

Prospects for the development of probiotics and prebiotics for oral applications

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There has been a paradigm shift towards an ecological and microbial community-based approach to understanding oral diseases. This has significant implications for approaches to therapy and has raised the possibility of developing novel strategies through manipulation of the resident oral microbiota and modulation of host immune responses. The increased popularity of using probiotic bacteria and/or prebiotic supplements to improve gastrointestinal health has prompted interest in the utility of this approach for oral applications. Evidence now suggests that probiotics may function not only by direct inhibition of, or enhanced competition with, pathogenic micro-organisms, but also by more subtle mechanisms including modulation of the mucosal immune system. Similarly, prebiotics could promote the growth of beneficial micro-organisms that comprise part of the resident microbiota. The evidence for the use of pro or prebiotics for the prevention of caries or periodontal diseases is reviewed, and issues that could arise from their use, as well as questions that still need to be answered, are raised. A complete understanding of the broad ecological changes induced in the mouth by probiotics or prebiotics will be essential to assess their long-term consequences for oral health and disease.

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In recent years, there have been significant changes with respect to the effectiveness of, and attitudes towards, conventional antimicrobial therapy to combat disease. With the threat of widespread antibiotic resistance rendering many antibiotics useless against important diseases, there is an increased necessity not only to minimise antibiotic use and develop novel non-antibiotic-based treatments, but also to raise the profile of disease prevention. There is a public appetite for new therapies that are perceived to be natural through, for example, manipulation of the resident microbiota by the ingestion of probiotic bacteria or prebiotics. These changing attitudes are also relevant to the prevention of dental diseases and there is an increased interest in the use of strategies that do not involve conventional antimicrobial agents for oral care (1–3).

There has been a paradigm shift away from treating dental diseases by targeting specific oral pathogens towards an ecological and microbial community-based approach to understand conditions, such as caries and periodontal diseases (4,5). These approaches recognise the importance of maintaining the natural balance of the

resident oral microbiota and the need to carefully modulate host immune responses to the microflora at a site.

One approach that has gained interest over recent years is the use of probiotic bacteria for oral applications. The rationale for their use in oral health care stems from the increase in evidence that supports their claims for benefit for a range of diseases, especially in the gastrointestinal tract (6–12). In this article, we will review the data on the use of probiotics for oral care or disease prevention, and discuss some of the issues that arise from their use, as well as identify questions that still need to be answered.

Probiotics and prebiotics

There is a long tradition, particularly in parts of Europe and Asia, of ingesting microbes or food products that affect the intestinal microbiota in ways that are believed to provide beneficial health effects, i.e. intake of probiotics and prebiotics. Probiotics are defined as viable micro-organisms that confer health benefit when administered in sufficient doses (6). The organisms that have been used as probiotics are primarily certain species of lactobacilli

and bifidobacteria, and *Saccharomyces* spp., but some streptococci, enterococci and commensal *Escherichia coli* have also been claimed to have beneficial effects in certain situations (1, 6, 13, 14). Prebiotics (e.g. inulin-type fructans, maltodextrin, fructooligosaccharides and galactooligosaccharides) have been defined as non-digestible oligosaccharides that affect the proliferation of resident commensal bacteria that may then exert probiotic effects (15). More recently, the definition has been refined to include selectively fermented ingredients that allow specific changes in the composition and/or activity of the resident microflora that confer benefits upon host well-being and health (16). Studies of prebiotics have mainly been focused on gastrointestinal microbiota and health benefits; there has been little work in the oral cavity.

Much of the evidence for the health benefits of probiotics and prebiotics has been anecdotal, but the last decade has seen some developments in establishing the scientific base for administration of such agents and in understanding the mechanisms underlying their effects. This is reflected in the proliferation of reviews in this area in recent years (1, 6–14, 17–21).

Current applications of probiotics and prebiotics

Most of the applications and research into the mechanisms of action of probiotics and prebiotics concentrate on their roles in influencing intestinal health and function. Although some of the experimental evidence and data from clinical trials is conflicting, there is growing evidence for their efficacy in protecting against acute diarrhoeal disease in children, gastroenteritis and antibiotic-associated diarrhoea, inflammatory bowel diseases and pouchitis (6, 7, 10, 12). There is also evidence to support further investigation of the use of probiotics and prebiotics in the treatment of illnesses affecting sites other than the intestinal tract, e.g. urinary tract infections, vaginal infections, arthritis, atopic eczema, pharyngitis and otitis media (6, 7, 11, 22). Recently, *Lactobacillus rhamnosus* GG (LGG) administered in yoghurt was reported to enhance faecal clearance of vancomycin-resistant enterococci (23). The possibilities of applying probiotic therapy for other medical conditions are being investigated, including recovery from haemorrhagic shock, recovery from burn injury, cholesterol reduction and protection from coronary heart disease, effects on breast cancer cells, enhancement of tolerance of food allergens, protection from respiratory tract infections, liver conditions, skin infections, enhancement of bone health and reduction of obesity (18, 20, 21). However, the evidence-base for many of these is relatively under-developed.

The potential applications of probiotic bacteria have been further expanded by the development of strains that

have been genetically engineered to produce the anti-inflammatory cytokine IL-10 (24), trefoil factor family proteins to enhance wound healing (25) or the 2D-CD4 receptor to try to reduce HIV infectivity (26).

Mechanisms of action

Probiotics

The diversity of conditions that may benefit from ingestion of probiotics illustrates the variety of mechanisms that may be involved in their actions and that some effects are systemic rather than only local. It is likely that these mechanisms vary according to the specific strain or combinations of strains used, the presence of prebiotics and the condition that is being treated, as well as the stage of the disease process in which the probiotic is administered (7). There are common themes emerging in studies of the modes of action of probiotics and numerous mechanisms have been proposed (7, 9–11, 13) including:

- Prevention of adhesion of pathogens to host tissues.
- Stimulation and modulation of the mucosal immune system, e.g. by reducing production of pro-inflammatory cytokines through actions on NF κ B pathways, increasing production of anti-inflammatory cytokines such as IL-10 and host defence peptides such as β -defensin 2, enhancing IgA defences and influencing dendritic cell maturation.
- Modulation of cell proliferation and apoptosis through cell responses to, for example, microbially produced short chain fatty acids.
- Improvement of intestinal barrier integrity and up-regulation of mucin production.
- Killing or inhibition of growth of pathogens through production of bacteriocins or other products, such as acid or peroxide, which are antagonistic towards pathogenic bacteria.

Prebiotics

The ability of certain oligosaccharides to enhance the growth of resident commensal gut bacteria, particularly bifidobacteria and lactobacilli, is well documented (17). Thus, the major mechanism of action of prebiotics is assumed to be indirect, i.e. facilitating the proliferation of beneficial components of the resident microflora, with probiotic effects resulting from the actions of these bacteria as described above. Cellobiose has the additional property of down-regulating virulence factors of *Listeria monocytogenes* (27). There is evidence that some prebiotics also exert direct effects on the host, independent of their effects on resident bacterial populations (8, 15). These include stimulation of expression of IL-10 and interferon γ , enhancement of IgA secretion, modulation of inflammatory responses to pathogens and stabilisation

of the gut mucosal barrier. Additionally, prebiotics with enhanced function have been designed. These oligosaccharide derivatives contain sugars that are specific epithelial cell receptors to which pathogens adhere and they, therefore, provide ‘decoy’ adhesion sites and cause pathogens to adhere to luminal contents rather than to epithelial cells (17).

