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The multifactorial interplay of diet, the microbiome and appetite control: current knowledge and future challenges

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5 Bernard M. Corfe^{1,*}, Charlotte J Harden¹, Matthew Bull ², Iveta Garaiova³

- 6 1. Molecular Gastroenterology Research Group, Academic Unit of Surgical Oncology, Department of
 7 Oncology, University of Sheffield, Beech Hill Road, Sheffield, S10 2RX, UK
- 8 2. Organisms and Environment Division, Cardiff School of Biosciences, Cardiff University, Cardiff,9 CF10 3AXUK
- 10
- 11 3. Cultech Ltd, Research Department, Port Talbot, SA12 7BZ, UK
- 12 * Author for Correspondence: Dr Bernard Corfe, Molecular Gastroenterology Research Group,
- 13 Academic Unit of Surgical Oncology, Department of Oncology, University of Sheffield, Beech Hill Road,
- 14 Sheffield, S10 2RX, UK; Tel: +44 (0) 114 271 3004; Email: b.m.corfe@shef.ac.uk
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- 16
- 17 Keywords: fibre, microbiome, appetite, obesity, short-chain fatty acids
- 18

19 Abstract

The recent availability of high-throughput nucleic acid sequencing technologies has rapidly advanced 20 21 approaches to analysing the role of the gut microbiome in governance of human health, including gut 22 health, but also metabolic, cardiovascular, and mental health, inter alia. Recent scientific studies suggest EI perturbations at population level cannot account for the current obesity epidemic, and significant work 23 24 is investigating the potential role of the microbiome, and in particular its metabolic products, notably 25 short-chain fatty acids, predominantly acetate, propionate and butyrate, the last of which is an energy 26 source for the epithelium of the large intestine. The energy yield from dietary residues may be a significant 27 factor influencing energy balance. This review posits that the contribution toward EI is governed by 28 energy intake, diet composition (not just fibre), by the composition of the microbiome, and by levels of 29 physical activity. Furthermore we hypothesize that these factors do not exist in a steady state, but rather 30 are dynamic, with both short- and medium-term effects on appetite regulation. We suggest that existing 31 modelling strategies for bacterial dynamics, specifically for growth in chemostat culture, are of utility in 32 understanding the dynamic interplay of diet, activity and microbiomic oraganisation. Such approaches 33 may be informative in optimising the application of dietary and microbial therapy to promote health.

35 1. Overview

36 The availability of high-throughput nucleic acid sequencing technologies has facilitated a range of new 37 approaches to analysing the role of the gut microbiome in governance of human health (1). Modern techniques suggest a role for the microbiome maintenance of, not only gut health but, systemic 38 39 conditions including cardiovascular health (2), mental health (3) and obesity (3). Despite wide media focus 40 on excess energy intake (EI), recent scientific studies suggest EI perturbations at population level cannot 41 account for the current obesity epidemic (4). The microbiome is responsible for production of a highly complex and highly dynamic metaexometabolome. Well known components of this include the short-42 43 chain fatty acids acetate, propionate and butyrate, the last of which is an energy source for the epithelium 44 of the large intestine (5), as well as an inhibitor of histone deacetylation (and thereby cell fate 45 determination) (6). The energy yield from dietary residues entering the large intestine may account for as 46 much as 10% of EI (7) and is therefore a significant factor influencing energy balance. The guiding 47 theme of this review is that this contribution toward EI is governed by energy intake, diet composition, 48 the composition of the microbiome, and levels of physical activity. Furthermore we hypothesize that 49 these factors do not exist in a steady state, but rather are dynamic, with both short- and medium-term 50 effects on appetite regulation. There is therefore potential to modulate this component of EI through a 51 range of modalities to promote health.

52

53 2. Fibre /Dietary Residue

54 2.1 Scope of definitions of dietary fibre

55 Fibre is a component of diet which is highly complex and inconsistently defined. Approaches to the definition vary from the biochemical, to the physiological, to the functional. The Englyst definition, for 56 57 example, is "non-starch polysaccharides" (8). This is in line with other definitions within nutrition, 58 although it is notable for the element of exclusion which places fibres in the general class of 59 polysaccharides outwith the subclass of starches. Fig 1 provides top-level indication of the potential 60 chemical complexity of this ontology (accessed from ChEBI 08.07.15). However, each endpoint within 61 this ontology masks further factors, including the degree of polymerisation: the nature and extent of 62 polymerisation of side-chains on any polysaccharide backbone. Against this rigid definition is the Association of Official Agricultural Chemists (AOAC)-adopted version by Prosky (9), that fibres are 63 64 "remnants of plant cells resistant to digestion by human digestive enzymes". This definition introduces a 65 physiological component, insofar as resistance to digestion implicates human physiology, but its relevance to non-humans and humans with abnormal digestive capacity is questionable. For example, is "fibre" 66 67 different for animals with different profiles of digestive enzymes? Furthermore, what is the relationship between fibre and personalised medicine? For example, in the case of an inborn error of metabolism 68 69 which may impair intraluminal digestion or absorption - is this definition personal, with each of us 70 potentially having a different profile of fibres? Finally, it introduces a source component, in this case 71 botanical, which raises the question of how fungi fit within this classification. The definition was further extended to include an aspect of functionality in the following Scientific Advisory Committee on 72 73 Nutrition (SACN) statement:

SACN consider that a material can be considered as dietary fibre if it is resistant to digestion and absorption in the small
intestine and has a demonstrable physiological effect potentially associated with health benefits in the body, such as increasing
stool bulk, decreasing intestinal transit time or decreasing post prandial glycaemia. Evidence only of increased fermentation in
the gut should not be included under this definition, since although this has a direct effect on the microflora, it must also be

78 shown to have a demonstrable benefit to the host to be considered as dietary fibre.

80 This extension to the Prosky definition includes and exemplifies health benefits of fibre, yet such 81 advantages are notoriously difficult to demonstrate and attribute. Additionally, it recognises that 82 functionalities may occur beyond the gut, implying indirect mechanisms, although other classes of 83 compound potentially yielding the same intermediate effectors would be excluded from this definition. 84 The SACN statement does not reflect the source (botanical or otherwise) of fibre, but does introduce 85 difficulties of defining fibres in potentially personalised terms.

