# Phenotypic Characterisation of *EIF2AK4* Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically with Pulmonary Arterial Hypertension

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#### **Abstract**

**Background**—Pulmonary arterial hypertension (PAH) is a rare disease with an emerging genetic basis. Heterozygous mutations in the gene encoding the bone morphogenetic protein receptor type 2 (*BMPR2*) are the commonest genetic cause of PAH, whereas biallelic mutations in the eukaryotic translation initiation factor 2 alpha kinase 4 gene (*EIF2AK4*) are described in pulmonary veno-occlusive disease and pulmonary capillary haemangiomatosis (PVOD/PCH). Here, we determined the frequency of these mutations and define the genotype-phenotype characteristics in a large cohort of patients diagnosed clinically with PAH.

*Methods*—Whole genome sequencing was performed on DNA from patients with idiopathic and heritable PAH, as well as PVOD/PCH recruited to the NIHR BioResource - Rare Diseases Study. Heterozygous variants in *BMPR2* and biallelic *EIF2AK4* variants with a minor allele frequency of < 1:10,000 in control data sets and predicted to be deleterious (by CADD, PolyPhen-2 and SIFT predictions) were identified as potentially causal. Phenotype data from the time of diagnosis were also captured.

**Results**—Eight hundred and sixty-four patients with idiopathic or heritable PAH and 16 with PVOD/PCH were recruited. Mutations in *BMPR2* were identified in 130 patients (14.8%). Biallelic mutations in *EIF2AK4* were identified in 5 patients with a clinical diagnosis of PVOD/PCH. Furthermore, 9 patients with a clinical diagnosis of PAH carried biallelic *EIF2AK4* mutations. These patients had a reduced transfer coefficient for carbon monoxide (KCO: 33 [IQR: 30 - 35] % predicted) and younger age at diagnosis (29 [23 - 38] years) as well as more interlobular septal thickening and mediastinal lymphadenopathy on computed tomography of the chest, compared to PAH patients without *EIF2AK4* mutations. However, radiological assessment alone could not accurately identify biallelic *EIF2AK4* mutation carriers. PAH patients with biallelic *EIF2AK4* mutations had a shorter survival.

**Conclusions**—Biallelic *EIF2AK4* mutations are found in patients classified clinically as idiopathic and heritable PAH. These patients cannot be identified reliably by CT, but a low KCO and a young age of diagnosis suggests the underlying molecular diagnosis. Genetic testing can identify these misclassified patients, allowing appropriate management and early referral for lung transplantation.

**Key Words:** genetics, human; pulmonary hypertension; prognosis; EIF2AK4, pulmonary veno-occlusive disease

#### **Clinical Perspective**

#### What is new?

- 1% of patients with a clinical diagnosis of PAH carry biallelic *EIF2AK4* mutations.
- Patients diagnosed clinically with PAH who had a KCO < 50% predicted and age of diagnosis < 50 years were more likely to carry biallelic *EIF2AK4* mutations. The diagnostic yield for genetic testing in this group was 53%.
- Radiological assessment was unable to distinguish reliably between these patients and idiopathic PAH patients.
- Histology from these patients may show predominately pulmonary arteriopathy, with subtle involvement of the pulmonary veins and capillaries.
- PAH patients with biallelic *EIF2AK4* mutations had a worse prognosis compared to other PAH patients.

#### What are the clinical implications?

- Younger patients diagnosed with idiopathic PAH, but with a low KCO, have a high frequency of biallelic *EIF2AK4* mutations.
- Such patients should be reclassified as pulmonary veno-occlusive disease/pulmonary capillary haemangiomatosis (PVOD/PCH).
- Similar to patients with PVOD/PCH these patients have a poor prognosis compared to other PAH patients.
- The spectrum of radiological and histological changes associated with biallelic *EIF2AK4* mutations is wider than previously assumed. The presence of only subtle or infrequent features associated with PVOD may lead to misclassification of these patients as PAH.
- Genetic testing allows early identification of these patients, facilitating appropriate management.

Pulmonary arterial hypertension (PAH) is a heterogeneous and rare disorder that can be classified into idiopathic and heritable forms, associated with an underlying condition, such as connective tissue disease or congenital heart disease, or related to specific drugs and toxins <sup>1, 2</sup>. In addition, pulmonary veno-occlusive disease (PVOD) and pulmonary capillary haemangiomatosis (PCH) are even rarer forms of pulmonary hypertension that are grouped together with PAH under the current classification system <sup>2</sup>.

Clinical features described in patients with PVOD/PCH include a low transfer coefficient for carbon monoxide (KCO) and oxygen desaturation on exertion, as well as the presence of centrilobular ground glass opacification, interlobular septal thickening and mediastinal lymphadenopathy on high resolution computed tomography (HRCT) of the lung parenchyma <sup>3, 4</sup>. However, these clinical and radiological features have also been reported in idiopathic PAH <sup>5-7</sup>. Consequently, the clinical distinction between PVOD/PCH and idiopathic PAH can be challenging. It has been estimated that 10% of patients with PVOD/PCH are misdiagnosed as idiopathic PAH <sup>8, 9</sup>. The diagnosis of PVOD/PCH is often only confirmed post mortem, or from explanted lungs, by histology.

The histological features of PVOD/PCH typically include pulmonary venous obstructions and pulmonary capillary proliferation, although the distribution of these changes within the lung can be heterogeneous <sup>10, 11</sup>. Pulmonary artery smooth muscle hypertrophy and intimal hyperplasia, similar to the changes observed in other forms of PAH, may also be present. Furthermore, pulmonary venous changes have been reported in cases of idiopathic PAH, scleroderma-associated PAH and those with *BMPR2* mutations, to varying extents <sup>12, 13</sup>.

A major advance in the molecular diagnosis of PVOD/PCH was the finding of biallelic mutations in the gene encoding the eukaryotic translation initiation factor 2 alpha kinase 4

(EIF2AK4) in both familial (100%) and sporadic (20-25%) cases of PVOD/PCH <sup>14, 15</sup>. EIF2AK4 is an activator of the integrated stress response (ISR) pathway, and responds to environmental stresses, including amino acid deprivation, by phosphorylating the alpha subunit of eukaryotic translation initiation factor 2 <sup>11, 16, 17</sup>. These discoveries suggest that *EIF2AK4* mutations are specific to PVOD/PCH and that finding biallelic *EIF2AK4* mutations in a patient with pulmonary hypertension would be diagnostic of PVOD/PCH. Patients with PVOD/PCH have a poor prognosis and risk fatal pulmonary oedema with the use of pulmonary artery vasodilator therapies <sup>4, 18-20</sup>. Consequently, early and accurate diagnosis is vital to guide clinical management.

Heterozygous mutations in the gene encoding the bone morphogenetic protein type 2 receptor (*BMPR2*) are the most common genetic cause of PAH. They are found in approximately 17% of individuals with idiopathic PAH and 82% with a family history of the disease <sup>21</sup>. However, mutations in *BMPR2* have also been reported in patients with histologically proven PVOD <sup>4, 22-24</sup>. Thus, there remains considerable uncertainty to what extent the finding of *EIF2AK4* or *BMPR2* mutations reliably predict the clinical phenotype and response to therapy in a population of patients with PAH.

Here we report the genetic and phenotypic characteristics of patients assessed for *BMPR2* and *EIF2AK4* mutations, through whole genome sequencing, within a large cohort (n=880) of PAH patients recruited to the National Institute of Health Research (NIHR) BioResource – Rare Diseases (BRIDGE) Study (Supplementary Table 1). The frequency of mutations in other previously reported genes associated with PAH will be reported in a future publication. In this study, we identified and characterised patients with a clinical and radiological diagnosis of idiopathic PAH who were found to possess biallelic *EIF2AK4* mutations. These patients had a

low KCO and were diagnosed at a younger age compared with idiopathic PAH patients without mutations in these genes. We show that, in common with patients diagnosed clinically with PVOD/PCH, PAH patients with biallelic *EIF2AK4* mutations have a shorter survival. We conclude that clinical assessment alone is inadequate for the accurate diagnosis of PVOD/PCH. Clinical genetic testing in younger patients presenting clinically with PAH but with a low KCO, will allow appropriate classification, leading to better risk stratification and management of these patients.

#### **Methods**

#### Ethical approval and consent

UK patients (621 [70.6%]) were recruited prospectively to the BRIDGE Study and provided written informed consent for genetic analysis and the capture of clinical data (NIHR BioResource - Rare Diseases Study 13/EE/0325). Additionally, the study included patients recruited retrospectively from non-UK centres (191 [21.7%]), and deceased UK patients (68 [7.7%]), if they had signed local tissue bank consent forms allowing genetic sequencing.

Explanted lung tissue from an individual undergoing lung transplantation for end stage PAH was

collected under Papworth Hospital Research Tissue Bank ethics (08/H0304/56).

#### **Recruitment and patients**

The BRIDGE Study is a prospective study recruiting both prevalent and incident patients with selected rare diseases. Recruitment to the BRIDGE PAH Study started in January 2013 and the last patient included in this analysis was recruited on 15/06/2016. Patients with idiopathic PAH, heritable PAH, PVOD and PCH, diagnosed according to international guidelines at specialist pulmonary hypertension centres in the United Kingdom, Netherlands and France, were recruited

(Figure 1 and Supplementary Table 2) <sup>2</sup>. This included 14 patients with confirmed mutations in *BMPR2*.

Throughout the manuscript, we classify patients recruited to the study as idiopathic PAH or familial PAH based on the absence or presence of a family history of the disease. The term heritable PAH does not distinguish between sporadic PAH patients with a mutation, and patients with a mutation where there is a family history. Therefore, the term "heritable PAH" is only used when referring to previous publications and guidelines.

Patients with other rare diseases and their unaffected relatives recruited to the BRIDGE Study (Supplementary Table 3) acted as non-PAH controls for the genetic analysis.

#### Whole genome sequencing and variant calling

Next generation sequencing using 100-150 base pair paired-end sequencing was performed on DNA libraries created from genomic DNA using Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, USA).

Reads were aligned against the Genome Reference Consortium human genome (build 37) (GRCh37) and variants were called using the Issac Aligner and Variant Caller respectively (version 2, Illumina Inc.). Variants in *BMPR2* and *EIF2AK4* were extracted and annotated using Ensembl's Variant Effect Predictor (VEP) v84 <sup>25</sup>. Deletions (resulting in the loss of more than 50bp) were identified by applying Isaac Copy Number Variant Caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina). Further information is provided in the supplemental materials.

Likely causal variants were identified based on minor allele frequency (MAF) and predicted deleteriousness. Variants were considered further if they had a MAF of less than 1 in 10,000 in unrelated non-PAH BRIDGE controls and the ExAC database <sup>26</sup>. The rare variants that

passed the MAF filtering were then assessed for deleteriousness. Variants were considered pathogenic based on a combined annotation dependent depletion (CADD) score of 15 or higher and PolyPhen-2 *or* SIFT predictions not classified as "benign" or "tolerated" respectively <sup>27-29</sup>.

#### **Over-representation analyses**

For comparison of variant frequencies between disease and control groups only variants from unrelated individuals were used. The PRIMUS software package was used to identify non-related individuals amongst both non-PAH BRIDGE controls and PAH patients <sup>30</sup>. The number of unrelated control subjects was maximised by including either patients with other rare diseases or their unaffected relatives. The frequency of rare and predicted deleterious heterozygous *EIF2AK4* variants in PAH index cases was also compared to publically available information in the ExAC database (http://exac.broadinstitute.org) <sup>26</sup>. This analysis provides the maximum estimate of the frequency of heterozygous *EIF2AK4* variants in the ExAC database as variants in ExAC were assumed not to be in a compound heterozygous state.

#### Phenotypic data capture and CT assessment

Paper and electronic patient records of PAH patients were reviewed to capture demographic and phenotypic variables from the time of diagnosis and follow up. Survival data for UK patients were obtained from recruiting centres through the NHS National Spine and local databases.

Anonymised information was captured securely online using the free OpenClinica® software, adapted for data capture specific to PAH.

CT images of the chest, where available, were reviewed independently by 2 cardiothoracic radiologists (AS and NS), with specialist imaging experience in pulmonary hypertension, blinded to the underlying diagnoses using a customised proforma. Further

information is provided in the supplemental materials, Supplementary Table 4 and Supplementary Table 5.

#### Statistical analysis

Statistical analysis was performed in R (<u>www.r-project.org</u>). Further information is provided in the supplemental materials.

Semi-parametric Cox-proportional hazard models were used to assess survival between groups using the "survival" package in R. Time from diagnosis to both death and death or transplantation was assessed. Age at diagnosis and gender were used as covariates in the models. To avoid immortal time bias arising from the inclusion of retrospectively recruited patients and prevalent patients, a sensitivity analysis was conducted. In this analysis only prospectively recruited patients from the UK were included and patients entered the risk set only from the time they consented to the study. Further information is provided in the supplemental materials.

