

promoting access to White Rose research papers



Universities of Leeds, Sheffield and York
<http://eprints.whiterose.ac.uk/>

This is a copy of the final published version of a paper published via gold open access in **Chemical Communications**

This open access article is distributed under the terms of the Creative Commons Attribution Licence (<http://creativecommons.org/licenses/by/3.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

White Rose Research Online URL for this paper:
<http://eprints.whiterose.ac.uk/88194>

Published paper

Lilley, M., Mambwe, B., Jackson, R.F.W. and Muimo, R. (2014) *4-Phosphothiophen-2-yl alanine: a new 5-membered analogue of phosphotyrosine*. *Chemical Communications*, 50 (66). 9343 - 9345

10.1039/c4cc03393k

4-Phosphothiophen-2-yl alanine: a new 5-membered analogue of phosphotyrosine†

Cite this: DOI: 10.1039/c4cc03393k

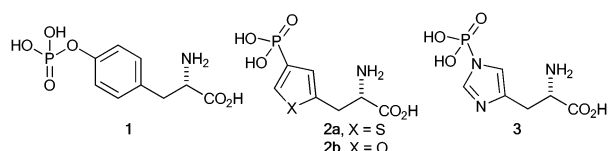
Received 6th May 2014,
Accepted 25th June 2014

DOI: 10.1039/c4cc03393k

www.rsc.org/chemcomm

Polyclonal antibodies raised against 4-phosphothiophen-2-yl alanine 2a, a novel five-membered ring analogue of phosphotyrosine, showed high selectivity for phosphotyrosine and no cross-reactivity with other phosphorylated amino acids. Western blots showed that the polyclonal was similarly effective, but different in selectivity, to a commercially available monoclonal antibody.

Protein tyrosine phosphatases (PTPs) catalyse the dephosphorylation of phosphotyrosine residues and together with protein tyrosine kinases (PTKs) regulate the tyrosine phosphoproteome in cells.¹ Several PTPs have been identified as attractive therapeutic targets in human diseases such as cancer, diabetes, obesity, neurodegenerative disorders and inflammation.^{2–4}



Since phosphotyrosine **1** is unstable under typical biological conditions, interest in the biological roles of PTPs concomitantly led to the development of stable analogues of phosphotyrosine.^{4,5} These analogues, azobenzene phosphonate (ABP),⁶ phosphotyramine,⁷ phosphonomethylphenylalanine (Pmp),⁸ and difluorophosphono-methylphenylalanine (F2Pmp)^{9,10} have been invaluable tools in unraveling the importance of phosphotyrosine in cellular processes. These benzenoid analogues have been useful not only as immunogens to generate antibodies that recognise phosphotyrosine with high specificity without cross-reactivity to other phosphoamino acids, but also as inhibitors and affinity ligands for PTPs and other pTyr binding proteins.

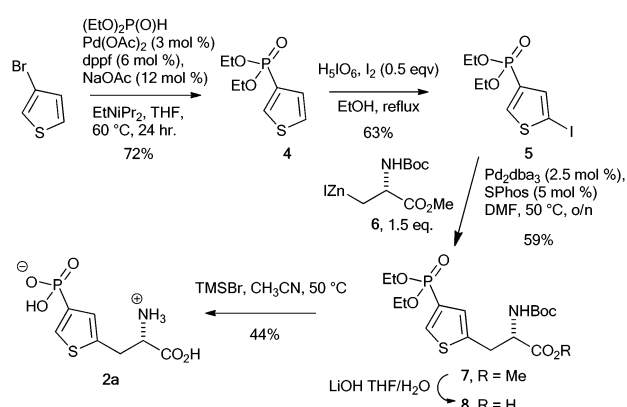
^a Department of Chemistry, The University of Sheffield, Dainton Building, Sheffield, S3 7HF, UK. E-mail: r.f.w.jackson@shef.ac.uk

^b Department of Infection and Immunity, Academic Unit of Respiratory Medicine, The University of Sheffield Medical School, Beech Hill Road, Sheffield, S10 2TH, UK. E-mail: r.muimo@shef.ac.uk

† Electronic supplementary information (ESI) available: Experimental procedures and NMR spectra for **2a**, **4**, **5**, **7** and **8**. See DOI: 10.1039/c4cc03393k

Significantly, incorporation of Pmp and F2Pmp into peptide sequences, has provided phospho-site specific antibodies and more potent or specific phosphatase inhibitors. These peptides have also been used as affinity ligands and in structural studies where they have provided invaluable information about protein–protein interactions.^{8–11} A major limitation is the absence of specific inhibitors against many PTPs. Consequently, PTP research has lagged behind PTK studies and the related field of serine/threonine protein phosphatases.^{12,13}