86 This extended cynicism about mainstream definitions could be coupled to a simple, unifying observation: 87 bacteria cannot read research papers or position statements. The extent of compounds which reach the 88 colon has been demonstrated, inter alia, in studies of differentially diced almond skins, which were found 89 to yield a range of macro- and micro-nutrients (10). It can therefore be argued that the colon environment 90 is not solely nourished by fibres, but by the totality of the ileo-caecal efflueate (ICE) - the material that 91 passes through the ileocaecal valve, whether intact or part-digested, whether of plant, animal or fungal 92 origin, whether polysaccharide or not. For the purposes of a review of interactions between fibres and the 93 microbiome, this definition facilitates the full scope of potential interaction between dietary factors and 94 the microbiome in understanding the production of the exometabolome. Our concept of ICE resembles 95 the definition of fibre proposed by Ha "Any dietary component that reaches the colon without being 96 absorbed in a healthy human gut" (11). The authors critically assimilate the overarching effects of fibre, 97 reproduced in Figure 2 - the division between fermentable and non-fermentable fibres. Fermentable 98 fibres are generally progressively degraded to metabolic endproducts including short-chain fatty acids

99 2,2 The nature of the exometabolome

Major products ensuing from this fermentation are the short chain fatty acids (SCFAs) acetate, butyrate 100 101 and propionate, which can be utilised for lipid or gluconeogenesis (12). SCFAs have been estimated to provide 10% of total dietary energy in humans, and host epithelial cells derive 60–70% of their energy 102 supply from SCFA, particularly butyrate (13). Acetate and propionate are transported across the mucosa 103 and into the hepatic portal and may be detected in the systemic circulation (14) although circulating 104 concentrations of butyrate are disproportionately depleted in the circulation due to mucosal metabolism. 105 106 Other key exometabolites include glucose, vitamins and precursors to neuropeptides. The GI tract has a panel of cell types sensing and responding to these molecules, this interaction is linked to the nervous 107 108 system, and thereby the gut-brain axis (15).

109 3. Microbiome

The human GI tract houses a very complex microbial ecosystem of more than 100 trillion 110 111 microorganisms, ten times greater than the total number of the human cells in the body. Human-112 associated bacteria are dominated by two phyla; Firmicutes and Bacteroidetes, with Proteobacteria, Actinobacteria and Verrucomicrobia present in minor proportions (16, 17), and each phyla containing 113 many different bacterial species (18). The gut microbiota plays an important role in metabolism, immune 114 function, protection of the host from pathogens and bidirectional communication between the GI tract 115 and the central nervous system (19). Dysbiosis, an aberrant state of imbalance of the gut microbiota, has 116 been associated with a diversity of diseases and syndromes such as inflammatory bowel disease, irritable 117 118 bowel syndrome, colorectal cancer, atopy, anxiety, depression, Type II diabetes and metabolic syndrome. The role of the gut microbiota in obesity has been of particular interest, especially given that the global 119 120 prevalence of obesity in both children and adults is rapidly increasing (20), and is a leading cause of 121 preventable disability and death. Obesity results from a sustained net positive energetic balance whereby 122 energy intake exceeds energy output. In addition, host differences in the ability to store and expend

- 123 energy contribute to obesity (21). A new but growing body of evidence suggests the gut microbiota,
- through its role as an interface between nutrients and the host, may assist body weight regulation. The gut
 microbiota can affect nutrient acquisition and energy harvest, as well as producing exometabolites that in
 turn may regulate host metabolic pathways (6, 22).

127 Early indications that the gut microbiota was involved in obesity came when metabolically obese mice, with a mutation in the leptin gene, were shown to have a significantly different microbiota compared to 128 mice without the mutation (23). Further investigation indicated that the ratio of Firmicutes to 129 Bacteroidetes in the gut microbiota of obese mice was shifted in favour of Firmicutes, whilst lean mice 130 were dominated by Bacteroidetes (24). In humans, the gut microbiota composition can respond to 131 132 changes in body weight and is altered in obese compared to non-obese individuals (18). Bacteroidetes 133 may be responsive to calorie intake because their levels increase when body weight is reduced following 134 a reduced calorie diet (25), although numerous human studies have failed to demonstrate a consistent 135 relationship between obesity and the ratio of Firmicutes to Bacteroidetes at both the phylum- and 136 species-level (26).

Hydrogen-producing Prevotellaceae and hydrogen-utilizing methanogenic Archaea were more abundant 137 138 in obese individuals suggesting a higher energy harvest in large intestine to hydrogen transfer between bacterial and archaeal species (27). Changes in the composition of the gut microbiota have been linked 139 140 with (i) suppression of intestinal fasting-induced adipocyte factor (Fiaf), which is a contributing factor to 141 enhanced fat deposition (28), (ii) increased capacity to harvest energy from food and (iii) low-grade inflammation due to activation of toll-like receptors (TLR4), endotoxin and proinflammatory cytokine 142 143 production (29, 30). Approximately 5% of ingested calories are lost in the stool and urine (31). Altered 144 nutrient load over a three-day period induced changes in the gut microbiota in both obese and non-obese 145 individuals, despite statistically significant differences in the composition of the lean and obese microbiome at baseline under a weight maintaining diet (32). In the case of lean subjects, a 20% increase 146 in Firmicutes (and a corresponding decrease in Bacteroidetes) was observed over the three-day period and 147 148 was associated with a ≈ 150 kcal increase in energy absorption.

SCFAs have been implicated in metabolic diseases, including obesity (33). Higher levels of faecal SCFAs, mainly butyrate and propionate, have been reported in obese adults (34) and children (35), compared to lean individuals. Changes in the concentration and proportion of individual SCFA may be in line with changes in the bacterial groups present (12, 35).

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154 3. Appetite control

There are two general definitions of appetite (36). The first relates to food preference, selection and 155 156 intake, and the motivation to eat, whilst the second refers to qualitative and sensory aspects of food, including the impact of environmental stimulation. These eclipse homeostatic theories which suggested 157 feeding corresponds to energy/nutrient deficit or excess (37), yet it is likely that a suite of homeostatic 158 and complex non-homeostatic factors determine the overall expression of appetite. Appetite is normally 159 160 described in terms of hunger, satiation and satiety. Hunger is associated with emptiness of the stomach, irritability and light-headedness (36). Humans can and do, however, display hunger for other reasons: the 161 162 smell, sight or even thought of food can initiate feeding (38). Eating triggers a cascade of metabolic signals that can suppress hunger and inhibit further consumption (39). Satiation is the point of 163 164 satisfaction that results in meal termination (38, 40, 41). Satiety is the (modifiable) post-ingestion period 165 of repletion which influences the time of the next eating occasion (42).

