

This is a repository copy of *The role of acroblast formation during Drosophila spermatogenesis*.

White Rose Research Online URL for this paper:

<https://eprints.whiterose.ac.uk/101995/>

Version: Accepted Version

---

**Article:**

Fari, Karolina, Takacs, Sandor, Ungar, Daniel [orcid.org/0000-0002-9852-6160](https://orcid.org/0000-0002-9852-6160) et al. (1 more author) (2016) The role of acroblast formation during *Drosophila* spermatogenesis. *Open Biology*. pp. 1102-1110. ISSN 2046-2441

<https://doi.org/10.1242/bio.018275>

---

**Reuse**

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

**Takedown**

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing [eprints@whiterose.ac.uk](mailto:eprints@whiterose.ac.uk) including the URL of the record and the reason for the withdrawal request.

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27

## **The role of acroblast formation during *Drosophila* spermatogenesis**

**Karolina Fári<sup>1\*</sup>, Sándor Takács<sup>1\*</sup>, Dániel Ungár<sup>2</sup>, Rita Sinka<sup>1</sup>**

<sup>1</sup> Department of Genetics, University of Szeged, Hungary

<sup>2</sup> Department of Biology, University of York, York, UK

\* These authors contributed equally to this work

Running title: acroblast-acrosome formation

Corresponding author: Rita Sinka, [rsinka@bio.u-szeged.hu](mailto:rsinka@bio.u-szeged.hu)

Key words: acrosome, acroblast, Golgi, *Drosophila*, spermatogenesis

28 **Summary Statement**

29 This study demonstrates that a functional acroblast is essential for normal nuclear elongation  
30 and acrosome formation during *Drosophila* spermatogenesis.

31

32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50

## Abstract

Protein recycling is important for maintaining homeostasis of the Golgi and its cisternae. The Vps54 (Scat) protein, a subunit of the GARP tethering complex, is a central factor in retrograde transport to the *trans*-Golgi. We found the *scat*<sup>l</sup> mutant to be male sterile in *Drosophila* with individualization problems during spermatogenesis. Another typically observed phenotype was the abnormal nuclear structure in elongated mutant cysts. Examining the structure and function of the Golgi a failure in acrosome formation and endosome-Golgi vesicular transport were found in the *scat*<sup>l</sup> mutant. This acrosome formation defect was due to a fault in the *trans*-Golgi side of the acroblast ribbon. Testing a mutation in a second retrograde transport protein, Fws, a subunit of the COG tethering complex, the acroblast structure was again disrupted. *fws*<sup>P</sup>, caused a similar albeit milder acrosome and sperm individualization phenotype as the *scat*<sup>l</sup> mutant. In the case of *fws*<sup>P</sup> the *cis* side of the acroblast ribbon was dispersed, in-line with the intra-Golgi retrograde function of COG. Our results highlight the importance of an intact acroblast for acrosome formation, nuclear elongation and therefore sperm maturation. Moreover, these results suggest the importance of retrograde tethering complexes in the formation of a functional Golgi ribbon.

## 51 **Introduction**

52

53 In most mammalian cell types the Golgi apparatus appears as a ribbon formed from  
54 interconnected stacks of cisternae. However, this arrangement is not universal. For example,  
55 in gastric parietal cells the Golgi appears in the form of mini-stacks dispersed throughout the  
56 cytoplasm (Gunn et al., 2011). This scattered arrangement is common in most cell types of  
57 the fruit fly *D. melanogaster* (Kondylis and Rabouille, 2009). Several possible roles have  
58 been proposed for the assembly of Golgi stacks into a ribbon. These include an increase in  
59 efficiency and uniformity of glycosylation (Puthenveedu et al., 2006), a necessity of the  
60 ribbon for the secretion of large cargoes (Lavieu et al., 2014), and importantly, a role in  
61 polarized secretion (Horton et al., 2005). Generation of the Golgi ribbon requires microtubule  
62 mediated transport of stacks or vesicles forming them into the vicinity of the microtubule  
63 organizing center (Wehland et al., 1983), followed by tethering and fusion of these into a  
64 ribbon (Marra et al., 2007). Specialized cell types in non-vertebrates can also present an  
65 assemblage of Golgi stacks in a perinuclear location. For example, the Golgi apparatus of  
66 developing *Drosophila* spermatids is in such a peri-nuclear location just prior to and during  
67 the nuclear elongation phase of spermatogenesis (Kondylis and Rabouille, 2009). This  
68 specialized Golgi assemblage, known as the acroblast, is likely needed to organize the  
69 secretory pathway in this highly polarized cell type of the fruit fly.

70 While it is clear that anterograde transport to the Golgi is essential for generation of a  
71 polarized assembly of Golgi stacks, the role of retrograde transport in this process is less well  
72 understood. Retrograde transport within and to the Golgi is coordinated by two multisubunit  
73 tethering complexes, the Golgi associated retrograde protein (GARP) (Bonifacino and Hierro,  
74 2011) and the conserved oligomeric Golgi (COG) (Miller and Ungar, 2012) complexes.  
75 GARP is a four subunit complex of the CATCHR (Complexes Associated with Tethering  
76 Containing Helical Rods) family (Hughson and Reinisch, 2010), composed of the Vps51,  
77 Vps52, Vps53 and Vps54 proteins (Conibear and Stevens, 2000). Its primary role in  
78 membrane trafficking is to direct retrograde carriers to the TGN (Conibear and Stevens,  
79 2000), such as vesicles that recycle the mannose-6-phosphate receptor (M6PR) (Perez-  
80 Victoria et al., 2008), or those carrying the SNARE protein Snc1 (Quenneville et al., 2006)  
81 from endosomes to the TGN. Lack of Vps54 in mice causes the *wobbler* phenotype, which  
82 manifests in progressive neurodegeneration and male sterility (Schmitt-John et al., 2005). In  
83 *Drosophila* the GARP complex has been shown to require the Arl5 GTPase for correct

84 localization, loss of which results in defective recycling of Lerp, the fly homolog of M6PR  
85 (Rosa-Ferreira et al., 2015). While COG is also involved in endosome to Golgi transport  
86 (Whyte and Munro, 2001), its main function is the intra-Golgi retrograde trafficking of  
87 resident Golgi proteins (Oka et al., 2004). The eight COG subunits can be grouped into two  
88 lobes, with subunits Cog1-4 forming lobe A, and Cog5-8 lobe B (Ungar et al., 2002). While  
89 loss of lobe A function causes defects in the recycling of early Golgi residents, lobe B is  
90 mainly involved in late Golgi homeostasis (Oka et al., 2004; Willett et al., 2013; Wu et al.,  
91 2004). Consequently, lobe A is essential for development of an organism, its loss is lethal in  
92 yeast (Whyte and Munro, 2001) and during early development in *Drosophila* (Schnorrer et  
93 al., 2010). In contrast, lobe B loss causes much milder phenotypes, for example loss of Cog5  
94 in a human patient was shown to lead to relatively mild psychomotor retardation (Paesold-  
95 Burda et al., 2009), while two loss of function alleles of its fly homolog, fws, cause male  
96 sterility due to incomplete cytokinesis during spermatogenesis (Farkas et al., 2003).  
97 Interestingly, COG interacts with the golgin TMF (Miller et al., 2013), which is a critical  
98 factor for vesicular transport during late stages of mouse spermatogenesis (Lerer-Goldshtein  
99 et al., 2010).

