Investigating the structural compaction of biomolecules upon transition to the gas-phase using ESI-TWIMS-MS

Paul W. A. Devine1, Henry C. Fisher1, Antonio N. Calabrese1, Fiona Whelan2, Daniel R. Higazi3, Jennifer R. Potts2, David C. Lowe4, Sheena E. Radford\*1, Alison E. Ashcroft\*1

1Astbury Centre for Structural Biology, School of Molecular and Cellular Biology, University of Leeds, Leeds, LS2 9JT, UK

2Department of Biology, University of York, York, YO10 5DD, UK

3Ipsen Ltd. UK, Wrexham Industrial Estate, 9 Ash Road North, Wrexham, LL13 9UF, UK

4MedImmune, Sir Aaron Klug Building, Granta Science Park, Cambridge, CB21 6GH, UK

**Abstract**

Collision cross-section (CCS) measurements obtained from ion mobility spectrometry-mass spectrometry (IMS-MS) analyses often provide useful information concerning a protein’s size and shape and can be complemented by modelling procedures. However, there have been some concerns about the extent to which certain proteins maintain a native-like conformation during the gas-phase analysis, especially proteins with dynamic or extended regions. Here we have measured the CCSs of a range of biomolecules including non-globular proteins and RNAs of different sequence, size and stability. Using travelling wave IMS-MS, we show that for the proteins studied the measured CCS deviates significantly from predicted CCS values based upon currently available structures. The results presented indicate that these proteins collapse to different extents varying on their elongated structures upon transition into the gas-phase. Comparing two RNAs of similar mass but different solution structures, we show that these biomolecules may also be susceptible to gas-phase compaction. Together the results suggest that caution is needed when predicting structural models based on CCS data for RNAs as well as proteins with non-globular folds.

**Introduction**

The advent of electrospray ionisation (ESI) transformed the field of mass spectrometry (MS) by providing the ability to routinely analyse not only large proteins but also non-covalently bound biomolecular complexes. In the three decades since this development, there has been a significant body of literature providing evidence of the native-like state of biomolecules measured by both ESI-MS and, more recently, ESI-ion mobility spectrometry-MS (ESI-IMS-MS) [1-3].

Ion mobility spectrometry (IMS) is a separation technique based on the gas-phase mobility of ions as they travel, under the influence of a weak electric current, through a drift tube filled with an inert gas [4-6]Department of Chemistry & Biochemistry, University of California, Santa Barbara, CA 93106-9510, United States

\*Tel.: +1 805 893 2893; fax.: +1 805 893 8703. Email: [ude.bscu.mehc@srewob](mailto:dev@null) (M.T. Bowers).

. Ions are separated based on their charge and shape: briefly, compact ions travel faster than extended ions carrying the same number of charges, whilst ions with a high number of charges travel faster than ions carrying a lower number of charges derived from the same precursor molecules. When coupled with MS, the data output is a 3D array of m/z *vs* intensity *vs* IMS drift time. The IMS drift time for ions can be converted to collision cross-section (CCS) directly if the IMS drift tube is a linear one [6-8], or indirectly following a calibration procedure [9-12] if the drift tube is of a travelling wave (TW) [13] design. The CCS of an ion corresponds to the averaged rotational 2D projection of the biomolecule’s 3D structure. Hence, ESI-IMS-MS is a unique and powerful tool that can separate and characterise biomolecules, providing both mass and shape (via CCS) information on individual species within an ensemble in a single, rapid, experiment. Indeed, ESI-IMS-MS has been employed to study the 3D architecture and conformational properties of many proteins and non-covalently bound biomolecular complexes [4-6, 9, 7, 8, 14-22].

In 1997, Joseph Loo stated there are three camps of opinion concerning the retention of native protein structure upon transition into the gas-phase: “believers, nonbelievers and undecided” [2], and quite possibly he was correct to hint at caution because, despite the high number of successes reported, there has been a slow, low level emergence of literature demonstrating the “collapse” of certain proteins upon transition into the gas-phase [23-25], one key example being antibodies [26-28]. Here, by systematic analysis of different non-globular proteins and RNA molecules using ESI-TWIMS-MS, we provide evidence of compaction in the gas-phase, highlighting a potential caveat in studying these specific biomolecules using this technique. The degree of compaction has been revealed by comparing the CCS values estimated from the ESI-IMS-MS data with CCS values calculated from the PDB structures of these biomolecules and also, in the case of the proteins, with *in vacuo* Molecular Dynamics (MD) simulations.

**Methods**

**Biomolecules**

The monoclonal antibody, mAb1, was supplied by Medimmune (Cambridge, UK); (I27)5 was expressed recombinantly as described elsewhere [29] with amino acid linkers in between each sub-unit (See Supporting Information Table S1); POTRAs were cloned and expressed recombinantly as described elsewhere [30]; a SasG construct (G51-G57-Strep-CysCys) was produced as described previously [31]. The RNA molecules were purchased from Integrated DNA Technologies BVBA, Leuven, Belgium.

**Protein mass spectrometry analyses**

All nanoESI-TWIMS-MS protein measurements were carried out using a Synapt HDMS mass spectrometer (Waters Corp., Wilmslow, UK). Samples were introduced to the mass spectrometer using in-house pulled borosilicate capillaries (Sutter Instrument Company, Novato, CA, USA) coated with palladium using a sputter coater (Polaron SC7620, Quorum Technologies Ltd, Kent, UK). All protein samples were analysed in positive ESI mode. The *m/z* scale was calibrated using 10 mg/mL aqueous caesium iodide (CsI) clusters across the acquisition range (typically *m/z* 500-15,000).