Appetite is controlled by multiple integrated physiological signals (See Figure 3). Short-term signals help 166 regulate meal initiation and termination whereas long-term, humoral signals play a central role in body 167 weight regulation (43). This conceptual framework for examining the impact of feeding is continually 168 updated to represent an increasing number of factors encompassing peripheral physiological and 169 170 metabolic events, and brain responses that play important roles in appetite control (44). The GI tract 171 responds to feeding in three integrated phases: cephalic, post-ingestive and post-absorptive, all of which 172 depend on parasympathetic nerve transmission. The cephalic phase occurs at the point of food selection and early ingestion, and is thus stimulated by conditioned processes and organoleptic factors (45, 46). It 173 174 is held that post-ingestive satiation signals arise largely from mechanical distention, while signals from the 175 GI tract derive predominantly from the chemical effects of food (47). In contrast, post-absorptive effects 176 are the result of interplay between hormones and the hypothalamic region of the brain that respond to 177 fluctuating concentrations of nutrients in the portal vein, plasma and brain.

178 3,1 Impact of the exometabolome on post-ingestive appetite regulation

179 Landmark human studies have shown intestinal nutrient infusions can reduce food intake with rapid 180 effects (48-50), indicating that satiation signals must originate from the gut as well as post-absorptively. 181 Numerous hormones, neurotransmitters and peptides stimulate orexigenic or anorexigenic responses. Many peptide hormones are produced in the gastrointestinal tract and released in response to nutritional 182 183 stimuli. Anorexigenic hormones include CCK, glucagon-like peptide-1 and -2 (GLP-1 and GLP-2), 184 glucose-dependent insulinotropic polypeptide (GIP), oxyntomodulin, PP, peptide histidine isoleucine, peptide histidine valine, peptide YY and somatostatin)(51, 52). Enteroendocrine (EE) cells represent less 185 186 than one percent of the mucosal cell population, yet form the largest endocrine system in the human (53), 187 and is populated by singly distributed enteroendocrine cells which release a very significant portion of appetite regulating hormones (54). (TABLE 1). EE cells have a characteristic flask-shaped morphology 188 and have been divided into at least sixteen cellular subtypes based on the major products they produce 189 190 and secrete (55), although this model is contested and a continuum of cell types has also been proposed 191 (56).

192 The primary EE cell types in the colon are D cells, L cells and EnteroChromaffin (EC) cells (57). Whilst 193 all cell types may be found along the colon, EC are the most abundant, and D cells the least, with a progressive increase in the proportion of L-cells along the caeco-rectal axis. As summarised in our review, 194 these cells harbour peptide/hormones involved in appetitive regulation including PYY, GLP-1, GLP-2 195 196 and oxyntomodulin. Intriguingly the EC subclass also contain 5HT (serotonin) and reports suggest that as much as 95% of the body's 5HT may exist in the gut (58) Serotonin has been implicated in appetitive 197 198 regulation, mood control and regulation of gut transit. This underwrites plausible links between luminal 199 content, motivation to eat and wider aspects of regulation of colorectal content through modulation of 200 transit time. These factors are explored in greater detail below.

SCFAs are important signalling components within the gut-brain axis, the system of communication between the gut and the brain (19, 59) which interacts directly with gut endocrine cells, and stimulates secretion of peptide YY (PYY) by activating two G-protein-coupled receptors (GPR41 and GPR43). EE carry free fatty acid receptors (FFARs) on their surface which have differential affinity for SCFAs and which signal the release of appetitive hormones from EEC (60). As components of the exometabolome, SCFAs therefore act as key molecules governing the sensing-signalling pathway linking luminal metabolism to appetite regulation.

208 Our group have recently identified a further plausible mechanism of action. A significant body of 209 literature suggests butyrate is a potent regulator of numbers of proliferating cells in the colon crypt. We 210 recently demonstrated an inverse association between SCFA and the numbers of EEC cells in the crypt

- 211 (61). Mathematical modelling suggests SCFA may modulate differentiation pathways on exit from the
- stem cell compartment (62). Taken together these data suggest two possible tiers of regulation of post-
- **213** ingestive appetite by the exometabolome: (1) an acute response in terms of regulating release of anorectic
- hormones; and (2) an adaptive modulation of numbers of EEC and thereby available pools of appetite-
- **215** regulatory hormones.

216 3,2 Impact of the exometabolome on post-absorbtive appetite regulation

Post-absorptive signals are stimulated by the entry of nutrients into the portal vein of the liver, or by fluctuating nutrient concentrations in the plasma and brain (63). These signals act (via the hypothalamic region of the brain and vagus nerve) on the periphery and central nervous system and also interact with long-acting adiposity hormones (such as leptin) that help regulate body weight *ibid*. Two key areas are impacted by the exometabolome: via intestinal gluconeogenesis and through pan-systemic propionate sensing.

- 223 Gluconeogenesis has until relatively recently been viewed as a primarily hepatic and renal phenomenon, 224 and is not positively associated with health, reflecting excess energy intake. Relatively recently the 225 intestine has been identified as a site gluconeogenesis (distinguished as Intestinal GlucoNeogenesis -226 IGN) (64). IGN is regulated by both butyrate and propionate. Butyrate acts to govern the levels of IGN enzymes in the mucosa. In contrast propionate is both a substrate for IGN and is a regulator of IGN 227 228 enzyme activity mediated via FFAR3 signalling (Fig 4) (65). This paper therefore also suggests emergent 229 distinctions between the fates and activities of SCFA. Intestinally-produced glucose is transported to the 230 HPV where it is directly sensed by sodium-coupled glucose co-transporter (66). Critically, in contrast to 231 hepatic and renal gluconeogenesis, IGN associated with positive health outcomes (65).
- Post-ingestive appetite regulation may also occur at the level of FFAR3 signalling. There is growing
 recognition that FFAR family receptors, including FFAR3 are expressed on a wide range of tissues
 including adipose, liver. The role of FFAR3 in non-gut tissue is reviewed elsewhere in this issue (67).