Motivated by a report on the corresponding furan **2b** that suggested it might be a non-hydrolysable and non-isomerisable analogue of phosphohistidine **3**,¹⁴ we synthesised 4-phosphothiophen-2-yl alanine, **2a**, with the aim of establishing whether the thiophene would better mimic the imidazole in **3** or the benzene ring in **1**. Hirao cross coupling¹⁵ between diethyl phosphite and 3-bromothiophene gave 3-diethoxyphosphothiophene **4**. Iodination of **4** gave, in addition to the 2,5-diodo-derivative (18%), the iodothiophene **5**, which was subjected to Negishi cross-coupling with zinc reagent **6** using our recently optimised conditions¹⁶ to give the protected amino acid **7**. Sequential deprotection then gave 4-phosphothiophen-2-yl alanine **2a**, via the corresponding acid **8** (Scheme 1).



Scheme 1 Synthesis of 4-phosphothiophen-2-yl alanine **2a**.

To explore its potential as a phosphoamino acid analogue, compound **2a** was used as a hapten to produce antibodies. Polyclonal antibodies were raised in rats using KLH-conjugated analogue **2a** as the immunogen. In one of the initial reports of a phosphotyrosine antibody generated using ABP as immunogen, Frackelton *et al.* observed cross-reactivity towards phosphohistidine **3**.⁶ In the course of this study, several non-hydrolysable analogues of phosphohistidine have been reported including phosphopyrrole,¹⁷ phosphoryltriazolylalanine isomers,^{18,19} a sulfonamide-based transition-state analogue of enzymatic phosphohistidine dephosphorylation²⁰ and phosphoryltriazolylethylamine.²¹ Use of phosphoryltriazolylalanine as a mimic of one phosphohistidine in a histone H4 peptide allowed the generation of antibodies that did not universally recognise phosphohistidine in proteins.¹⁸ Antibodies subsequently generated to phosphoryltriazolylethylamine showed cross-reactivity towards phosphohistidine and phosphotyrosine.²¹ The reactivity of the antibodies generated to **2a** was initially tested using dot-blots spotted with BSA-conjugated histidine, phosphoserine, phosphothreonine, phosphotyrosine and τ -phosphohistidine. Antibodies showed strong reactivity towards phosphotyrosine without any cross-reactivity for τ -phosphohistidine, histidine, tyrosine or other phosphohydroxy amino acids (results not shown). This suggested that **2a** was a potential analogue for phosphotyrosine. The antibodies were therefore rescreened against tyrosine, phosphotyrosine, phosphoserine, phosphothreonine and τ -phosphohistidine (Fig. 1, entry A). The antibodies showed high selectivity for phosphotyrosine with minimal side reactivity towards BSA-conjugated τ -phosphohistidine (Fig. 1, A lane v). However, this apparent τ -phosphohistidine cross-reactivity was simply removed by affinity depletion using a histidine-KLH glutaraldehyde conjugated Sepharose column (Fig. 1, entry B) suggesting that the cross linker itself produced a reasonable immune response.

The thiophene in analogue **2a** is evidently a poor mimic of the corresponding imidazole present in τ -phosphohistidine, but a good mimic of the benzene ring present in phosphotyrosine, which has resulted in a polyclonal serum that detects phosphotyrosine in preference to τ -phosphohistidine.

Competition studies, performed by ELISA on the purified polyclonal antibody, showed no binding or inhibition of the antisera by any of the amino acids apart from phosphotyrosine (Fig. 2). This indicates that while analogue **2a** may not be a

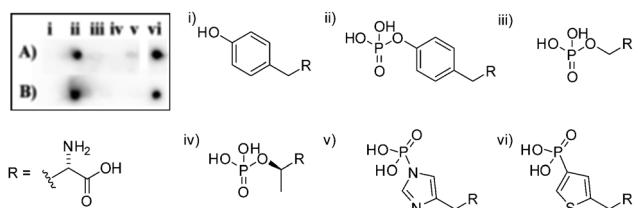


Fig. 1 Dot-blots of 4-phosphothiophen-2-yl alanine generated antibody using (A) crude polyclonal antibody and (B) polyclonal antibody affinity depleted with Sepharose bound/glutaraldehyde linked histidine-KLH when probed for (i) tyrosine, (ii) phosphotyrosine, (iii) phosphoserine, (iv) phosphothreonine, (v) τ -phosphohistidine, (vi) 4-phosphothiophen-2-yl alanine.

