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# The morphological diversity of *Osedax* worm borings (Annelida: Siboglinidae)

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*Marine worms in the genus Osedax, have specialized 'root' tissues used to bore into the bones of decomposing vertebrate skeletons and obtain nutrition. We investigated the borings of nine Osedax species, using micro computed tomography to quantitatively describe the morphology of the borings and provide three-dimensional reconstructions of the space occupied by Osedax root tissues inside the bone. Each Osedax species displayed a consistent boring morphology in any given bone, but these differed between bones. In bones where multiple species coexisted there was limited evidence for spatial niche partitioning by Osedax root tissues inside the bones investigated here. The new morphological data may be applied to Osedax traces in fossil bones, showing that borings can be used to indicate minimum species richness in these bones.*

**Keywords:** bioerosion, deep-sea, whale-fall, Polychaeta, taphonomy, trace fossil, computed tomography

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## INTRODUCTION

Animals that excavate hard substrates (known as bioeroders) play an important functional role in many communities by modifying the physical environment and facilitating its use by other organisms. Bioerosion is a widespread behaviour, occurring in at least twelve marine phyla (Warme, 1975). In most instances, boring into hard substrates has evolved to provide protection and a stable domicile. However, in the gutless worms belonging to the genus *Osedax* (Figure 1), the hard substrate itself is used as a food source. All described species in the siboglinid genus *Osedax* subsist on the skeletons of deceased, decomposing vertebrates on the seafloor (Rouse *et al.*, 2004, 2008; Glover *et al.*, 2005, 2013; Fujikura *et al.*, 2006). These worms have soft root-like tissues that release enzymes and acid secretions to invade and erode the bones (Higgs *et al.*, 2011a; Tresguerres *et al.*, 2013). The trunk section of the worm's body, which extends into the water, is crowned with long respiratory palps (Huusgaard *et al.*, 2012). Initial studies suggest that *Osedax* species obtain nutrition from the bone via heterotrophic symbionts that are housed in their root tissues (Goffredi *et al.*, 2005, 2007; Verna *et al.*, 2010), although the precise mechanisms of this process remain unclear (Fujikura *et al.*, 2006; Katz *et al.*, 2010). In addition to nutrition, the borings protect the ovaries and provide a refuge into which the reproductive

females can partially or wholly retract (Glover *et al.*, 2005). Borings are only created by females, since all species investigated to date exhibit extreme male dwarfism, with males living as paedomorphs inside the female tube (Rouse *et al.*, 2008).

To date, only the borings of *Osedax mucofloris* Glover *et al.*, 2005 have been documented in detail (Higgs *et al.*, 2010, 2011a), whilst those of two other species have been shown in profile (e.g. Kiel *et al.*, 2010; Higgs *et al.*, 2012). In this study we examine the range of morphology in borings for nine *Osedax* species and use micro computed tomography to investigate how each species excavates the bone. Our goals are to examine the diversity of *Osedax* borings to help improve our knowledge about their morphological taxonomy, to test the hypothesis that the local diversity of *Osedax* at whale-falls is linked to their ability to differentiate their niches based on boring behaviour and to further improve our knowledge about their trace fossils. We present three-dimensional reconstructions of borings with quantitative details regarding cross-sections, volume and surface area, where possible, for each species.

## MATERIALS AND METHODS

### Provenance of samples

Most of the bone samples investigated in this study were collected during previously reported expeditions (see references in Table 1), so detailed narratives will only be given here for

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Fig. 1. *Osedax rubiplumus*: (A) *i [in] situ* on whale phalanx at time of collection; (B) dissected specimen (courtesy of Greg Rouse). Scale bar: B, 1 cm.

samples not included in previous reports. Unless otherwise stated, bones were collected using remotely operated vehicles (ROVs) from the skeletons of dead whales that had either arrived on the seafloor naturally or were implanted experimentally. Non-cetacean bones were collected from experimentally deployed bone packages and were used when it was not possible to obtain CITES permits for the shipment of whale bones to the UK.