235 3.3 Impact of the exometabolome on cephalic phase of appetite regulation

The impact of exometabolites upon cephalic phase of appetite has not been well explored however it is 236 237 reasonable to hypothesize that it does contribute to the wider mechanisms of appetite control as 238 precedents have been shown in microbiome-mood interactions. For example: perturbations of the gut flora have been associated with schizophrenia and depression (68, 69); probiotic interventions in mouse 239 240 models have demonstrated anxiolytic potential of microbial intervention (70); probiotic interventions 241 have also shown impact upon brain activity (71) and on cognitive outcome (72). Recent reviews have 242 suggested potential mechanisms of action, including modulation of afferent signalling by SCFA, cytokine-243 mediated responses triggered through TLRs in the mucosa responding to the microbiome, and 244 modulation of GABA-mediated signalling (15). As a strong evidence-base is emerging for a role of the microbiome and exometabolome in governance of mood and cognition, it seems likely that this will in 245 246 time extend through to cephalic phase appetite control.

248 4. Modification of the microbiome by alteration of transit (the chemostat analogy)

249 Although obesity and obesity-related disorders have been linked with alterations in the gut microbiota,

less attention has been directed towards investigating lifestyle aspects of obesity, such as exercise and diet,and their effect on the microbial and physical environment of the gastrointestinal tract (73). In a recent

study, elite athletes had a significantly more diverse gut microbiota compared to non-athletic size matched

²⁴⁷

- (high body mass index (BMI) \approx 30) and age/gender matched (BMI <25) control groups (74). As the elite athlete group also consumed a significantly different diet, which provided more calories per day from carbohydrates, proteins and fat compared to the control groups, this study suggested that both diet and exercise were driving factors in changing gut microbial diversity. Exercise has also been shown to decrease transit time, particularly through the descending colon (74, 75). Previous reports have suggested however, that physical activity does not necessarily improve overall gastrointestinal transit (76).
- 259 It may be convenient therefore to view the colon as a chemostat, a commonly used form of bioreactor which has been applied in microbiological settings for the determination of growth parameters. (Fig 5). In 260 this simple model the ecosystem is fed at a specific rate (the dilution rate) which is also reflected in the 261 rate of effluent production. The population within this system will have a growth rate (μ) proportional to 262 263 the dilution rate (D). At a certain dilution rate μ_{max} is reached – the maximal growth rate for a particular 264 species (in the context of an ecosystem this will be for a specific species as each will have a unique μ_{max}), 265 at this point the species will start to dilute from the system. The dilution rate therefore represents an extremely strong selective pressure upon the microbiome. As discussed in sections above, fibre intake as 266 267 well as physical activity levels will influence transit time, which is analogous to the dilution rate in a chemostat. Data suggest that individuals on high-fibre diets lose more energy in faecal material than those 268 on lower-fibre diets with an equivalent energy content (77), supporting a model whereby reduced energy 269 270 harvest associates with a factor affecting transit.

We therefore argue that a contributing longtundinal effect of high fibre intakes, or high physical activity, or the combination thereof is the modification of the microbiome by exerting a specific selective pressure. Contrastingly, excessive slow values for dilution rate, D, will provide opportunities for these microbial products to interact with the host epithelium, potentially increasing host energy harvest in the case of SCFAs, and elevating exposure to pro-inflammatory signalling and cytotoxic molecules.

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277 6. Summary and future directions

The question of whether alterations in gut microbiota are a cause or a consequence of obesity still 278 remains unclear, although evidence from observational and intervention studies in humans appears to 279 suggest that both the microbiota and diet play a significant role in body weight regulation, beginning at 280 birth. Although the utility of animal models for conducting more controlled experiments investigating the 281 differences between the obese and lean microbiota has been established, translation to research in 282 283 humans has proved less fruitful in providing a clear consensus concerning the role played by the balance 284 between the most abundant bacterial phyla in the human gut. Indeed, the emerging evidence indicates 285 that even the effect of individual bacterial species cannot be disregarded from study. This means that moving towards the use of high-resolution, standardised analytical techniques for surveying the gut 286 microbiota, combined with well-designed human studies taking all of the confounding variables (e.g. age, 287 sex, ethnicity, diet and genetic factors) into account, may allow us to identify a specific consortium of 288 289 microbes that contribute to obesity, elucidate their modes of action via host and diet interactions, and evaluate novel strategies to regulate energy balance in obese individuals. Such strategies may for example 290 include approaches to modify (or restore "normality" to) the mircobiota in order to restore energy 291 292 balance. Changes in gut microbiota composition have been observed after consumption of a calorie 293 restricted diet in overweight and obese subjects (26). Inconclusive evidence exists on the effect of supplementation with lactobacilli and bifidobacteria, alone or in combination with prebiotics, on weight 294 295 management in humans (78-80). As such, intervention strategies are an attractive approach to appetite 296 management through restoration of ecological balance in the gut.

297 7. Key conclusions and areas for future research

- Fibres are inconsistently defined and an oversight of the totality of nutrients entering the large
 bowel may be more informative
- Perturbations in the microbiome associate with obesity and increased energy harvest. The
 relationship between the diet and microbiome and host health is mediated considerably by the
 exometabolome.
- Most studies to date are associative and greater emphasis needs to be placed on longitudinal or
 prospective trials
- The relationship between the exometabolome and the host is dynamic and multifactorial, reductionist approaches are unlikely to yield an insight into health benefits.

