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Article

Greenness Assessment and Synthesis for the Bio-Based Production of the Solvent 2,2,5,5-Tetramethyloxolane (TMO)

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Abstract: 2,2,5,5-tetramethyloxolane (TMO) has recently been identified and demonstrated as a safer solvent to replace toluene, THF, and hydrocarbons in a handful of applications. Herein, several bio-based routes to TMO are presented and assessed for greenness, assisted by the CHEM21 Metrics Toolkit and *BioLogicTool* plots. Using glucose as a common starting point, two chemocatalytic routes and two biochemical routes to TMO were identified and the pathways compared using the aforementioned tools. In addition, bio-based TMO was synthesised via one of these routes; from methyl levulinate supplied by Avantium, a by-product of the sugar dehydration step during the production of 2,5-furandicarboxylic acid. First, methyl levulinate underwent triple methylation using methyl magnesium chloride (MeMgCl) to yield 2,5-dimethylhexane-2,5-diol (DHL) in high yields of 89.7%. Then DHL was converted to high purity TMO (>98.5%) by cyclodehydration using H-BEA zeolites based on the previously reported approach. Bio-based content of this TMO was confirmed by ASTM D6866-20 Method B and found to have 64% bio-based carbon, well above the threshold of 25% set by CEN/TC 411 standards and matching the anticipated content. This study represents the first demonstration of a bio-based synthesis of TMO and confirmation of bio-content by accepted standards.

Keywords: bio-based solvents; safer solvent; green metrics; platform molecules; solvent substitution



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1. Introduction

In 2017, the synthesis and testing of 2,2,5,5-tetramethyloxolane (TMO) from 2,5-dimethylhexane-2,5-diol (DHL) was published, alongside rationale for its selection as a new non-polar solvent [1]. TMO is a cyclic ether but has properties and performance remarkably similar to toluene in many applications. Importantly, unlike traditional ethers such as diethyl ether, cyclopentylmethylether (CPME) and tetrahydrofuran (THF), TMO does not form peroxides, even when exposed to bubbling air, UV light, and heat [1]. TMO been shown to behave more like toluene than other traditional ethers, 1,4-dioxane, THF and 2-methyltetrahydrofuran (2-MeTHF), in radically-initiated polymerisations [1], Grignard reactions [1], amidation reactions [1], enzyme-catalysed esterification [2] and polycondensation reactions [3]. Solute partitioning experiments showed a stronger affinity for lipophilic protic solutes (fatty acids, fatty alcohols, and phenols) compared to toluene [4,5]. A summary of TMO's properties, as reported in the 2017 article are shown in Table 1.

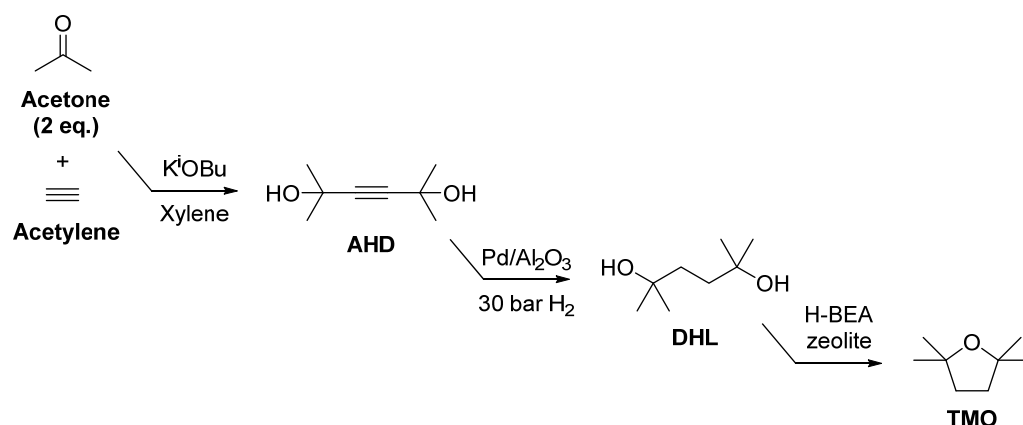
TMO was initially synthesised from petroleum-derived DHL using H-BEA zeolites, achieving very high yield and selectivity of >98.5% in a reactive distillation set-up [1]. The key DHL precursor is currently produced industrially by the reaction of acetylene and acetone, catalysed by potassium isobutoxide (KOiBu, Scheme 1), in a process patented by BASF [6].

Table 1. Properties of TMO compared to toluene and THF.

