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**Article:**

Grewal, Jaspreet, McLuskey, Karen, Das, Debanu et al. (9 more authors) (2016) PNT1 is a C11 Cysteine Peptidase Essential for Replication of the Trypanosome Kinetoplast. *Journal of Biological Chemistry*. pp. 9492-9500. ISSN 1083-351X

<https://doi.org/10.1074/jbc.M116.714972>

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## PNT1 is a C11 Cysteine Peptidase Essential for Replication of the Trypanosome Kinetoplast

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Running Head: *Trypanosoma brucei* C11 cysteine peptidase

Keywords: Parasite, protease, RNAi, kinetoplast

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### ABSTRACT

The structure of a C11 peptidase PmC11 from the gut bacterium, *Parabacteroides merdae*, has recently been determined, enabling the identification and characterization of a C11 orthologue, PNT1, in the parasitic protozoan *Trypanosoma brucei*. A phylogenetic analysis identified PmC11 orthologues in bacteria, archaea, Chromerids, Coccidia and Kinetoplastida, the latter being the most divergent. A primary sequence alignment of PNT1 with clostripain and PmC11 revealed the position of the characteristic His-Cys catalytic dyad (H99 and C136), and an Asp (D134) in the potential S<sub>1</sub> binding site. Immunofluorescence and cryo-electron microscopy revealed that PNT1 localizes to the kinetoplast, an organelle containing the parasite's mitochondrial genome

(kDNA), with an accumulation of the protein at or near the antipodal sites. Depletion of PNT1 by RNAi in the *T. brucei* bloodstream form was lethal both in *in vitro* culture and *in vivo* in mice and the induced population accumulated cells lacking a kinetoplast. In contrast, overexpression of PNT1 led to cells having mislocated kinetoplasts. RNAi depletion of PNT1 in a kDNA independent cell line resulted in kinetoplast loss but was viable, indicating that PNT1 is required exclusively for kinetoplast maintenance. Expression of a recoded wild type *PNT1* allele, but not of an active site mutant restored parasite viability after induction *in vitro* and *in vivo* confirming that the peptidase activity of PNT1 is essential for parasite survival. These data provide evidence that PNT1 is a cysteine peptidase that is required exclusively for maintenance of the trypanosome kinetoplast.

### INTRODUCTION

*Trypanosoma brucei* is a kinetoplastid protozoan parasite and the causative agent of human African trypanosomiasis (sleeping sickness) and nagana in cattle. Sleeping sickness causes widespread human morbidity and death in sub-

Saharan Africa. Nagana leads to cattle mortality, lower meat and milk production and lower calving rate. The parasite has a complex life cycle spanning both the tsetse fly and the mammalian host. The metacyclic trypomastigote is transmitted to the mammalian host by the bite

of the tsetse fly. Once inside the human host, these parasites transform into bloodstream form (BSF) trypomastigotes that divide and multiply in blood and lymph, and which is followed by invasion of the parasites to other organs and the central nervous system. The sleep-wake cycle gets disrupted and if the disease is left untreated, the infected individual enters coma and eventually dies (1). Approximately 60 million people are at risk of being infected worldwide with this disease (WHO fact sheet May 2015). Currently no vaccines are available and the drugs in use are becoming ineffective and are toxic (2,3). It is therefore imperative that new drug targets are identified against the protozoan parasite.

Cysteine peptidases of parasitic protozoa are associated with important biological processes such as, invasion of the host cells (in case of intracellular parasites) and subsequent pathogenesis (4,5). Clan CD is comprised of peptidase families that have a protein fold similar to the caspase family (C14). Clan CD is exemplified by several important cysteine peptidases such as GPI8 (family C13), a component of the GPI:proteintransamidase complex (6); metacaspase (family C14B) (7,8); separase (family C50) (9) and a relatively less characterized family C11 (clostripain) (10). In *T. brucei* the GPI:proteintransamidase is required for anchoring proteins to the plasma membrane. Among these proteins the most prominent is VSG (variant surface glycoprotein), which forms a monolayer on the parasites surface and functions in the evasion of the host's immune system (6). *T. brucei* has five metacaspases, including MCA4, which is a pseudopeptidase and virulence factor (11) and MCA2, which is a calcium-dependent enzyme associated with RAB11 positive endosomes and does not require processing for activation (12,13). Another *T. brucei* cysteine peptidase, separase, functions in segregation of mini-chromosomes and proper mitotic assembly (14).

Recently, the first crystal structure of a family C11 peptidase, PmC11, was determined from the gut bacterium *Parabacteroides merdae* (15). This structure facilitated the identification of an important *T. brucei* protein, PNT1 (Puf Nine Target 1) as a C11 orthologue and a potential cysteine peptidase. The PNT1 transcript was previously described, in the insect procyclic form of the parasite, as the target of the RNA

binding protein PUF9 (16), with PNT1 localizing to the kinetoplast, a unique organelle containing the organism's mitochondrial DNA. The kinetoplast DNA (kDNA) of *T. brucei* is composed of a few dozen maxicircles (23kb) and several thousand minicircles (~1kb) (17). The maxicircles encode essential mitochondrial proteins including the respiratory chain complex subunits. The minicircles encode guide-RNAs that function in editing the RNA encoded by the maxicircles. The division of the mitochondrial DNA is coordinated with cytokinesis (18) and the presence of essential genes on the kinetoplast makes it imperative for each of the *T. brucei* progeny cells to inherit a kinetoplast (19). Related species, *T. equiperdum* and *T. evansi*, do not have a fully functional kinetoplast and are, therefore, known as dyskinetoplastic (20). RNAi studies on kinetoplast regulatory genes have shown that loss of the kinetoplast in *T. brucei* is lethal; hence, the organelle is considered an important drug target (19,21). Our functional analysis suggests that PNT1 is a peptidase that plays an essential role in the maintenance of the kinetoplast, and that a catalytically active PNT1 is required for the survival of *T. brucei*. This work also demonstrates that depletion of PNT1 leads to the generation of an akinetoplastic cell line, whereas overexpression leads to formation of mislocated kinetoplasts thus, inhibitors of PNT1 could be of interest for drug discovery programmes.

## EXPERIMENTAL PROCEDURES

*Phylogenetic analysis*—The peptidase C11 hidden Markov model (HMM) profile (ACC PF03415.9) was obtained from Pfam ([www.pfam.org](http://www.pfam.org)). The C11 peptidases are members of clan CD ([www.merops.org](http://www.merops.org)), a group characterised by a doublet [H(G|A)] and triplet [D(A|S)C] motif accounting for the catalytic histidine/cysteine dyad (22). The UniProtKB FASTA set (04/29/2015) was downloaded from the UniProt site ([www.uniprot.org](http://www.uniprot.org)). This sequence set was searched for peptides containing domains consistent with the model (conformant proteins) by use of the HMM search program of HMMER3 (<http://hmmer.org/>) using default parameters. The hits (full sequence E-value <0.0001) were filtered as follows: Sequences from prokaryotic and archaeal species were excluded; eukaryotic alignments were examined and only those with at least a minimum alignment to the cysteine containing

motif [D(A|S)C] retained. Full peptide sequences for the identified eukaryotic sequences were subsequently retrieved from UniprotKB; and a representative sample of bacterial and archaeal C11 peptidase sequences were obtained from the MEROPS database ([www.merops.org](http://www.merops.org)) and added to the sequence set.

The meta-method for assembling multiple sequence alignments was performed on the sequence set using the M-Coffee (23) mode of T-Coffee (24). The output alignment (color coded for alignment quality) was examined visually to determine the domain containing the doublet [H(G|A)] and triplet [D(A|S)C]. The alignment was trimmed to this domain, and an HMM profile generated (C11\_euk), using the HMMER3 program hmmbuild. This HMM profile was used to search proteome sets of a variety of eukaryotic genomes for C11\_euk conformant sequences. The domain sequences were extracted, the sequence for metacaspase TbMCA2 (*Trypanosoma brucei* Tb927.6.940, MEROPS family C14, also in clan CD) was added as an outgroup and a multiple sequence alignment was generated as above. Low quality (<4) and low occupancy (<30%) sites were removed from the alignment and a neighbour-joining tree generated from the refined alignment (SplitsTree4 (25)), [www.splitstree.org](http://www.splitstree.org)).