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Tables and Figure Legends

Table 1: The secretory products of enteroendocrine cells of the colon and rectum and theiractions

311	actions		
	Peptide	Actions	
	5-HT	Intestinal motility; intestinal secretion; visceral sensation; appetite reduction	
	Glicentin	Stimulates mucosal enterocyte proliferation; inhibits gastric emptying	
	GLP-1	Incretin effect; delays gastric emptying; postprandial satiety, inhibits energy intake	
	GLP-2	Stimulates mucosal enterocyte proliferation, enhances digestive and absorptive capacities of intestine, inhibits gastric secretion	
	Oxyntomodulin	Inhibits gastric emptying, reduces gastric motility, inhibits food intake	
	РҮҮ	Inhibits gastric emptying and intestinal motility; inhibits gastric acid secretion and pancreatic exocrine function; suppresses appetite; stimulates mucosal enterocyte proliferation	
	Somatostatin	Major inhibitory hormone for digestive endocrine and exocrine function; stimulates colonic peristalsis; potential for reducing food intake	
312 313 314 315	PYY, peptide YY; (GLP-1, glucagon-like peptide 1; GLP-2, glucagon-like peptide 2.	
	Table taken from Gunarwardene Corfe & Staton CA (2011) with additional information from (81-83)		
316			
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318			

319	Figure 1 A chemical Ontology for "Fibre"
320	Accessed from ChEBI (www.chebi.ebi.ac.uk), 08.07.14
321	
322	Figure 2 An alternative definition of "fibre"
323 324 325	Based on Ha et al (2000) this definition encompasses all material able to enter the colon (ICE – Ileo Caecal Effulent), as available for microbial metabolism. Some components are readily metabolised, some highly resistant to metabolism.
326	
327	Fig 3 Tiers of appetite regulation by short-chain fatty acids
328	
329	Figure 4 Intestinal Gluconeogenesis Pathway
330	
331	Figure 5 Analogy between the Chemostat and the Colon
332	Chemostat image: chemistry.about.com, colon image www.clker.com
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340 2006;13(25):3005-21. 341 3. Grenham S, Clarke G, Cryan JF, Dinan TG. Brain-gut-microbe communication in health and 342 disease. Frontiers in physiology. 2011 2011;2:94. 343 England PH. National Diet and Nutrition Survey: Results from Years 1 - 4 (combined) of the 4. 344 Rolling Programme (2008/2009 – 2011/12). 2014. 345 Roediger WEW. UTILIZATION OF NUTRIENTS BY ISOLATED EPITHELIAL-CELLS OF THE RAT 5. 346 COLON. Gastroenterology. [Article]. 1982;83(2):424-9. 347 Corfe BM. Hypothesis: butyrate is not an HDAC inhibitor, but a product inhibitor of 6. 348 deacetylation. Mol Biosyst. [10.1039/c2mb25028d]. 2012;8(6):1609-12. 349 McNeil NI. THE CONTRIBUTION OF THE LARGE-INTESTINE TO ENERGY SUPPLIES IN MAN. 7. 350 American Journal of Clinical Nutrition. [Article]. 1984;39(2):338-42. Englyst HN, Quigley ME, Hudson GJ, Cummings JH. DETERMINATION OF DIETARY FIBER AS 351 8. 352 NONSTARCH POLYSACCHARIDES BY GAS-LIQUID-CHROMATOGRAPHY. Analyst. 1992;117(11):1707-353 14. 354 9. Prosky L. What is dietary fiber? Journal of Aoac International. 2000;83(4):985-7. 355 10. Mandalari G, Faulks RM, Rich GT, Lo Turco V, Picout DR, Lo Curto RB, et al. Release of 356 protein, lipid, and vitamin E from almond seeds during digestion. J Agric Food Chem. [Article]. 2008 May;56(9):3409-16. 357 358 11. Ha MA, Jarvis MC, Mann JI. A definition for dietary fibre. European Journal of Clinical 359 Nutrition. [Review]. 2000 Dec;54(12):861-4. 360 Schwiertz A, Taras D, Schafer K, Beijer S, Bos NA, Donus C, et al. Microbiota and SCFA in Lean 12. 361 and Overweight Healthy Subjects. Obesity. [Article]. 2010 Jan;18(1):190-5. 362 Scheppach W, Sommer H, Kirchner T, Paganelli GM, Bartram P, Christl S, et al. EFFECT OF 13. 363 BUTYRATE ENEMAS ON THE COLONIC MUCOSA IN DISTAL ULCERATIVE-COLITIS. Gastroenterology. 364 1992;103(1):51-6. 365 14. Knudsen KEB, Serena A, Canibe N, Juntunen KS. New insight into butyrate metabolism. Proceedings of the Nutrition Society. [Article; Proceedings Paper]. 2003 Feb;62(1):81-6. 366 367 15. Forsythe P, Sudo N, Dinan T, Taylor VH, Bienenstock J. Mood and gut feelings. Brain Behav 368 Immun. [Review]. 2010 Jan;24(1):9-16. 369 16. Costello EK, Lauber CL, Hamady M, Fierer N, Gordon JI, Knight R. Bacterial Community 370 Variation in Human Body Habitats Across Space and Time. Science. [Article]. 2009 371 Dec;326(5960):1694-7. 372 Pflughoeft KJ, Versalovic J. Human Microbiome in Health and Disease. Annu Rev Pathol-17. 373 Mech Dis. [Review; Book Chapter]. 2012;7:99-122. 374 Tremaroli V, Backhed F. Functional interactions between the gut microbiota and host 18. 375 metabolism. Nature. [Review]. 2012 Sep;489(7415):242-9. 376 19. Chen VL, Surana NK, Duan JY, Kasper DL. Role of Murine Intestinal Interleukin-1 Receptor 1-377 Expressing Lymphoid Tissue Inducer-Like Cells in Salmonella Infection. Plos One. [Article]. 2013 378 Jun;8(6). 379 Nguyen DM, El-Serag HB. The Epidemiology of Obesity. Gastroenterology Clinics of North 20. 380 America. 2010 Mar;39(1):1-+. Clement K, Ferre P. Genetics and the pathophysiology of obesity. Pediatr Res. [Review]. 2003 381 21. 382 May;53(5):721-5.

Huttenhower C, Gevers D, Knight R, Abubucker S, Badger JH, Chinwalla AT, et al. Structure,

function and diversity of the healthy human microbiome. Nature. [Article]. 2012 Jun;486(7402):207-

Fava F, Lovegrove JA, Gitau R, Jackson KG, Tuohy KM. The gut microbiota and lipid

metabolism: Implications for human health and coronary heart disease. Curr Med Chem. [Review].

22. Donohoe DR, Collins LB, Wali A, Bigler R, Sun W, Bultman SJ. The Warburg Effect Dictates the
Mechanism of Butyrate-Mediated Histone Acetylation and Cell Proliferation. Molecular Cell. 2012
Nov 30;48(4):612-26.

