.⁹

Persistence in *Leishmania* may also be a concern. Persistent L. mexicana and L. major parasites have been reported in mouse models,^{10,11} although similar forms have not yet been rough in animals for L. donovani or L. infantum. However, nonreplicating L. donovani were identified in a macrophage model, and these could represent persister-type cells, the existence of which is implicated through the recrudescence that can occur following VL treatment being manifest as PKDL.¹²

IMMUNE MODULATION HAS PROMISE IN ANTIPARASITIC THERAPY

234 Kinetoplastid infections provoke robust innate and adaptive 235 immune reactions, which can be protective or disease-236 promoting. This provides a rationale for investigating host-237 directed strategies such as immune modulators as an add-on to 238 antiparasitic therapy. Lessons from immuno-oncology may 239 offer a roadmap. Indeed, there are similarities in the dynamics 240 of host-tumor and host-parasite interactions. Both tumor 241 cells and cells harboring intracellular parasites are perceived by 242 the immune system as foreign, both retain features of normal 243 cells that could trigger an immune tolerance, and both can 244 create a microenvironment that facilitates immune escape and 245 promotes disease progression. In certain types of cancer, 246 adding immunotherapy to cytotoxic chemotherapy improves 247 survival. For example, a monoclonal antibody that binds to T 248 cells and blocks their inhibition by tumor cells is now part of 249 the first-line treatment for some lung cancers.¹³ An analogous 250 approach could conceivably help antiparasitic medicines to 251 work more quickly, more effectively, or with less variability.

Harnessing the immune system to treat kinetoplastid diseases is not a new idea. Beginning in the early 1990s, diseases is not a new idea. Beginning in the early 1990s, combination with antimony), with mixed results. Interleukin combination with antimony), with mixed results. Interleukin discontinue parasite growth, and experimental models suggest that that he IL-10 blockade can reduce disease progression.^{14,15} A so clinical trial with a humanized anti-IL-10 antibody was planned but later withdrawn due to problems in securing quality drug for the study (NCT01437020). IL-10 neutralization may also provide a benefit in CL.¹⁶ Additionally, TLR9 agonist CpG D35 is currently undergoing preclinical development in combine the problem of the problem of the study of the preparation for clinical trials for CL.⁶

In Chagas disease, the association of several pro- and antiinflammatory cytokines has been observed with cardiac and indeterminate forms of the disease, respectively.¹⁷ Some immunomodulation strategies postulated for Chagas disease include limiting regulatory T cells and increasing IL-17,¹⁸ rough overall there is less evidence to support immune mune mune 21 modulation in Chagas disease compared with leishmaniasis.

272 Naturally, there are challenges to testing and deploying 273 immunotherapies. Success or failure in experimental models is 274 not necessarily predictive of outcomes in humans. Patients 275 with leishmaniasis are at high risk of coinfection with bacteria; 276 therefore, modulating immunity in these populations will 277 require careful safety monitoring. Even if immunotherapy is 278 successful in enhancing the response to treatment in acute 279 disease, there are no tools to definitively assess latent infection. Finally, the immune system is dynamic and changes with age, 280 pregnancy, coinfections, and other conditions, which would 281 need to be considered. 282

ADJUNCT THERAPIES ARE ALSO NECESSARY IN 283 THE CLINICAL ARMORY 284

Adjunct therapies that improve outcomes including quality of 285 life should be pursued in parallel with work to discover 286 parasite-specific agents. These include medicines to improve 287 wound healing in CL, to address nutrition or coinfection 288 complications in VL, and to better manage cardiovascular and 289 gastrointestinal complications in Chagas disease. This also 290 applies to preventative or therapeutic vaccines, which are in 291 various phases of development. 292

