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- 1 Landmark Papers: No. 7
- 2

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Commentary on the impact of Smith et al. (2003): by G. Guggenberger, B. Ludwig & M. Menon

9

# 10 Introduction

Smith et al. (2003) published their review on the interactions of soil physical factors and 11 biological processes controlling the exchange of greenhouse gases between soil and atmosphere 12 13 at a time when global change was already considered to be one of the most important challenges of mankind (IPCC, 2001). In the Climate Change 2014 Synthesis Report (IPCC, 2014) a global 14 15 warming of 0.7°C between 1951 and 2010 was reported and further warming and long-lasting changes in all components of the climate systems forecasted. Smith K.A. et al. (2003) and later 16 Smith P. et al. (2008) emphasized that about one third of CH<sub>4</sub> and two thirds of N<sub>2</sub>O emitted 17 globally to the atmosphere per year derive from soil processes, while soil is considered a small 18 CO<sub>2</sub> sink, which may change with increasing warming (Crowther et al., 2016). This is reason 19 enough to analyse the processes that lead to this net emission of gases to the atmosphere. While 20 21 biological processes produce or consume these greenhouse gases, the size of the fluxes is 22 strongly controlled by soil physical factors. However, the controlling factors on the interaction between the controlling physical factors and biological processes in the exchange of greenhouse 23 gases between the soil and atmosphere had not been widely considered. Keith Smith and his 24 co-authors were pioneering in this field (e.g. Smith, 1980; Ball et al., 1997a, Ball et al., 1997b; 25

Conen et al., 2000; Dobbie & Smith, 2001), which finally led to the review of Smith et al.
(2003).

28

29 Controlling factors for CO<sub>2</sub> emissions

Smith et al. (2003) summarized that the release of CO<sub>2</sub> by aerobic respiration can be described 30 by a non-linear function of temperature over a wide range of water contents. The link between 31 microbial processes and physical factors, in addition to availability of substrate and chemical 32 factors (e.g. soil pH), is of substantial importance because of the direct and indirect effects of 33 physical factors on the production of CO<sub>2</sub> by microorganisms and roots. The non-linear 34 35 response of CO<sub>2</sub> as a function of temperature has been confirmed in several recent studies (e.g. Schaufler et al., 2010). The factors affected by water content that were discussed by Smith et 36 al. (2003) are also now well established; water is important for gas diffusivity (Ball, 2013) and 37 38 substrate supply to soil microorganisms (Schindlbacher et al., 2004). Notably, Schaufler et al. (2010) reported that maximum CO<sub>2</sub> emissions from European soils under different land uses 39 occur at intermediate soil moisture, which accords well with the summarizing synthesis by 40 Smith et al. (2003). 41

Smith et al. (2003) reported a marked scatter of Q<sub>10</sub> values for CO<sub>2</sub> emissions and pointed 42 43 out the need for standardization and accurate interpretation of temperature responses of the soil's CO<sub>2</sub> emissions at greater depths. They emphasized that for accurate determinations and 44 interpretations of Q<sub>10</sub> values, diurnal temperature changes, thermal conductivities and thermal 45 diffusivities of the soil need to be considered in greater detail in future studies. In fact, a later 46 study by Pavelka et al. (2007) also addressed this important issue and recommended 47 measurement of soil temperature at a very shallow soil depth to determine useable values of 48 Q<sub>10</sub>, and suggested a procedure to standardize Q<sub>10</sub> values for soil temperatures measured at 49 different depths. 50

### 52 Controlling factors for N<sub>2</sub>O emissions

53 For N<sub>2</sub>O, Smith et al. (2003) focused on the important microbiological processes of nitrification of ammonium and denitrification of nitrate in soil, and the governing processes for the 54 respective rates. In particular, they elucidated soil conditions, e.g. structure, wetness, O<sub>2</sub> content 55 of pores and soil depth, being responsible for the release of N<sub>2</sub>O to the atmosphere or further 56 reduction to N<sub>2</sub>. Nitrate ammonification and nitrifier denitrification as additional processes 57 leading to the formation of N2O have been discussed since in greater detail by Baggs & Phillipot 58 (2010) and Smith (2017). The merit of the review by Smith et al. (2003) lies again in the 59 important emphasis of the link between microbial processes and physical factors in addition to 60 61 other factors, such as substrate availability and chemical factors such as soil pH (e.g. Weslien et al., 2009). This link is crucial for an understanding and prognosis of N<sub>2</sub>O emissions. 62