100 During *Drosophila* spermatogenesis, following meiotic division the 64 spermatids  
101 undergo a dramatic differentiation program that leads to formation of the highly elongated  
102 flagellated mature sperm (Fig. 1A). This process starts with rearrangement and fusion of  
103 mitochondria to form the Nebenkern from two mitochondrial derivatives (Fig. 1A) (Tokuyasu  
104 et al., 1972). At the same time the basal body is embedded into the nuclear envelope to  
105 polarize the nucleus (Vogt et al., 2006). The Golgi apparatus, which is normally a collection  
106 of scattered stacks throughout the cytosol, is then recruited to the nucleus at the opposing side  
107 to the basal body (Fig. 1A,B) (Fuller, 1993). This polarization event is thought to be essential  
108 for subsequent nuclear elongation. The change in nuclear shape is coincident with a major  
109 reorganization of chromatin, which manifests in the replacement of histones with protamines.  
110 This histone to protamine switch is critical for the proper elongation of the nuclei (Raja and  
111 Renkawitz-Pohl, 2005). During the later stages of nuclear elongation the specialized Golgi  
112 structure, the acroblast, is converted into the acrosome and the actin-based investment cones  
113 are formed (Fig. 1A). These investment cones are also involved in the individualization of the  
114 mature sperm when an enormous amount of new membrane is used for elongation, which  
115 concludes spermatogenesis (Fabian and Brill, 2012).

116 The acroblast described above, contains all the markers of a typical Golgi apparatus,  
117 such as the glycosylation enzyme mannosidase II (Farkas et al., 2003), the golgins

118 Golgin245, GM130 (Hirst and Carmichael, 2011) and Lava lamp (Farkas et al., 2003), the  
119 COPI vesicle coat (Kitazawa et al., 2012) and the COG complex (Farkas et al., 2003). In  
120 addition, the lysosomal protein Lamp1, and the acrosomal protein Sneaky also localize to the  
121 acroblast (Wilson et al., 2006) (Fig. S1G,H). Yet, the acroblast is unusual in *Drosophila*, as it  
122 forms a ribbon as opposed to the scattered stacks typical for Golgi architecture in other fruit  
123 fly cells (Kondylis and Rabouille, 2009). The molecular determinants of acroblast formation  
124 and its breakdown upon acrosome formation are not very well understood, but the Golgi  
125 architecture leading to acrosome formation has been recently documented (Yasuno et al.,  
126 2013). After meiosis the Golgi is organized around the nucleus and participates in the  
127 formation of the acroblast (Fig. 1A,B). Once nuclei elongate the acroblast disassembles and  
128 some of the Golgi components, such as Sneaky, together with lysosomal components  
129 generate the acrosome, which maintains an apical positioning next to the nucleus (Fig. 1A,D).  
130 At the same time the remaining Golgi components migrate to the posterior side of the nucleus  
131 and appear as scattered stacks akin to somatic *Drosophila* cells (Fig. 1A,C). The known  
132 molecular players that have so far been associated with the formation of the acroblast, and its  
133 later breakdown have all been found to affect meiotic division as well (Belloni et al., 2012;  
134 Farkas et al., 2003). It is therefore often difficult to tease out direct effects on Golgi  
135 architecture from secondary effects due to delays in spermatogenesis and associated defects  
136 in polarization. Such factors include microtubules (Yasuno et al., 2013), the  
137 phosphatidylinositol transfer protein Giotto (Giansanti et al., 2006), the small GTPase Rab11  
138 (Giansanti et al., 2007), the TRAPP II complex (Robinett et al., 2009), as well as the Cog5  
139 and Cog7 subunits of COG (Belloni et al., 2012; Farkas et al., 2003).

140 Here we have analyzed two different male sterile P-element insertion mutations; one of  
141 the GARP subunit Vps54 (*scat*), the other of the Cog5 (*fws*) subunit of COG. These mutants  
142 have no defects in the meiotic phase of sperm development, but nuclear elongation and  
143 acrosome formation are both affected. Mutant spermatids of *scat*<sup>l</sup> and *fws*<sup>P</sup> do not  
144 individualize and therefore do not mature. We show that the main defect of these mutants is  
145 in the organization of the acroblast and the ensuing completion of the spermatogenic  
146 differentiation program. These results highlight an essential function of the GARP and COG  
147 mediated retrograde transport processes in the establishment of a polarized Golgi ribbon,  
148 which is important in nuclear elongation, individualization and acrosome formation during  
149 *Drosophila* spermatogenesis.

150

## 151 **Results**

152

### 153 **The *scat*<sup>1</sup> mutant has a male fertility defect**

154

155 In order to probe the function of vesicle tethering complexes that act at the Golgi during  
156 spermatogenesis, we investigated the Vps54 subunit of the GARP complex, encoded by the  
157 *scat* gene. The *scat*<sup>1</sup> allele was identified as a male sterile mutant with scattered nuclei in a P  
158 element screen (Castrillon et al., 1993). The P element is incorporated in the third exon of the  
159 gene (Fig. 2A). Genetic characterization showed that homozygous *scat*<sup>1</sup> males were 100%  
160 sterile and their seminal vesicle was devoid of mature sperm (Fig. S1 A,B). In contrast, all  
161 females were fertile. We tested the *scat*<sup>1</sup> allele in complementation analysis and found male  
162 sterility in a hemizygous combination with an overlapping *Df(2L)ED680* deficiency. The  
163 male sterility of *scat*<sup>1</sup> was completely reversed by precise excision of the P element. To verify  
164 the involvement of the *scat* gene a C-terminally RFP tagged *scat* transgene expressed from a  
165 *P{UASp}* vector was used to rescue the male sterile phenotype. Expression of the *P{UASp-*  
166 *Scat-RFP}* fusion protein using the germ line specific *Bam-Gal4* driver completely rescued  
167 male sterility (Fig. S1 G-L). This proves that the P element insertion within *scat* is indeed  
168 responsible for the male sterility and the Scat-RFP fusion protein correctly incorporates into  
169 the GARP complex. A polyclonal antibody raised against Scat recognizes the protein at the  
170 predicted molecular weight of 105 kDa, as well as the Scat-RFP fusion protein in extracts  
171 from wild type or Scat-RFP transgenic testes (Fig. 2B first two lanes). In contrast, in  
172 homozygous *scat*<sup>1</sup> mutant testis extracts the protein was absent from the immunoblot,  
173 confirming that *scat*<sup>1</sup> is a null mutant (Fig. 2B right lane).