Protein samples were dialysed into 150 mM aqueous ammonium acetate before being infused into the Synapt G1 instrument. nESI-MS and nESI-TWIMS-MS experiments were conducted under the following settings: capillary voltage, 1.5 kV; sample cone, 30 V; extraction cone, 4 V; source temperature, 60-80 ⁰C; backing pressure, 3.0-5.0 mBar; trap voltage, 10-40 V; trap/transfer gas flow, 1.5 mL/min; IMS nitrogen gas flow, 20 mL/min, IMS wave height (ramped), 5-30 V and travelling wave speed, 300 ms.

All data were processed and analysed with the MassLynx v4.1 and Driftscope software, supplied with the mass spectrometer.

**ESI-TWIMS-MS Collision Cross-Section (CCS) calibrations for proteins**

ESI-TWIMS-MS experiments were carried out on a Synapt HDMS mass spectrometer using travelling wave IMS. Calibration of the travelling wave drift cell was carried out using a previously published method [11]. The calibrant proteins used were: beta-lactoglobulin, concanavalin A, alcohol dehydrogenase, and pyruvate kinase, taken from the Clemmer/Bush database [12]. Calibrant proteins were dissolved at a concentration of 10 µM in 200 mM ammonium acetate before being analysed under the same conditions as the protein analytes.

Calibrant proteins were corrected for mass-dependant flight time using Equation 1 [11]:

Equation 1

Where t’D is the corrected drift time, tD the measured drift time of the analyte, *m/z* the mass-to-charge ratio of the ion, and CEDC the enhanced duty cycle (EDC) delay coefficient of the instrument (in this case 1.57). The corrected drift times were plotted against the reduced cross-sections (Ω’) as outlined in [11], and the plot fitted to a linear relationship (Equation 2):

Equation 2

where A is a fit determined constant and X the exponential factor.

The calibrations were converted to linear plots to allow for straightforward extrapolation for measurements of unknown proteins and complexes. For this, a new corrected drift time was calculated using Equation 3:

Equation 3

where µ is the reduced mass of the ion.

The new corrected drift time (t’’D) was then plotted against the cross-section of the calibrant proteins (taken from the database [12]) to generate the calibration plots (see Supporting Information: Figures S1, S2, and S3).

**RNA mass spectrometry analyses**

All nanoESI-TWIMS-MS RNA measurements were carried out using a Synapt G2-S mass spectrometer (Waters Corp., Wilmslow, UK). Samples were introduced to the mass spectrometer using in-house pulled borosilicate capillaries (Sutter Instrument Company, Novato, CA, USA) coated with palladium using a sputter coater (Polaron SC7620, Quorum Technologies Ltd, Kent, UK). All RNA samples were analysed in negative ESI mode. The *m/z* scale was calibrated using 10 mg/mL aqueous caesium iodide (CsI) clusters across the acquisition range (typically *m/z* 500-15,000).

On receipt, the two 35-nucleotide RNAs (PDB structures 2PCV and 2DRB) were diluted to 200 µM in RNAse-free MilliQ water, separated into 100 µL aliquots and stored at -80 oC. When ready for analysis, these samples were diluted with 50 mM ammonium acetate to a concentration of 40 µM, desalted using spin columns (BioRad, Hemel Hempstead, UK) and then diluted to 8 µM with 50 mM ammonium acetate. nESI-TWIMS-MS experiments were conducted under the following settings: capillary voltage, -0.8 kV; sample cone, 30 V; source temperature, 60-80 ⁰C; IMS sheath gas, He 2.42e-2 bar; IMS buffer gas, Ar 3.01 bar; IMS wave height 40 V; IMS wave velocity 400-750 m/s per scan.

**ESI-TWIMS-MS Collision Cross-Section (CCS) calibration for RNAs**

ESI-TWIMS-MS experiments for the RNAs were carried out on a Synapt G2-S mass spectrometer using travelling wave ion mobility separation with negative ionisation electrospray. Calibration of the travelling wave drift cell was carried out using the method described previously in this document for protein samples but with an enhanced duty cycle delay coefficient (CEDC) of 1.41. The calibrant was a DNA polythymine of ten nucleotides (d[T]10) whose CCS had been measured and reported by Clemmer [32] (see Supporting Information: Figure S4).

**Theoretical CCS calculation**

MOBCAL software was used to calculate the theoretical CCSs for the samples studied and was implemented using a Linux operating system. The MOBCAL Projection Approximation value [33] was used to generate the Projection Superposition Approximation (PSA) as outlined in [34]. Equation 4 was used for this:

Equation 4

***In vacuo* Molecular Dynamics (MD) simulations**

MD simulations were run using the NAMD software (NAMD 2.9), using the CHARMM force field [35]. Structures were simulated in a solvent free system. For the simulation, a constant temperature of 300 K with Langevin thermostat was used and a time-step of 2.0 fs with a radial cut-off distance of 12 Å used throughout. Energy minimization *in vacuo* was implemented for a total of 0.5 ns before an equilibration of 10 ns, the cut-off distance, force field and time step remained as described above throughout the simulation. Visual Molecular Dynamics (VMD) [36] was then used to visualise the simulation and individual frames were saved as PDB coordinates in order to compute the CCS using MOBCAL. The VMD software was also used to calculate the root mean square deviation (RMSD) and radius of gyration (Rg). Analysis of the root-mean-square deviation (RMSD) revealed whether a protein had equilibrated by the end of the 10 ns simulation; any sample that had not finished equilibrating was re-submitted for a further 10 ns until equilibration was reached. The NAMD and VMD software was operated under a Linux operating system.

**Results and Discussion**

**Insights into the gas-phase collapse of monoclonal antibodies**

Using ESI-TWIMS-MS under non-denaturing conditions to characterise an IgG1 monoclonal antibody, mAb1, we observed that, despite presenting a narrow ESI charge state distribution (+21 to +25 ions) usually indicative of a “native-like” protein, the experimentally estimated CCS value of the lowest charge state (68.2 nm2) was significantly lower (32.4 %) than the computationally determined CCS (101 nm2) based on the published structure (PDB = 1IGY [37]) (**Figures 1a i, ii, iii**). Similar behaviour of monoclonal antibodies (mAbs) has been reported by others [27, 28], and Pacholarz *et al*. carried out *in vacuo* MD simulations to interrogate the observed compaction of IgG molecules in the gas-phase, demonstrating that the protein likely collapsed around the hinge region in between the fragment antigen-binding (Fab) and the fragment crystallisable(Fc) regions [28].