Smith et al. (2003) emphasized that the anaerobic volume is affected by increases in the 63 water-filled pore space (WFPS), where an increase in WFPS may also result in an exponential 64 increase in N<sub>2</sub>O emissions. There is still some controversy about which physical soil property 65 is most useful for estimating N<sub>2</sub>O emissions; for example the ratio of gas diffusivity within the 66 soil to that in free air, the degree of aggregation and compaction, matric potential, WFPS and 67 volumetric water content (for a discussion see Ball, 2013 and Smith, 2017). Smith et al. (2003) 68 69 indicated that N<sub>2</sub>O emissions also increase markedly with temperature. They attributed this to increases in the anaerobic volume fraction. An increase in temperature results in an increase in 70 the size of the anaerobic zones because of increased respiration, which causes larger gradients 71 72 in O<sub>2</sub>. In addition, increased temperatures are also likely to lead to increased rates of denitrification per unit anaerobic volume. Both increases then favour a dramatic increase in 73 N<sub>2</sub>O emissions. In fact, the concept of anaerobic zones is a key feature of the process-based 74 DNDC (denitrification-decomposition) model, for which there are several versions for different 75 land uses. This model has a kinetic scheme for the anaerobic volumetric fraction (an 'anaerobic 76 balloon') that is implemented to calculate the anaerobic fraction of soil in a given soil layer in 77

relation to O<sub>2</sub> diffusion and the respiratory activity of soil micro-organisms and roots (for a
summary of the DNDC model see Gilhespy et al., 2014). Overall, there is no doubt that soil
temperature and soil moisture are important for explaining much of the temporal variation in
N<sub>2</sub>O emissions within a site (e.g. Pilegaard et al. 2006).

82

83 Controlling factors for CH<sub>4</sub> emissions

For CH<sub>4</sub> production and transport, Smith et al. (2003) reported that ebullition and diffusion 84 through the aerenchyma of rice and plants in natural wetlands contribute substantially to the 85 emission of CH<sub>4</sub> and that the proportion of the emissions taking place by each pathway varies 86 87 seasonally.. The oxidation of atmospheric CH<sub>4</sub> to CO<sub>2</sub> is controlled by gas diffusivity, whereas the effect of temperature is small (Smith et al., 2003). Ball (2013) suggested that the control of 88 gas diffusivity on the oxidation of CH<sub>4</sub> might not hold for all sites and that the effect of pH, 89 90 moisture, temperature, and nitrogen and type of organic matter and content might be pronounced. The role of nitrogen as a regulatory factor of CH<sub>4</sub> oxidation has been addressed in 91 92 detail by Bodelier & Laanbroek (2004), who discussed the inhibiting role of additions of nitrogenous fertilizer. The effect of WFPS on CH<sub>4</sub> oxidation may be seen as a hump-shaped 93 function where the optimum oxidation occurs at 20-50% WFPS. At smaller water contents, 94 95 desiccation stress and at larger water contents diffusion limitation might be inhibiting CH<sub>4</sub> oxidation (Dunfield, 2007). Thus, moist, well-aerated soil favours CH<sub>4</sub> oxidation and CO<sub>2</sub> 96 exchange (Ball, 2013). 97

98

99 Soil structure, microbial communities and greenhouse gas emissions

Smith et al. (2003) emphasized that although the greenhouse gases are produced by microbial processes, the size of their fluxes between soil and atmosphere depends largely on soil physical factors. The transport of gases within the soil and the gas exchange between soil and atmosphere is a function of gas diffusivity, which depends on the air-filled porosity or, inversely, with the

