

This is a repository copy of *In vitro activities of MCB3681 and 8 comparators against Clostridium difficile isolates with known ribotypes and diverse geographical spread.*

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/111852/

Version: Accepted Version

Article:

Freeman, J, Pilling, S, Vernon, J et al. (1 more author) (2017) In vitro activities of MCB3681 and 8 comparators against Clostridium difficile isolates with known ribotypes and diverse geographical spread. Antimicrobial Agents and Chemotherapy, 61 (3). e02077-16. ISSN 0066-4804

https://doi.org/10.1128/AAC.02077-16

(c) 2016, Freeman et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Reuse

Unless indicated otherwise, fulltext items are protected by copyright with all rights reserved. The copyright exception in section 29 of the Copyright, Designs and Patents Act 1988 allows the making of a single copy solely for the purpose of non-commercial research or private study within the limits of fair dealing. The publisher or other rights-holder may allow further reproduction and re-use of this version - refer to the White Rose Research Online record for this item. Where records identify the publisher as the copyright holder, users can verify any specific terms of use on the publisher's website.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

| 1 | |
|----|--|
| 2 | In vitro activities of MCB3681 and 8 comparators against |
| 3 | Clostridium difficile isolates with known ribotypes and |
| 4 | diverse geographical spread |
| 5 | |
| 6 | *J Freeman, ^{1,2} S Pilling ² , J Vernon ² , MH Wilcox ^{1,2} |
| 7 | |
| 8 | |
| 9 | Microbiology, Leeds Teaching Hospitals Trust ¹ & Healthcare Associated Infections |
| 10 | Research Group, ² Leeds Institute for Biomedical and Clinical Sciences, University of |
| 11 | Leeds Leeds LIK |
| 40 | |
| 12 | |
| 13 | |
| 14 | Running title: C. difficile susceptibility to MCB3681 and comparators |
| 15 | |
| 10 | |
| 16 | |
| 17 | *Corresponding author: Dr Jane Freeman Jane.freeman4@nhs.net |
| 18 | |
| 19 | |
| 20 | |
| 21 | |
| 22 | |

23 Abstract

- 24 Treatments for Clostridium difficile infection remain limited, despite the introduction
- 25 of fidaxomicin, and development of new agents is necessary.
- 26 We determined the in vitro susceptibilities of 199 prevalent or emerging Clostridium
- 27 difficile PCR ribotypes to MCB3681, a novel investigational quinolonyl-oxazolidinone,
- 28 and 8 comparators (metronidazole, vancomycin, fidaxomicin, moxifloxacin,
- 29 ciprofloxacin, clindamycin, tigecycline and linezolid).
- 30 MCB3681 showed good activity against C. difficile with no evidence of MCB3681
- 31 resistance in isolates showing either or both moxifloxacin and linezolid resistance.32
- 33 C. difficile infection (CDI) is a major burden on healthcare resources. CDI is thought
- to arise following the depletion of gut microflora by antimicrobial action, allowing the
- 35 organism to proliferate and cause disease. Antimicrobial treatments for CDI are
- 36 currently limited to metronidazole, vancomycin and fidaxomicin. Metronidazole has
- 37 more recently been associated with treatment failures, while promotion of
- 38 glycopeptide resistance within the host microflora is a risk associated with
- 39 vancomycin therapy. ¹ Symptomatic recurrence is common following treatment with
- 40 these agents, ² requiring further episodes of antimicrobial therapy. Further treatment
- 41 options are highly desirable to broaden the range of therapeutic choice and
- 42 strengthen antimicrobial stewardship.
- 43 MCB3681 is a novel small molecule with structural elements of an oxazolidinone and
- 44 a quinolone showing good activity against C. difficile, including isolates that were
- 45 resistant to linezolid, ciprofloxacin, moxifloxacin and clindamycin.³ It achieves high
- 46 faecal concentrations after intravenous infusions and has shown activity against
- 47 Gram positive components of the gut microflora in a clinical Phase 1 study⁴. The
- 48 development of an intravenous treatment agent achieving high faecal concentrations
- 49 would circumvent issues of rapid gut transit, or impaired delivery of orally
- 50 administered agents due to ileus, particularly in patients with severe or
- 51 protracted/multiple recurrent diarrhoeal episodes.
- 52 We determined the in vitro activities of MCB3681 and 8 comparators (metronidazole,
- 53 vancomycin, moxifloxacin, ciprofloxacin, clindamycin, tigecycline, linezolid, and
- 54 fidaxomicin) against a panel of 200 Clostridium difficile isolates of known PCR
- ribotypes (RT) from 21 European countries (selected from the ClosER study July
- 56 2011-April 2013, by kind permission of Astellas Pharma Europe). ⁵
- 57