Progress toward adjunct therapies is hampered in part by 293 our limited understanding of many features of disease biology. 294 There are, however, illustrative examples. In CL, the disease is 295 in large part mediated by the inflammatory immune response. 296 For example, lesions from L. braziliensis patients have few 297 parasites but severe ulceration, and experimental studies 298 indicate that a blockade of IL-1 β or the NLRP3 inflammasome 299 may ameliorate the disease in these patients.¹⁹ There is 300 evidence that wound treatment with pharmaceutical sodium 301 chlorite 0.045% or radio-frequency-induced heat therapy has 302 clinical benefits for CL.²⁰ In chronic Chagas cardiomyopathy, 303 patients generally suffer worse outcomes than those with heart 304 failure from other causes, despite the fact that Chagas disease 305 patients are usually younger and have fewer comorbidities.²¹ 306 Recently, angiotensin receptor-neprilysin inhibition was found 307 to reduce mortality and hospitalization in a large group of heart 308 failure patients with a reduced ejection fraction, including a 309 subgroup of patients with Chagas disease.²² Future work will 310 be needed to determine the specific implications for Chagas 311 disease patients. 312

SUMMARY

While there have been substantial advances in recent years to 314 address kinetoplastid diseases, on the whole these conditions 315 remain severely neglected across the domains of health policy, 316 advocacy, funding, and research. For HAT, more work is 317 needed to ensure that the gains realized are not lost. With 318 respect to finding safe and effective new therapies for Chagas 319 disease and leishmaniasis, we highlight several key priorities. 320 To start, the fundamental pathobiology of these diseases must 321 be further demystified to pave the way for targeted treatments; 322 the discovery of persister parasites is a sobering reminder that 323 we have much to learn before definitive medicines can be 324 generated. Novel tools will be needed to successfully validate 325 clinical candidates in patients, including biomarkers capable of 326 measuring intracellular parasite clearance and predicting the 327 clinical benefit without the need for extended follow-up. The 328 potential benefits in kinetoplastid diseases for immune 329 modulation and adjunct therapies need to be carefully 330 evaluated. Finally, we advocate continued and even greater 331 multidisciplinary collaboration. In the face of limited resources, 332 with an all too small scientific and medical community focused 333 on these complex diseases, harmonized research and develop- 334 ment strategies will be essential to accelerating progress toward 335 the common good of transformative new therapies for patients. 336

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355 **REFERENCES**

(1) Aufderheide, A. C., Salo, W., Madden, M., Streitz, J., Buikstra, J.,
Guhl, F., Arriaza, B., Renier, C., Wittmers, L. E., Jr., Fornaciari, G., and
Allison, M. (2004) A 9,000-year record of Chagas' disease. *Proc. Natl. Acad. Sci. U. S. A.* 101 (7), 2034–9.

(2) Mesu, V., Kalonji, W. M., Bardonneau, C., Mordt, O. V., Blesson,
S., Simon, F., Delhomme, S., Bernhard, S., Kuziena, W., Lubaki, J. F.,
Vuvu, S. L., Ngima, P. N., Mbembo, H. M., Ilunga, M., Bonama, A. K.,
Heradi, J. A., Solomo, J. L. L., Mandula, G., Badibabi, L. K., Dama, F.
R., Lukula, P. K., Tete, D. N., Lumbala, C., Scherrer, B., StrubWourgaft, N., and Tarral, A. (2018) Oral fexinidazole for late-stage
African Trypanosoma brucei gambiense trypanosomiasis: a pivotal
multicentre, randomised, non-inferiority trial. *Lancet 391* (10116),
144–154.

369 (3) Morillo, C. A., Marin-Neto, J. A., Avezum, A., Sosa-Estani, S., 370 Rassi, A., Jr., Rosas, F., Villena, E., Quiroz, R., Bonilla, R., Britto, C., 371 Guhl, F., Velazquez, E., Bonilla, L., Meeks, B., Rao-Melacini, P., 372 Pogue, J., Mattos, A., Lazdins, J., Rassi, A., Connolly, S. J., Yusuf, S., 373 and Investigators, B. (2015) Randomized Trial of Benznidazole for 374 Chronic Chagas' Cardiomyopathy. *N. Engl. J. Med.* 373 (14), 1295– 375 1306.