#### Results

#### **Study patients**

Whole genome sequencing was performed on 932 patients recruited to the NIHR BRIDGE PAH Study and 7134 non-PAH control subjects recruited to other NIHR BRIDGE Study cohorts. Fifty-two patients were excluded from further analysis because they did not have a clinical diagnosis of idiopathic PAH, heritable PAH, PVOD or PCH (Figure 1). The remaining 880 patients (of which 872 were defined as unrelated index cases) consisted of 16 patients (1.8%) with a clinical diagnosis of PVOD/PCH, 56 (6.4%) with PAH and a family history of the disease (referred to as familial PAH) and 808 (91.8%) with idiopathic PAH and no known family

history. One of the 16 patients with a clinical diagnosis of PVOD/PCH had an affected sister, whereas the remainder had the sporadic form of the disease.

#### BMPR2 mutations in the PAH cohort

Rare and predicted deleterious *BMPR2* mutations (single nucleotide variants, indels and larger deletions) were found in 41 patients (73.2%) with familial PAH and 89 patients (11.0%) with idiopathic PAH. No *BMPR2* mutations were found in patients with a clinical diagnosis of PVOD/PCH.

#### Rare and predicted deleterious EIF2AK4 variants in the PAH cohort

Sixty-nine rare and predicted deleterious *EIF2AK4* single nucleotide variants and indels were present in the NIHR BRIDGE Study. No large deletions were found that affected the *EIF2AK4* gene locus. The variants are summarised in Supplementary Table 6. Five of the 16 patients (31.3%) with clinically diagnosed PVOD/PCH carried biallelic *EIF2AK4* mutations (2 homozygotes and 3 compound heterozygotes).

Twenty-five *EIF2AK4* variants were also found in 19 patients (2.2%) diagnosed clinically with PAH, in whom there was no clinical diagnosis of PVOD/PCH (5 homozygotes, 4 compound heterozygotes and 10 heterozygotes; Supplementary Table 7). One of these patients with a homozygous *EIF2AK4* mutation (c.3097C>T creating a premature stop codon) had a sister who had died of PAH. There was no reported family history of PVOD/PCH.

The remaining rare *EIF2AK4* variants were found in a heterozygous state in 36 control subjects (0.5%). Four of these variants appeared in more than 1 non-PAH control subject and none were shared with PAH patients.

## Over-representation of rare heterozygous EIF2AK4 variants in idiopathic PAH patients compared to control subjects

The proportion of patients with a clinical diagnosis of idiopathic PAH carrying heterozygous rare EIF2AK4 variants (1.2%) was significantly greater than the non-PAH control subjects (0.5%; p = 0.030). A similar over-representation in idiopathic PAH patients was observed when compared to allele frequencies in the ExAC database (0.6%; p = 0.042). Two idiopathic PAH patients with heterozygous rare EIF2AK4 variants also carried a rare and predicted deleterious BMPR2 mutation.

#### Phenotype of patients with a clinical diagnosis of PAH and biallelic EIF2AK4 mutations

Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations presented at a younger age (median [IQR]: 29 [23 - 38] years) compared to patients without these variants (51 [37 - 65] years; p = 0.024) (Table 1). Mean pulmonary artery pressure, cardiac output and pulmonary vascular resistance were not significantly different between PAH patients with biallelic *EIF2AK4* mutations and the other groups. As previously reported, haemodynamic variables were significantly more severe in patients with *BMPR2* mutations compared to those without any mutations in these genes.

The PAH patients with biallelic EIF2AK4 mutations exhibited a reduced KCO (33 [30 - 35] % predicted) compared to BMPR2 mutation carriers (81 [73 - 92] % predicted, p < 0.001) and PAH patients with no identified mutation (71 [51 - 85] % predicted, p = 0.001). PAH patients with biallelic EIF2AK4 mutations had no obstructive or restrictive deficit on spirometry. These differences remained after exclusion of patients with abnormal spirometry in the other groups (FEV<sub>1</sub> < 80% or FVC < 80%) (Supplementary Table 8).

Digital clubbing was over-represented amongst patients with biallelic *EIF2AK4* mutations diagnosed clinically with PAH (42%; p=0.002). Eleven percent of patients with a clinical diagnosis of PVOD were clubbed.

Only one patient with a heterozygous rare and predicted deleterious *EIF2AK4* variant (c.2516T>C) had a reduced KCO (54% predicted) with normal spirometry (FEV<sub>1</sub> 102% predicted, FVC 98% predicted and TLC 100% predicted). Although, there was mild paraseptal emphysema on thoracic CT (< 5% of the lung parenchyma affected). This patient, a 44-year-old Caucasian male diagnosed with idiopathic PAH, also carried a rare and deleterious *BMPR2* splice acceptor mutation (c.853-2A>G).

We questioned whether KCO was a predictor of biallelic *EIF2AK4* mutations in the wider cohort. However, amongst PAH patients with no mutations and normal spirometry (n=255), a reduced KCO (< 50% predicted) was present in 65 patients (25.5%). In these patients with a reduced KCO and preserved spirometry, 90.8% were aged over 50 years at diagnosis and 69.2% had a history of coronary artery disease, left ventricular dysfunction or cardiovascular risk factors (diabetes mellitus, systemic hypertension or hyperlipidaemia).

Given the high prevalence of a low KCO with preserved spirometry in the wider cohort, we restricted an analysis to patients under the age of 50 years, who at the time of diagnosis had normal spirometry (n=164). Even, in this group a significant proportion (15, 9.1%) had a KCO < 50% predicted (Figure 2). Eight of these 15 patients carried biallelic *EIF2AK4* mutations. One patient with biallelic *EIF2AK4* mutations was aged 70 years at diagnosis and subsequently did not meet this cut-off.

Amongst patients with normal spirometry, the presence of a KCO < 50% predicted and age at diagnosis < 50 years had a high sensitivity (0.889) and specificity (0.977) for identifying

patients who carry biallelic *EIF2AK4* mutations, the positive predictive value was low (0.533). Nevertheless, in terms of the diagnostic yield, while genetic testing for biallelic *EIF2AK4* mutations in the entire cohort of patients diagnosed clinically with PAH yielded a 1% detection rate, the presence of biallelic *EIF2AK4* mutations in PAH patients with a KCO < 50% with normal spirometry and aged under 50 at diagnosis was 53%.

#### CT features of EIF2AK4 mutation carriers

Centrilobular ground glass opacification extent, mediastinal lymphadenopathy and interlobular septal thickening are considered suggestive of PVOD/PCH. However, we found subtle or gross centrilobular ground glass opacification in 38% of patients diagnosed clinically with PAH and carrying no mutations (n=21) and 67% of PAH patients with *BMPR2* mutations (n=21). This was not significantly different compared to patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations (86%, n=7) and patients with a clinical diagnosis of PVOD (50%, n=14). Gross interlobular septal thickening and mediastinal lymphadenopathy was significantly more frequent amongst patients with PAH and biallelic *EIF2AK4* mutations (29% and 57% respectively) and those with PVOD (64% and 79%) compared to patients with PAH and no mutation (5% and 0%) or *BMPR2* mutations (5% and 10%). A radiological suspicion of PVOD/PCH was raised in 71% of those with PVOD, 57% of patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations, 14% of PAH patients with no mutation, and 5% of those with *BMPR2* mutations (Table 2).

A further CT analysis comparing patients with biallelic *EIF2AK4* mutations (with a clinical diagnosis of PVOD/PCH or PAH; n=11) and those with a clinical diagnosis of PVOD but not carrying biallelic *EIF2AK4* mutations (n=10) was made (Supplementary Table 9).

Patients with biallelic *EIF2AK4* mutations were younger at diagnosis (27 [IQR: 23 - 34] years)

compared to those with PVOD and no *EIF2AK4* mutations (68 [64 - 72] years, p=0.001). The patients with biallelic *EIF2AK4* mutations also had a lower KCO (32 [29 – 33] % predicted) compared to patients with PVOD and no *EIF2AK4* mutations (41.4 [37 – 54] % predicted, p=0.013). Centrilobular ground glass opacification appeared more extensive in those with biallelic *EIF2AK4* mutations (82%) compared to those without a mutation (10%; p=0.012). However, pleural effusions were more common amongst those without a mutation (40%) compared to patients with biallelic *EIF2AK4* mutations (0%, p=0.035). This may suggest that patients with biallelic *EIF2AK4* mutations have a distinct radiological phenotype compared to patients with PVOD and no biallelic *EIF2AK4* mutations.

#### Response to pulmonary artery vasodilator therapies

The response to pulmonary artery vasodilator therapies at 1 and 3 years was assessed for patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations as well as the other PAH patients included in the CT analysis. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations did not improve their functional class at either 1 year or 3 years post diagnosis unlike the other PAH groups (Supplementary Table 10).

#### Histological features of biallelic EIF2AK4 mutation carrier

The explanted lungs of one patient diagnosed with idiopathic PAH but found to have a homozygous *EIF2AK4* missense mutation (c.1795G>C, p.G599R) were assessed. The predominant histological feature was pulmonary arterial vasculopathy. The pulmonary arteries predominantly showed concentric and eccentric intimal fibrosis. No plexiform lesions were observed. Although infrequent, there was some fibrosis of the septal veins and venules, some of which were nearly completely occluded. Although there was evidence of capillary congestion, no capillary hemangiomatosis was observed (Figure 3). The missense variant carried by this patient

was not reported in the ExAC database, occurs in a conserved area of the genome (GERP score 5.5) and was predicted to be deleterious (CADD score 32, PolyPhen-2 prediction of "probably damaging [1]", SIFT prediction of "deleterious [0]"). The same homozygous mutation was also found in a second unrelated patient with a clinical diagnosis of idiopathic PAH.

#### Impact of genotype on survival

Eight hundred and fifty-eight patients were included in the Cox proportional hazards model (Supplementary Table 11, Supplemental Figure 1). Patients diagnosed clinically as PAH with biallelic EIF2AK4 mutations had a shorter survival time from diagnosis compared to the BMPR2 mutation carriers (p < 0.001) and those without any variants in PAH associated genes (p < 0.001). Age (p < 0.001) and gender (p = 0.001) also had a significant effect on survival, with male sex and an older age at diagnosis associated with shorter survival in the model. Similar results were obtained when assessing the time to death or transplantation (Supplementary Tables 12). In the sensitivity analysis, including only prospectively recruited UK patients, only 2 events occurred in the biallelic EIF2AK4 group. Thus no significant difference was observed in mortality between patients diagnosed clinically as PAH with biallelic EIF2AK4 mutations and patients with BMPR2 mutations (p = 0.215), or patients without any variants in PAH associated genes (p = 0.282; Supplementary Table 13).

#### **Discussion**

This is the first study to analyse the frequency of *EIF2AK4* rare variation in a large cohort of PAH patients and make detailed phenotypic and radiological assessments. Previously the presence of biallelic *EIF2AK4* mutations were reported in patients with a clear clinical diagnosis of PVOD/PCH as well as a large kindred and a single family with a possible diagnosis of PAH <sup>20</sup>,

<sup>31, 32</sup>. As expected, we identified a high frequency of biallelic *EIF2AK4* mutations in patients with a clear clinical presentation of PVOD/PCH. However, we also found biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH.

The discovery of biallelic *EIF2AK4* mutations in PVOD/PCH raised the possibility of rapid molecular diagnosis in the majority of patients with familial, and up to 25% of patients with sporadic PVOD/PCH <sup>14, 15</sup>. In the present study, the presence of biallelic *EIF2AK4* mutations was associated with a poor prognosis, even in patients who have a clinical diagnosis of PAH, and who did not develop pulmonary oedema in response to pulmonary artery vasodilator therapies. Therefore, early identification of these patients through genetic testing may prompt early referral for lung transplantation similar to patients with clinically diagnosed PVOD/PCH <sup>18</sup>.

The presence of biallelic *EIF2AK4* mutations in patients with a clinical diagnosis of PAH raises the question whether *EIF2AK4* mutations can cause classical idiopathic PAH, or whether there are cases of PVOD/PCH caused by *EIF2AK4* mutations that are wrongly classified even by expert centres. We further show that phenotypic, radiological and histological assessments can be difficult to interpret. The presence of subtle or infrequent features may lead to an incorrect diagnosis of PAH in patients with biallelic *EIF2AK4* mutations. This study suggests that patients with pathogenic biallelic *EIF2AK4* mutations may present with a spectrum of phenotypic, radiological and histological features that can overlap with PAH.

PAH patients with biallelic *EIF2AK4* mutations demonstrated a reduced KCO despite normal spirometry, which is characteristic of patients with PVOD/PCH. The reduced KCO likely reflects widespread reduction in alveolar gas exchange due to endothelial proliferation and patchy thickening of the blood gas barrier by the process of capillary haemangiomatosis.

Ultrastructural thickening of the capillary basement membrane may also play a role <sup>33</sup>. In

keeping with previous reports in PVOD/PCH we also show that PAH patients with biallelic mutations in *EIF2AK4* are younger at diagnosis than patients with either *BMPR2* mutations or no known mutation <sup>14, 20</sup>. However, the presence of these characteristic features has a low positive predictive value for the identification of patients with biallelic *EIF2AK4* mutations.

In contrast to previous descriptions of patients with PVOD, none of the patients with clinically diagnosed PAH and biallelic *EIF2AK4* mutations developed pulmonary oedema in response to pulmonary artery vasodilator therapies. For example, intravenous prostanoids were used in 50% of these patients. In classical PVOD patients, pulmonary oedema with intravenous prostanoids has been reported in up to 44% of patients after a median treatment duration of just 9 days <sup>4</sup>. Presumably the extent and severity of the pulmonary venous involvement in these patients might underlie the differing responses to prostanoids.