Molecular modelling is a useful tool to aid the study of biomolecules in the gas-phase. Although CCSs measured using ESI-TWIMS-MS methods can be compared directly to solved X-ray crystal or NMR structures from the Protein Data Bank (PDB), it is becoming clearer that this is not suitable for all proteins. For example, the conditions used to crystallise proteins can be very different to the conditions used for mass spectrometric analysis. Further, some proteins are inherently flexible or disordered, and may not have a PDB structure with which to compare the measured CCS, and additionally a subset of structures in the PDB consist only of fragments of the full protein in question. *In vacuo* modelling, therefore, allows us to achieve a glimpse of how such proteins may behave within the gas-phase. Adopting a similar *in vacuo* MD simulation approach as used by [28], we also observe a collapse around the hinge region of mAb1 such that the measured CCS of the mAb is substantially less than both the predicted CCS from its crystal structure [37] and the *in vacuo* MD simulation (**Figure 1a iii**).

To understand the role of the hinge region and determine whether this flexible linker was the main attributor to the collapse observed, we released the Fab and Fc regions of mAb1 using Lys-C proteolysis and analysed the two fragments independently (**Figures 1b, 1c**). The CCS values determined by ESI-TWIMS-MS, estimated from the PDB co-ordinates, and indicated by *in vacuo* MD simulations are compared for both the Fab and the Fc regions (**Figures 1b** and **1c**, respectively). The MD simulations indicated that both proteins collapsed to some extent *in vacuo* compared with their crystal structures, with the Fab region collapsing 11 % compared with the 17 % collapse of the Fc region. Furthermore, the CCS of the Fab region measured by ESI-TWIMS-MS was closer in agreement to the CCS predicted from its PDB structure than with its equilibrated collapsed MD structure, while the CCS of the Fc region measured by ESI-TWIMS-MS was closer to that of its collapsed MD structure than with its crystal structure. Although both of these fragments consist of four Ig domains, the Fc region retains the majority of the hinge region, supporting the notion that the flexible hinge plays a prominent role in the gas-phase collapse observed.

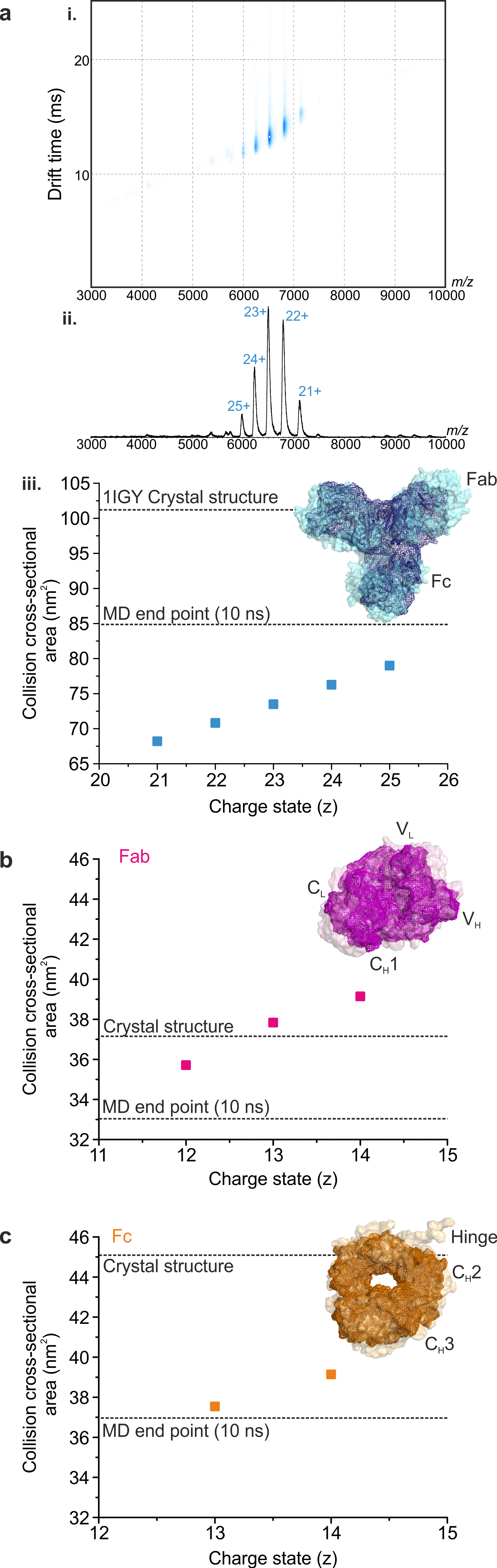


Figure 1: Monoclonal antibodies collapse around the flexible hinge region in the gas-phase. (a) ESI-TWIMS-MS (i) Driftscope plot of *m/z* *vs* TWIMS drift time *vs* intensity and (ii) mass spectrum for mAb1 with (iii) the TWIMS CCSs for each charge state compared with the CCS calculated from the protein’s PDB co-ordinates and the CCS estimated from the end-point of the *in vacuo* MD simulation. The ESI-TWIMS-MS CCS data for (b) the Fab and (c) the Fc regions of mAb1 isolated by lys-C digestion: the calculated CCS values from the PDB co-ordinates and those after *in vacuo* MD equilibration are reported for both proteins (dotted lines). Insets show the PDB structures (transparent surface) alongside the *in vacuo* MD equilibrated structures (solid wire frame).