174

### 175 **Scat is Golgi localized throughout spermatogenesis**

176

177 Mouse Vps54 was shown to localize to both endosomes and Golgi, and to incorporate  
178 into the fully developed acrosome (Berruti et al., 2010). This is in contrast with other Golgi  
179 trafficking proteins, such as Golgin95 or Golgin97, which localize to the developing  
180 acrosome only during the early steps of acrosomogenesis but do not label the testicular  
181 spermatozoa (Moreno et al., 2000). We therefore tested the subcellular localization of Scat-  
182 RFP during different stages of *Drosophila* spermatogenesis. Both in the early premeiotic and  
183 in the late postmeiotic stages Scat localized to the Golgi (Fig. 2C-E), as is typical for the



184 GARP complex (Conibear et al., 2003). This was confirmed by co-staining with anti-  
185 dGM130, a protein known to be restricted to the *cis*-Golgi cisternae, just as its mammalian  
186 orthologue (Fig. 2C-E) (Sinka et al., 2008). We found that the Scat-RFP signal localized  
187 close but slightly displaced from dGM130 in all stages of spermatogenesis, suggesting that  
188 Scat is localized to the *trans* side of the Golgi (Fig. 2C-E). Early in the development process,  
189 in primary spermatocytes, the RFP stained Golgi is randomly distributed throughout the  
190 cytoplasm, similarly to the distribution found for GM130 and other medial/*trans* Golgi  
191 markers (Yasuno et al., 2013) (Fig. 2C). Interestingly, after meiosis, during acroblast  
192 formation the RFP staining marking the *trans* side of the Golgi was always positioned in the  
193 proximity of the nuclei as opposed to the more distally positioned *cis*-Golgi side (Fig. 2D).  
194 During nuclear elongation the Golgi localized Sneaky and the lysosomal Lamp then localized  
195 to the acrosome (Fig. 3I, FigS2A), but Scat, like the *cis*-Golgi specific dGM130, and the  
196 *trans*-Golgi specific dGolgin245 did not. Rather, Scat localizes with the rest of the Golgi  
197 markers in the scattered Golgi-stacks that move to the basal side of the nucleus and are later  
198 removed in the cystic bulge with the majority of the cytosol (Fig. 2E).

199

## 200 **Nuclear elongation is disrupted in *scat*<sup>1</sup> males**

201

202 All early steps of spermatogenesis, such as the maintenance of germ stem cells, the  
203 formation of primary spermatocytes, and meiotic divisions were normal in *scat*<sup>1</sup> testes.  
204 Nucleus to Nebenkern ratio was 1:1 in all round spermatids of the *scat*<sup>1</sup> mutant, suggesting  
205 normal cytokinesis (Fig. S1 D,E). We therefore focussed on the post meiotic stages of  
206 spermatogenesis to understand how the loss of Scat function perturbs spermatogenesis.  
207 Investigating the elongating spermatid nuclei their majority were found in late canoe stage  
208 and hardly any were observed as needle shaped in the *scat*<sup>1</sup> mutant cysts (Fig. 3A,B,D,E,G,  
209 H, I, J). The lack of needle shaped, fully elongated nuclei correlated with the appearance of  
210 scattered spermatid bundles in the *scat*<sup>1</sup> mutant post-meiotic cysts (Fig. 3A,B). Elongation  
211 and chromatin condensation occur parallel to each other. As in mammals, chromatin  
212 condensation is achieved by a histone to protamine switch during nuclear elongation in  
213 *Drosophila* (Raja and Renkawitz-Pohl, 2005). This switch is normal in the *scat*<sup>1</sup> mutant (Fig.  
214 3D, E), suggesting that the observed nuclear elongation defect is independent of the  
215 chromatin condensation process. The scattering of nuclei could also be caused by defects in  
216 basal body formation (Texada et al., 2008). However, visualization of the basal body with

217 GFP-PACT failed to reveal any abnormalities in elongated cysts of the *scat*<sup>1</sup> mutant (Fig.  
218 3G,H).

219 Vps54 mutant mice that are male sterile are missing acrosomes (Paiardi et al., 2011).  
220 These are normally formed during the later stages of nuclear elongation, so we wondered  
221 whether acrosome formation was normal in *scat*<sup>1</sup> mutants. Two different acrosomal markers,  
222 Snky-GFP and Lamp1-GFP (Fabian and Brill, 2012; Wilson et al., 2006) both showed  
223 acrosomal localization at the tips of elongated nuclei in WT spermatids (Fig. 3I and FigS2  
224 A). Yet the GFP signal was diffuse without any recognizable acrosome staining in the same  
225 stage of *scat*<sup>1</sup> mutant spermatids (Fig. 3J and FigS2 B).

226 Individualization starts with the formation of 64 actin-rich investment cones adjacent to  
227 the nuclei, which move together towards the distal end of the individualizing cyst (Fig 3A)  
228 (Fabrizio et al., 1998). In the case of the *scat*<sup>1</sup> mutant we hardly observed any investment  
229 cones and the process of individualization did not start. Occasionally we could detect a very  
230 faint Phalloidin signal, which could be due to investment cone remnants or the investment  
231 cones in the process of degradation, but these were always scattered (Fig. 3B). Following  
232 failed individualization the elongated cysts lost their integrity, the cells scattered and died.

233 Thus the earliest defect in spermatogenesis in *scat*<sup>1</sup> mutants is their failure to fully  
234 elongate the nuclei. While this is not accompanied with a defect in chromatin condensation, it  
235 does lead to a defect in individualization.

236

### 237 **Acrosome defects are the consequence of the abnormal acroblast formation**

238

239 The failures in nuclear elongation, acrosome formation and individualization all point to  
240 a defect following acroblast disassembly. This could be caused by an inherent defect of the  
241 acroblast itself in the mutants. The GARP complex is known to contribute to the recycling of  
242 M6PR between endosomes and the TGN (Perez-Victoria et al., 2008) in mammals, and  
243 therefore we wondered if this trafficking pathway was for example defective at the acroblast.  
244 A transgenic line was established with testis specific expression of the *Drosophila* M6PR,  
245 *P{tv3-GFP-Lerp}*. This showed acroblast localization in WT spermatids (Fig. 4A), while it  
246 had a more dispersed localization in the *scat*<sup>1</sup> mutant (Fig. 4B). The *trans*-Golgi marker  
247 Golgin245 showed co-localization with GFP-Lerp in WT, but not in the *scat*<sup>1</sup> mutant  
248 postmeiotic round spermatids (Fig. 4A,B). Thus similar to effects in mammalian cells, a  
249 faulty GARP complex causes defective retrograde trafficking of M6PR from endosomes to

250 the TGN. Such a defect in the endosome-Golgi trafficking route marked by the M6PR could  
251 affect acroblast integrity.

252 The distribution of *cis*- and *trans*-Golgi markers was therefore tested throughout  
253 spermatogenesis. In the early stage the *cis*- and *trans*-Golgi markers GM130 and Golgin245  
254 are localized close to each other in both WT and *scat*<sup>1</sup> mutant spermatocytes (FigS2 D,F). In  
255 WT post-meiotic spermatids these *cis*- and *trans*-Golgi markers appear closely apposed to  
256 each other in perinuclear localization, consistent with the perinuclear ribbon like Golgi  
257 formed by the acroblast (Fig. 4C). However, in the *scat*<sup>1</sup> mutants of the same stage,  
258 localization of the *trans*-Golgi marker is much more diffuse (Fig. 4D). The failure of GM130  
259 and Golgin245 to co-localize in the *scat*<sup>1</sup> mutants persists during nuclear elongation when the  
260 Golgi travels to the basal side of the nucleus (FigS. 2E,G). However, the high degree of cell  
261 death at this stage precludes far reaching conclusions to be drawn from this last stage. Overall  
262 these results suggest that a functional GARP complex is necessary for normal acroblast  
263 organization. Our data also imply that proper perinuclear organization of the acroblast is  
264 required for completion of spermatogenesis, including individualization, acrosome formation  
265 and the final stage of nuclear elongation.