WFPS. Most soils develop a three-dimensional architecture with pedogenesis, which is 104 105 characterized by the aggregate size distribution. The distribution of aggregates largely controls almost every process in soil. This refers to the air-filled porosity or WFPS at a given matric 106 107 potential (Ball, 2013) as well as to the distribution of microbial populations in soil (Nunan et al., 2003). Therefore, soil structure controls the habitat of the actors involved in the production 108 of greenhouse gases and determines the diffusion of  $O_2$  and dissolved organic matter (DOM) to 109 110 fuel aerobic microbes. Consequently, inter- and intra-aggregate pore space needs to be considered. Sey et al. (2008) compared the greenhouse gas emissions from various aggregate 111 size classes (<0.25 mm, 0.25-2 mm and 2-6 mm) and from 2-mm sieved bulk soil at different 112 113 WFPS (20, 40, 80 and 80%). They found that denitrification was responsible for 95% of N<sub>2</sub>O emissions in microaggregates, whereas nitrification was responsible for 97-99% of N<sub>2</sub>O 114 production in macroaggregates. This inferred that diffusion of O<sub>2</sub> was largely inhibited in 115 116 microaggregates when the WFPS was 80%, whereas macroaggregates maintained aerobic conditions. 117

The interrelations between soil structure and greenhouse gas emissions can be readily 118 investigated when the natural soil structure and size distribution of aggregates are disrupted due 119 to external forces (e.g. compaction), which in turn can alter the pore size distribution and 120 hydraulic properties (Menon et al., 2015). Beare et al. (2009) showed that the production of 121 N<sub>2</sub>O was 67 times greater in compacted than uncompacted soil at field moisture contents, and 122 they demonstrated the effect of soil moisture on emissions of N<sub>2</sub>O and CO<sub>2</sub>. Deurer et al. (2012) 123 124 reported enhanced carbon sequestration under the wheel tracks, probably because of reduced microbial decomposition of organic matter. Bessou et al. (2010) also found that compacted soil 125 had smaller emissions of CO<sub>2</sub>, but at the same time larger N<sub>2</sub>O emissions by inducing anoxic 126 conditions favourable for denitrification activity. 127

Experiments with compacted soil also help to elucidate the relation between microbial communities and greenhouse gas emissions depending on soil physical factors. So Nadian et al. (1998) reported a significant decline in vesicular-arbuscular mycorrhizal fungi biomass at
higher bulk density, and Peacock et al. (2001) found a significant reduction in microbial
biomass for heavy traffic treatments. Schnurr-Pütz et al. (2006) observed that fungi, in
particular, are negatively affected by soil compaction, whereas denitrifiers and methanogens
appear to be more prominent. From that, the links between soil physical properties and
greenhouse gas emissions can be conceptualized as in Fig. 1.

136

### 137 New developments in linking soil physical factors to biological processes

In their landmark paper on the interactions of soil physical factors and biological processes, 138 Smith et al. (2003) focus on gas diffusivity, which affects soil aeration and the capacity of the 139 soil microbial community to produce or consume CO<sub>2</sub>, N<sub>2</sub>O and CH<sub>4</sub>. The concept of hotspots 140 and hot moments (Kuzyakov & Blagodatskaya, 2015) adds the supply of the organic substrates, 141 which is also linked partly to soil physical factors. Transport of the labile OM sources to the 142 microbial community occurs largely through biotic activities such as the release of root 143 144 exudates (Jones et al. 2004) and the detritus of soil animals (Schrader et al. 2007), but also as 145 DOM leached from the O and A horizons (Qualls & Haines 1992). Translocation of DOM to the subsoil depends strongly on the flow paths in soil and on soil structure and precipitation 146 events (Leinemann et al. 2016). Because DOM is mainly translocated in the inter-aggregate 147 pore space of the soil, it is retained on aggregate surfaces, which are enriched in OM (Amelung 148 et al. 2002), thus creating a hotspot. At the same time, the inter-aggregate pore space usually 149 150 enables good aeration, leading to the release of CO<sub>2</sub> with microbial decomposition of the substrate. In otherwise aerobic soil, strong microenvironments may exist that are important 151 152 sources of N<sub>2</sub>O and CH<sub>4</sub> (Keiluweit et al. 2016). Hotspots of denitrification and methanogenesis in the intra-aggregate pore space results from slow diffusion of O<sub>2</sub>, whereas in the rhizosphere 153 this is caused by the inflow of very available OM from root exudation (Henry et al. 2008). This, 154 155 once again, emphasizes the complex interplay of soil physical factors and biological processes

in the production of greenhouse gases in soil and their exchange between soil and atmosphere(Smith et al. 2003).