It is generally considered that HRCT imaging is a useful non-invasive test to assist in the diagnosis of suspected PVOD/PCH <sup>11</sup>. Although there was an increased prevalence of mediastinal lymphadenopathy and interlobular septal thickening in PAH patients with biallelic *EIF2AK4* mutations, we found that radiological features at the time of diagnosis could not accurately determine the underlying genotype <sup>6</sup>. The differing radiological features of all patients with biallelic *EIF2AK4* mutations compared with PVOD cases without mutations is of interest. This may reflect differences between the younger onset genetic cases of PVOD, compared with the predominantly older group of patients without *EIF2AK4* mutations in whom other nongenetic factors, such as exposure to inorganic solvents, may play an important role <sup>34</sup>.

Histological examination (usually post mortem or from explanted lungs) is often considered essential for diagnostic confirmation of PVOD/PCH but may be confounded by the heterogeneous nature of vascular pathology <sup>35</sup>. Surgical biopsy of the lung in patients with severe

PAH is contraindicated and a limitation of this study is that lung tissue from only one patient with biallelic *EIF2AK4* mutations was available for analysis. This patient had a rare and predicted deleterious homozygous missense mutation in *EIF2AK4*. The predominant feature on assessment of the explanted lung tissue was of pulmonary arteriopathy, as usually seen in PAH. Although only infrequent, fibrosis of the septal venules and the possible presence of siderophages in the alveolar space were observed. These features are found in patients with PVOD/PCH. This case supports the hypothesis that patients with biallelic *EIF2AK4* mutations may present with a spectrum of venous and arterial involvement.

There are increasing reports of phenotypic, radiological and histological similarities between PAH and PVOD/PCH <sup>6, 12, 13</sup>. Tenorio et al. reported a homozygous missense mutation in *EIF2AK4* in a large kindred of Iberian Romani with apparent heritable PAH <sup>31</sup>. This kindred is likely to have PVOD/PCH as these diagnoses were not confirmed histologically and PVOD was suspected in half the patients. More recently, Best et al. also report two sisters with apparent heritable PAH carrying biallelic *EIF2AK4* mutations <sup>32</sup>. These patients also had a reduced KCO but had not had HRCT assessment of their lung parenchyma which may have altered their clinical diagnosis. Taken together, these previous reports are compatible with the findings in this larger cohort, that patients with a clinical presentation of idiopathic or heritable PAH may in fact have underlying PVOD/PCH as determined by genetic analysis.

A strength of this study is the centralised reporting of radiographic features. However, the data collection was retrospective and incomplete in some cases. Assessing rare diseases, such as PAH and PVOD/PCH, with a prospective study recruiting incident cases would take a prohibitively long time. This is especially true when assessing survival and response to therapy. In this study including prevalent and retrospectively recruited patients, we demonstrated a worse

prognosis in patients with a clinical diagnosis of PAH and biallelic *EIF2AK4*. However, the inclusion of prevalent and retrospectively recruited patients can introduce bias such as immortal time bias, when there are long periods between diagnosis and enrolment in the study. The effect of immortal time bias and other confouders such as the inclusion of prevalent and incident cases can be difficult to predict. In all groups there are likely to be patients who died prior to study enrolment, and thus would not feature in any analysis. When we attempted to eliminate these sources of bias in a sensitivity analysis restricted to prospectively recruited patients from the UK, the study did not have sufficient power to show a difference in survival between different genotypes. Further studies of survival and response to therapy will be needed to definitively show whether "misclassified" PAH patients with biallelic *EIF2AK4* mutations have a similarly poor prognosis as classical PVOD patients with these mutations.

The genetic architecture of idiopathic and heritable PAH remains to be fully elucidated. Ongoing analysis of whole genome sequence data in our cohort is likely to reveal novel rare variation underlying this condition. Mutations in *BMPR2* account for approximately 17% of idiopathic PAH patients and other known PAH genes account for approximately 1-2% of all cases <sup>21, 36</sup>. In the present study *BMPR2* mutations were found in 11% of patients without a family history of PAH. It is worth noting that patients with the sporadic form of the disease with no reported family history represent a higher burden of *BMPR2* mutations (n=89) compared to those with a family history (n=49). This has important implications for clinical genetic testing in patients with sporadic as well as familial disease.

In previous studies mutations in both *EIF2AK4* alleles are required to cause PVOD and PCH <sup>14, 15</sup>. In autosomal recessive disorders, it is unusual for the heterozygous state to manifest the disease phenotype and thus heterozygous *EIF2AK4* variants would not be expected to be

pathogenic. In this study, we found a significant over-representation of heterozygous rare and predicted deleterious *EIF2AK4* variants in PAH compared to control subjects and report 2 patients with rare variants in both *BMPR2* and *EIF2AK4*. Recently, the possibility that heterozygous *EIF2AK4* variants influence the penetrance of *BMPR2* mutations has been raised in a single family with PAH <sup>37</sup>. Further studies are required to determine whether heterozygous *EIF2AK4* variants contribute to aetiology in PAH.

In summary, we demonstrate that biallelic *EIF2AK4* mutations are found in patients diagnosed clinically with idiopathic and familial PAH. These patients may have subtle features suggestive of PVOD/PCH on close inspection and are likely to have underlying PVOD/PCH. The spectrum of phenotypic, radiological and histological features found in patients with biallelic *EIF2AK4* mutations made by current clinical assessments is wider and less clear cut than previously recognised. This may lead to misclassification of patients as PAH rather than PVOD and hinders accurate risk stratification. Ascertaining the *EIF2AK4* mutation status of patients through clinical genetic testing provides additional information to aid risk stratification and guide management. In a young patient presenting with apparent PAH, the presence of a low KCO with normal spirometry strongly suggests the presence of underlying biallelic *EIF2AK4* mutations. Patients with an apparent clinical diagnosis of PAH and biallelic *EIF2AK4* mutations have a worse prognosis compared to patients with *BMPR2* mutations and those without these mutations. Clinical genetic testing should aid identification of this high-risk group and facilitate early referral for lung transplantation and appropriate management.

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#### References

- 1. Simonneau G, Gatzoulis MA, Adatia I, Celermajer D, Denton C, Ghofrani A, Gomez Sanchez MA, Krishna Kumar R, Landzberg M, Machado RF, Olschewski H, Robbins IM and Souza R. Updated clinical classification of pulmonary hypertension. *J Am Coll Cardiol*. 2013;62:D34-41.
- 2. Galie N, Humbert M, Vachiery JL, Gibbs S, Lang I, Torbicki A, Simonneau G, Peacock A, Vonk Noordegraaf A, Beghetti M, Ghofrani A, Gomez Sanchez MA, Hansmann G, Klepetko W, Lancellotti P, Matucci M, McDonagh T, Pierard LA, Trindade PT, Zompatori M and Hoeper M. 2015 ESC/ERS Guidelines for the diagnosis and treatment of pulmonary hypertension: The Joint Task Force for the Diagnosis and Treatment of Pulmonary Hypertension of the European Society of Cardiology (ESC) and the European Respiratory Society (ERS): Endorsed by: Association for European Paediatric and Congenital Cardiology (AEPC), International Society for Heart and Lung Transplantation (ISHLT). *Eur Respir J*. 2015;46:903-75.
- 3. Resten A, Maitre S, Humbert M, Rabiller A, Sitbon O, Capron F, Simonneau G and Musset D. Pulmonary hypertension: CT of the chest in pulmonary venoocclusive disease. *AJR Am J Roentgenol*. 2004;183:65-70.
- 4. Montani D, Achouh L, Dorfmuller P, Le Pavec J, Sztrymf B, Tcherakian C, Rabiller A, Haque R, Sitbon O, Jais X, Dartevelle P, Maitre S, Capron F, Musset D, Simonneau G and Humbert M. Pulmonary veno-occlusive disease: clinical, functional, radiologic, and hemodynamic characteristics and outcome of 24 cases confirmed by histology. *Medicine* (*Baltimore*). 2008;87:220-33.
- 5. Trip P, Girerd B, Bogaard HJ, de Man FS, Boonstra A, Garcia G, Humbert M, Montani D and Vonk-Noordegraaf A. Diffusion capacity and BMPR2 mutations in pulmonary arterial hypertension. *Eur Respir J.* 2014;43:1195-8.
- 6. Rajaram S, Swift AJ, Condliffe R, Johns C, Elliot CA, Hill C, Davies C, Hurdman J, Sabroe I, Wild JM and Kiely DG. CT features of pulmonary arterial hypertension and its major subtypes: a systematic CT evaluation of 292 patients from the ASPIRE Registry. *Thorax*. 2015;70:382-7.

- 7. Trip P, Nossent EJ, de Man FS, van den Berk IA, Boonstra A, Groepenhoff H, Leter EM, Westerhof N, Grunberg K, Bogaard HJ and Vonk-Noordegraaf A. Severely reduced diffusion capacity in idiopathic pulmonary arterial hypertension: patient characteristics and treatment responses. *Eur Respir J.* 2013;42:1575-85.
- 8. Mandel J, Mark EJ and Hales CA. Pulmonary veno-occlusive disease. *Am J Respir Crit Care Med*. 2000;162:1964-73.
- 9. Pietra GG, Edwards WD, Kay JM, Rich S, Kernis J, Schloo B, Ayres SM, Bergofsky EH, Brundage BH and Detre KM. Histopathology of primary pulmonary hypertension. A qualitative and quantitative study of pulmonary blood vessels from 58 patients in the National Heart, Lung, and Blood Institute, Primary Pulmonary Hypertension Registry. *Circulation*. 1989;80:1198-206.
- 10. Lantuejoul S, Sheppard MN, Corrin B, Burke MM and Nicholson AG. Pulmonary veno-occlusive disease and pulmonary capillary hemangiomatosis: a clinicopathologic study of 35 cases. *Am J Surg Pathol*. 2006;30:850-7.
- 11. Montani D, Lau EM, Dorfmuller P, Girerd B, Jais X, Savale L, Perros F, Nossent E, Garcia G, Parent F, Fadel E, Soubrier F, Sitbon O, Simonneau G and Humbert M. Pulmonary veno-occlusive disease. *Eur Respir J.* 2016;47:1518-34.
- 12. Dorfmuller P, Humbert M, Perros F, Sanchez O, Simonneau G, Muller KM and Capron F. Fibrous remodeling of the pulmonary venous system in pulmonary arterial hypertension associated with connective tissue diseases. *Hum Pathol.* 2007;38:893-902.
- 13. Ghigna MR, Guignabert C, Montani D, Girerd B, Jais X, Savale L, Herve P, Thomas de Montpreville V, Mercier O, Sitbon O, Soubrier F, Fadel E, Simonneau G, Humbert M and Dorfmuller P. BMPR2 mutation status influences bronchial vascular changes in pulmonary arterial hypertension. *Eur Respir J.* 2016;48:1668-1681.
- 14. Eyries M, Montani D, Girerd B, Perret C, Leroy A, Lonjou C, Chelghoum N, Coulet F, Bonnet D, Dorfmuller P, Fadel E, Sitbon O, Simonneau G, Tregouet DA, Humbert M and Soubrier F. EIF2AK4 mutations cause pulmonary veno-occlusive disease, a recessive form of pulmonary hypertension. *Nat Genet*. 2014;46:65-9.
- 15. Best DH, Sumner KL, Austin ED, Chung WK, Brown LM, Borczuk AC, Rosenzweig EB, Bayrak-Toydemir P, Mao R, Cahill BC, Tazelaar HD, Leslie KO, Hemnes AR, Robbins IM and Elliott CG. EIF2AK4 mutations in pulmonary capillary hemangiomatosis. *Chest*. 2014;145:231-6.
- 16. Dever TE, Feng L, Wek RC, Cigan AM, Donahue TF and Hinnebusch AG. Phosphorylation of initiation factor 2 alpha by protein kinase GCN2 mediates gene-specific translational control of GCN4 in yeast. *Cell*. 1992;68:585-96.
- 17. Harding HP, Zhang Y, Zeng H, Novoa I, Lu PD, Calfon M, Sadri N, Yun C, Popko B, Paules R, Stojdl DF, Bell JC, Hettmann T, Leiden JM and Ron D. An integrated stress response regulates amino acid metabolism and resistance to oxidative stress. *Mol Cell*. 2003;11:619-33.
- 18. Wille KM, Sharma NS, Kulkarni T, Lammi MR, Barney JB, Bellot SC, Cantor RS, Naftel DC, Diaz-Guzman E and McGiffin DC. Characteristics of patients with pulmonary venoocclusive disease awaiting transplantation. *Ann Am Thorac Soc.* 2014;11:1411-8.
- 19. Palmer SM, Robinson LJ, Wang A, Gossage JR, Bashore T and Tapson VF. Massive pulmonary edema and death after prostacyclin infusion in a patient with pulmonary veno-occlusive disease. *Chest.* 1998;113:237-40.
- 20. Montani D, Girerd B, Jais X, Levy M, Amar D, Savale L, Dorfmuller P, Seferian A, Lau EM, Eyries M, Le Pavec J, Parent F, Bonnet D, Soubrier F, Fadel E, Sitbon O, Simonneau G and

