



This is a repository copy of *The Carpathians hosted extra-Mediterranean refugia-within-refugia during the Pleistocene Ice Age: genomic evidence from two newt genera.*

White Rose Research Online URL for this paper:  
<http://eprints.whiterose.ac.uk/124388/>

Version: Accepted Version

---

**Article:**

Wielstra, B. [orcid.org/0000-0002-7112-5965](https://orcid.org/0000-0002-7112-5965), Zieliński, P. and Babik, W. (2017) The Carpathians hosted extra-Mediterranean refugia-within-refugia during the Pleistocene Ice Age: genomic evidence from two newt genera. *Biological Journal of the Linnean Society*, 122 (3). pp. 605-613. ISSN 0024-4066

<https://doi.org/10.1093/biolinnean/blx087>

---

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.



[eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk)  
<https://eprints.whiterose.ac.uk/>

1 Running head: **Carpathian refugia-within-refugia**

2

3 Title: **The Carpathians hosted extra-Mediterranean refugia-within-refugia during the Pleistocene**

4 **Ice Age: genomic evidence from two newt genera**

5

6 Ben Wielstra<sup>1,2,3,\*</sup>, Piotr Zieliński<sup>4</sup> and Wiesław Babik<sup>4</sup>

7

8 <sup>1</sup> Department of Animal and Plant Sciences, University of Sheffield, S10 2TN Sheffield, UK. <sup>2</sup>

9 Department of Ecology and Evolutionary Biology, University of California, Los Angeles, CA 90095,

10 USA. <sup>3</sup> Naturalis Biodiversity Center, P.O. Box 9517, 2300 RA Leiden, The Netherlands. <sup>4</sup> Institute of

11 Environmental Sciences, Jagiellonian University, ul. Gronostajowa 7, 30-387, Kraków, Poland.

12

13 \* Corresponding author:

14 Department of Ecology and Evolutionary Biology

15 University of California, Los Angeles

16 CA 90095, USA

17 Email: [ben.wielstra@naturalis.nl](mailto:ben.wielstra@naturalis.nl)

18

19 **Abstract**

20 Part of Europe's temperate species survived the Pleistocene glacial cycles in refugia north of the  
21 Mediterranean peninsulas. For one such extra-Mediterranean refugia, the Carpathians, an intricate  
22 'refugia-within-refugia' scenario has been suggested, involving species surviving in multiple discrete  
23 glacial refugia. We test the Carpathian refugia-within-refugia hypothesis, employing genome-wide  
24 multilocus datasets for two newt species (*Triturus cristatus* and *Lissotriton montandoni*). We first use  
25 Bayesian clustering to delineate intraspecific evolutionary lineages. The number of intraspecific  
26 lineages identified, and the allocation of localities to these lineages, were used to construct testable  
27 hypotheses on the spatial arrangement of glacial refugia in both newt species. Next we employ  
28 approximate Bayesian computation to date whether these lineages are of Holocene (< 12 Ka) or  
29 Pleistocene (> 12 Ka) origin. We identify three intraspecific evolutionary lineages for *T. cristatus* and  
30 two for *L. montandoni*. For both newt species, intraspecific divergence is rooted in the Pleistocene, in  
31 line with species survival in distinct range fragments during the last glacial period. Hence, our findings  
32 firmly support the Carpathian refugia-within-refugia hypothesis. Furthermore, we show that  
33 mitochondrial DNA overestimates the age of intraspecific evolutionary lineages and we urge caution in  
34 basing refugia-within-refugia scenarios on mitochondrial DNA alone.

35

36 **Keywords:** Approximate Bayesian computation; Bayesian clustering; Historical biogeography;  
37 *Lissotriton montandoni*; Next-generation sequencing; *Triturus cristatus*; Quaternary

38

## 39 **Introduction**

40 The climate oscillations of the Pleistocene Ice Age moulded intraspecific genetic structuring by  
41 repeatedly reducing temperate species' ranges during glacial cycles (Hewitt, 2000). The refugia-within-  
42 refugia concept addresses the evolution of intraspecific geographical genetic structuring, as species  
43 survive glacial conditions in fragmented pockets of suitable habitat within a single, wider refugial area  
44 (Abellán & Svenning, 2014; Gómez & Lunt 2007). Refugia-within-refugia have been reported from  
45 Europe's canonical glacial refugia: the Iberian (Gómez & Lunt 2007), Italian (Canestrelli et al., 2014)  
46 and Balkan (Poulakakis et al., 2015) Peninsulas. As regions situated north of Europe's southern  
47 peninsula are increasingly appreciated as sources of postglacial recolonization of temperate Europe  
48 (Schmitt & Varga, 2012; Stewart et al., 2010), this raises the question whether such areas also facilitated  
49 intraspecific Pleistocene differentiation. The Carpathians are arguably the most significant extra-  
50 Mediterranean refugium and accumulating phylogeographic studies suggest a refugia-within-refugia  
51 scenario applies (Mráz & Ronikier, 2016). We test this hypothesis here, using two newt species from  
52 different genera as a system.