158

159 Methodological progress

The landmark paper of Smith et al. (2003) on these interactions also triggered substantially the 160 161 methodological development with respect to the visualization of pores of different size, to 162 measurement of microbial activity and the resulting O<sub>2</sub> and CO<sub>2</sub> partial pressures at small scales, and the development of physical and biophysical models. In the last decade much 163 progress in the understanding of soil structure and the associated pore-space architecture has 164 165 been? gained by X-ray computed tomography (CT), which enables an in-situ and real-time 3-D mapping at scales of a few microns. Measured properties include porosity, pore-size 166 distribution, tortuosity and topology (Naveed et al. 2013; Vogel et al. 2010). Peth et al. (2014) 167 168 showed that synchrotron-based X-ray CT in combination with osmium staining is not only suitable for describing soil structure, but also for identifying the location of organic matter in 169 170 soil, e.g. in the intra-aggregate pore space. Neutron radiography emerged as a useful method to map the water distribution within soil and its temporal changes (Oswald et al. 2008; Carminati 171 et al. 2010), whereas the 2-D distribution of oxygen concentration can be analysed by 172 173 fluorescence imaging with planar optodes (Blossfeld et al. 2011). Rudolph-Mohr et al. (2017) emphasized the great potential of combining neutron radiography with fluorescence imaging 174 to investigate the effect of different soil moisture conditions on the oxygen patterns in soil. Such 175 176 analyses may provide important input parameters for geometry-based mechanistic models.

Keith Smith also pioneered modelling of microbial respiration and denitrification at the aggregate scale by systematically incorporating factors such as oxygen supply and nitrogen concentration (Smith, 1980). Ebrahimi & Or (2015, 2016) have built on that and developed a 3-D pore-scale model that simulates the aerobic and anaerobic microbial communities within aggregates together with rates of production of N<sub>2</sub>O and CO<sub>2</sub> along the aggregate radius. This

model considers substrate and oxygen diffusion processes and is integrated with individual cellbased models that link soil physical processes with microbial community dynamics. Ebrahimi
& Or (2016) upscaled this modelling framework to quantify depth-resolved rates of production
of CO<sub>2</sub> and N<sub>2</sub>O depending on small-scale environmental conditions. In a very recent model,
this approach was used to quantify methane production in thawing permafrost soil, based on
the microbial activity dynamics in pore networks with? consideration of transport dynamics and
physiological aspects of the cells (Ebrahimi & Orr, 2017).

189

# 190 Conclusions

191 Smith and co-authors expressed hope that their review would demonstrate the key roles played by soil physical factors in controlling the biological processes responsible for the exchange of 192 greenhouse gases between soil and atmosphere. Indeed, the authors convincingly built a bridge 193 194 between soil physics and soil biology. From this landmark publication and some other manuscripts, soil biophysics has developed as an emerging field within the soil sciences. Inter-195 196 and intra-aggregate pore architecture is decisive in the control of the availability of O<sub>2</sub> and organic substrates to microorganisms. It is thus of utmost importance not only for the 197 production of the different greenhouse gases, but also for organic matter stabilization and biotic 198 199 redox processes associated with mineral weathering and mineral transformation. The effect of biota on soil physical factors has also received increasing interest recently. This concerns, for 200 example, the formation of aggregates by living and dead organic agents, which affects soil 201 202 structure and associated pore architecture, or the rhizosphere, where water uptake by the roots 203 strongly modifies the WFPS. Novel instrumental and modelling approaches will allow an understanding of the multiple interactions between soil physical and biotic processes in soil in 204 relation to soil functioning and ecosystem services. This is only possible by crossing the 205 boundaries in soils science, which is what this landmark paper emphasized. 206