Samples obtained from the Southern California basins were collected in 1995 and 1998 as part of a multi-decade programme of research on whale-fall ecosystems and details of their collection have been reported elsewhere (Smith *et al.*, 2002; Smith & Baco, 2003; Schuller *et al.*, 2004). In October 2008, subsamples were taken from frozen and formalin-fixed bones collected during these previous investigations and transferred to a buffered 4% formaldehyde solution for shipment to the UK. Samples were shipped to the UK and stored in 80% ethanol after CITES approval was obtained. Similarly, samples from Monterey Canyon were collected

during previously reported expeditions (Braby *et al.*, 2007; Lundsten *et al.*, 2010) and sent to the Natural History Museum, London in 80% ethanol from the Scripps Institute of Oceanography under a CITES exemption programme. Details of whale bones experimentally deployed in Antarctica are given in Glover *et al.* (2013).

Pig bone samples were experimentally deployed in January 2010 at 234 m depth near a site of previous whale-fall studies off Cape Nomamsaki, Japan (Fujiwara *et al.*, 2007) and retrieved in May 2010. The bones with *Osedax* living on them were kept in tanks until 2 June 2010 and then fixed in a buffered 4% formaldehyde solution before being transferred to 70% ethanol. Other samples from Sagami Bay, Japan were deployed in May 2009 and collected 8.5 months later (January 2010). Bones with living *Osedax* were kept in tanks for 6 months, after which, they were fixed and treated in the same way as those from Cape Nomamsaki. Tissue samples were taken from individual *Osedax* specimens for genetic analysis prior to fixation. All samples from Japan were then sent to the UK for investigation in July 2010.

### *Osedax* identification

Specimens of *Osedax* were identified to species level upon collection, based on visually identifiable morphological characters or on nucleic acid sequences obtained from tissue samples (e.g. Vrijenhoek *et al.*, 2009). Only seven *Osedax* species have been formally described (Rouse *et al.*, 2004, 2008; Glover *et al.*, 2005, 2013; Fujikura *et al.*, 2006), but operational taxonomic units (OTUs) have been designated for over 20 other genetically distinct forms (Vrijenhoek *et al.*, 2009). Here we use the place-holder names given to these OTUs in

Table 1. Bone samples analysed in this study. Voxel size indicates resolution of computed tomography (CT) scans.

Locality	Depth (m)	Vertebrate species	Bone type	Exposure (months)	CT voxel size ( $\mu\text{m}$ )	<i>Osedax</i> species on bone	References
Antarctica							
Smith Island	500	<i>Balaenoptera acutorostrata</i>	Vertebra	14	20.8	<i>O. antarcticus</i>	Glover <i>et al.</i> (2013)
Monterey Bay							
Monterey Canyon	1820	<i>Eschrichtius robustus</i>	Phalanx	18	32.2	<i>O. rubiplumus</i>	Braby <i>et al.</i> (2007)
Monterey Canyon	2893	<i>Bos</i> sp.	Femur	–	76.0	<i>O. frankpressi</i> , <i>O.</i> ‘nude-plap #20’, <i>Osedax</i> sp.	Jones <i>et al.</i> (2008)
Monterey Canyon	–	Cetacean sp.	Tympanic bulla	–	54.5	<i>Osedax</i> sp.	Lundsten <i>et al.</i> (2010)
Southern California							
San Diego Trough	1220	<i>Eschrichtius robustus</i>	Rib	24	61.5	<i>Osedax</i> sp.	Smith <i>et al.</i> (2002); Smith & Baco (2003)
San Nicolas slope	960	<i>Balaenoptera</i> sp.	Epiphyseal disc	~910	–	? <i>Osedax</i> sp.	
Japan							
Sagami Bay	930	<i>Sus</i> sp.	Pelvis	9	52.4	<i>O.</i> ‘yellow palp’, <i>O.</i> ‘nude palp C’, <i>O.</i> ‘nude palp E’	
Sagami Bay	930	<i>Sus</i> sp.	Limb	9	81.4	<i>O.</i> ‘yellow palp’, <i>O.</i> ‘white-collar’, <i>O.</i> ‘nude palp C’, <i>O.</i> ‘nude palp E’, <i>O.</i> ‘nude palp M’	
Cape Nomamisaki	234	<i>Sus</i> sp.	Limb	3	81.5	<i>O. japonicus</i>	Fujiwara <i>et al.</i> (2007)