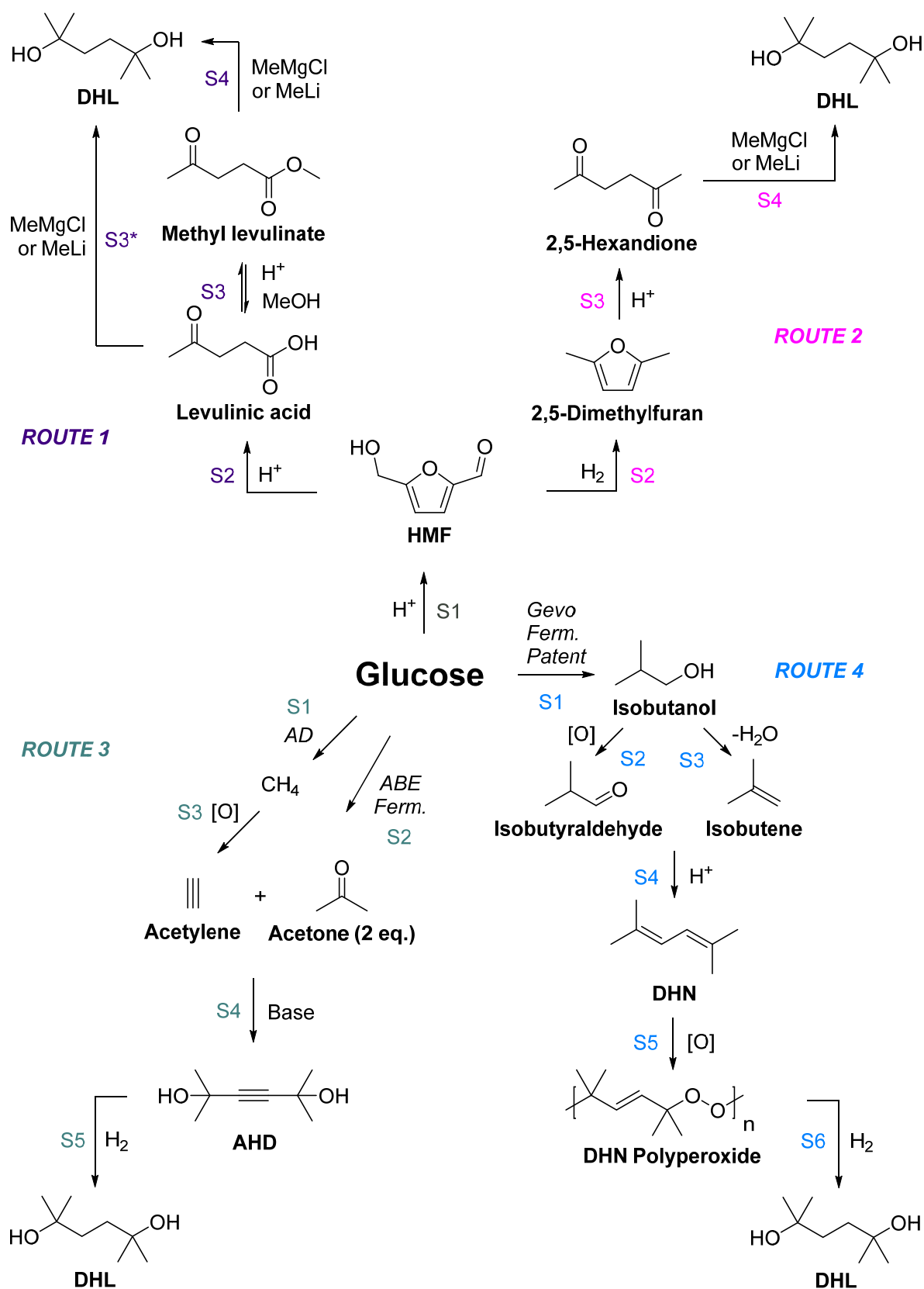
Property	TMO	Toluene	THF
Mw/g mol ^{−1}	128.25	92.14	72.11
Boiling point/°C	112 ^a	111 ^b	66 ^b
Melting point/°C	<−90 ^a	−95 ^b	−108 ^b
Density/g mL ^{−1}	0.802 ^a	0.867 ^b	0.883 ^b
Mol. Vol./cm ^{−3} mol ^{−1}	151.1 ^c	106.6 ^c	81.9 ^c
Autoignition temperature/°C	417 ^a	522 ^a	321 ^a
Lower explosion limit/v/v%	0.9 ^a	1.1 ^a	2.0 ^a
δ _D ^d	15.6 ^c	18.0 ^c	16.8 ^c
δ _P ^e	2.3 ^c	1.4 ^c	5.7 ^c
δ _H ^f	2.4 ^c	2.0 ^c	8.0 ^c
α ^g	0.00 ^j	0.00 ^j	0.00 ^j
B ^h	0.77 ^a	0.10 ^a	0.58 ^a
π* ⁱ	0.35 ^a	0.51 ^a	0.59 ^a
logP _(o/w)	1.92 ^a	2.73 ^a	0.46 ^a

^a Byrne et al., 2017, previously named 2,2,5,5-tetramethyltetrahydrofuran and abbreviated to TMTHF [1].

^b PubChem database. ^c HSPiP (5th Edition 5.1.08). ^d Hansen solubility parameter for dispersion forces. ^e Hansen solubility parameter for permanent dipole. ^f Hansen solubility parameter for hydrogen-bonding. ^g Kamlet-Abboud-Taft parameter for hydrogen-bond donating ability. ^h Kamlet-Abboud-Taft parameter for hydrogen-bond accepting ability. ⁱ Kamlet-Abboud-Taft parameter for polarity/polarizability. ^j Assumed value.

**Scheme 1.** Synthesis of 2,2,5,5-tetramethyloxolane (TMO) starting from acetone and acetylene.

The bio-based synthesis of TMO was one of the goals of the BBI-JU funded project, ReSolve [7]. Herein, alternative routes to bio-based TMO to the one described above are proposed and compared in terms of greenness with the aid of the CHEM21 Metrics Toolkit [8], and the recently presented *BioLogicTool* plots [9]. Literature data have been used for many reaction steps, while the authors have carried out certain synthetic steps experimentally in an effort to close any data gaps in the proposed routes (Scheme 2). Finally, bio-based TMO was synthesised using one of the suggested routes, via methyl levulinate, and carbon-14 quantification subsequently confirmed the bio-based content of the DHL and TMO samples (ASTM D6866-20 Method B). Avantium is in the progress of building a global first FDCA plant in Delfzijl, the Netherlands, and therefore industrially relevant quantities of methyl levulinate will become available 2024 onwards. Many potential applications have been identified for levulinic acid and its corresponding methyl ester, but the presented research in this paper is novel to the best of our knowledge [10].



Scheme 2. Four proposed bio-based synthetic routes to DHL, the precursor to TMO, and all routes from a common glucose feedstock. S3* = two possible reactions (using either MeMgCl or MeLi) were investigated for this step as part of Route 1; AD = anaerobic digestion; ABE *ferm.* = acetones, butanol and ethanol fermentation.

2. Materials and Methods

2.1. Materials

2,5-dimethylhexa-2,4-diene (DHN), was purchased from Santa Cruz Biotechnology. Methyl levulinate was supplied by Avantium. Methyl lithium solution (1.6 M in diethyl ether), 3 Å molecular sieves, methylmagnesium bromide (3 M solution in tetrahydrofuran), ammonium chloride, diethyl ether, 2,5-hexanedione and 10% Pd/C were purchased from Sigma Aldrich. Sodium chloride and magnesium sulfate and hydrochloric acid were purchased from Fischer Scientific. H-BEA zeolite catalyst (Si/Al ratio 30:1) was supplied by Clariant.