The oral microbiota in health and disease

To be able to develop probiotic or prebiotic interventions for applications in oral health care and to understand their mechanisms of action and potential risks, it is essential to have a clear understanding of the oral microbiota and their functions in oral health and disease. This is not always easy, given the complexity of the oral microbiota; more than 700 species have been detected in the human mouth and the resident microbiota of one individual may comprise 30 to >100 species (28–30).

A wide variety of sites in the mouth are heavily colonised. Supragingival and subgingival plaque form through sequential and specific adhesive interactions that result in a complex climax community (5, 31). The tongue is heavily colonised and micro-organisms on the dorsum of the tongue are reservoirs for supragingival and subgingival plaque and salivary microbial populations (32–34). Many oral bacteria, especially streptococci, also survive within buccal epithelial cells (35, 36).

Functions of the resident microbiota

The main focus of research has been defining the micro-organisms and their traits that are responsible for disease, but there is an increased awareness that the resident microbiota does not play merely a passive role, but actively contributes to the maintenance of health. The large, diverse resident microbial communities that colonise mucosal sites co-exist with a host, with harmful effects only if the host becomes immunocompromised, if the resident microbial populations are suppressed or if micro-organisms reach sites to which they do not normally have access (e.g. through trauma). Studies, mostly of gastrointestinal bacteria, have shown that resident microbial populations contribute to host protection through blocking of colonisation by pathogens (37, 38), development of cell structure and function (39, 40), development of the immune system (41) and modulation of inflammatory responses (42–49). Evidence is accumulating to support a similar role for oral commensal bacteria in the development of the immune system (50), the maintenance of healthy oral tissue by influencing expression of mediators such intracellular adhesion molecule 1 (ICAM-1), E-selectin and IL-8 (51), modulating immune responses and enhancing cellular homeostatic mechanisms (52, 53).

Defining the resident microbiota

Technological improvements in the detection of culturable and non-culturable micro-organisms has led to the identification of increasing numbers of taxa in the mouth (54, 55) and have confirmed that resident oral microbial populations are site-specific as well as highly diverse, and the profile of the microbial community may be specific to an individual (28, 29, 33, 56). Species that predominate in disease can often be present in low numbers at healthy sites (5, 31).

In recent years there has been a greater emphasis, not only on defining resident microbial populations more fully, but also on identifying those that are significantly positively associated with health in an effort to better understand the processes that eventually lead to disease and the ways in which microbial populations may be manipulated to maintain host–microbe homeostasis and to develop novel prevention strategies. Kilian et al. (57) list the following species as ‘true’ oral commensal micro-organisms: *Streptococcus mitis*, *Streptococcus oralis*, *Actinomyces naeslundii*, *Fusobacterium nucleatum*, *Haemophilus parainfluenzae*, *Eikenella corrodens* and some species of *Prevotella*. Other studies have generated an increasingly long list of culturable and unculturable bacteria with a significant association with healthy sites (28, 30, 58–60).

Microbial populations associated with oral disease

The most common oral diseases are caries and periodontitis, which result from a shift in the balance of the resident microbiota at a site. The types and proportions of bacteria found in plaque taken from sites diagnosed with either caries or periodontal disease differ from one another and both are distinct from those that predominate at healthy sites. In caries, there are increases in acidogenic and acid-tolerating species such as mutans streptococci and lactobacilli, although other bacteria with similar properties can also be found and bifidobacteria, non-mutans streptococci, *Actinomyces* spp., *Propionibacterium* spp., *Veillonella* spp. and *Atopobium* spp. have also been implicated as significant in the aetiology of this disease (30, 61–65).

In periodontal diseases, there is an increase in plaque mass and a shift towards increases in obligately anaerobic and proteolytic bacteria, many of which are Gram negative and currently unculturable. The host damage that occurs during periodontal disease arises through the combined activities of subgingival biofilms and the host responses to these diverse bacterial populations. A number of reviews give excellent overviews of periodontal microbiology (5, 54, 57, 66–69) and these illustrate the significant paradigm shift that has occurred, away from concentrating on the roles of individual specific pathogens to recognising that periodontal disease results from the activities of successive consortia of organisms.

Other common oral infections also result from the activities of micro-organisms that are found in the resident microbiota. *Candida albicans* and other *Candida* species are present in low levels in oral microbial communities and can cause oral candidiasis and denture-associated stomatitis (70, 71). Halitosis is most often the result of production of malodorous metabolic end-products (especially volatile sulphur compounds) by oral bacteria, in particular Gram negative anaerobes (72, 73).

The potential for probiotics in prevention and control of oral diseases

Probiotics in prevention of caries

The oral health applications of either probiotics or 'replacement therapy' with *Streptococcus mutans* strains of attenuated virulence and increased competitiveness were first suggested for prevention of dental caries more than 20 years ago (74). Despite this, and the fact that some products have reached the market, there remains a paucity of clinical evidence to support the effectiveness of probiotics to prevent or treat caries (2, 3).

Many early studies concentrated on utilising bacteria that expressed bacteriocins or bacteriocin-like inhibitory substances (BLIS) that specifically prevented the growth of cariogenic bacteria (11, 74). Another approach has been to identify food grade and probiotic bacteria that may have potential in caries prevention. These have been selected because of their likely ability to colonize teeth and influence the supragingival plaque; *in vitro* models for this selection have included adhesion to hydroxyapatite, as a surrogate for colonisation of teeth, and mixed species biofilm models (75, 76). Also, strains have been screened for suitable antagonistic activity against relevant oral bacteria (77). *In vitro* studies of the antibacterial activity of live yoghurts showed inhibition of *S. mutans* but not some other oral streptococci, including *Streptococcus sobrinus*; this activity was heat-sensitive implying that the effect was not simply due to acid (77). Recently, oral lactobacilli have also been screened for their utility as potential probiotic strains (78–80) and strains of oral lactobacilli have been isolated that are inhibitory against *S. mutans*, *Aggregatibacter actinomycetemcomitans*, *Porphyromonas gingivalis* and *Prevotella intermedia*, as well as being tolerant of relevant environmental stresses (81). Another approach utilised a recombinant strain of *S. mutans* expressing urease, which was shown to reduce the cariogenicity of plaque in an animal model (82). Similarly, genetically modified probiotics with enhanced properties can be developed ('designer probiotics'). For example, a recombinant strain of *Lactobacillus* that expressed antibodies targeting one of the major adhesins of *S. mutans* (antigen I/II) was able to reduce both the viable counts of *S. mutans* and the caries score in a rat model (83).

Clinical studies have indicated that bacteria with established probiotic effects (lactobacilli and bifidobacteria) have some promise for prevention of caries. LGG ingested in dairy products (milk, cheese) reduced salivary mutans streptococcal counts in adults and protected against caries in children (84, 85). Other lactobacilli have also been shown to reduce mutans streptococcal counts in saliva. *Lactobacillus reuteri*, when delivered by yoghurt (86), straw or tablet (87), by chewing gum (88) or as a lozenge (89), significantly reduced the counts of mutans streptococci in saliva ($p < 0.05$). The short-term consumption of yoghurt (90) or ice cream (91) containing *Bifidobacterium* spp. resulted in a significant reduction in salivary mutans streptococci ($p < 0.05$) but not in lactobacilli. Other studies have reported reductions in mutans streptococci levels in saliva following use of probiotic-containing yoghurts (92).