53         The Northern crested newt *Triturus cristatus* (Laurenti, 1768) is a species of lowland and hills,  
54 distributed over much of temperate Europe and adjacent Asia, while the Carpathian newt *Lissotriton*  
55 *montandoni* (Boulenger, 1880) is a montane species, endemic to the Carpathians (Fig. 1a). Despite their  
56 ecological differences (Speybroeck et al., 2016), both species survived the Pleistocene glaciations in  
57 the Carpathians (Wielstra, Babik & Arntzen, 2015; Wielstra et al., 2013; Zieliński et al., 2014a;  
58 Zieliński et al., 2013). As genetic data show geographical sub-structuring and species distribution  
59 models suggest glacial range fragmentation within the Carpathians, these species are particularly  
60 suitable to test the Carpathian refugia-within-refugia hypothesis. We sequence several dozen nuclear  
61 DNA markers and use Bayesian clustering to delineate intraspecific geographical structure within each  
62 species. Subsequently, we test in an approximate Bayesian computation framework whether the  
63 observed intraspecific structure indeed arose during the Pleistocene, which would indicate species  
64 survival in multiple discrete glacial refugia and thus support the Carpathian refugia-within-refugia  
65 hypothesis.

66

## 67 **Material and methods**

68

### 69 **Sampling**

70 For *T. cristatus* we included 28 Carpathian breeding ponds (Fig. 1b) and an additional seven positioned  
71 in postglacially colonized area and sampled up to three (2.9 on average) individuals per pond (see Table  
72 S1 in Appendix S1). For *L. montandoni* we included 31 Carpathian breeding ponds (Fig. 1c) and  
73 sampled up to three (1.3 on average) individuals per pond (Table S2 in Appendix S1). Individual ponds  
74 were treated as localities.

75

### 76 **Summary of sequencing**

77 For *T. cristatus* we newly sequenced 52 nuclear markers. See Wielstra et al. (2014) for a detailed  
78 description of the laboratory and bioinformatics protocol. In brief, we amplified markers of c. 140 bp  
79 in length (excluding primers), positioned in 3' untranslated regions of protein-coding genes, in five  
80 multiplex PCRs. We pooled the multiplexes for each individual and ligated unique tags to be able to  
81 recognize the amplicons belonging to each individual. We sequenced the amplicons on the Ion Torrent  
82 next-generation sequencing platform and processed the output with a bioinformatics pipeline that filters  
83 out poor quality reads, identifies alleles and converts data to a genotypic format directly usable for  
84 population genetic analysis. Mean coverage was 777 reads (range 0-13,622) per marker-individual  
85 combination. Marker-individual combinations with  $\geq 20$  reads (1.73%) were considered successful.

86 For *L. montandoni* we sequenced 74 nuclear markers. See Zieliński et al. (2014b) for a detailed  
87 description of the laboratory and bioinformatics protocol. In brief, we amplified markers of ca. 500 bp  
88 in length (excluding primers), positioned in 3' untranslated regions of protein-coding genes, in ten  
89 multiplex PCRs. Again, multiplexes for each individual were pooled and given unique tags. We  
90 sequenced the amplicons on the Illumina MiSeq next-generation sequencing platform to the average  
91 per base coverage of  $1017 \pm$  (SD) 1181. Sequence data were further processed using standard, freely  
92 available bioinformatic tools, producing phase-resolved variants. Fasta files were obtained from vcf  
93 files using custom python script. Marker-individual combinations with  $< 10$  reads were considered  
94 failed. These data were previously used in another study (Zieliński et al., 2016).

95

96 Bayesian clustering analysis – constructing hypotheses

97 Triturus and Lissotriton newts hybridize with congeneric species at their contact zones (Arntzen,  
98 Wielstra & Wallis, 2014; Zieliński et al., 2016) and while introgression of mitochondrial DNA in *T.*  
99 *crystatus* is restricted to the contact zone with congeneric species (Wielstra et al., 2015), it has led to the  
100 complete replacement of the original mitochondrial DNA of *L. montandoni* (Babik et al., 2005;  
101 Zieliński et al., 2013). Including individuals showing recent genetic admixture with another species  
102 (early generation hybrids) could have unpredictable effects in downstream analyses of intraspecific  
103 genetic divergence, while limited nuclear DNA introgression (via ancient hybridization) simply  
104 constitutes a part of intraspecific genetic diversity and as such does not require separate treatment in  
105 our models.

106 To confirm there were no early generation hybrids present in our dataset we took a two-step  
107 approach. We first used STRUCTURE 2.3.4 (Pritchard, Stephens & Donnelly, 2000) to confirm that our  
108 set of *T. cristatus* and *L. montandoni* individuals did not show significant genetic admixture  
109 (STRUCTURE  $Q \geq 0.05$ ) with Triturus or Lissotriton species with abutting ranges. We did so by enforcing  
110 the number of gene pools ( $k$ ) to 2 in pairwise species comparisons (Tables S1-S2 in Appendix S1).  
111 Entire haplotypes were treated as alleles at each locus. We used the admixture model in combination  
112 with the correlated allele frequency model with 100,000 iterations, after 50,000 iterations of burn-in,  
113 and ran five replicates, which were summarized with CLUMPAK (Kopelman et al., 2015). As *T. cristatus*  
114 has parapatric range boundaries with four other Triturus species, namely *T. carnifex*, *T. dobrogicus*, *T.*  
115 *ivanbureschi* and *T. macedonicus*, we took reference data for these species, four localities per species  
116 with three individuals per locality, from Wielstra et al. (2014). The only species that *L. montandoni* has  
117 a parapatric range boundary with is *L. vulgaris* and we took reference data for this species, 45  
118 individuals from 38 localities, from Zieliński et al. (2016).

