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Dissecting conformational contributions to glycosidase catalysis and inhibition

Gaetano Speciale¹, Andrew J Thompson², Gideon J Davies² and Spencer J Williams¹



Glycoside hydrolases (GHs) are classified into >100 sequence-based families. These enzymes process a wide variety of complex carbohydrates with varying stereochemistry at the anomeric and other ring positions. The shapes that these sugars adopt upon binding to their cognate GHs, and the conformational changes that occur along the catalysis reaction coordinate is termed the conformational itinerary. Efforts to define the conformational itineraries of GHs have focussed upon the critical points of the reaction: substrate-bound (Michaelis), transition state, intermediate (if relevant) and product-bound. Recent approaches to defining conformational itineraries that marry X-ray crystallography of enzymes bound to ligands that mimic the critical points, along with advanced computational methods and kinetic isotope effects are discussed.

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Edited by **Harry J Gilbert** and **Harry Brumer**

For a complete overview see the [Issue](#) and the [Editorial](#)

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Glycoside hydrolases catalyze the hydrolytic cleavage of the glycosidic bond. They are enzymes of enduring interest owing to the ubiquity of carbohydrates in nature and their importance in human health and disease, the food, detergent, oil & gas and biotechnology industries. Glycoside hydrolases generally, but not quite exclusively, perform catalysis with a net retention or inversion of anomeric stereochemistry. The gross mechanisms of glycosidases were postulated by Koshland in 1953 [1^{••}], and his prescient insights remain largely true to this day. The glycoside hydrolases are an immensely varied group of enzymes and are usefully classified on the basis of sequence according to the CAZy system (www.cazy.org; see also Cazypedia: www.cazypedia.org), which reveals a growing and formidable diversity of proteins (133 families

as of 2014) [2]. What continues to occupy the attention of mechanistic enzymologists is a complete description of the fine details of the overall reaction coordinate. The free energy profile of catalysis is a composite of terms including: bond-making and breaking; the establishment and disbandment of stereoelectronic effects; and conformational effects. Conformational interactions include substrate-based: vicinal (e.g. eclipsing, gauche, $\Delta 2$), 1,3-diaxial, and 1,4-bridgehead; and enzyme-based: local and global conformational changes of the enzyme that occur on the time-scale of catalysis [3].

Two major areas of inquiry are active in the area of conformation and glycoside hydrolases:

1. What are the conformational changes that occur during catalysis upon substrate binding, at the transition state(s), intermediates (if relevant), and product? Aside from the elemental interest in this question, there is the potential for utilizing this information to develop glycosidase inhibitors that take advantage of the considerable amounts of energy used to selectively bind the transition state (for a glycosidase with a catalytic rate enhancement of 10^{17} , the calculated transition state affinity is 10^{-22} M [4]), with the enticing possibility that differences in transition state conformation may allow the development of glycosidase-selective inhibitors.
2. Once transition-state structural information is acquired and used to inspire inhibitor development, do the resulting inhibitors actually bind by utilizing the same interactions that are used to stabilize the transition state — that is, are they genuine transition state mimics? The answers to this question speak to our abilities to realize this unique form of rational inhibitor design.

In this review we cover recent developments in the understanding of conformational reaction coordinates and how such information is acquired; and what constitutes good transition state mimicry by inhibitors. This work extends two recent comprehensive reviews [5,6[•]].

Contortions along the reaction coordinate

Substantial evidence has accrued that retaining and inverting glycoside hydrolases perform catalysis through an oxocarbenium ion-like transition state with significant bond breakage to the departing group and limited bond formation to the attacking nucleophile (Figure 1a) [7]. On

the basis of the four idealized half-chair and boat conformations expected for the transition state (see [Side Panel A](#)), four ‘classical’ conformational itineraries may be identified ([Figure 1b](#)). In these simplified presentations, it is apparent that C1 scribes an arc along the conformational reaction coordinate as it undergoes an electrophilic migration from the leaving group to a nucleophile. However, other ring atoms also change positions, in particular O5 and C2. The subtle change in the

position of O5 has little mechanistic consequence other than to allow development of the partial double bond. Interactions at C2 are usually (but not always, see: [8]) significant and for the β -glucosidase Abg from *Agrobacterium* sp. or for α -glucosidase of *Saccharomyces cerevisiae* [9] have been shown to contribute 18–22 kJ mol⁻¹ to transition state stabilization [10], highlighting that the repositioning of C2 and its substituent and other electronic changes accompanying formation of the oxocarbenium

Side panel A Theoretical considerations of the transition state conformation

It is a stereoelectronic requirement that the development of a partial double bond between O5 and C1 results in flattening of the system C2–C1–O5–C5, with the remaining pyranose C3 and C4 atoms having freedom to move [11]. The possible idealized transition state structures that satisfy these requirements are ³H₄, ⁴H₃, ^{2,5}B and ^B_{2,5} (and the closely related but usually higher energy ⁴E, ³E, ^E₄ and ^E₃). As the oxocarbenium ion-like transition states of glycosidases are ‘central’ transition states it is appropriate to invoke the principle of least nuclear motion, which states

that elementary reactions that involve the least change in atomic position and electronic configuration will be favoured [12,13]. Accordingly, the conformational reaction coordinate will most likely involve ground state conformations (corresponding to Michaelis, intermediate and product complexes) that are close neighbors to the transition state conformations. The conformational relationships of pyranose rings [14] may be conveniently summarized using a Cremer-Pople sphere [15] or its equivalent Mercator and polar projections ([Figure 1](#)).

