



UNIVERSITY OF LEEDS

This is a repository copy of *Aragonite formation in confinements: A step toward understanding polymorph control*.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/139261/>

Version: Accepted Version

Article:

Xu, Y orcid.org/0000-0001-5180-8892 and Sommerdijk, NAJM (2018) Aragonite formation in confinements: A step toward understanding polymorph control. *Proceedings of the National Academy of Sciences of the United States of America*, 115 (34). pp. 8469-8471. ISSN 0027-8424

<https://doi.org/10.1073/pnas.1811696115>

© 2018. Published under the PNAS license. This is an author produced version of a paper published in *Proceedings of the National Academy of Sciences of the United States of America*. Uploaded in accordance with the publisher's self-archiving policy.

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk
<https://eprints.whiterose.ac.uk/>

Aragonite formation in confinements: a step towards understanding polymorph control

Yifei Xu^{1,2} and Nico A. J. M. Sommerdijk^{1,2}

1. Department of Chemical Engineering and Chemistry, Laboratory of Materials and Interface Chemistry and Centre for Multiscale Electron Microscopy, Eindhoven University of Technology, PO Box 513, 5600 MB, Eindhoven, The Netherlands

2. Institute for Complex Molecular Systems, Eindhoven University of Technology, PO Box 513, 5600 MB, Eindhoven, The Netherlands

Correspondence to Nico A. J. M. Sommerdijk: n.sommerdijk@tue.nl

Calcium carbonate (CaCO_3) is one of the most common minerals on the earth, which not only forms rocks like limestone or marble, but is also a main component of biominerals such as pearls, the nacre of sea shells and sea urchin skeletons.(1) Despite many years of research, the polymorphism of CaCO_3 is still far from being understood. CaCO_3 has three anhydrous crystalline forms: calcite, aragonite and vaterite, with a decreasing thermodynamic stability under aqueous ambient conditions (calcite>aragonite>vaterite).(2) While vaterite is rare in Nature, calcite and aragonite are both frequently found in rocks or biominerals.(1) A well-known example is the aragonite structure of nacre,(3) where the organization of the crystals leads to extraordinary mechanical performances. However, in synthetic systems, crystallization experiments only generate a small fraction of aragonite compared to calcite at ambient conditions and in the absence of additives.(4) So how is the formation of aragonite facilitated in Nature, especially in biominerals? In their recent PNAS paper, Zeng et al. shed light on this matter by showing that aragonite formation is dramatically promoted within confinements.(5)

In recent decades, great efforts have been spent towards understanding the strategies exploited by organisms to regulate aragonite formation, and several key factors have been identified. Up to now, the effect of Mg^{2+} additive is most well-established. Mg^{2+} is abundant in seawater and is expected to be present during the formation of many marine biominerals.(6, 7) At high $\text{Mg}^{2+}:\text{Ca}^{2+}$ ratios, aragonite forms as the major crystalline form instead of calcite at room temperature. This was recently explained by Sun et al. in PNAS,(8) who show that Mg^{2+} can significantly increase the surface energy of calcite and raise its nucleation barrier, while aragonite is much less affected. Meanwhile, insoluble organic matrices and soluble acidic macromolecules extracted from aragonite-forming tissues also favor aragonite formation to different extents.(9, 10) Detailed mechanisms of the effects remain unclear, but were generally attributed to the interaction between the acidic functional groups of the biomacromolecules and the mineral components. Additionally it was reported that a macromolecular hydrogel-like 3D network is formed prior to the mineralization of nacre,(11) which may play a role in the crystallization process by confining the crystallization to defined small volumes. Nonetheless, confinement has never been directly correlated with aragonite formation in biominerals, although its capability on controlling crystal orientation and polymorphism has been shown in many recent reports.(12-15)

Now, Zeng et al. explored for the first time the impact of confinement on aragonite formation.(5) By precipitating CaCO_3 within the cylindrical pores of track-etched membranes (Fig. 1), they investigated the relationship between pore size and the polymorphism of CaCO_3 . Strikingly, a high level of aragonite formation was detected within these nanosized confinements. Using the same concentrations of Mg^{2+}

and SO_4^{2-} as additives, the aragonite proportion in bulk solution was only 7%, while this value increased to 69% in 200 nm sized nanopores, and reached 100% in 50 nm sized pores. When even smaller sized pores were used (25 nm), pure aragonite crystals were obtained even in the absence of any additives. More than that, the aragonite crystallized within these nanopores were mainly single crystal rods, highly oriented along the c-axis.