119 Next, we used STRUCTURE to determine the number of intraspecific evolutionary lineages in  
120 both newt species. We used the same settings as before but tested over multiple values of  $k$ . The upper  
121  $k$  limit was 35 for *T. cristatus* and 31 for *L. montandoni*, as defined by the total number of localities  
122 included. We used Evanno's  $\Delta k$  criterion (Evanno, Regnaut & Goudet, 2005) to select the optimum  $k$

123 value. The number of intraspecific lineages identified, and the allocation of localities to these lineages,  
124 were used to construct testable hypotheses on the spatial arrangement of glacial refugia in both newt  
125 species.

126

#### 127 Approximate Bayesian computation – rationale

128 Using approximate Bayesian computation (ABC) we evaluated the existence of three *T. cristatus* and  
129 two *L. montandoni* glacial refugia in the Carpathians (as suggested by STRUCTURE) by testing models  
130 assuming: 1) a Holocene (< 12 Ka) and 2) a Pleistocene (> 12 Ka) origin of intraspecific evolutionary  
131 lineages. Support for the latter model would imply that the evolutionary lineages diverged prior to the  
132 end of the last glacial maximum and hence must have survived glaciations in separate refugia. Therefore  
133 our ABC modelling can be considered an explicit test of the refugia-within-refugia hypothesis. Within  
134 species, all parameter priors (except topology for *T. cristatus*) were identical for the tested models and  
135 no demographic changes and historical gene flow were allowed to keep models as simple as possible.

136

#### 137 Approximate Bayesian computation – data preparation

138 For *T. cristatus* we focus on localities in the Carpathian area only (1-28). According to the STRUCTURE  
139 results, crested newt localities were assigned to three lineages, one within (**TcB**), one east of (**TcE**) and  
140 one south of (**TcS**) the Carpathian mountain belt (figure 1b). We considered three topologies: 1) (**TcE**,  
141 **TcS**) **TcB** – supported by a drift tree based on allele frequency data, 2) **TcB**, **TcE**, **TcS** – a polytomy,  
142 and 3) (**TcB**, **TcS**) **TcE** - supported by the nucleotide distance between evolutionary lineages (see Fig.  
143 S1 in Appendix S2). Two localities (2 and 9) showing a high degree of admixture between evolutionary  
144 lineages ( $0.3 < Q < 0.7$ ) were not analysed to exclude the effect of ongoing hybridization and early-  
145 generation admixture. Seven markers (agl, clasp, gys, samdb, smo, taf8, and usp) in which more than  
146 5% of individuals did not amplify were removed. Furthermore, 11 individuals for which one or more  
147 of the retained markers did not amplify were excluded. Next, alignment columns with missing data (i.e.  
148 indels) were removed. We assume that newt breeding ponds correspond to discrete demes which may  
149 undergo extinction and recolonization. To minimize the confounding effects of current population  
150 structure we randomly subsampled one gene copy per locality. It has been shown (Wakeley, 2004;

151 Wakeley & Aliacar, 2001) that if one gene copy per locus is sampled per deme in a metapopulation  
152 composed of a large number of demes, the ancestral process producing such a sample is identical to the  
153 unstructured coalescent process. Our final ABC dataset contained one gene copy per locus from 25  
154 localities, distributed over the three evolutionary lineages as follows: 7 **TcB**, 9 **TcE** and 9 **TcS**.

155 For *L. montandoni* STRUCTURE suggested two lineages: one south (**LmS**) and one north (**LmN**)  
156 of approximately the centre of the Eastern Carpathians, roughly the Ukrainian/Romanian border (figure  
157 1c). As there are only two evolutionary lineages in *L. montandoni* we only had to consider a single  
158 topology: **LmN**, **LmS** (Fig. S2 in Appendix S2). We excluded two individuals from localities with  
159 considerable admixture between evolutionary lineages ( $0.3 < Q < 0.7$ ). For the ABC analysis, we  
160 excluded eight markers that were fully coding or amplified inconsistently so that the final data set  
161 included 66 markers. Furthermore, five individuals for which one or more of the retained markers did  
162 not amplify were excluded. Next, alignment columns with missing data were removed. As explained  
163 above for *T. cristatus*, one gene copy per marker was sampled per locality. Our final ABC dataset  
164 contained one gene copy per locus from 24 localities, distributed over the two evolutionary lineages as  
165 follows: 9 **LmN** and 15 **LmS**.

