

This is a repository copy of TRPM2: Shredding the mitochondrial network..

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/121498/

Version: Accepted Version

Article:

Li, F, Abuarab, N and Sivaprasadarao, A orcid.org/0000-0002-6755-3502 (2017) TRPM2: Shredding the mitochondrial network. Channels, 11 (6). pp. 1-3. ISSN 1933-6950

https://doi.org/10.1080/19336950.2017.1376982

© 2017, Taylor & Francis. This is an Accepted Manuscript of an article published by Taylor & Francis in Channels on 6 September 2017, available online: http://dx.doi/10.1080/19336950.2017.1376982 Uploaded in accordance with the publisher's self-archiving policy

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/







ISSN: 1933-6950 (Print) 1933-6969 (Online) Journal homepage: http://www.tandfonline.com/loi/kchl20

TRPM2: Shredding the mitochondrial network

Fangfang Li, Nada Abuarab & Asipu Sivaprasadarao

To cite this article: Fangfang Li, Nada Abuarab & Asipu Sivaprasadarao (2017): TRPM2: Shredding the mitochondrial network, Channels, DOI: 10.1080/19336950.2017.1376982

To link to this article: http://dx.doi.org/10.1080/19336950.2017.1376982



Accepted author version posted online: 06 Sep 2017.



Submit your article to this journal 🗗



Article views: 109



View related articles



View Crossmark data 🗹

Full Terms & Conditions of access and use can be found at http://www.tandfonline.com/action/journalInformation?journalCode=kchl20

TRPM2: Shredding the mitochondrial network

Fangfang Li¹, Nada Abuarab¹, and Asipu Sivaprasadarao^{1,2,*}

¹School of Biomedical Sciences, Faculty of Biological Sciences, University of Leeds, LS2 9JT,

Leeds, U.K.

²Multidisciplinary Cardiovascular Research Centre, University of Leeds, LS2 9JT, Leeds, U.K.

Corresponding author:

Professor Asipu Sivaprasadarao

School of Biomedical Sciences

G6.44d, Garstang Building

University of Leeds

Leeds

LS2 9JT UK

E-mail: a.sivaprasadarao@leeds.ac.uk

Autocommentary to: Li F, Munsey TS, Sivaprasadarao A. Asipu SivaprasadaraoTRPM2mediated rise in mitochondrial Zn2+ promotes palmitate-induced mitochondrial fission and pancreatic β-cell death in rodents. Cell Death Differ. 2017 Jul 28. doi: 10.1038/cdd.2017.118. PMID: 28753206 Submitted: August 30, 2017

Accepted: August 31, 2017

ACCEPTED MANUSCRIPT

Keywords

Mitochondrial dynamics, TRPM2, oxidative stress, reactive oxygen species, calcium, zinc, pancreatic β -cell death, diabetes, obesity, fatty acid

² ACCEPTED MANUSCRIPT

Mitochondria, traditionally known for their role in power (ATP) generation, are important signalling organelles. To support these vital functions, mitochondria need to undergo continuous remodelling- a process known as 'mitochondrial dynamics'¹. Failure to do so compromises their function, leading to a spectrum of age-related human diseases, including diabetes, cardiovascular (ischaemia), neuronal (stroke, Parkinson's and Alzheimer's) and renal diseases, and cancer². It is generally acknowledged that a rise in oxidative stress, characterised by an increased production of reactive oxygen species (ROS), is responsible for abnormal mitochondrial dynamics. However, how ROS signal abnormal mitochondrial dynamics was unclear.