**Investigating the gas-phase collapse of other non-globular proteins**

To investigate the generality of the role of flexible hinge regions in gas-phase protein collapse, using ESI-TWIMS-MS we analysed an I27 concatamer, (I27)5 [29] (**Figure 2a**) and the POTRA domains from BamA [38] (**Figure 2b**). (I27)5 is a mechanically robust pentamer, whose folded Ig subunits are connected by flexible linkers of 4-6 amino acids. This construct is used widely for AFM and mechanical stability studies [29, 39]. Furthermore, poly-Ig domains as well as I27 polyproteins have been shown to be flexible in solution and can adopt various conformations as revealed by electron microscopy [40, 39, 41]. The POTRA domains were chosen as, similar to (I27)5, the protein consists of five subunits (POTRAS 1-5) connected by short linker regions [42, 43].

The mass spectrum of (I27)5 indicated a narrow charge state distribution (13+ to 16+ ions) (**Figure 2a i**). As neither a crystal nor NMR structure was available for (I27)5, a model was built based on the solution structure of the I27 monomer (1TIT, [44]) and building in the 4-6 residue linker regions (see Supporting Information) (**Figure 2a i**). This enabled a theoretical CCS for the five-domain construct to be established and formed the starting point for the *in vacuo* MD simulations (see Supporting Information). The measured CCS for (I27)5 (39.8 nm2) [45] is lower than both the modelled value predicted for the native structure (63.1 nm2) and the MD simulation end point (49.4 nm2) (**Figure 2a ii**). Upon *in vacuo* minimisation and equilibration, the protein undergoes compaction then collapses around the flexible linker regions between the individual subunits, which is reflected by the CCS at the end of the simulation.

ESI-TWIMS-MS analysis of the combined POTRA domains from BamA again produced a mass spectrum with a narrow charge state distribution (12+ to 16+ ions) (**Figure 2b i**). The ESI-TWIMS-MS data indicate that the CCS (35.1 nm2) obtained for the lowest charge state ions (+12) is closer to the predicted CCS of the *in vacuo*-equilibrated structure (37.1 nm2) than the CCS value predicted from the crystal structure (5D0O; 45.3nm2) (**Figure 2b ii**). The MD collapse observed for the POTRA domains is attributable to compaction around the short hinge regions between the individual domains, as well as to an overall collapse with POTRA1 moving towards POTRA5, resulting in a more ring-like structure in the equilibrated molecule (**Figure 2b iii**).

Together, the data presented for mAb1, (I27)5 and the POTRA domains suggest that non-globular proteins with flexible linker or hinge regions are susceptible to gas-phase collapse. To determine how linear, elongated molecules without any distinct linker regions behave upon transition to the gas-phase, we studied the protein SasG (**Figure 2c**). SasG consists of repeats of two domains (G5 and E), in which the C-terminus of any given domain is directly connected to the N-terminus of the subsequent domain (**Figure 2c**). Furthermore, SasG (G51-G57) has been shown to form long, elongated fibrillary structures that maintain a highly extended conformation in solution, with no evidence of compaction [31]. The ESI-MS data indicate a native-like conformation, centred on the 20+ and 21+ charge state ions, together with a highly charged, more unfolded conformation (centred on the 48+ charge state ions) (**Figure 2c i**). The ESI-TWIMS-MS CCS of the compact conformation was measured at 57.7 nm2 (18+ ions). In comparison, the predicted CCS based on the structure obtained from SAXS data [43] was 137.8 nm2, while the *in vacuo* MD simulations indicate that the protein collapses in the absence of solvent to a species with a CCS of 80.7 nm2 (**Figure 2c ii, iii**). Thus, an elongated linear protein, with no obvious linker or hinge regions can also undergo significant compaction in the gas-phase.

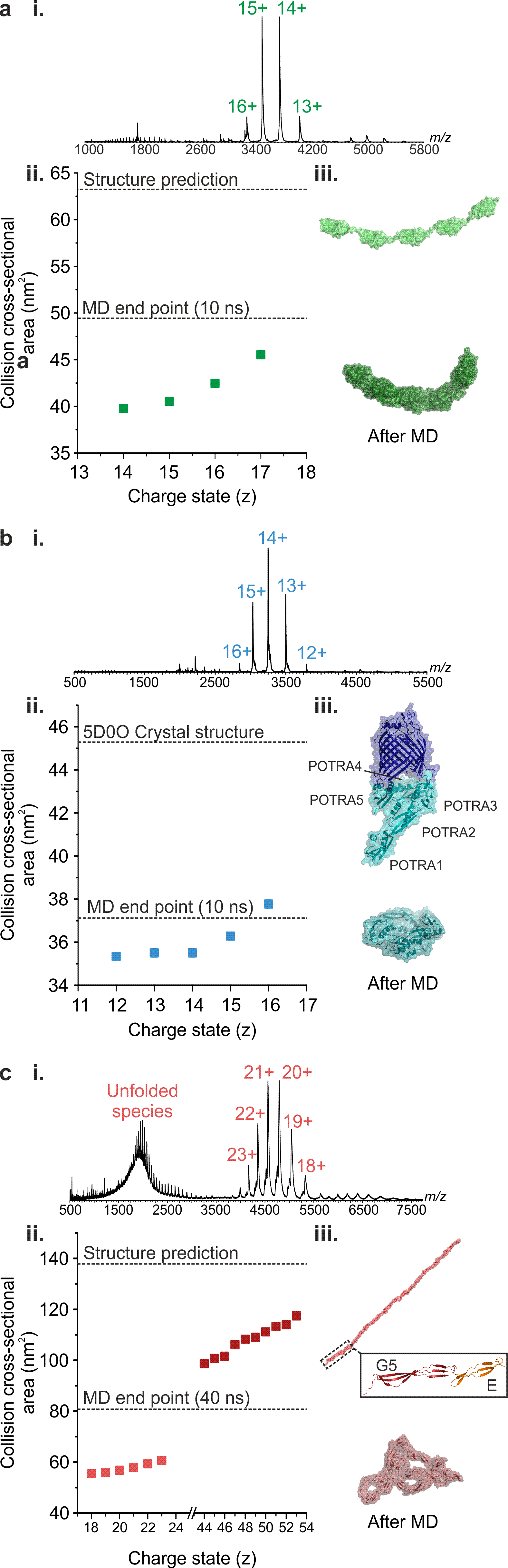


Figure 2: Compaction in the gas-phase is observed for a range of non-globular proteins: (a) (I27)5, (b) BamA POTRA domains and (c) SasG. ESI-TWIMS-MS (i) mass spectra and (ii) CCS data for each protein are shown together with the theoretical CCS values before (from the PDB co-ordinates) and after *in vacuo* MD equilibration (dotted lines). The starting and equilibrated structures are shown for comparison.