266

### 267 **The intra-Golgi retrograde transport factor Fws is also necessary for acroblast integrity** 268 **and completion of the late stages of spermatogenesis**

269

270 Involvement of GARP-dependent trafficking in acrosome formation has been  
271 demonstrated in mice (Paiardi et al., 2011). The fruit fly spermatogenesis model provides a  
272 unique opportunity to study the involvement of Golgi ribbon biogenesis in acrosome integrity  
273 and formation, since the acroblast is the only true ribbon-like Golgi structure in the  
274 developing *Drosophila* sperm. Generation of a Golgi-ribbon is known to require the  
275 microtubule mediated transport of Golgi elements to the perinuclear region (Wehland et al.,  
276 1983). Our finding that GARP complex function is required for the correct formation of the  
277 peri-nuclear Golgi known as acroblast raised the intriguing possibility that other known  
278 retrograde transport factors could also be important contributors of Golgi-ribbon formation. A  
279 second retrograde trafficking factor, the Fws subunit of the intra-Golgi transport specific  
280 COG complex was therefore also investigated. Fws has previously been characterised during  
281 spermatogenesis using two EMS alleles (*fws*<sup>z-0161</sup> and *fws*<sup>z-1201</sup>). Transheterozygotes of these  
282 two alleles were shown to manifest in spermatocyte cytokinesis and spermatid elongation  
283 defects (Farkas et al., 2003). Given the strong defect in these EMS mutants during

284 cytokinesis, it is unclear whether the spermatid elongation and associated acrosome formation  
285 defects are secondary consequences of the meiotic defect. We therefore decided to investigate  
286 the phenotype of a new P element insertion line  $fws^{KG02853}$  ( $fws^P$ ).

287  $fws^P$  contains a P element insertion in the first exon of the  $fws$  gene. This disruption of  
288 the  $fws$  gene results in 74% male sterility in homozygotes and 90% in hemizygotes over the  
289 *DfBSC148* deficiency. These numbers show that this mutant is possibly a hypomorphic allele  
290 of  $fws$ . The sterile homozygous  $fws^P$  mutants' seminal vesicles were devoid of mature sperm  
291 (Fig1S C). Remobilisation of the P element in  $fws^P$  reverted the male sterile phenotype to  
292 recover complete fertility. The male sterile phenotype of  $fws^P$  was also rescued with the wild  
293 type GFP- $fws$  genomic rescue construct (Farkas et al., 2003), suggesting that the P element  
294 insertion in  $fws$  is indeed responsible for the male sterile phenotype of  $fws^P$  (FigS1 M-O).  
295 Importantly, in contrast to the previously reported EMS alleles, meiotic cytokinesis in  $fws^P$   
296 homozygous testes was normal (Fig. S1D,F) and proper elongation of the post meiotic cysts  
297 was observed (Fig. S1F). Phenotypic characterisation of developing spermatids showed  
298 scattered nuclei and investment cones in the  $fws^P$  mutant cysts (Fig. 3C), but similarly to the  
299 *scat<sup>l</sup>* mutant the histone protamine transition was again found to be normal (Fig. 3F). Using  
300 the acrosomal markers Snky-GFP and Lamp1-GFP showed that the majority of the elongated  
301 spermatids do not form acrosomes in the  $fws^P$  mutant (Fig. 3K and FigS. 2C). Yet, in contrast  
302 to the *scat<sup>l</sup>* mutant, in some cases we could observe an acrosomal signal with both transgenes  
303 (Fig. 3K and FigS. 2C arrowhead), some of the acrosomes decorating non-scattered nuclei.  
304 This is in line with the fertility results suggesting that the  $fws^P$  mutant has an overall milder  
305 defect than the *scat<sup>l</sup>* mutant, likely due to  $fws^P$  being a hypomorphic rather than a null-allele.  
306 Given the good correlation between the extent of the fertility and acrosome defects it seems  
307 that the most sensitive effect of  $fws$  disruption is on acrosome formation and nuclear  
308 elongation rather than cytokinesis (Farkas et al., 2003).

309 Finally, to test if the primary defect in the  $fws^P$  mutant, as in the *scat<sup>l</sup>* mutant, is in  
310 acroblast organization, the distribution of *cis*- and *trans*-Golgi markers was investigated in  
311  $fws^P$  from early stages up to cyst elongation (Fig. 4E and FigS. 2H,I). Similar to the *scat<sup>l</sup>*  
312 mutant we found a defect in the perinuclear acroblast (Fig. 4E). However, in this instance it  
313 was the GM130 marker that showed a more dispersed staining, while Golgin245 remained  
314 compact (Fig. 4E). This is in line with the COG complex, involved in intra-Golgi transport  
315 (Miller and Ungar, 2012), affecting more the formation of the *cis* side of the Golgi ribbon,  
316 while the GARP complex, involved in retrograde transport to the late Golgi (Conibear and  
317 Stevens, 2000), affecting the *trans* side.

318

## 319 **Discussion**

320

321 Our work sheds light on the interplay between vesicle trafficking, Golgi structure,  
322 acrosome formation and sperm development. The primary defect during sperm development  
323 in the analyzed mutants is disruption of the acroblast structure, which in turn causes defects  
324 in acrosome formation, nuclear elongation and individualization. As opposed to previously  
325 characterized Golgi trafficking mutants that have been shown to exhibit acroblast  
326 abnormalities (Farkas et al., 2003; Giansanti et al., 2006; Robinett et al., 2009), the *scat<sup>l</sup>* and  
327 *fws<sup>P</sup>* mutants exhibit no meiotic cytokinesis defects at all. The Giotto, and TRAPP mutants,  
328 as well as the EMS mutagenesis alleles of *fws* did show acroblast defects, but this could have  
329 been caused by a generic delay and consequent disruption of spermatogenesis due to  
330 defective cytokinesis. Our study is therefore the first clear demonstration that both Scat and  
331 Fws and therefore the GARP and COG complexes are essential for establishing correct  
332 acroblast morphology. Consequently, the lack of acrosomes in the here described mutants is,  
333 again, a more direct demonstration of the need for acroblast homeostasis in order to generate  
334 the acrosome and complete nuclear elongation, since other spermatogenesis stages up to the  
335 canoe stage of nuclear elongation are normal. The acrosome has been shown to contain  
336 elements of the TGN as well as the late endosome/lysosome (Herms et al., 2010). We find  
337 that the correct organization of the rest of the Golgi, including both the *cis* and *trans* sides is  
338 essential for acrosome formation, despite these Golgi components not being incorporated into  
339 the acrosome (Fig. 5). The mutants we describe will therefore be valuable tools in the future  
340 to study acrosome biogenesis during *Drosophila* spermatogenesis.