2.2. Synthesis of 2,5-Dimethylhexane-2,5-Diol from Methyl Levulinate by the Grignard Reaction—Route 1

Bio-based methyl levulinate (6.83 mL, 50 mmol), which had been dried using 3 Å molecular sieves, was added to a 100 mL round-bottomed flask along with dry tetrahydrofuran (10 mL) and a magnetic stirring bar. The flask was fitted with a Suba-Seal rubber septum and the mixture was degassed. A balloon was fitted to the rubber septum and the temperature of the mixture was reduced to 0 °C, at which point methylmagnesium bromide (3 M solution in tetrahydrofuran, 53.3 mL, 3.2 equivalents) was added dropwise using a syringe. The reaction mixture was stirred for 2 h at room temperature, after which the reaction was quenched using ammonium chloride solution. The mixture was transferred to a 250 mL separating funnel along with 25 mL diethyl ether. The organic layer was collected and washed twice with brine (10 mL). The aqueous layer was then back-extracted twice using diethyl ether (10 mL). The organic fractions were combined, dried with magnesium sulfate, and the solvent was removed in vacuo to yield crude 2,5-dimethyl-2,5-hexandiol (DHL) as an off-white powder. The powder could be recrystallised in diethyl ether to produce clear needles of 2,5-dimethyl-2,5-hexandiol with a yield of 89%. ¹H NMR (400 MHz, CDCl₃): δ = 1.18 (s, 12 H, 4x(-CH₃)), 1.53 (s, 4 H, 2x(-CH₂-)); ¹³C NMR (400 MHz, CDCl₃): δ = 29.35, 39.17, 71.37; MS (ESI): m/z 147.14 [1⁺], 169.12 [Na⁺]; Bio-based carbon content: 64%.

2.3. Synthesis of 2,5-Dimethylhexane-2,5-Diol from 2,5-Hexanedione Using Methyl lithium—Route 2

To a flame-dried, argon-filled round-bottomed flask was added 2,5-hexanedione (1 mL, 9 mmol) and 20 mL dry and degassed diethyl ether. The mixture was further purged with argon using the freeze-thaw method. The temperature was reduced to −78 °C and methyl lithium solution (1.6 M in diethyl ether, 10 mL, 16 mmol methyl lithium) was added dropwise. The reaction mixture was allowed to stir for 2 h at −78 °C before being brought to room temperature and stirred overnight. The reaction was then quenched by slowly adding water at 0 °C, before 10 mL of 2 M hydrochloric acid was added dropwise. The mixture was transferred to a separating funnel where the organic layer was collected and washed with brine (2 × 10 mL) and the combined aqueous fractions were back-extracted using diethyl ether (2 × 10 mL). The combined organic fractions were dried using magnesium sulphate and concentrated in vacuo to yield crude 2,5-dimethyl-2,5-hexandiol as an off-white powder. The powder could be recrystallised in diethyl ether to produce clear needles of 2,5-dimethyl-2,5-hexandiol with a yield of 71%. ¹H NMR (400 MHz, CDCl₃): δ = 1.24 (s, 12 H, 4x(-CH₃)), 1.58 (s, 4 H, 2x(-CH₂-)); ¹³C NMR (400 MHz, CDCl₃): δ = 29.54, 37.99, 70.81.

2.4. Oxidation of 2,5-Dimethylhexa-2,4-Diene (DHN)—Route 4

DHN (5 g, 45.4 mmol), assumed petrochemical derived, was added to a 25 mL round-bottomed flask along with 2,2,5,5-tetramethyloxolane (TMO, 10 mL) and a magnetic stirring bead. The flask was sealed with a rubber septum which was pierced with a long syringe needle connected to a compressed air source (air inlet). This syringe needle was submerged in the reaction mixture. A second and third syringe needle were then inserted into the rubber septum (air outlet) but kept above the reaction mixture level. The mixture was stirred and an air flow of 60 mL min^{−1} was passed through the air. As solvent may be

lost by evaporation out the air outlet during the reaction, fresh solvent can be added as required to maintain a viscosity that can be stirred by the magnetic stirring bead. Reaction progress was monitored by ^1H NMR spectroscopy. Upon completion of the reaction, the solvent was removed in vacuo to yield DHN polyperoxide with a yield of 97.5%. ^1H NMR (400 MHz, CDCl_3): δ 5.66 (s, 2H, $2\times(-\text{CH} =)$), 1.26 (s, 12H, $4\times(-\text{CH}_3)$).

2.5. Hydrogenation of DHN Polyperoxide—Route 4

DHN polyperoxide (1 g), 2,2,5,5-tetramethyloxolane (10 mL) and 10% Pd/C (500 mg) were added to a 20 mL Hastelloy steel autoclave. The autoclave was purged with hydrogen and then pressurised with hydrogen (10 bar). The reaction mixture was stirred for 18 h at 18 °C before being depressurised, filtered, and analysed by NMR spectroscopy.

2.6. Synthesis of 2,2,5,5-Tetramethyloxolane (TMO) from 2,5-Dimethylhexane-2,5-Diol (DHL)

2,5-dimethylhexane-2,5-diol (DHL, 10 g, 68.4 mmol) was added to a 25 mL round-bottomed flask fitted with Vigreux distillation apparatus along with H-BEA zeolite catalyst (Si/Al ratio 30:1) and a magnetic stirring bead. The reaction mixture was heated to 140 °C, at which point the 2,5-dimethylhexane-2,5-diol melted and began to react. Upon the formation of 2,2,5,5-tetramethyloxolane and water, the mixture began to reflux. The distillate was collected as two phases. The organic phase was further purified by distillation to yield 2,2,5,5-tetramethyloxolane with a purity of 99% as confirmed by GC-FID. ^1H NMR (400 MHz, CDCl_3): δ 1.81 (s, 4H, $2\times(-\text{CH}_2-)$), 1.21 (s, 12H, $4\times(-\text{CH}_3)$); ^{13}C NMR (400 MHz, CDCl_3): δ 29.75, 38.75, 80.75; MS (ESI): m/z 129.13 [1^+]; Bp: 112 °C; bio-based carbon content: 64%.

2.7. Bio-Based Carbon Content Testing

2,5-dimethylhexane-2,5-diol (DHL) and 2,2,5,5-tetramethyloxolane (TMO) from Route 1 above were tested according to ASTM D6866-20 Method B at the laboratory of BETA Analytics in Florida.