58 In vitro susceptibility testing was performed using a Wilkins-Chalgren agar incorporation method, as previously described.^{5,7} Briefly, C. difficile test isolates and 59 60 control strains (C. difficile ATCC 750057, C. difficile E4 PCR ribotype 010, 61 Bacteroides fragilis ATCC 25285, Enterococcus faecalis ATCC 29212 and 62 Staphlyococcus aureus ATCC 29213 were cultured anaerobically at 37°C for 24h in 63 Schaedler's anaerobic broths prior to dilution to 0.5 McFarland standard 64 equivalence) in pre-reduced sterile saline and inoculation onto antibiotic-containing and control Wilkins-Chalgren agar plates. Inoculated plates were incubated 65 66 anaerobically at 37°C for 48h.

67

68 MCB3681 is a quinolonyl- oxazolidinone antibacterial which has previously

69 demonstrated good activity against C. difficile. ³ All the CDI treatment agents,

including MCB3681 showed good activity against the isolates tested (Table 1).

71 Fidaxomicin was the most active treatment agent (Kruskal-Wallis p<0.0001;

geometric mean (GM) MIC=0.05 mg/L), followed by MCB3681 (p<0.0001; GM=0.12

73 mg/L), then metronidazole (p<0.0001; GM=0.33 mg/L), with no evidence of

resistance to any of these compounds (Table 1). Vancomycin was the least active

75 (p<0.0001; GM=1.02 mg/L), but resistance was very scarce (1.5%;

76 breakpoint>8mg/L). Reduced metronidazole susceptibility (4 mg/L) was observed in

only 1% of isolates. GM metronidazole MICs were elevated in RT027 (0.96 mg/L)

and RT106 (0.74 mg/L) vs GM metronidazole MICs for all isolates tested (0.33

79 mg/L), in agreement with previous data.⁴

80

81 All isolates were resistant to ciprofloxacin according to the breakpoints defined

82 (Table 1), and 48% of isolates showed moxifloxacin resistance, including at least one

isolate in each RT group tested. Highly elevated MICs to both moxifloxacin (\geq 32

mg/L) and ciprofloxacin (\geq 128 mg/L) were prevalent in RT001, RT027 and RT356.

85 Clindamycin MICs were highest in RT001, RT017 and RT126 (GM MICs = 61.11

86 mg/L; 64 mg/L and 38.05 mg/L, respectively), but there was evidence of clindamycin

87 resistance in all RTs tested (Table 1). There was no evidence of tigecycline

resistance (range=0.03-0.125mg/L; GM=0.05mg/L), in agreement with previous data

89 (Table 1).⁴ The majority of isolates (78.9%) were sensitive to linezolid (Table 1), with

90 a GM MIC of 5.16 mg/L. RT001 and RT017 showed the highest GM linezolid MICs

91 (10.08 mg/L and 7.03 mg/L, respectively). This is also in agreement with previous