*Trypanosome cell culture*—*T. brucei* 2T1 bloodstream form (BSF) cells were maintained in HMI-9 medium supplemented with 10% FBS (low TET, GIBCO), 100 U mL<sup>-1</sup> penicillin 100 µg mL<sup>-1</sup> streptomycin. The parasites were transfected as described (26) and clones were obtained by limiting dilution. Appropriate concentrations of antibiotics were used for the selection and maintenance of the parasites: 2 µg mL<sup>-1</sup> puromycin, 2.5 µg mL<sup>-1</sup> phleomycin, 5 µg mL<sup>-1</sup> hygromycin B and 10 µg mL<sup>-1</sup> blasticidin (all from InvivoGen). RNAi was induced using 1 µg mL<sup>-1</sup> tetracycline. Cells were grown in 24-well plates in triplicate for growth curve experiments.

*Generation of the RNAi, recoded and Myc-tagged PNT1 constructs*—a PNT1 RNAi construct (pTL232) was generated using pGL2084, a gateway compatible version of pRPa<sup>SLi</sup> (27,28). The oligonucleotides used for the generation of sense and antisense PNT1 inserts for pTL232 were (OL4052:GGGGACAAGTTTGTACAAAAA GCAGGCTCGTCAGGACATGCTTGAAGA, and

OL4053:GGGGACCACTTTGTACAAGAAAG CTGGGTTTTGCAATTCTTTCTGTGCG).

pTL232 was digested with *AscI* and the stem loop cassette was transfected in bloodstream form *T. brucei* 2T1. A recoded gene (encoding mRNA that is refractory to RNAi) of PNT1 was designed in CLC genomics workbench (CLCbio) using the codon usage table of *T. brucei* and subsequently synthesized by GeneArt (Thermo Fisher Scientific). The recoded PNT1 gene was cloned in frame with a C-terminal HA tag in pGL2243 (29), which integrates the gene of interest at the tubulin locus for constitutive expression. Site-directed mutagenesis was carried out on the recoded PNT1 to generate an active site recoded mutant protein PNT1HA<sup>C136A</sup>. The mutations were carried out using the Quick-change Site-directed Mutagenesis kit (Stratagene) as per the manufacturer's protocol. The identifiers of PNT1 HA recoded and the mutated constructs are pGL2278 and pGL2320 respectively. The oligonucleotides used to generate the mutated recoded PNT1 were (OL4280:CGATGGTTTTTGATAGTAGTTTCA TGGCGAGCCTGG and OL4281: CCAGGCTCGCCATGAAACTACTATCAAAA ACCATCG). PNT1 was cloned in frame in a tetracycline inducible 6X C-Myc tag vector (pGL2220) (27) for generating a PNT1Myc cell line.

*Transfection of the parasites*— $1 \times 10^7$  log phase BSF cells were suspended in 100 µL of ice-cold Amaxa Human T-Cell solution (Amaxa, Koeln, Germany) and transfection carried out according to the manufacturer's protocol. Briefly, 10 µg of linearized DNA plasmid was added to the cells and transfected with the Amaxa Human Nucleofector II, using program X-001. The cells were transferred to pre-warmed HMI 11 medium and plated out in 24-well plates at 1:20 and 1:40 dilutions. The selection antibiotic was added after 6 h of transfection and cells incubated at 37 °C. Transfectants were selected after a week. F<sub>1</sub>F<sub>0</sub>-ATPase subunit γL262P mutants (20) were generated in the PNT1 RNAi cell line using gene replacement constructs (Matt Gould and Achim Schnauffer, unpublished). Briefly, two rounds of transfections were carried out to replace both the endogenous subunit γ alleles with γL262P mutant alleles. The cell lines were tested for kDNA independence by treating with 10 nM ethidium bromide for four passages, which resulted in generation of viable akinetoplastic cells.

*Analysis of trypanosome growth after induction of RNAi*—The PNT1 RNAi cell lines

were seeded in triplicate at  $1 \times 10^5$  cells per well in 24 well plates (HMI 11 hygromycin-pleuromycin media) in the presence and absence of tetracycline ( $1 \mu\text{g ml}^{-1}$ ). The cell numbers were counted using a haemocytometer every 24 h and cell seeded at  $1 \times 10^5$  cells for the subsequent time point. Infections in mice were carried out by initially inoculating a donor mouse with  $1 \times 10^5$  parasites and monitoring blood parasitemia over 72 h. Four experimental mice were infected with  $4 \times 10^4$  parasites harvested from the donor mouse. Doxycycline was added to the drinking water of two of the mice on the day of inoculation. Parasitaemia in the blood of the mice was calculated daily using a haemocytometer after drawing the blood by tail venepuncture and diluting the blood in 0.83% ammonium chloride. Cumulative growth curves were plotted for the *in vivo* and *in vitro* cell counts using Prism6 (GraphPad Software). The experiments were done in triplicate for *in vitro* and in duplicate for *in vivo* experiments.

**Real time PCR**— $2 \times 10^5$  BSF cells were grown for 24 h in the presence and absence of tetracycline, harvested and RNA was prepared using the RNeasy kit (Qiagen). First strand cDNA was prepared using  $4 \mu\text{g}$  of RNA for each sample (Invitrogen Superscript: First strand synthesis kit). The cDNA was treated with RNase H (Invitrogen) to degrade the template RNA. PCR was performed using SYBR green PCR master mix (Applied Biosystems) and GPI8 was used as an endogenous control. Oligonucleotides OL4119: CACTTGGGTGGGCTCTCTTC and OL4120: CAGCGGCTTCGACAATGG were used for amplifying PNT1, and OL2272: CGAAGCGCATTTGGATAGC and OL2273: AGCGCGTGATGACAGTGAAG to amplify GPI8. The qPCR was run and analysed on a 7300 Real time PCR system (Applied Biosystems).

**SDS-PAGE and Western blotting of *T. brucei* cell lysates**— $1 \times 10^7$  BSF parasites were lysed in sample buffer (NuPAGE sample buffer) and analysed on a 4-12% NuPAGE Novex Bis-Tris gel using MES buffer. Subsequently, the samples were electro blotted on nitrocellulose membrane (GE healthcare) and probed with the respective antibodies. The PNT1HA cell line was probed with anti-HA antibody (1:4000, Roche). The PNT1Myc cell line was probed with anti-Myc antibody (1:2000, Millipore). Anti-EF1-alpha antibody (1:5000, Millipore) was used for the loading controls.

**Immunofluorescence**—DAPI staining was carried out using  $10^6$  cells from exponential phase cultures. The cells were washed with vPBS (22,28), added to a 12-well glass slide (Thermo Fisher) pre-coated in poly-L lysine (0.1% w/v in  $\text{H}_2\text{O}$ , Sigma), and allowed to adhere for 4 min. Cells were fixed with 4% para-formaldehyde (PFA) for 5 min. The wells were washed twice with PBS (pH 7.4) and excess PFA was quenched with 100 mM glycine for 10 min. The cells were permeabilized with 0.1% Triton X-100 for 10 min. The wells were washed three times with PBS and overlaid with a drop of SlowFade Gold Antifade Reagent with DAPI (Invitrogen). Coverslips were added and the slides were kept at  $4^\circ\text{C}$  until required. Staining of PNT1HA was performed using  $1 \times 10^7$  cells from an exponential phase culture. The cells were resuspended in  $200 \mu\text{l}$  PBS followed by addition of  $300 \mu\text{L}$  of 4% PFA to fix overnight at  $4^\circ\text{C}$ . The cells were washed with PBS and resuspended in 100 mM glycine for 15 min. The cells were then centrifuged at 600 g for 10 min and resuspended in 0.1% Triton X-100 (Sigma) for 5 min. Cells were washed twice with PBS and incubated on ice for 1 h in 1% BSA (in PBS). The cells were centrifuged for 600 g for 10 min and incubated with anti-HA antibody (1:200, Rat monoclonal, Roche) for 1 h on ice. After washing with 0.1% BSA (PBS) cells were resuspended in anti-Rat Alexa-488 (1:2000, Molecular Probes) and incubated in the dark on ice for 1 h. The cells were washed with 0.1% BSA (PBS) before  $0.5 \mu\text{l}$  DAPI (100  $\mu\text{g/ml}$  stock) was added to the cells and incubated for 5 min. The cells were washed once with 0.1% BSA (PBS) and once with PBS. The final cell pellet was resuspended in final volume of PBS (30 – 50  $\mu\text{L}$ ) and stored at  $4^\circ\text{C}$  until ready to image. The staining of the PNT1Myc cell line was also carried out as described above using an anti Myc antibody conjugated to Alexa Fluor 488 (Millipore) at 1:500 dilution. Stained cells were spread on a slide and overlaid with a square coverslip. The images were acquired on a DeltaVision Core microscope using excitation wavelength of 405-489 nm and images were analysed and processed in Adobe Photoshop and FIJI.