Ley RE, Backhed F, Turnbaugh P, Lozupone CA, Knight RD, Gordon JI. Obesity alters gut
microbial ecology. Proceedings of the National Academy of Sciences of the United States of America.
2005 Aug 2;102(31):11070-5.

Turnbaugh PJ, Ley RE, Mahowald MA, Magrini V, Mardis ER, Gordon JI. An obesity-associated
 gut microbiome with increased capacity for energy harvest. Nature. [Article]. 2006
 Des: 444(7122):1027-21

391 Dec;444(7122):1027-31.

25. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology - Human gut microbes associated
with obesity. Nature. 2006 Dec 21;444(7122):1022-3.

39426.Tagliabue A, Elli M. The role of gut microbiota in human obesity: Recent findings and future395perspectives. Nutrition Metabolism and Cardiovascular Diseases. 2013 Mar;23(3):160-8.

Zhang H, DiBaise JK, Zuccolo A, Kudrna D, Braidotti M, Yu Y, et al. Human gut microbiota in
obesity and after gastric bypass. Proceedings of the National Academy of Sciences of the United
States of America. 2009 Feb 17;106(7):2365-70.

Backhed F, Manchester JK, Semenkovich CF, Gordon JI. Mechanisms underlying the
resistance to diet-induced obesity in germ-free mice. Proceedings of the National Academy of
Sciences of the United States of America. 2007 Jan 16;104(3):979-84.

402 29. Cani PD, Bibiloni R, Knauf C, Neyrinck AM, Delzenne NM, Burcelin R. Changes in gut
403 microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity
404 and diabetes in mice. Diabetes. 2008 Jun;57(6):1470-81.

30. Zhao LP. The gut microbiota and obesity: from correlation to causality. Nat Rev Microbiol.
406 [Article]. 2013 Sep;11(9):639-47.

407 31. Heymsfield SB, Pietrobelli A. Individual differences in apparent energy digestibility are larger
408 than generally recognized. American Journal of Clinical Nutrition. 2011 Dec;94(6):1650-1.

32. Jumpertz R, Duc Son L, Turnbaugh PJ, Trinidad C, Bogardus C, Gordon JI, et al. Energybalance studies reveal associations between gut microbes, caloric load, and nutrient absorption in
humans. American Journal of Clinical Nutrition. 2011 Jul;94(1):58-65.

33. Nicholson JK, Holmes E, Kinross J, Burcelin R, Gibson G, Jia W, et al. Host-Gut Microbiota
Metabolic Interactions. Science. 2012 Jun 8;336(6086):1262-7.

41434.Schwiertz A, Taras D, Schaefer K, Beijer S, Bos NA, Donus C, et al. Microbiota and SCFA in415Lean and Overweight Healthy Subjects. Obesity. 2010 Jan;18(1):190-5.

416 35. Payne AN, Chassard C, Zimmermann M, Mueller P, Stinca S, Lacroix C. The metabolic activity
417 of gut microbiota in obese children is increased compared with normal-weight children and exhibits
418 more exhaustive substrate utilization. Nutrition & Diabetes. 2011 Jul;1.

419 36. Blundell J. Making claims: functional foods for managing appetite and weight. Nature
420 Reviews Endocrinology. 2010 Jan;6(1):53-6.

421 37. Mayer EA, Naliboff BD, Craig ADB. Neuroimaging of the brain-gut axis: From basic
422 understanding to treatment of functional G1 disorders. Gastroenterology. 2006 Dec;131(6):1925-42.

422 understanding to treatment of functional G1 disorders. Gastroenterology. 2006 Dec;131(6):1925-4.
 423 38. Yeomans MR, Gray RW. Effects of naltrexone on food intake and changes in subjective

appetite during eating: Evidence for opioid involvement in the appetizer effect. Physiology &
 Behavior. 1997 Jul;62(1):15-21.

426 39. Reid M, Hetherington M. Relative effects of carbohydrates and protein on satiety - A review
427 of methodology. Neuroscience and Biobehavioral Reviews. 1997 May;21(3):295-308.

428 40. Weenen H, Stafleu A, de Graaf C. Dynamic aspects of liking: post-prandial persistence of 429 sensory specific satiety. Food Quality and Preference. 2005 Sep;16(6):528-35.

430 41. Sclafani A. Gut-brain nutrient signaling. Appetition vs. satiation. Appetite. 2013 Dec431 1;71:454-8.

432 42. Williams RA, Roe LS, Rolls BJ. Comparison of three methods to reduce energy density. Effects433 on daily energy intake. Appetite. 2013 Jul 1;66:75-83.