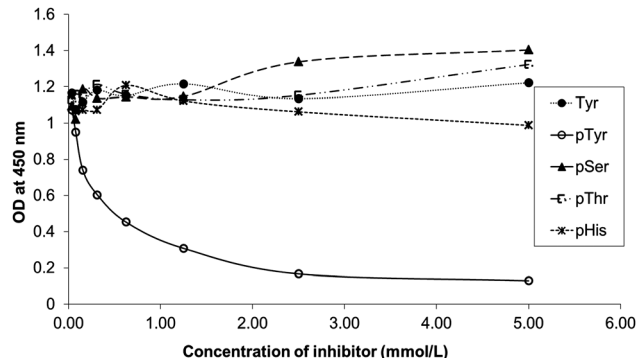


Fig. 2 Competitive inhibition ELISA study using anti-sera generated from 4-phosphothiophen-2-yl alanine, **2a**.

suitable analogue for τ -phosphohistidine it was likely to be a useful analogue for phosphotyrosine. Competitive inhibition ELISA analysis using *O*-phospho-L-tyrosine (pTyr) or **2a**, demonstrated that the antibody generated from **2a** has a higher affinity towards **2a** compared to phosphotyrosine (Fig. 3, see inset), as expected.

In Western blot analysis, using cytosolic fractions obtained from sheep tracheal epithelia, the antibody generated from **2a** gave a staining profile comparable to that of a commercially available anti-phosphotyrosine monoclonal antibody (pY99) – albeit with varying degrees of staining or detection intensity for various protein bands by either antibody (Fig. 4A). This indicates that the antibody generated by **2a** is able to detect phosphotyrosine containing proteins and recognises many protein bands that are also detected by the anti-phosphotyrosine pY99. Interestingly, the antibody generated from **2a** also detected proteins that were not detected by the pY99 (and *vice versa*). This is consistent with previous studies where different types of antibodies provided varied results in the same systems.²²

Furthermore, in total lysates of human bronchial epithelial cell lines (16HBE14o-) treated with Na_3VO_4 (a broad spectrum PTP inhibitor) for 45 min, both the antibody generated from **2a** and pY99 detect enhanced phosphorylation of a high molecular

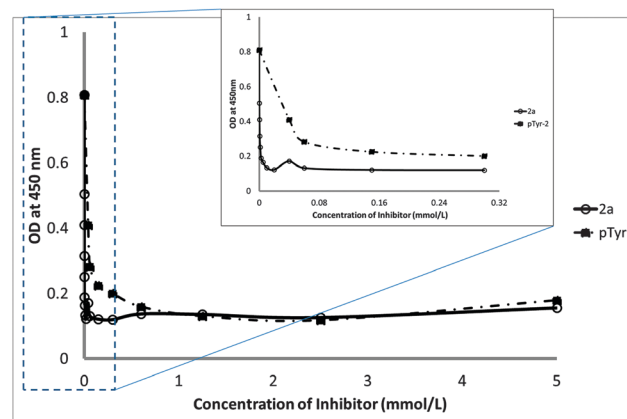


Fig. 3 A competitive inhibition ELISA study involving *O*-phospho-L-tyrosine (pTyr) and 4-phosphothiophen-2-yl alanine, **2a** using anti-sera generated from analogue, **2a**.

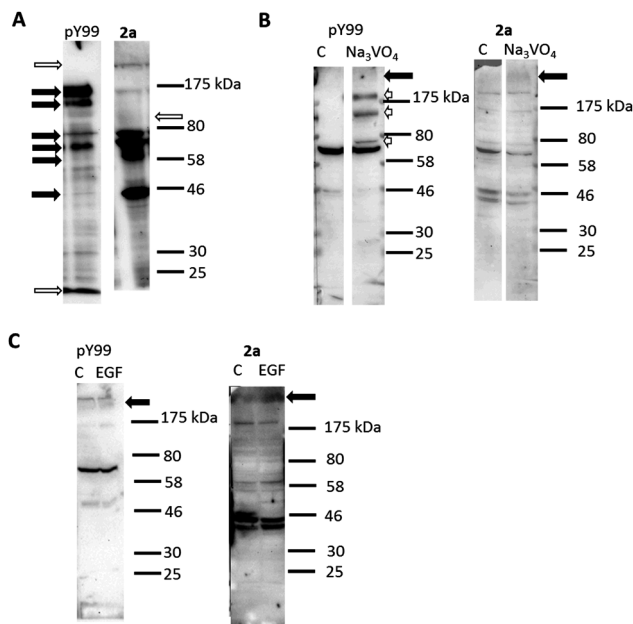


Fig. 4 Comparison of antibody generated to **2a** and commercially available anti-phosphotyrosine monoclonal antibody (pY99). (A) Western blots of cytosolic fraction from sheep tracheal epithelia probed with pY99 or **2a** antibody. Arrows highlight similarities (filled) or differences (unfilled). Western blots of lysates of human bronchial epithelial cell lines (16HBE14o-) treated \pm Na₃VO₄ (10 mM) for 45 min (B) or \pm EGF (1 nM) for 4 hours (C) and probed with pY99 (left panel) or the antibody generated from **2a** (right panel), (C = control, untreated). Arrows/arrowheads highlight similarities (filled) or differences (unfilled).

