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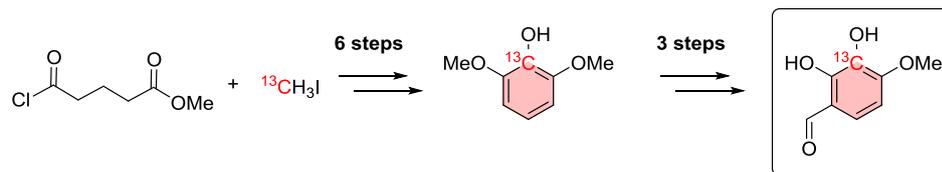
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Graphical abstract



Synthesis of [3-¹³C]-2,3-dihydroxy-4-methoxybenzaldehyde

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Abstract: An efficient synthesis of [3-¹³C]-2,3-dihydroxy-4-methoxybenzaldehyde, an isotopically labelled probe of a common intermediate used in the synthesis of a number of biologically relevant molecules, has been achieved in 9 steps from an acyclic, non-aromatic precursor. A ¹³C label for molecular imaging was introduced in a linear synthesis from the reaction of [¹³C]-labelled methyl iodide with glutaric monomethyl ester chloride. Cyclisation then aromatisation gave 1,3-dimethoxybenzene and an additional methoxy group was introduced by a formylation/Baeyer-Villiger/hydrolysis/methylation sequence. Subsequent *ortho*-formylation and selective demethylation yielded the desired [3-¹³C]-2,3-dihydroxy-4-methoxybenzaldehyde.

Keywords: Dynamic Nuclear Polarisation; ¹³C-Labeling; Baeyer-Villiger

2,3-Dihydroxy-4-methoxybenzaldehyde **1** can be used as a key intermediate in a number of natural products/drugs that have a range of biological activities (Figure 1), including the vascular disrupting agents CA1P **2** and BNC 105P **3** that are currently in Phase I and II clinical trials, respectively.¹⁻³ Another anti-cancer compound that can use this compound as a key intermediate is narciclasine **4**,⁴ shown *in vivo* to be effective against primary brain cancers and brain metastases.⁵ Amongst other molecules of interesting biological properties, thalimonine **5** is a natural product that has shown to be active against influenza and herpes simplex virus,⁶⁻⁸ whilst stenocephlavone **6** exhibits significant inhibitory activity against acetylcholinesterase, butyrylcholinesterase and lipoxygenases,⁹ and 2'-hydroxy-3',4',3,4-tetramethoxychalcone **7** shows a topical anti-inflammatory effect in mice.¹⁰ Whilst the biological activity of such compounds is usually easy to obtain, real-time molecular imaging can provide vital information on molecular targets and the metabolic fates of

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biologically active species. Dynamic nuclear polarization (DNP) is a hyperpolarization based magnetic resonance technique that can significantly increase the sensitivity of ^{13}C nuclei;¹¹ when used in conjunction with specifically labelled ^{13}C samples, the relative sensitivity is further increased, allowing *in vivo* metabolism to be investigated.

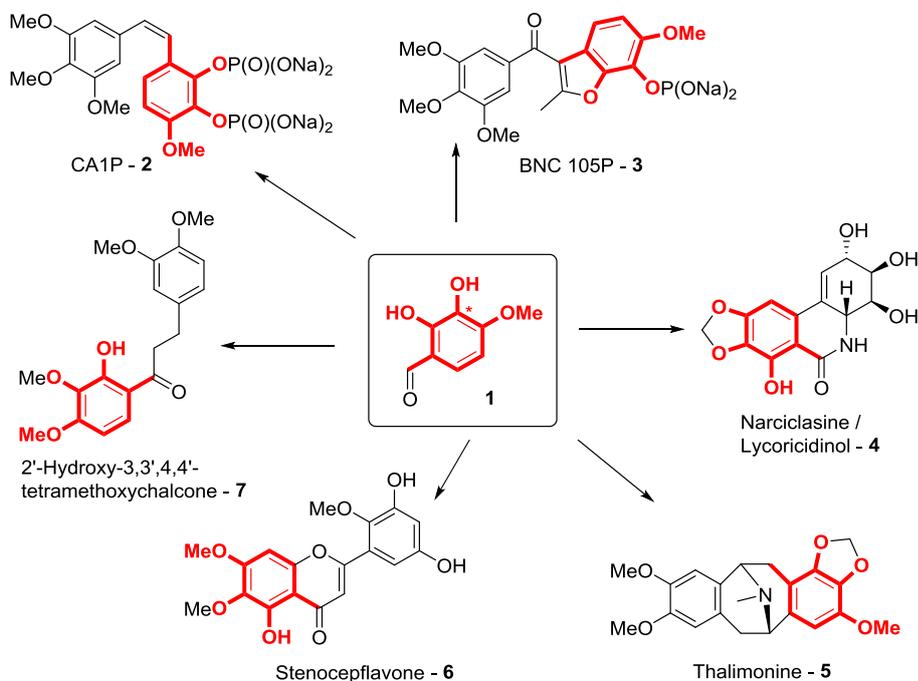
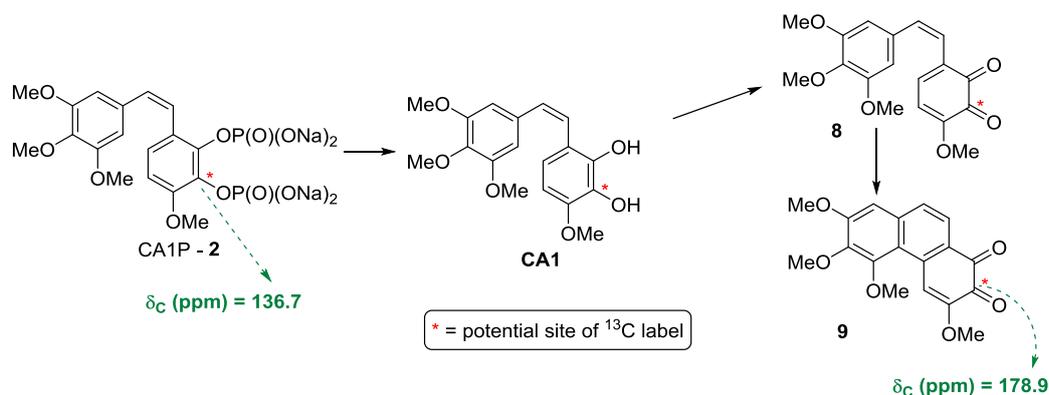