341 Several steps during post-meiotic spermatogenesis occur in parallel or close succession.  
342 These include chromatin condensation, basal body formation, acroblast formation, nuclear  
343 elongation and acrosome formation. The mutants characterized in this study allow us to place  
344 these in a hierarchy of dependence. It is clear that acroblast formation is not required for  
345 chromatin condensation, basal body formation and the initial phase of nuclear elongation. At  
346 the same time, formation of the acrosome and the elongation to needle shape nuclei cannot  
347 proceed even where chromatin has condensed and the basal body formed unless the acroblast  
348 is fully functional (Fig. 5). The most important function of the acroblast's intact ribbon  
349 during nuclear elongation and acrosome formation is its influence on the polarization of the  
350 cyst, which ultimately leads to normal individualization. However, molecular details of the

351 links between acroblast formation and function and the process of individualization remain to  
352 be identified.

353 The acroblast is a very special form of the Golgi apparatus in *Drosophila*, since it forms  
354 a perinuclear ribbon as opposed to the scattered stacks found in other cells of the fruit fly  
355 (Kondylis and Rabouille, 2009). The two mutants analyzed in this study show normal Golgi  
356 distribution in cells where the scattered stack morphology is predominant (FigS 2F, H). This  
357 implies that the retrograde transport routes defined by GARP and COG are not essential for  
358 the formation and maintenance of Golgi stacks in spermatocytes. While Golgi defects are  
359 common in mammalian COG mutants (Reynders et al., 2009; Ungar et al., 2002), a loss of  
360 Golgi stacks is not observed. Similarly, the ribbon of the acroblast is seriously malformed in  
361 both the *scat*<sup>1</sup> and the *fws*<sup>P</sup> mutants. This implies that ribbon formation may indeed need the  
362 retrograde transport pathways established for GARP (Bonifacino and Hierro, 2011) and COG  
363 (Miller and Ungar, 2012) in addition to the well-known contributions of microtubule  
364 mediated anterograde transport (Wehland et al., 1983). The fact that the observed disruption  
365 in the acroblast is most prominent on either the *trans* side (for GARP) (Conibear and Stevens,  
366 2000) or the *medial/cis* side (for COG) (Miller et al., 2013) is in line with the respective  
367 known destinations of the transported vesicles (Bonifacino and Hierro, 2011; Willett et al.,  
368 2013). Several candidates for the associated machinery that could act together with COG  
369 have already been flagged up by other studies, such as the golgins TMF (Schmitt-John et al.,  
370 2005) and GMAP210 (Kierszenbaum et al., 2011) that are both essential for acrosome  
371 formation in mouse testes. Yet, future studies are required to address what it is that has to be  
372 delivered to the particular Golgi areas by COG and GARP mediated retrograde transport in  
373 order to generate specific parts of the ribbon: Is it the whole vesicle that is needed, is it a very  
374 specific transport factor or factors that have to be recycled, or is it the general protein  
375 homeostasis within cisternae, maintained through recycling, that is essential for ribbon  
376 maintenance in the acroblast?

377

## 378 **Materials and methods**

379

### 380 **Fly stocks, mutants and transgenes**

381 Flies were crossed and maintained on standard cornmeal agar medium at 25°C. Oregon-  
382 R stock was used as wild type control. Fertility tests were performed by crossing single males  
383 with four wild type females. The progeny was counted in every tube and an average  
384 calculated from 30-50 males.

385 The following lines were obtained from the Bloomington Stock Center: *scat*<sup>1</sup>,  
386 *Df(2L)ED680*, *fws*<sup>P</sup> (*fws*<sup>KG02853</sup>), *Df(2L)ED1175*, *P(His2Av-EGFP.C2)*, *P(protamineB-eGFP)*.  
387 Flies carrying the *Snky-GFP*, *GFP-PACT*, *Lamp1-GFP*, *bam-Gal4* and *GFP-fws* transgenes  
388 have been described previously (Farkas et al., 2003; Martinez-Campos et al., 2004; Wilson et  
389 al., 2006). *Snky-GFP*, *Lamp1-GFP* and *GFP-fws* transgenes were recombined onto the 2<sup>nd</sup>  
390 chromosome with *scat*<sup>1</sup> and *fws*<sup>P</sup>. Remobilization of the P element in *scat*<sup>1</sup> and *fws*<sup>P</sup> was done  
391 according to Engels et al., 1990 (Engels et al., 1990). Revertant lines were tested for fertility  
392 and the precise excisions of the P elements were confirmed by PCR. The C-terminal  
393 *P{UASp-Scat-RFP}* construct was generated using the Gateway® cloning system  
394 (Invitrogen) according to the manufacturer's instructions, using *scat* cDNA. Transgenic lines  
395 were established and the *bam-Gal4* testis specific driver was used to express the transgene in  
396 wild type and *scat*<sup>1</sup> mutant backgrounds. The *P{tv3-Arl1-mCherry}* and *P{tv3-GFP-Lerp}*  
397 transgenic constructs were generated by amplifying the *arl1* and *lerp* cDNAs, and cloning the  
398 PCR fragments into a modified testis-vector3 (Wong et al., 2005) containing an insertion of  
399 mCherry or GFP to create a C- or N-terminal fusion protein. Transgenic flies were generated  
400 on a *w*<sup>1118</sup> background.

401

### 402 **Staining and microscopy**

403 For immunostaining, intact or partially squashed testes from 2-4 days old wild type and  
404 mutant flies were processed as described earlier (White-Cooper, 2004). DAPI (1µg/ml) was  
405 used for DNA staining and Texas Red®-X Phalloidin (Invitrogen) was used in 1:250 dilution  
406 for actin visualization. Primary antibodies used were: rabbit anti-dGM130 (Abcam) and goat  
407 anti-dGolgin245 (1:100, gift of Sean Munro, Riedel et al., 2016).

408 Alexa Fluor 488 and Alexa Fluor 594 conjugated anti-rabbit secondary antibodies were  
409 from Invitrogen. The samples were mounted in Fluoromount (Southern Biotech) and imaging  
410 was done with an Olympus BX51 fluorescent microscope or an Olympus FV 1000 confocal  
411 microscope. Nuclear length was measured by ImageJ, statistical significance of differences  
412 determined using a one-way ANOVA on ranks with a Tukey post-hoc test.

413

### 414 **Antibody generation and western blotting**

415 Polyclonal antibody was raised in guinea pigs immunised using purified His-tagged  
416 fusion protein containing the N-terminal 200 residues of Scat expressed from the pET28b  
417 vector (Novagen). For immunoblotting analysis adult testes from 40 individual males of each

418 genotype were homogenised in 100 µl of Lysis buffer (10 mM Tris-HCl pH 7.5, 150 mM  
419 NaCl, 0.5 mM EDTA, 0.5% NP40, 1 mM PMSF, 1× Protease inhibitor Cocktail) at 4°C.  
420 Samples were separated on 8% SDS polyacrylamide gels (Bio-Rad) and transferred to PVDF  
421 membrane for immunoblotting. Blocking and antibody incubations were in Tris-buffered  
422 saline (Sigma-Aldrich) with 0.05% Tween-20 (TBST) containing 4% nonfat dry milk.  
423 Primary antibodies were anti-Scat diluted 1:2000 and anti-tubulin (Abcam). HRP-linked  
424 secondary antibodies (Millipore) were used at 1:5000. After incubation with the antibodies,  
425 blots were washed in TBST and imaged on X-ray film using an ECL detection kit (GE  
426 Healthcare).