92 observations.⁷ Three RT017, and two RT027 isolates showed dual quinolone-

93 oxzolidinone resistant phenotype, and showed MCB3681 MICs of 0.5 mg/L. We

94 have previously reported that these isolates showed high level resistance to chloramphenicol (Table 2).^{5,8} Marin et al. reported linezolid, chloramphenicol, 95 96 erythromycin, and clindamycin resistance associated with the presence of the 97 multidrug resistance gene, cfr, in C. difficile RT017, RT078 and RT126 isolates.¹¹ 98 The MIC₅₀ and MIC₉₀ values reported here for MCB3681 are similar to those recently described for cadazolid, another quinolonyl-oxazolidinone molecule.⁹ A previous 99 100 study investigating susceptibility of C. difficile to cadazolid and comparators, reported 101 an association between resistance to either moxifloxacin or linezolid and 102 moxifloxacin/linezolid double-resistant mutants, and 2- or 4-fold higher cadazolid MICs in mono- or double-resistant isolates, respectively.¹⁰ However, the highest 103 104 MCB3681 MIC was 0.5 mg/L, and we also found isolates with moxifloxacin, 105 ciprofloxacin, linezolid and chloramphenicol resistance that demonstrated very low 106 MCB3681 MICs (0.008 mg/L) (Table 2). We did not investigate the molecular basis 107 of resistance in these isolates, but the results do not suggest a link between this 108 phenotype and MCB3681 MICs. The results shown here, in conjunction with those previously reported^{8, 11} would also indicate that other modes of resistance to linezolid 109 110 (23s rRNA alterations, ribosomal protein modifications) may be at play in 111 combination with quinolone resistance mechanisms.

112

113 Rashid et al. reported MICs of MCB3681 for C. difficile ranged from 0.008-0.5 mg/L,³ 114 which were similar to our results (range 0.008-0.5 mg/L). However, in the present 115 study, MIC₅₀ and MIC₉₀ values were 0.125 and 0.25 mg/L, respectively, which were 116 marginally higher than those reported previously, but within 2 doubling dilutions (0.03) 117 and 0.06 mg/L, respectively). This may be explained by methodological/agar or C. 118 difficile strain distribution differences. The influence of testing media and 119 components therein on MICs has previously been reported and may have been a 120 factor in the differences observed.^{7,12} We used a Wilkins-Chalgren agar incorporation 121 method to determine MICs, since is superior to CLSI-recommended Brucella blood 122 agar (BBA) in the detection of reduced susceptibility to metronidazole in C. difficile.⁷ 123

124 This study builds on this previous data of Rashid et al by substantially expanding the

125 diversity of ribotypes examined to include, in particular, RT027 and several RTs

126 already noted for resistance to multiple antimicrobials: RT001, RT017, RT018,

127 RT027 and RT356.,^{5,8} There was no evidence of MCB3681 resistance among them.

128 MCB3681 achieves faecal concentrations of 99-226mg/kg after intravenous

| 129 | infusions, far in excess of MIC ranges for C. difficile reported here. MCB3681 has |
|------------|---|
| 130 | been reported to be active against Gram-positive gut microflora bacteria, but sparing |
| 131 | of Gram-negative organisms in human volunteer studies with intravenous |
| 132 | administration over 5 days. Further data are needed to assess the impact of |
| 133 | MCB3681 on C. difficile and the gut microflora over a longer duration. |
| 134 | |
| 135 | In summary, MCB3681 showed good activity against C. difficile isolates from |
| 136 | emerging or prevalent European PCR ribotypes with no evidence of resistance. The |
| 137 | presence of quinolone and/or linezolid resistance did not influence MCB3681 MICs. |
| 138 | |
| 139 | Funding |
| 140 | This research was funded by Morphochem AG, Munich, Germany |
| 141 | |
| 142 | Acknowledgements |
| 143 | We are grateful to Dr Chris Longshaw and Astellas Pharma Europe for kind |
| 144 | permission to use C. difficile isolates collected during The ClosER study. |
| 145 | |
| 146 | Transparency Declaration |
| 147 | JF has grant/research funding outside this work from Astellas and Melinta |
| 148 | Therapeutics. |
| 149 | MHW has received grant/research funding outside this work from Abbott, Actelion, |
| 150 | Alere, Astellas, Biomerieux, Cerexa, Cubist, Da Volterra, European Tissue |
| 151 | Symposium, Merck, Sanofi-Pasteur, Summit, The Medicines Company and Qiagen |
| 152 | and consultancies and/or lecture honoraria from Actelion, Alere, Astellas, Astra- |
| 153 | Zeneca, Basilea, Bayer, Cubist, Durata, European Tissue Symposium, J&J, Merck, |
| 154 | Nabriva, Novacta, Novartis, Optimer, Pfizer, Roche, Sanofi-Pasteur and Seres and |
| 155 | has been a member of a speaker's bureau for Pfizer. |
| 156 | JV and SP have nothing to declare. |
| 157 | |
| 158 159 | References |
| 160 | |
| 162 | 1. Al-Nassir WN, Sethi AK, Li Y, Peltz MJ, Riggs MM, Sonskey CJ. 2008. Both oral |
| 163 | metronidazole and oral vancomycin promote persistent overgrowth of vancomycin-resistant |
| 164 | enterococci during treatment of Clostridium difficile-associated disease. Antimicrob Agents |
| 165 | Chemother. 52 (7): 2403-6 |