Probiotics in prevention of periodontal diseases

There are fewer experimental studies exploring probiotic use in periodontal diseases, partly reflecting a poorer understanding of the precise aetiology of the disease and of the conditions that promote health. However, patients with moderate to severe gingivitis who were given either one of two *L. reuteri* formulations had reduced plaque and gingivitis scores compared to a placebo group (93). Similarly, the regular (three times daily for eight weeks) intake of tablets containing *Lactobacillus salivarius* resulted in benefits in terms of pocket probing depth and plaque index in individuals at high risk of periodontal disease (smokers) compared to a placebo control group (94). Other studies have aimed to identify organisms that have the potential for probiotic action that may protect against periodontal diseases. Some oral strains of lactobacilli and streptococci (81, 95–97) and bifidobacteria (98) have been reported to have *in vitro* inhibitory activity against periodontal pathogens, while others are more active against mutans streptococci (79–81). The subgingival application of beneficial oral bacteria (e.g. *Streptococcus sanguinis*, *Streptococcus salivarius* and *S. mitis*) (replacement therapy) has been shown to delay recolonisation by periodontal pathogens, reduce inflammation, and improve bone density and bone levels in a beagle dog model (99, 100). Koll-Klais et al. (97) observed that *Lactobacillus gasseri* strains isolated from periodontally healthy subjects were more efficient at inhibiting the growth of *A. actinomycetemcomitans* than strains from periodontally diseased subjects, and also inhibited the growth of *P. gingivalis* and *P. intermedia*; this correlated with an inverse relationship between carriage of homofermentative lactobacilli and subgingival colonisation by *A. actinomycetemcomitans*, *P. gingivalis* and *P. intermedia*. Ishikawa et al. (96) observed *in vitro* inhibition of *P. gingivalis*, *P. intermedia* and *Prevotella nigrescens* by *L. salivarius*. Daily ingestion of

L. salivarius-containing tablets resulted in reduced salivary counts of these black pigmented anaerobes.

The mechanisms of inhibition of periodontal pathogens have not been fully clarified. The inhibitory activity displayed by homofermentative lactobacilli against periodontal pathogens was principally related to their production of acid, not to H₂O₂ or bacteriocin production (97). Hojo et al. (98) suggested that bifidobacteria inhibit some black pigmented anaerobes by competing for an essential growth factor, vitamin K, although there was no significant relationship between higher bifidobacterial counts and lower black-pigmented anaerobe counts. Recently, a bacteriocin purified from *Lactobacillus casei* killed *P. gingivalis* but its use was proposed as a novel chemotherapeutic agent rather than as strain development for probiotic applications (101).

Probiotics in prevention of other oral diseases

Probiotics have also been reported to reduce yeast counts in the elderly, and may be a route to control *Candida* spp. and hyposalivation in this age group (102). There have also been clinical and laboratory studies of their potential for preventing halitosis. Peroxide production by strains of *Weissella cibaria* (commonly present in fermented foods) isolated from the mouths of healthy children, inhibited production of volatile sulphur compounds that contribute to oral malodour by *F. nucleatum in vitro* and in exhalations following mouth-rinsing by adult volunteers with a suspension of *W. cibaria* (103). The success of *W. cibaria* in reducing malodour may have also been because it coaggregated efficiently with *F. nucleatum* (103) and therefore competed with other late/secondary colonisers for adhesion sites. Thus, *W. cibaria* may have probiotic activities with potential for prevention of periodontal disease. Volatile sulphur compounds, such as H₂S and mercaptoethanol, are produced by a range of periodontal anaerobes (104). The inhibition of these micro-organisms by peroxide from *W. cibaria* may help reduce subgingival plaque pathogenicity while its competition for coaggregation sites may reduce the reservoir of micro-organisms available for transmission into plaque.

S. salivarius is one of the earliest colonisers of epithelial surfaces in the human mouth and nasopharynx, and its primary habitat is the dorsum of the healthy tongue (28, 73). *S. salivarius* K12 produces salivaricin, a lantibiotic with inhibitory activity towards most *Streptococcus pyogenes* (11). This strain has been commercially promoted as a probiotic that is reported to be protective against throat infections and oral malodour (11, 105). *S. salivarius* K12 displays other activities, not related to salivaricin production, that most likely contribute to its probiotic properties. It down-regulated IL-8 secretion by epithelial cells in response to pathogenic bacteria and to the immunomodulatory host defence peptide LL-37, and also influenced numerous cellular homeostatic pathways

(53). *Streptococcus gordonii* was recently shown to also inhibit epithelial cell IL-6 and IL-8 secretion (52).

Many other strains of *S. salivarius* are reported to produce bacteriocins or BLIS, leading to suggestions for their usefulness as oral probiotics (106). Two salivaricin-producing strains, when administered to children in milk, promoted salivaricin A-like inhibitory activity in the indigenous, resident *S. salivarius* populations (107). The importance of strain selection for probiotic use is illustrated by the fact that some *S. salivarius* strains differ from K12 in some important activities; one strain increased production of malodorous products by facilitating *P. gingivalis* metabolism of salivary mucins (108) and another up-regulated IL-8 secretion by oral epithelial cells (109) in contrast to the down-regulation observed in response to K12.

Outstanding questions

Can probiotics colonise the oral cavity?

Most evidence indicates that probiotics in the gut are not able to permanently become part of the resident gastrointestinal microbiota and they disappear from faeces very soon after probiotic ingestion ends. Previous studies of the oral microbiota would indicate that it is very difficult to alter the composition of established plaque microbial communities (57). A number of studies of oral colonisation following probiotic ingestion have found similar patterns of lack of colonisation to those in the gut, in that ingested lactobacilli colonised only transiently and disappeared soon after administration of the probiotic ended (92, 110, 111). Colonisation with *L. reuteri* was achieved in the majority of periodontal patients given a probiotic, but the study only ran for 14 days (93). Stable and long-term colonisation by probiotic lactobacilli appears to have only been observed in an individual who received LGG probiotic therapy at the age of 10 years (111). The resident microbiota of children seems to be less stable and more subject to flux than resident microbial communities in adults (57), and perhaps it is in childhood that long-term influences on resident populations will be achieved.

Is colonisation of plaque essential for protection against caries or periodontitis?

There is some evidence that colonisation of the gut by probiotics may have beneficial systemic effects, enabling these organisms to provide protection against diseases at distant sites (22). Studies of the influence of *L. reuteri* ATCC55730 on salivary mutans streptococci and lactobacilli indicated that the benefits seen may have been due to systemic effects rather than to the colonisation of the mouth by the probiotic bacterium (87). Probiotics have been effective in some chronic inflammatory diseases that involve deregulation of the immune responses, e.g. arthritis

and Crohn's disease. Some of the systemic effects claimed for probiotics are immunomodulation, alteration of mucin production, stabilisation of mucosal barriers, enhancement of IgA defences, effects on neutrophils and dendritic cells, (7, 9–11, 13) and enhancement of bone health through influencing bone mineral content and structure (20). Chronic inflammation and bone resorption contribute significantly to the pathogenesis of periodontal diseases, and it is possible to speculate that some of these systemic effects might provide concomitant protection against periodontal diseases. However, no studies have been carried out providing evidence for this.