**Transmission electron microscopy**—Cells were washed in PBS, fixed with 2.5% glutaraldehyde and 4% paraformaldehyde in 0.1 M sodium cacodylate buffer (pH 7.2), and post-fixed with 1% osmium tetroxide and 2.5% potassium ferrocyanide in 0.1 M sodium cacodylate buffer (pH 7.3) for 45 min in the dark.

Samples were *en bloc* stained with 2% aqueous uranyl acetate, dehydrated in a series of acetone solutions (30, 50, 70, 90 and 100%) and finally embedded in Epon resin. Collected ultrathin sections were stained with lead citrate prior to observation in a Tecnai T20 transmission electron microscope (FEI, Netherlands). All images were analysed and processed in Adobe Photoshop and FIJI (30). Acquired images were processed and the intensity plot profile acquired for kinetoplast density analysis. Statistical analyses were performed using Prism6, with one-way ANOVA analysis. Immuno-gold labelling of cells was done by fixing cells overnight in 4% paraformaldehyde in phosphate buffer, and dehydrating in ascending ethanol solutions. The material was progressively infiltrated with LR White resin, and polymerization was carried out in gelatine capsules under ultraviolet light. Formvar-coated nickel grids with ultrathin sections were incubated with blocking solution (3% BSA, 0.02% Tween 20, in phosphate buffer, pH 8) for 1 h. The grids were then incubated with the primary antibody anti-HA (1:200 rat monoclonal, Roche) diluted in blocking buffer for 1 h followed by several washes in blocking buffer. The grids were then incubated with protein A conjugated to immunogold (Aurion, Netherlands) for 1 h. After several washes with blocking buffer and phosphate buffer, the grids were stained with uranyl acetate and lead citrate prior to observation in a Tecnai T20 transmission electron microscope (FEI, Netherlands).

## RESULTS

*Phylogeny of the C11 cysteine peptidase family*—To determine the phylogenetic distribution of C11 proteins, the UniProt database was queried with a bespoke C11 Hidden Markov Model (HMM) domain profile. This analysis identified clostripain-like proteins in a wide range of bacterial and archaeal species and in a limited range of eukaryotic species, which could be divided into distinct clades (branches) (Fig 1A). One clade, containing a subset of species from *Chromerida*, *Coccidia* and *Chlorophyta*, is monophyletic with the archaeal and bacterial species, as exemplified by *P. merdae* PmC11 (15) and *Clostridium histolyticum* clostripain, and suggests a lateral gene transfer event from a non-eukaryotic species to a plastid-containing organism ancestral to these phyla (31). The sparse distribution of the C11 domain is indicative of frequent loss from genomes in successive rounds

of evolution, as there is no evidence of C11 proteins among species of *Babesia* or *Plasmodium*, whereas C11 proteins are found in other apicomplexan genera including *Toxoplasma*, *Eimeria* and *Neospora*. A second clade (kin) is present in kinetoplastids and is atypical in comparison to other eukaryotic species and bacterial/archaeal domains. The C11 domain is found in *T. brucei* (PNT1), *Leishmania* and *Bodo saltans*, but not in *Phytomonas*, *Leptomonas* or *Paratrypanosoma*, indicating secondary loss of the PNT1 gene in kinetoplastid groups.

*Active PNT1 is essential for Trypanosoma brucei viability*—PNT1 is the ortholog of PmC11 in *T. brucei* (Fig. 1A). A primary sequence alignment of PmC11, clostripain and PNT1 (Fig. 1B) led to the identification of the conserved His-Cys catalytic dyad (H99 and C136, respectively). PmC11 is somewhat shorter than the other two proteins but large sections of all three overlay well, covering most of the PmC11 residues with particularly good alignment around the active site. This suggests that the secondary structural elements identified in PmC11 will form the core structural elements of clostripain and PNT1.

Attempts to obtain soluble recombinant PNT1, for testing protease activity *in vitro*, were unsuccessful. However, it was possible to investigate peptidase activity and its importance in parasite survival indirectly. Initially, a *PNT1* RNAi line was generated in the bloodstream-form (BSF) of *T. brucei*, where induction by tetracycline resulted in loss of cell viability in the *in vitro* culture (Fig. 2A). PNT1 was also observed to be essential *in vivo* with doxycycline induction leading to loss of detectable parasitaemia in mice after 96 h (Fig. 2B).

To test whether catalytically active PNT1 is required for the survival of the parasites, a recoded version of *PNT1*, containing a C-terminal HA tag (*PNT1HA*, which is refractory to RNAi) was constitutively expressed in the *PNT1* RNAi line. Expression of the PNT1HA recoded protein compensated for the loss of endogenous PNT1 and restored viability of tetracycline induced parasites both *in vitro* culture (Fig. 2C) and *in vivo* (Fig. 2D). A cell line in which the recoded *PNT1HA* had the predicted active site cysteine mutated to serine PNT1HA<sup>C136S</sup> was not viable after induction *in vitro* (Fig. 2E) and *in vivo* (Fig. 2F). Quantitative RT-PCR confirmed the depletion of PNT1 mRNA in each of the cell lines (insets of Fig's.

2A,C and E) and Western blotting confirmed that PNT1HA<sup>C136S</sup> was expressed at a similar level to PNT1HA (Fig. 3A). Another interesting observation from the Western blots was the absence of processed fragments of recoded PNT1HA suggesting that PNT1 might not require processing for activity. This is in contrast to PmC11, which requires intramolecular processing for full activation (15). This observation is further supported by the primary sequence alignment of PNT1 with PmC11 and clostripain, where the cleavage site Lys (K147) in PmC11 aligns with a reported cleavage site Arg (R190) in clostripain (32), whereas in PNT1 there is an Ala at the equivalent position (Fig. 1B). Additionally, phylogenetic analysis (Fig. 1A) shows that trypanosomatid C11 orthologues are quite divergent from bacterial and archaea C11 family members. These results suggest that PNT1 is different from PmC11 in terms of activation and that the peptidase activity of PNT1 is essential for viability of *T. brucei*.