376 (4) Viotti, R., Vigliano, C., Lococo, B., Bertocchi, G., Petti, M., 377 Alvarez, M. G., Postan, M., and Armenti, A. (2006) Long-term cardiac 378 outcomes of treating chronic Chagas disease with benznidazole versus 379 no treatment: a nonrandomized trial. *Ann. Intern. Med.* 144 (10), 380 724–34.

(5) Khare, S., Nagle, A. S., Biggart, A., Lai, Y. H., Liang, F., Davis, L.
2C., Barnes, S. W., Mathison, C. J., Myburgh, E., Gao, M. Y., Gillespie,
3J. R., Liu, X., Tan, J. L., Stinson, M., Rivera, I. C., Ballard, J., Yeh, V.,
4Groessl, T., Federe, G., Koh, H. X., Venable, J. D., Bursulaya, B.,
Shapiro, M., Mishra, P. K., Spraggon, G., Brock, A., Mottram, J. C.,
Buckner, F. S., Rao, S. P., Wen, B. G., Walker, J. R., Tuntland, T.,
Molteni, V., Glynne, R. J., and Supek, F. (2016) Proteasome
inhibition for treatment of leishmaniasis, Chagas disease and sleeping
sickness. *Nature 537* (7619), 229–233.

(6) DNDi, Drugs for Neglected Diseases Initiative Research and
 391 Development Portfolio; https://www.dndi.org/diseases-projects/
 392 portfolio/, 2018.

(7) Lewis, M. D., Francisco, A. F., Taylor, M. C., and Kelly, J. M.
(2015) A new experimental model for assessing drug efficacy against
Trypanosoma cruzi infection based on highly sensitive in vivo
imaging. J. Biomol. Screening 20 (1), 36–43.

(8) Morillo, C. A., Waskin, H., Sosa-Estani, S., Del Carmen Bangher,
398 M., Cuneo, C., Milesi, R., Mallagray, M., Apt, W., Beloscar, J., Gascon,
399 J., Molina, I., Echeverria, L. E., Colombo, H., Perez-Molina, J. A.,
400 Wyss, F., Meeks, B., Bonilla, L. R., Gao, P., Wei, B., McCarthy, M.,

Yusuf, S., and Investigators, S.-C. (2017) Benznidazole and 401 Posaconazole in Eliminating Parasites in Asymptomatic T. Cruzi 402 Carriers: The STOP-CHAGAS Trial. J. Am. Coll Cardiol 69 (8), 939–403 947. 404

(9) Sanchez-Valdez, F. J., Padilla, A., Wang, W., Orr, D., and 405 Tarleton, R. L., Spontaneous dormancy protects Trypanosoma cruzi 406 during extended drug exposure. *Elife* 2018, 7, DOI: 10.7554/ 407 eLife.34039. 408

(10) Mandell, M. A., and Beverley, S. M. (2017) Continual renewal 409 and replication of persistent Leishmania major parasites in 410 concomitantly immune hosts. *Proc. Natl. Acad. Sci. U. S. A. 114* (5), 411 E801–E810. 412

(11) Kloehn, J., Saunders, E. C., O'Callaghan, S., Dagley, M. J., and 413 McConville, M. J. (2015) Characterization of metabolically quiescent 414 Leishmania parasites in murine lesions using heavy water labeling. 415 *PLoS Pathog.* 11 (2), e1004683. 416

(12) Tegazzini, D., Diaz, R., Aguilar, F., Pena, I., Presa, J. L., Yardley, 417 V., Martin, J. J., Coteron, J. M., Croft, S. L., and Cantizani, J. (2016) A 418 Replicative In Vitro Assay for Drug Discovery against Leishmania 419 donovani. *Antimicrob. Agents Chemother.* 60 (6), 3524–32. 420