166

#### 167 Approximate Bayesian computation – summary statistics

168 We focused on a set of basic summary statistics, likely to be informative about the time of the split  
169 between intraspecific evolutionary lineages, and other demographic parameters. For each evolutionary  
170 lineage we calculated average and variance of: number of fixed polymorphisms (SF), number of shared  
171 polymorphisms (SS), number of private polymorphisms (SP), nucleotide  $F_{ST}$  ( $F_{ST\_nuc}$ ) calculated  
172 between evolutionary lineages and between a particular evolutionary lineage and the remaining ones  
173 pooled (in three-lineage models), Tajima's D (D), nucleotide diversity ( $\Pi$ ), number of haplotypes  
174 (nHap), haplotype diversity (HapW),  $d_{xy}$  calculated between lineages ( $\Pi_A$ ) and the number of  
175 haplotypes shared between all lineages and lineage pairs (n\_shared\_hap). Additionally we calculated  
176 average and variance of nHap, HapW, D,  $\Pi$  and the overall number of segregating sites (S) for the  
177 whole dataset. Summary statistics for both observed and simulated data sets were calculated on  
178 polymorphic biallelic sites only. Positions with more than two segregating variants were excluded as

179 departing from the infinite sites model. For each statistic, mean and variance across all loci were  
180 calculated using MSSTATSPOP v.0.998980-beta (Ramos-Onsins et al., unpublished) and custom Python  
181 scripts.

182

183 Approximate Bayesian computation – simulations and analysis

184 Coalescent simulations were performed using FASTSIMCOAL2.01 (Excoffier et al., 2013). We simulated  
185 data using the finite site mutation model (as our data did not fit the infinite site model) and a single,  
186 fixed mutation rate of  $= 5.7 \times 10^{-9}$  per site, per generation, as previously estimated for smooth and  
187 Carpathian newts using fossil-based dating of divergence within genus *Lissotriton* (Pabijan et al., 2015;  
188 Zieliński et al., 2016). Considering that *Triturus* and *Lissotriton* are relatively closely related (Zhang et  
189 al., 2008) and we use highly similar genetic markers (Wielstra et al., 2014; Zieliński et al., 2014b), we  
190 considered it appropriate to use the same mutation rate for both systems. These markers are known to  
191 be unlinked in both newt systems (Wielstra et al., 2017; Zieliński et al., 2016). Loci were simulated as  
192 independent chromosomes. The ABC analysis was performed within the ABCTOOLBOX (Wegmann et  
193 al., 2010). We used a generation time of 4 years based on the synthesis of the literature (Nadachowska  
194 & Babik, 2009) and assumed it appropriate to use this value for *Triturus* as well (Duellman & Trueb,  
195 1994). Our recombination rate priors were based on a previous estimate for smooth and Carpathian  
196 newts (Zieliński et al., 2016) (Tables S3-S4 in Appendix S1). Parameter values were sampled from  
197 uniform prior distributions, priors for population sizes were uniform on a log<sub>10</sub> scale (Tables S3-S4 in  
198 Appendix S1) and were set to cover biologically plausible values. Analyses were based on  $10^6$  datasets  
199 simulated under each demographic model. We retained the 0.1% ( $10^3$ ) best simulations for each model  
200 and computed the marginal likelihood of the observed and retained datasets under a Generalized Linear  
201 Model (Leuenberger & Wegmann, 2010). For each species we compared all models in a single model  
202 selection procedure and selected the best fitting ones based on posterior probabilities. We inspected  
203 posterior probability curves and the fraction of retained simulations with the marginal likelihood smaller  
204 or equal to that of the observed data (observed P-value) to determine if models could faithfully  
205 reproduce the observed data. The best fitting model was selected based on Bayes factors (ratios of model  
206 marginal densities). To estimate the power to distinguish between models we generated 1,000 pseudo-

207 observed datasets for each model and checked how often the ABC model choice procedure correctly  
208 predicted the true model (the one that produced the dataset). Each pseudo-observed dataset was treated  
209 as the observed data and used to calculate marginal densities of all compared models. Bayes factors  
210 were then used to select the best model. As we were interested in both the overall power to identify the  
211 true model as well as the power in the observed summary statistics space, the pseudo-observed datasets  
212 for each model were chosen from both random and retained simulations. To check whether the marginal  
213 posterior distributions estimated from the best models were biased, we generated 1,000 pseudo-  
214 observed data sets for each best model and tested uniformity of the posterior quantile distributions (the  
215 position of the true values within the posterior distribution) for each parameter with a Kolmogorov–  
216 Smirnov test. If the parameter values for these pseudo-observed data were randomly chosen from the  
217 prior distribution, we expect the posterior quantiles to be uniformly distributed. Because for *T. cristatus*  
218 (while a Holocene divergence was confidently rejected) the posterior validation suggested potential  
219 overestimation of divergence time in the preferred model, we further explored this matter by rerunning  
220 the preferred model 1) without a fixed lower prior boundary for split time and 2) without a fixed lower  
221 prior boundary for split time and with an upper prior boundary for split time fixed to 0.5 Ma.

222

## 223 **Results**

224

225 Bayesian clustering analysis – intraspecific evolutionary lineages

226 STRUCTURE confirmed our set of *T. cristatus* and *L. montandoni* individuals showed no significant  
227 recent genetic admixture with congeners. STRUCTURE suggested  $k = 3$  as the most likely number of  
228 gene pools for *T. cristatus* and  $k = 2$  for *L. montandoni* (Tables S1-S2 in Appendix S1). The three *T.*  
229 *cristatus* lineages roughly correspond to within (**TcB**), east of (**TcE**) and south of (**TcS**) the Carpathian  
230 mountain belt (figure 1b). Lineage **TcB** is also the one that postglacially colonized temperate Eurasia.  
231 The two *L. montandoni* lineages show a different geographical configuration, with an evolutionary  
232 lineage south (**LmS**) and north (**LmN**) of approximately the centre of the Eastern Carpathians, roughly  
233 the Ukrainian/Romanian border (figure 1c).