The authors subsequently examined several possible origins for the preferred formation of aragonite within these confinements. Confinement is known to increase the incubation time for crystallization nucleation, inhibiting the formation of thermodynamically stable phases (e.g., calcite) and in favor of metastable phases.(13) This is, however, unlikely the reason for aragonite formation here since aragonite is seldom seen as a precursor to calcite. Computations demonstrated that the reaction was also not affected by a variation of diffusion rates. Hence, the only reasonable cause for the promotion of aragonite formation seems to be the influence of the pore surface on crystal nucleation. Indeed, aragonite formation was further promoted when smaller sized pores were used and larger pore surfaces were generated. Zeng et al. suggest that the pore surface may modulate ion activity and thus facilitate aragonite formation. Unfortunately, experimental confirmation was hindered by the difficulty to directly measure the ionic profiles within the nanopores.

By showing that pure aragonite single crystals can be synthesized at ambient condition by only using nanosized confinements, the work of Zeng et al. provides a route for CaCO_3 polymorph control that may possibly also be applied by biomineralizing organisms. In particular, it points to the importance of surface effects in controlling aragonite formation. This also aligns with the computational work of Sun et al.,(8) showing that the presence of Mg^{2+} favors aragonite formation by modulating the surface energy of the crystals. Nevertheless, the details of the mechanism by which surface effects facilitate aragonite growth for now remain unknown. The surface interactions may modulate the CaCO_3 polymorph through tuning the distribution of ions, as proposed by the authors; alternatively, the effect may be due to a lower interfacial energy between the pore surface and aragonite, as compared to calcite. Another intriguing question is how the confinement promotes the formation of oriented single crystals, as was also reported for other mineral systems.(14-16) Clearly, future investigations of crystallization within porous membranes are likely to give us more important insights on the control of crystal orientation and polymorphism with possible relevance for both synthetic and biological systems.

Meanwhile, to what extent organisms indeed deploy nano-sized confinements to promote aragonite formation is a question that needs further discussion. One interesting system is nacre where lamellar

organic matrices were found between adjacent ~500 nm thick aragonite layers.(17) As this dimension is still a bit above the largest effective pore size (200 nm) reported by Zeng et al., it is likely that in addition to confinement also the interaction between the mineral and biomacromolecular matrix(9-11) plays a role in the preferred aragonite formation in nacre.

A similar fundamental question remains on how confinement is correlated to the selection of the aragonite polymorph. Zeng et al. show that the two main volume effects of confinement, i.e. inhibiting nucleation and limiting diffusion rate, are both irrelevant to the preferred aragonite formation, and the main role of confinement appears to be enhancing the surface effect.(5) Indeed, when track-etch membranes from different manufacturers were used, different CaCO₃ polymorphs form within the same sized pores.(15) This was attributed to possible differences in the density or conformation of the chemical species lining the membrane pores, as only minor differences of surface roughness and no differences in composition were detected for these membranes. Hence, maybe not only aragonite, but also other polymorphs of CaCO₃ can be selected by nano-sized confinements with the appropriate surface chemistry. This could certainly be a strategy employed in biomineralization, but could also potentially provide a new window for controlling polymorphism in the synthesis of crystalline materials. In conclusion, the results of Zeng et al. are of great significance for the understanding of polymorph control, which has seen some recent interesting advances,(18, 19) but is still in its infancy despite its importance, in particular in the preparation of pharmaceuticals.

Author Contributions

Y. X and N. A. J. M. S co-wrote the paper.

Conflict of Interest

The author declares no conflict of interest.