It is also possible that colonisation of one site may provide indirect protection at other sites by mechanisms other than systemic effects. For example, reduction of colonisation of the tongue may reduce reservoirs for colonisation of plaque. Supragingival plaque and subgingival plaque are intimately connected, as supragingival plaque extends down the tooth to form subgingival plaque, so changes in supragingival plaque will influence the future composition of subgingival plaque. Lactobacilli associated with periodontal health were only rarely isolated from subgingival samples (97). However, these bacteria were found to inhibit the growth of certain periodontal pathogens; it was proposed that they may reduce the levels of these pathogens on the tongue, which constitutes a major reservoir for their transmission, and thereby indirectly reduce the colonisation of subgingival plaque by periodontal pathogens.

Are current approaches targeting the right micro-organisms?

Most oral diseases are polymicrobial in nature and result from complex ecological shifts in the resident microbiota. In caries, the ability of bacteria to colonize plaque, produce acid and survive under low pH conditions are of over-arching importance and these properties are not restricted to a few species (112). Thus, highly targeted approaches may have limited success as there are so many other micro-organisms that can occupy a similar niche. Also, there has been an emphasis on identifying probiotics that will have an effect on bacteria that have strong associations with established or severe disease; for example, strains are proposed to be potentially useful against periodontal disease if they have inhibitory activity against *P. gingivalis* or *A. actinomycetemcomitans*. By the time these species are prevalent, the disease is well established and the site is already in crisis; a more effective therapeutic approach than to target these late pathogens might be to inhibit the growth or activity of those microbial consortia that are associated with the transition from health to disease. The advances that have occurred in the technologies used to detect and characterise microbial populations are leading to a more detailed characterisation of the microbiota associated

with specific phases of health and disease so this approach is becoming a realistic possibility. Finally, there is a widespread acceptance of the importance of oral ecology and maintenance of host–microbe homeostasis in oral health and disease (5, 113, 114). Recognition of the activities of bacteria that contribute to disease (e.g. acid production in caries, induction of inflammation and bone resorption in periodontal disease) may lead to therapies that target such activities, rather than certain species.

Are prebiotics a viable alternative or adjunct?

For a rational approach to the development of oral prebiotics and the manipulation of the resident microbiota, it is essential to know which species can be considered to promote health and to gain some understanding of their metabolic needs and interactions. It is recognised that the resident oral microbiota persists by catabolising endogenous nutrients such as salivary proteins and glycoproteins (115) and gingival crevicular fluid.

Clearly poor diet has an impact on oral health as well as general health, and controlling refined sugar intake has been used for many years to control the oral ecology and protect against caries. Similarly, xylitol has been used to reduce acid production by mutans streptococci, although this made no difference to the effectiveness of a probiotic-containing chewing gum (88). Algal lectins and cranberry juice have been suggested to reduce adhesion of oral streptococci (116, 117). Cocoa polyphenols can reduce the viability and acid production by cariogenic bacteria (118). However, while these all use (or suggest the use of) dietary components to influence the oral microbiota, they are not prebiotics as they inhibit potential pathogens rather than stimulate beneficial resident micro-organisms.

We know little about the impact of dietary components on subgingival plaque composition. In the gut there is some evidence collected over many years for the beneficial effects of promoting populations of bifidobacteria and lactobacilli. Koll-Klais et al. (97) found that homofermentative lactobacilli, particularly *L. gasseri*, were more prevalent in healthy rather than periodontally diseased sites; their presence was inversely associated with clinical parameters related to chronic periodontitis and also to subgingival colonisation by periodontal pathogens. Hojo et al. (98) also found *L. gasseri*, as well as *L. salivarius* and *Lactobacillus fermentum*, to be more prevalent in healthy sites but not exclusive to health. The same study found that *Bifidobacterium* spp., although not predominant in the mouth, were isolated from 80% of periodontally healthy subjects and *Bifidobacterium adolescentis* was isolated from 40% of healthy subjects and no periodontitis subjects. Counts of bifidobacteria were particularly high in a group of well-maintained ex-periodontitis subjects, indicating perhaps

that these bacteria are better able to colonise sites that have undergone plaque removal. Thus, it is possible that prebiotic therapies that promote the growth of certain bifidobacteria and lactobacilli may enhance periodontal health. However, lactobacilli and bifidobacteria are themselves linked to caries aetiology, and it is also difficult to envisage how a prebiotic approach to enhance their growth would not encourage a general increase in prevalence of aciduric and acidogenic populations that are associated with an increased risk of dental caries.

Are there potential risks?

It is worth sounding a note of caution concerning the use of probiotics for the purpose of preventing oral disease. Different strains of a species may not all possess characteristics that enable them to be probiotics and rigorous strain selection for the disease concerned is complex but essential (7, 18). Some probiotic strains have been in use for many years and have excellent safety records (119–121). Most probiotic bacteria are weakly proteolytic and, for example, *Lactobacillus bulgaricus*, was shown to be incapable of degrading some host tissue components (122). However, there have been some cases of bacteraemia and fungaemia associated with probiotic use, although these have been in subjects who are immunocompromised (123, 124), or who suffer from chronic disease (119) or short gut syndrome (125). Other predisposing factors include prior prolonged hospitalisation and prior surgical intervention (124). An individual who had been taking *L. rhamnosus* in a probiotic preparation developed *Lactobacillus* endocarditis following dental treatment (126). In Finland, however, there has not been an increase in bacteraemia associated with probiotic lactobacilli following the increase in the use of these products since 1990 (127).

The species that most commonly exhibit probiotic benefits are lactobacilli and other lactic acid bacteria, and the production of acid is often thought to be an important component of their protection against pathogenic colonisation. However, *Lactobacillus* spp. and acid production by acidogenic plaque populations play a significant part in the development of caries, and a probiotic strain of *L. salivarius* has been shown to be cariogenic in a rat model (128). A number of probiotic lactobacilli and bifidobacteria produce acid from fermentation of dietary sugars *in vitro* (129). There are conflicting data on the salivary lactobacilli levels following probiotic usage. Some studies have reported no effects (91), others have found trends for an increase (1, 87), while others have detected statistically significant rises in counts of salivary lactobacilli (130). There is a converse risk in that the control or prevention of caries may indirectly affect periodontal pathogens. It has been known for many years that streptococci, through production of hydrogen peroxide, inhibit the growth of putative

periodontal pathogens, leading to early proposals that interactions between groups of micro-organisms within plaque can influence the development of disease or actively contribute to the maintenance of health (131), and lactobacilli and bifidobacteria also inhibit the growth of a range of periodontal pathogens (81, 95–98, 131).

It is clear that careful selection of the strain to be ingested for a particular disease is essential and the mode and timing of administration can be crucial, as well as the age and health of the individual taking the probiotic. There is a sufficient knowledge base for major and minor risk factors to have been proposed for administration of probiotics to prevent intestinal conditions (119), but this knowledge base for oral applications is clearly more distant. One of the biggest problems to overcome may be that the probiotic activities and micro-organisms that protect against oral disease could increase the risk of development of dental caries. Therefore, a prebiotic-type approach to enhance endogenous beneficial commensals may be more attractive. It will also be a challenge to ensure that modes of delivery are developed that provide sufficient retention and exposure times in the mouth that will allow probiotics to colonise plaque or prebiotics to enter into plaque or mucosal biofilms and influence microbial metabolism within them.