*PNT1 localizes on the kinetoplast*—PNT1 localizes to the kinetoplast in BSF parasites and was enriched either within or near the antipodal sites, which are protein assemblies located on opposite sides of the kDNA disc involved in minicircle replication (33,34). This localization was more prominent in dividing kinetoplasts (Fig. 3B). Immunogold-labelling of the PNT1HA cell line confirmed that PNT1HA is located on the kinetoplast (Fig. 3C (i, ii)). The status of the kinetoplast in PNT1-depleted cells was investigated using TEM, which showed a lower electron density of the kinetoplast in RNAi-induced cells compared to uninduced cells (Fig's. 3(iii, iv)). This indicates that PNT1 depletion results in a reduction of material within the kinetoplast. It has been demonstrated that RNAi-mediated depletion of proteins closely associated with the kinetoplast, can cause the loss of kDNA (35,36). In order to observe the effects of downregulating PNT1 on number of nuclei and kinetoplasts, the *PNT1* RNAi line was stained with DAPI at different time points after induction. Depletion of PNT1 by RNAi resulted in increased number of cells lacking a kinetoplast (akinetoplasic, 1NOK) after 2 days (Fig. 3D). The number of 1NOK cells increased from 16% on day 2 to 46% on day 4, after RNAi induction. This increase in akinetoplasic cells suggests that PNT1 is involved in the maintenance of the kinetoplast in *T. brucei*. Interestingly, inducible overexpression of a Myc-tagged PNT1 (PNT1Myc) leads to

formation of an additional kinetoplast anterior to the nucleus, a phenotype similar to that observed in the procyclic form of *T. brucei* (16). In addition, 36% of the induced PNT1Myc cell line had a mislocated kinetoplast after 48h of induction (Fig. 4A,B). PNT1 was also localized on these incorrectly positioned kinetoplasts (Fig. 4B (lower panel)).

Overexpression of PNT1 led to an increase of 1N2K cells from 12% to 39% after 24 h of induction, compared to 12% observed in uninduced cells (Fig. 4C). However, the number of 1N2K cells in the induced culture decreased to 14% and 8% at 48 h and 72 h time points, respectively. The *T. brucei* cell cycle is a well-co-ordinated event in which the kinetoplast is placed posterior to the nucleus during cytokinesis. In cells overexpressing PNT1, mislocated kinetoplasts found anterior to the nucleus were observed in cells at all stages of the cell cycle. The decrease in the number of kinetoplasts might be due to continuous cytokinesis of cells having mislocated kinetoplasts. PNT1 overexpression by tetracycline induction also leads to loss of cell viability in BSF *T. brucei* (Fig. 4D). Importantly, cell death is observed when PNT1 is either downregulated or overexpressed, indicating that an optimal amount of PNT1 peptidase activity is essential for viability of the parasite.

*PNT1 is required exclusively for maintenance of the kinetoplast*—A kinetoplast-independent *PNT1* RNAi cell line was generated to investigate whether PNT1 is required exclusively for kinetoplast maintenance or whether it has other essential mitochondrial functions. It has recently been demonstrated that an L262P mutation in the F1 $\gamma$  subunit of the F1-F0 ATPase complex allows BSF *T. brucei* to survive loss of the kinetoplast (20). The *PNT1* RNAi line was modified to replace both of the wild-type  $\gamma$  alleles with L262P mutant alleles. The *PNT1* RNAi was then induced with tetracycline. No growth defect was observed after induction (Fig.5A, left panel) despite greater than 90% depletion of the *PNT1* transcript (Fig. 5A, right panel). Additionally, depletion of PNT1 resulted in a 100% akinetoplasic cell population after 4 days of induction. Thus, PNT1 is redundant in the absence of a requirement for the kinetoplast for cell viability (Fig. 5B). Taken together, these data indicate that PNT1 is exclusively required for kinetoplast maintenance and becomes

redundant in the absence of a cellular requirement for the organelle.

## DISCUSSION

The structure of a bacterial C11 protein (PmC11) (15) allowed the catalytic dyad of the PNT1 orthologue to be identified and probed. Despite being larger than PmC11, a basic primary sequence alignment suggests that the secondary structural elements identified in PmC11 will form the core structural elements of PNT1 (Fig 1B). Our data and others (16) indicate that PNT1 is present at the ends of the kinetoplast, which could be the antipodal sites where newly replicated minicircles reattach to the network for repairing discontinuities during replication (such as gap filling and ligation) (37,38). Several proteins have been identified at the antipodal sites of the kinetoplast including helicases (33,39), a minicircle origin binding protein p38 (40), DNA polymerase  $\beta$  (41), and a DNA primase PRI1 (42), among others. Depletion of p38 (40), PIF8 (33) and PRI1 (42) also result in loss of kDNA, a similar phenotype to PNT1 depletion; however, there is no evidence to suggest that these proteins require proteolytic processing whereas it is the peptidase activity of PNT1 that is essential. To date, only one other protease, HslVU (43) has been found to regulate kDNA maintenance in *T. brucei*. Unlike PNT1, downregulation of HslVU leads to over-replication of minicircles and formation of large kDNA networks. Moreover, HslVU localizes

throughout the mitochondrion, suggesting that HslVU could have other roles in the organelle, whereas PNT1 is required exclusively for kDNA maintenance. The appearance of an additional kinetoplast in an abnormal position observed after PNT1 overexpression is reminiscent of the ‘ancillary’ kinetoplasts reported after ablation of, DNA polymerase IC (44) or the acyl carrier protein (45), and of those observed in candidate gametes of *T. brucei* (46). Phylogenetic analysis of PNT1 orthologues suggests that kinetoplastid PNT1 genes are very divergent from orthologues of other organisms. The sequence alignment of PNT1 with PmC11 revealed the presence of conserved structural elements, but also showed novel elements indicating that they have evolved to perform some kinetoplastid-specific function. Not all kinetoplastids have PNT1; *Phytomonas* and *Leptomonas* appear to lack a PmC11 orthologue, which might indicate that there are alternative regulatory mechanisms for kDNA replication in these organisms. It is also interesting to note that *T. evansi* and *T. equiperdum* have a PNT1 gene, even though these organisms are dyskinetoplastic (47). It is possible that PNT1 is required for maintenance of minicircles in these organisms, although there are no data currently to support this hypothesis. Further work on the identification of PNT1 substrates would provide exciting new insights into the mechanism of kinetoplast replication and segregation in *T. brucei* as well as guiding inhibitor and possibly drug discovery.

**ACKNOWLEDGEMENTS.** This work was supported by the Medical Research Council (MR/K019384) and the Wellcome Trust (091790, 104111). We thank Torsten Ochsenreiter for helpful discussions.

**Conflict of interest:** The authors declare that they have no conflicts of interest with the contents of this article.

**Author contributions:** JSG, KM, DD, JCM designed research; JSG, KM, EM, JW, EB, LL performed research; JSG, KM, EM, JW, LL, RB, GHC, AS, and JCM analysed data; and JSG, KM, GHC, and JCM wrote the paper.