3. Results

Several synthetic routes to DHL, the precursor to TMO, from biomass were identified and can be seen in Scheme 2. A high yielding, essentially quantitative, synthesis of TMO from DHL has previously been reported [1], so no further comments are made about that final step of the synthesis in this work. Glucose is illustrated as the starting point for simplicity, but fructose, starch, cellulose or lignocellulosic biomass could also have been shown [11,12]. Indeed, lignocellulosic biomass would be the ideal source of glucose as it is a second-generation biomass source, although an additional pre-treatment step to what is shown in Scheme 2 would be required [13]. Two routes (Routes 1 and 2) proceed chemocatalytically via 5-(hydroxymethyl)furfural (HMF) and two routes (Routes 3 and 4) proceed predominately biochemically. The biochemical conversion of glucose to platform molecules methane, acetone and isobutanol in Routes 3 and 4 will not be assessed for greenness in this report due to insufficient data being available, but they will be included in the step count in Table 2. Table 2 shows the yield, conversion, selectivity, atom economy (AE), reaction mass efficiency (RME) and process mass intensity (PMI). Below is a detailed description of each Route 1-4 followed by a discussion of their accompanying *BioLogicTool* [9] and metric assessments. Metric assessment was performed using the CHEM21 Metric Toolkit [8]. The *BioLogicTool* is a new graphical tool that plots pathways from feedstock through to products against axis of % heteroatom content by mass versus molar mass [9]. The *BioLogicTool* allows for comparisons of potential routes prior to synthesis (Figure 1) and specifically looks at how rational each route is in terms of optimum use of the abundant heteroatoms in the biomass feedstock.

Table 2. Reaction metrics of each step (SX) of each Route (1 to 4) to DHL and subsequently to TMO as set out in Scheme 2.

	Yield/%	Conversion/%	Selectivity/%	AE */%	RME */%	PMI *
Route 1						
S1	n/a	n/a	n/a	n/a	n/a	n/a
S2	n/a	n/a	n/a	n/a	n/a	n/a
S3	n/a	n/a	n/a	97	1	78
S4	90	100	90	41	37	23
Route 2						
S1	56	88	64	70	39	36
S2	71	100	71	75	53	69
S3	99	100	99	100	99	12
S4	71	100	71	92	63	110
Route 3						
S1	n/a	n/a	n/a	n/a	n/a	n/a
S2	n/a	n/a	n/a	n/a	n/a	n/a
S3	63	70	90	81	51	2
S4	92	98	94	100	94	8
S5	96	100	99	100	99	3
Route 4						
S1	n/a	n/a	n/a	n/a	n/a	n/a
S2	50	50	100	97	49	2
S3	96	99	97	76	73	1
S4 @	95 (33) @	100 (35) @	95 (95) @	86 (86) @	82 (26) @	1 (3.5) @
S5	98	98	100	100	98	2
S6	87	100	87	100	84	8
DHL to TMO						
	99	100	99	88	87	1

* Metrics calculated using the CHEM21 Metrics Toolkit, the spreadsheet used for these calculations is available as a separate supplementary information file titled "CHEM21 Metrics—All Routes". AE = atom economy; RME = reaction mass efficiency; PMI = process mass intensity. @ The numbers outside the brackets are for a hypothetical multi-pass through the quoted flow reactor of Route 4 S4, the number inside the brackets are for a single pass through the flow reactor.

3.1. Route 1—Via Levulinic Acid or Methyl Levulinate

Route 1 can start both from levulinic acid as well as from methyl levulinate (ML). Levulinic acid is a desired platform molecule on its own [10,14], while ML is produced by Avantium as a by-product from their proprietary YXY[®] process to produce 2,5-furandicarboxylic acid (FDCA) from fructose [15]. Triple methylation of the carbonyl groups of levulinic acid yields DHL. In addition, levulinic acid can be esterified prior to methylation, which helps to maintain dry conditions during the reaction. Route 1 requires few steps, but the methylation of carbonyl groups raises an issue from a green perspective, as outlined below.

Levulinic acid is produced easily from biomass and there are many reports of its synthesis in the literature, either from saccharides and polysaccharides, HMF or as a by-product in the production of other chemicals [14]. However, sufficient data for the metrics calculation of its synthesis from glucose are unavailable in the literature, as only conversions based on HPLC analysis are presented [14]. In the current work, ML was supplied by Avantium, this produced as a by-product in the sugar dehydration step in their YXY process for FDCA production. Again, this makes traditional green metrics more complicated to apply, but benefits can clearly be seen [16]. Most obvious, an underutilized potential side-stream is valorised, reducing the waste of the YXY process. Furthermore, the inputs (catalysts, solvents, etc.) can be divided by the relative masses of the products for the calculation of RME and PMI for each product.