439 normal subjects. Appetite. 2000 Aug;35(1):45-55. 440 Powley TL. Vagal circuitry mediating cephalic-phase responses to food. Appetite. 2000 46. 441 Apr;34(2):184-8. 442 47. Powley TL, Phillips RJ. Gastric satiation is volumetric, intestinal satiation is nutritive. 443 Physiology & Behavior. 2004 Aug;82(1):69-74. 444 48. French SJ, Conlon CA, Mutuma ST, Arnold M, Read NW, Meijer G, et al. The effects of 445 intestinal infusion of long-chain fatty acids on food intake in humans. Gastroenterology. 2000 446 Oct;119(4):943-8. 447 49. Feltrin KL, Little TJ, Meyer JH, Horowitz M, Smout A, Wishart J, et al. Effects of intraduodenal 448 fatty acids on appetite, antropyloroduodenal motility and plasma CCK and GLP-1 in humans are 449 dependent on their chain length. Gastroenterology. 2004 Apr;126(4):A524-A. 450 50. Maljaars PWJ, Symersky T, Kee BC, Haddeman E, Peters HPF, Masclee AAM. Effect of ileal fat 451 perfusion on satiety and hormone release in healthy volunteers. International Journal of Obesity. 452 2008 Nov;32(11):1633-9. 453 51. Hussain SS, Bloom SR. The regulation of food intake by the gut-brain axis: implications for 454 obesity. International Journal of Obesity. 2013 May;37(5):625-33. 455 52. Perry B, Wang Y. Appetite regulation and weight control: the role of gut hormones. Nutrition 456 & Diabetes. 2012 Jan;2. 457 53. Rehfeld JF. Beginnings: A reflection on the history of gastrointestinal endocrinology. 458 Regulatory Peptides. 2012 Aug 10;177:S1-S5. 459 Cuomo R, D'Alessandro A, Andreozzi P, Vozzella L, Sarnelli G. Gastrointestinal regulation of 54. 460 food intake: do gut motility, enteric nerves and entero-hormones play together? Minerva 461 Endocrinologica. 2011 Dec;36(4):281-93. 55. 462 Choi S, Lee M, Shiu AL, Yo SJ, Hallden G, Aponte GW. GPR93 activation by protein 463 hydrolysate induces CCK transcription and secretion in STC-1 cells. American Journal of Physiology-464 Gastrointestinal and Liver Physiology. 2007 May;292(5):G1366-G75. 465 56. Gribble FM. The gut endocrine system as a coordinator of postprandial nutrient 466 homoeostasis. Proceedings of the Nutrition Society. [Article; Proceedings Paper]. 2012 467 Nov;71(4):456-62. 468 57. Gunawardene AR, Corfe BM, Staton CA. Classification and functions of enteroendocrine cells 469 of the lower gastrointestinal tract. International Journal of Experimental Pathology. 2011 470 Aug;92(4):219-31. 471 Spiller R. Serotonergic agents and the irritable bowel syndrome: what goes wrong? Current 58. 472 Opinion in Pharmacology. 2008 Dec;8(6):709-14. 473 59. Cryan JF, Dinan TG. Mind-altering microorganisms: the impact of the gut microbiota on brain 474 and behaviour. Nature Reviews Neuroscience. 2012 Oct;13(10):701-12. 475 60. Darzi J, Frost GS, Robertson MD. Postgraduate Symposium Do SCFA have a role in appetite 476 regulation? Proceedings of the Nutrition Society. 2011 Feb;70(1):119-28. 477 61. Yu DC, Bury JP, Tiernan J, Waby JS, Staton CA, Corfe BM. Short-chain fatty acid level and field 478 cancerization show opposing associations with enteroendocrine cell number and neuropilin 479 expression in patients with colorectal adenoma. Mol Cancer. [10.1186/1476-4598-10-27]. 480 2011;10:27. 481 Smallbone K, Corfe BM. A mathematical model of the colon crypt capturing compositional 62. 482 dynamic interactions between cell types. International Journal of Experimental Pathology. 2014 483 Feb;95(1):1-7.

Blundell JE, Lawton CL, Cotton JR, Macdiarmid JI. Control of human appetite: Implications for

Nederkoorn C, Smulders FTY, Jansen A. Cephalic phase responses, craving and food intake in

Dalton M, Finlayson G. Hedonics, satiation and satiety. In: Blundell JE, Bellisle F, editors.

the intake of dietary fat. Annual Review of Nutrition. 1996 1996;16:285-319.

Satiation, Satiety and the Control of Food Intake: Theory and Practice2013. p. 221-37.

434

435

436

437

438

43.

44.

45.

484 63. Cummings DE, Overduin J. Gastrointestinal regulation of food intake. Journal of Clinical
485 Investigation. 2007 Jan;117(1):13-23.

486 64. Croset M, Rajas F, Zitoun C, Hurot JM, Montano S, Mithieux G. Rat small intestine is an
487 insulin-sensitive gluconeogenic organ. Diabetes. 2001 Apr;50(4):740-6.

488 65. De Vadder F, Kovatcheva-Datchary P, Goncalves D, Vinera J, Zitoun C, Duchampt A, et al.
489 Microbiota-Generated Metabolites Promote Metabolic Benefits via Gut-Brain Neural Circuits. Cell.
490 2014 Jan 16;156(1-2):84-96.

66. Delaere F, Duchampt A, Mounien L, Seyer P, Duraffourd C, Zitoun C, et al. The role of
sodium-coupled glucose co-transporter 3 in the satiety effect of portal glucose sensing. Molecular
metabolism. 2012 2012;2(1):47-53.

494 67. Chambers E. S MDJ, Frost G. . Proceedings of the Nutrition Society in press.

495 68. Drexhage RC, Weigelt K, van Beveren N, Cohen D, Versnell MA, Nolen WA, et al. IMMUNE
496 AND NEUROIMMUNE ALTERATIONS IN MOOD DISORDERS AND SCHIZOPHRENIA. In: Guest PC, Bahn
497 S, editors. Biomarkers of Neurological and Psychiatric Disease2011. p. 169-201.

498 69. Foster JA, Neufeld K-AM. Gut-brain: how the microbiome influences anxiety and depression.
499 Trends in Neurosciences. 2013 May;36(5):305-12.

500 70. Messaoudi M, Lalonde R, Violle N, Javelot H, Desor D, Nejdi A, et al. Assessment of 501 psychotropic-like properties of a probiotic formulation (Lactobacillus helveticus R0052 and

Bifidobacterium longum R0175) in rats and human subjects. British Journal of Nutrition. 2011
 Mar;105(5):755-64.

Tillisch K, Labus J, Kilpatrick L, Jiang Z, Stains J, Ebrat B, et al. Consumption of Fermented Milk
Product With Probiotic Modulates Brain Activity. Gastroenterology. 2013 Jun;144(7):1394-U136.

506 72. Owen LJ, Reinders, M.J., Narramore, R., Marsh, A.M.R., Gar Lui, F., Baron, R., Plummer,
507 S.F., Garaiova, I. & Corfe, B.M. . A double blind, placebo controlled, randomised pilot study
508 examining the effects of probiotic administration on mood and cognitive function in healthy young
509 adults. . European Journal of Nutrition.submitted.

51073.Hold GL. Western lifestyle: a 'master' manipulator of the intestinal microbiota? Gut. 2014511Jan;63(1):5-6.

512 74. Clarke SF, Murphy EF, O'Sullivan O, Lucey AJ, Humphreys M, Hogan A, et al. Exercise and
513 associated dietary extremes impact on gut microbial diversity. Gut. 2014.

514 75. Song BK, Cho KO, Jo Y, Oh JW, Kim YS. Colon Transit Time According to Physical Activity Level 515 in Adults. Journal of Neurogastroenterology and Motility. 2012 Jan;18(1):64-9.