### 208 **References**

- 209 Amelung, W., Kaiser, K., Kammerer, G. & Sauer, G. 2002. Organic carbon at soil particle
- surfaces—Evidence from X-ray photoelectron spectroscopy and surface abrasion. Soil
  Science Society of America Journal, 66, 1526–1530.
- Baggs, E. & Phillipot, L. 2010. Microbial terrestrial pathways to nitrous oxide. In: Nitrous
  Oxide and Climate Change (ed. K.A. Smith), pp. 4–35. Earthscan, London.
- Ball, B.C. 2013. Soil structure and greenhouse gas emissions: a synthesis of 20 years of
  experimentation. European Journal of Soil Science, 64, 357–373.
- 216 Ball, B.C., Dobbie, K.E., Parker, J.PO. & Smith, K.A. 1997a. The influence of gas transport
- and porosity on methane oxidation in soils. Journal of Geophysical Research, 102, 23309–
  23317.
- Ball, B.C., Smith, K.A., Klemedtsson, L., Brumme, R., Sitaula, B.K., Hansen, S. et al. 1997b.
  The influence of soil gas transport properties on methane oxidation in a selection of northern
  European soils. Journal of Geophysical Research, 53, 29–39.
- 222 Benckiser, G., Schartel, T. & Weiske, A. 2015. Control of  $NO_3^-$  and  $N_2O$  emission in 223 agroecosystems: a review. Agronomy for Sustainable Development, **35**, 1059–1074.
- Bessou, C., Mary, B., Leonard, J., Roussel, M., Grehan, E. & Gabrielle, B. 2010. Modelling
- soil compaction impacts on nitrous oxide emissions in arable fields. European Journal of
  Soil Science, 61, 348–363.
- Blossfeld, S., Gansert, D., Thiele, B., Kuhn, A.J. & Lösch, R. 2011. The dynamics of oxygen
  concentration, pH value, and organic acids in the rhizosphere of Juncus spp. Soil Biology &
- Biochemistry, **43**, 1186–1197.
- Bodelier, P.L.E. & Laanbroek, H.J. 2004. Nitrogen as a regulatory factor of methane oxidation
  in soils and sediments. FEMS Microbiology Ecology, 47, 265–277.
- 232 Carminati, A., Moradi, A.B., Vetterlein, D., Vontobel, P., Lehmann, E., Weller, U., et al. 2010.
- 233 Dynamics of soil water content in the rhizosphere. Plant and Soil, **332**, 163–176.

- Conen, F., Dobbie, K.E. & Smith, K.A. 2000. Predicting N<sub>2</sub>O emissions from agricultural land
   through related soil parameters. Global Change Biology, 6, 417–426.
- 236 Crowther, T.W., Todd-Brown, K.E.O., Rowe, C.W., Wieder, W.R., Carey, J.C., Machmuller,
- M.B. et al. 2016. Quantifying global soil carbon losses in response to warming. Nature, 540,
  104–108.
- Dobbie, K.E. & Smith, K.A. 2001. The effects of temperature, water-filled pore space and land
  use on N<sub>2</sub>O emissions from an imperfectly drained gleysol. European Journal of Soil
  Science, 52, 667–673.
- 242 Dunfield, P.F. 2007. The soil methane sink. In: Greenhouse Gas Sinks (eds D.S. Reay, C.N.
- Hewitt, K.A. Smith & J. Grace), pp. 152–157. CAB International, Wallingford, UK.
- Ebrahimi, A. & Or, D. 2015. Hydration and diffusion processes shape microbial community
  organization and function in model soil aggregates. Water Resources Research, 51, 9804–
- **246** 9827.
- Ebrahimi, A. & Or, D. 2016. Microbial community dynamics in soil aggregates shape
  biogeochemical gas fluxes from soil profiles—Upscaling an aggregate biophysical model.
- 249 Global Change Biology, **22**, 3141–3156.
- 250 Ebrahimi, A. & Or, D. 2017. Mechanistic modeling of microbial interactions at pore to profile
- scale resolve methane emission dynamics from permafrost soil. Journal of Geophysical
  Research: Biogeosciences, **122**, 1216–1238.
- 253 Gilhespy, S.L., Anthony, S., Cardenas, L., Chadwick, D., del Prado, A., Li, C.S., Misselbrook
- T. et al. 2014. First 20 years of DNDC (DeNitrification DeComposition): model evolution.
  Ecological Modelling, 292, 51–62.
- Henry, S., Texier, S., Hallet, S., Bru, D., Dambreville, C., Chèneby, D. et al. 2008.
  Disentangling the rhizosphere effect on nitrate reducers and denitrifiers: insight into the role
  of root exudates. Environmental Microbiology, 10, 3082–3092.