Figure 1

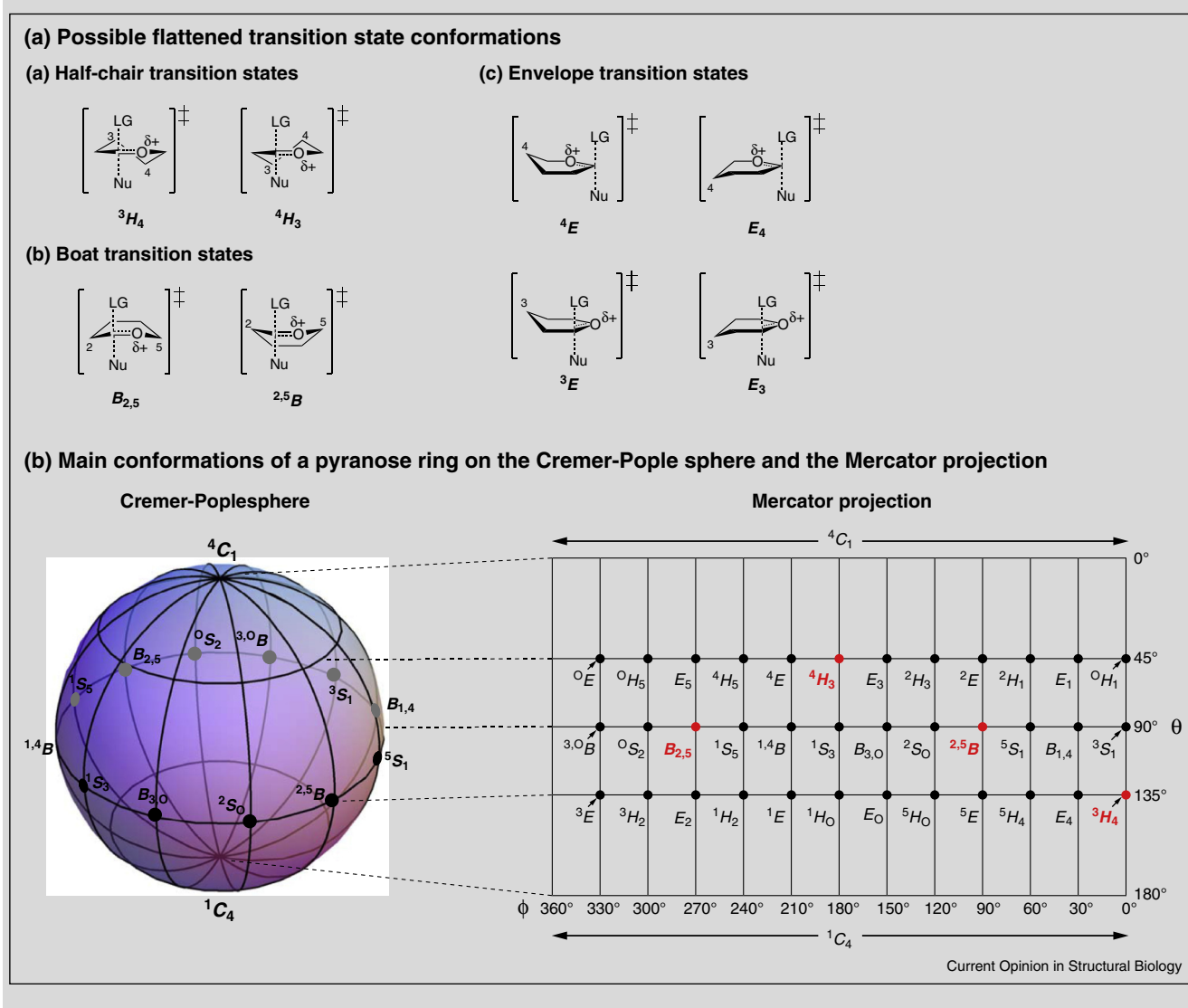
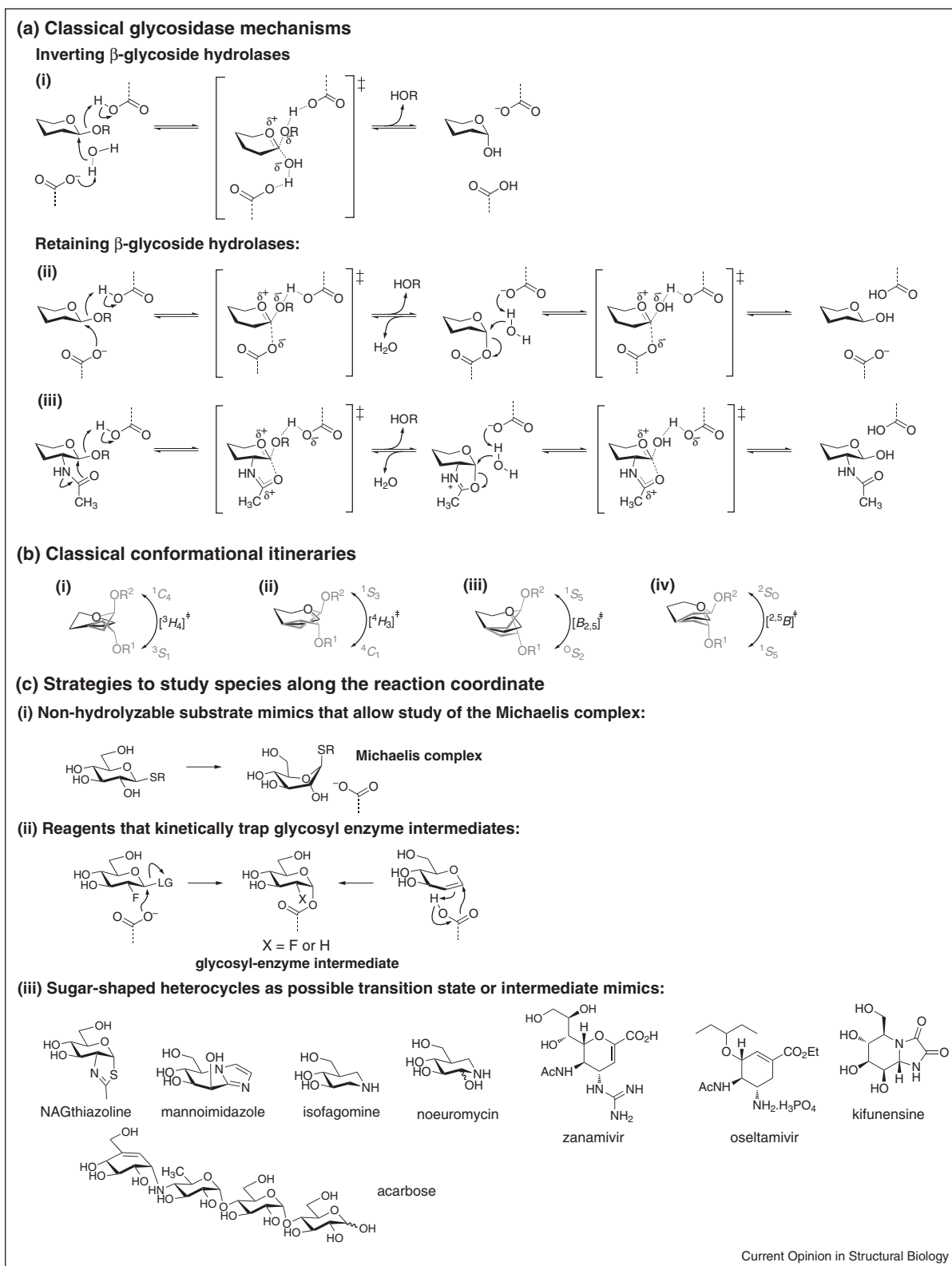


Figure 1



(a) Mechanisms of classical (i) inverting and retaining glycosidases that utilize (ii) an enzymic nucleophile or (iii) substrate-assisted catalysis. **(b)** Classical conformational itineraries around planar, oxocarbenium ion-like transition states in (i,ii) half-chair (*H*) or (iii,iv) boat (*B*) conformations. **(c)** Strategies and reagents used to study key species along the reaction coordinate.

ion-like transition state can provide substantial amounts of stabilization energy. The ground state conformations and those of intermediates and transition states need not sit squarely on the graticules of the major meridians and latitudes but may be located within the conformational space nearby (see [Side Panel A](#)).

Powerful computing resources allow the calculation of the energy of every possible conformation of individual sugars providing a so-called free energy landscape (FEL). Each carbohydrate stereoisomer possesses a unique FEL, owing to the presence of various substituents, the resulting hydrogen-bonding interactions, local steric interactions, and the contribution of the anomeric effect. This computational approach was first applied to β -glucopyranose and 9 energetic minima were identified [16]. Aside from the global minimum of 4C_1 , the remaining 8 local minima were approximate *B* and *S* conformations; however these differed from the canonical *B* and *S* conformations owing to the lack of rigidity of the ring, and the presence of attractive (hydrogen bonding) and repulsive (eclipsing and 1,3-diaxial) interactions. Several of these conformations were identified as pre-activated for catalysis, with pseudo-axial C1–O1 bonds leading to a lengthening of the C1–O1 bond and a shortening of the O5–C1 bond owing to the developing anomeric effect, and partial charge at C1 (see [Side Panel B](#)). Enticingly, the pre-activated conformations are those that are most frequently observed in so-called Michaelis complexes (representing E.S complexes) studied by X-ray crystallography for GHs that process β -glucosides. FEL analysis has been extended to include α -glucopyranose, β -xylopyranose, β -mannopyranose, and β -*N*-acetylglucopyranosamine [17], and α -mannopyranose

Side panel B On substrate distortion and Michaelis complexes

The concept of substrate distortion upon binding to an enzyme in the enzyme-substrate (Michaelis) complex has long been proposed since the earliest X-ray crystallographic data of glycosidases became available. Four major principles are invoked to justify the need for a substrate to distort from a ground-state and less reactive conformation to a distorted and typically higher energy conformation: (1) the principle of least nuclear motion, which favours distorted conformations that require less nuclear movement to achieve the transition state (see [Side Panel A](#)); (2) the geometric demand that nucleophilic attack on the anomeric centre needs to be 'in line' with the departing nucleophile, which arises from the stereoelectronic requirement that the electrons derived from the nucleophile will populate the σ^* orbital of the glycosidic bond; (3) the stereoelectronic requirement for development of a partial double bond at the oxocarbenium ion-like transition state, which requires electron donation by a suitably located n-type lone pair on the endocyclic oxygen; and (4) the fact that general acid catalytic residues of glycosidases are located within the plane of the ring, either syn or anti to the C1–O5 bond [20]. Although often invoked as a rationale for substrate distortion, Sinnott has persuasively argued that the 'antiperiplanar lone pair hypothesis', viz. that sp^3 lone pairs of electrons on a heteroatom direct the departure of a leaving group from an adjacent tetrahedral carbon centre, requires implausible contortions of the pyranose ring [21].

[18**], with the data supporting the conclusion that for these sugars the catalytically relevant conformations are frequently the energetically predisposed distorted structures. These observations are harmonious with the suggestion of Wolfenden that the fact that many enzymes achieve rates approaching the diffusion limit suggests that they have evolved to recognize species that are reasonably populous in solution [19].

Defining the conformational coordinate: Is seeing believing?

X-ray crystallography is a powerful technique that provides a detailed molecular description of the catalytic machinery of an enzyme. While unliganded (apo) structures provide some information that can be used to help understand mechanism, the acquisition of complexes, with substrates (or substrate analogues), sugar-shaped inhibitors, mechanism-based inhibitors, or products have the potential to reveal intricate details of the amino acid residues involved in catalysis and the conformations of enzyme-bound species ([Figure 1c](#)). Three main strategies for the acquisition of Michaelis (E.S)-like complexes of retaining and inverting glycosidases are: firstly, co-crystallization of non-hydrolyzable substrate mimics with wild-type enzyme ([Figure 1c\(i\)](#)) [22], or secondly, co-crystallization of substrate with catalytically inactive mutant enzymes, or thirdly, co-crystallization of substrate and wildtype enzyme at a pH at which it is inactive. Occasionally, Michaelis complexes have been obtained serendipitously at pH values under which the enzyme is active; the reason for lack of hydrolysis in these cases is unclear [23,24].