- Humbert M. Clinical phenotypes and outcomes of heritable and sporadic pulmonary veno-occlusive disease: a population-based study. *Lancet Respir Med.* 2017;5:125-134.
- 21. Evans JD, Girerd B, Montani D, Wang XJ, Galie N, Austin ED, Elliott G, Asano K, Grunig E, Yan Y, Jing ZC, Manes A, Palazzini M, Wheeler LA, Nakayama I, Satoh T, Eichstaedt C, Hinderhofer K, Wolf M, Rosenzweig EB, Chung WK, Soubrier F, Simonneau G, Sitbon O, Graf S, Kaptoge S, Di Angelantonio E, Humbert M and Morrell NW. BMPR2 mutations and survival in pulmonary arterial hypertension: an individual participant data meta-analysis. *Lancet Respir Med*. 2016;4:129-37.
- 22. Runo JR, Vnencak-Jones CL, Prince M, Loyd JE, Wheeler L, Robbins IM, Lane KB, Newman JH, Johnson J, Nichols WC and Phillips JA, 3rd. Pulmonary veno-occlusive disease caused by an inherited mutation in bone morphogenetic protein receptor II. *Am J Respir Crit Care Med*. 2003;167:889-94.
- 23. Aldred MA, Vijayakrishnan J, James V, Soubrier F, Gomez-Sanchez MA, Martensson G, Galie N, Manes A, Corris P, Simonneau G, Humbert M, Morrell NW and Trembath RC. BMPR2 gene rearrangements account for a significant proportion of mutations in familial and idiopathic pulmonary arterial hypertension. *Hum Mutat*. 2006;27:212-3.
- 24. Machado RD, Aldred MA, James V, Harrison RE, Patel B, Schwalbe EC, Gruenig E, Janssen B, Koehler R, Seeger W, Eickelberg O, Olschewski H, Elliott CG, Glissmeyer E, Carlquist J, Kim M, Torbicki A, Fijalkowska A, Szewczyk G, Parma J, Abramowicz MJ, Galie N, Morisaki H, Kyotani S, Nakanishi N, Morisaki T, Humbert M, Simonneau G, Sitbon O, Soubrier F, Coulet F, Morrell NW and Trembath RC. Mutations of the TGF-beta type II receptor BMPR2 in pulmonary arterial hypertension. *Hum Mutat*. 2006;27:121-32.
- 25. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P and Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol.* 2016;17:122.
- 26. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB, Tukiainen T, Birnbaum DP, Kosmicki JA, Duncan LE, Estrada K, Zhao F, Zou J, Pierce-Hoffman E, Berghout J, Cooper DN, Deflaux N, DePristo M, Do R, Flannick J, Fromer M, Gauthier L, Goldstein J, Gupta N, Howrigan D, Kiezun A, Kurki MI, Moonshine AL, Natarajan P, Orozco L, Peloso GM, Poplin R, Rivas MA, Ruano-Rubio V, Rose SA, Ruderfer DM, Shakir K, Stenson PD, Stevens C, Thomas BP, Tiao G, Tusie-Luna MT, Weisburd B, Won HH, Yu D, Altshuler DM, Ardissino D, Boehnke M, Danesh J, Donnelly S, Elosua R, Florez JC, Gabriel SB, Getz G, Glatt SJ, Hultman CM, Kathiresan S, Laakso M, McCarroll S, McCarthy MI, McGovern D, McPherson R, Neale BM, Palotie A, Purcell SM, Saleheen D, Scharf JM, Sklar P, Sullivan PF, Tuomilehto J, Tsuang MT, Watkins HC, Wilson JG, Daly MJ and MacArthur DG. Analysis of protein-coding genetic variation in 60,706 humans. *Nature*. 2016;536:285-91.
- 27. Adzhubei IA, Schmidt S, Peshkin L, Ramensky VE, Gerasimova A, Bork P, Kondrashov AS and Sunyaev SR. A method and server for predicting damaging missense mutations. *Nat Methods*. 2010;7:248-9.
- 28. Ng PC and Henikoff S. Predicting deleterious amino acid substitutions. *Genome Res.* 2001;11:863-74.
- 29. Kircher M, Witten DM, Jain P, O'Roak BJ, Cooper GM and Shendure J. A general framework for estimating the relative pathogenicity of human genetic variants. *Nat Genet*. 2014;46:310-5.

- 30. Staples J, Qiao D, Cho M, Silverman E, Nickerson D and Below J. PRIMUS: Rapid Reconstruction of Pedigrees from Genome-wide Estimates of Identity by Descent. *Am J Hum Genet*. 2014;95:553-64.
- 31. Tenorio J, Navas P, Barrios E, Fernandez L, Nevado J, Quezada CA, Lopez-Meseguer M, Arias P, Mena R, Lobo JL, Alvarez C, Heath K, Escribano-Subias P and Lapunzina P. A founder EIF2AK4 mutation causes an aggressive form of pulmonary arterial hypertension in Iberian Gypsies. *Clin Genet*. 2015;88:579-83.
- 32. Best DH, Sumner KL, Smith BP, Damjanovich-Colmenares K, Nakayama I, Brown LM, Ha Y, Paul E, Morris A, Jama MA, Dodson MW, Bayrak-Toydemir P and Elliott CG. EIF2AK4 Mutations in Patients Diagnosed with Pulmonary Arterial Hypertension. *Chest.* 2016.
- 33. Villaschi S and Pietra GG. Alveolo-capillary membrane in primary pulmonary hypertension. *Appl Pathol.* 1986;4:132-7.
- 34. Montani D, Lau EM, Descatha A, Jais X, Savale L, Andujar P, Bensefa-Colas L, Girerd B, Zendah I, Le Pavec J, Seferian A, Perros F, Dorfmuller P, Fadel E, Soubrier F, Sitbon O, Simonneau G and Humbert M. Occupational exposure to organic solvents: a risk factor for pulmonary veno-occlusive disease. *Eur Respir J.* 2015;46:1721-31.
- 35. Pietra GG, Capron F, Stewart S, Leone O, Humbert M, Robbins IM, Reid LM and Tuder RM. Pathologic assessment of vasculopathies in pulmonary hypertension. *J Am Coll Cardiol*. 2004;43:25s-32s.
- 36. Machado RD, Southgate L, Eichstaedt CA, Aldred MA, Austin ED, Best DH, Chung WK, Benjamin N, Elliott CG, Eyries M, Fischer C, Graf S, Hinderhofer K, Humbert M, Keiles SB, Loyd JE, Morrell NW, Newman JH, Soubrier F, Trembath RC, Viales RR and Grunig E. Pulmonary Arterial Hypertension: A Current Perspective on Established and Emerging Molecular Genetic Defects. *Hum Mutat*. 2015;36:1113-27.
- 37. Eichstaedt CA, Song J, Benjamin N, Harutyunova S, Fischer C, Grunig E and Hinderhofer K. EIF2AK4 mutation as "second hit" in hereditary pulmonary arterial hypertension. *Respir Res.* 2016;17:141.

**Table 1.** Phenotypic summary of *EIF2AK4* variant carriers. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations are younger at diagnosis and have a significantly reduced KCO compared to other groups.

	RMPR2 mutations *	mutations in PAH associated genes	heterozygous	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	p
n	130	704	8	9	16	
Age (years)	39 [31 - 52]	51 [37 - 65]	49 [36 - 67]	29 [23 - 38]	57 [41 - 69]	< 0.001
Gender (n female [%])	85 [65.4%]	494 [70.2%]	7 [87.5%]	4 [44.4%]	9 [56.2%]	0.18
Ethnicity (n white Caucasian [%])	108 [83.1%]	551 [78.5%]	5 [62.5%]	2 [22.2%]	13 [81.2%]	0.002
Digital clubbing (n [%])	6 [9.7%]	10 [3.4%]	0 [0%]	3 [42.9%]	1 [11.1%]	0.002
BMI	28 [24 - 33]	28 [24 - 33]	26 [23 - 28]	24 [20 - 27]	27 [24 - 31]	0.216
mPAP (mmHg)	57 [51 - 69]	52 [44 - 61]	44 [42 - 52]	52 [46 - 65]	48 [40 - 58]	< 0.001
CO (L/min)	3 [3 - 4]	4 [3 - 5]	3 [3 - 5]	5 [3 - 6]	4 [3 - 4]	< 0.001
PVR (WU)	15 [11 - 20]	10 [7 - 14]	9 [6 - 10]	9 [8 - 13]	10 [9 - 12]	< 0.001
Vasoresponders (n [%])	0 [0%]	28 [17.5%]	0 [0%]	0 [0%]	0 [0%]	0.011
FEV <sub>1</sub> (%pred)	90 [78 - 99]	84 [72 - 95]	83 [71 - 94]	94 [85 - 100]	85 [70 - 95]	0.031
FVC (%pred)	97 [86 - 109]	95 [82 - 106]	96 [75 - 98]	100 [86 - 119]	97 [81 - 103]	0.310
KCO (%pred)	81 [73 - 92]	71 [51 - 85]	81 [72 - 95]	33 [30 - 35]	37 [32 - 47]	< 0.001
Resting S <sub>A</sub> O <sub>2</sub> (%)	96 [94 - 97]	96 [93 - 97]	98 [98 - 98]	91 [90 - 94]	94 [91 - 95]	0.010
S <sub>A</sub> O <sub>2</sub> post walk test (%)	94 [90 - 97]	92 [85 - 96]	94 [84 - 96]	78 [75 - 82]	88 [85 - 89]	< 0.001

BMI - body mass index, mPAP - mean pulmonary artery pressure, PVR - pulmonary vascular resistance, FEV1 - forced expiratory volume in 1 second, FVC - forced vital capacity, KCO - transfer coefficient for carbon monoxide. \* Also includes the 2 patients with a heterozygous EIF2AK4 variant and a BMPR2 variant. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

Table 2. Radiological features and consensus radiological diagnosis of PAH patients in the CT substudy

	Group	PAH patients with BMPR2 mutations	PAH patients with no mutations in the previously reported PAH genes	PAH patients with heterozygous EIF2AK4 variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD	р
	n	21	21	4	7	14	
Contribute to a second of the	None	7 [33.3%]	13 [61.9%]	2 [50.0%]	1 [14.3%]	7 [50.0%]	0.122
Centrilobular ground glass	Subtle	12 [57.1%]	5 [23.8%]	0 [0.0%]	2 [28.6%]	3 [21.4%]	
opacification density	Present	2 [9.5%]	3 [14.3%]	2 [50.0%]	4 [57.1%]	4 [28.6%]	
	None	8 [38.1%]	13 [61.9%]	2 [50.0%]	1 [4.3%]	8 [57.1%]	0.077
	<5%	0 [0.0%]	3 [14.3%]	0 [0.0%]	1 [14.3%]	1 [7.1%]	
Centrilobular ground glass	5-25%	2 [9.5%]	0 [0.0%]	1 [25.0%]	2 [28.6%]	1 [7.1%]	
opacification extent	25-50%	2 [9.5%]	4 [19.0%]	0 [0.0%]	0 [0.0%]	2 [14.3%]	
1	50-75%	5 [23.8%]	1 [4.8%]	0 [0.0%]	2 [28.6%]	0 [0.0%]	
	75-100%	4 [19.0%]	0 [0.0%]	1 [25.0%]	1 [14.3%]	2 [14.3%]	
Interdahadan santal	None	17 [81.0%]	18 [85.7%]	4 [100.0%]	5 [71.4%]	4 [28.6%]	0.001
Interlobular septal	Subtle	3 [14.3%]	2 [9.5%]	0 [0.0%]	0 [0.0%]	1 [7.1%]	
thickening	Present	1 [4.8%]	1 [4.8%]	0 [0.0%]	2 [28.6%]	9 [64.3%]	
Mediastinal	None	19 [90.5%]	21 [100.0%]	4 [100.0%]	3 [42.9%]	3 [21.4%]	ر د0 001
lymphadenopathy	Present	2 [9.5%]	0 [0.0%]	0 [0.0%]	4 [57.1%]	11 [78.6%]	< 0.001
Diamed official	None	17 [81.0%]	21 [100.0%]	3 [75.0%]	7 [100.0%]	10 [71.4%]	0.048
Pleural effusion	Small	4 [19.0%]	0 [0.0%]	1 [25.0%]	9 [0.0%]	4 [28.6%]	
Negrocaylarity	None	12 [57.1%]	18 [85.7%]	4 [100.0%]	6 [85.7%]	13 [92.9%]	0.077
Neovascularity	Present	9 [42.9%]	3 [14.3%]	0 [0.0%]	1 [14.3%]	1 [7.1%]	0.077
	PAH	20 [95.2%]	18 [85.7%]	3 [75.0%]	3 [42.9%]	4 [28.6%]	
CT diagnosis	Possible PVOD/PCH	1 [4.8%]	3 [14.3%]	1 [25.0%]	4 [57.1%]	10 [71.4%]	

Data presented as n [%].