427

### 428 **Acknowledgements**

429 We are grateful for the members of the Sinka group for comments on the manuscript and  
430 for their help in image processing. We are indebted to S. Munro, H. White-Cooper, J. A.  
431 Brill, M. Fuller, J. Raff and H. Kramer for fly stocks and reagents. We thank Nia Bryant for  
432 helpful comments on the manuscript. This work was supported by grants from EMBO  
433 (Installation Grant (No1825)), the Hungarian Scientific Research funds (OTKA: NF101001),  
434 and a collaborative grant from the Royal Society awarded to DU and RS. RS also thanks the  
435 Hungarian Academy of Science's Bolyai Scholarship for funding.

436

### 437 **Competing interest**

438 We have no competing interests.

439

### 440 **Author Contributions**

441 Conceived and designed the experiments: RS, DU. Performed the experiments: KF, ST,

442 Wrote the paper: RS, DU.

443



444 **References**

- 445 **Belloni, G., Sechi, S., Riparbelli, M. G., Fuller, M. T., Callaini, G. and Giansanti, M.**  
446 **G.** (2012). Mutations in Cog7 affect Golgi structure, meiotic cytokinesis and sperm  
447 development during Drosophila spermatogenesis. *J. Cell Sci.* doi: **10.1242/**  
448 **jcs.108878.**
- 449 **Berruti, G., Ripolone, M. and Ceriani, M.** (2010). USP8, a regulator of endosomal  
450 sorting, is involved in mouse acrosome biogenesis through interaction with the  
451 spermatid ESCRT-0 complex and microtubules. *Biol Reprod.* **82**, 930-939.
- 452 **Bonifacino, J. S. and Hierro, A.** (2011). Transport according to GARP: receiving  
453 retrograde cargo at the trans-Golgi network. *Trends Cell Biol.* **21**, 159-167.
- 454 **Castrillon, D. H., Gonczy, P., Alexander, S., Rawson, R., Eberhart, C. G.,**  
455 **Viswanathan, S., DiNardo, S. and Wasserman, S. A.** (1993). Toward a molecular  
456 genetic analysis of spermatogenesis in Drosophila melanogaster: characterization of  
457 male-sterile mutants generated by single P element mutagenesis. *Genetics.* **135**,  
458 489-505.
- 459 **Conibear, E., Cleck, J. N. and Stevens, T. H.** (2003). Vps51p mediates the  
460 association of the GARP (Vps52/53/54) complex with the late Golgi t-SNARE Tlg1p.  
461 *Mol. Biol. Cell.* **14**, 1610-1623.
- 462 **Conibear, E. and Stevens, T. H.** (2000). Vps52p, Vps53p, and Vps54p form a novel  
463 multisubunit complex required for protein sorting at the yeast late Golgi. *Mol. Biol.*  
464 *Cell.* **11**, 305-323.
- 465 **Engels, W. R., Johnson-Schlitz, D. M., Eggleston, W. B. and Sved, J.** (1990). High-  
466 frequency P element loss in Drosophila is homolog dependent. *Cell.* **62**, 515-525.
- 467 **Fabian, L. and Brill, J. A.** (2012). Drosophila spermiogenesis: Big things come from  
468 little packages. *Spermatogenesis.* **2**, 197-212.
- 469 **Fabrizio, J. J., Hime, G., Lemmon, S. K. and Bazinet, C.** (1998). Genetic dissection of  
470 sperm individualization in Drosophila melanogaster. *Development.* **125**, 1833-1843.
- 471 **Farkas, R. M., Giansanti, M. G., Gatti, M. and Fuller, M. T.** (2003). The Drosophila  
472 Cog5 homologue is required for cytokinesis, cell elongation, and assembly of  
473 specialized Golgi architecture during spermatogenesis. *Mol. Biol. Cell.* **14**, 190-200.
- 474 **Fuller, M. T.** (1993). Spermatogenesis. *In* The Development of Drosophila  
475 melanogaster. M. Bate and A.M. Arias, editors. Cold Spring Harbor Laboratory  
476 Press. 71-148.
- 477 **Giansanti, M. G., Belloni, G. and Gatti, M.** (2007). Rab11 is required for membrane  
478 trafficking and actomyosin ring constriction in meiotic cytokinesis of Drosophila  
479 males. *Mol Biol Cell.* **18**, 5034-5047.
- 480 **Giansanti, M. G., Bonaccorsi, S., Kurek, R., Farkas, R. M., Dimitri, P., Fuller, M. T.**  
481 **and Gatti, M.** (2006). The class I PITP giotto is required for Drosophila cytokinesis.  
482 *Curr Biol.* **16**, 195-201.
- 483 **Gunn, P. A., Gliddon, B. L., Londrigan, S. L., Lew, A. M., van Driel, I. R. and**  
484 **Gleeson, P. A.** (2011). The Golgi apparatus in the endomembrane-rich gastric

- 485 parietal cells exist as functional stable mini-stacks dispersed throughout the  
486 cytoplasm. *Biol. Cell.* **103**, 559-572.
- 487 **Hermo, L., Pelletier, R. M., Cyr, D. G. and Smith, C. E.** (2010). Surfing the Wave,  
488 Cycle, Life History, and Genes/Proteins Expressed by Testicular Germ Cells. Part 2:  
489 Changes in Spermatid Organelles Associated With Development of Spermatozoa.  
490 *Microsc Res Techniq.* **73**, 279-319.
- 491 **Hirst, J. and Carmichael, J.** (2011). A potential role for the clathrin adaptor GGA in  
492 *Drosophila* spermatogenesis. *BMC Cell Biol.* **12**, 22.
- 493 **Horton, A. C., Racz, B., Monson, E. E., Lin, A. L., Weinberg, R. J. and Ehlers, M. D.**  
494 (2005). Polarized secretory trafficking directs cargo for asymmetric dendrite growth  
495 and morphogenesis. *Neuron.* **48**, 757-771.
- 496 **Hughson, F. M. and Reinisch, K. M.** (2010). Structure and mechanism in membrane  
497 trafficking. *Curr. Opin. Cell Biol.* **22**, 454-460.
- 498 **Kierszenbaum, A. L., Rivkin, E., Tres, L. L., Yoder, B. K., Haycraft, C. J., Bornens,  
499 M. and Rios, R. M.** (2011). GMAP210 and IFT88 Are Present in the Spermatid Golgi  
500 Apparatus and Participate in the Development of the Acrosome-Acroplaxome  
501 Complex, Head-Tail Coupling Apparatus and Tail. *Dev Dynam.* **240**, 723-736.
- 502 **Kitazawa, D., Yamaguchi, M., Mori, H. and Inoue, Y. H.** (2012). COPI-mediated  
503 membrane trafficking is required for cytokinesis in *Drosophila* male meiotic divisions.  
504 *J Cell Sci.* **125**, 3649-3660.
- 505 **Kondylis, V. and Rabouille, C.** (2009). The Golgi apparatus: lessons from *Drosophila*.  
506 *FEBS Lett.* **583**, 3827-3838.
- 507 **Lavieu, G., Dunlop, M. H., Lerich, A., Zheng, H., Bottanelli, F. and Rothman, J. E.**  
508 (2014). The Golgi ribbon structure facilitates anterograde transport of large cargoes.  
509 *Mol. Biol. Cell.* **25**, 3028-3036.
- 510 **Lerer-Goldshtein, T., Bel, S., Shpungin, S., Pery, E., Motro, B., Goldstein, R. S.,  
511 Bar-Sheshet, S. I., Breitbart, H. and Nir, U.** (2010). TMF/ARA160: A key regulator  
512 of sperm development. *Dev. Biol.* **348**, 12-21.
- 513 **Marra, P., Salvatore, L., Mironov, A., Jr., Di Campli, A., Di Tullio, G., Trucco, A.,  
514 Beznoussenko, G., Mironov, A. and De Matteis, M. A.** (2007). The biogenesis of  
515 the Golgi ribbon: the roles of membrane input from the ER and of GM130. *Mol. Biol.  
516 Cell.* **18**, 1595-1608.
- 517 **Martinez-Campos, M., Basto, R., Baker, J., Kernan, M. and Raff, J. W.** (2004). The  
518 *Drosophila* pericentrin-like protein is essential for cilia/flagella function, but appears  
519 to be dispensable for mitosis. *Journal of Cell Biology.* **165**, 673-683.
- 520 **Miller, V. J., Sharma, P., Kudlyk, T. A., Frost, L., Rofe, A. P., Watson, I. J., Duden,  
521 R., Lowe, M., Lupashin, V. V. and Ungar, D.** (2013). Molecular insights into vesicle  
522 tethering at the Golgi by the Conserved Oligomeric Golgi (COG) complex and the  
523 golgin TMF. *J. Biol. Chem.* **288**, 4229-4240.
- 524 **Miller, V. J. and Ungar, D.** (2012). Re'COG'nition at the Golgi. *Traffic.* **13**, 891-897.