**Gas-phase collapse of other biomolecules**

Recent ESI-TWIMS-MS studies indicated that the DNA duplex (d(GCGAAGC)) is a dynamic ensemble in the gas-phase [46], in contrast to earlier work on G-complexes of ≥20 nucleotides which suggested that their chemical topology remained unaltered in the gas-phase [47]. Here, we carried out ESI-TWIMS-MS analyses on two RNAs, each of 35 nucleotides and of very similar mass, but different sequences and secondary structures (2PCV [48]; 11,217 Da and 2DRB ([49]; 11,219 Da) (**Table 1;** **Figure 3a**) to determine if their 3D structures were preserved in the gas-phase and hence if it was possible to differentiate between the two. An NMR solution structure has been published for 2PCV [48] and a crystal structure for 2DRB [49] and these were used to calculate CCS values (**Figure 3b**).

|  |  |  |  |
| --- | --- | --- | --- |
| **RNA** | **Mass** | **Mobcal PA** | **Sequence** |
| 2PCV | 11,217 Da | 1,445 Å2 | 5’- GGA CCC GCC ACU GCA GAG AUG CAA UCC AGU GGU CC -3’ |
| 2DRB | 11,219 Da | 1,146 Å2 | 5’- GGC CCG GGG CGG UUC GAU UCC GUU CUG GGC CAC CA -3’ |

**Table 1:** The molecular masses, sequences and predicated CCSs of the two 35-nucleotide RNAs: 2PCV and 2DRB.

ESI-TWIMS-MS analysis of the RNAs yielded identical CCSs for all of the corresponding charge state ions (4- to 7- ions; CCS ~10-11 nm2) (**Figure 3b**). Comparing the TWIMS CCS values with the CCSs estimated from the PDB structures, the TWIMS CCS data were significantly lower than the predicted values for either 2PCV (14.46 nm2) or 2DRB (11.46 nm2). For example, in the case of the 5- ions, TWIMS CCSs of 10.21 nm2 for 2PCV and 10.16 nm2 for 2DRB were measured, thus indicating both RNAs undergo gas-phase collapse. It may be argued that the ESI-MS solution conditions (50 mM aqueous ammonium acetate) differ from the crystallography conditions used for 2DRB (50 mM HEPES, 80 mM ammonium sulfate (n.b. some crystals were detected in the absence of the sulfate ions), 0.2 M tri-lithium citrate and 20% PEG4000 [49]) and from the NMR solution conditions used for 2PCV (5 mM cacodylate, 50 mM NaCl, and 0.1 mM EDTA [48]), and that this may have affect the CCS values obtained from the three biophysical techniques. While beyond the scope of this study, a systematic analysis of the effects of counterions, pH, oligonucleotide length and sequence on collapse in the gas-phase with parallel MD simulations [46, 50] could be informative to cast more light on the response of RNA molecules in the gas-phase in general. However here, the collapse of both of the RNAs to a similar degree in the gas-phase is evident.

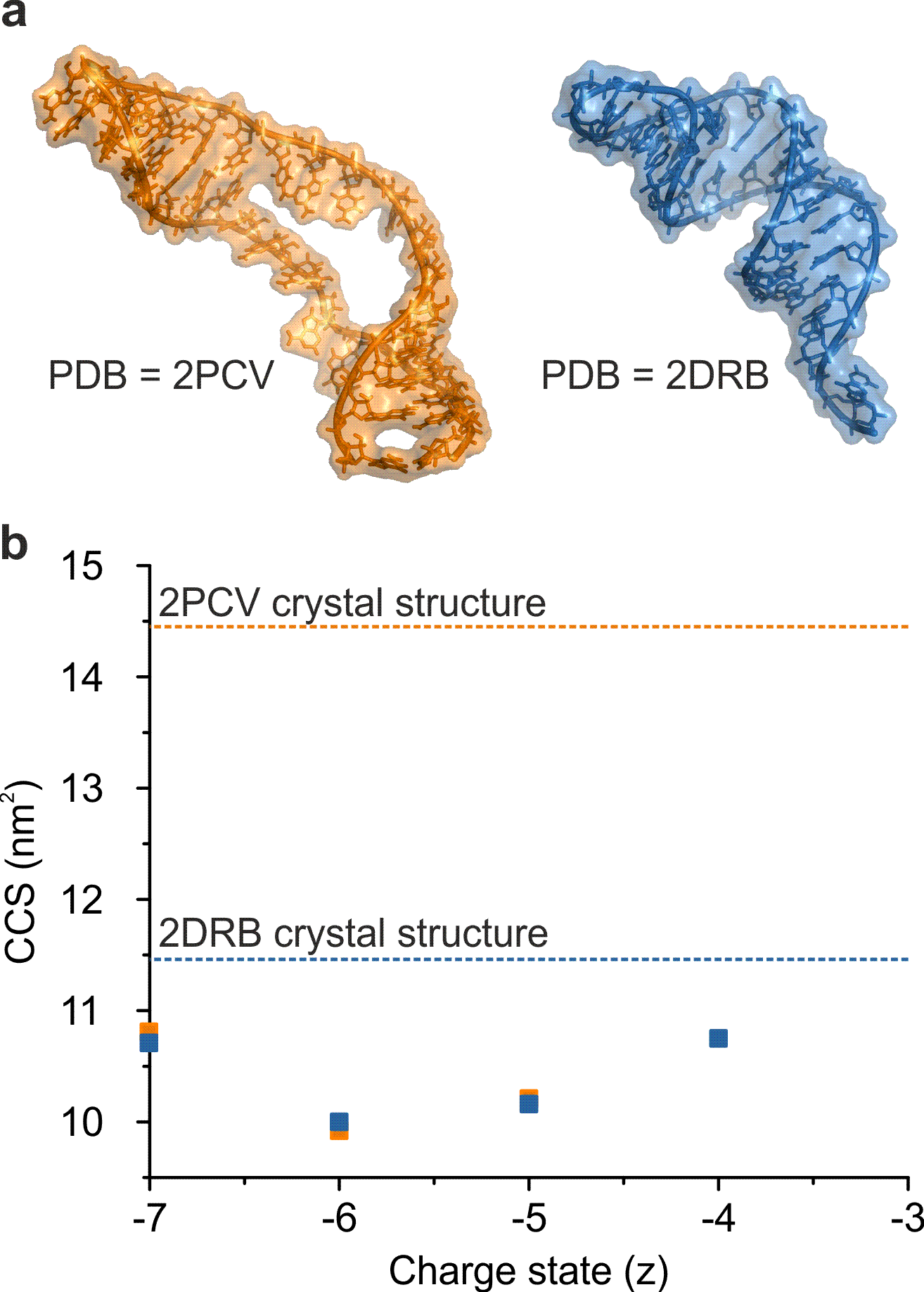
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Figure 3: Observed gas-phase collapse of RNAs. (a) The structures of two 35-nucleotide RNA molecules: 2PCV [48] (orange) and 2DRB [49] (blue). (b) ESI-TWIMS-MS CCS data for the 4- to 7- charge state ions of the two RNAs, together with the predicted values derived from the respective PDB co-ordinates of the RNAs RNA (dotted lines).