| 166 | 2. | Debast SB, Bauer MP, Kuijper EJ. European Society of Clinical Microbiology and |
|-----|-----|---|
| 167 | | Infectious Diseases. 2014. European Society of Clinical Microbiology and Infectious |
| 168 | | Diseases: update of the treatment guidance document for Clostridium difficile infection. Clin |
| 169 | | Microbiol Infect.; 20: suppl 2: 1-26 |
| 170 | 3. | Rashid M-U, Dalhoff A, Weintraub A, Nord CE. 2014. In vitro activity of MCB3681 against |
| 171 | | Clostridium difficile strains. Anaerobe; 28: 216-219 |
| 172 | 4. | Dalhoff A, Rashid M-U, Kapsner T, Panagiotidis G, Weintraub A, Nord CE. 2015. Analysis |
| 173 | | of effects of MCB3681, the antibacterially active substance of prodrug MCB3837, on human |
| 174 | | resident microflora as proof of principle. Clin Microbiol Infect: 21: 767.e1-767.e4 |
| 175 | 5. | Freeman J, Vernon J, Morris K, Nicholson S, Todhunter S, Lonshaw C, Wilcox MH. 2015 |
| 176 | | Pan-European longitudinal surveillance of antimicrobial resistance among prevalent |
| 177 | | Clostridium difficile ribotypes. Clinical Microbiol and Infect; 21(3):248.e9-248.e16 |
| 178 | 6. | Clinical Laboratory Standards Institute. 2014 Principles and Procedures for Detection of |
| 179 | | Anaerobes in Clinical Specimens; Approved Guideline M56-A. Wayne, PA, USA, |
| 180 | 7. | Baines SD, O'Connor R, Freeman J, Fawley WN, Harmanus C, Mastrantonio P, Kuijper |
| 181 | | EJ, Wilcox MH.2008. Emergence of reduced susceptibility to metronidazole in Clostridium |
| 182 | | difficile. J Antimicrob Chemother: 62 :1046-1052. |
| 183 | 8. | Freeman J, Vernon JJ, Vickers R, Wilcox MH. 2015. Susceptibility of Clostridium difficile |
| 184 | | isolates of varying antimicrobial resistance phenotypes to SMT19969 and 11 comparators. |
| 185 | | Antimicrob Agents Chemother: 60 (1): 689-92 |
| 186 | 9. | Gerding DN, Hecht DW, Louie T, Nord CE, Talbot GH, Cornely OA, Buitrago M, Best E, |
| 187 | | Sambol S, Osmolski JR, Kracker H, Locher HH, Charef P, Wilcox M. 2016 Susceptibility |
| 188 | | of Clostridium difficile isolates from a Phase 2 clinical trial of cadazolid and vancomycin in C. |
| 189 | | difficile infection. J. Antimicrob Chemother. 71(1): 213-9 |
| 190 | 10 | Locher HH. Seiler P, Chen X Schroeder S, Pfaff P, Enderlin M, Lkenk A, Fournier E, |
| 191 | | Hubschwerlen C, Ritz D, Kelly CP, Keck W. 2014 In vitro and in vivo antibacterial |
| 192 | | evaluation of cadazolid, a new antibiotic for treatment of Clostridium difficile infection. |
| 193 | | Antimicrob Agents Chemother. 58(2):892-900 |
| 194 | 11. | . Marin M, Martin A, Alcala L,Cercenado E, Iglesias, Reigadas E, Bouza E. 2015. |
| 195 | | Clostridium difficile isolates with high linezolid MICs harbour the multiresistance gene, cfr. |
| 196 | | Antimicrob Agents Chemother; 59 (1): 586-9. |
| 197 | 12 | . Wu X, Hurdle JG. 2015. Hemin modulates metronidazole susceptibility of Clostridium difficile. |
| 198 | | Abstr. C-576. Presented at the 55th Interscience Conference on Antimicrobial Agents and |
| 199 | | Chemotherapy. San Diego, CA. |
| 200 | | |
| 201 | | |
| 202 | | |
| 203 | | |
| 204 | | |