234

235 Approximate Bayesian computation – polymorphism and observed summary statistics

236 The *T. cristatus* ABC dataset included 45 markers of the average length 139 bp (6,248 bp). There were  
237 106 haplotypes out of which 51 (48%) were shared between evolutionary lineages. We observed 67  
238 polymorphic sites out of which four (6%) were private to **TcB**, 19 (28%) to **TcE** and 23 (34%) to **TcS**.  
239 The *L. montandoni* ABC data set comprised of 66 markers of the average length of 484 bp (31,929 bp).  
240 There were 391 haplotypes out of which 105 (27%) were shared between evolutionary lineages. We  
241 observed 652 polymorphic sites out of which 156 (24%) were private to **LmN**, 283 (43%) to **LmS**. In  
242 both species the percent of sites segregating in all lineages was similar, 31% in *T. cristatus* and 33% in  
243 *L. montandoni*. We found no fixed differences between lineages in either species (Tables S5-S6 in  
244 Appendix S1).

245

246 Approximate Bayesian computation – model choice for *Triturus cristatus*

247 The P-values calculated under the Generalized Linear Model were used to check whether tested models  
248 were able to reproduce the observed data (Table S5 in Appendix S1). For all *T. cristatus* models  
249 assuming a Pleistocene split (M2, M4, M6), the observed data fell well within the distribution of  
250 retained simulated data (Table S7 in Appendix S1). Models assuming a Pleistocene divergence were  
251 always favoured and the polytomy model (M4) had the highest posterior probability (PP=0.95) (Table  
252 S7 in Appendix S1). The mean power to identify the true model was 0.59 and in the case of the preferred  
253 M4 model it was 0.74 (Table S9 in Appendix S1). Although within the observed summary statistics  
254 space the M4 model power decreased to 0.37, there was no case in which simulations produced under  
255 other models would choose M4 as the true model more often than the model of origin. Importantly,  
256 only simulations under other models of a Pleistocene divergence selected M4 more often than expected  
257 by chance (Table S9 in Appendix S1). The selected M4 model indicates a Middle Pleistocene 172 Ka  
258 (77-281 Ka) divergence between lineages (Fig. S3 in Appendix S2, Table S3 in Appendix S1).

259 According to the posterior validation (Fig. S5 in Appendix S2, Table S3 in Appendix S1),  
260 divergence time might be overestimated for model M4, so the estimates should be treated with caution.  
261 It needs to be stressed here, however, that hypothesis testing was based on model selection, not on  
262 parameter estimates. Therefore, the bias in the divergence time estimates does not affect the main results

263 of our test, which firmly supports the Pleistocene divergence and rejects a Holocene divergence. Still,  
264 to interpret whether this bias affected the actual divergence time estimate within the preferred  
265 Pleistocene divergence model, we reran the preferred model without a fixed lower prior boundary for  
266 split time (M7) and without a fixed lower prior boundary for split time and with an upper prior boundary  
267 for split time fixed to 0.5 Ma (M8). While M7 showed a similar bias as M4, bias was considerably  
268 reduced in M8 (details on Dryad). Yet, the inferred divergence time was almost identical (details on  
269 Dryad). Hence we conclude that the divergence time estimated in M4 is reliable.

270

271 Approximate Bayesian computation – model choice for *Lissotriton montandoni*

272 A model assuming Pleistocene divergence (M2) performed better than one assuming post-Pleistocene  
273 divergence (M1; Table S8 in Appendix S1). The power to correctly predict the true model was high for  
274 both models, regardless of statistics space (Table S10 in Appendix S1). According to the Kolmogorov–  
275 Smirnov test results (Table S4 in Appendix S1) all parameter distributions were biased. However,  
276 visual inspection of the distributions of divergence time posterior quantiles (Fig. S6 in Appendix S2)  
277 suggests that the true values were more often found in the centre of the distribution, which is a  
278 consequence of overly wide priors. Importantly, this kind of bias may only slightly decrease precision  
279 of the estimates. Hence, while our simple models were not able to faithfully reproduce the observed  
280 data (Table S6, S8 in Appendix S1), we nevertheless consider it safe to interpret the estimated  
281 divergence time from the best performing model. The selected M2 model again indicates a Middle  
282 Pleistocene 202 Ka (54–347 Ka) divergence between lineages (Fig. S4 in Appendix S2, Table S4 in  
283 Appendix S1).

284

## 285 **Discussion**

286 While the importance of the Carpathians as a glacial refugium has by now become well established, a  
287 more intricate pattern of refugia-within-refugia is still emerging (Mráz & Ronikier, 2016). We here  
288 tested the Carpathian refugia-within-refugia hypothesis, based on next-generation phylogeography and  
289 ABC analysis for two newt species of different genera. For both species, models assuming a Pleistocene  
290 divergence were strongly preferred, even though disparate patterns of intraspecific genetic structure

291 highlight that species had idiosyncratic responses to glacial cycles (Fig. 1). The build-up of deep  
292 intraspecific differentiation in ecologically distinct species provides strong support for a scenario in  
293 which multiple discrete regions within the Carpathians acted as glacial refugia, for a broad range of  
294 species. Our findings emphasize the key role that the Carpathians played in Pleistocene survival and  
295 radiation of temperate Eurasia's biodiversity.