In conclusion, the use of probiotics for use in oral care applications is gaining momentum. There is increasing evidence that the use of existing probiotic strains can deliver oral health benefits. Further work will be needed to fully optimise and quantify the extent of this benefit. In parallel, the potential of prebiotics to maintain and enhance the benefits provided by the resident oral microbiota will be investigated. However, whether considering probiotics or prebiotics, it will be essential to develop an understanding of the broad ecological changes induced in the mouth by their ingestion and the long-term consequences of their use on oral health and disease.

References

1. Caglar E, Kargul B, Tanboga I. Bacteriotherapy and probiotics' role on oral health. *Oral Dis* 2005; 11: 131–7.
2. Meurman JH. Probiotics: do they have a role in oral medicine and dentistry? *Eur J Oral Sci* 2005; 113: 188–96.
3. Meurman JH, Stamatova I. Probiotics: contributions to oral health. *Oral Dis* 2007; 13: 443–51.
4. Marsh PD. Are dental diseases examples of ecological catastrophes? *Microbiol* 2003; 149: 279–94.
5. Socransky SS, Haffajee AD. Periodontal microbial ecology. *Periodontology* 2000 2005; 38: 135–87.
6. Reid G, Jass J, Sebulsky MT, McCormick JK. Potential uses of probiotics in clinical practice. *Clin Microbiol Rev* 2003; 16: 658–66.
7. Geier MS, Butler RN, Howarth GS. Inflammatory bowel disease: current insights into pathogenesis and new therapeutic

- options; probiotics, prebiotics and synbiotics. *Int J Food Microbiol* 2007; 115: 1–11.
8. Kleessen B, Blaut M. Modulation of gut mucosal biofilms. *Br J Nutr* 2005; 93: S35–40.
 9. Marco ML, Pavan S, Kleerebezem M. Towards understanding molecular modes of probiotic action. *Curr Opin Biotechnol* 2006; 17: 204–10.
 10. Nomoto K. Prevention of infections by probiotics. *J Biosci Bioeng* 2005; 100: 583–92.
 11. Tagg JR, Dierksen KP. Bacterial replacement therapy: adapting ‘germ warfare’ to infection prevention. *Trends Biotechnol* 2003; 21: 217–23.
 12. Weng MQ, Walker WA. Bacterial colonization, probiotics, and clinical disease. *J Pediatr* 2006; 149: S107–14.
 13. Picard C, Fioramonti J, Francois A, Robinson T, Neant F, Matuchansky C. Review article: bifidobacteria as probiotic agents – physiological effects and clinical benefits. *Aliment Pharmacol Ther* 2005; 22: 495–512.
 14. Moreno MRF, Sarantinopoulos P, Tsakalidou E, De Vuyst L. The role and application of enterococci in food and health. *Int J Food Microbiol* 2006; 106: 1–24.
 15. Forchielli ML, Walker WA. The role of gut-associated lymphoid tissues and mucosal defence. *Br J Nutr* 2005; 93: S41–8.
 16. Roberfroid M. Prebiotics: the concept revisited. *J Nutr* 2007; 137: 830S–7S.
 17. Gibson GR, McCartney AL, Rastall RA. Prebiotics and resistance to gastrointestinal infections. *Br J Nutr* 2005; 93: S31–4.
 18. Reid G, Kim SO, Kohler GA. Selecting, testing and understanding probiotic microorganisms. *FEMS Immunol Med Microbiol* 2006; 46: 149–57.
 19. Salminen S, Benno Y, de Vos W. Intestinal colonisation, microbiota and future probiotics? *Asia Pac J Clin Nutr* 2006; 15: 558–62.
 20. Scholz-Ahrens KE, Ade P, Marten B, Weber P, Timm W, Asil Y, et al. Prebiotics, probiotics, and synbiotics affect mineral absorption, bone mineral content, and bone structure. *J Nutr* 2007; 137: 838S–46S.
 21. Walker WA, Goulet O, Morelli L, Antoine JM. Progress in the science of probiotics: from cellular microbiology and applied immunology to clinical nutrition. *Eur J Nutr* 2006; 45: 1–18.
 22. Lenoir-Wijnkoop I, Sanders ME, Cabana MD, Caglar E, Corthier G, Rayes N, et al. Probiotic and prebiotic influence beyond the intestinal tract. *Nutr Rev* 2007; 65: 469–89.
 23. Manley KJ, Fraenkel MB, Mayall BC, Power DA. Probiotic treatment of vancomycin-resistant enterococci: a randomised controlled trial. *Med J Aust* 2007; 186: 454–7.
 24. Steidler L, Hans W, Schotte L, Neiryck S, Obermeier F, Falk W, et al. Treatment of murine colitis by *Lactococcus lactis* secreting interleukin-10. *Science* 2000; 289: 1352–5.
 25. Vandenbroucke K, Hans W, Van Huysse J, Neiryck S, Demetter P, Remaut E, et al. Active delivery of trefoil factors by genetically modified *Lactococcus lactis* prevents and heals acute colitis in mice. *Gastroenterol* 2004; 127: 502–13.
 26. Chang TLY, Chang CH, Simpson DA, Xu Q, Martin PK, Lagenaur LA, et al. Inhibition of HIV infectivity by a natural human isolate of *Lactobacillus jensenii* engineered to express functional two-domain CD4. *Proc Natl Acad Sci USA* 2003; 100: 11672–7.
 27. Park SF, Kröll RG. Expression of listeriolysin and phosphatidylinositol-specific phospholipase-C is repressed by the plant-derived molecule cellobiose in *Listeria monocytogenes*. *Molec Microbiol* 1993; 8: 653–61.
 28. Aas JA, Paster BJ, Stokes LN, Olsen I, Dewhirst FE. Defining the normal bacterial flora of the oral cavity. *J Clin Microbiol* 2005; 43: 5721–32.
 29. Paster BJ, Olsen I, Aas JA, Dewhirst FE. The breadth of bacterial diversity in the human periodontal pocket and other oral sites. *Periodontol* 2000 2006; 42: 80–7.
 30. Aas JA, Griffen AL, Dardis SR, Lee AM, Olsen I, Dewhirst FE, et al. Bacteria of dental caries in primary and permanent teeth in children and young adults. *J Clin Microbiol* 2008; 46: 1407–17.
 31. Kolenbrander PE, Palmer RJ, Rickard AH, Jakubovics NS, Chalmers NI, Diaz PI. Bacterial interactions and successions during plaque development. *Periodontol* 2000 2006; 42: 47–79.
 32. Gibbons RJ. Bacterial adhesion to oral-tissues-a model for infectious-diseases. *J Dent Res* 1989; 68: 750–60.
 33. Mager DL, Ximenez-Fyvie LA, Haffajee AD, Socransky SS. Distribution of selected bacterial species on intraoral surfaces. *J Clin Periodontol* 2003; 30: 644–54.
 34. Beighton D, Rippon HR, Thomas HEC. The distribution of *Streptococcus mutans* serotypes and dental caries in a group of 5-year-old to 8-year-old Hampshire schoolchildren. *Br Dent J* 1987; 162: 103–6.
 35. Rudney JD, Chen R, Sedgewick GJ. *Actinobacillus actinomycetemcomitans*, *Porphyromonas gingivalis*, and *Tannerella forsythensis* are components of a polymicrobial intracellular flora within human buccal cells. *J Dent Res* 2005; 84: 59–63.
 36. Rudney JD, Chen R, Zhang G. Streptococci dominate the diverse flora within buccal cells. *J Dent Res* 2005; 84: 1165–71.
 37. Mead GC, Barrow PA. Salmonella control in poultry by competitive-exclusion or immunization. *Lett Appl Microbiol* 1990; 10: 221–7.
 38. Roos K, Hakansson EG, Holm S. Effect of recolonisation with “interfering” alpha streptococci on recurrences of acute and secretory otitis media in children: randomised placebo controlled trial. *Br Med J* 2001; 322: 210–2.
 39. Hooper LV, Falk PG, Gordon JI. Analyzing the molecular foundations of commensalism in the mouse intestine. *Curr Opin Microbiol* 2000; 3: 79–85.
 40. Freitas M, Axelsson LG, Cayuela C, Midtvedt T, Trugnan G. Microbial-host interactions specifically control the glycosylation pattern in intestinal mouse mucosa. *Histochem Cell Biol* 2002; 118: 149–61.
 41. Cebra JJ. Influences of microbiota on intestinal immune system development. *Am J Clin Nutr* 1999; 69: 1046S–51S.
 42. Neish AS, Gewirtz AT, Zeng H, Young AN, Hobert ME, Karmali V, et al. Prokaryotic regulation of epithelial responses by inhibition of I kappa B-alpha ubiquitination. *Science* 2000; 289: 1560–3.
 43. Cario E, Podolsky DK. Intestinal epithelial TOLLerance versus inTOLLerance of commensals. *Molec Immunol* 2005; 42: 887–93.
 44. Sansonetti PJ. War and peace at mucosal surfaces. *Nature Rev Immunol* 2004; 4: 953–64.
 45. Tran AX, Lester ME, Stead CM, Raetz CRH, Maskell DJ, McGrath SC, et al. Resistance to the antimicrobial peptide polymyxin requires myristoylation of *Escherichia coli* and *Salmonella typhimurium* lipid A. *J Biol Chemistry* 2005; 280: 28186–94.
 46. Kelly D, Campbell JI, King TP, Grant G, Jansson EA, Coutts AGP, et al. Commensal anaerobic gut bacteria attenuate inflammation by regulating nuclear-cytoplasmic shuttling of PPAR-gamma and RelA. *Nature Immunol* 2004; 5: 104–12.
 47. Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by