**Conclusion**

The question remains: can the solution structure of proteins be retained upon transfer into the gas-phase? For stable, globular proteins the answer is undoubtedly “yes”, backed by an impressive number of literature examples. However, here we have presented a small number of protein examples from our 14 years’ experience with ESI-IMS-MS where we have found that the CCS values measured underestimate the physical size of the solution structure and modelled data of the biomolecule under scrutiny. This phenomenon has been reported elsewhere in the case of antibodies [26-28], but here we have shown by studying isolated regions of an antibody that the Fc region, which contains the majority of the flexible hinge region, is more prone to gas-phase compaction than the Fab region. Other proteins we have identified that undergo gas-phase compaction include those with flexible hinge regions in between more structured domains, such as an engineered concatamer, (I27)5, in addition to the BamA complex with its extended array of POTRA domains. Other non-globular proteins such as SasG, an elongated linear protein, can also exhibit this behaviour. Gas-phase compaction is not limited to proteins, as illustrated with reference to two 35-nucleotide RNA molecules of similar mass but different shape. Both RNAs appeared from the ESI-TWIMS-MS data to be significantly smaller than expected from their 3D crystal or solution structures.

We do not intend this report to be perceived as a negative message to the use of ESI-TWIMS-MS. Indeed, the advantages of this technique far outweigh any disadvantages. However, there are certain classes of biomolecules for which due caution should be employed when interpreting the results.

**Acknowledgements**

We thank theBiotechnology and Biological Sciences Research Council (BBSRC) and Medimmune for funding PWAD (BB/J011819/1) and the Engineering and Physical Sciences Research Council (EPSRC) and GSK for funding HCF (EP/I501495/1). The Synapt HDMS mass spectrometer was funded by the BBSRC (BB/E012558/1) and the Synapt G2-S donated by Drs M. Morris and K. Giles, Waters UK Ltd. SER and AEA also acknowledge funding from the European Research Council under the European Union's Seventh Framework Programme (FP7/2007-2013; ERC grant 322408). We thank Drs D. R. Brockwell and B. Schiffin for providing (I27)5 and the BAM POTRA domains.

**References**

1. Leney, A. C., Heck, A. J.: Native Mass Spectrometry: What is in the Name? J. Am. Soc. Mass Spectrom., **28**, 5-13 (2017).

2. Loo, J. A.: Studying noncovalent protein complexes by electrospray ionization mass spectrometry. Mass Spectrom. Rev., **16**, 1-23 (1997).

3. Marcoux, J., Robinson, C. V.: Twenty years of gas phase structural biology. Structure, **21**, 1541-1550 (2013).

4. Bohrer, B. C., Merenbloom, S. I., Koeniger, S. L., Hilderbrand, A. E., Clemmer, D. E.: Biomolecule analysis by Ion Mobility Spectrometry. Annu. Rev. Anal. Chem., **1**, 1-10 (2008).

5. Bowers, M. T.: Ion mobility spectrometry: A personal view of its development at UCSB. Int. J. Mass Spectrom., **370**, 75-96 (2014).

6. Clemmer, D. E., Jarrold, M. F.: Ion Mobility measurements and their applications to clusters and biomolecules. Mass Spectrom., **32**, 577-592 (1997).

7. Mosier, P. D., Counterman, A. E., Jurs, P. C., Clemmer, D. E.: Prediction of peptide ion collision cross sections from topological molecular structure and amino acid parameters. Anal. Chem., **74**, 1360-1370 (2002).

8. Revercomb, H. E., Mason, E. A.: Theory of plasma chromatography/gaseous electrophoresis: a review. Anal. Chem., **47**, 970-983 (1975).

9. Knapman, T. W., Morton, V. L., Stonehouse, N. J., Stockley, P. G., Ashcroft, A. E.: Determining the topology of virus assembly intermediates using ion mobility spectrometry-mass spectrometry. Rapid Commun. Mass Spectrom., **24**, 3033-3042 (2010).