- 259 IPCC. 2001. Climate Change 2001: The Scientific Basis. Contribution of Working Group I to
- the Third Assessment Report of the Intergovernmental Panel on Climate Change (eds J.T.
- Houghton, Y. Ding, D.J. Griggs, M. Noguer, P.J. van der Linden, X. Dai et al.) Cambridge
  University Press, Cambridge, United Kingdom and New York, NY, USA, .
- 263 IPCC. 2014. Climate Change 2014: Synthesis Report. Contribution of Working Groups I, II
- and III to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change
- 265 (eds Core Writing Team, R.K. Pauchauri & L.A. Meyer). IPCC, Geneva, Switzerland.
- Jones, D.L., Hodge, A. & Kuzyakov, Y. 2004. Plant and mycorrhizal regulation of
  rhizodeposition. New Phytologist, 163, 459–480.
- Keiluweit, M., Nico, P.S., Kleber, M. & Fendorf, S. 2016. Are oxygen limitations under
  recognized regulators of organic carbon turnover in upland soils? Biogeochemistry, 127,
  157–171.
- Leinemann, T., Mikutta, R., Kalbitz, K., Schaarschmidt, F. & Guggenberger, G. 2016. Small
  scale variability of vertical water and dissolved organic matter fluxes in sandy Cambisol
  subsoils as revealed by segmented suction plates. Biogeochemistry, 131, 1–15.
- 274 Menon, M., Jia, X., Lair, G.J., Fraj, P.H. & Blaud, A. 2015. Analysing the impact of compaction
- of soil aggregates using X-ray microtomography and water flow simulations. Soil & Tillage
  Research, 150, 147–157.
- Nadian, H., Smith, S.E., Alston, A.M., Murray, R.S. & Siebert, B.D. 1998. Effects of soil
  compaction on phosphorus uptake and growth of Trifolium subterraneum colonized by four
- species of vesicular-arbuscular mycorrhizal fungi. New Phytologist, **140**, 155–165.
- 280 Naveed, M., Moldrup, P, Arthur, E., Wildenschild, D., Eden, M., Lamande, M. et al. 2013.
- 281 Revealing soil structure and functional macroporosity along a clay gradient using X-ray
- computed tomography. Soil Science Society of America Journal, **77**, 403–411.