#### **Figure Legends**

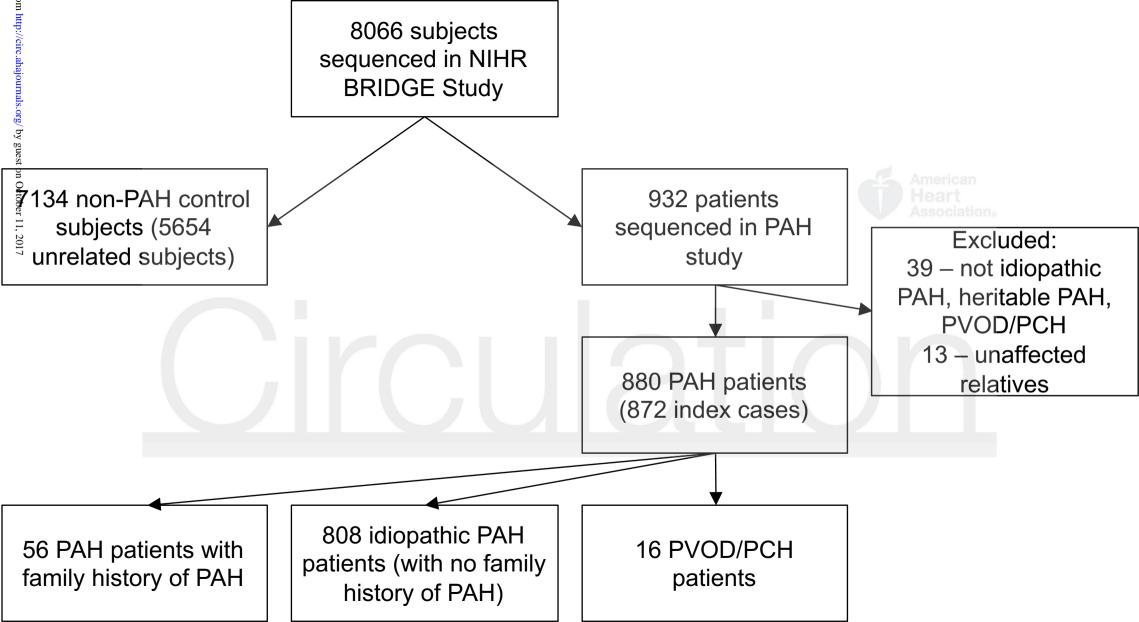
Figure 1. Subjects recruited to the NIHR BioResource – Rare Diseases Study and the clinical diagnostic categories of PAH patients included in this study.

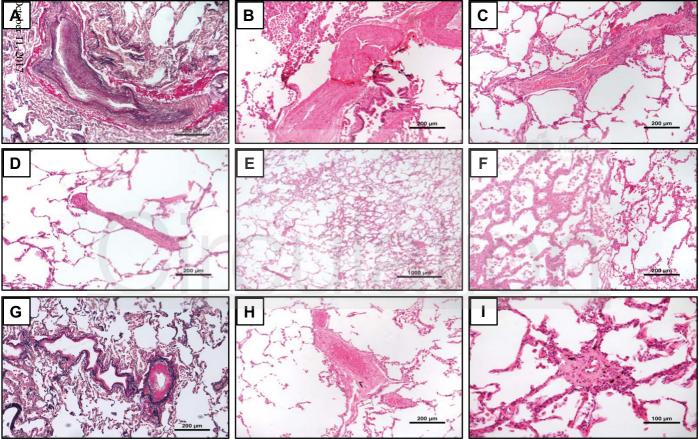
Figure 2. The transfer coefficient for carbon monoxide (KCO) is influenced by genotype in pulmonary arterial hypertension. Patients with  $FEV_1 < 80$  % predicted and FVC < 80 % predicted and diagnosed with PAH or PVOD/PCH after 50 years of age excluded from the plot.

Figure 3. Representative histopathological images from one patient with clinically and diagnosed idiopathic PAH but found to have a rare (not reported in the ExAC database) and predicted deleterious (CADD score 32) homozygous EIF2AK4 missense variant (c.1795G>C). The patient was of Pakistani origin and did not have a family history of PAH or PVOD. At presentation, he was 22 years old and had a reduced KCO (31% predicted) despite preserved spirometry. HRCT of his chest showed subtle but extensive (50-75% involvement) ground glass opacification. No interlobular septal thickening or mediastinal lymphadenopathy was observed. No suspicion of PVOD/PCH was raised based on radiological appearances. Histopathology was reviewed by two independent pathologists each confirming the predominant histological pattern to be one of pulmonary arterial vasculopathy. The pulmonary arteries showed eccentric and concentric intimal fibrosis and medial hypertrophy (A, B) as well as some lesions with features of recanalised thrombus (C). Several concentrically muscularised arterioles were also observed (D). No complex plexiform lesions were present. There was patchy thickening of the alveolar septa with capillary congestion and pigmented intra-alveolar

macrophages similar to PCH (E, F). Venous remodelling was difficult to trace and infrequent, but present. Fibrous thickening of the intima in septal veins (G, I) and a micro-vessel (H).







### <u>Circulation</u>



### Phenotypic Characterisation of EIF2AK4 Mutation Carriers in a Large Cohort of Patients Diagnosed Clinically with Pulmonary Arterial Hypertension

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The NIHR BioResource - Rare Diseases Consortium & UK National Cohort Study of Idiopathic and Heritable PAH

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Natalie Canham	London North West Healthcare NHS Trust	UK
Emma Wakeling	London North West Healthcare NHS Trust	UK
Susan Holder	London North West Healthcare NHS Trust	UK
Neeti Ghali	London North West Healthcare NHS Trust	UK
Angie Brady	London North West Healthcare NHS Trust	UK
Virginia Clowes	London North West Healthcare NHS Trust	UK
Robert MacLaren	Moorfields Eye Hospital NHS Foundation Trust	UK
	Moorfields Eye Hospital NHS Foundation	
Andrew Webster	Trust/University College London	UK

	Moorfields Eye Hospital NHS Foundation	
Anthony Moore	Trust/University College London	UK
Gavin Arno	Moorfields Eye Hospital NHS Foundation Trust/University College London Moorfields Eye Hospital NHS Foundation	UK
Michel Michaelides	Trust/University College London	UK
Julia Rankin	Royal Devon & Exeter NHS Foundation Trust UCL Great Ormond Street Institute of Child	UK
Manju Kurian	Health University College London Hospitals NHS	UK
Elaine Murphy	Foundation Trust	UK
Keren Carss	University of Cambridge	UK
Alba Sanchis-Juan	University of Cambridge	UK
Marie Erwood	University of Cambridge	UK
Eleanor Dewhurst	University of Cambridge	UK
	University of Cambridge (CIMR Medical	
Detelina Grozeva	Genetics)	UK
	University of Cambridge/Cambridge University	
F Lucy Raymond	Hospitals	UK
5 0 11	University of Cambridge/Cambridge University	1112
Evan Reid	Hospitals NHS Foundation Trust	UK
Geoff Woods	University of Cambridge/Cambridge University Hospitals NHS Foundation Trust	UK
Geoff Woods	University of Cambridge/Cambridge University	OK
Marc Tischkowitz	Hospitals NHS Foundation Trust	UK
	University of Cambridge/Cambridge University	
Richard Sandford	Hospitals NHS Foundation Trust	UK
PAH		
	University of Cambridge/Cambridge University	
Nicholas Morrell	Hospitals	UK
Stefan Gräf	University of Cambridge	UK

	Department of Medicine, University of	
Marta Bleda	Cambridge	UK
	Department of Medicine, University of	
Charaka Hadinnapola	Cambridge	UK
	Department of Medicine, University of	
Matthias Haimel	Cambridge	UK
	Cambridge University Hospitals NHS	
Simon Holden	Foundation Trust	UK
	Department of Medicine, University of	
Jennifer Martin	Cambridge	UK
Sonia Ali	Imperial and Hammersmith	UK
Harm Boggard	VU University Medical Center, Amsterdam	Netherlands
Colin Church	Golden Jubilee National Hospital	UK
Paul Corris	Newcastle Freeman	UK
Gerry Coghlan	Royal Free	UK
Amanda Creaser-Myers	Sheffield CRF, Royal Hallamshire	UK
Victoria Cookson	GOSH	UK
Rosa DaCosta	Royal Brompton	UK
Natalie Dormand	Royal Brompton	UK
Pavandeep K Ghataorhe	Imperial and Hammersmith	UK
Simon Gibbs	Imperial and Hammersmith	UK
Alan Greenhalgh	Newcastle Freeman	UK
Marc Humbert	University of South Paris	France
Anna Huis in't Veld	VU University Medical Center, Amsterdam	Netherlands
Fiona Kennedy	Golden Jubilee National Hospital	UK
David Kiely	Sheffield CRF, Royal Hallamshire	UK
Allan Lawrie	Sheffield CRF, Royal Hallamshire	UK
Rob Mackenzie Ross	Bath	UK
Rajiv Machado	University of Lincoln	UK
Larahmie Masati	Imperial and Hammersmith	UK
Sharon Meehan	Imperial and Hammersmith	UK

Shahin Moledina	GOSH	UK
Shokri Othman	Imperial and Hammersmith	UK
Andrew Peacock	Golden Jubilee National Hospital	UK
Joanna Pepke-Zaba	Papworth Hospital	UK
Val Pollock	Golden Jubilee National Hospital	UK
Gary Polwarth	Papworth Hospital	UK
Christopher J Rhodes	Imperial and Hammersmith	UK
Kevin Rue-Albrecht	Imperial and Hammersmith	UK
Gwen Schotte	VU University Medical Center, Amsterdam	Netherlands
Debbie Shipley	Newcastle Freeman	UK
Laura Southgate	Kings College, London	UK
Respiratory Nurse Specialists	Bath	UK
Jay Suntharalingam	Bath	UK
Yvonne Tan	Royal Free	UK
Mark Toshner	Papworth Hospital	UK
	Department of Medicine, University of	
Carmen Treacy	Cambridge	UK
Richard Trembath	Kings College, London	UK
Anton Vonk Noordegraaf	VU University Medical Center, Amsterdam	Netherlands
Ivy Wanjiku	Imperial and Hammersmith	UK
John Wharton	Imperial and Hammersmith	UK
Martin Wilkins	Imperial and Hammersmith	UK
John Wort	Royal Brompton	UK
John Wharton	Imperial and Hammersmith	UK
PID		
Kenneth Smith	University of Cambridge	UK
Taco Kuijpers	Emma Children's Hospital, Amsterdam UCL Great Ormond Street Institute of Child	Netherlands
Adrian Thrasher	Health	UK
James Thaventhiran	University of Cambridge	UK

Matthew Brown	University of Cambridge	UK
Hana Lango Allen	University of Cambridge	UK
Ilenia Simeoni	University of Cambridge	UK
	University of Cambridge/Cambridge University	
Emily Staples	Hospitals NHS Foundation Trust	UK
Crina Samarghitean	University of Cambridge	UK
Hana Alachkar	Salford Royal NHS Foundation	UK
Richard Antrobus	University Hospitals Birmingham	UK
Gururaj Arumugakani	Leeds Teaching Hopsital	UK
	UCL Great Ormond Street Institute of Child	
Chiara Bacchelli	Health	UK
Helen Baxendale	Papworth Hospital	UK
Claire Bethune	Plymouth Hopsital	UK
	UCL Great Ormond Street Institute of Child	
Shahnaz Bibi	Health	UK
	UCL Great Ormond Street Institute of Child	
Claire Booth	Health	UK
Michael Browning	Leicester Royal Infirmary	UK
Siobhan Burns	Royal Free Hospital	UK
	Cambridge University Hospitals NHS	
Anita Chandra	Foundation Trust	UK
Nichola Cooper	Imperial College Healthcare NHS Trust	UK
Coulty Do to	Cambridge University Hospitals NHS	1.117
Sophie Davies	Foundation Trust	UK
Lisa Devlin	Royal Hospitals Belfast	UK
Rainer Doffinger	University of Cambridge	UK
Elizabeth Drewe	Nottingham University Hospitals NHS Trust	UK
David Edgar	Royal Hospitals Belfast	UK
William Egner	Sheffield Teaching Hospitals	UK
Rohit Ghurye	Barts Health NHS Trust	UK

	UCL Great Ormond Street Institute of Child	
Kimberley Gilmour	Health	UK
Sarah Goddard	University Hospitals of North Midlands	UK
Pavel Gordins	Hull & East Yorkshire Hospitals NHS Trust	UK
Sofia Grigoriadou	Barts Health NHS Trust	UK
Scott Hackett	Birmingham Heartlands	UK
	Royal Hospital for Children, NHS Greater	
Rosie Hague	Glasgow and Clyde	UK
	Epsom & St Helier University Hospitals NHS	
Grant Hayman	Trust	UK
Archana Herwadkar	Salford Royal NHS Foundation	UK
Aarnoud Huissoon	Birmingham Heartlands	UK
Stephen Jolles	University Hospital Wales	UK
Peter Kelleher	Imperial College Healthcare NHS Trust	UK
	Cambridge University Hospitals NHS	
Dinakantha Kumararatne	Foundation Trust	UK
Sara Lear	Norforlk & Norwich University Hospital	UK
Hilary Longhurst	Barts Health NHS Trust	UK
Lorena Lorenzo	Barts Health NHS Trust	UK
	UCL Great Ormond Street Institute of Child	
Jesmeen Maimaris	Health	UK
	Cambridge University Hospitals NHS	
Ania Manson	Foundation Trust	UK
Elizabeth McDermott	Nottingham University Hospitals NHS Trust	UK
	Gartnavel General Hospital, NHS Greater	
Sai Murng	Glasgow and Clyde	UK
Sergey Nejentsev	University of Cambridge	UK
Sadia Noorani	Sandwell and West Birmingham Hospitals	UK
Eric Oksenhendler	Hopital St Louis, Paris	France
Mark Ponsford	University Hospital Wales	UK

	UCL Great Ormond Street Institute of Child	
Waseem Qasim	Health	UK
Isabella Quinti	Sapienza Universita di Roma	Italy
Alex Richter	University Hospitals Birmingham	UK
Ravishankar Sargur	Sheffield Teaching Hospitals	UK
Sinisa Savic	Leeds Teaching Hopsital	UK
Suranjith Seneviratne	Royal Free Hospital	UK
Carrock Sewell	Scunthorpe General Hospital	UK
Hans Stauss	Royal Free Hospital	UK
	Gartnavel General Hospital, NHS Greater	
Moira Thomas	Glasgow and Clyde	UK
Steve Welch	Birmingham Heartlands	UK
	Cambridge University Hospitals NHS	
Lisa Willcocks	Foundation Trust	UK
Nigel Yeatman	Barts Health NHS Trust	UK
Patrick Yong	Frimley Park Hospital	UK

## **SUPPLEMENTAL MATERIAL:**

Phenotypic characterisation of *EIF2AK4* mutation carriers in a large cohort of patients diagnosed clinically with pulmonary arterial hypertension

Hadinnapola et al.