## REFERENCES

1. Kennedy, P. G. (2013) Clinical features, diagnosis, and treatment of human African trypanosomiasis (sleeping sickness). *Lancet Neurol.* **12**, 186-194
2. Steinmann, P., Stone, C. M., Sutherland, C. S., Tanner, M., and Tediosi, F. (2015) Contemporary and emerging strategies for eliminating human African trypanosomiasis due to *Trypanosoma brucei gambiense*: review. *Trop. Med. Int. Health* **20**, 707-718
3. Simarro, P. P., Jannin, J., and Cattand, P. (2008) Eliminating human African trypanosomiasis: where do we stand and what comes next? *PLoS Med.* **5**, e55
4. Klemba, M., and Goldberg, D. E. (2002) Biological roles of proteases in parasitic protozoa. *Annu. Rev. Biochem.* **71**, 275-305
5. Mottram, J. C., Helms, M. J., Coombs, G. H., and Sajid, M. (2003) Clan CD cysteine peptidases of parasitic protozoa. *Trends Parasitol.* **19**, 182-187
6. Lillico, S., Field, M. C., Blundell, P., Coombs, G. H., and Mottram, J. C. (2003) Essential roles for GPI-anchored proteins in African trypanosomes revealed using mutants deficient in GPI8. *Mol. Biol. Cell* **14**, 1182-1194
7. McLuskey, K., Rudolf, J., Proto, W. R., Isaacs, N. W., Coombs, G. H., Moss, C. X., and Mottram, J. C. (2012) Crystal structure of a *Trypanosoma brucei* metacaspase. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 7469-7474
8. Szallies, A., Kubata, B. K., and Duzenko, M. (2002) A metacaspase of *Trypanosoma brucei* causes loss of respiration competence and clonal death in the yeast *Saccharomyces cerevisiae*. *FEBS Lett.* **517**, 144-150
9. Uhlmann, F., Wernic, D., Poupart, M. A., Koonin, E. V., and Nasmyth, K. (2000) Cleavage of cohesin by the CD clan protease separin triggers anaphase in yeast. *Cell* **103**, 375-386
10. Kocholaty, W., Weil, L., and Smith, L. (1938) Proteinase secretion and growth of *Clostridium histolyticum*. *Biochem. J.* **32**, 1685-1690
11. Proto, W. R., Castanys-Munoz, E., Black, A., Tetley, L., Moss, C. X., Juliano, L., Coombs, G. H., and Mottram, J. C. (2011) *Trypanosoma brucei* metacaspase 4 is a pseudopeptidase and a virulence factor. *J. Biol. Chem.* **286**, 39914-39925
12. Moss, C. X., Westrop, G. D., Juliano, L., Coombs, G. H., and Mottram, J. C. (2007) Metacaspase 2 of *Trypanosoma brucei* is a calcium-dependent cysteine peptidase active without processing. *FEBS Lett.* **581**, 5635-5639
13. Helms, M. J., Ambit, A., Appleton, P., Tetley, L., Coombs, G. H., and Mottram, J. C. (2006) Bloodstream form *Trypanosoma brucei* depend upon multiple metacaspases associated with RAB11-positive endosomes. *J. Cell Sci.* **119**, 1105-1117
14. Bessat, M., and Ersfeld, K. (2009) Functional characterization of cohesin SMC3 and separase and their roles in the segregation of large and minichromosomes in *Trypanosoma brucei*. *Mol. Microbiol.* **71**, 1371-1385
15. McLuskey, K., Grewal, J.S., Das, D., Godzik, A., Lesley, S.A., Deacon, A.M., Coombs, G.H., Elsliger, M.A., Wilson, I.A., and Mottram, J.C. (2016) Crystal structure and activity studies of the C11 cysteine peptidase from *Parabacteroides merdae* in the human gut microbiome. *J. Biol. Chem.*
16. Archer, S. K., Luu, V. D., de Queiroz, R. A., Brems, S., and Clayton, C. (2009) *Trypanosoma brucei* PUF9 regulates mRNAs for proteins involved in replicative processes over the cell cycle. *PLoS Pathog.* **5**, e1000565
17. Klingbeil, M. M., and Englund, P. T. (2004) Closing the gaps in kinetoplast DNA network replication. *Proc. Natl. Acad. Sci. U. S. A.* **101**, 4333-4334
18. Gluenz, E., Povelones, M. L., Englund, P. T., and Gull, K. (2011) The kinetoplast duplication cycle in *Trypanosoma brucei* is orchestrated by cytoskeleton-mediated cell morphogenesis. *Mol. Cell. Biol.* **31**, 1012-1021
19. Jensen, R. E., and Englund, P. T. (2012) Network news: the replication of kinetoplast DNA. *Annu. Rev. Microbiol.* **66**, 473-491

20. Dean, S., Gould, M. K., Dewar, C. E., and Schnauffer, A. C. (2013) Single point mutations in ATP synthase compensate for mitochondrial genome loss in trypanosomes. *Proc. Natl. Acad. Sci. U. S. A.* **110**, 14741-14746
21. Baker, N., Hamilton, G., Wilkes, J. M., Hutchinson, S., Barrett, M. P., and Horn, D. (2015) Vacuolar ATPase depletion affects mitochondrial ATPase function, kinetoplast dependency, and drug sensitivity in trypanosomes. *Proc. Natl. Acad. Sci. U. S. A.* **112**, 9112-9117
22. Nolan, D. P., Jackson, D. G., Biggs, M. J., Brabazon, E. D., Pays, A., Van Laethem, F., Paturiaux-Hanocq, F., Elliott, J. F., Voorheis, H. P., and Pays, E. (2000) Characterization of a novel alanine-rich protein located in surface microdomains in *Trypanosoma brucei*. *J. Biol. Chem.* **275**, 4072-4080
23. Wallace, I. M., O'Sullivan, O., Higgins, D. G., and Notredame, C. (2006) M-Coffee: combining multiple sequence alignment methods with T-Coffee. *Nucleic Acids Res.* **34**, 1692-1699
24. Notredame, C., Higgins, D. G., and Heringa, J. (2000) T-Coffee: A novel method for fast and accurate multiple sequence alignment. *J. Mol. Biol.* **302**, 205-217
25. Huson, D. H., and Bryant, D. (2006) Application of phylogenetic networks in evolutionary studies. *Mol. Biol. Evol.* **23**, 254-267
26. Monnerat, S., Almeida Costa, C. I., Forkert, A. C., Benz, C., Hamilton, A., Tetley, L., Burchmore, R., Novo, C., Mottram, J. C., and Hammarton, T. C. (2013) Identification and Functional Characterisation of CRK12:CYC9, a Novel Cyclin-Dependent Kinase (CDK)-Cyclin Complex in. *PLoS One* **8**, e67327
27. Alsford, S., and Horn, D. (2008) Single-locus targeting constructs for reliable regulated RNAi and transgene expression in *Trypanosoma brucei*. *Mol. Biochem. Parasitol.* **161**, 76-79
28. Jones, N. G., Thomas, E. B., Brown, E., Dickens, N. J., Hammarton, T. C., and Mottram, J. C. (2014) Regulators of *Trypanosoma brucei* cell cycle progression and differentiation identified using a kinome-wide RNAi screen. *PLoS Pathog.* **10**, e1003886
29. Moss, C. X., Brown, E., Hamilton, A., Van der Veken, P., Augustyns, K., and Mottram, J. C. (2015) An essential signal peptide peptidase identified in an RNAi screen of serine peptidases of *Trypanosoma brucei*. *PLoS One* **10**, e0123241
30. Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., Rueden, C., Saalfeld, S., Schmid, B., Tinevez, J. Y., White, D. J., Hartenstein, V., Eliceiri, K., Tomancak, P., and Cardona, A. (2012) Fiji: an open-source platform for biological-image analysis. *Nat. Methods* **9**, 676-682
31. Bhattacharya, D., Yoon, H. S., and Hackett, J. D. (2004) Photosynthetic eukaryotes unite: endosymbiosis connects the dots. *Bioessays* **26**, 50-60
32. Witte, V., Wolf, N., and Dargatz, H. (1996) Clostripain linker deletion variants yield active enzyme in *Escherichia coli*: a possible function of the linker peptide as intramolecular inhibitor of clostripain automaturation. *Curr. Microbiol.* **33**, 281-286
33. Wang, J., Englund, P. T., and Jensen, R. E. (2012) TbPIF8, a *Trypanosoma brucei* protein related to the yeast Pif1 helicase, is essential for cell viability and mitochondrial genome maintenance. *Mol. Microbiol.* **83**, 471-485
34. Ferguson, M., Torri, A. F., Ward, D. C., and Englund, P. T. (1992) In situ hybridization to the *Crithidia fasciculata* kinetoplast reveals two antipodal sites involved in kinetoplast DNA replication. *Cell* **70**, 621-629
35. Beck, K., Acestor, N., Schulfer, A., Anupama, A., Carnes, J., Panigrahi, A. K., and Stuart, K. (2013) *Trypanosoma brucei* Tb927.2.6100 is an essential protein associated with kinetoplast DNA. *Eukaryot. Cell* **12**, 970-978