previous literature and in GenBank (Table 1; Glover *et al.*, 2013).

### CT scanning

All samples were scanned with a Metris X-Tek HMX ST 225 cone beam micro computed tomography (CT) system operated at the Natural History Museum in London. A total of 3142 angular projections were collected at  $0.11^\circ$  angular intervals in a single  $360^\circ$  rotation for each scanned sample. The radial projections were reconstructed into a three-dimensional matrix of isotropic voxels. Voxel dimensions for each sample are given in Table 1. The volumes were reconstructed using CT Pro Version 2.0 (Nikon Metrology, Tring, UK) and rendered using VG Studio Max 2.0 (Volume Graphics, Heidelberg, Germany).

*Osedax* borings were identified as xenogeneic voids in the bone material that were coincidental with areas colonised by *Osedax*. Voids were isolated and delineated using the region growing tools of the VG Studio software, based on the grey

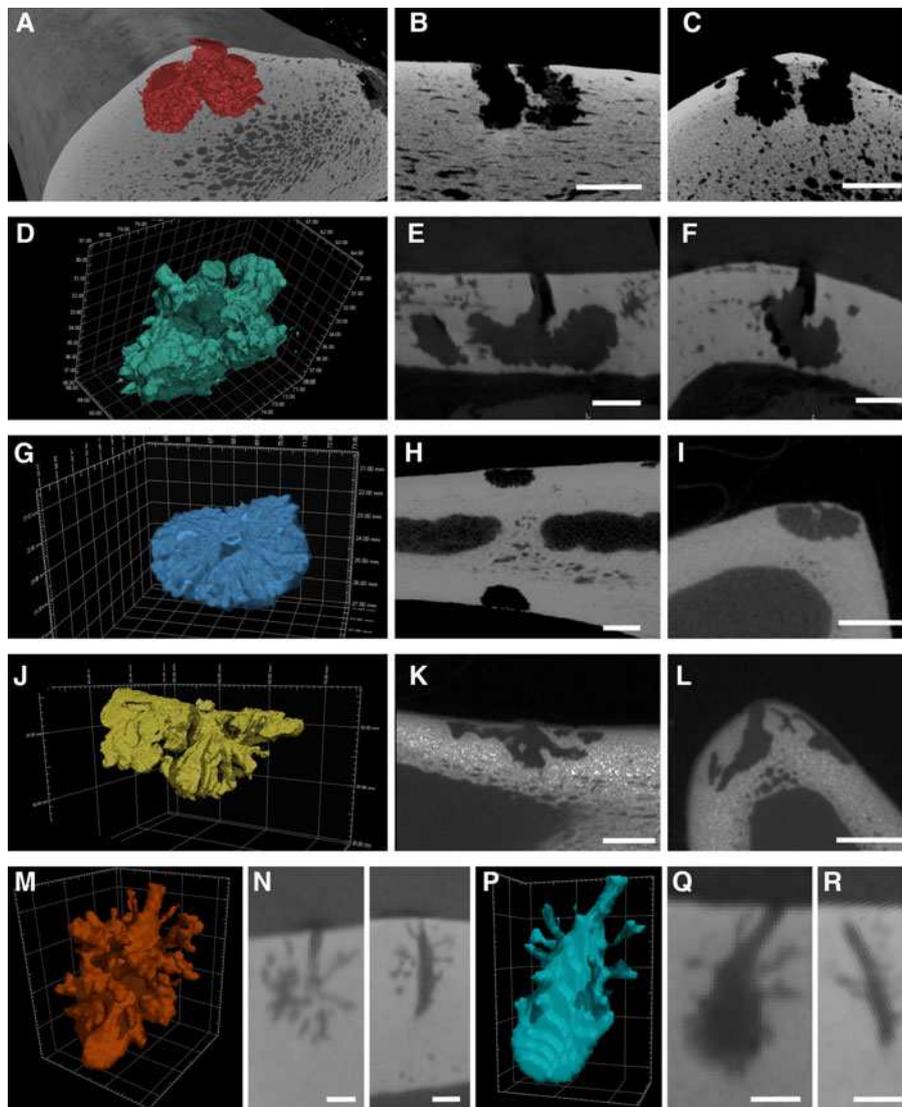
values of voxels making up the void. Voids were then reconstructed as virtual objects and measurements taken using tools of the VG Studio software.