- 525 **Moreno, R. D., Ramalho-Santos, J., Sutovsky, P., Chan, E. K. and Schatten, G.**  
526 (2000). Vesicular traffic and golgi apparatus dynamics during mammalian  
527 spermatogenesis: implications for acrosome architecture. *Biol Reprod.* **63**, 89-98.
- 528 **Oka, T., Ungar, D., Hughson, F. M. and Krieger, M.** (2004). The COG and COPI  
529 complexes interact to control the abundance of GEARs, a subset of Golgi integral  
530 membrane proteins. *Mol. Biol. Cell.* **15**, 2423-2435.
- 531 **Paesold-Burda, P., Maag, C., Troxler, H., Foulquier, F., Kleinert, P., Schnabel, S.,**  
532 **Baumgartner, M. and Hennet, T.** (2009). Deficiency in COG5 causes a moderate  
533 form of congenital disorders of glycosylation. *Hum. Mol. Genet.* **18**, 4350-4356.
- 534 **Paiardi, C., Pasini, M. E., Gioria, M. and Berruti, G.** (2011). Failure of acrosome  
535 formation and globozoospermia in the wobbler mouse, a Vps54 spontaneous  
536 recessive mutant. *Spermatogenesis.* **1**, 52-62.
- 537 **Perez-Victoria, F. J., Mardones, G. A. and Bonifacino, J. S.** (2008). Requirement of  
538 the human GARP complex for mannose 6-phosphate-receptor-dependent sorting of  
539 cathepsin D to lysosomes. *Mol. Biol. Cell.* **19**, 2350-2362.
- 540 **Puthenveedu, M. A., Bachert, C., Puri, S., Lanni, F. and Linstedt, A. D.** (2006).  
541 GM130 and GRASP65-dependent lateral cisternal fusion allows uniform Golgi-  
542 enzyme distribution. *Nat Cell Biol.* **8**, 238-248.
- 543 **Quenneville, N. R., Chao, T. Y., McCaffery, J. M. and Conibear, E.** (2006). Domains  
544 within the GARP subunit Vps54 confer separate functions in complex assembly and  
545 early endosome recognition. *Mol. Biol. Cell.* **17**, 1859-1870.
- 546 **Raja, S. J. and Renkawitz-Pohl, R.** (2005). Replacement by *Drosophila melanogaster*  
547 protamines and Mst77F of histones during chromatin condensation in late  
548 spermatids and role of sesame in the removal of these proteins from the male  
549 pronucleus. *Mol Cell Biol.* **25**, 6165-6177.
- 550 **Reynders, E., Foulquier, F., Leao Teles, E., Quelhas, D., Morelle, W., Rabouille, C.,**  
551 **Annaert, W. and Matthijs, G.** (2009). Golgi function and dysfunction in the first  
552 COG4-deficient CDG type II patient. *Hum. Mol. Genet.* **18**, 3244-3256.
- 553 **Riedel, F., Gillingham, A. K., Rosa-Ferreira, C., Galindo, A. and Munro, S.** (2016).  
554 An antibody toolkit for the study of membrane traffic in *Drosophila melanogaster*. *Biol*  
555 *Open.*
- 556 **Robinett, C. C., Giansanti, M. G., Gatti, M. and Fuller, M. T.** (2009). TRAPP II is  
557 required for cleavage furrow ingression and localization of Rab11 in dividing male  
558 meiotic cells of *Drosophila*. *J Cell Sci.* **122**, 4526-4534.
- 559 **Rosa-Ferreira, C., Christis, C., Torres, I. L. and Munro, S.** (2015). The small G  
560 protein Arl5 contributes to endosome-to-Golgi traffic by aiding the recruitment of the  
561 GARP complex to the Golgi. *Biol Open.* **4**, 474-481.
- 562 **Schmitt-John, T., Drepper, C., Mussmann, A., Hahn, P., Kuhlmann, M., Thiel, C.,**  
563 **Hafner, M., Lengeling, A., Heimann, P., Jones, J. M., et al.** (2005). Mutation of  
564 Vps54 causes motor neuron disease and defective spermiogenesis in the wobbler  
565 mouse. *Nat Genet.* **37**, 1213-1215.