Acknowledgements

Research of the authors is financially supported by TOPPUNT from the Netherlands Organization of Scientific Research (NWO).

References

1. Lowenstam HA & Weiner S (1989) *On biomineralization* (Oxford University Press, New York).
2. Plummer LN & Busenberg E (1982) The solubilities of calcite, aragonite and vaterite in CO₂-H₂O solutions between 0 and 90 C, and an evaluation of the aqueous model for the system CaCO₃-CO₂-H₂O. *Geochim. Cosmochim. Acta* 46(6):1011-1040.
3. Jackson A, Vincent JF, & Turner R (1988) The mechanical design of nacre. *Proc. R. Soc. Lond. B* 234(1277):415-440.
4. Ogino T, Suzuki T, & Sawada K (1987) The formation and transformation mechanism of calcium carbonate in water. *Geochim. Cosmochim. Acta* 51(10):2757-2767.
5. Zeng M, *et al.* (2018) Confinement generates single-crystal aragonite rods at room temperature. *Proc. Nat. Acad. Sci. USA*
6. Morse JW, Wang Q, & Tsio MY (1997) Influences of temperature and Mg: Ca ratio on CaCO₃ precipitates from seawater. *Geology* 25(1):85-87.
7. Loste E, Wilson RM, Seshadri R, & Meldrum FC (2003) The role of magnesium in stabilising amorphous calcium carbonate and controlling calcite morphologies. *J. Cryst. Growth* 254(1-2):206-218.
8. Sun W, Jayaraman S, Chen W, Persson KA, & Ceder G (2015) Nucleation of metastable aragonite CaCO₃ in seawater. *Proc. Nat. Acad. Sci. USA*
9. Suzuki M, *et al.* (2009) An acidic matrix protein, Pif, is a key macromolecule for nacre formation. *Science* 325(5946):1388-1390.
10. Falini G, Albeck S, Weiner S, & Addadi L (1996) Control of aragonite or calcite polymorphism by mollusk shell macromolecules. *Science* 271(5245):67-69.
11. Nudelman F, *et al.* (2008) Forming nacreous layer of the shells of the bivalves *Atrina rigida* and *Pinctada margaritifera*: An environmental- and cryo-scanning electron microscopy study. *J. Struct. Biol.* 162(2):290-300.
12. Jiang Q & Ward MD (2014) Crystallization under nanoscale confinement. *Chem. Soc. Rev.* 43(7):2066-2079.
13. Stephens CJ, Ladden SF, Meldrum FC, & Christenson HK (2010) Amorphous calcium carbonate is stabilized in confinement. *Adv. Funct. Mater.* 20(13):2108-2115.

14. Cantaert B, Beniash E, & Meldrum FC (2013) Nanoscale confinement controls the crystallization of calcium phosphate: relevance to bone formation. *Chem.--Eur. J.* 19(44):14918-14924.
15. Schenk AS, Albarracin EJ, Kim Y-Y, Ihli J, & Meldrum FC (2014) Confinement stabilises single crystal vaterite rods. *Chem. Commun.* 50(36):4729-4732.
16. Kim YY, *et al.* (2011) Capillarity Creates Single - Crystal Calcite Nanowires from Amorphous Calcium Carbonate. *Angew. Chem. Int. Ed.* 50(52):12572-12577.
17. Lia A, Derk J, Fabio N, & Steve W (2006) Mollusk Shell Formation: A Source of New Concepts for Understanding Biomineralization Processes. *Chem.--Eur. J.* 12(4):980-987.
18. Van Driessche AE, *et al.* (2018) Molecular nucleation mechanisms and control strategies for crystal polymorph selection. *Nature* 556(7699):89.
19. Nanev CN (2018) Recent Insights into Protein Crystal Nucleation. *Crystals* 8(5):219.

Figure Captions

Fig. 1. Scheme of CaCO_3 crystallization within the cylindrical nanopores of track-etched membranes. While crystals formed in the bulk solution is mainly calcite, aragonite single crystals oriented in c-axis are formed within the nanopores.