For retaining glycoside hydrolases that proceed through a glycosyl enzyme intermediate, fairly effective methods have been developed to allow the trapping of kinetically competent intermediates [25]; the general principle is to rapidly access the glycosyl enzyme intermediate using a good leaving group, but to modify the sugar such that its turnover to product is slowed, allowing its accumulation and study. The initial work by Legler involved addition to glycals to generate 2-deoxyglycosyl enzymes [26] or use of aryl 2-deoxyglycosides [27], and was elegantly extended by Withers to 2-deoxy-2-fluoro-, 2-deoxy-2,2-difluoro- and 5-fluoro glycosyl fluorides (and closely related 2,4-dinitrophenyl, and other activated glycosides) [25] ([Figure 1c\(ii\)](#)). For reasons that are not entirely clear, for α -glycosidases the use of C5-inverted 5-fluoro-glycosyl fluorides for the corresponding enzymes usually yield better trapping results than for the stereochemically-matched alternative. For retaining enzymes that proceed by anchimeric assistance from a 2-acylamido group, sulfur mimics of the proposed oxazoline (or oxazolinium ion) intermediate, most notably NAGthiazoline [28,29], have proved effective as inhibitors and informative as mechanistic probes for crystallographically studying the intermediate ([Figure 1c\(iii\)](#)).

It is important to recognize that all X-ray structures of protein–ligand complexes are by their very nature not catalytically competent and thus care must be taken in how to interpret the important clues they provide in the proposal of a credible conformational itinerary. Kinetically trapped species recapitulate the major bond-forming and breaking events but the structural modifications made to allow kinetic trapping may perturb substrate interactions that are important for defining the conformational reaction coordinate. Occasionally, crystallization efforts at non-optimal pH have led to the acquisition of apparently bonafide Michaelis complexes; however even these constitute complexes with catalytically-incompetent enzymes and the interpretation of these structures must recognize that these do not lie on the reaction coordinate. Complexes with substrate and enzyme may be ‘pre-Michaelis’ complexes that represent enzyme-bound species that precede the formation of the true Michaelis complex, or may be catalytically irrelevant species that are actively misleading. Product bound to enzyme may have relaxed from its first formed conformation as the lack of a sizeable anomeric substituent prevents the enzyme from utilizing +1 subsite interactions to stabilize its conformation. Nonetheless, with some exceptions, most pseudo-Michaelis, glycosyl enzyme intermediate or thiazoline intermediate, and product complexes are sufficiently akin to hypothetical bonafide species on the reaction coordinate to allow cautious but probably reasonable insights into mechanism.

Caution must also be exercised when studying complexes with sugar-shaped heterocycles that function as competitive inhibitors (Figure 1c(iii)). While superficially these compounds resemble aspects of the proposed transition state, there are intrinsic limitations of what can be mimicked in a chemically stable compound, including hybridization changes, partial charge development, and fractional bond orders. A study of the FELs of two inhibitors that display superficial transition state mimicry: isofagomine and mannoimidazole revealed dramatic differences (Figure 2a) [30^{••}]. Isofagomine is strongly biased toward a 4C_1 conformation, with potential transition state mimicking 4H_3 and $B_{2,5}$ conformations lying 12 and 8 kcal mol⁻¹ higher, respectively, and importantly with a significant barrier to attaining those conformations. On the other hand while mannoimidazole prefers 4H_3 and 3H_4 conformations (with a 1 kcal mol⁻¹ preference for the latter), the $B_{2,5}$ conformation is also energetically accessible. Overlaying the observed conformations of isofagomine-type and mannoimidazole-type inhibitors from X-ray structures with mannose-processing enzymes of various GH families reveals all isofagomine complexes adopt a 4C_1 conformation, whereas for mannoimidazole the $B_{2,5}$ conformation is observed on enzymes of families GH2, 26, 38, 92 and 113, implying a ${}^1S_5 \rightarrow B_{2,5}^\ddagger \rightarrow {}^0S_2$ conformational itinerary. One interesting footnote is that

non-ground state conformations of isofagomine-type inhibitors have been observed on family GH6 cellulases in either ${}^{2,5}B/{}^2S_0$ or 2S_0 conformations [31,32]. The energetic difficulties in attaining such conformations highlight their special significance when seen and in these cases they reflect the proposed ${}^2S_0 \rightarrow {}^{2,5}B^\ddagger \rightarrow {}^5S_1$ itinerary.

While the FEL of an isolated carbohydrate is often biased toward those conformations pre-activated for catalysis, further substrate distortion presumably occurs upon binding to enzyme. Quantum mechanics/molecular mechanics calculations of α -mannopyranose revealed that a FEL determined within the constraints of a GH47 α -mannosidase is moulded by the enzyme to dramatically limit the conformations accessible by the substrate to a previously inaccessible region of the FEL for the substrate off-enzyme (Figure 2b) [18^{••}]. In support of the theoretical predictions, X-ray analysis of ‘snapshot’ complexes of the enzyme with a substrate analogue, transition state, and product mimics supported a ${}^3S_1 \rightarrow {}^3H_4^\ddagger \rightarrow {}^1C_4$ conformational itinerary predicted on the basis of the FEL remodelling.

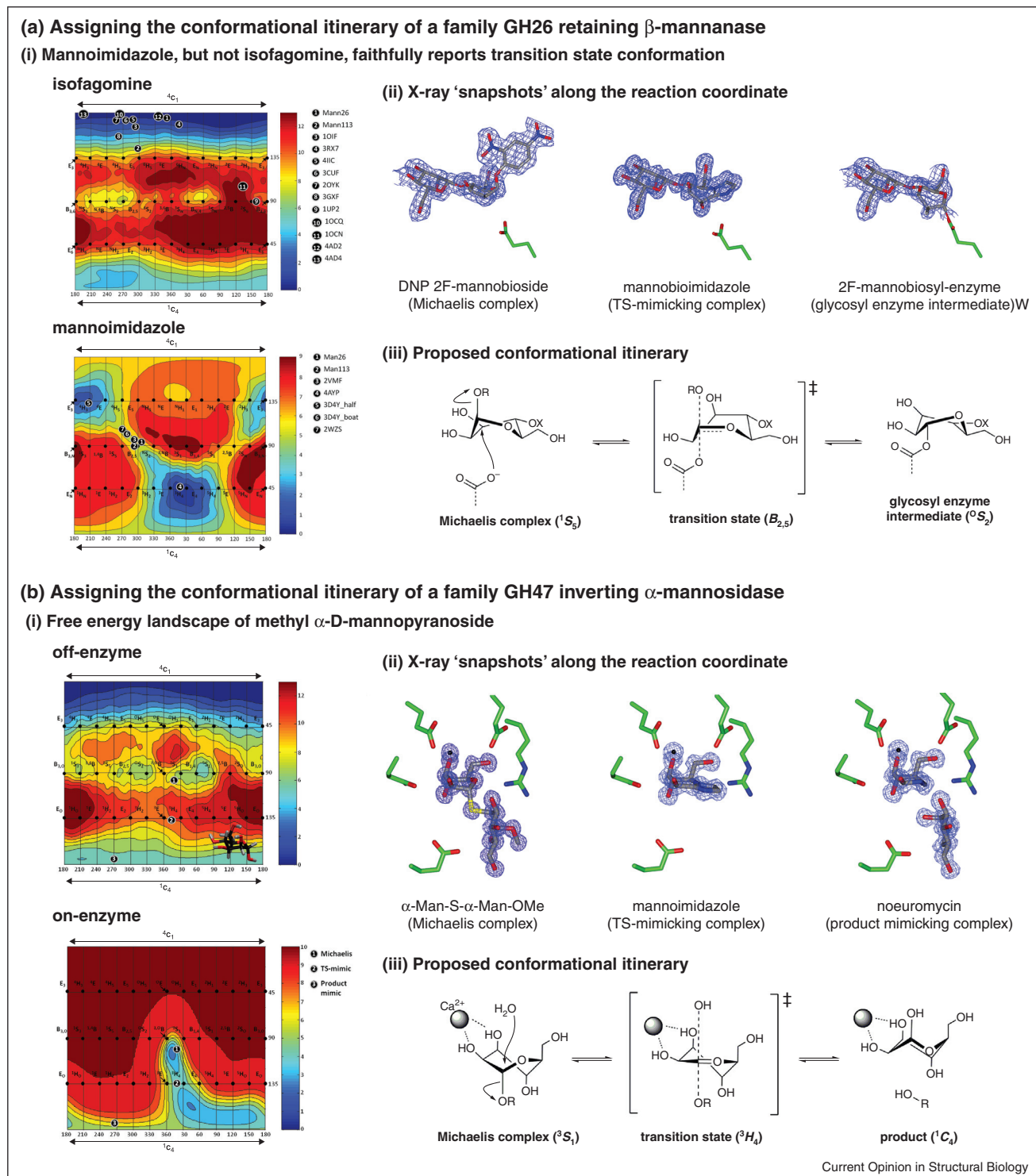
Sugars getting into shape: News dispatches from the families

Table 1 summarizes well-defined conformational itineraries for a range of GH enzymes (see Supporting Information for a more detailed listing). We present a few highlights from the last two years that are not covered in detail elsewhere.