76. Robertson G, Meshkinpour H, Vandenberg K, James N, Cohen A, Wilson A. EFFECTS OF
EXERCISE ON TOTAL AND SEGMENTAL COLON TRANSIT. Journal of Clinical Gastroenterology. 1993
Jun;16(4):300-3.

519 77. Beyer PL, Flynn MA. EFFECTS OF HIGH-FIBER AND LOW-FIBER DIETS ON HUMAN FECES.
520 Journal of the American Dietetic Association. 1978 1978;72(3):271-7.

52178.Kadooka Y, Sato M, Imaizumi K, Ogawa A, Ikuyama K, Akai Y, et al. Regulation of abdominal522adiposity by probiotics (Lactobacillus gasseri SBT2055) in adults with obese tendencies in a523adiposity by probiotics (Lactobacillus gasseri SBT2055) in adults with obese tendencies in a

randomized controlled trial. European Journal of Clinical Nutrition. 2010 Jun;64(6):636-43.

524 79. Gobel RJ, Larsen N, Jakobsen M, Molgaard C, Michaelsen KF. Probiotics to Adolescents With
525 Obesity: Effects on Inflammation and Metabolic Syndrome. Journal of Pediatric Gastroenterology
526 and Nutrition. 2012 Dec;55(6):673-8.

52780.Sanz Y, Rastmanesh R, Agostonic C. Understanding the role of gut microbes and probiotics in528obesity: How far are we? Pharmacological Research. 2013 Mar;69(1):144-55.

52981.Dakin CL, Gunn I, Small CJ, Edwards CMB, Hay DL, Smith DM, et al. Oxyntomodulin inhibits530food intake in the rat. Endocrinology. 2001 Oct;142(10):4244-50.

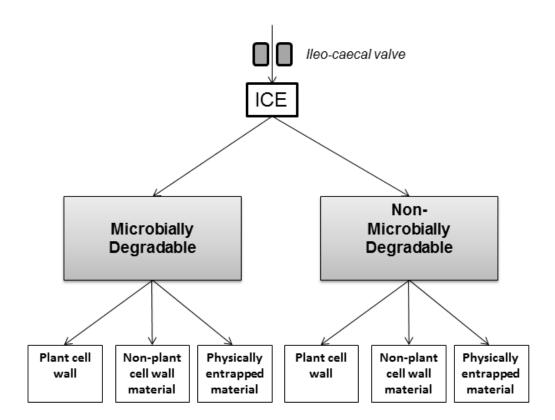
531 82. Dakin CL, Small CJ, Park AJ, Seth A, Ghatei MA, Bloom SR. Repeated ICV administration of

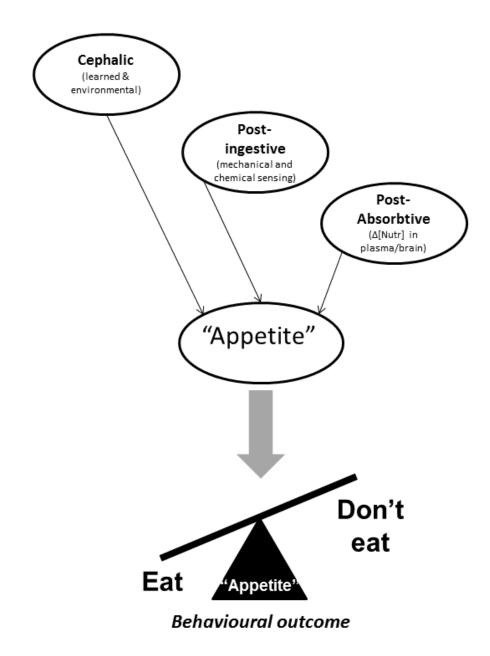
532 oxyntomodulin causes a greater reduction in body weight gain than in pair-fed rats. American

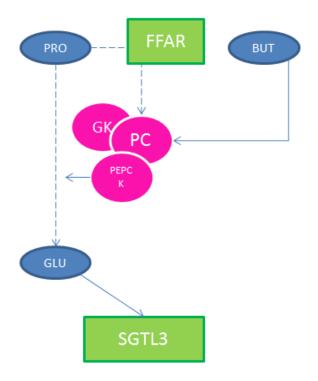
Journal of Physiology-Endocrinology and Metabolism. 2002 Dec;283(6):E1173-E7.

- 83. Verdich C, Flint A, Gutzwiller JP, Naslund E, Beglinger C, Hellstrom PM, et al. A meta-analysis
- of the effect of glucagon-like peptide-1 (7-36) amide on ad libitum energy intake in humans. Journal
- of Clinical Endocrinology & Metabolism. 2001 Sep;86(9):4382-9.

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p-block molecular entity organochalcogen compound
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organic molecular entity
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[3]-alpha-L-Araf (1->3)-6d-alpha-L-gulo-Hepp-(1->)n
heterogiycan alpha-D-giucos/4-1-9-2/12-96-beta-D-fructos/fin
[3]-alpha-L-Rhap-(1→2)-alpha-L-Rhap-(1→]n
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(3)-aipha-D-Fuop-(1→2)-beta-U-Rhap-(1→jn
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(4)-alpha-D-Galp-(1-x3)-beta-D-Galf-(1-x]n S. Pullorum strain 77 O-specific polysaccharide
[2]-beta-D-Manp-(1≫4)-alpha-L-Rhap-(1≫3)-alpha-D-Galp-(1⇒)n
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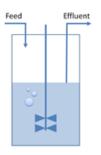






Chemostat

Continuous culture bioreactor



$$D = \frac{\text{Medium flow rate}}{\text{Culture volume}} = \frac{\text{F}}{\text{V}}$$

- Any given bacterial species will grow at a rate µ which is a function of D and the nature of the nutrient
- When µ_{max} is reached the species can no longer compete with D and will be progressively diluted from the system



Also a continuous culture bioreactor



- Increase faecal bulk
- F is a function of rate of ICE
- F is additionally a function of rate of absorption
- V is variable, but any given individual will have a Vmin and Vmax
- Insol Fibre will affect F and V and so have an effect on D
- Consequent selective pressure and impact upon the composition of the microbiome