#### **Supplemental Methods:**

Whole genome sequencing

Genomic DNA was extracted from whole blood samples prior to assessment of concentration by Qubit, and quality by gel electrophoresis. After fragmentation of DNA into 200bp fragments (Covaris E220, Covaris Inc, Woburn, USA) DNA libraries were created using Tru SeqDNA LT Prep kit (Illumina Inc, San Diego, USA). The libraries underwent next generation sequencing using 100-150 base pair paired-end sequencing using Illumina HiSeq 2500 and HiSeq X (Illumina Inc, San Diego, USA).

#### Variant calling

Reads were aligned against the Genome Reference Consortium human genome (build 37) (GRCh37) and variants were called using the Issac Aligner and Variant Caller respectively (version 2, Illumina Inc.). Genebuilds for *BMPR2* and *EIF2AK4* genes were based on Ensembl v75. Variants from these genes were extracted and annotated using Ensembl's Variant Effect Predictor (VEP) v84 <sup>1</sup>. VEP was also used to annotate data from the Exome Aggregation Consortium's (ExAC) database <sup>2</sup>.

Deletions (resulting in the loss of more than 50bp) were identified by applying Isaac copy number variant caller (Canvas, Illumina) and Isaac Structural Variant Caller (Manta, Illumina).

To be called by both Canvas and Manta deletions required a reciprocal overlap of  $\geq$  20%. Overlapping deletions represented in the Zarrei dataset with a reciprocal overlap of  $\geq$  50% and deletions with a non-PAH BRIDGE control frequency of more than 1 in 1,000 were excluded  $^3$ .

#### Analysis of computed tomographic images of the chest

CT images of the chest, where available, were reviewed independently by 2 cardiothoracic radiologists (AS and NS), with specialist imaging experience in pulmonary hypertension, blinded to the underlying diagnoses using a customised proforma (Supplemental Table 4). In addition to CT scans of patients with EIF2AK4 mutations or with a clinical diagnosis of PVOD in the cohort, CT scans of patients from Papworth Hospital and the Royal Hallamshire Hospital with normal spirometry (FEV<sub>1</sub> > 80% predicted and FVC > 80% predicted) and either BMPR2 mutations (n=21) or no variants in the known PAH genes (n=21) were analysed (Supplemental Table 5). A consensus read was undertaken for individual CT features and a mutually agreed overall radiological diagnosis was recorded.

#### Histology

The explanted lung tissue of one patient with a clinical diagnosis of idiopathic PAH and biallelic *EIF2AK4* mutations was available for further analysis. Four micrometre (µm) tissue sections were cut from formalin-fixed paraffin wax embedded blocks from the explanted lung tissue. Representative sections from each lobe of both lungs were stained with Elastic-Van Gieson and Haemotoxylin and Eosin stains. Two expert histopathologists examined the sections independently by light microscopy.

## Statistical analysis

Statistical analysis was performed in R (www.r-project.org).

Differences between groups of categorical variables were assessed using the Fisher Exact test. Where one of the variables was an ordinal the Cochran-Armitage test was applied using the chisq\_test function from the "coin" package <sup>4</sup>. Differences in continuous variables were assessed using the Mann–Whitney U test (2 comparator groups) and the Kruskal-Wallis test (3 or more comparator groups). Post-hoc pairwise comparisons were performed using Dunn's Test for multiple testing.

Semi-parametric Cox-proportional hazards models were used to assess survival between groups using the "survival" package in R <sup>5</sup>. Survival time from diagnosis to death and diagnosis to death or transplantation was assessed. Patients were censored at the date of transplantation for the primary survival analysis. Age at diagnosis and gender were used as covariates in the models.

The proportional hazards assumptions were tested by assessing Schoenfeld residuals over log time  $^6$ . The goodness of fit of the model was assessed by plotting the log of cumulative hazard of Cox-Snell residuals against the log of time and confirming the simple regression has 0 intercept and slope of 1  $^7$ .

The inclusion of retrospectively recruited and prevalent patients in a survival analysis assessing time from diagnosis to death/transplantation can cause immortal time bias. The immortal time is the period between diagnosis and enrolment in the study and so patients

had to have survived till this point. Patients with worse prognosis diagnosed at a similar time may not have survived long enough to enrol in the study. To further explore this potential bias, a sensitivity analysis was performed including only on UK patients recruited prospectively to the study. In this multivariate Cox-proportional hazards model, the survival period was defined as the time period from date of diagnosis to date of death and patients only entered the risk set after enrolment into the study (consent date).

# **Supplemental Tables**

Supplemental Table 1. NIHR BioResource – Rare Diseases Collaboration. See spreadsheet.

Centre	Principle	Clinicians and research staff
	Investigator	
Freeman Hospital, Newcastle,	Paul A Corris	Alan Greenhalgh, Debbie Shipley,
UK		Margaret Day
Golden Jubilee National	Andrew	Colin Church, Val Irvine, Fiona Kennedy
Hospital, Glasgow, UK	Peacock	
Great Ormond Street	Shahin	Victoria Cookson
Hospital, London, UK	Moledina	
Hammersmith Hospital and	Martin R	Simon Gibbs, John Wharton, Sonia Ali,
Imperial College, London, UK	Wilkins	Larahmie Masati, Sharon Meehan, Ivy
		Wanjiku, Shokri Othman
Papworth Hospital,	Joanna Pepke-	Mark Toshner, Gary Polwarth
Cambridge, UK	Zaba	
Royal Brompton Hospital,	Stephen J Wort	Rosa DaCosta, Natalie Dormand, Alice
London, UK		Parker
Royal Free Hospital, London,	Gerry Coghlan	Yvonne Tan, Dipa Ghedia
UK		
Royal Hallamshire Hospital,	David G Kiely	Robin Condliffe, Amanda Creaser-Myers,
Sheffield, UK		Stephen Roney, Sara Walker
Royal United Hospitals Bath	Jay	Robert MacKenzie Ross, Mark Grover, Ali
NHS Foundation Trust, Bath,	Suntharalingam	Grove, Jill Peel, Ann Coy
UK		
University of South Paris	Marc Humbert	David Montani, Florent Soubrier, Barbara
		Girerd, Mélanie Eyries
VU University Medical Center,	Anton Vonk	Harm Bogaard, Anna Huis in't Veld, Gwen
Amsterdam, Netherlands	Noordegraaf	Schotte, Ale Struiksma
Supplemental Table 2. Special	ist pulmonary hyp	ertension centres participating in the study

Recruiting cohorts	n						
Genomics England	1965						
Specialist Pathology: Evaluating Exomes in	1356						
Diagnostics							
Primary Immune Disorders	1299						
Bleeding and Platelet Disorders	1004						
Pulmonary Arterial Hypertension	932						
Multiple Primary Malignant Tumours	376						
Hypertrophic Cardiomyopathy	187						
Cerebral Small Vessel Diseases	183						
Steroid Resistant Nephrotic Syndrome	161						
Intrahepatic Cholestasis of Pregnancy	140						
Stem Cell & Myeloid Disorders	132						
Primary Membranoproliferative Glomerulonephritis	128						
Neuropathic Pain Disorder	114						
Leber Hereditary Optic Neuropathy	59						
Control	15						
Ehlers-Danlos Syndromes	15						
Supplemental Table 3. NIHR BioResource - Rare Dise	ases Study						
recruiting cohorts and GEL							

Parameter	Response
ID	
Date of birth	
Unenhanced CT	(Y/N)
СТРА	(Y/N)
HRCT	(Y/N)
Expiratory CT	(Y/N)
Pulmonary artery diameter (cm)	
Aorta diameter (cm)	
Ground glass opacification centrilobular pattern DENSITY	(None / Subtle / Present)
Ground glass centrilobular pattern EXTENT	(0, <5%, 5-25, 25-50, >50)
Ground glass DISTRIBUTION	(central (C)/peripheral (P)/zonal (Z) or diffuse (D))
Non-specific mosaic pattern / GGO	
Neovascularity vessels	(Y/N)
Arterio-venous malformations	(Y/N)
Bronchial arteries	(Y/N)
Largest bronchial artery size	
Interlobular septal thickening	(None, Subtle, Present)
Mediastinal lymphadenopathy	(Y/N)
Emphysema	(Y/N) and % of parenchyma involved
Fibrosis	(Y/N) and % of parenchyma involved
Pleural effusion	(Y/N)
Air trapping	(Y/N)
Comments	
Likely diagnosis	Any suspicion of PVOD or PCH / PAH
Supplemental Table 4. Proforma used in	n analysis of CT scans

Group	n
PAH patients with BMPR2 variants	21
PAH patients with biallelic <i>EIF2AK4</i>	7
variants	,
PVOD patients	14
PAH patients with heterozygous EIF2AK4	4
variants	4
PAH patients with no variants in the	21
previously reported PAH genes	21

**Supplemental Table 5**. CT scans of patients with PVOD and patients with PAH carrying biallelic *EIF2AK4* mutations were reassessed by radiologists blinded to the diagnosis. For comparison CT scans of PAH patients with normal spirometry (FEV $_1$  > 80 % predicted and FVC > 80 % predicted) who either had no mutations in the previously reported PAH genes or carried *BMPR2* mutations were assessed.

			Su	pplemental	Table 6. Pa	age 1/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.292C>G	missense variant	p.L98V	0	1	0.00001656	probably damaging (0.999)	deleterious (0)	25.7	Heterozygous variant
BRIDGE control	c.354_355delTG	frameshift variant	p.C118Wfs*7	0	2	Not found in ExAC			35	Heterozygous variant
BRIDGE control	c.745C>T	stop gained & splice region variant	p.R249*	0	1	0.00007451			39	Heterozygous variant
BRIDGE control	c.746G>A	missense variant & splice region variant	p.R249Q	0	1	2.48E-05	probably damaging (0.999)	deleterious (0.02)	34	Heterozygous variant
BRIDGE control	c.767G>T	missense variant	p.C256F	0	1	1.66E-05	possibly damaging (0.904)	deleterious (0.02)	28.4	Heterozygous variant
BRIDGE control	c.985G>A	missense variant	p.E329K	0	1	Not found in ExAC	probably damaging (0.981)	deleterious (0.01)	34	Heterozygous variant
BRIDGE control	c.1153dupG	frameshift variant	p.V385Gfs*30	0	1	0.00003308			32	Heterozygous variant
BRIDGE control	c.1190T>A	missense variant	p.l397N	0	1	Not found in ExAC	possibly damaging (0.67)	deleterious (0)	32	Heterozygous variant

			S	upplement	al Table 6.	Page 2/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen-2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.1215C>G	stop gained	p.Y405*	0	2	Not found in ExAC			29.4	Heterozygou variant
BRIDGE control	c.1331A>G	missense variant	p.Y444C	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28.7	Heterozygou variant
BRIDGE control	c.1345C>T	missense variant	p.R449C	0	1	0.00001654	probably damaging (1)	deleterious (0)	35	Heterozygou variant
BRIDGE control	c.2249T>A	missense variant & splice region variant	p.L750Q	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	28	Heterozygou variant
BRIDGE control	c.2298delG	frameshift variant	p.N767Tfs*24	0	1	Not found in ExAC			28.3	Heterozygou variant
BRIDGE control	c.2720A>T	missense variant	p.Y907F	0	4	1.66E-05	probably damaging (1)	deleterious (0)	31	Heterozygou variant
BRIDGE control	c.2828C>T	missense variant	p.T943M	0	1	0.00003311	probably damaging (1)	deleterious (0)	34	Heterozygou variant
BRIDGE control	c.3104_3106delT CT	inframe deletion	p.F1035del	0	1	Not found in ExAC			22	Heterozygou variant