β -Hexosaminidases of family GH3 perform catalysis through a two step mechanism with the initial substitution step occurring by an enzymatic nucleophile to afford a glycosyl enzyme intermediate (Figure 1a(ii)). Insight into the reaction coordinate has been obtained through trapping a glycosyl enzyme intermediate in a 4C_1 conformation using the mechanism based inhibitor 5F-GlcNAcF, suggesting a ${}^1S_3 \rightarrow {}^4H_3^\ddagger \rightarrow {}^4C_1$ itinerary [33[•]]. Interestingly, a complex with an inactive mutant with substrate, and of wild-type with product, also revealed 4C_1 conformations. In this case there is strong evidence that the complex with substrate is not a bonafide Michaelis complex as a loop containing the putative histidine general acid/base undergoes a dramatic movement. In the product complex it appears that the sugar has relaxed to a more stable conformation.

Family GH39 α -L-iduronidase (IDUA) is a retaining lysosomal enzyme that assists in the stepwise degradation of heparin sulfate and dermatan sulfate, and which is of interest for enzyme replacement therapy of the associated lysosomal storage disorder (LSD) mucopolysaccharidosis type I [34[•]]. Michaelis complexes with iduronate analogues in 2S_0 conformations, and the trapping of a glycosyl enzyme on IDUA in a ${}^5S_1/{}^{2,5}B$ conformation using 2-deoxy-2-fluoro- α -L-idopyranosyluronic

Figure 2



Computational studies, in concert with X-ray crystallography and inhibitor design and synthesis, assist in assigning conformational itineraries. **(a)** Assigning the conformational itinerary of *Cellvibrio japonicus* GH26 β -mannanase Man26C. (i) Free energy landscapes reveal mannoimidazole, unlike isofagomine, is able to attain the conformations relevant to glycosidase catalysis; (ii) X-ray structures of a Michaelis complex (1GVY), glycosyl enzyme intermediate (1GW1), and transition state mimicking β -mannosyl-1,4-mannoimidazole complex (4CD5); (iii) proposed conformational itinerary. **(b)** Assigning the conformational itinerary of *Caulobacter* strain K31 GH47 α -mannosidase. (i) Free energy landscapes highlight substrate preactivation off-enzyme, and reshaping of the available conformations on-enzyme; (ii) X-ray structures of Michaelis complex (4AYP), transition state mimicking mannoimidazole complex (4AYQ), and product mimicking noeuromycin complex (4AYR); (iii) proposed conformational itinerary.

Table 1

Conformational itineraries around various transition state conformations. Listed are families for which strong evidence in support of a conformational itinerary is available. For a more comprehensive listing see Supporting Information Table S1

Transition state conformation	GH families	Configuration of substrate	Enzymatic activities
${}^3H_4^\ddagger$ (${}^4H_5^\ddagger$ for sialidases)	29, 33, 34, 47	L-fuco (=L-galacto) sialic acid, D-manno	α -Fucosidase α -Mannosidase
${}^4H_3^\ddagger$	1, 2, 3, 5, 7, 10, 12, 16, 20, 22, 26, 27, 30, 84	D-gluco/D-manno D-galacto D-gluco D-xylo D-gluco/galacto	β -Glucosidase/cellulase/lichenases/ b-mannosidase β -Galactosidase α -Glucosidase/ α -glucanase β -Xylosidase/xylanase β -Hexosaminidase
${}^{2,5}B^\ddagger$	6, 8, 11	D-gluco D-xylo	β -Glucosidase, chitinase Xylanase
$B_{2,5}^\ddagger$	2, 26, 38, 92, 113	D-manno	β -Mannosidase, β -mannanase, α -mannosidase
${}^4E^\ddagger$	117	3,6-anhydro-L-galacto	α -1,3-L-Neogaroobiase

acid fluoride, imply a ${}^2S_O \rightarrow {}^{2,5}B^\ddagger \rightarrow {}^5S_1$ conformational itinerary (Figure 3a) [35**].

Family GH59 β -galactocerebrosidase (GALC) degrades glycosphingolipids and its deficiency leads to another LSD, Krabbe disease. X-ray ‘snapshots’ of a Michaelis complex (using 4-nitrophenyl β -D-galactopyranoside), a 2-deoxy glycosyl enzyme (from galactal addition), and product (with galactose) showed the sugar ring in a 4C_1 conformation in all structures [36*]. This surprising result, reminiscent of that seen with the GH2 β -galactosidase LacZ [37], was interpreted as suggesting that no distortion of the ring occurs along the reaction coordinate. Interestingly, the acid/base residue in the substrate complex is incorrectly positioned suggesting that this is not a Michaelis complex; additionally, the product complex may have relaxed from its first-formed conformation.

Family GH117 α -1,3-L-neogaroobiase has been described as a keystone enzyme owing to its role in agarose degradation, which provides the capability for the human gut microbiota to degrade seaweed diets [38**]. These interesting (probably inverting) enzymes act on 3,6-anhydro- α -L-galactosides (Figure 3b). The 3,6-anhydro bridge imparts significant rigidity on the sugar to prefer 4C_1 and ${}^{1,4}B$ conformations. A Michaelis complex of an inactive mutant with neogaroobiase highlighted a histidine residue as a potential catalytic general acid and revealed a ${}^{1,4}B$ conformation, suggestive of a ${}^{1,4}B \rightarrow {}^4E^\ddagger \rightarrow {}^4C_1$ conformational itinerary.

GH11 xylanases are an as yet unresolved case. Early structures of intermediate complexes trapped with 2-fluoro sugars were interpreted as ${}^{2,5}B$ conformations [39,40], suggesting a possible ${}^2S_O \rightarrow {}^{2,5}B^\ddagger \rightarrow {}^5S_1$ itinerary. Very recently a long sought apparent Michaelis complex of xylohexaose bound to the xylanase XynII from *Trichoderma reesi* revealed a slightly distorted 4C_1 conformation [41]. Confounding this issue, product complexes with

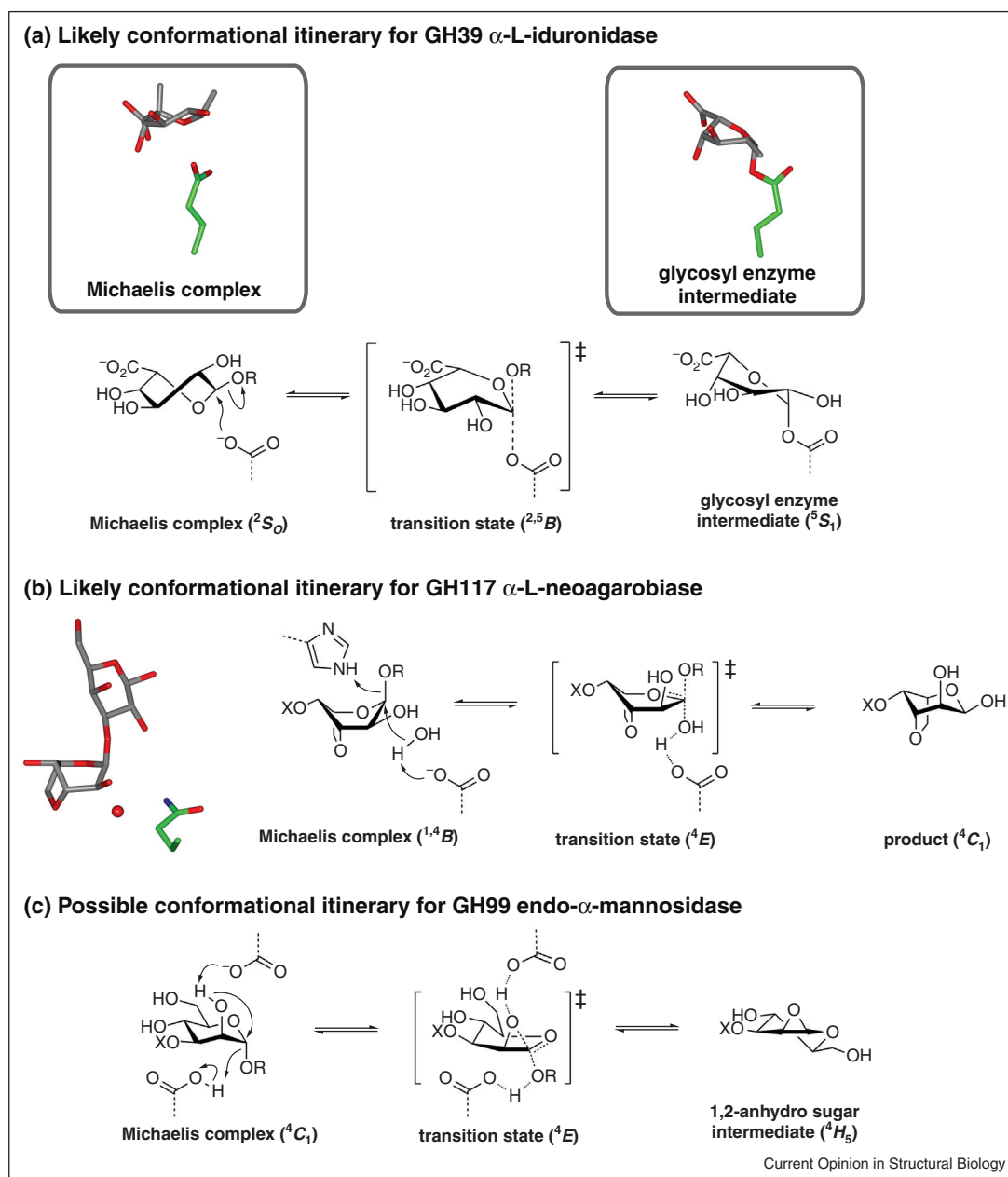
GH11 enzymes reveal a range of different conformations that are inconsistent with the proposed itinerary.