36. Liu, B., Yildirim, G., Wang, J., Tolun, G., Griffith, J. D., and Englund, P. T. (2010) TbPIF1, a *Trypanosoma brucei* mitochondrial DNA helicase, is essential for kinetoplast minicircle replication. *J. Biol. Chem.* **285**, 7056-7066
37. Englund, P. T. (1978) The replication of kinetoplast DNA networks in *Crithidia fasciculata*. *Cell* **14**, 157-168
38. Perez-Morga, D. L., and Englund, P. T. (1993) The attachment of minicircles to kinetoplast DNA networks during replication. *Cell* **74**, 703-711
39. Liu, B., Wang, J., Yildirim, G., and Englund, P. T. (2009) TbPIF5 is a *Trypanosoma brucei* mitochondrial DNA helicase involved in processing of minicircle Okazaki fragments. *PLoS Pathog.* **5**, e1000589
40. Liu, B., Molina, H., Kalume, D., Pandey, A., Griffith, J. D., and Englund, P. T. (2006) Role of p38 in replication of *Trypanosoma brucei* kinetoplast DNA. *Mol. Cell. Biol.* **26**, 5382-5393
41. Saxowsky, T. T., Choudhary, G., Klingbeil, M. M., and Englund, P. T. (2003) *Trypanosoma brucei* has two distinct mitochondrial DNA polymerase beta enzymes. *J. Biol. Chem.* **278**, 49095-49101
42. Hines, J. C., and Ray, D. S. (2010) A mitochondrial DNA primase is essential for cell growth and kinetoplast DNA replication in *Trypanosoma brucei*. *Mol. Cell. Biol.* **30**, 1319-1328
43. Li, Z., Lindsay, M. E., Motyka, S. A., Englund, P. T., and Wang, C. C. (2008) Identification of a bacterial-like HslVU protease in the mitochondria of *Trypanosoma brucei* and its role in mitochondrial DNA replication. *PLoS Pathog.* **4**, e1000048
44. Klingbeil, M. M., Motyka, S. A., and Englund, P. T. (2002) Multiple mitochondrial DNA polymerases in *Trypanosoma brucei*. *Mol. Cell* **10**, 175-186
45. Clayton, A. M., Guler, J. L., Povelones, M. L., Gluenz, E., Gull, K., Smith, T. K., Jensen, R. E., and Englund, P. T. (2011) Depletion of mitochondrial acyl carrier protein in bloodstream-form *Trypanosoma brucei* causes a kinetoplast segregation defect. *Eukaryot. Cell* **10**, 286-292
46. Peacock, L., Bailey, M., Carrington, M., and Gibson, W. (2014) Meiosis and haploid gametes in the pathogen *Trypanosoma brucei*. *Curr. Biol.* **24**, 181-186
47. Schnauffer, A. (2010) Evolution of dyskinetoplastic trypanosomes: how, and how often? *Trends Parasitol.* **26**, 557-558

## FIGURE LEGENDS

**FIGURE 1.** Phylogenetic analysis of the clan CD C11 peptidase family. *A*, Unrooted neighbour joining tree of the C11 family from bacteria and archaeal species (Bact/Arch), Coccidia (Cocc), Chromerida (Chro), Chlorophyta (Chl) and Kinetoplastids (Kin; *A. Trypanosoma*, *B. Leishmania/Endotrypanum/Crithidia*, *C. Bodonida*). Clostripain (+)Uniprot ID: P09870, PmC11(\*)Uniprot ID: A7A9N3, and PNT1 (#)Uniprot ID:Q385B4 are marked. Key: 1; K2PE90, *Trypanosoma cruzi*. 2; A0A061IYW5 *T. rangeli*. 3; F9W8X5 *T. congolense*. 4; Tb927.11.6550 *T. brucei*. 5; G0UBL1 *T. vivax*. 6; Q4QH06 *Leishmania major*. 7; A0FS01001278.1 2 2 2134 *Endotrypanum monterogeii*. 8; CfaC1 13 0800 *Crithidia fasciculata*. 9; BS39455 *Bodo saltans*. 10; EQ973215 *Bacteroides fragilis*. 11; GL883824 *Paraprevotella xylaniphila*. 12; AHHG01000029 *Elizabethkingia anophelis*. 13; P09870 *Clostridium histolyticum*. 14; ADLW01000010 *Dysgonomonas mossii*. 15; AE010299 *Methanosarcina acetivorans*. 16; SNAP00000001341 *Eimeria tenella*. 17; B6ABJ5|*Cryptosporidium muris*. 18; MER037012 *Cryptosporidium parvum*. 19; Cvel 15101 *Chromera velia*. 20; A0A086LCT5 *Toxoplasma gondii*. 21 V4ZRY6 *Toxoplasma gondii*. 22; F0VJH3 *Neospora caninum*. 23; U6G6U0 *Eimeria praecox*. 24; MER494436 *Vitrella brassicaformis*. 25; D8TW66 *Volvox carteri*. 26; A8I9P1 *Chlamydomonas reinhardtii*. 27; A0A0D2MVH2 *Monoraphidium neglectum*. 28; NC 018015 *Thermococcus* sp. CL1. 29; ABYK01000024 *Arthrospira maxima*. 30; DS990529 *Aciduliprofundum boonei*. 31; A7A9N3 *Parabacteroides merdae*. 32; AJ437303 *Trypanosoma brucei* MCA2 (outgroup). *B*, Primary amino acid sequence alignment of PmC11 (from *Parabacteroides merdae*), PNT1 (from *T. brucei*) and clostripain (from *Clostridium histolyticum*) with identical residues highlighted in grey shading. The assigned secondary structure from the crystal structure of PmC11 (15) is mapped onto its sequence with the position of the PmC11 catalytic dyad and auto-catalytic cleavage site (K147) highlighted by a red star and a red triangle, respectively. Connecting loops are colored gray/black, the main  $\beta$ -sheet is in orange, with other strands in olive,  $\alpha$ - helices in blue and the nonapeptide linker of clostripain underlined in red. Sequences around the catalytic site of clostripain, PmC11, and PNT1 are aligned where identical residues are highlighted in grey.

**FIGURE 2.** Growth of *PNT1* RNAi cell lines *in vitro* and *in vivo*. *A* and *B*, *PNT1* RNAi cell line. *C* and *D* *PNT1* RNAi cell line expressing recoded PNT1HA. *E* and *F*, *PNT1* RNAi cell line expressing recoded PNT1HA<sup>C136S</sup>. *A*, *C* and *E* representative growth curves show cumulative cell counts after induction of RNAi with tetracycline *in vitro* (mean of three independent experiments) (inset: *PNT1* mRNA depletion monitored by qRT-PCR). *B*, *D* and *F*, 4 mice were inoculated intraperitoneally with  $4 \times 10^4$  parasites and RNAi was induced with doxycycline in 2 mice at the time of inoculation. Uninduced mice were culled when the parasitemia levels were higher than  $10^8$  cells/ml. The parasitemia in blood samples were plotted over time.

**FIGURE 3.** Phenotype analysis of *PNT1* RNAi, PNT1HA and PNT1Myc cell lines. *A*, Expression of recoded PNT1HA in the *PNT1* RNAi cell line. Lysates of *PNT1* RNAi cells expressing recoded PNT1HA (lanes 1 and 2) or recoded PNT1HA<sup>C136S</sup> (lanes 3 and 4) before (lanes 1 and 3) and after (lanes 2 and 4) tetracycline induction were immunoblotted and probed with anti-HA antibody. EF1 $\alpha$  was used as loading control. *B*, Immunofluorescence of PNT1HA expressed in tetracycline induced *PNT1* RNAi cell line (lower panel) shows localization within or near antipodal sites. Uninduced *PNT1* RNAi cell lines (upper panel) were used as a control. The cells were stained with anti-HA antibody (green) and DAPI (blue). A stained, nearly duplicated kinetoplast (yellow box) is shown in magnified view. Scale bar; 5 $\mu$ m. *C*, (i, ii) Immuno gold labelling of PNT1HA cell line using anti-HA antibody after 72 h of RNAi induction. Scale bar; 100 nm. (iii, iv) Electron micrographs of *PNT1* RNAi uninduced (iii) and 72 h tetracycline induced (iv) cells (scale bar; 150 nm). *D*, *PNT1* RNAi cells induced with tetracycline (Tet+) were stained with DAPI and the number of nuclei (N) and kinetoplasts (K) per cell was counted (in 200 cells) at the indicated time points and plotted as a percentage of the total. Each histogram represent mean  $\pm$ SD (n=3).