### Petrographic thin sections

Bone samples from an epiphysis disc collected from a whale-fall in the San Nicolas slope (Table 1) could not be adequately CT scanned, owing to the extreme contrast of bone dimensions in perpendicular planes. Instead bone samples were sent for petrographic thin sectioning at the University of Leeds, as described by Higgs *et al.* (2012).

### RESULTS

In this section we report on the morphology of *Osedax* borings found in the bone samples listed in Table 1. We begin with those species that were positively identified and have been formally described, then proceed to describe borings created by



**Fig. 2.** Micro computed tomography scans of *Osedax* borings: (A–C) *O. rubiplumus*; (D–F) *O. frankpressi*; (G–I) *O. japonicus*; (J–L) *Osedax* ‘yellow palp’; (M–R) *Osedax* ‘nude palp #20’. Borings are shown as three-dimensional reconstructions (A, D, G, J, M and P), in longitudinal section (B, E, H, K, N and R) and transverse cross-section (C, F, I, L, O and R). Scale mesh size: D, G, M and P, 1 mm; J, 5 mm. Scale bars: B, C, E, F, 3 mm; H, I, 4 mm; K, L, 5 mm; N–Q, 1 mm.

species that were positively identified, but are not yet formally described. Finally, we show borings created by *Osedax* species that could not be positively identified, but appear to show unique boring morphologies. Borings are described according to the terminology set out by Pirrone *et al.* (2014).

### *Osedax rubiplumus*

Five borings made by *Osedax rubiplumus* Rouse *et al.*, 2004 were documented from a single cetacean carpal bone. Individuals in this sample were identified visually upon collection, based on their size and brilliant red palps (Figure 1A). The *O. rubiplumus* borings are all approximately spherical and appear to expand just below the external bone surface (Figure 2A–C). Some borings have a defined tube section before expansion into the bones begins, but this feature is short when present, making up a small proportion of the boring. The bulk of the boring consists of a chamber section ranging in diameter from 3.2 to 4.6 mm. The surficial openings (apertures) of the borings are 1.3–1.6 mm across.