10. Smith, D. P., Knapman, T. W., Campuzano, I., Malham, R. W., Berryman, J. T., Radford, S. E., Ashcroft, A. E.: Deciphering drift time measurements from travelling wave ion mobility spectrometry - mass spectrometry studies. Eur. J. Mass Spectrom., **15**, 113-130 (2009).

11. Ruotolo, B. T., Benesch, J. L., Sandercock, A. M., Hyung, S. J., Robinson, C. V.: Ion mobility-mass spectrometry analysis of large protein complexes. Nat. Protoc., **3**, 1139-1152 (2008).

12. Bush, M. F., Hall, Z., Giles, K., Hoyes, J., Robinson, C. V., Ruotolo, B. T.: Collision cross sections of proteins and their complexes: a calibration framework and database for gas-phase structural biology. Anal. Chem., **82**, 9557-9565 (2010).

13. Giles, K., Pringle, S. D., Worthington, K. R., Little, D., Wildgoose, J. L., Bateman, R. H.: Applications of a travelling wave-based radio-frequency-only stacked ring ion guide. Rapid Commun. Mass Spectrom., **18**, 2401-2414 (2004).

14. Bernstein, S. L., Dupuis, N. F., Laz, N. D., Wyttenbach, T., Condron, M. M., Bitan, G., Teplow, B. D., Shea, J., Ruotolo, B. T., Robinson, C. V., Bowers, M. T.: Amyloid-β protein oligomerization and the importance of tetramers and dodecamers in the aetiology of Alzheimer's disease. Nat. Chem., **1:326**, (2009).

15. Beveridge, R., Chappius, Q., Macphee, C., Barran, P.: Mass spectrometry methods for intrinsically disordered proteins. Analyst, **138**, 32-42 (2013).

16. Hopper, J. T., Yu, Y. T., Li, D., Raymond, A., Bostock, M., Liko, I., Mikhailov, V., Laganowsky, A., Benesch, J. L., Caffrey, M., Nietlispach, D., Robinson, C. V.: Detergent-free mass spectrometry of membrane protein complexes. Nat. Methods, **10**, 1206-1208 (2013).

17. Konijnenberg, A., Butterer, A., Sobott, F.: Native ion mobility-mass spectrometry and related methods in structural biology. Biochim. Biophys. Acta, **1834**, 1239-1256 (2013).

18. Laganowsky, A., Reading, E., Allison, T. M., Ulmschneider, M. B., Degiacomi, M. T., Baldwin, A. J., Robinson, C. V.: Membrane proteins bind lipids selectively to modulate their structure and function. Nature, **510**, 172-175 (2014).

19. Ruotolo, B. T., Giles, K., Campuzano, I., Sandercock, A. M., Bateman, R. H., Robinson, C. V.: Evidence for macromolecular protein rings in the absence of bulkwater. Science, **310**, 1658-1661 (2005).

20. Schmidt, M., Zhou, M., Marriott, H., Morgner, N., Politis, A., Robinson, C. V.: Comparative cross-linking and mass spectrometry of an intact F-type ATPase suggest a role for phosphorylation. Nat. Commun., **4**, 1985 (2013).

21. Van Dujin, E., Barendregt, A., Synowsky, S., Versluis, C., Heck, A. J.: Chaperonin complexes monitored by ion mobility mass spectrometry. J. Am. Chem. Soc., **131**, 1452-1459 (2009).

22. Zhou, M., Politis, A., Davies, R. B., Liko, I., Wu, K.-J., Stewart, A., Stock, D., Robinson, C. V.: Ion mobility mass spectrometry of a rotary ATPase reveals ATP-induced reduction in conformational flexibility. Nat. Chem., **6**, 208-215 (2014).

23. Hogan, C. J., Ruotolo, B. T., Robinson, C. V., Fernandez de la Mora, J.: Tandem differential mobility analysis-mass spectrometry reveals partial gas-phase collapse of the GroEL complex. J. Phys. Chem., **115**, 3614-3621 (2011).

24. Jenner, M., Ellis, J., Huang, W. C., Lloyd Raven, E., Roberts, G. C., Oldham, N. J.: Detection of a protein conformational equilibrium by electrospray ionisation-ion mobility-mass spectrometry. Angew. Chem. Int. Ed. Engl., **50**, 8291-8293 (2011).

25. Jurneczko, E., Barran, P. E.: How useful is ion mobility mass spectrometry for structural biology? The relationship between protein crystal structures and their collision cross sections in the gas phase. Analyst, **136**, 20-28 (2011).

26. Campuzano, I. D. G., Larriba, C., Bagal, D., Schnier, P. D.: Ion Mobility and Mass Spectrometry Measurements of the Humanized IgGk NIST Monoclonal Antibody. Chapter 4, ACS Symposium Series, Vol. 1202, State-of-the-Art and Emerging Technologies for Therapeutic Monoclonal Antibody Characterization Volume 3. Defining the Next Generation of Analytical and Biophysical Techniques, 75-112 (2015).

27. Debaene, F., Boeuf, A., Wagner-Rousset, E., Colas, O., Ayoub, D., Corvaia, N., Van Dorsselaer, A., Beck, A., Cianferani, S.: Innovative native MS methodologies for antibody drug conjugate characterization: High resolution native MS and IM-MS for average DAR and DAR distribution assessment. Anal. Chem., **86**, 10674-10683 (2014).

28. Pacholarz, K. J., Porrini, M., Garlish, R. A., Burnley, R. J., Taylor, R. J., Henry, A. J., Barran, P. E.: Dynamics of intact immunoglobulin G explored by drift-tube ion-mobility mass spectrometry and molecular modeling. Angew. Chem. Int. Ed. Engl., **53**, 7765-7769 (2014).