Mitochondrial dynamics involves continuous fission and fusion of the mitochondrial tubular network^{1, 2}. Fission is initiated by the ER-assisted constriction of the mitochondrial tubules followed by their cleavage by Drp1 (dynamin-related protein1) and dynamin 2. Drp1 is normally localised to the cytoplasm where it needs to undergo modifications before it can be recruited to mitochondria. One such modification requires a Ca^{2+} signal from an undetermined source. By activating calcineurin, Ca^{2+} removes the inhibitory phosphate group from Drp1, thereby promoting its recruitment to mitochondria. Fission results in the generation of functional and dysfunctional fragments. The latter are removed by mitophagy. Functional mitochondrial fragments merge with the healthy mitochondrial network, with the help of mitofusin-1/2 (MFN-1/2) and OPA-1(Optic atrophy type 1), which catalyse the fusion of the outer and inner membranes of mitochondria respectively.

Previous studies have shown that free fatty acids (FFAs), whose levels increase in obesity and contribute to type 2 diabetes, cause extensive fragmentation of the mitochondrial network in pancreatic β -cells by stimulating cellular ROS production³. In our recent publication⁴, we tested

³ ACCEPTED MANUSCRIPT

the possibility that the rise in ROS would activate Ca^{2+} channels to provide the Ca^{2+} required for Drp1 recruitment to mitochondria. We focussed our attention on TRPM2 (transient receptor potential melastatin2) channels because these are activated by ROS and conduct cations including Ca^{2+5} .

Consistent with our prediction, inhibition of TRPM2 channels by chemical, siRNA and gene knockout approaches prevented FFA-induced mitochondrial fragmentation as well as β -cell death⁴. ROS required for mitochondrial fission are provided by the FFA-mediated activation of cytoplasmic NADPH-oxidase-2. At first, these findings seemed to support our initial hypothesis that TRPM2 activation provides the Ca²⁺ required for Drp1 recruitment and the subsequent mitochondrial fragmentation. However, we have decided to test the role of Zn²⁺ because our previous study demonstrated that Zn²⁺ chelation prevents TRPM2-mediated β -cell death⁶, and other studies suggested that mitochondrial fragmentation precedes β -cell death³. Surprisingly, chelating Zn²⁺ alone was sufficient to prevent mitochondrial fission. On probing further, we found that free Zn²⁺ found largely in the lysosomes was transferred to mitochondria. In a related study, we have demonstrated that TRPM2-mediated Ca²⁺ entry causes lysosomal membrane permeabilisation and Zn²⁺ release⁷. Although we do not know how this Zn²⁺ is transferred to mitochondrial membrane potential, promoting Drp1 recruitment to mitochondria and β -cell apoptosis.

This study has thus led to the discovery of a novel signalling pathway where Ca^{2+} and Zn^{2+} collaborate to transduce the signal from FFA to the mitochondria of pancreatic β -cells to cause cell death (Figure 1). Importantly, the two ions facilitate communication between different cellular compartments. Thus extracellular Ca^{2+} enters the cytoplasm via ROS-activated plasma

⁴ ACCEPTED MANUSCRIPT

membrane TRPM2 channels. The resultant rise in cytosolic Ca^{2+} triggers escape of lysosomal free Zn^{2+} to mitochondria. Zn^{2+} , being an inhibitor of the electron transport chain, leads to the loss of mitochondrial membrane potential required for the recruitment cytoplasmic Drp1 to mitochondria. Drp1 then induces excessive mitochondrial fission, leading to increased apoptotic β -cell death.

These new findings raise a number of unanswered questions: (1) How does the rise in cytosolic Ca^{2+} induce lysosomal Zn^{2+} release? (2) How does Zn^{2+} enter mitochondria? Is it via the mitochondrial uniporter (MCU) or other transport molecules? (3) How does Zn²⁺ promote Drp1 recruitment? Which of the multiple posttranslational mechanisms required for mitochondrial Drp1 recruitment does Zn²⁺ affect? (4) Does Drp-1 recruitment to mitochondria involve Ca²⁺ signalling? If so, where does this Ca²⁺ come from? Interestingly, a recent study suggested Ca²⁺calcineurin signalling occurs at the ER-mitochondria junction.⁸ Answers to these key questions are important if we are to understand how the oxidative stress signals are translated into mitochondrial fragmentation, which is a common feature of most age-related human illnesses. In conclusion, our findings highlight a previously unappreciated role for ionic signalling, in particular a role for Zn²⁺ signalling, in mitochondrial dynamics. Furthermore, they suggest that ionic signalling is not limited to communication between just two organelles (for example, between the plasma membrane and the ER, and mitochondria and the ER), but could involve multiple organelles. Our findings emphasise the importance of an integrated approach to investigate how abnormal inter-organelle signalling can affect organelle homeostasis and contribute to human diseases. Such approaches may reveal hitherto unknown therapeutic targets