### *Osedax frankpressi*

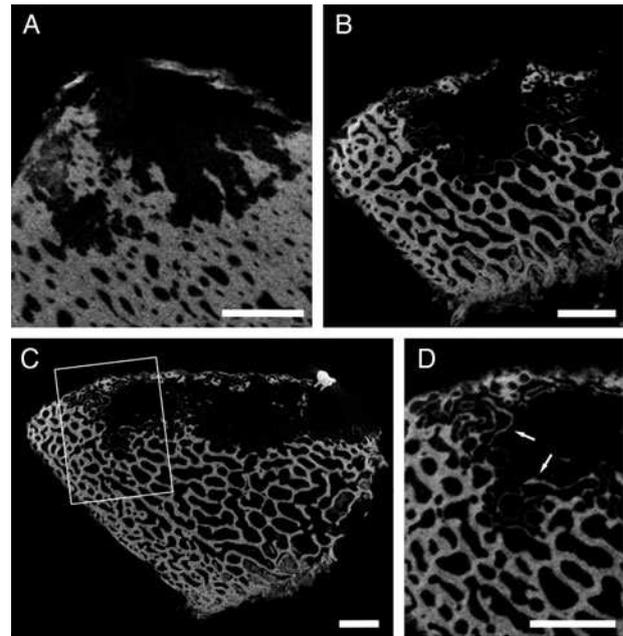
Only one boring of *Osedax frankpressi* Rouse *et al.*, 2004 was available for analysis, from an experimentally deployed cow bone (Table 1). The large boring consists of a relatively deep tunnel (60% total boring depth) penetrating into the bone, with an irregular chamber extending distally from its base (Figure 2D–F). The edge of the chamber is not smooth but shows a serrated pattern where the bone has been eroded. The round aperture of the boring is 1.3 mm in diameter, which continues as a uniformly thick tube for 3.4 mm into the bone before tapering into a 9.4 mm wide chamber.

### *Osedax japonicus*

Numerous borings of *Osedax japonicus* Fujikura *et al.*, 2006 were found in two pig bones and all consistently showed the same morphology, although many had merged. Individual borings are generally hemispherical with an aperture located at the centre of the boring (Figure 2G–I). They lack a tube section and generally sit just underneath the bone surface, covered by a layer of bone <1 mm in thickness. The top of the reconstructed boring shows a corrugated surface of ridges and troughs radiating from the centre of the borings. Several of the borings show a wedge of bone situated just below the aperture of the boring.

### *Osedax antarcticus*

Small samples (<4 × 4 cm) of whale bone containing several individuals of *Osedax antarcticus* Glover *et al.*, 2013 were CT scanned, but it was not possible to identify intact individual borings because of the small bone samples. Nevertheless, cross-sections of the borings allow some comparisons to other boring morphologies (Figure 3). Outlines of individual and merged borings show that there is a short tube section leading from the surface aperture (1.1 mm wide) to a globular chamber (4.7 mm across). At its periphery the chamber is divided into distinct lobes that are more pronounced in the denser bone sample. The small size of the samples facilitated high resolution CT scanning (Table 1) that was able to show



**Fig. 3.** *Osedax antarcticus*: (A–C) cross-sections of borings in segments of whale vertebra; (D) enlargement of boxed area in (C), highlighting bone trabeculae that have been hollowed out by root tissues (arrows). Scale bars: A–D, 2 mm.

the hollowed-out bone trabeculae at the margins of the *Osedax* borings (Figure 3D), as previously described from *O. mucofloris* borings (Higgs *et al.*, 2011a).

### *Osedax* ‘yellow-palp’

*Osedax* ‘yellow-palp’ is an undescribed species known from 980 m in Sagami Bay, Japan. Samples of their borings were found in pig bones and specimens were identified as a genetically distinct OTU (‘Sagami 5’, FM995539 & FM998083 on GenBank). The borings of *O.* ‘yellow-palp’ are predominantly composed of a multi-annexed chamber that expands from a central aperture (Figure 2J–L). The boring extends laterally under the bone surface and is wider than it is deep, giving it a relatively high surface area to volume ratio. The annexes are narrow nearest the central aperture but fan-out distally. CT scans of a small individual show that the boring initially starts out as a shallow flat chamber before branching off into separate annexes.