29. Brockwell, D. J., Beddard, G. S., Clarkson, J., Zinober, R. C., Blake, A. W., Trinick, J., Olmstead, P. D., Smith, D. A., Radford, S. E.: The effect of core destabilization on the mechanical resistance of I27. Biophys. J., **83**, 458-472 (2002).

30. Iadanza, M. G., Higgins, A. J., Schriffen, B., Calabrese, A. N., Brockwell, D. J., Ashcroft, A. E., Radford, S. E., Ranson, N. A.: Lateral opening in the intact beta-barrel assembly machinery captured by cryo-EM. Nat. Commun., **7:12865**, (2016).

31. Gruska, D. T., Whelan, F., Farrance, O. E., Fung, H. K., Paci, E., Jeffries, C. M., Svergun, D. I., Baldock, C., Baumann, C. G., Brockwell, D. J., Potts, J. R., Clarke, J.: Cooperative folding of intrinsically disordered domains drives assembly of a strong elongated protein. Nat. Commun., **6:7271**, (2015).

32. Hoagland, C. S., Liu, Y. H., Ellington, A. D., Pagel, M., Clemmer, D. E.: Gas-phase DNA: Oligothymine ion conformers. J. Am. Chem. Soc., **119**, 9051-9052 (1997).

33. Mesleh, M. F., Hunter, J. M., Shvartsburg, A. A., Schatz, G. C., Jarrold, M. F.: Structural information from oion mobility measurements: Effects of the long-range potential. J. Phys. Chem., **100**, 16082-16086 (1996).

34. Bleiholder, C., Wyttenbach, T., Bowers, M. T.: A novel projection approximation algorithm for the fast and accurate computation of molecular collision cross sections (I). Method. Int. J. Mass Spectrom., **308**, 1-10 (2011).

35. Brooks, B. R., Bruccoleri, R. E., Olafson, B. D., States, D. J., Swaminathan, S., Karplus, M.: CHARMM: A program for macromolecular energy, minimization, and dynamics calculations. J. Comput. Chem., **4**, (1983).

36. Humphrey, W., Dalke, D., Schulten, K.: VMD: visual molecular dynamics. J. Mol. Graph., **14**, 27-28 (1996).

37. Harris, L. J., Skaletsky, E., McPherson, A.: Crystallographic structure of an intact IgG1 monoclonal antibody. J. Mol. Biol., **275**, 861-872 (1998).

38. Fleming, P. J., Patel, D. S., Wu, E. L., Qi, Y., Yeom, M. S., Sousa, M. C., Fleming, K. G., Im, W.: BamA POTRA Domain Interacts with a Native Lipid Membrane Surface. Biophys. J., **110**, 2698-2709 (2016).

39. Oberhauser, A. F., Marszalek, P. E., Carrion-Vazquez, M., Fernandez, J. M.: Single protein misfolding events captured by atomic force microscopy. Nat. Struct. Biol., **6**, 1025-1028 (1999).

40. Li, H., Oberhauser, A. F., Redick, S. D., Carrion-Vazquez, M., Erickson, H. P., Fernandez, J. M.: Multiple conformations of PEVK proteins detected by single-molecule techniques. Proc. Natl. Acad. Sci. USA, **98**, 10682-10686 (2001).

41. von Castelmur, E., Marino, M., Svergun, D. I., Kreplak, L., Ucurum-Fotiadis, Z., Konarev, P. V., Urzhumtsev, A., Labeit, D., Labeit, S., Mayans, O.: A regular pattern of Ig super-motifs defines segmental flexibility as the elastic mechanism of the titin chain. Proc. Natl. Acad. Sci. USA, **105**, 1186-1191 (2008).

42. Gatzeva-Topalova, P. Z., Warner, L. R., Pardi, A., Sousa, M. C.: Structure and flexibility of the complete periplasmic domain of BamA: the protein insertion machine of the outer membrane. Structure, **18**, 1492-1501 (2010).

43. Knowles, T. J., Jeeves, M., Bobat, S., Dancea, F., McClelland, D., Palmer, T., Overduin, M., Henderson, I. R.: Fold and function of polypeptide transport-associated domains responsible for delivering unfolded proteins to membranes. Mol. Microbiol., **68**, 1216-1227 (2008).

44. Improta, S., Politou, A. S., Pastore, A.: Immunoglobulin-like modules from titin I-band: extensible components of muscle elasticity. Structure, **4**, 323-337 (1996).

45. Smith, D. P., Radford, S. E., Ashcroft, A. E.: Elongated oligomers in beta(2)-microglobulin amyloid assembly revealed by ion mobility spectrometry-mass spectrometry. Proceedings Of The National Academy Of Sciences Of The United States Of America, **107**, 6794-6798 (2010).

46. Arcella, A., Dreyer, J., Ippoliti, E., Ivani, I., Portella, G., Gabelica, V., Carloni, P., Orozco, M.: Structure and dynamics of oligonucleotides in the gas phase. Angew. Chem. Int. Ed. Engl., **54**, 467-471 (2015).

47. Baker, E. S., Bernstein, S. L., Gabelica, V., De Pauw, E., Bowers, M. T.: G-quadruplexes in telomeric repeats are conserved in a solvent-free environment. Int. J. Mass Spectrom., **253**, 225-237 (2006).

48. Jin, H., Loria, J. P., Moore, P. B.: Solution structure of an rRNA substrate bound to the pseudouridylation pocket of a box H/ACA snoRNA. Mol. Cell, **26**, 205-215 (2007).

49. Tomita, K., Ishitani, R., Fukai, S., Nureki, O.: Complete crystallographic analysis of the dynamics of CCA sequence addition. Nature, **443**, 956-960 (2006).

50. Li, R., Macnamara, L. M., Leuchter, J. D., Alexander, R. W., Cho, S. S.: MD Simulations of tRNA and Aminoacyl-tRNA Synthetases: Dynamics, Folding, Binding, and Allostery. Int. J. Mol. Sci., **16**, 15872-15902 (2015).