- toll-like receptors is required for intestinal homeostasis. *Cell* 2004; 118: 229–41.
48. Tien MT, Girardin SE, Regnault B, Le Bourhis L, Dillies MA, Coppee JY, et al. Anti-inflammatory effect of *Lactobacillus casei* on *Shigella*-infected human intestinal epithelial cells. *J Immunol* 2006;176:1228–37.
 49. Collier-Hyams LS, Sloane V, Batten BC, Neish AS. Cutting edge: bacterial modulation of epithelial signaling via changes in neddylation of cullin-1. *J Immunol* 2005; 175: 4194–8.
 50. Dixon DR, Reife RA, Cebra JJ, Darveau RP. Commensal bacteria influence innate status within gingival tissues: a pilot study. *J Periodontol* 2004; 75: 1486–92.
 51. Dixon DR, Bainbridge BW, Darveau RP. Modulation of the innate immune response within the periodontium. *Periodontol* 2000 2004; 35: 53–74.
 52. Hasegawa Y, Mans JJ, Mao S, Lopez MC, Baker HV, Handfield M, et al. Gingival epithelial cell transcriptional responses to commensal and opportunistic oral microbial species. *Infect Immun* 2007; 75: 2540–7.
 53. Cosseau C, Devine DA, Dullaghan E, Gardy JL, Chikhatmarla A, Gellatly S, et al. The commensal *Streptococcus salivarius* down-regulates immune responses of human epithelial cells and promotes host-microbe homeostasis. *Infect Immun* 2008; 76: 4163–75.
 54. Paster BJ, Olsen I, Aas JA, Dewhirst FE. The breadth of bacterial diversity in the human periodontal pocket and other oral sites. *Periodontology* 2000 2006; 42: 80–7.
 55. Kroes I, Lepp PW, Relman DA. Bacterial diversity within the human subgingival crevice. *Proc Natl Acad Sci USA* 1999; 96: 14547–52.
 56. Diaz PI, Chalmers NI, Rickard AH, Kong C, Milburn CL, Palmer RJ, Jr., et al. Molecular characterization of subject-specific oral microflora during initial colonization of enamel. *Appl Environ Microbiol* 2006; 72: 2837–48.
 57. Kilian M, Frandsen EVG, Haubek D, Poulsen K. The etiology of periodontal disease revisited by population genetic analysis. *Periodontol* 2000 2006; 42: 158–79.
 58. Ledder RG, Gilbert P, Huws SA, Aarons L, Ashley MP, Hull PS, et al. Molecular analysis of the subgingival microbiota in health and disease. *Appl Environ Microbiol* 2007; 73: 516–23.
 59. Kumar PS, Griffen AL, Barton JA, Paster BJ, Moeschberger ML, Leys EJ. New bacterial species associated with chronic periodontitis. *J Dent Res* 2003; 82: 338–44.
 60. Stingu C-S, Eschrich K, Rodloff AC, Schaumann R, Jentsch H. Periodontitis is associated with a loss of colonization by *Streptococcus sanguinis*. *J Med Microbiol* 2008; 57: 495–9.
 61. Tanner ACR, Milgrom PM, Kent R, Mokeem SA, Page RC, Riedy CA, et al. The microbiota of young children from tooth and tongue samples. *J Dent Res* 2002; 81: 53–7.
 62. Boyar RM, Bowden GH. The microflora associated with the progression of incipient carious lesions in teeth of children living in a water-fluoridated area. *Caries Res* 1985; 19: 298–306.
 63. Babaahmady KG, Marsh PD, Challacombe SJ, Newman HN. Variations in the predominant cultivable microflora of dental plaque at defined subsites on approximal tooth surfaces in children. *Arch Oral Biol* 1997; 42: 101–11.
 64. Beighton D. The complex oral microflora of high-risk individuals and groups and its role in the caries process. *Com Dent Oral Epidemiol* 2005; 33: 248–55.
 65. Becker MR, Paster BJ, Leys EJ, Moeschberger ML, Kenyon SG, Galvin JL, et al. Molecular analysis of bacterial species associated with childhood caries. *J Clin Microbiol* 2002; 40: 1001–9.
 66. Loesche WJ, Grossman NS. Periodontal disease as a specific, albeit chronic, infection: diagnosis and treatment. *Clin Microbiol Rev* 2001; 14: 727–52.
 67. Nishihara T, Koseki T. Microbial etiology of periodontitis. *Periodontol* 2000 2004; 36: 14–26.
 68. Sakamoto M, Umeda M, Benno Y. Molecular analysis of human oral microbiota. *J Periodontol Res* 2005; 40: 277–85.
 69. Haffajee AD, Teles RP, Socransky SS. The effect of periodontal therapy on the composition of the subgingival microbiota. *Periodontol* 2000 2006; 42: 219–58.
 70. Redding SW, Dahiya MC, Kirkpatrick WR, Coco BJ, Patterson TF, Fothergill AW, et al. *Candida glabrata* is an emerging cause of oropharyngeal candidiasis in patients receiving radiation for head and neck cancer. *Oral Surg Oral Med Oral Pathol Oral Radiol Endodont* 2004; 97: 47–52.
 71. Ramage G, Tomsett K, Wickes BL, Lopez-Ribot JL, Redding SW. Denture stomatitis: a role for *Candida* biofilms. *Oral Surg Oral Med Oral Pathol Oral Radiol Endodont* 2004; 98: 53–9.
 72. Loesche WJ, Kazor C. Microbiology and treatment of halitosis. *Periodontol* 2000 2002; 28: 256–79.
 73. Kazor CE, Mitchell PM, Lee AM, Stokes LN, Loesche WJ, Dewhirst FE, et al. Diversity of bacterial populations on the tongue dorsa of patients with halitosis and healthy patients. *J Clin Microbiol* 2003; 41: 558–63.
 74. Hillman JD. Genetically modified *Streptococcus mutans* for the prevention of dental caries. *Antonie Van Leeuwenhoek* 2002; 82: 361–6.
 75. Comelli EM, Guggenheim B, Stingle F, Nesser JR. Selection of dairy bacterial strains as probiotics for oral health. *Eur J Oral Sci* 2002; 110: 218–24.
 76. Haukioja A, Yli-Knuutila H, Loimaranta V, Kari K, Ouweland AC, Meurman JH, et al. Oral adhesion and survival of probiotic and other lactobacilli and bifidobacteria in vitro. *Oral Microbiol Immunol* 2006; 21: 326–32.
 77. Petti S, Tarsitani G, Simmonetti D'Arca A. Antibacterial activity of yoghurt against viridans streptococci in vitro. *Arch Oral Biol* 2008; 53: 985–90.
 78. Chung J, Ha ES, Park HR, Kim S. Isolation and characterization of *Lactobacillus* species inhibiting the formation of *Streptococcus mutans* biofilm. *Oral Microbiol Immunol* 2004; 19: 214–6.
 79. Simark-Mattsson C, Emilson CG, Hakansson EG, Jacobsson C, Roos K, Holm S. Lactobacillus-mediated interference of mutans streptococci in caries-free vs. caries-active subjects. *Eur J Oral Sci* 2007; 115: 308–14.
 80. Strahinic I, Busarcevic M, Pavlica D, Milasin J, Golic N, Topisirovic L. Molecular and biochemical characterizations of human oral lactobacilli as putative probiotic candidates. *Oral Microbiol Immunol* 2007; 22: 111–7.
 81. Koll P, Mandar R, Marcotte H, Leibur E, Mikelsaar M, Hammarstro L. Characterization of oral lactobacilli as potential probiotics for oral health. *Oral Microbiol Immunol* 2008; 23: 139–47.
 82. Clancy KA, Pearson S, Bowen WH, Burne RA. Characterization of recombinant, ureolytic *Streptococcus mutans* demonstrates an inverse relationship between dental plaque ureolytic capacity and cariogenicity. *Infect Immun* 2000; 68: 2621–9.
 83. Kruger C, Hu YZ, Pan Q, Marcotte H, Hultberg A, Delwar D, et al. In situ delivery of passive immunity by lactobacilli producing single-chain antibodies. *Nature Biotech* 2002; 20: 702–6.
 84. Nase L, Hatakka K, Savilahti E, Saxelin M, Ponka A, Poussa T, et al. Effect of long-term consumption of a probiotic bacterium, *Lactobacillus rhamnosus* GG, in milk on dental caries and caries risk in children. *Caries Res* 2001; 35: 412–20.
 85. Ahola AJ, Yli-Knuutila H, Suomalainen T, Poussa T, Ahlstrom A, Meurman JH, et al. Short-term consumption of probiotic-containing cheese and its effect on dental caries risk factors. *Arch Oral Biol* 2002; 47: 799–804.