weight protein (Fig. 4B, filled arrow). However, pY99 additionally detects enhanced phosphorylation of three other proteins (approx. 75, 100, 180 kDa) in Na₃VO₄ treated cells (Fig. 4B, left panel, unfilled arrow heads). Interestingly, both antibodies also detect several other proteins that were insensitive to Na₃VO₄ and in this regard, pY99 antibodies detected two, whereas the antibody generated from **2a** detected six, of these proteins. In lysates from cells stimulated with epidermal growth factor (EGF, 1 nM for 4 hours), both the antibody generated from **2a** and pY99 detected enhanced phosphorylation of a high molecular weight protein (Fig. 4C, filled arrow).

In conclusion, we have synthesized 4-phosphothiophen-2-yl alanine, **2a** and demonstrated its potential as a novel 5-membered

ring analogue of phosphotyrosine. Our data suggests that analogue **2a**, and resulting polyclonal antibody, may potentially be a useful tool to both characterize the biological effects of phosphotyrosine and the role of specific PTP(s) in cells.

We thank the EPSRC Life Sciences Interface DTC (ML). The 16HBE14o- cells were provided by Dr D. Gruenert (California Pacific Medical Center, CA).

Notes and references

- 1 T. Hunter, *Cell*, 2000, **100**, 113–127.
- 2 J. A. Ardura and P. A. Friedman, *Pharmacol. Rev.*, 2011, **63**, 882–900.
- 3 L. Bialy and H. Waldmann, *Angew. Chem., Int. Ed.*, 2005, **44**, 3814–3839.
- 4 A. Noeren-Mueller, I. Reis-Correa, Jr., H. Prinz, C. Rosenbaum, K. Saxena, H. J. Schwalbe, D. Vestweber, G. Cagna, S. Schunk, O. Schwarz, H. Schiewe and H. Waldmann, *Proc. Natl. Acad. Sci. U. S. A.*, 2006, **103**, 10606–10611.
- 5 K. Shen, *Methods*, 2007, **42**, 234–242.
- 6 A. R. Frackelton, A. H. Ross and H. N. Eisen, *Mol. Cell. Biol.*, 1983, **3**, 1343–1352.
- 7 G. Steinhilber, M. W. Sproll, J. M. Wolff and F. A. Anderer, *Anticancer Res.*, 1990, **10**, 907–911.
- 8 Z. Y. Zhang, D. Maclean, D. J. McNamara, T. K. Sawyer and J. E. Dixon, *Biochemistry*, 1994, **33**, 2285–2290.
- 9 M. R. Groves, Z. J. Yao, P. P. Roller, T. R. Burke and D. Barford, *Biochemistry*, 1998, **37**, 17773–17783.
- 10 M. Akamatsu, P. P. Roller, L. Chen, Z. Y. Zhang, B. Ye and T. R. Burke, *Bioorg. Med. Chem.*, 1997, **5**, 157–163.
- 11 K. Shen, L. X. Qi and L. Stiff, *Curr. Pharm. Des.*, 2010, **16**, 3101–3117.
- 12 J. J. Fernandez, M. L. Candelas, M. L. Souto, M. M. Trujillo and M. Norte, *Curr. Med. Chem.*, 2002, **9**, 229–262.
- 13 C. F. B. Holmes, J. T. Maynes, K. R. Perreault, J. F. Dawson and M. N. G. James, *Curr. Med. Chem.*, 2002, **9**, 1981–1989.
- 14 C. Schenkels, B. Erni and J. L. Reymond, *Bioorg. Med. Chem. Lett.*, 1999, **9**, 1443–1446.
- 15 M. Kalek, M. Jezowska and J. Stawinski, *Adv. Synth. Catal.*, 2009, **351**, 3207–3216.
- 16 A. J. Ross, H. L. Lang and R. F. W. Jackson, *J. Org. Chem.*, 2010, **75**, 245–248.
- 17 P. V. Attwood, M. J. Piggott, X. L. Zu and P. G. Besant, *Amino Acids*, 2007, **32**, 145–156.
- 18 J.-M. Kee, B. Villani, L. R. Carpenter and T. W. Muir, *J. Am. Chem. Soc.*, 2010, **132**, 14327–14329.
- 19 T. E. McAllister, M. G. Nix and M. E. Webb, *Chem. Commun.*, 2011, **47**, 1297–1299.
- 20 M. F. Eerland and C. Hedberg, *J. Org. Chem.*, 2012, **77**, 2047–2052.
- 21 J. M. Kee, R. C. Oslund, D. H. Perlman and T. W. Muir, *Nat. Chem. Biol.*, 2013, **9**, U416–U428.
- 22 J. Y. J. Wang, *Anal. Biochem.*, 1988, **172**, 1–7.