- 566 **Schnorrer, F., Schonbauer, C., Langer, C. C., Dietzl, G., Novatchkova, M.,**  
567 **Schernhuber, K., Fellner, M., Azaryan, A., Radolf, M., Stark, A., et al.** (2010).  
568 Systematic genetic analysis of muscle morphogenesis and function in *Drosophila*.  
569 *Nature*. **464**, 287-291.
- 570 **Sinka, R., Gillingham, A. K., Kondylis, V. and Munro, S.** (2008). Golgi coiled-coil  
571 proteins contain multiple binding sites for Rab family G proteins. *J. Cell Biol.* **183**,  
572 607-615.
- 573 **Texada, M. J., Simonette, R. A., Johnson, C. B., Deery, W. J. and Beckingham, K.**  
574 **M.** (2008). Yuri gagarin is required for actin, tubulin and basal body functions in  
575 *Drosophila* spermatogenesis. *J Cell Sci.* **121**, 1926-1936.
- 576 **Tokuyasu, K. T., Peacock, W. J. and Hardy, R. W.** (1972). Dynamics of  
577 spermiogenesis in *Drosophila melanogaster*. I. Individualization process. *Z Zellforsch*  
578 *Mikrosk Anat.* **124**, 479-506.
- 579 **Ungar, D., Oka, T., Brittle, E. E., Vasile, E., Lupashin, V. V., Chatterton, J. E.,**  
580 **Heuser, J. E., Krieger, M. and Waters, M. G.** (2002). Characterization of a  
581 mammalian Golgi-localized protein complex, COG, that is required for normal Golgi  
582 morphology and function. *J. Cell Biol.* **157**, 405-415.
- 583 **Vogt, N., Koch, I., Schwarz, H., Schnorrer, F. and Nusslein-Volhard, C.** (2006). The  
584 gamma TuRC components Grip75 and Grip128 have an essential microtubule-  
585 anchoring function in the *Drosophila* germline. *Development.* **133**, 3963-3972.
- 586 **Wehland, J., Henkart, M., Klausner, R. and Sandoval, I. V.** (1983). Role of  
587 microtubules in the distribution of the Golgi apparatus: effect of taxol and  
588 microinjected anti-alpha-tubulin antibodies. *Proc. Natl. Acad. Sci. USA.* **80**, 4286-  
589 4290.
- 590 **White-Cooper, H.** (2004). Spermatogenesis: analysis of meiosis and morphogenesis.  
591 *Methods Mol Biol.* **247**, 45-75.
- 592 **Whyte, J. R. C. and Munro, S.** (2001). The Sec34/35 Golgi Transport Complex is  
593 related to the exocyst, defining a family of complexes involved in multiple steps of  
594 membrane traffic. *Dev. Cell.* **1**, 527-537.
- 595 **Willett, R., Kudlyk, T., Pokrovskaya, I., Ungar, D., Duden, R. and Lupashin, V.**  
596 (2013). COG complexes forms spatial landmarks for distinct SNARE complexes.  
597 *Nature Communications.* **4**, 1553.
- 598 **Wilson, K. L., Fitch, K. R., Bafus, B. T. and Wakimoto, B. T.** (2006). Sperm plasma  
599 membrane breakdown during *Drosophila* fertilization requires sneaky, an acrosomal  
600 membrane protein. *Development.* **133**, 4871-4879.
- 601 **Wong, R., Hadjiyanni, I., Wei, H. C., Polevoy, G., McBride, R., Sem, K. P. and Brill,**  
602 **J. A.** (2005). PIP2 hydrolysis and calcium release are required for cytokinesis in  
603 *Drosophila* spermatocytes. *Current Biology.* **15**, 1401-1406.
- 604 **Wu, X., Steet, R. A., Bohorov, O., Bakker, J., Newell, J., Krieger, M., Spaapen, L.,**  
605 **Kornfeld, S. and Freeze, H. H.** (2004). Mutation of the COG complex subunit gene  
606 COG7 causes a lethal congenital disorder. *Nat. Med.* **10**, 518-523.

607 **Yasuno, Y., Kawano, J., Inoue, Y. H. and Yamamoto, M. T.** (2013). Distribution and  
608 morphological changes of the Golgi apparatus during *Drosophila* spermatogenesis.  
609 *Dev Growth Differ.* **55**, 635-647.

610

611

612

613

614 **Figure legends**

615

616 **Figure 1. Post-meiotic *Drosophila* spermatogenesis.**

617 (A) Schematic representation of spermatogenesis highlighting round and individualizing  
618 spermatids. (B) Fluorescent images of spermatids co-expressing Arl1-mCherry (acroblast)  
619 (red), GFP-PACT (basal body) (green) and Histone-GFP (nucleus) (green). (C) Confocal  
620 images of elongated cysts with antibody staining of the *cis*-Golgi marker GM130 (red)  
621 localizing at the basal end of the cysts. (D) Fluorescent images of Snky-GFP (green) in  
622 elongated 64 cell cysts marking the formed acrosome. Nuclei labelled with DAPI (blue).  
623 Scale bars, 10 $\mu$ m.

624

625 **Figure 2. *Scat*<sup>l</sup> is a null mutant and Scat protein shows Golgi and acroblast localization**  
626 **in *Drosophila* testis.**

627 (A) Schematic representation of the *scat* gene with the inserted P element in the third exon.  
628 (B) Immunoblots of testis lysates from WT, *scat*<sup>l</sup> mutant, and WT expressing a Scat-RFP  
629 transgene.  $\alpha$ -Tubulin is used as a loading control. (C-E) Confocal images of Scat-RFP (red)  
630 expressing testes immunostained for dGM130 (green). (C) primary spermatocytes, (D)  
631 meiotic spermatids, (E) elongated spermatids. Nuclei labelled with DAPI (blue). Scale bars,  
632 10 $\mu$ m.

633

634 **Figure 3. The main defects are the failure to complete nuclear elongation followed by**  
635 **scattering of the nuclei and defective acrosome organization in *scat*<sup>l</sup> and *fws*<sup>P</sup> mutants.**

636 (A-C) Investment cones (arrow) visualized by confocal microscopy using Phalloidin staining  
637 (red) and DAPI (blue) in WT, *scat*<sup>l</sup> and *fws*<sup>P</sup> spermatids containing elongated nuclei. (D-F)  
638 Protamine-GFP (green) expressed in elongating spermatids visualized in WT (D), *scat*<sup>l</sup> (E)  
639 and *fws*<sup>P</sup> (F) mutants using confocal microscopy. (G, H) Basal bodies visualized with GFP-  
640 PACT (green) in WT (G) and *scat*<sup>l</sup> mutant (H) cysts. (I-K) Fluorescence images of elongated  
641 spermatids visualized with Snky-GFP in WT (I), *scat*<sup>l</sup> (J) and *fws*<sup>P</sup> (K) mutants. Arrowheads  
642 label acrosomes in the WT and *fws*<sup>P</sup> mutant. Arrows label the lack of acrosome in *scat*<sup>l</sup> and  
643 *fws*<sup>P</sup> mutant. Nuclei labelled with DAPI (blue). Scale bars, 10 $\mu$ m. (L) Measurement of  
644 nuclear length in elongated spermatids. n=100 in each genotype,\*\*\*: p<0.001. Length is  
645 indicated in  $\mu$ m.

646

647 **Figure 4. Integrity of the Golgi ribbon is compromised at the acroblast stage in the *scat*<sup>1</sup>**  
648 **and *fws*<sup>P</sup> mutants.**

649 (A, B) Confocal micrographs of round spermatids expressing GFP-Lerp (green)  
650 immunostained for dGolgin245 (red) in WT (A) and in *scat*<sup>1</sup> (B) mutant. (C-E) Confocal  
651 micrographs of WT (C), *scat*<sup>1</sup> (D) and *fws*<sup>P</sup> (E) round spermatids stained with the Golgi  
652 markers Golgin245 (green) and GM130 (red). Nuclei are marked with DAPI (blue). Scale  
653 bars, 10µm. Arrowheads label acroblast.

654

655 **Figure 5. Schematics depicting the involvement of tethering complexes in acroblast**  
656 **integrity.**

657 Under wild type conditions (top) the functions of the COG and GARP complexes in  
658 retrograde traffic are needed for the proper morphology of the Golgi ribbon known as the  
659 acroblast during spermatogenesis. This functional acroblast is then used to form the  
660 acrosome. When COG or GARP are non-functional, such as in the *fws*<sup>P</sup> and *scat*<sup>1</sup> mutants,  
661 the Golgi ribbon spreads probably due to a lack of appropriate retrograde traffic – at the *cis*  
662 side in COG mutants, or the *trans* side in GARP mutants. This results in defective acrosome  
663 formation, nuclear elongation and ultimately failed spermatogenesis.