On the importance of being mannose

The majority of common, naturally occurring sugars in their ground-state chair conformation either have an equatorial hydroxyl at C2 (galactosides, glucosides, xylosides, fucosides), or no substituent (sialosides; formally C3). Mannosides and rhamnosides, bearing axial 2-hydroxyls in the ground state conformations, provide exceptions that have interesting and significant consequences for reactivity that lies at the heart of what has been described the recalcitrant chemistry of mannose. For α -mannosides in a 4C_1 conformation, in addition to the existence of the stabilizing anomeric effect that dissuades substrate distortion, the presence of the strongly electron-withdrawing OH at C2 results in opposing dipoles at C1 and C2 that provide additional ground state stabilization. For β -mannosides in a 4C_1 conformation, the anomeric effect provides little stabilization. In addition, other destabilizing effects are operative. Collectively, these can be described as a $\Delta 2$ effect, a term first coined by Reeves [42]. The $\Delta 2$ effect describes the destabilizing effect of an oxygen on one carbon that bisects two oxygens substituted on an adjacent carbon, aligning two dipoles and causing gauche-gauche interactions between the vicinal oxygens. Nucleophilic substitutions at C1 of α -mannosides have to contend with a developing $\Delta 2$ effect. Reflecting these complexities, nature has devised some remarkable strategies for enzymes to solve these problems.

α -Mannosidases of families GH38, 47 and 92 are metal dependent, with crystallographic evidence for the divalent metal cation (Zn^{2+} or Ca^{2+}) binding O2 and O3. This interesting observation may provide a means to overcome the high stability of the unreactive 4C_1 conformation of α -mannosides, and encourage contraction of the ground state O2–C2–C3–O3 torsion angle within the 4C_1 conformation

Figure 3



New glycosidase conformational assignments. **(a)** A likely conformational itinerary for the GH39 human α -L-iduronidase based on structures of a Michaelis complex (4KGJ) and a glycosyl enzyme intermediate (4KH2). **(b)** A likely conformational itinerary for an α -L-neoagarobiase based on a Michaelis complex (4AK7). **(c)** A possible conformational itinerary for a GH99 endo- α -mannosidase based on a proposed mechanism that proceeds through a 1,2-anhydro sugar intermediate.

of 60° toward the $0\text{--}15^\circ$ angle expected at the $B_{2,5}$ transition state [43]. In addition the flexible coordination number and geometry of calcium may allow coordination and delivery of the nucleophilic water, thereby providing a way to overcome a developing $\Delta 2$ effect [43]. A computational study of a GH38 α -mannosidase suggested that Zn^{2+} coordination may stabilize charge that develops on O2 at the oxocarbenium ion like transition state [44].

Remarkably, this calculation revealed that the charge on zinc varies reciprocally with the charge developing on the oxocarbenium ion-like TS.

There is now compelling evidence that family GH2, 26 and 113 retaining β -mannosidases, and GH38 retaining and GH92 inverting α -mannosidases, utilize $B_{2,5}$ transition states with Michaelis complexes in a 1S_5 (for β -) or

0S_2 (for α -), and thus operate through ${}^1S_5 \leftrightarrow B_{2,5}^\ddagger \leftrightarrow {}^0S_2$ conformational itineraries [30**]. The Michaelis complex conformation provides a pseudo axial arrangement of the anomeric leaving group and permits inline attack of the nucleophile; and importantly the 1S_5 conformation relieves the $\Delta 2$ effect. One question that logically arises from studies of transition state conformation is whether all enzymes within a family operate with the same conformational itinerary? Family GH26 contains enzymes that act on β -mannosides, β -glucosides and β -xylosides: lichenases (which hydrolyse the mixed linkage β -1,3-; β -1,4-glucan lichenan), β -mannanase and β -1,3-xylanases. Studies of Michaelis complexes and trapped glycosyl enzymes provides good evidence for an alternative ${}^1S_3 \rightarrow {}^4H_3^\ddagger \rightarrow {}^4C_1$ itinerary for lichenases [45] and β -1,3-xylanases [46*]. The different conformations of the transition state of the D-*gluco*/D-*xylo* and D-*manno* configured substrates result in the substituents at C2 being pseudo-equatorial in both cases and lying at essentially the same place in space, explaining how the conserved catalytic machinery of different GH26 family members can tolerate differently configured sugars, with the specificity arising from a large difference in the positions of the C3 substituents [45], a relationship which is highlighted by the common inhibition of β -mannosidases and β -glucosidases by isofagomine lactam [47].

Uncertainty surrounds the conformational itineraries of α -mannosidases of families GH76, 99 and 125. No complexes are available for GH76 that could provide any insight into a possible itinerary. For GH99, which contains retaining endo-acting α -mannosidases, the only complexes available are with isofagomine and deoxymannojirimycin-derived inhibitors, and these bind in 4C_1 conformations which match the ground state of the inhibitors, so it is not clear whether these complexes represent enzyme-induced or substrate-biased conformations. However, on the basis of an inability to identify a catalytic nucleophile in the complex with α -glucosyl-1,3-isofagomine, a neighboring group participation mechanism for GH99 was suggested that proceeded through a 1,2-anhydro sugar [48*]. This proposal implies the intermediate adopts a 4H_5 conformation, and least nuclear motion would predict a ${}^4C_1 \rightarrow {}^4E^\ddagger \rightarrow {}^4H_5$ itinerary (Figure 3c). For the inverting GH125 α -mannosidases, a pseudo Michaelis complex is available which has the -1 sugar in an undistorted 4C_1 conformation, which matches that observed with a complex with deoxymannojirimycin [49]. The lack of distortion for enzyme bound to the non-hydrolyzable substrate is surprising.

Neuraminidases: of conformational itineraries and transition state mimicry by inhibitors

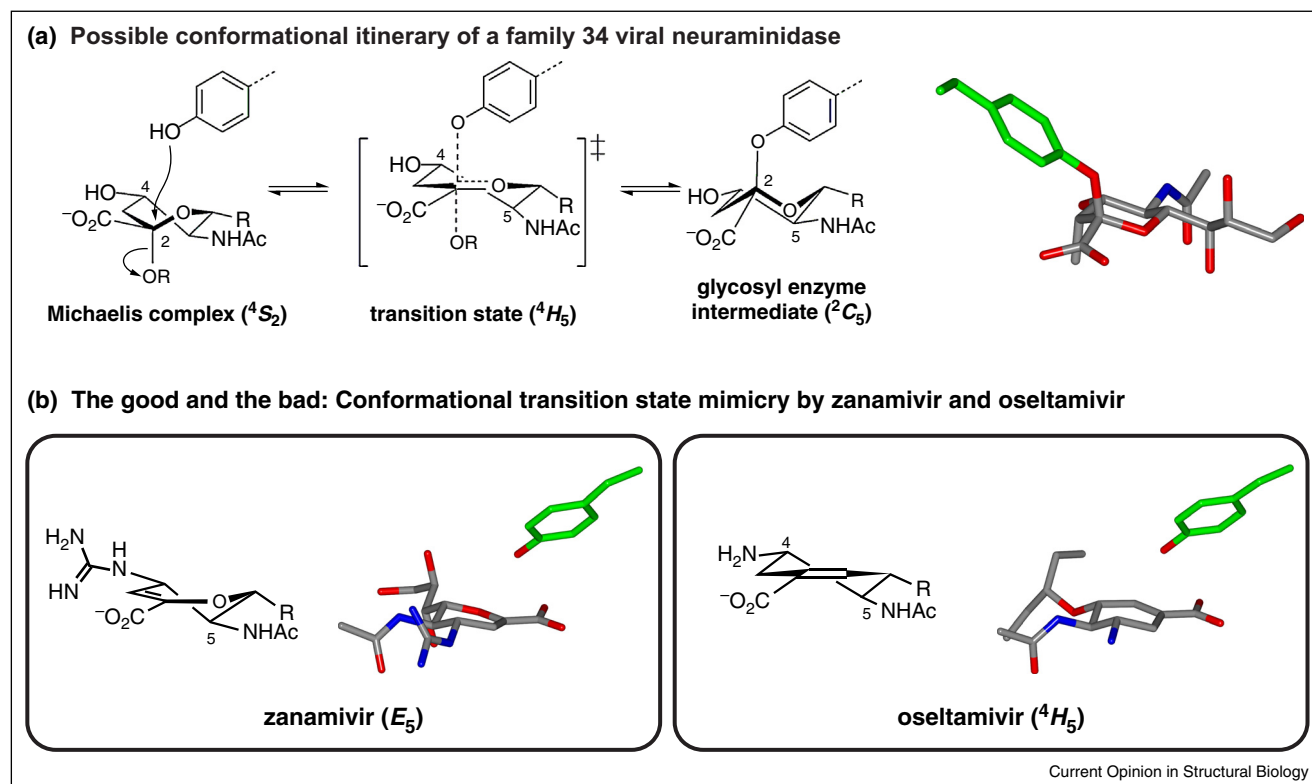
Neuraminidases (sialidases) are glycosidases that cleave sialic acid residues, with the family GH34 viral surface, retaining neuraminidases being significant as the eponymous enzymes in the HXNY classification system of influenza viruses. Influenza virus neuraminidases play