86. Nikawa H, Makihira S, Fukushima H, et al. *Lactobacillus reuteri* in bovine milk fermented decreases the oral carriage of mutans streptococci. *Int J Food Microbiol* 2004; 95: 219–23.
87. Caglar E, Cildir SK, Ergeneli S, Sandalli N, Twetman S. Salivary mutans streptococci and lactobacilli levels after ingestion of the probiotic bacterium *Lactobacillus reuteri* ATCC 55730 by straws or tablets. *Acta Odontol Scand* 2006; 64: 314–8.
88. Caglar E, Kavaloglu SC, Kuscü OO, Sandalli N, Holgerson PL, Twetman S. Effect of chewing gums containing xylitol or probiotic bacteria on salivary mutans streptococci and lactobacilli. *Clin Oral Investig* 2007; 11: 425–9.
89. Caglar E, Kuscü OO, Cildir SK, Kuvvetli SS, Sandalli N. A probiotic lozenge administered medical device and its effect on salivary mutans streptococci and lactobacilli. *Int J Paediatr Dent* 2008; 18: 35–9.
90. Caglar E, Sandallii N, Twetman S, Kavaloglu S, Ergeneli S, Selvi S. Effect of yogurt with *Bifidobacterium* DN-173 010 on salivary mutans streptococci and lactobacilli in young adults. *Acta Odontol Scand* 2005; 63: 317–20.
91. Caglar E, Kuscü OO, Kuvvetli SS, Cildir SK, Sandalli N, Twetman S. Short-term effect of ice-cream containing *Bifidobacterium lactis* Bb-12 on the number of salivary mutans streptococci and lactobacilli. *Acta Odontol Scand* 2008; 66: 154–8.
92. Petti S, Tarsitani G, D'Arca AS. A randomized clinical trial of the effect of yoghurt on the human salivary microflora. *Arch Oral Biol* 2001; 46: 705–12.
93. Krasse P, Carlsson B, Dahl C, Paulsson A, Nilsson A, Sinkiewicz G. Decreased gum bleeding and reduced gingivitis by the probiotic *Lactobacillus reuteri*. *Swed Dent J* 2006; 30: 55–60.
94. Shimauchi H, Mayanagi G, Nakaya S, Minamibuchi M, Ito Y, Yamaki K, et al. Improvement of periodontal condition by probiotics with *Lactobacillus salivarius* WB21: a randomized, double-blind, placebo-controlled study. *J Clin Periodontol* 2008; 35: 897–905.
95. Sookkhee S, Chulasiri M, Prachyabrued W. Lactic acid bacteria from healthy oral cavity of Thai volunteers: inhibition of oral pathogens. *J Appl Microbiol* 2001; 90: 172–9.
96. Ishikawa H, Aiba Y, Nakanishi M, Oh-Hashi Y, Koga Y. Suppression of periodontal pathogenic bacteria by the administration of *Lactobacillus salivarius* T12711. *J Jap Soc Periodontol* 2003; 45: 105–12.
97. Koll-Klais P, Mandar R, Leibur E, Marcotte H, Hammarstrom L, Mikelsaar M. Oral lactobacilli in chronic periodontitis and periodontal health: species composition and antimicrobial activity. *Oral Microbiol Immunol* 2005; 20: 354–61.
98. Hojo K, Mizoguchi C, Taketomo N, Ohshima T, Gomi K, Arai T, et al. Distribution of salivary *Lactobacillus* and *Bifidobacterium* species in periodontal health and disease. *Biosci Biotech Biochem* 2007; 71: 152–7.
99. Teughels W, Newman MG, Coucke W, Haffajee AD, van der Mei HC, Haake SK, et al. Guiding periodontal pocket recolonization: a proof of concept. *J Dent Res* 2007; 86: 1078–82.
100. Nackaerts O, Jacobs R, Quirynen M, Rober M, Sun Y, Teughels W. Replacement therapy for periodontitis: pilot radiographic evaluation in a dog model. *J Clin Periodontol* 2008; 35: 1048–52.
101. Pangsomboon K, Kaewnopparat S, Pitakpornpreecha T, Srichana T. Antibacterial activity of a bacteriocin from *Lactobacillus paracasei* HL32 against *Porphyromonas gingivalis*. *Arch Oral Biol* 2006; 51: 784–93.
102. Hatakka K, Ahola AJ, Yli-Knuutila H, Richardson M, Poussa T, Meurman JH, et al. Probiotics reduce the prevalence of oral Candida in the elderly-A randomized controlled trial. *J Dent Res* 2007; 86: 125–30.
103. Kang MS, Kim BG, Chung J, Lee HC, Oh JS. Inhibitory effect of *Weissella cibaria* isolates on the production of volatile sulphur compounds. *J Clin Periodontol* 2006; 33: 226–32.
104. Persson S, Edlund MB, Claesson R, Carlsson J. The formation of hydrogen-sulfide and methyl mercaptan by oral bacteria. *Oral Microbiol Immunol* 1990; 5: 195–201.
105. Burton JP, Chilcott CN, Moore CJ, Speiser G, Tagg JR. A preliminary study of the effect of probiotic *Streptococcus salivarius* K12 on oral malodour parameters. *J Appl Microbiol* 2006; 100: 754–64.
106. Wescombe PA, Upton M, Dierksen KP, Ragland NL, Sivabalan S, Wirawan RE, et al. Production of the lantibiotic salivaricin A and its variants by oral streptococci and use of a specific induction assay to detect their presence in human saliva. *Appl Environ Microbiol* 2006; 72: 1459–66.
107. Dierksen KP, Moore CJ, Inglis MA, Wescombe PA, Tagg JR. The effect of ingestion of milk supplemented with salivaricin A-producing *Streptococcus salivarius* on the bacteriocin-like activity of the streptococcal population on the tongue. *FEMS Microbiol Ecol* 2006; 59: 584–91.
108. Sterer N, Rosenberg M. *Streptococcus salivarius* promotes mucin putrefaction and malodor production by *Porphyromonas gingivalis*. *J Dent Res* 2006; 85: 910–4.
109. Mostefaoui Y, Bart C, Frenette M, Rouabhia M. *Candida albicans* and *Streptococcus salivarius* modulate IL-6, IL-8, and TNF-alpha expression and secretion by engineered human oral mucosa cells. *Cell Microbiol* 2004; 6: 1085–96.
110. Busscher HJ, Mulder A, van der Mei HC. In vitro adhesion to enamel and in vivo colonization of tooth surfaces by lactobacilli from a bio-yoghurt. *Caries Res* 1999; 33: 403–4.
111. Yli-Knuutila H, Snall J, Kari K, Meurman JH. Colonization of *Lactobacillus rhamnosus* GG in the oral cavity. *Oral Microbiol Immunol* 2006; 21: 129–31.
112. de Soet JJ, Nyvad B, Kilian M. Strain-related acid production by oral streptococci. *Caries Res* 2000; 34: 486–90.
113. Marsh PD. Microbial ecology of dental plaque and its significance in health and disease. *Adv Dent Res* 1994; 8: 263–71.
114. Marsh PD. Plaque as a biofilm: pharmacological principles of drug delivery and action in the sub- and supragingival environment. *Oral Dis* 2003; 9: 16–22.
115. Beighton D, Smith K, Hayday H. The growth of bacteria and the production of exoglycosidic enzymes in the dental plaque of macaque monkeys. *Arch Oral Biol* 1986; 31: 829–35.
116. Teixeira EH, Napimoga MH, Carneiro VA, de Oliveira TM, Nascimento KS, Nagano CS, et al. In vitro inhibition of oral streptococci binding to the acquired pellicle by algal lectins. *J Appl Microbiol* 2007; 103: 1001–6.
117. Yamanaka A, Kimizuka R, Kato T, Okuda K. Inhibitory effects of cranberry juice on attachment of oral streptococci and biofilm formation. *Oral Microbiol Immunol* 2004; 19: 150–4.
118. Percival RS, Devine DA, Duggal MS, Chartron S, Marsh PD. The effect of cocoa polyphenols on the growth, metabolism, and biofilm formation by *Streptococcus mutans* and *Streptococcus sanguinis*. *Eur J Oral Sci* 2006; 114: 343–8.
119. Boyle RJ, Robins-Browne RM, Tang MLK. Probiotic use in clinical practice: what are the risks? *Am J Clin Nutr* 2006; 83: 1256–64.
120. Reid G. Safe and efficacious probiotics: what are they? *Trends Microbiol* 2006; 14: 348–52.