| mg/L | MCB3681 | FDX | MTZ | VAN | MXF | CIP | CLI | TGC | LZD |
|--------------------|------------------|--------|---------|-------|-------|--------------------|-------|-------------------|------------------|
| | S<4; | S <1; | S<2; | S<2; | S<2; | S <8; | S<2; | S<4; | S<4; |
| | | | I=4; | I=4; | I=4; | | I=4; | | |
| Breakpoints | R>4 ² | RS >14 | R>84 | R>84 | R>84 | RS >8 ² | R>84 | RS>4 ⁴ | R>4 ⁷ |
| %S | 100 | 100 | 99 | 96 | 50.5 | - | 5.5 | 100 | 78.9 |
| %I | - | - | 1 | 2.5 | 1 | - | 29.5 | - | - |
| %R | - | - | 0 | 1.5 | 48 | 100 | 54 | - | 21.1 |
| MIC ₅₀ | 0.125 | 0.06 | 0.25 | 1 | 2 | 64 | 16 | 0.06 | 4 |
| MIC ₉₀ | 0.25 | 0.125 | 1 | 2 | 32 | 256 | 128 | 0.06 | 8 |
| | | 0.004- | <0.125- | | 1- | | | | |
| range | 0.008-0.5 | 0.25 | 4 | 0.5-8 | >64 | 8->128 | 1->64 | 0.03-0.125 | 2->64 |
| Geometric mean | | | | | | | | | |
| MIC (mg/L) | | | | | 1 | | | 1 | |
| RT001 (15) | 0.07 | 0.02 | 0.42 | 0.79 | 16.00 | 111.43 | 61.11 | 0.03 | 10.08 |
| RT002 (14) | 0.11 | 0.06 | 0.19 | 0.87 | 1.82 | 27.86 | 12.13 | 0.04 | 4.39 |
| RT005 (16) | 0.14 | 0.06 | 0.29 | 1.16 | 2.00 | 37.12 | 9.28 | 0.04 | 5.66 |
| RT014 (16) | 0.11 | 0.07 | 0.28 | 0.88 | 3.36 | 39.74 | 10.37 | 0.05 | 4.36 |
| RT015 (15) | 0.14 | 0.06 | 0.25 | 0.87 | 1.91 | 26.60 | 7.29 | 0.04 | 4.19 |
| RT017 (16) | 0.15 | 0.04 | 0.26 | 0.74 | 12.88 | 86.67 | 64.00 | 0.06 | 7.03 |
| RT018 (14) | 0.12 | 0.06 | 0.41 | 1.49 | 6.90 | 110.33 | 8.83 | 0.04 | 4.42 |
| RT020 (15) | 0.10 | 0.06 | 0.25 | 0.75 | 2.59 | 36.44 | 11.31 | 0.05 | 4.76 |
| RT027 (16) | 0.16 | 0.09 | 0.96 | 1.14 | 21.67 | 206.14 | 19.87 | 0.05 | 5.19 |
| RT078 (16) | 0.11 | 0.05 | 0.26 | 0.92 | 2.38 | 34.90 | 12.34 | 0.05 | 5.42 |
| RT106 (14) | 0.11 | 0.09 | 0.74 | 1.10 | 7.61 | 81.98 | 10.77 | 0.04 | 4.42 |
| RT126 (16) | 0.12 | 0.06 | 0.32 | 1.00 | 8.35 | 72.88 | 38.05 | 0.06 | 4.56 |
| RT356 (16) | 0.08 | 0.04 | 0.27 | 2.28 | 29.34 | 245.15 | 12.88 | 0.04 | 4.76 |
| All isolates (199) | 0.12 | 0.05 | 0.33 | 1.02 | 5.87 | 66.27 | 16.17 | 0.05 | 5.16 |