296 Accuracy of our divergence time estimates, crucial for the interpretation of this test, could be  
297 affected by 1) the mutation rate and generation time used to convert ABC estimates into calendar years,  
298 and 2) gene flow between evolutionary lineages. Only a several-fold underestimation of mutation time  
299 or overestimation of generation time could lead to erroneous support for Pleistocene divergence, but,  
300 as the values used are well supported, we consider this unlikely. Furthermore, gene flow would cause  
301 under- rather than an overestimation of divergence time, yet post-Pleistocene divergence was still  
302 rejected. Hence, we conclude that our ABC analysis strongly supports a pre-Pleistocene divergence of  
303 evolutionary lineages and provides robust evidence for Carpathian refugia-within-refugia, illustrating  
304 the added value of ABC analysis in Carpathian phylogeography (see also Kolář et al., 2016).

305 Our nuclear DNA results suggest that the intraspecific structuring observed today originated  
306 during the penultimate glacial period (130-200 Ka). This is an order of magnitude younger than the  
307 divergence of the three mitochondrial DNA lineages present in *T. cristatus* (with even the most  
308 conservative interpretation based on confidence intervals suggesting divergence well over a million  
309 years ago), which have a similar distribution as the evolutionary lineages identified in the present study  
310 (Wielstra et al., 2015). It should be noted that no comparable mitochondrial DNA data is available for  
311 *L. montandoni*, as its native mitochondrial DNA relatively recently got replaced with that of a congener,  
312 via mitochondrial DNA capture (Zieliński et al., 2013). Nuclear gene flow upon secondary contact  
313 during Pleistocene interglacials would cause fusion of intraspecific lineages, a realistic scenario given  
314 the historical instability of phylogeographic patterns (Hofreiter et al., 2004), and in fact genetic  
315 admixture is observed where evolutionary lineages meet today. Phylogeographic structure is often  
316 retained longer in mitochondrial DNA than in the nuclear genome (Petit & Excoffier, 2009). While the  
317 long-term persistence of geographically structured mitochondrial DNA clades could be interpreted as  
318 evidence that the same areas acted as refugia during multiple glacial periods (Hewitt, 2011), our findings

319 underline that the stability and historical isolation of refugia-within-refugia delineated based on  
 320 mitochondrial DNA alone could well be overestimated. Considering the strong bias in phylogeographic  
 321 surveys towards mitochondrial DNA (Riddle, 2016), we suggest that proposed refugia-within-refugia  
 322 scenarios require re-evaluation with nuclear data.

323

### 324 **Acknowledgements**

325 BW initiated this work as a Newton International Fellow. This project has received funding from the  
 326 European Union's Horizon 2020 research and innovation programme under the Marie Skłodowska-  
 327 Curie grant agreement No. 655487. PZ and WB were supported by the Polish National Science Centre  
 328 grants 8171/B/P01/2011/40 and 2014/15/B/NZ8/00250 and the Jagiellonian University grant  
 329 DS/WBiNoZ/INoS/762/16. M. Samples were collected under permits: DOPozgiz-4200/II-  
 330 78/3702/10/JRO provided by the Polish General Director for Environmental Protection, 03.04.12 No.  
 331 67 issued by the National Academy of Sciences of Ukraine and 3256/9.07.2010 provided by the  
 332 Romanian Commission for Protection of Natural Monuments. Samples were collected with institutional  
 333 animal ethics approval (number 101/2009), issued by the First Local Ethical Committee on Animal  
 334 Testing at the Jagiellonian University in Krakow. J.W. Arntzen, M. Bonk, D. Cogălniceanu, S-D.  
 335 Covaciu-Marcov, K. Dudek, M. Herdegen, M. Liana, K. Nadachowska-Brzyska, Z. Prokop, J. Radwan  
 336 and M. Stuglik helped with sampling. P. Kania and A. Fijarczyk provided python scripts. We thank two  
 337 anonymous reviewers for their helpful comments.

338

### 339 **References**

- 340 **Abellán P, Svenning J-C. 2014.** Refugia within refugia – patterns in endemism and genetic divergence  
 341 are linked to Late Quaternary climate stability in the Iberian Peninsula. *Biological Journal of*  
 342 *the Linnean Society* **113**: 13-28.
- 343 **Arntzen JW, Wielstra B, Wallis GP. 2014.** The modality of nine *Triturus* newt hybrid zones, assessed  
 344 with nuclear, mitochondrial and morphological data. *Biological Journal of the Linnean Society*  
 345 **113**: 604–622.
- 346 **Babik W, Branicki W, Crnobrnja-Isailovic J, Cogălniceanu D, Sas I, Olgun K, Poyarkov NA,**  
 347 **Garcia-Paris M, Arntzen JW. 2005.** Phylogeography of two European newt species -  
 348 Discordance between mtDNA and morphology. *Molecular Ecology* **14**: 2475-2491.
- 349 **Boulenger GA. 1880.** Sur une forme intéressante de Triton provenant de Moldavie et observations sur  
 350 le genre *Pelonectes* Lataste. *Bulletin de la Société Zoologique de France* **5**: 37-40.
- 351 **Canestrelli D, Bisconti R, Sacco F, Nascetti G. 2014.** What triggers the rising of an intraspecific  
 352 biodiversity hotspot? Hints from the agile frog. *Scientific Reports* **4**.
- 353 **Duellman WE, Trueb L. 1994.** *Biology of amphibians.* JHU press.