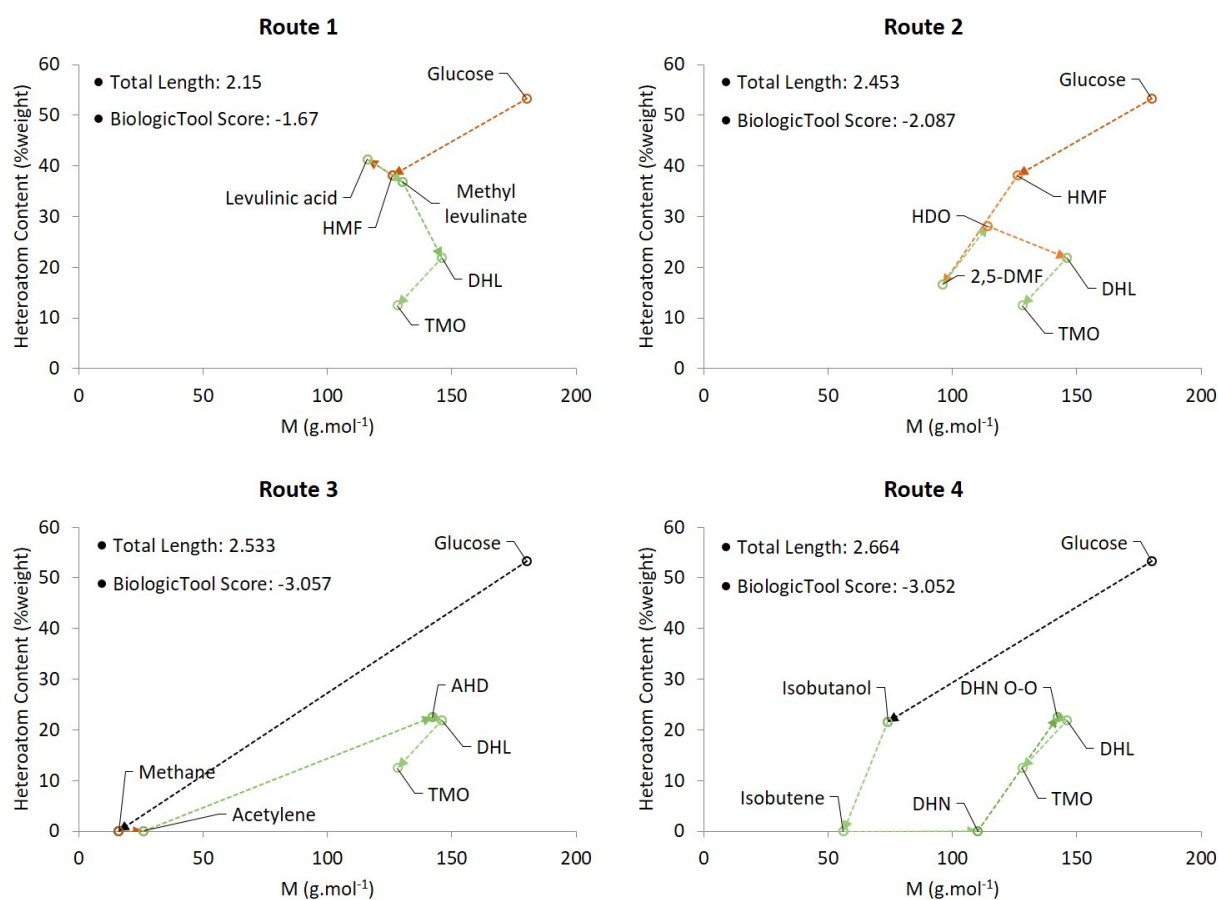


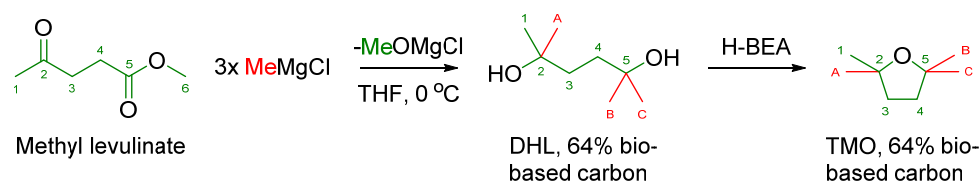
Figure 1. Plots generated by the *BioLogicTool*, showing the pathways of each route 1–4 to TMO from a common glucose feedstock for each of the four routes. The colour of the arrow is determined by the yield of that step (see Table 2); green being the highest, red being the lowest, and black where no or insufficient data was available. The *BioLogicTool* does not consider the possibility that a synthetic pathway requires the merging of two routes at any point in the synthesis. As such, the larger numerical value (ignoring minus sign) for the *BioLogicTool* score and Total Length for any convergent pathway has been presented, this deemed more representative where a convergent synthesis is used. *BioLogicTool* plots were prepared using the spreadsheet available as supplementary information accompanying the original 2019 article by Lie et al. [9].

In aqueous conditions, levulinic acid is formed instead of ML. Levulinic acid can react with alcohols to produce levulinate esters such as ML [17]. However, compared with levulinic acid, ML is easier to isolate and purify due to its significantly lower boiling point. ML is therefore the preferred starting material because it is easier to maintain dry conditions compared to levulinic acid in the subsequent moisture sensitive steps. Metrics for the esterification process are given in Table 2 based on the work of Pan et al. [18].

There are many methods of methylation of carbonyl groups, with the Grignard reaction being among the most commonly used at lab-scale. Complete conversion of the starting material was observed by the Grignard reaction and a DHL yield of 90% was achieved (Scheme 3). Subsequent ring closure of DHL to produce TMO was performed without further purification. A selectivity to TMO of >98.5% was obtained in a continuous reactive distillation, catalysed by H-BEA zeolites as previously described [1], this resulted in the isolation of the partially bio-based TMO of high purity as confirmed by NMR spectroscopy (ESI, Figure S1)

Even if the methide anion was petroleum-derived, the bio-based content of DHL produced by this route would be 62.5% (five out of eight carbons in the final structure are bio-based as can be seen in Scheme 3), well above the 25% bio-based carbon required by the CEN/TC 411 (Bio-based products). Bio-based carbon content was measured by ASTM

D6866-20 Method B and showed 64% bio-based carbon content in both the DHL and TMO subsequently produced, consistent with theoretical calculations.



Scheme 3. Synthesis of bio-based TMO via DHL from methyl levulinate. Bio-based carbon is shown in green, petroleum-derived carbon is shown in red.

While the Grignard reaction allows the production of bio-based DHL in high yields and high purity (by NMR spectroscopy, ESI Figures S2 and S3), the AE and RME are quite low at 41% and 37% respectively (ESI, CHEM21 Metrics spreadsheet, tab R1 S4). In particular, large amounts of hazardous solvent were required and stoichiometric amounts of magnesium salts produced which are difficult to recover and reuse. Furthermore, dry conditions were required, adding to the complexity of this reaction, but partially overcome through the use of ML that it is easier to dry than levulinic acid. Ultimately, Route 1 will only become suitable for efficient large-scale production of bio-based TMO when more efficient and preferably bio-based methylation routes are developed.

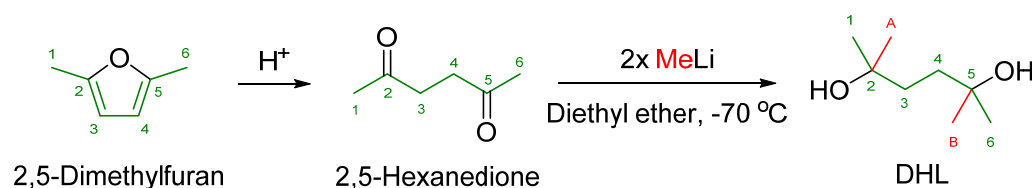
3.2. Route 2—Via 2,5-Dimethylfuran

Route 2 is a chemocatalytic route in which glucose is first dehydrated to HMF, which in turn is hydrogenated to 2,5-dimethylfuran. 2,5-Dimethylfuran can be easily converted via hydrolysis to 2,5-hexanedione (HDO) by treatment with acid and water. Methylation of HDO forms DHL which can be ring-closed to generate TMO.