**FIGURE 4.** Overexpression of PNT1 *A*, Upper panel, Western blot showing expression of PNT1Myc using anti-Myc antibody at different time points after tetracycline induction (lanes 2,3 and 4 represent induced samples after 24, 48 and 72 h respectively). A 24 h uninduced PNT1Myc sample in lane 1 served as negative control. EF1 $\alpha$  was used as the loading control. Lower panel, Percentage of cells having anterior kinetoplasts after tetracycline induction at different time points (quantitation from 200 cells, histogram represents mean  $\pm$  SD (n=3)). *B*, Representative images showing the DAPI-stained, PNT1Myc-tagged cell line before (top panel) and after (middle panel) induction with tetracycline for 24 h. Bottom panel, Staining of PNT1Myc cell line with anti-Myc antibody conjugated to Alexa fluor 488 (green). Scale bar; 5 $\mu$ m. *C*, Counts of DAPI stained kinetoplast (K) and nuclei (N) of the PNT1Myc cell line showing a higher percentage of 1N2K cells at 24 h after induction. The number of 1N2K cells fall after 48 and 72 h (n=3, mean $\pm$ SD). *D*, cumulative growth curve of PNT1Myc cell line showing reduced cell viability after PNT1 overexpression.

**FIGURE 5.** PNT1 is exclusively required for kinetoplast function. *A*, Left panel, cumulative growth curve of PNT1  $\gamma$ L262P RNAi line over time  $\pm$  tetracycline (n=3). Right panel, the mRNA levels of PNT1 transcript were evaluated in the induced and uninduced  $\gamma$ L262P *PNT1* RNAi cell line using qRT-PCR. Each histogram represents the mean  $\pm$  SD (n=3). *B*, Loss of kinetoplast in  $\gamma$ L262P *PNT1* RNAi cell line after addition of tetracycline to downregulate PNT1. The cells were grown for 120 h  $\pm$  tetracycline and stained with DAPI. Scale bar; 5 $\mu$ m.

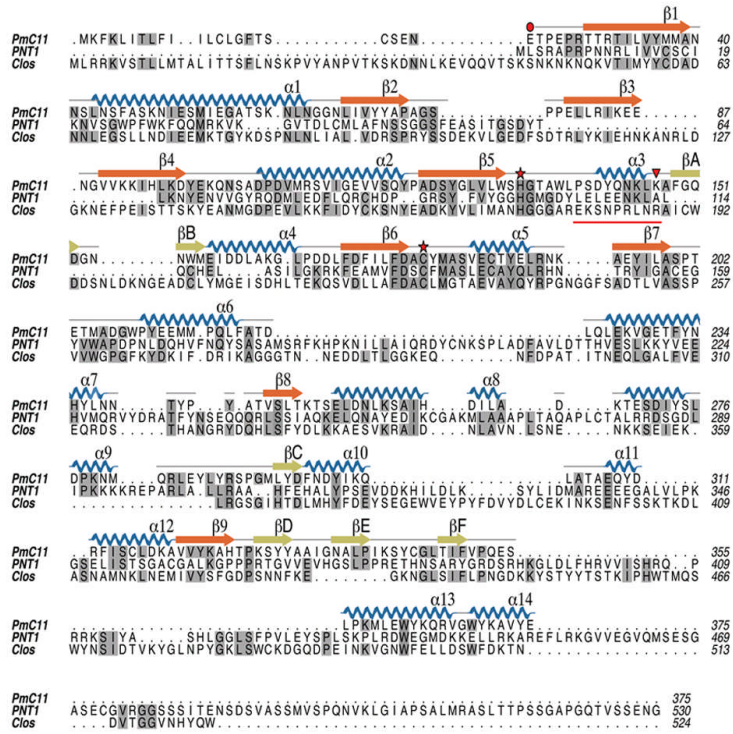
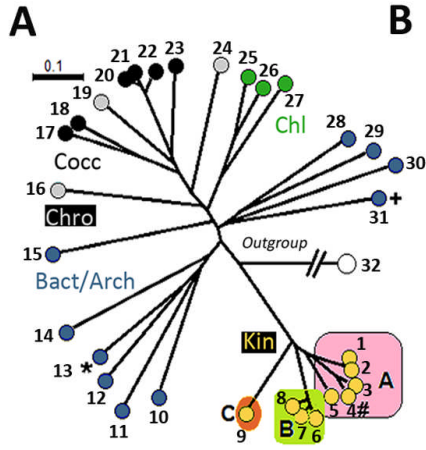