Figure 1. Potential natural products derived from key intermediate **1**

The design criteria for a ^{13}C labelled probe for use in DNP studies is critical, since this needs to be at a site in the molecule with a long T_1 relaxation time (essential for extending the lifetime of hyperpolarization), ideally on a quaternary centre remote from any neighboring spin $\frac{1}{2}$ nuclei, which cannot undergo any destructive metabolism that would remove the label from the probe. Furthermore, the label need to be located at a site where metabolic activity occurs in order to observe a change in the chemical shift of the enhanced ^{13}C signal that would be indicative of any new metabolite formed. For example, combretastatin A1P **2** is known to metabolise into *ortho*-quinone species **8** and **9** *in vivo* (Scheme 1), but the biological role of this metabolite has not yet been established.¹² The chemical shift of the highlighted ^{13}C in CA1P **2** is significantly lower than that on the same site in the *ortho*-quinone metabolite **9** [δ_c (ppm) 136.7 vs. 178.9],^{13,14} so by hyperpolarising [^{13}C]-CA1P **2**, signals for both species can easily be distinguished. Thus development of an efficient route to prepare and investigate real-time [3- ^{13}C]-2,3-dihydroxy-4-methoxybenzaldehyde **1** would provide an avenue to prepare and investigate real-time

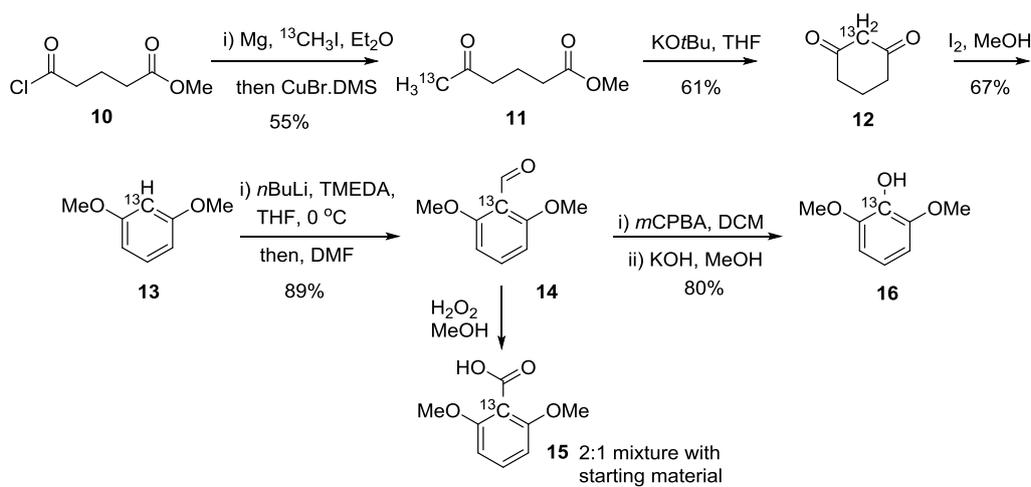
biological processes for these molecules of potential therapeutic interest. Furthermore, routes to pyrogallol derivatives of this type with site-specific incorporation of an aromatic ^{13}C label have not been reported.