- 354 **Evanno G, Regnaut S, Goudet J. 2005.** Detecting the number of clusters of individuals using the  
355 software structure: a simulation study. *Molecular Ecology* **14**: 2611-2620.
- 356 **Excoffier L, Dupanloup I, Huerta-Sánchez E, Sousa VC, Foll M. 2013.** Robust demographic  
357 inference from genomic and SNP data. *PLoS Genetics* **9**: e1003905.
- 358 **Gómez A, Lunt DH. 2007.** In: Weiss S and Ferrand N, eds. *Phylogeography of Southern European*  
359 *refugia*. Dordrecht: Springer.
- 360 **Hewitt G. 2000.** The genetic legacy of the Quaternary ice ages. *Nature* **405**: 907-913.
- 361 **Hewitt GM. 2011.** Mediterranean peninsulas: the evolution of hotspots. In: Zachos FE and Habel JC,  
362 eds. *Biodiversity hotspots: distribution and protection of conservation priority areas*. Berlin:  
363 Springer. 123-147.
- 364 **Hofreiter M, Serre D, Rohland N, Rabeder G, Nagel D, Conard N, Münzel S, Pääbo S. 2004.** Lack  
365 of phylogeography in European mammals before the last glaciation. *Proceedings of the*  
366 *National Academy of Sciences of the United States of America* **101**: 12963-12968.
- 367 **Kolář F, Fuxová G, Závěská E, Nagano AJ, Hyklová L, Lučanová M, Kudoh H, Marhold K. 2016.**  
368 Northern glacial refugia and altitudinal niche divergence shape genome-wide differentiation in  
369 the emerging plant model *Arabidopsis arenosa*. *Molecular Ecology* **25**: 3929-3949.
- 370 **Kopelman NM, Mayzel J, Jakobsson M, Rosenberg NA, Mayrose I. 2015.** Clumpak: a program for  
371 identifying clustering modes and packaging population structure inferences across K.  
372 *Molecular Ecology Resources* **15**: 1179–1191.
- 373 **Laurenti JN. 1768.** *Specimen Medium, Exhibens Synopsis Reptilium Emendatam cum Experimentis*  
374 *circa Venena et Antidota Reptilium Austriacorum*. J. Thomae Trattner: Vienna.
- 375 **Leuenberger C, Wegmann D. 2010.** Bayesian computation and model selection without likelihoods.  
376 *Genetics* **184**: 243-252.
- 377 **Mráz P, Ronikier M. 2016.** Biogeography of the Carpathians: evolutionary and spatial facets of  
378 biodiversity. *Biological Journal of the Linnean Society* **119**: 528-559.
- 379 **Nadachowska K, Babik W. 2009.** Divergence in the face of gene flow: the case of two newts  
380 (*Amphibia*: Salamandridae). *Molecular Biology and Evolution* **26**: 829-841.
- 381 **Pabijan M, Zieliński P, Dudek K, Chloupek M, Sotiropoulos K, Liana M, Babik W. 2015.** The  
382 dissection of a Pleistocene refugium: phylogeography of the smooth newt, *Lissotriton vulgaris*,  
383 in the Balkans. *Journal of Biogeography* **42**: 671–683.
- 384 **Petit RJ, Excoffier L. 2009.** Gene flow and species delimitation. *Trends in Ecology & Evolution* **24**:  
385 386-393.
- 386 **Poulakakis N, Kapli P, Lymberakis P, Trichas A, Vardinoyiannis K, Sfenthourakis S, Mylonas**  
387 **M. 2015.** A review of phylogeographic analyses of animal taxa from the Aegean and  
388 surrounding regions. *Journal of Zoological Systematics and Evolutionary Research* **53**: 18-32.
- 389 **Pritchard JK, Stephens M, Donnelly P. 2000.** Inference of population structure using multilocus  
390 genotype data. *Genetics* **155**: 945-959.
- 391 **Ramos-Onsins SE, Ferretti L, Raineri E, Jené J, Marmorini G, Burgo W, Vera G. unpublished.**  
392 *Mstatspop: statistical analysis using multiple populations for genomic data.*  
393 (<https://bioinformatics.cragenomica.es/numgenomics/people/sebas/software/software.html>).
- 394 **Riddle BR. 2016.** Comparative phylogeography clarifies the complexity and problems of continental  
395 distribution that drove A. R. Wallace to favor islands. *Proceedings of the National Academy of*  
396 *Sciences* **113**: 7970-7977.
- 397 **Schmitt T, Varga Z. 2012.** Extra-Mediterranean refugia: the rule and not the exception? *Frontiers in*  
398 *Zoology* **9**: 22.
- 399 **Speybroeck J, Beukema W, Bok B, Van Der Voort J. 2016.** *Field Guide to the Amphibians and*  
400 *Reptiles of Britain and Europe*. Bloomsbury Publishing.
- 401 **Stewart JR, Lister AM, Barnes I, Dalen L. 2010.** Refugia revisited: individualistic responses of  
402 species in space and time. *Proceedings of the Royal Society B-Biological Sciences* **277**: 661-  
403 671.
- 404 **Wakeley J. 2004.** Metapopulation models for historical inference. *Molecular Ecology* **13**: 865-875.
- 405 **Wakeley J, Aliacar N. 2001.** Gene Genealogies in a Metapopulation. *Genetics* **159**: 893-905.
- 406 **Wegmann D, Leuenberger C, Neuenchwander S, Excoffier L. 2010.** ABCtoolbox: a versatile  
407 toolkit for approximate Bayesian computations. *BMC Bioinformatics* **11**: 116.