that may be common to a wide range of human diseases.

⁵ ACCEPTED MANUSCRIPT

References

Friedman JR, Nunnari J. Mitochondrial form and function. Nature 2014; 505:335-43;
PMID:24429632; ; doi. 10.1038/nature12985

Archer SL. Mitochondrial dynamics--mitochondrial fission and fusion in human diseases.
N Engl J Med 2013; 369:2236-5110. PMID:24304053; doi:10.1056/NEJMra1215233

3. Molina AJ, Wikstrom JD, Stiles L, Las G, Mohamed H, Elorza A, et al. Mitochondrial networking protects beta-cells from nutrient-induced apoptosis. Diabetes 2009; 58:2303-15; PMID:19581419; doi.10.2337/db07-1781

4. Li F, Munsey TS, Sivaprasadarao A. TRPM2-mediated rise in mitochondrial Zn2+ promotes palmitate-induced mitochondrial fission and pancreatic beta-cell death in rodents. Cell Death Differ 201710.1; PMID:28753206; doi.10.1038/cdd.2017.118

5. Sumoza-Toledo A, Penner R. TRPM2: a multifunctional ion channel for calcium signalling. Journal of Physiology-London 2011; 589:1515-2510; PMID:21135052; doi. 10.1113/jphysiol.2010.201855

 Manna PT, Munsey TS, Abuarab N, Li F, Asipu A, Howell G, et al. TRPM2 mediated intracellular Zn2+ release triggers pancreatic beta cell death. The Biochemical journal 2015; 466:537-46; PMID:25562606;doi.10.1042/BJ20140747

7. Abuarab N, Munsey TS, Jiang LH, Li J, Sivaprasadarao A. High glucose-induced ROS activates TRPM2 to trigger lysosomal membrane permeabilization and Zn2+-mediated mitochondrial fission. Sci Signal 2017; 10;PMID:28765513;doi.<u>10.1126/scisignal.aal4161</u>

⁶ ACCEPTED MANUSCRIPT

8. Mehta S, Aye-Han NN, Ganesan A, Oldach L, Gorshkov K, Zhang J. Calmodulincontrolled spatial decoding of oscillatory Ca2+ signals by calcineurin. Elife 2014; 3:e03765; PMID:25056880;doi. <u>10.7554/eLife.03765</u>

7 ACCEPTED MANUSCRIPT

Figure legend

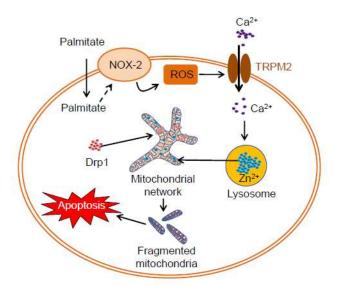


Figure 1. Schematic of how palmitate induces mitochondrial fragmentation in pancreatic β cells. Palmitate activates NOX-2 (NADPH oxidase-2) to induce ROS (reactive oxygen species) production. ROS activates TRPM2 channel leading to extracellular Ca²⁺ (purple dots) entry, Ca²⁺-induced lysosomal Zn²⁺ (blue dots) transfer to mitochondria. Rise in mitochondrial Zn²⁺ induces Drp1 (red dots) recruitment, breakdown of the mitochondrial network and eventually β cell death.

⁸ ACCEPTED MANUSCRIPT