key roles in the infection of cells by the virus and the ability of progeny virions to detach from an infected cell and infect new cells. In two related studies, Withers and co-workers [50**] and Gao and co-workers [51*] designed a series of neuraminidase inhibitors that combine features of the deoxyfluorosugar inhibitors modified to incorporate structural features of the clinically-approved drugs zanamivir (Relenza) and oseltamivir (Tamiflu). X-ray structures of an elusive tyrosyl enzyme intermediate revealed a 2C_5 conformation (Figure 4a). While a ${}^4S_2 \rightarrow {}^4H_5^\ddagger \rightarrow {}^2C_5$ (equivalent to a ${}^3S_1 \rightarrow {}^3H_4^\ddagger \rightarrow {}^1C_4$ for a hexopyranose) is consistent with this data and was proposed for neuraminidases of GH33 [52], Bennet reported kinetic isotope effect analysis of the GH33 *Micromonospora viridifaciens* sialidase that implied a Michaelis complex in a 6S_2 (5S_1 for aldose) conformation [53], a conformation also seen in the Michaelis complex of a GH33 transialidase from *Trypanosoma cruzi*, and consistent with a ${}^6S_2 \rightarrow {}^4H_5^\ddagger \rightarrow {}^2C_5$ (equivalent to a ${}^5S_1 \rightarrow {}^3H_4^\ddagger \rightarrow {}^1C_4$ for an aldose) conformational itinerary [52].

Zanamivir and oseltamivir are potent competitive inhibitors of viral neuraminidases and bear some similarity to the proposed transition state of neuraminidase, yet it is not clear whether either compound achieves its potency through transition state mimicry. As espoused by Wolfenden [54] and Thompson [55] and elegantly summarized by Bartlett [56], critical analysis of transition-state mimicry can be achieved by comparing the effects of equivalent structural perturbations on the affinity of the true transition state (via effects on substrate k_{cat}/K_M) and on the affinity of the transition state analogue (via K_I values), plotted as a linear free energy relationship. Two approaches may be used to introduce perturbations: firstly, modifications to the inhibitor and the corresponding substrate and measurement of kinetic parameters with the wild-type enzyme, or secondly, mutation of enzymatic active site residues to afford mutant enzymes, which are studied with the same inhibitor and substrate. A limitation of the former method is the effort that needs to be expended on synthesis of derivatives, but which allow atomic level modifications to be made limited only by the imagination and synthetic chemistry. Limitations of the latter include the rather blunt tool of site-directed mutagenesis which is restricted to the genetically encoded natural amino acids, and the possibility that mutational perturbations may affect the fundamental reaction mechanism of the enzyme.

Zanamivir was designed to improve the known sialidase inhibitor neuraminic acid glycol (Neu5Ac2en) by the rational inclusion of an enzyme-specific guanidine group targeting a negatively charged pocket near the active site [57]. Neu5Ac2en bears some similarity to the transition state by virtue of sp^2 hybridization at C2. It can be argued that a transition state mimicking inhibitor has the potential to provide potent inhibitors that should be resistant to

Figure 4



Conformational itinerary of influenza GH34 neuraminidases and conformational transition state mimicry by inhibitors. **(a)** The conformational itinerary of influenza GH34 neuraminidases informed by an X-ray structure of a glycosyl enzyme intermediate (4H_5). **(b)** Complexes of anti-influenza drugs with influenza neuraminidases reveals that oseltamivir (2HU4), unlike zanamivir (1NNC), provides good conformational mimicry of the proposed sialidase transition state.

mutations within the active site, as mutations that affect the ability of the inhibitor to bind should also affect the catalytic proficiency of the enzyme to similar degrees, resulting in loss of fitness for the virus. By making alterations in the structure of zanamivir at the 4-position and relating the effects of these changes upon inhibitor K_I values to the equivalent changes to the substrate and their effect upon k_{cat}/K_M or K_M Bennet and co-workers showed that zanamivir is not a transition state analogue and is better considered a ground state analogue [58^{*}]. Notably, this is confluent with the observation that influenza strains resistant to zanamivir possess reduced binding avidity for this drug but still possess catalytic competence. With impressive foresight, this possibility was suggested in the earliest publication describing the invention of zanamivir [57]. The failure of zanamivir in this Bartlett analysis is perhaps not overly surprising. Zanamivir has a double bond between C2=C3, and cannot adopt the 4H_5 conformation predicted for the transition state of GH33 sialidases; indeed an E_5 conformation is observed for zanamivir in complexes (Figure 4b). On the other hand oseltamivir (Tamiflu) is a carbocycle with a double bond located at the appropriate position to mimic the partial C2–O5 double bond at the transition state, and is

observed to bind to sialidases in a 4H_5 conformation matching that of the transition state [59]. It will be interesting to see if Bartlett analysis applied to oseltamivir provides evidence of transition state mimicry. This situation is worth comparing with the powerful α -glucosidase inhibitor acarbose, which has a double bond C5=C6 (using pyranose numbering) and cannot adopt a planar conformation matching that expected for a glycosidase transition state; Bartlett analysis of acarbose with a GH14 cyclodextrin glycosyltransferase gives good correlation of $\log K_I$ with $\log k_{cat}/K_M$, but also good correlation with $\log K_M$ suggesting both substrate and transition state mimicry [60].

Conclusions

A sophisticated view of glycoside hydrolase catalysis is now evident in which conformational changes occur that predispose substrates to react through oxocarbenium ion like transition states that are in accord with stereoelectronic and least nuclear motion principles. The challenges of these studies include the fact that X-ray crystal structures in complex with ligands by their very nature result in perturbation of the system for species nominally on or near the reaction coordinate, and for species off the

reaction coordinate, great care needs to be taken to ensure that ground state conformational preferences do not bias interpretations. Kinetic isotope effect measurements and computational analysis can provide much needed help in assigning conformational itineraries. Compelling data is now available to assign conformational itineraries for a large number of GH families, yet as highlighted above, there are examples in which crystallographic data alone do not allow proposal of conformational itineraries. In these cases application of KIE analyses and theoretical approaches may help reveal a likely itinerary.

There is a growing need for glycosidase inhibitors that exhibit selectivity against specific glycosidases, both to enable chemical biology approaches in glycobiology such as unravelling the roles of specific glycoside hydrolases in complex biochemical pathways [61], and in translational applications, for example, as folding chaperones for treatment of lysosomal storage disorders [62], and as enzyme inhibitors targeting aberrant glycosylation [63]. One of the long-term goals of conformational analysis of the glycosidase reaction coordinate is the hope that such information can inform the design of potent inhibitors, and in addition that these may be specific for particular conformational itineraries. While using such information in the design of inhibitors is not the primary focus of this review it is probably fair to say that as a general rule while the destination is now clear, the path to achieve this is not. Some success has been achieved with inhibitors that have particular conformational biases such as the selectivity of kifunensine for GH47 α -mannosidases. However, the crude attempts to achieve unusual conformations by structural means such as the introduction of bridges across the molecule are typically not tolerated by glycosidase active sites, although the 3,6-anhydrosugars processed by the GH117 α -L-neoagarobiose might constitute a logical target for applying such an approach. More generally, smarter, less intrusive ways are needed to control the conformation of inhibitors through the application of stereoelectronic principles and hybridization. A better understanding of the intrinsic conformational preferences of existing glycosidase inhibitors would greatly assist in directing these efforts.

Conflicts of interest

None declared.

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Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at <http://dx.doi.org/10.1016/j.sbi.2014.06.003>.