- 408 **Wielstra B, Babik W, Arntzen JW. 2015.** The crested newt *Triturus cristatus* recolonized temperate  
 409 Eurasia from an extra-Mediterranean glacial refugium. *Biological Journal of the Linnean*  
 410 *Society* **114**: 574-587.
- 411 **Wielstra B, Burke T, Butlin RK, Avci A, Üzüm N, Bozkurt E, Olgun K, Arntzen JW. 2017.** A  
 412 genomic footprint of hybrid zone movement in crested newts. *Evolution Letters*:  
 413 10.1002/evl1003.1009.
- 414 **Wielstra B, Crnobrnja-Isailović J, Litvinchuk SN, Reijnen BT, Skidmore AK, Sotiropoulis K,**  
 415 **Toxopeus AG, Tzankov N, Vukov T, Arntzen JW. 2013.** Tracing glacial refugia of *Triturus*  
 416 newts based on mitochondrial DNA phylogeography and species distribution modeling.  
 417 *Frontiers in Zoology* **10**: 13.
- 418 **Wielstra B, Duijm E, Lagler P, Lammers Y, Meilink WRM, Ziermann JM, Arntzen JW. 2014.**  
 419 Parallel tagged amplicon sequencing of transcriptome-based genetic markers for *Triturus* newts  
 420 with the Ion Torrent next-generation sequencing platform. *Molecular Ecology Resources* **14**:  
 421 1080-1089.
- 422 **Zhang P, Papenfuss TJ, Wake MH, Qu LH, Wake DB. 2008.** Phylogeny and biogeography of the  
 423 family Salamandridae (Amphibia: Caudata) inferred from complete mitochondrial genomes.  
 424 *Molecular Phylogenetics and Evolution* **49**: 586-597.
- 425 **Zieliński P, Dudek K, Stuglik MT, Liana M, Babik W. 2014a.** Single nucleotide polymorphisms  
 426 reveal genetic structuring of the Carpathian newt and provide evidence of interspecific gene  
 427 flow in the nuclear genome. *PLoS ONE* **9**: e97431.
- 428 **Zieliński P, Nadachowska-Brzyska K, Dudek K, Babik W. 2016.** Divergence history of the  
 429 Carpathian and smooth newts modelled in space and time. *Molecular Ecology* **25**: 3912-3928.
- 430 **Zieliński P, Nadachowska-Brzyska K, Wielstra B, Szkotak R, Covaciu-Marcov S, Cogălniceanu**  
 431 **D, Babik W. 2013.** No evidence for nuclear introgression despite complete mtDNA  
 432 replacement in the Carpathian newt (*Lissotriton montandoni*). *Molecular Ecology* **22**: 1884-  
 433 1903.
- 434 **Zieliński P, Stuglik MT, Dudek K, Konczal M, Babik W. 2014b.** Development, validation and high  
 435 throughput analysis of sequence markers in non-model species. *Molecular Ecology Resources*  
 436 **14**: 352-360.

#### 437 **Supporting information**

439 **Appendix S1** Supplementary Tables S1-S10.

440 **Appendix S2** Supplementary Figures S1-6.

441

#### 442 **Data accessibility**

443 Sequence data and files associated with analyses are available from the Dryad Digital Repository entries

444 doi:10.5061/dryad.83k00 and [xxxToBeAddedxxx].

445

#### 446 **Figure legends**

447

448 **Figure 1.** Distribution of and Bayesian clustering results for *Triturus cristatus* and *Lissotriton*  
 449 *montandoni*. In (a) rough outlines of the ranges of both species are shown, with the range of *L.*  
 450 *montandoni* (in blue) superimposed on that of *T. cristatus* (in red). In (b) the preferred model for each

451 species in the approximate Bayesian computation analysis is shown. Codes for evolutionary lineages  
452 are explained in Results and colours correspond to the gene pools identified in the Structure analysis.  
453 In (c) pie diagrams represent the allocation by Structure of localities to different gene pools (k) for *T.*  
454 *cristatus* (k = 3; red tones) and *L. montandoni* (k = 2; blue tones) and pie diameter reflects sample size  
455 of localities (n = 1 or n = 3). Grey shading denotes elevation.