The first step of this route required the production of HMF from a hexose carbohydrate source, such as glucose, cellulose or starch. The production of HMF is not trivial due to its instability in air and light, but AVA BioChem has begun production on a 20 tonne per annum scale [19]. The reaction is carried out at 220 °C and 22 bar in acidic conditions, and an in situ biomass power plant is proposed to provide energy for the chemistry [19]. The exact metrics of AVA's process cannot be calculated due to lack of data, but literature values are used to give approximate metrics in supplementary CHEM21 spreadsheet, tab R2 S1 [20]. In any case, the large-scale production of HMF represents a major breakthrough in bio-based commodity production and should be viewed positively overall due to the opportunities it will open [12].

2,5-Dimethylfuran can be easily converted to HDO in excellent isolated yields in acidic aqueous conditions. Yang et al. reported 99% yields using a biphasic system of methyl isobutyl ketone (MIBK) and acidic water [21]. Sulfuric acid was found to be the best catalyst and MIBK could extract HDO from the acidic aqueous phase prior to self-aldol reactions occurring [21]. This step is highly efficient, achieving an atom economy of 100% and RME of 99% (supplementary CHEM21 spreadsheet, tab R2 S3), meaning no purification is required prior to the next step (Table 2).

HDO can be converted to DHL in a similar manner to methyl levulinate—by methylation at the carbonyl groups (Scheme 4). HDO has the benefit of requiring only two equivalents of methylating agent, which can be sourced from methylating agents such as Grignard reagents or methyllithium. This results in an AE of 92.4% and RME of 62.5% (supplementary CHEM21 spreadsheet, tab R2 S4). The AE is significantly better than when a Grignard reagent is used, but more controlled conditions complicate the process significantly compared to the Grignard reaction.



Scheme 4. Synthesis of 2,5-dimethyl-2,5-hexanediol (DHL) via 2,5-dimethylfuran and 2,5-hexanedione.

When performing this route in the lab we selected methyllithium as the methide ion source to compare the process to that of the aforementioned Grignard reaction chosen for Route 1. HDO was used in a slight excess as we observed that methyllithium would act like a base and deprotonate HDO, causing aldol-type reactions to occur. It was found that the conditions required for methyllithium were much more sensitive than for the Grignard reaction. Dry glassware was required, reagents needed to be carefully dried and degassed, and low reaction temperatures of $-78\ ^\circ C$ were required to obtain high selectivity ($\sim 90\%$). If the stringency of the conditions was eased even slightly, by not flame drying glassware or increasing temperature ($-20\ ^\circ C$), then selectivity dramatically suffered as shown by NMR spectroscopy (ESI, Figures S4 and S5). In addition, like the Grignard reaction, a stoichiometric amount of lithium containing waste is produced. As such, this route is unlikely to be suitable for commercial production of TMO. Although HDO is readily bio-derivable, large amounts of hydrogen gas are required to form the 2,5-dimethylfuran from HMF, and the source of HDO from chemical suppliers is unknown so the DHL and TMO produced by this route was not tested for bio-based content.

3.3. Route 3—Via Bio-Acetylene and Bio-Acetone

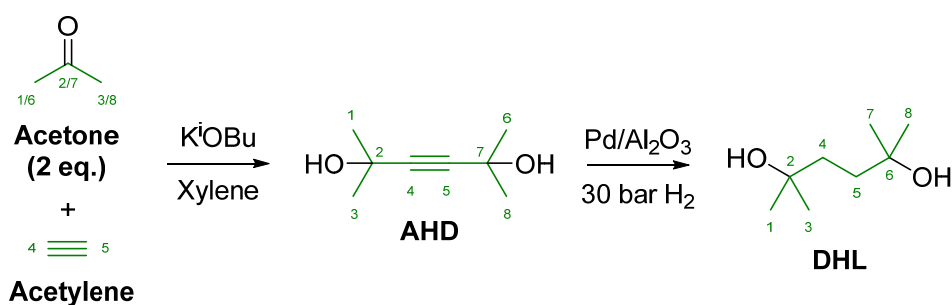
Route 3 is a bio-based equivalent to the established petroleum derived synthesis of DHL (Scheme 2). Acetone (likely sourced from the cumene process) [22,23] and methane (likely sourced from natural gas) [24] are the current starting materials for this process. Conveniently, both of these compounds are two of the most established bio-based platform chemicals—acetone is one product of the ABE (acetone-butanol-ethanol) fermentation process [25,26] (this was the primary source of acetone prior to the first world war) while methane is a product of anaerobic digestion (AD) [27,28]. A slight increase in cost between petroleum-derived and bio-based versions of acetone and methane can be expected initially, but increasing legislation, economies of scale, carbon tax and improving technology will likely make the bio-based production of acetone and methane competitive over time.

In this hypothetical bio-based route, methane is first produced by AD and then partially oxidised, generating acetylene (and syngas) [29,30]. Acetylene is reacted with two equivalents of ABE fermentation-sourced acetone using xylene as a solvent and potassium isobutoxide (KO^iBu) as a recoverable base to yield an alkyne hexanediol (AHD, Scheme 5). AHD is hydrogenated to produce 100% bio-based DHL. The benefit of this route is that the processes are all established, and a simple switch to bio-based methane and acetylene is all that is required.

The first reactions in this route involve the biochemical production of methane and acetone from glucose. Biochemical processes will not be assessed in this work as other publications with assessments of biochemical production of chemicals are available in the literature [31–33]. The oxidation of methane is a well-established process for the production of acetylene (Route 3 S1) [34]. Although acetylene yields are modest ($\sim 25\%$), work continues on the improvement of oxidative coupling of methane today using new reactor technology. For example, Khoe Dinh et al. reported methane conversions of over 70% with over 90% selectivity for acetylene using a novel rotating arc driven by AC electrical power [29].