References and recommended reading

Papers of particular interest, published within the period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Koshland DE Jr: **Stereochemistry and mechanism of enzymic reactions**. *Biol Rev Cambridge Philos Soc* 1953, **28**:416-436. This timeless review demands close reading by beginning and advanced students of glycosidase mechanism.
 2. Lombard V, Golaconda Ramulu H, Drula E, Coutinho PM, Henrissat B: **The carbohydrate-active enzymes database (CAZy) in 2013**. *Nucleic Acids Res* 2014, **42**:D490-D495.
 3. Schramm VL: **Transition states, analogues, and drug development**. *ACS Chem Biol* 2012, **8**:71-81.
 4. Wolfenden R, Lu X, Young G: **Spontaneous hydrolysis of glycosides**. *J Am Chem Soc* 1998, **120**:6814-6815.
 5. Vocadlo DJ, Davies GJ: **Mechanistic insights into glycosidase chemistry**. *Curr Opin Chem Biol* 2008, **12**:539-555.
 6. Davies GJ, Planas A, Rovira C: **Conformational analyses of the reaction coordinate of glycosidases**. *Acc Chem Res* 2012, **45**:308-316. A recent, comprehensive review of the assignment of conformational itineraries to glycoside hydrolases.
 7. Zechel DL, Withers SG: **Glycosidase mechanisms: anatomy of a finely tuned catalyst**. *Acc Chem Res* 2000, **33**:11-18.
 8. Planas A, Nieto J, Abel M, Segade A: **Unusual role of the 3-OH group of oligosaccharide substrates in the mechanism of Bacillus 1,3-1,4- β -glucanase**. *Biocatal Biotransform* 2003, **21**:223-231.
 9. Tanaka KSE, Zhu J, Huang XC, Lipari F, Bennet AJ: **Glycosidase-catalyzed hydrolysis of 2-deoxyglucopyranosyl pyridinium salts: effect of the 2-OH group on binding and catalysis**. *Can J Chem* 2000, **78**:577-582.
 10. Namchuk MN, Withers SG: **Mechanism of Agrobacterium beta-glycosidase: kinetic analysis of the role of noncovalent enzyme/substrate interactions**. *Biochemistry* 1995, **34**:16194-16202.
 11. Sinnott ML: **Catalytic mechanisms of enzymatic glycosyl transfer**. *Chem Rev* 1990, **90**:1171-1202.
 12. Hine J: **The principle of least nuclear motion**. In *Adv Phys Org Chem*. Edited by Gold V, Bethel D. Academic Press; 1978:1-61.
 13. Sinnott ML: **The principle of least nuclear motion and the theory of stereoelectronic control**. *Adv Phys Org Chem* 1988, **24**:113-204.
 14. Stoddart JF: *Stereochemistry of Carbohydrates*. Canada: John Wiley & Sons; 1971, .
 15. Cremer D, Pople JA: **General definition of ring puckering coordinates**. *J Am Chem Soc* 1975, **97**:1354-1358.
 16. Biarnes X, Ardevol A, Planas A, Rovira C, Laio A, Parrinello M: **The conformational free energy landscape of β -D-glucopyranose. Implications for substrate preactivation in beta-glucoside hydrolases**. *J Am Chem Soc* 2007, **129**:10686-10693.
 17. Mayes HB, Broadbelt LJ, Beckham GT: **How sugars pucker: electronic structure calculations map the kinetic landscape of five biologically paramount monosaccharides and their implications for enzymatic catalysis**. *J Am Chem Soc* 2013, **136**:1008-1022.
 18. Thompson AJ, Dabin J, Iglesias-Fernandez J, Ardevol A, Dinev Z, Williams SJ et al.: **The reaction coordinate of a bacterial GH47 α -mannosidase: a combined quantum mechanical and structural approach**. *Angew Chem Int Ed* 2012, **51**:10997-11001. A comprehensive study combining theory, inhibitor design and X-ray crystallography to assign the conformational itinerary of a GH47 α -mannosidase. Atomic resolution structures of a Michaelis complex, transition state mimicking mannoimidazole and product mimic show how the enzyme distorts the substrate and transition state. QM/MM

calculations reveal how the free energy landscape of isolated α -D-mannose is molded on enzyme to only allow one energetically accessible conformational itinerary.

19. Wolfenden R: **Transition state analog inhibitors and enzyme catalysis.** *Annu Rev Biophys Bioeng* 1976, **5**:271-306.
 20. Heightman TD, Vasella AT: **Recent insights into inhibition, structure and mechanism of configuration retaining glycosidases.** *Angew Chem Int Ed* 1999, **38**:750-770.
 21. Sinnott ML: **On the antiperiplanar lone pair hypothesis and its application to catalysis by glycosidases.** *Biochem J* 1984, **224**:817-821.
 22. Driguez H: **Thiooligosaccharides as tools for structural biology.** *ChemBioChem* 2001, **2**:311-318.
 23. Tews I, Perrakis A, Oppenheim A, Dauter Z, Wilson KS, Vorgias CE: **Bacterial chitinase structure provides insight into catalytic mechanism and the basis of Tay-Sachs disease.** *Nat Struct Biol* 1996, **3**:638-648.
 24. Cartmell A, Topakas E, Ducros VM-A, Suits MDL, Davies GJ, Gilbert HJ: **The *Cellvibrio japonicus* mannanase CjMan26C displays a unique exo-mode of action that is conferred by subtle changes to the distal region of the active site.** *J Biol Chem* 2008, **283**:34403-34413.
 25. Rempel BP, Withers SG: **Covalent inhibitors of glycosidases and their applications in biochemistry and biology.** *Glycobiology* 2008, **18**:570-586.
 26. Legler G, Roeser KR, Illig HK: **Reaction of beta-D-glucosidase A3 from *Aspergillus wentii* with D-glucal.** *Eur J Biochem* 1979, **101**:85-92.
 27. Roeser K-R, Legler G: **Role of sugar hydroxyl groups in glycoside hydrolysis. Cleavage mechanism of deoxyglucosides and related substrates by β -glucosidase A3 from *Aspergillus wentii*.** *Biochem Biophys Acta* 1981, **657**:321-333.
 28. Martinez-Fleites C, He Y, Davies GJ: **Structural analyses of enzymes involved in the O-GlcNAc modification.** *Biochem Biophys Acta* 2010, **1800**:122-133.
 29. Macauley MS, Vocadlo DJ: **Increasing O-GlcNAc levels: an overview of small-molecule inhibitors of O-GlcNAcase.** *Biochim Biophys Acta* 2010, **1800**:107-121.
 30. Williams RJ, Iglesias-Fernandez J, Stepper J, Jackson A, Thompson AJ, Lowe EC *et al.*: **Combined inhibitor free-energy landscape and structural analysis reports on the mannosidase conformational coordinate.** *Angew Chem Int Ed* 2014, **53**:1087-1091.
- Glycosidase inhibitors are often used to assign conformations to species along the reaction coordinate but it is not obvious whether it is appropriate to do so. Computational studies reveal that mannoimidazole is energetically poised to report faithfully on half-chair and boat conformations relevant to the transition state of glycosidases, whereas for isofagomine the equivalent conformations are difficult to achieve. A complex with a β -mannosyl-1,4-mannoimidazole revealed a $B_{2,5}$ conformation for β -mannanases of GH26 and GH113.
31. Varrot A, Macdonald J, Stick RV, Pell G, Gilbert HJ, Davies GJ: **Distortion of a cellobio-derived isofagomine highlights the potential conformational itinerary of inverting beta-glucosidases.** *Chem Commun* 2003:946-947.
 32. Varrot A, Leydier S, Pell G, Macdonald JM, Stick RV, Henrissat B *et al.*: ***Mycobacterium tuberculosis* strains possess functional cellulases.** *J Biol Chem* 2005, **280**:20181-20184.
 33. Bacik JP, Whitworth GE, Stubbs KA, Vocadlo DJ, Mark BL: **Active site plasticity within the glycoside hydrolase NagZ underlies a dynamic mechanism of substrate distortion.** *Chem Biol* 2012, **19**:1471-1482.
- This study reports a series of GH3 retaining β -hexosaminidase X-ray structures in complex with substrate, a glycosyl enzyme and product. Remarkably all structures possess the same 4C_1 conformation. A large protein loop containing the putative histidine general acid/base residue is dislocated in the complex with substrate suggesting that this is not a bonafide Michaelis complex.
34. Maita N, Tsukimura T, Taniguchi T, Saito S, Ohno K, Taniguchi H *et al.*: **Human alpha-L-iduronidase uses its own N-glycan as a substrate-binding and catalytic module.** *Proc Natl Acad Sci U S A* 2013, **110**:14628-14633.
- This and the next study provide a comprehensive structural view of the lysosomal storage disorder-associated human family GH39 iduronidase (IDUA). Structures of a Michaelis complex and glycosyl enzyme reveal important conformations allowing assignment of a conformational itinerary. A remarkable feature of IDUA is that an N-glycan comprises part of the active site, explaining the requirement for correct glycosylation to maintain enzymatic activity.
35. Bie H, Yin J, He X, Kermod AR, Goddard-Borger ED, Withers SG *et al.*: **Insights into mucopolysaccharidosis I from the structure and action of alpha-L-iduronidase.** *Nat Chem Biol* 2013, **9**:739-745.
- See annotation to Ref. [34*].
36. Hill CH, Graham SC, Read RJ, Deane JE: **Structural snapshots illustrate the catalytic cycle of β -galactocerebrosidase, the defective enzyme in Krabbe disease.** *Proc Natl Acad Sci U S A* 2013, **110**:20479-20484.
- A detailed structural picture of GH59 human β -galactocerebrosidase, associated with a lysosomal storage disorder.
37. Juers DH, Heightman TD, Vasella A, McCarter JD, Mackenzie L, Withers SG *et al.*: **A structural view of the action of *Escherichia coli* (lacZ) beta-galactosidase.** *Biochemistry* 2001, **40**:14781-14794.
 38. Hehemann J-H, Smyth L, Yadav A, Vocadlo DJ, Boraston AB: **Analysis of keystone enzyme in agar hydrolysis provides insight into the degradation (of a polysaccharide from) red seaweeds.** *J Biol Chem* 2012, **287**:13985-13995.
- This landmark paper describes a structural study of a fascinating GH117 enzyme that plays a keystone role in the ability of the human microbiota to degrade the seaweed polysaccharide agarose. Owing to the structural constraint imposed by the 3,6-anhydro- α -L-galactose bridge, this enzyme appears to go via a rare 4E transition state conformation.
39. Sidhu G, Withers SG, Nguyen NT, McIntosh LP, Ziser L, Brayer GD: **Sugar ring distortion in the glycosyl-enzyme intermediate of a family GH11 Xylanase.** *Biochemistry* 1999, **38**:5346-5354.
 40. Sabini E, Sulzenbacher G, Dauter M, Dauter Z, Jorgensen PL, Schulein M *et al.*: **Catalysis and specificity in enzymatic glycoside hydrolysis: a 2,5B conformation for the glycosyl-enzyme intermediate revealed by the structure of the *Bacillus agaradhaerens* family 11 xylanase.** *Chem Biol* 1999, **6**:483-492.
 41. Wan Q, Zhang Q, Hamilton-Brehm S, Weiss K, Mustyakimov M, Coates L *et al.*: **X-ray crystallographic studies of family 11 xylanase Michaelis and product complexes: implications for the catalytic mechanism.** *Acta Crystallogr D Biol Crystallogr* 2014, **70**:11-23.
 42. Reeves RE: **The shape of pyranoside rings.** *J Am Chem Soc* 1950, **72**:1499-1506.
 43. Zhu Y, Suits MDL, Thompson AJ, Chavan S, Dinev Z, Dumon C *et al.*: **Mechanistic insights into a calcium-dependent family of α -mannosidases in a human gut symbiont.** *Nat Chem Biol* 2010, **6**:125-132.
 44. Petersen L, Ardevol A, Rovira C, Reilly PJ: **Molecular mechanism of the glycosylation step catalyzed by Golgi alpha-mannosidase II: a QM/MM metadynamics investigation.** *J Am Chem Soc* 2010, **132**:8291-8300.
 45. Money VA, Smith NL, Scaffidi A, Stick RV, Gilbert HJ, Davies GJ: **Substrate distortion by a lichenase highlights the different conformational itineraries harnessed by related glycoside hydrolases.** *Angew Chem Int Ed* 2006, **45**:5136-5140.
 46. Goddard-Borger ED, Sakaguchi K, Reitingner S, Watanabe N, Ito M, Withers SG: **Mechanistic insights into the 1,3-xylanases: useful enzymes for manipulation of algal biomass.** *J Am Chem Soc* 2012, **134**:3895-3902.
- This paper describes a structural study of a GH26 1,3-xylanase, including the trapping of a 2-fluoroglycosyl enzyme, identification of the catalytic nucleophile and the proposal of a conformational itinerary.
47. Vincent F, Gloster TM, Macdonald J, Morland C, Stick RV, Dias FMV *et al.*: **Common inhibition of both β -glucosidases and β -mannosidases by isofagomine lactam reflects different**