Figure 1

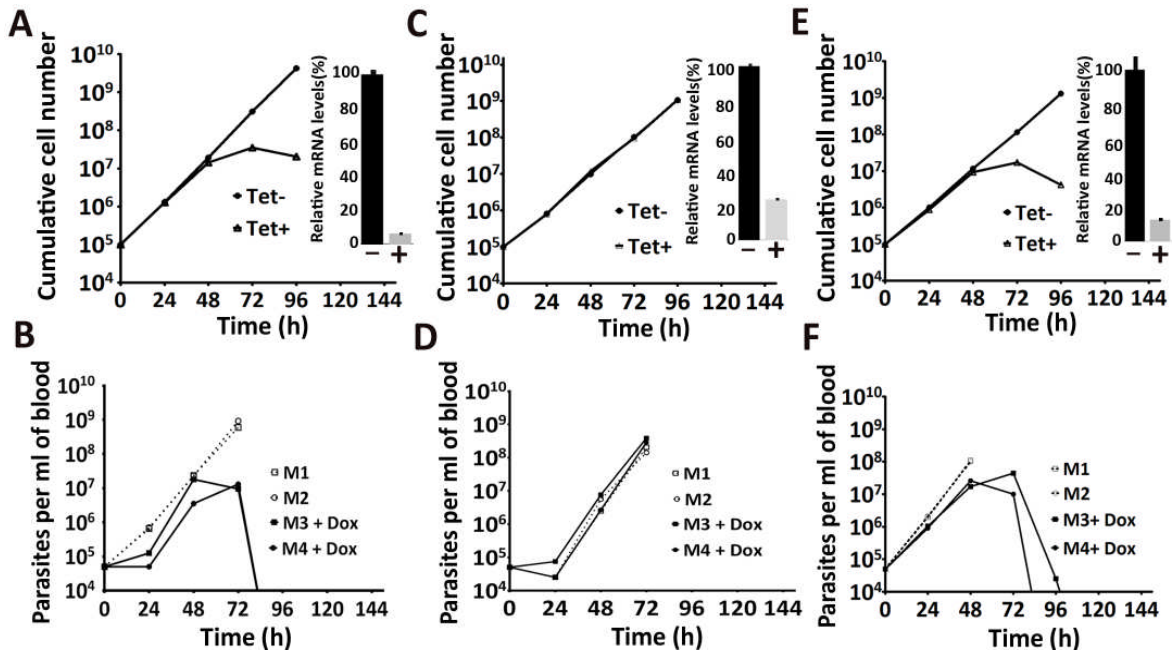


Figure 2

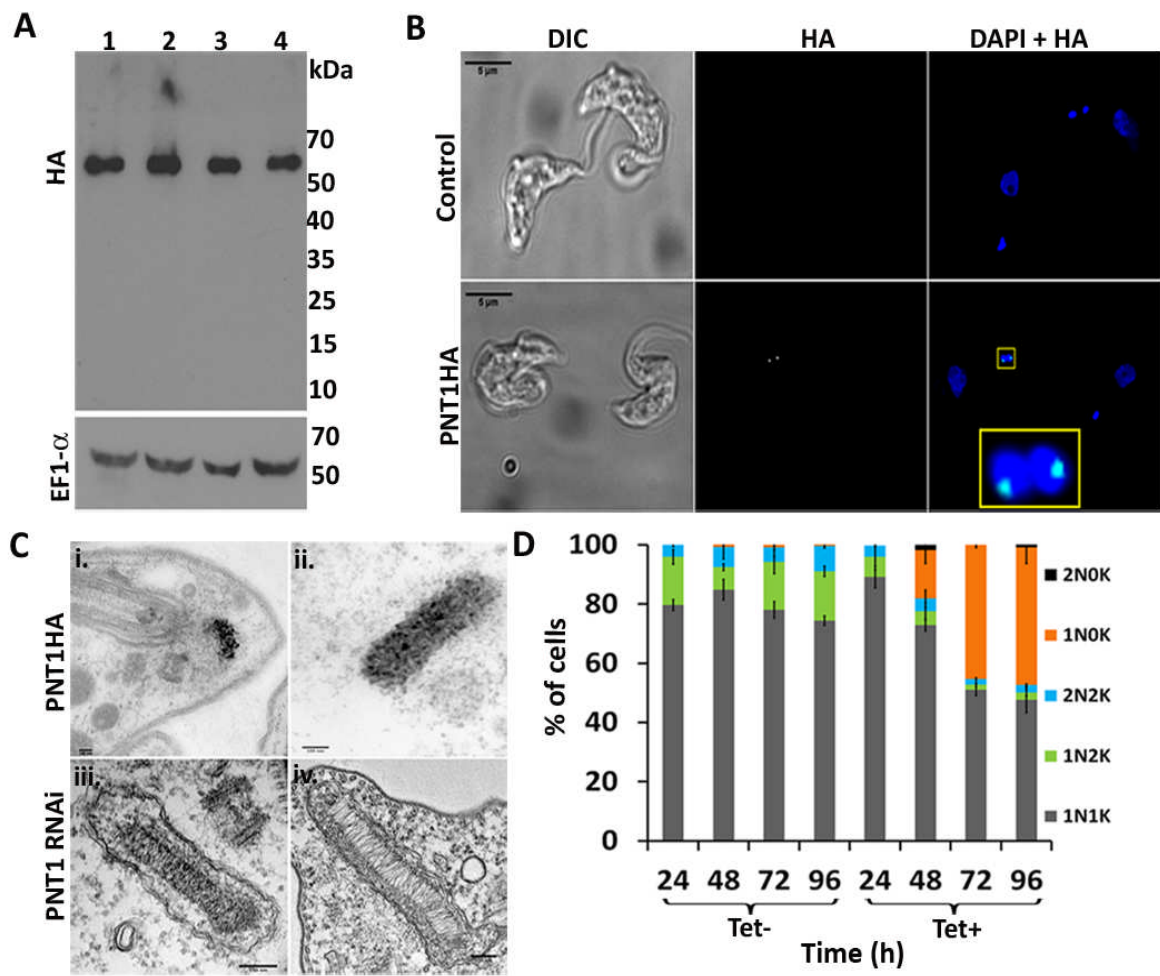


Figure 3



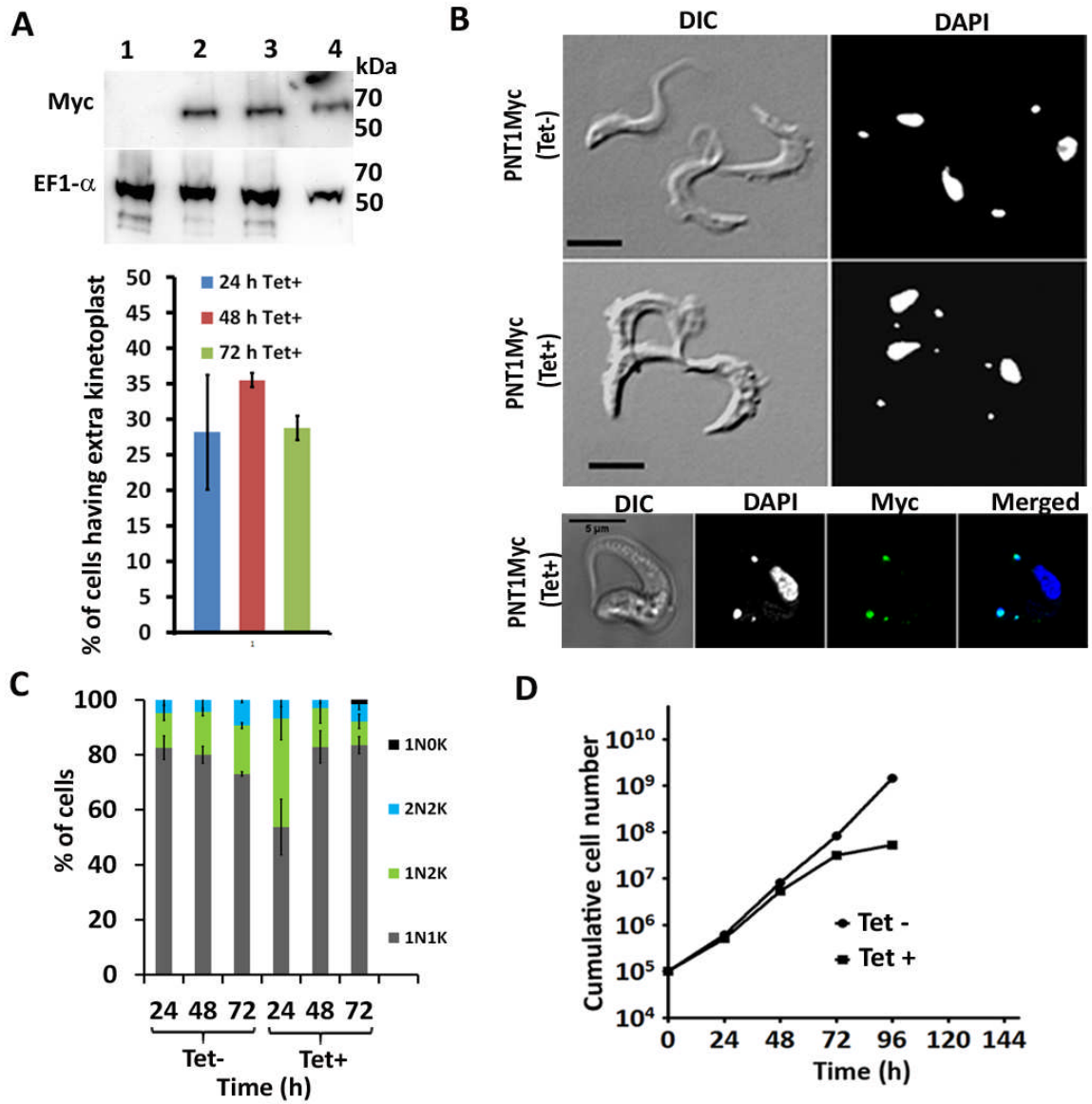


Figure 4

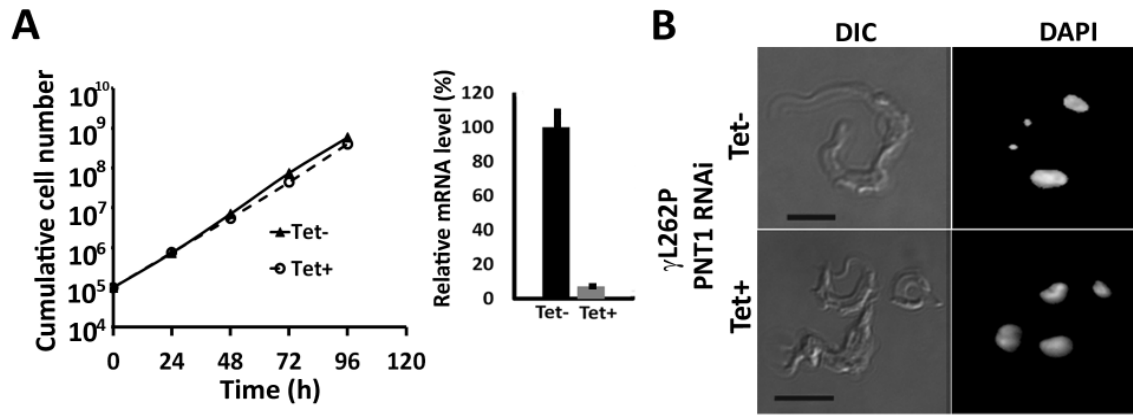


Figure 5