The production of DHL from acetylene and acetone was patented by BASF in 2005 (Route 3 S2) [6]. This involved the coupling of acetylene and two equivalents of acetone to produce an alkyne, AHD (2,5-dimethylhex-3-yne-2,5-diol), followed by hydrogenation of AHD to yield DHL [6]. The yield and RME of the coupling step are excellent at 92% and

93%, respectively, and AE is 100% (Table 2). KOⁱBu in a suspension of xylene is used as the base in the reaction. Although stoichiometric amounts of base are required to deprotonate the acetylene, a simple process for the recovery and reuse of KOⁱBu is also covered in the patent [6]. This means that the potential production of huge amounts of waste is eliminated. It appears that xylene is selected as a solvent due to its polarity and relative inertness under reaction conditions. If this is the case, there are many greener solvents with similar solubility properties that could be used instead, the most obvious being TMO itself. The hydrogenation of AHD was reported as being done in flow in solventless conditions and used Pd/Al₂O₃ as the catalyst [6]. A yield of 99% was achieved with 100% atom economy (Table 2). Finally, the ring closure of DHL to produce TMO is completed as previously reported.



Scheme 5. Synthesis of DHL from acetone and acetylene via AHD.

Route 3 to TMO involves six steps, with the first two being the biochemical conversion of glucose to methane and acetone in well-established processes. Once acetone and acetylene are produced, the coupling reaction, hydrogenation reaction, and final ring closure to produce TMO can all be carried out in series, without the need for purification in between. All steps are either catalytic or use recoverable and reusable reagents (KOⁱBu) which significantly increases the likelihood of commercial viability of the overall process. However, an inability to source bio-based acetone and methane meant we were unable to perform this reaction and isolate bio-based TMO from this route.

3.4. Route 4—Via Isobutanol

Route 4 also relies on a key biochemical step, but one which now uses the platform molecule isobutanol produced via fermentation (Scheme 2). Gevo and Butamax have been developing the biosynthetic pathway to isobutanol for the past 10 years for use as a transport fuel. Furthermore, isobutanol can be used as a platform molecule to produce a wide range of other chemicals such as isobutene [35], isobutyraldehyde [36], isobutyric acid [37], isooctane [38] and para-xylene [39].

Route 4 requires both the partial oxidation of isobutanol to isobutyraldehyde and the dehydration of isobutanol to isobutene (Scheme 2) [40]. Isobutyraldehyde and isobutene can then be coupled to produce a fully bio-based diene, 2,5-dimethylhexa-2,4-diene (DHN) [40]. DHN is then oxidised to produce peroxide oligomers that can be hydrogenated to yield DHL precursor of TMO (this work).

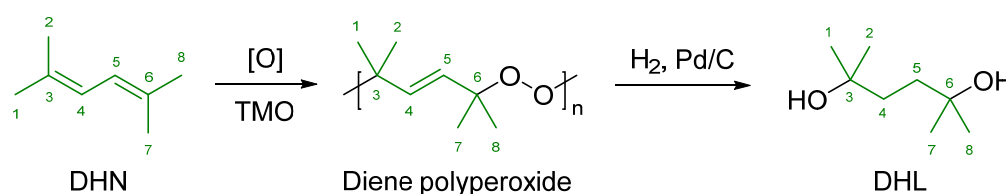
Underpinning the proposed Route 4 is a Gevo patent from 2014 reporting the synthesis of DHN in a series of high yielding flow processes [40]. In the first set of flow reactions, isobutanol is partially oxidised to isobutyraldehyde using a supported CuO catalytic system and leaving some residual unreacted isobutanol (Route 4 S1). Only partial conversion is required and complete selectivity for the desired product is achieved, with a high AE of 97.3% (Table 2). The small loss of AE is due to the elimination of hydrogen during the oxidation process. However, it will be seen that this hydrogen can in theory be recovered and reused in a later step.

The isobutanol/isobutyraldehyde mixture is then passed over a solid dehydration catalyst generating a highly pure mixture of isobutene and isobutyraldehyde (Route 4 S2).

Again, almost complete conversion and selectivity for isobutene is achieved, with the AE only being affected by the loss of water.

Isobutene and isobutyraldehyde are coupled using niobic acid to produce DHN [Route 4 S3]. Reported conversion is low at 35%, but as selectivity is high (95%) (Table 2 and CHEM21 spreadsheet, tab “R4 S3—one pass”). If unreacted starting material is simply recycled back via a flow system then a high atom economy and RME of 82% (Table 2 and CHEM21 spreadsheet, tab “R4 S3—multi-pass”). However, it does not appear that Gevo are currently pursuing this synthesis as DHN is not listed in their catalogue of commercial products at the time of writing.

DHN is an interesting molecule in that it can oxidise in air at room temperature to form oligomers of a stable polyperoxide (DHN polyperoxide, Scheme 6) [S4]. No solvent was used in the oxidation step as reported by Griesbaum et al. [41], but an extraction using methanol was required to remove unreacted diene. As TMO does not easily oxidise (unlike other common ethers such as THF and diethyl ether) [1], we successfully used TMO itself as the polymerisation solvent in the current work, usually commercially available and assumed petrochemical-derived DHN (NMR spectra in ESI, Figure S6. Our approach using TMO as solvent maintained a low viscosity of the mixture and eliminated the need of a methanol extraction. The process involved the slow bubbling of air through the mixture slowly overnight, obtaining almost complete conversion of DHN (97.5%) and minimal TMO losses. The atom economy of this step is 100%, and the polyperoxide was isolated in high purity as shown by NMR spectroscopy (ESI, Figure S7)



Scheme 6. Oxidation of 2,5-dimethylhexane-2,4-diene (DHN) and subsequent hydrogenation to produce DHL.

Hydrogenation of the DHN polyperoxide yields DHL at almost complete conversion, as described by Griesbaum et al. [41]. In that work, THF was used as the solvent for the hydrogenation step with yields of 90%. However, TMO could be a suitable solvent for the hydrogenation in a neat system where the final product is used as the solvent in several reaction steps. However, when this mixture was transferred to an autoclave along with Pd/C (10%) catalyst and pressurised with hydrogen gas (10 bar) in similar conditions to those reported by Griesbaum et al. [41], no reaction was observed. The reasons for this are unclear as there is no obvious barrier to the reaction occurring, but further research is clearly required to develop this pathway to DHL. It is also possible that hydrogen produced from the first step of this route (oxidation of isobutanol) can be recovered and used directly in this hydrogenation step. Finally, the DHL/TMO mixture can be exposed to H-BEA catalysts without the need for DHL isolation and still produce more TMO, completing a neat system where TMO is the solvent for its own production in several steps.