conformational itineraries for pyranoside hydrolysis.*ChemBioChem* 2004, **5**:1596-1599.

48. Thompson AJ, Williams RJ, Hakki Z, Alonzi DS, Wennekes T,
 • Gloster TM *et al.*: **Structural and mechanistic insight into N-glycan processing by endo- α -mannosidase.** *Proc Natl Acad Sci U S A* 2012, **109**:781-786.

This work describes the first structural study of a bacterial GH99 enzyme as a model for the mammalian endo- α -mannosidase that cleaves glucose-substituted mannose residues in N-linked glycans. On the basis of an inability to identify a candidate nucleophile in a complex with an isofagomine derived inhibitor, a mechanism involving neighboring group participation and a 1,2-anhydro sugar intermediate is proposed.

49. Gregg KJ, Zandberg WF, Hehemann JH, Whitworth GE, Deng L, Vocadlo DJ *et al.*: **Analysis of a new family of widely distributed metal-independent α -mannosidases provides unique insight into the processing of N-linked glycans.** *J Biol Chem* 2011, **286**:15586-15596.

50. Kim J-H, Resende R, Wennekes T, Chen H-M, Bance N, Buchini S
 •• *et al.*: **Mechanism-based covalent neuraminidase inhibitors with broad-spectrum influenza antiviral activity.** *Science* 2013, **340**:71-75.

In this and Ref. [51*], a series of 3-fluoro sialic acid inhibitors are developed that act as specific, mechanism based anti-influenza drugs, by accumulating a covalent intermediate on the GH34 influenza neuraminidase. X-ray structures reveal the conformation of the covalent glycosyl tyrosine allowing the assignment of a conformational itinerary. These compounds are active in cell-based assays and in animal models, with efficacies comparable to that of the neuraminidase inhibitor zanamivir and with broad-spectrum activity against drug-resistant strains.

51. Vavricka CJ, Liu Y, Kiyota H, Sriwilaijaroen N, Qi J, Tanaka K *et al.*:
 • **Influenza neuraminidase operates via a nucleophilic mechanism and can be targeted by covalent inhibitors.** *Nat Commun* 2013, **4**:1491.

See annotation to Ref. [50**].

52. Amaya MF, Watts AG, Damager I, Wehenkel A, Nguyen T, Buschiazzi A *et al.*: **Structural insights into the catalytic mechanism of *Trypanosoma cruzi* trans-sialidase.** *Structure (London, England: 1993)* 2004, **12**:775-784.

53. Chan J, Lu A, Bennet AJ: **Turnover is rate-limited by deglycosylation for *Micromonospora viridifaciens***

sialidase-catalyzed hydrolyses: conformational implications*for the Michaelis complex.* *J Am Chem Soc* 2011, **133**:2989-2997.

54. Westerik JOC, Wolfenden R: **Aldehydes as inhibitors of papain.** *J Biol Chem* 1972, **247**:8195-8197.

55. Thompson RC: **Use of peptide aldehydes to generate transition-state analogs of elastase.** *Biochemistry* 1973, **12**:47-51.

56. Mader MM, Bartlett PA: **Binding energy and catalysis: the implications for transition-state analogs and catalytic antibodies.** *Chem Rev* 1997, **97**:1281-1301.

57. von Itzstein M, Wu W-Y, Kok GB, Pegg MS, Dyason JC, Jin B *et al.*: **Rational design of potent sialidase-based inhibitors of influenza virus replication.** *Nature* 1993, **363**:418-423.

58. Shidmoosavee FS, Watson JN, Bennet AJ: **Chemical insight into the emergence of influenza virus strains that are resistant to relenza.** *J Am Chem Soc* 2013, **135**:13254-13257.

In this paper the authors apply the Bartlett linear free energy approach to test whether the anti-influenza drug Relenza achieves inhibition of neuraminidase by transition state analogy. The negative result has implications for the acquisition of drug resistance by mutations that reduce inhibitor binding but maintain catalytic activity.

59. Russell RJ, Haire LF, Stevens DJ, Collins PJ, Lin YP, Blackburn GM *et al.*: **The structure of H5N1 avian influenza neuraminidase suggests new opportunities for drug design.** *Nature* 2006, **443**:45-49.

60. Mosi R, Sham H, Uitdehaag JC, Ruitkamp R, Dijkstra BW, Withers SG: **Reassessment of acarbose as a transition state analogue inhibitor of cyclodextrin glycosyltransferase.** *Biochemistry* 1998, **37**:17192-17198.

61. Gloster TM, Vocadlo DJ: **Developing inhibitors of glycan processing enzymes as tools for enabling glycobiology.** *Nat Chem Biol* 2012, **8**:683-694.

62. Parenti G: **Treating lysosomal storage diseases with pharmacological chaperones: from concept to clinics.** *EMBO Mol Med* 2009, **1**:268-279.

63. Dalziel M, Crispin M, Scanlan CN, Zitzmann N, Dwek RA: **Emerging principles for the therapeutic exploitation of glycosylation.** *Science* 2014, **343**:1235681.