Scheme 1. Known ortho-quinone metabolites of CA1P 2

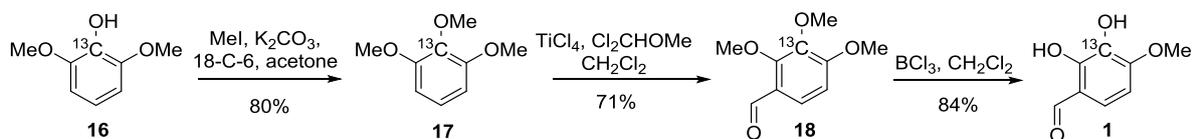
The synthesis of [3- ^{13}C]-2,3-dihydroxy-4-methoxybenzaldehyde **1** started with the installation of the ^{13}C label by modifying an existing literature method.¹⁵ In this work, Botting and co-workers treated glutaric monomethyl ester chloride **10** with [^{13}C]-lithium dimethyl cuprate, generated from [^{13}C]-methyl iodide, lithium, and copper(I) iodide, which resulted in formation of methyl 5-oxo-[6- ^{13}C]-hexanoate **11** in 43% yield. However, in the route developed herein, an improved yield of 55% for the ester **11** was obtained when the acid chloride **10** was treated with [^{13}C]-methyl magnesium iodide and copper(I) bromide dimethyl sulfide, where the [^{13}C] labelled Grignard reagent was accessed from commercially available [^{13}C]-methyl iodide (Scheme 2). This method was found to be highly reproducible and easier to operate than the original cuprate procedure. Furthermore, this method circumvents the sometimes difficult generation of [^{13}C]-labelled methyl lithium and uses stoichiometric quantities of [^{13}C]-methyl iodide, rather than losing an additional unreactive equivalent through the cuprate. Cyclisation of methyl 5-oxo-[6- ^{13}C]-hexanoate **11** to [2- ^{13}C]-cyclohexane-1,3-dione **12** was achieved in 61% yield *via* an intramolecular Claisen reaction using potassium *tert*-butoxide as the base. The product was observed as a mixture of both keto and enol tautomers in the ^1H NMR spectrum. Aromatisation of [2- ^{13}C]-cyclohexane-1,3-dione **12** to give [2- ^{13}C]-resorcinol has previously been achieved by catalytic dehydrogenation using Pd/C in refluxing xylene or triglyme with yields ranging from 45 – 88%.^{16–18} However, an alternate, more efficient route was developed that encompassed both the aromatisation and subsequent methylation steps in a one-pot reaction. Thus, dione **12** was heated at reflux with iodine in methanol to give the aromatised 1,3-dimethoxy-[2- ^{13}C]-benzene **13** in 67% yield.¹⁹ Next, the regioselective addition of a hydroxyl group in the 2-position was accomplished using a three-step strategy. Regioselective lithiation of 1,3-dimethoxy-[2- ^{13}C]-

benzene **13** with *n*BuLi at 0 °C in the presence of *N,N,N',N'*-tetramethylethylenediamine followed by treatment with dimethylformamide and gave [1-¹³C]-2,6-dimethoxybenzaldehyde **14** in 89% yield.²⁰ This was then treated with hydrogen peroxide in methanol to affect a Baeyer-Villiger reaction but unfortunately this failed to yield the desired formate ester, instead giving carboxylic acid **15** in a 2:1 ratio with starting material, formed by hydride-migration in the rearrangement step of the Baeyer-Villiger reaction. However, when aldehyde **14** was treated with *m*CPBA, the desired formate ester was formed, which was hydrolyzed using potassium hydroxide to give phenol **16** in 84% yield over the three-steps from methyl ether **13**.



Scheme 2. Synthesis of [2-¹³C]-2-hydroxy-1,3-dimethoxybenzene **16**

[2-¹³C]-2,6-Dimethoxyphenol **16** was then methylated using potassium carbonate and methyl iodide to yield [2-¹³C]-1,2,3-trimethoxybenzene **17** in 80% yield, that underwent *ortho*-formylation using Rieche conditions (titanium tetrachloride and dichloromethyl methyl ether) to give the aldehyde **18** in 71% yield (Scheme 3). Treatment of [3-¹³C]-2,3,4-trimethoxybenzaldehyde **18** with two equivalents of boron trichloride led to selective demethylation of the methyl ethers *ortho* and *meta* to the carbonyl group, providing [3-¹³C]-2,3-dihydroxy-4-methoxybenzaldehyde **1** in 84% yield. The regiochemistry of this deprotection was confirmed by correlation of the ¹³C NMR data with previously reported unlabeled material, with all chemical shifts matching to within 0.3 ppm.²¹



Scheme 3. Synthesis of [3-¹³C]-2,3-dihydroxy-4-methoxybenzaldehyde **4**

In summary, we have adapted and developed an efficient route to install a ^{13}C label at a strategically important site of a common intermediate that can be used to construct a number of biologically relevant molecules. Ongoing work is looking to exemplify the use of such an intermediate in the synthesis of the target molecules illustrated.