- Nunan, N., Wu, K., Young, I.M., Crawford, J.W. & Ritz, K. 2003. Spatial distribution of
  bacterial communities and their relationships with the micro-architecture of soils. FEMS
  Microbiology Ecology, 44, 203–215.
- Oswald, S.E., Menon, M., Carminati, A., Vontobel, P., Lehmann, E. & Schulin, R. 2008.
  Quantitative imaging of infiltration, root growth, and root water uptake via neutron
  radiography. Vadose Zone Journal, 7, 1035–1047.
- Pavelka, M., Acosta, M., Marek, M.V., Kutsch, W. & Janous, D. 2007. Dependence of the Q<sub>10</sub>
  values on the depth of the soil temperature measuring point. Plant and Soil, 292, 171–179
- 291 Peth, S., Chenu, C., Leblond, N., Mordhorst, A., Garnier, P., Nunan, N. et al. 2014. Localization
- of soil organic matter in soil aggregates using synchrotron-based X-ray microtomography.
- 293 Soil Biology & Biochemistry, **78**, 189–194.
- Pilegaard, K., Skiba, U., Ambus, P., Beier, C., Brueggemann, N., Butterbach-Bahl, K. et al.
  2006. Factors controlling regional differences in forest soil emission of nitrogen oxides (NO
  and N<sub>2</sub>O). Biogeosciences, 3, 651–661.
- Qualls, R.G. & Haines, B.L. 1992. Biodegradability of dissolved organic matter in forest
  throughfall, soil solution, and stream water. Soil Science Society of America Journal, 56,
  578–586.
- Rudolph-Mohr, N., Tötzke, C., Kardjilov, N. & Oswald, S.E. 2017. Mapping water, oxygen,
  and pH dynamics in the rhizosphere of young maize roots. Journal of Plant Nutrition and
  Soil Science, 180, 336–346.
- Schaufler, G., Kitzler, B., Schindlbacher, A., Skiba, U., Sutton, M.A. & ZechmeisterBoltenstern, S. 2010. Greenhouse gas emissions from European soils under different land
  use: effects of soil moisture and temperature. European Journal of Soil Science, 61, 683–
  696.

- Schindlbacher, A., Zechmeister-Boltenstern, S. & Butterbach-Bahl, K. 2004. Effects of soil
  moisture and temperature on NO, NO<sub>2</sub>, and N<sub>2</sub>O emissions from European forest soils.
  Journal of Geophysical Research, 109, 1–12.
- Sey, B.K., Maceur, A.M., Wahlen, J.K., Gregorich, E.G. & Rochette, P. 2008. Small-scale
- 311 heterogeneity in carbon dioxide, nitrous oxide and methane production from aggregates of a
- cultivated sandy-loam soil. Soil Biology & Biochemistry, **40**, 2468–2473.
- Smith, K.A. 1980. A model of the extent of anaerobic zones in aggregated soils, and its potential
  application to estimates of denitrification. Journal of Soil Science, **31**, 263–277.
- Smith, K.A., 2017. Changing views of nitrous oxide emissions from agricultural soil: key
- 316 controlling processes and assessment at different spatial scales. European Journal of Soil
  317 Science, 68, 137–155.
- Smith, K.A., Ball, T., Conen, F., Dobbie, K.E., Massheder, J. & Rey, A. 2003. Exchange of
  greenhouse gases between soil and atmosphere: interactions of soil physical factors and
  biological processes. European Journal of Soil Science, 54, 779–791.
- Smith, P., Fang, C., Dawson, J.J.C. & Moncrieff, J.B. 2008. Impact of global warming on soil
  organic carbon. Advances in Agronomy, 97, 1–43.
- Tiunov, A.V. & Scheu, S. 2000. Microbial biomass, biovolume and respiration in Lumbricus
  terrestris L. cast material of different age. Soil Biology & Biochemistry, 32, 265–275.
- 325 Schnurr-Pütz, S., Bååth, E., Guggenberger, G., Drake, H. & Küsel, K. 2006. Compaction of
- forest soil by logging machinery favours occurrence of prokaryotes. FEMS Microbiology
  Ecology, 58, 503–516.
- Vogel, H.-J., Weller, U. & Schlüter, S. 2010. Quantification of soil structure based on
  Minkowksi function. Computational Geosciences, 36, 1236–1245.
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# 333 Figure

334

Figure 1. Conceptual model on the link between soil physical properties and greenhouse gas 335 emission depending on soil compaction (Menon and Blaud, unpublished). Soil compaction 336 337 leads to changes in soil structure (e.g. porosity), which will affect the flow of air and water, 338 and thereby create a more anaerobic environment in soil. This may lead to a shift in the relative abundance and functions of the microbial population, shown here as effects on the 339 340 C and N cycles. Abundance of nitrifiers and aerobic degraders are given by dashed lines and 341 abundance of denitrifiers and methanogens are given by solid lines. 342 343