			Sı	upplementa	Table 6. Pa	age 3/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.3217C>T	missense variant	p.R1073C	0	1	0.0000166	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3223T>G	missense variant	p.F1075V	0	1	0.0000083	probably damaging (0.997)	deleterious (0)	32	Heterozygous variant
BRIDGE control	c.3344C>T	missense variant	p.P1115L	0	1	8.26E-06	probably damaging (1)	deleterious (0)	35	Heterozygous variant
BRIDGE control	c.3358-3C>T	splice region variant & intron variant	p.NA	0	1	Not found in ExAC			17.15	Heterozygous variant
BRIDGE control	c.3406C>T	stop gained & splice region variant	p.R1136*	0	1	Not found in ExAC			40	Heterozygous variant
BRIDGE control	c.3430A>T	missense variant	p.R1144W	0	1	0.0000248	probably damaging (1)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.3986T>C	missense variant	p.F1329S	0	1	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Heterozygous variant

			Su	pplemental	Table 6. Pa	age 4/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
BRIDGE control	c.3992T>C	missense variant	p.F1331S	0	1	8.28E-06	possibly damaging (0.872)	deleterious (0.01)	28.4	Heterozygous variant
BRIDGE control	c.4039G>A	missense variant	p.A1347T	0	1	8.28E-05	probably damaging (1)	deleterious (0)	34	Heterozygous variant
BRIDGE control	c.4388_4389+12 delAGGTAAAGAC GTCA	splice donor variant & coding sequence variant & intron variant	p.NA	0	1	Not found in ExAC			36	Heterozygous variant
BRIDGE control	c.4397C>A	missense variant	p.S1466Y	0	2	Not found in ExAC	probably damaging (0.988)	deleterious (0)	33	Heterozygous variant
BRIDGE control	c.4729G>A	missense variant & splice region variant	p.V1577M	0	1	Not found in ExAC	probably damaging (0.999)	deleterious (0)	29.6	Heterozygous variant
BRIDGE control	c.4751dupT	frameshift variant	p.L1585lfs*11	0	1	Not found in ExAC			34	Heterozygous variant
BRIDGE control	c.4920_4931delT AGAGATGACTA	inframe deletion	p.R1641_Y1644 del	0	1	Not found in ExAC			23	Heterozygous variant

	1101/0		1101/0				5   5		6455	E150 A17 -
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	EXAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.44C>T	missense variant	p.P15L	1	0	8.32E-06	unknown (0)	deleterious low confidence (0.03)	23.5	Heterozygous variant
РАН	c.220G>A	missense variant	p.D74N	1	0	1.66E-05	possibly damaging (0.954)	deleterious (0)	32	Heterozygous variant
PAH	c.1072_1073dup GT	frameshift variant	p.V359*	1	0	Not found in ExAC			32	Heterozygous variant
PAH	c.1660G>T	missense variant & splice region variant	p.D554Y	1	0	Not found in ExAC	probably damaging (0.966)	deleterious (0)	28	Heterozygous variant
PAH	c.2446C>T	stop gained	p.Q816*	1	0	Not found in ExAC			41	Heterozygous variant
PAH	c.2516T>C	missense variant	p.I839T	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	28.9	Heterozygous variant
PAH	c.3218G>T	missense variant	p.R1073L	1	0	Not found in ExAC	probably damaging (0.995)	deleterious (0.01)	35	Heterozygous variant
PAH	c.3604C>T	missense variant	p.H1202Y	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	29.7	Heterozygous variant

			Su	pplemental	Table 6. Pa	age 6/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.3711_3713del GAG	inframe deletion	p.R1238del	1	0	0.0000083			21.6	Heterozygous variant
PAH	c.3722A>G	missense variant	p.E1241G	1	0	Not found in ExAC	probably damaging (0.971)	deleterious (0)	27.2	Heterozygous variant
PAH	c.4646G>A	missense variant	p.R1549H	1	0	0.0000910	probably damaging (0.998)	deleterious (0.01)	35	Heterozygous variant
РАН	c.145-2A>G	splice acceptor variant	p.NA	1	0	Not found in ExAC			23.9	Additional second (likely trans) variant identified
PAH	c.257+4A>C	splice region variant & intron variant	p.NA	1	0	8.28E-06			15.5	Additional second (likely trans) variant identified
PAH	c.1392delT	frameshift variant	p.R465Vfs*38	1	0	2.48E-05			35	Additional second (likely trans) variant identified
PAH	c.1739dupA	frameshift variant	p.R581Efs*9	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified

			Su	pplemental	Table 6. Pa	ige 7/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.1820T>G	missense variant & splice region variant	p.V607G	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	27.3	Additional second (likely trans) variant identified
РАН	c.2727C>G	missense variant	p.S909R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	33	Additional second (likely trans) variant identified
PAH	c.2827A>G	missense variant	p.T943A	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	26.4	Additional second (likely trans) variant identified
PAH	c.2841delG	frameshift variant	p.1948Sfs*35	1	0	Not found in ExAC			35	Additional second (likely trans) variant identified
PAH	c.3055_3064delC TGACCAACG	frameshift variant	p.L1019Wfs*9	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Additional second (likely trans) variant identified

			Su	pplemental	Table 6. Pa	ige 8/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.3325G>A	missense variant	p.G1109R	1	0	0.0000082	probably damaging (1)	deleterious (0.02)	35	Additional second (likely trans) variant identified
PAH	c.3884T>G	missense variant	p.L1295R	1	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Additional second (likely trans) variant identified
PAH	c.4400dupT	frameshift variant	p.E1468Rfs*14	1	0	Not found in ExAC			36	Additional second (likely trans) variant identified
PAH	c.4418_4421delC AGA	frameshift variant	p.T1473Rfs*17	1	0	0.0000083			36	Additional second (likely trans) variant identified
PAH	c.4769delT	frameshift variant	p.L1590*	1	0	0.0000083			33	Additional second (likely trans) variant identified
PAH	c.281dupA	frameshift variant	p.N94Lfs*8	2	0	Not found in ExAC			35	Homozygous variant
PAH	c.1159_1160delC T	frameshift variant	p.L387Cfs*27	2	0	Not found in ExAC			29.6	Homozygous variant

			Sı	pplemental	Table 6. Pa	age 9/9				
Project	HGVSc	Consequence	HGVSp	Allele count PAH patients	Allele count non-PAH BRIDGE controls	ExAC MAF	PolyPhen- 2	SIFT	CADD Phred Score	EIF2AK4 genotype
PAH	c.1795G>C	missense variant	p.G599R	4	0	Not found in ExAC	probably damaging (1)	deleterious (0)	32	Homozygous variant
PAH	c.3097C>T	stop gained	p.Q1033*	3	0	8.24E-06			45	Homozygous variant
PAH	c.3605A>T	missense variant	p.H1202L	2	0	Not found in ExAC	probably damaging (1)	deleterious (0)	31	Homozygous variant
PAH	c.4392dupT	frameshift variant & splice region variant	p.K1465*	2	0	Not found in ExAC			35	Homozygous variant

	Supplemental Table 7. Page 1/4																		
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	BMPR2 mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV <sub>1</sub> (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
22		c.3884T>G	missense variant	Clist			F2	2.2	2	07	110	22	V	Possible		PDE5i +	Na		
23	IVI	British	c.3055_30 64delCTGA CCAACG	frameshift variant	C Het			52	3.3	3	97	119	33	Yes	PVOD / PCH		ERA + IV Prostanoid	No	
18	M	Other	c.4400dup T	frameshift variant	C Het			46	6.4	3	116	120	45	No	CT not available		ERA + PDE5i +	No	
40	48 M Other	Other	c.1739dup A	frameshift variant	Criet			40	0.4	3	110	120	43	NO	for analysis		inhaled Prostanoid	NO	
		c.2827A>G	missense variant											CT not					
38	F	Other Asian	c.4418_44 21delCAGA	frameshift variant	C Het			40	4.5	2				No	available for analysis		ERA + PDE5i	No	
			c.145- 2A>G	splice acceptor variant											anaiysis				

**Supplemental Table 7.** Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV<sub>1</sub> – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 7. Page 2/4																		
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	BMPR2 mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV <sup>1</sup> (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
			c.1392del T	frameshift variant											Possible		PDE5i +		
70	F	British	c.257+4A >C	splice region variant & intron variant	C Het			76	6.6	З	101	127	33	Unk	PVOD / PCH		ERA + inhaled Prostanoid	No	
36	F	Indian	c.3605A> T	missense variant	Hom			44	2.7	3	73	83	40	Yes	Possible PVOD / PCH		ERA + PDE5i + inhaled Prostanoid	No	
22	М	Pakistani	c.1795G> C	missense variant	Hom			65	3.0	3	92	93	31	Yes	РАН		ERA + PDE5i + IV Prostanoid	No	Yes
29	Δ	Pakistani	c.3097C> T	stop gained	Hom			50	4.9	3	99	107	27	Unk	РАН	Sister died from PAH	PDE5i	No	
18	М	Not stated	c.1159_1 160delCT	frameshift variant	Hom			92		3	86	82	28	No	Possible PVOD / PCH		ERA + IV Prostanoid	No	
25	F	Pakistani	c.1795G> C	missense variant	Hom			57	5.6	3	82	87	33	No	РАН		PDE5i + ERA	No	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with EIF2AK4 variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV<sub>1</sub> – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 7. Page 3/4																		
Age (years)	Gender	Ethnicity	<i>EIF2AK4</i> variant HGVSc	Consequence type	EIF2AK4 genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV <sub>1</sub> (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
24	F	Not	c.2446C> T	stop gained	Het (both on			60	5.2	3	96	97	81	Unk	CT not available	Father and sister	Unk	Unk	
24	24 F	stated	c.3218G> T	missense variant	same allele) *			0	5.2	3	30	37	01	Olik	for analysis	died of PAH	Olik	OTIK	
39	F	British	c.1072_1 073dupG T	frameshift variant	Het			54	3.0	2	87	98	72	No	CT not available for analysis		ERA	No	
40	F	British	c.44C>T	missense variant	Het		c.4303- 50delT	43	5.6	2	99	96	109	Unk	Possible PVOD / PCH		ERA	No	
44	М	British	c.2516T> C	missense variant	Het	c.853- 2A>G (splice acceptor variant)	c.361- 180A>G	53	3.8	3	102	98	54	Unk	РАН		PDE5i + ERA	No	
25	F	British	c.3722A> G	missense variant	Het					3	53	49	41	No	CT not available for analysis		PDE5i + ERA + IV Prostanoid	No	

**Supplemental Table 7.** Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with *EIF2AK4* variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV<sub>1</sub> – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown, \*maternally inherited

	Supplemental Table 7. Page 4/4																		
Age (years)	Gender	Ethnicity	EIF2AK4 variant HGVSc	Consequence type	EIF2AK4 genotype	<i>BMPR2</i> mutation	Non-protein coding <i>EIF2AK4</i> variant	mPAP (mmHg)	Cardiac output (L/min)	FC	FEV <sub>1</sub> (% pred)	FVC (% pred)	KCO (% pred)	Digital clubbing	CT diagnosis	Family history PAH	Pulmonary artery vasodilator therapy	Pulmonary oedema with treatment	Histology assessed
66	F	Not stated	c.4646G> A	missense variant	Het			44	2.1	3	79	100		Unk	РАН		PDE5i + ERA	No	
72	М	British	c.1660G> T	missense variant & splice region variant	Het			30	2.8	3				No	РАН		IV Prostanoid	No	
59	F	Other	c.3711_3 713delGA G	inframe deletion	Het			41	3.4	3	68	68	95	Unk	РАН		ERA + PDE5i	No	
48	F	British	c.3604C>	missense variant	Het	c.2695C>T (stop gained)		57	4.4	4	90	100	61	Unk	РАН		PDE5i + ERA	No	
70	F	Other White	c.220G>A	missense variant	Het			42	5.4	2				Unk	CT not available for analysis		ERA	Unk	

Supplemental Table 7. Phenotypic and genotypic description of patients with a clinical diagnosis of PAH with EIF2AK4 variants. mPAP – mean pulmonary artery pressure, FC – functional class, FEV<sub>1</sub> – forced expiratory volume in 1s, FVC - forced vital capacity, Kco – transfer coefficient for carbon monoxide, PDE5i – phosphodiesterase type 5 inhibitor, ERA – endothelin receptor antagonist, C Het – compound heterozygous, Hom – homozygous, Het – heterozygous, Unk – unknown

	Supplemental Table 8. Page 1/2												
	PAH patients with BMPR2 mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with  EIF2AK4 heterozygous  variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	р							
n	64	255	3	7	5								
Age (years)	42 [31 - 52]	53 [39 - 67]	39 [32 - 40]	25 [23 - 38]	63 [27 - 76]	<0.001							
Gender (n female [%])	45 [70.3%]	179 [70.2%]	3 [100%]	2 [28.6%]	4 [80%]	0.161							
Ethnicity (n white Caucasian [%])	50 [78.1%]	226 [88.6%]	2 [66.7%]	2 [28.6%]	4 [80%]	<0.001							
Digital clubbing (n [%])	5 [13.2%]	3 [2.2%]	0 [0%]	2 [40%]	0 [0%]	0.004							
ВМІ	28 [25 - 33]	27 [24 - 31]	24 [24 - 25]	24 [21 - 27]	27 [24 - 32]	0.202							

Supplemental Table 8. Phenotype summary of patients with preserved spirometry (FEV<sub>1</sub> > 80 % predicted and FVC > 80 % predicted). PAH patients with biallelic *EIF2AK4* mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups.

mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV<sub>1</sub> – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. \* Also includes the 2 patients with heterozygous *EIF2AK4* variants and a *BMPR2* mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

		Supplemental	Table 8. Page 2/2			
	PAH patients with <i>BMPR2</i> mutations *	PAH patients with no mutations in PAH associated genes	PAH patients with  EIF2AK4 heterozygous  variants	PAH patients with biallelic <i>EIF2AK4</i> mutations	PVOD/PCH patients	р
mPAP (mmHg)	56 (15)	51 (18)	54 (8)	57 (20)	57 (7)	0.00
CO (L/min)	3 [3 - 4]	4 [3 - 5]	5 [4 - 5]	5 [4 - 6]	3 [3 - 3]	<0.0
PVR (WU)	14 [10 - 18]	10 [7 - 14]	8 [7 - 9]	9 [8 - 15]	14 [11 - 19]	<0.0
Vasoresponders (n [%])	0 [0%]	18 [21.7%]	0 [0%]	0 [0%]		0.01
FEV <sub>1</sub> (%pred)	97 [88 - 102]	93 [87 - 101]	96 [92 - 97]	97 [89 - 100]	98 [94 - 106]	0.52
FVC (%pred)	102 [96 - 113]	103 [96 - 112]	97 [96 - 98]	107 [90 - 120]	109 [101 - 113]	0.70
KCO (%pred)	80 [71 - 93]	68 [46 - 84]	81 [76 - 95]	33 [30 - 33]	33 [28 - 37]	<0.0
Resting S <sub>A</sub> O <sub>2</sub> (%)	96 [94 - 98]	96 [93 - 98]	98 [98 - 99]	91 [90 - 92]	95 [91 - 95]	0.02
S <sub>A</sub> O <sub>2</sub> post walk test (%)	95 [90 - 98]	91 [85 - 96]	94 [87 - 96]	80 [75 - 84]	85 [85 - 88]	<0.0

Supplemental Table 8. Phenotype summary of patients with preserved spirometry (FEV<sub>1</sub> > 80 % predicted and FVC > 80 % predicted). PAH patients with biallelic *EIF2AK4* mutations are still younger at diagnosis and have a significantly reduced KCO compared to other groups.

mPAP – mean pulmonary artery pressure, CO – cardiac output, PVR – pulmonary vascular resistance, FEV<sub>1</sub> – forced expiratory volume in 1 second, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide, BMI – body mass index. \* Also includes the 2 patients with heterozygous *EIF2AK4* variants and a *BMPR2* mutation. Data presented as median [IQR] unless indicated. Percentages were calculated using the number of patients for whom data were available as the denominator.

	Supplementa	al Table 9. Page 1/2		
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no EIF2AK4 mutation	р
n		11	10	
Age (years)		26.8 [22.5 - 34.3]	68.3 [63.9 - 72.1]	0.001
Gender (n female [%])		6 [54.5%]	5 [50.0%]	1.000
Ethnicity (n white Caucasian [%])		5 [45.5%]	9 [90.0%]	0.063
mPAP (mmHg)		52 [47 - 63]	48 [42 - 57]	0.342
PCWP (mmHg)		11 [7.5 - 12]	11.5 [9.0 – 12.2]	0.560
FEV <sub>1</sub> (% pred)		93.1 [82.8 - 98.5]	79.0 [72.3 – 91.0]	0.236
FVC (% pred)		95.5 [84.6 - 108.5]	96.0 [73.0 – 101.0]	0.720
KCO (% pred)		32.0 [28.7 – 33.0]	41.4 [36.8 – 54.0]	0.013
Contribute to the state of	None	2 [18.2%]	6 [60.0%]	
Centrilobular ground glass opacification density	Subtle	2 [18.2%]	3 [30.0%]	0.012
opacification density	Present	7 [63.6%]	1 [10.0%]	

**Supplemental Table 9.** Phenotypic and radiological characteristics of biallelic *EIF2AK4* mutation carriers compared to patients with a clinical diagnosis of PVOD and no *EIF2AK4* mutation.

mPAP – mean pulmonary artery pressure, PCWP – pulmonary capillary wedge pressure,  $FEV_1$  – forced expiratory volume 1 s, FVC – forced vital capacity, KCO – transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.

	Supplementa	l Table 9. Page 2/2		
Group		All biallelic <i>EIF2AK4</i> mutation carriers	PVOD with no EIF2AK4 mutation	р
	None	2 [18.2%]	7 [70.0%]	
	<5%	1 [9.1%]	1 [10.0%]	
Centrilobular ground glass	5-25%	2 [18.2%]	1 [10.0%]	0.007
opacification extent	25-50%	1 [9.1%]	1 [10.0%]	0.007
	50-75%	2 [18.2%]	0 [0.0%]	
	75-100%	3 [27.3%]	0 [0.0%]	
	None	7 [63.6%]	2 [20.0%]	
Interlobular septal thickening	Subtle	0 [0.0%]	1 [10.0%]	0.068
	Present	4 [36.4%]	7 [70.0%]	
Mediastinal	None	4 [36.4%]	2 [20.0%]	0.625
lymphadenopathy	Present	7 [63.6%]	8 [80.0%]	0.635
Diamed offusion	None	11 [100.0%]	6 [60.0%]	0.025
Pleural effusion	Small	0 [0.0%]	4 [40.0%]	0.035
Nie europeulo witeu	None	10 [90.9%]	9 [90.0%]	1 000
Neovascularity	Present	1 [9.1%]	1 [10.0%]	1.000
CT diagnosis	PAH	4 [36.4%]	3 [30.0%]	
CT diagnosis	Possible PVOD/PCH	7 [63.6%]	7 [70.0%]	

**Supplemental Table 9.** Phenotypic and radiological characteristics of biallelic *EIF2AK4* mutation carriers compared to patients with a clinical diagnosis of PVOD and no *EIF2AK4* mutation.

mPAP - mean pulmonary artery pressure, PCWP - pulmonary capillary wedge pressure,  $FEV_1$  - forced expiratory volume 1 s, FVC - forced vital capacity, KCO - transfer coefficient for carbon monoxide. Data presented as median [IQR] unless stated.

Group	Time to assessment 1 (days)	n	Change in 6mwd (m)	Change in FC	Time to assessment 2 (days)	n	Change in 6mwd (m)	Change in FC	Number on prostanoid therapy before the 2 <sup>nd</sup> assessment [%]
PAH <i>BMPR2</i>	357 [314 - 386]	21	+69 [20 - 100]	-1 [-11]	1120 [1055 - 1174]	18	+45 [31 - 115]	-1 [-10.5]	5 [23%]
PAH biallelic EIF2AK4	358 [335 -388]	9	+28 [-13 - 77]	0 [-1 - 0]	1102 [1090 – 1112]	5	+62 [-8 - 132]	0 [0 - 0]	1 [10%]
PAH no mutation	387 [340 - 414]	16	+81 [61 - 151]	-1 [-1 - 0]	1118 [1105 - 1159]	9	+104 [20 - 144]	-1 [-1 - 0]	4 [17%]
р	0.295		0.343	0.039	0.730		0.748	0.044	0.816

**Supplemental Table 10.** Response to pulmonary artery vasodilator therapies at 1 and 3 years after diagnosis compared to baseline. 6mwd - six-minute walk test distance, FC - functional class. Drop in number of patients between assessment 1 and 2 due to death, transplantation or lack of sufficient follow up time. Data presented as median [IQR] unless stated.

Variable	Hazard Ratio [95% confidence interval]	р
PAH BMPR2 mutation*	0.148 [0.055 - 0.396]	<0.001
PAH no mutation*	0.179 [0.073 - 0.440]	<0.001
PVOD*	0.393 [0.075 - 2.065]	0.27
Age at diagnosis	1.043 [1.033 - 1.053]	<0.001
Male gender	1.631 [1.222 - 2.179]	<0.001

**Supplemental Table 11.** Cox proportional hazards model assessing time to death. Patients with a clinical diagnosis of PAH and biallelic *EIF2AK4* mutations had an increased risk of death compared to other PAH patients. Number of patients = 858. Events = 194.

\* compared to the PAH biallelic *EIF2AK4* mutation carriers

Variable	Hazard Ratio [95% confidence interval]	р
PAH <i>BMPR2</i> mutation*	0.175 [0.066 - 0.462]	<0.001
PAH no mutation*	0.203 [0.083 - 0.501]	<0.001
PVOD*	0.840 [0.222 - 3.193]	0.798
Age at diagnosis	1.036 [1.027 - 1.046]	<0.001
Male gender	1.542 [1.165 - 2.042]	0.002

**Supplemental Table 12.** Cox proportional hazards model assessing time to death or transplantation. Number of patients = 858. Events = 208.

Variable	Hazard Ratio [95% confidence interval]	р
PAH BMPR2 mutation*	0.376 [0.080 - 1.763]	0.215
PAH no mutation*	0.456 [0.109 - 1.905]	0.282
PVOD*	1.029 [0.133 - 7.953]	0.978
Age at diagnosis	1.034 [1.020 - 1.046]	<0.001
Male gender	1.515 [1.000 - 2.296]	0.051

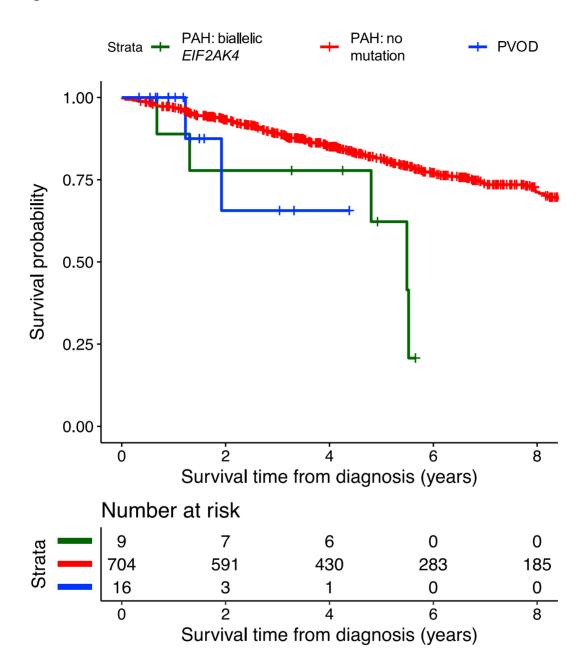
**Supplemental Table 13.** Sensitivity analysis including only prospectively recruited UK patients. Cox proportional hazards model assessing time to death. Number of patients = 608. Events = 95.

<sup>\*</sup> compared to the PAH biallelic EIF2AK4 mutation carriers

<sup>\*</sup> compared to the PAH biallelic *EIF2AK4* mutation carriers

# **Supplemental Figures**

Figure S1



# **Supplemental Figure Legends:**

Figure S1: Kaplan – Meier survival curves showing survival time (time to death) for patients with a clinical diagnosis of PAH or PVOD.

## **Supplemental References**

- 1. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GR, Thormann A, Flicek P and Cunningham F. The Ensembl Variant Effect Predictor. *Genome Biol.* 2016;17:122.
- 2. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, O'Donnell-Luria AH, Ware JS, Hill AJ, Cummings BB, Tukiainen T, Birnbaum DP, Kosmicki JA, Duncan LE, Estrada K, Zhao F, Zou J, Pierce-Hoffman E, Berghout J, Cooper DN, Deflaux N, DePristo M, Do R, Flannick J, Fromer M, Gauthier L, Goldstein J, Gupta N, Howrigan D, Kiezun A, Kurki MI, Moonshine AL, Natarajan P, Orozco L, Peloso GM, Poplin R, Rivas MA, Ruano-Rubio V, Rose SA, Ruderfer DM, Shakir K, Stenson PD, Stevens C, Thomas BP, Tiao G, Tusie-Luna MT, Weisburd B, Won HH, Yu D, Altshuler DM, Ardissino D, Boehnke M, Danesh J, Donnelly S, Elosua R, Florez JC, Gabriel SB, Getz G, Glatt SJ, Hultman CM, Kathiresan S, Laakso M, McCarroll S, McCarthy MI, McGovern D, McPherson R, Neale BM, Palotie A, Purcell SM, Saleheen D, Scharf JM, Sklar P, Sullivan PF, Tuomilehto J, Tsuang MT, Watkins HC, Wilson JG, Daly MJ and MacArthur DG. Analysis of protein-coding genetic variation in 60,706 humans. *Nature*. 2016;536:285-91.
- 3. Zarrei M, MacDonald JR, Merico D and Scherer SW. A copy number variation map of the human genome. *Nat Rev Genet*. 2015;16:172-83.
- 4. Hothorn T, Hornik K, Wiel MAvd and Zeileis A. A Lego System for Conditional Inference. *The American Statistician*. 2012;60:257-263.
- 5. Therneau T and Grambsch P. *Modeling Survival Data: Extending the Cox Model.* 1 ed. New York: Springer-Verlag 2000.
- 6. Grambsch P and Therneau H. Proportional hazards tests and diagnostics based on weighted residuals. *Biometrika*. 1994;81:515-526.
- 7. Collett D. *Modelling Survival Data in Medical Research.* 3rd ed. London: Chapman & Hall/CRC; 2014.