Overall, despite requiring a greater number of steps than the other routes, all steps are catalytic and many of the steps are carried out in flow without the need for any purification in between. When combined with the general benignity of each step, this route has many green aspects, as can be seen in Table 2.

4. Discussion

The greenness of any synthetic route to any product should be viewed holistically, i.e., no one metric is enough to rate the greenness of a process. In this section we assess the greenness using all relevant tools available. Table 2 shows the CHEM21 metrics for each step of the four routes to TMO, while the *BioLogicTool* shows that the BioLogic score and

Total BioLogic Length in Figure 1. Further questions should be asked however, which are not measurable by a score:

- Are toxic reagents used?
- Is the synthesis moisture- or air-sensitive?
- Are critical elements required?
- How quickly does the reaction proceed?
- Can it be carried out in flow?

Some of these questions are answered in the CHEM21 Metrics Toolkit spreadsheet, and can be seen viewed as a supplementary file with a separate tab for each step of each route. The total yield of final product as a percentage of biomass would be another highly informative metric for the mass efficiency of the processes, but unfortunately values for the yield is not available for each step of all routes, so a comparative overall route yield could not be calculated.

From the four routes to TMO proposed in this work, two proceeded chemocatalytically and two involved biochemical transformations. The two chemocatalytic routes require fewer steps from glucose, which is usually a good indicator of greenness as each additional unit operation likely carries greater energy and material demands. The *BioLogicTool* shows that the BioLogic score (the tortuosity of a route) and Total BioLogic Length (summation of the path lengths) of the chemocatalytic routes are better (i.e., lower values, noting that the minus symbol describes the vector direction, not length) than the biochemical routes. These conclusions from the *BioLogicTool* are roughly consistent with the fewer steps required from the shared glucose feedstock (Figure 1). Despite the better appearance of the chemocatalytic routes at this point, the synthesis as a whole must be considered, including the metrics shown on Table 2.

The two chemocatalytic routes rely on wasteful methylation steps which impact the greenness and efficiency of the synthesis to the point where it would not be commercialisable (Step 4 (S4) in Routes 1 and 2, Table 2). While AE and RME are better when methylolithium is used (92% and 63%, respectively) compared to when a Grignard reagent is used (41% and 37% respectively), the former requires more stringent dry and air-free conditions.

This leaves Routes 3 and 4 as the favoured synthetic routes to TMO. Route 3 has the benefits of being an already established process for the production of DHL, albeit with petroleum-derived starting materials. It is also analogous to the early steps of the synthesis of NMP, suggesting that the economics of the process will not be an issue. Route 4 is uncommercialised to the best of our knowledge and requires extra steps. However, little or no purification is required at any stage of Route 4, benign conditions are required throughout, and yields are high. One difficulty with Route 4 is the synthesis of 2,5-dimethylhexa-2,4-diene (DHN), Step 4, as the producer, Gevo, no longer advertise it on their product catalogue.

Overall, Route 3 is likely to be the best route to TMO due to it being an established process, requiring five steps from glucose, and having relatively good CHEM21 metrics (Table 2 and the supplementary CHEM21 spreadsheet) and BioLogic scores (Figure 1). In addition, the bio-based platform molecules used in the synthesis are acetone and methane; two of the most established bio-based molecules. Despite this assessment, it was only from Routes 1 and 2 that a sample of partially bio-based TMO could be prepared at this moment in time. Samples gathered from Route 1 were assessed via ASTM D6866-20 Method B and showed 64% bio-based carbon content in both the DHL and TMO subsequently produced.

5. Conclusions

Several routes to bio-based 2,2,5,5-tetramethyloxolane (TMO) have been proposed and assessed for greenness using the CHEM21 Metrics Toolkit and *BioLogicTool*. Two routes proceed chemocatalytically and two routes proceed predominately biochemically. TMO with a bio-based content of 64% was obtained by the methylation of bio-based methyl levulinate, surpassing the 25% required by the CEN/TC 411 bio-based content

standard. However, despite high yields (90%), atom economy, and process mass efficiency, stoichiometric amounts of magnesium salts are produced as a waste stream, meaning the coupling of acetone and acetylene is currently likely to be the greenest synthesis option, assuming bio-based acetone and acetylene are used. This study not only highlights the most logical pathway to bio-based TMO but also acts as an exemplar for the utility of the CHEM21 Metrics Toolkit and the *BioLogicTool* in comparing differing pathways to target bio-based compounds.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/suschem2030023/s1>, Figure S1: ¹H NMR spectrum of partially bio-based 2,2,5,5-tetramethyloxolane (TMO) produced by Route 1, CDCl₃ solvent, Figure S2: ¹H NMR spectrum of partially bio-based 2,5-dimethyl-2,5-hexandiol (DHL) produced by Route 1, d-MeOD solvent, Figure S3: ¹³C NMR spectrum of partially bio-based 2,5-dimethyl-2,5-hexandiol (DHL) produced by Route 1, d-MeOD solvent, Figure S4: ¹H NMR of 2,5-dimethyl-2,5-hexanediol as produced by Route 2, CDCl₃ solvent, Figure S5: ¹³C NMR of 2,5-dimethyl-2,5-hexanediol as produced by Route 2, CDCl₃ solvent, Figure S6: ¹H NMR spectrum of 2,5-dimethylhexane-2,4-diene, d-MeOD solvent, Figure S7: ¹H NMR spectrum of the polyperoxide of 2,5-dimethylhexane-2,4-diene (DHN polyperoxide), CDCl₃ solvent, Figure S8: ASTM D6866-20 Method B for bio-based carbon testing results for 2,5-dimethyl-2,4-hexanediol, Figure S9: ASTM D6866-20 Method B for bio-based carbon testing results for TMO. A supplementary spreadsheet titled “CHEM21 Metrics–All Routes” is the CHEM21 Metric Toolkit populated with available data to access the metrics for each proposed route with a separate tab for each step of each route, this was used to compile the data given in Table 2 of the main manuscript. A supplementary spreadsheet titled “BioLogicTool–All Routes” is the *BioLogicTool* spreadsheet available from the original 2019 article by Lie et al. [9] and has been populated with data for each route, this was used to compile Figure 1 of the main manuscript.

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