(13) Gandhi, L., Rodriguez-Abreu, D., Gadgeel, S., Esteban, E., 421 Felip, E., De Angelis, F., Domine, M., Clingan, P., Hochmair, M. J., 422 Powell, S. F., Cheng, S. Y., Bischoff, H. G., Peled, N., Grossi, F., 423 Jennens, R. R., Reck, M., Hui, R., Garon, E. B., Boyer, M., Rubio- 424 Viqueira, B., Novello, S., Kurata, T., Gray, J. E., Vida, J., Wei, Z., Yang, 425 J., Raftopoulos, H., Pietanza, M. C., Garassino, M. C., and 426 Investigators, K. (2018) Pembrolizumab plus Chemotherapy in 427 Metastatic Non-Small-Cell Lung Cancer. N. Engl. J. Med. 378 (22), 428 2078–2092. 429

(14) Murray, H. W., Lu, C. M., Mauze, S., Freeman, S., Moreira, A. 430 L., Kaplan, G., and Coffman, R. L. (2002) Interleukin-10 (IL-10) in 431 experimental visceral leishmaniasis and IL-10 receptor blockade as 432 immunotherapy. *Infect. Immun.* 70 (11), 6284–93. 433

(15) Gautam, S., Kumar, R., Maurya, R., Nylen, S., Ansari, N., Rai, 434 M., Sundar, S., and Sacks, D. (2011) IL-10 neutralization promotes 435 parasite clearance in splenic aspirate cells from patients with visceral 436 leishmaniasis. J. Infect. Dis. 204 (7), 1134–7. 437

(16) Belkaid, Y., Hoffmann, K. F., Mendez, S., Kamhawi, S., Udey, 438 M. C., Wynn, T. A., and Sacks, D. L. (2001) The role of interleukin 439 (IL)-10 in the persistence of Leishmania major in the skin after 440 healing and the therapeutic potential of anti-IL-10 receptor antibody 441 for sterile cure. *J. Exp. Med.* 194 (10), 1497–506. 442

(17) Dutra, W. O., Menezes, C. A., Magalhaes, L. M., and Gollob, K. 443
J. (2014) Immunoregulatory networks in human Chagas disease. 444 *Parasite Immunol.* 36 (8), 377–387. 445

(18) Magalhaes, L. M., Villani, F. N., Nunes Mdo, C., Gollob, K. J., 446 Rocha, M. O., and Dutra, W. O. (2013) High interleukin 17 447 expression is correlated with better cardiac function in human Chagas 448 disease. J. Infect. Dis. 207 (4), 661–665. 449

(19) Novais, F. O., Carvalho, A. M., Clark, M. L., Carvalho, L. P., 450 Beiting, D. P., Brodsky, I. E., Carvalho, E. M., and Scott, P. (2017) 451 CD8+ T cell cytotoxicity mediates pathology in the skin by 452 inflammasome activation and IL-1beta production. *PLoS Pathog.* 13 453 (2), e1006196. 454

(20) David, J. R. (2018) The successful use of radiofrequency- 455 induced heat therapy for cutaneous leishmaniasis: a review. *Para*- 456 sitology 145 (4), 527–536.

(21) Shen, L., Ramires, F., Martinez, F., Bodanese, L. C., Echeverria, 458 L. E., Gomez, E. A., Abraham, W. T., Dickstein, K., Kober, L., Packer, 459 M., Rouleau, J. L., Solomon, S. D., Swedberg, K., Zile, M. R., Jhund, P. 460 S., Gimpelewicz, C. R., McMurray, J. J. V., and Paradigm, H. F. 461 Contemporary Characteristics and Outcomes in Chagasic Heart 462 Failure Compared With Other Nonischemic and Ischemic Cardiomy- 463 opathy. *Circ. Heart Fail.* 2017, *10* (11), DOI: 10.1161/CIRCH- 464 EARTFAILURE.117.004361.

(22) McMurray, J. J., Packer, M., Desai, A. S., Gong, J., Lefkowitz, 466 M. P., Rizkala, A. R., Rouleau, J. L., Shi, V. C., Solomon, S. D., 467 Swedberg, K., Zile, M. R., and Investigators, P.-H.; Committees 468 469 (2014) Angiotensin-neprilysin inhibition versus enalapril in heart 470 failure. *N. Engl. J. Med.* 371 (11), 993–1004.