121. Snyderman DR. The safety of probiotics. Workshop on scientific and regulatory challenges of development of probiotics as foods and drugs. Adelphi, MD: University of Chicago Press; 2006. pp. S104–11.
122. Stamatova I, Meurman JH, Kari K, Tervahartiala T, Sorsa T, Baltadjieva M. Safety issues of *Lactobacillus bulgaricus* with respect to human gelatinases in vitro. FEMS Immunol Med Microbiol 2007; 51: 194–200.
123. Husni RN, Gordon SM, Washington JA, Longworth DL. *Lactobacillus* bacteremia and endocarditis: Review of 45 cases. Clin Infect Dis 1997; 25: 1048–55.
124. Salminen MK, Rautelin H, Tynkkynen S, Poussa T, Saxelin M, Valtonen V, et al. Lactobacillus bacteremia, clinical significance, and patient outcome, with special focus on probiotic *L. rhamnosus* GG. Clin Infect Dis 2004; 38: 62–9.
125. De Groote MA, Frank DN, Dowell E, Glode MP, Pace NR. *Lactobacillus rhamnosus* GG bacteremia associated with probiotic use in a child with short gut syndrome. Pediatr Infect Dis J 2005; 24: 278–80.
126. Mackay AD, Taylor MB, Kibbler CC, Hamilton-Miller JM. *Lactobacillus* endocarditis caused by a probiotic organism. Clin Microbiol Inf 1999; 5: 290–2.
127. Salminen MK, Tynkkynen S, Rautelin H, Saxelin M, Vaara M, Ruutu P, et al. *Lactobacillus* bacteremia during a rapid increase in probiotic use of *Lactobacillus rhamnosus* GG in Finland. Clin Infect Dis 2002; 35: 1155–60.
128. Matsumoto M, Tsuji M, Sasaki H, Fujita K, Nomura R, Nakano K, et al. Cariogenicity of the probiotic bacterium *Lactobacillus salivarius* in rats. Caries Res 2005; 39: 479–83.
129. Haukioja A, Soderling E, Tenovuo J. Acid production from sugars and sugar alcohols by probiotic lactobacilli and bifidobacteria in vitro. Caries Res 2008; 42: 449–53.
130. Montalto M, Vastola M, Marigo L, Covino M, Graziosetto R, Curigliano V, et al. Probiotic treatment increases salivary counts of lactobacilli: a double-blind, randomized, controlled study. Digestion 2004; 69: 53–6.
131. Hillman JD, Socransky SS, Shivers M. The relationships between streptococcal species and periodontopathic bacteria in human dental plaque. Arch Oral Biol 1985; 30: 791–5.

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