 Table 1. Susceptibility of 199 C. difficile isolates to MCB3681 and 8 comparators.

 FDX= fidaxomicin; MTZ=metronidazole, VAN= vancomcyin; MXF=moxifloxacin;CIP=ciprofloxacin; CLI=clindamycin; TGC=tigecycline; LZD=linezolid

 S=sensitive; I=intermediate; R=resistant; RS=reduced susceptibility

| | MIC (mg/L) | | | | | | | | |
|-----|------------|------|---------|-----|-----|---------|--|--|--|
| RT | MXF CIP | | LZD CLI | | CHL | MCB3681 | | | |
| 001 | 16 | 128 | 32 | >64 | 32 | 0.008 | | | |
| 001 | 32 | 128 | 32 | >64 | 32 | 0.015 | | | |
| 001 | 32 | 128 | 32 | >64 | 32 | 0.015 | | | |
| 001 | 16 | 64 | 16 | >64 | 8 | 0.25 | | | |
| 001 | 16 | >128 | 32 | >64 | 8 | 0.25 | | | |
| 001 | 16 | >128 | 32 | >64 | 32 | 0.25 | | | |
| 001 | 16 | >128 | 32 | >64 | 32 | 0.25 | | | |
| 014 | 16 | >128 | 32 | 16 | 16 | 0.06 | | | |
| 017 | 32 | 128 | 64 | >64 | 2 | 0.06 | | | |
| 017 | 16 | 64 | 16 | >64 | 32 | 0.25 | | | |
| 017 | 32 | 64 | 16 | >64 | 64 | 0.5 | | | |
| 017 | 32 | 64 | 32 | >64 | >64 | 0.5 | | | |
| 017 | 32 | 128 | 32 | >64 | 64 | 0.5 | | | |
| 018 | 16 | >128 | 8 | 8 | 4 | 0.03 | | | |
| 018 | 32 | >128 | 8 | 8 | 2 | 0.06 | | | |
| 027 | 32 | >128 | 32 | >64 | 64 | 0.5 | | | |
| 027 | 32 | >128 | 32 | >64 | 64 | 0.5 | | | |
| 078 | 8 | 128 | 8 | 4 | 4 | 0.125 | | | |
| 078 | 16 | >128 | 8 | 4 | 64 | 0.125 | | | |
| 106 | 16 | 128 | 8 | 8 | 4 | 0.06 | | | |
| 126 | 16 | 64 | 16 | >64 | 4 | 0.125 | | | |
| 356 | 32 | >128 | 8 | 8 | 4 | 0.03 | | | |
| 356 | 32 | >128 | 8 | 8 | 8 | 0.06 | | | |
| 356 | 32 | >128 | 8 | 8 | 4 | 0.25 | | | |
| 356 | >64 | 128 | 8 | 16 | 4 | 0.25 | | | |

Table 2 MCB3681 MICs (mg/L) in C. difficile isolates with dual quinolone-oxazolidinone resistance (highlighting indicates resistance). Clindamycin and

chloramphenicol MICs⁴ are also shown.

MXF=moxifloxacin; CIP=ciprofloxacin; LZD=linezolid; CLI=clindamycin; CHL=chloramphenicol