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Supplementary data

Supplementary data (experimental procedures and characterization data) associated with this article can be found in the online version.

References

- (1) Pettit, G. R.; Singh, S. B.; Niven, M. L.; Hamel, E.; Schmidt, J. M. *J. Nat. Prod.* **1987**, *50*, 119–131.
- (2) Tozer, G. M.; Kanthou, C.; Parkins, C. S.; Hill, S. A. *Int. J. Exp. Pathol.* **2002**, *83*, 21–38.
- (3) Kremmidiotis, G.; Leske, A. F.; Lavranos, T. C.; Beaumont, D.; Gasic, J.; Hall, A.; O’Callaghan, M.; Matthews, C. A.; Flynn, B. *Mol. Cancer Ther.* **2010**, *9*, 1562–1573.
- (4) Kornienko, A.; Evidente, A. *Chem. Rev.* **2008**, *108*, 1982–2014.
- (5) Goietsenoven, G. Van; Mathieu, V.; Lefranc, F.; Kornienko, A.; Evidente, A.; Kiss, R. *Med. Res. Rev.* **2013**, *33*, 439–455.
- (6) Velcheva, M. P.; Petrova, R. R. *J. Nat. Prod.* **1992**, *5*, 679–680.
- (7) Serkedjieva, J.; Velcheva, M. *Antivir. Chem. Chemother.* **2003**, *14*, 75–80.
- (8) Varadinova, T. L.; Shishkov, S. A.; Ivanovska, N. D.; Velcheva, M. P.; Danghaagin, S.; Samadanghiin, Z.; Yansanghiin, Z. *Phyther. Res.* **1996**, *10*, 414–417.
- (9) Shafiq, N.; Riaz, N.; Ahmed, S.; Ashraf, M.; Ejaz, S. A.; Ahmed, I.; Saleem, M.; Touseef, M. I.; Tareen, R. B.; Jabbar, A. *J. Asian Nat. Prod. Res.* **2013**, *15*, 286–293.

- (10) Ballesteros, J. F.; Sanz, M. J.; Ubeda, a; Miranda, M. a; Iborra, S.; Payá, M.; Alcaraz, M. J. *J. Med. Chem.* **1995**, *38*, 2794–2797.
- (11) Keshari, K. R.; Wilson, D. M. *Chem. Soc. Rev.* **2014**, *43*, 1627–1659.
- (12) Aprile, S.; Zaninetti, R.; Del Grosso, E.; Genazzani, A. A.; Grosa, G. *J. Pharm. Biomed. Anal.* **2013**, *78-79*, 233–242.
- (13) Pettit, G. R.; Lippert III., J. W. Preparation of combretastatin A-1 phosphate and combretastatin B-1 phosphate prodrugs with increased solubility. WO2001081355A1, November 1, 2001.
- (14) Folkes, L. K.; Christlieb, M.; Madej, E.; Stratford, M. R. L.; Wardman, P. *Chem. Res. Toxicol.* **2007**, *20*, 1885–1894.
- (15) Marshall, L. J.; Cable, K. M.; Botting, N. P. *J. Label. Compd. Radiopharm.* **2010**, *53*, 601–604.
- (16) Oldfield, M. F.; Chen, L.; Botting, N. P. *Tetrahedron* **2004**, *60*, 1887–1893.
- (17) Boyce, S. D.; Barefoot, A. C.; Hornig, J. F. *J. Label. Compd. Radiopharm.* **1983**, *20*, 243–256.
- (18) Botting, N. Synthesis of ¹³C-Labelled Estrogen Analogues. WO2004069774 (A3), 2004.
- (19) Kim, J. M.; Lee, K. Y.; Kim, J. N. *Bull. Korean Chem. Soc* **2003**, *24*, 1057–1058.
- (20) Haight, A. R.; Bailey, A. E.; Baker, W. S.; Cain, M. H.; Copp, R. R.; DeMattei, J. A.; Ford, K. L.; Henry, R. F.; Hsu, M. C.; Keyes, R. F.; King, S. A.; McLaughlin, M. A.; Melcher, L. M.; Nadler, W. R.; Oliver, P. A.; Parekh, S. I.; Patel, H. H.; Seif, L. S.; Staeger, M. A.; Wayne, G. S.; Wittenberger, S. J.; Zhang, W. *Org. Process Res. Dev.* **2004**, *8*, 897–902.
- (21) Pettit, G. R.; Thornhill, A.; Melody, N.; Knight, J. C. *J. Nat. Prod.* **2009**, *72*, 380–388.