### *Osedax* ‘nude palp #20’

One species of unidentified *Osedax* that was found on the cow bone from Monterey Canyon, living alongside *O. frankpressi* (Table 1), was very different in morphology from any other borings examined to date. Based on DNA retrieved from some of the borings, it seems that they were created by *Osedax* ‘nude palp #20’, another unnamed OTU from the 2893 m site (S. Johnson, unpublished data), but this could not be confirmed visually because of their small size. The boring consisted of a narrow aperture (0.38 mm) that expanded into a laterally broad, but vertically flattened, pouch-shaped chamber (Figure 2M–R). Small micro-tunnels emanated from this

chamber, probably representing exploratory root tendrils. Assuming that this is a true representation of root morphology (see Higgs *et al.*, 2011a), its surface area to volume ratio would be more than double that of any other species investigated (Table 2).

### *Osedax* 'nude palp' species: C, E and M

These three species of *Osedax* were found in two pig bones from Sagami Bay, Japan (Table 1), but two (C and E) are also known from Monterey Canyon on the opposite side of the Pacific Ocean (Vrijenhoek *et al.*, 2009). The three species are similar in external morphology and at the time of collection were thought to represent a single species. Genetic sequencing of several of these individuals revealed three distinct species of *Osedax* (F. Pradillon, unpublished data). Since a limited number of individuals were sequenced, only a few borings could be definitely attributed to one species and these often occurred in close proximity with each other, so the borings of these three species are presented together. In all instances where borings could definitely be attributed to one particular species the borings were very shallow and laterally expansive, occurring just under the bone (Figure 4). This morphology was observed for all three of the 'nude palp' species from Sagami Bay. The aperture of most of these borings was located in the centre of the boring.

### Additional *Osedax* borings from the San Diego Trough

Higgs *et al.* (2012) briefly described *Osedax* borings in the rib of a gray whale from the San Diego Trough off Southern California from an undescribed species of *Osedax*. It has not been possible to obtain sufficient genetic information from these specimens to positively identify them, since they were preserved in formalin.

Unlike most other *Osedax* borings, those seen in this bone penetrate deeply into the cancellous region of the bone (Figure 5). There is a long tube section (3.64 mm in the specimen illustrated in Figure 5) that passes through the dense cortical bone layer before expanding into a pouch-shaped (*sensu* Bromley, 1994) chamber section (Table 2). The chamber is substantially wider than the tube section in one axis but nearly identical in width when viewed on the perpendicular axis (Figure 5B, C).

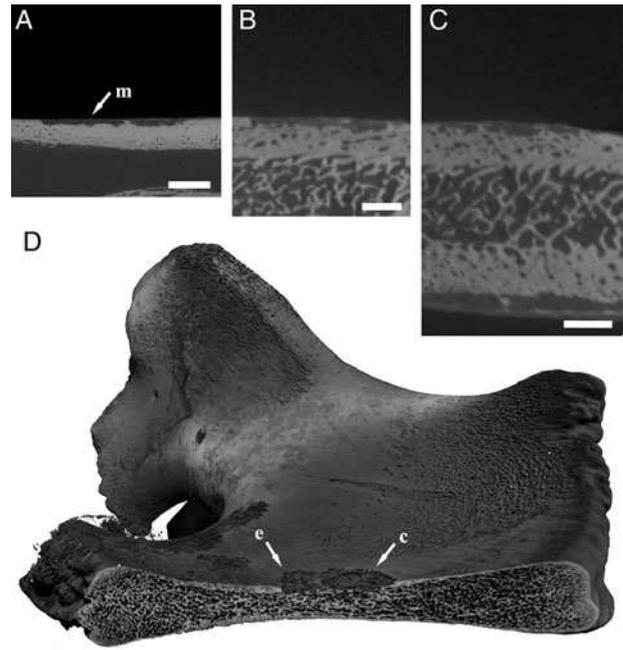


Fig. 4. *Osedax* 'nude palp' species: (A) boring made by *O.* 'nude palp M' (arrowed); (B, C) borings made by *O.* 'nude palp C'; (D) pig pelvis showing overlapping borings of *O.* 'nude palp' species (C) and (E) (arrowed). Scale bars: A-C, 2 mm.

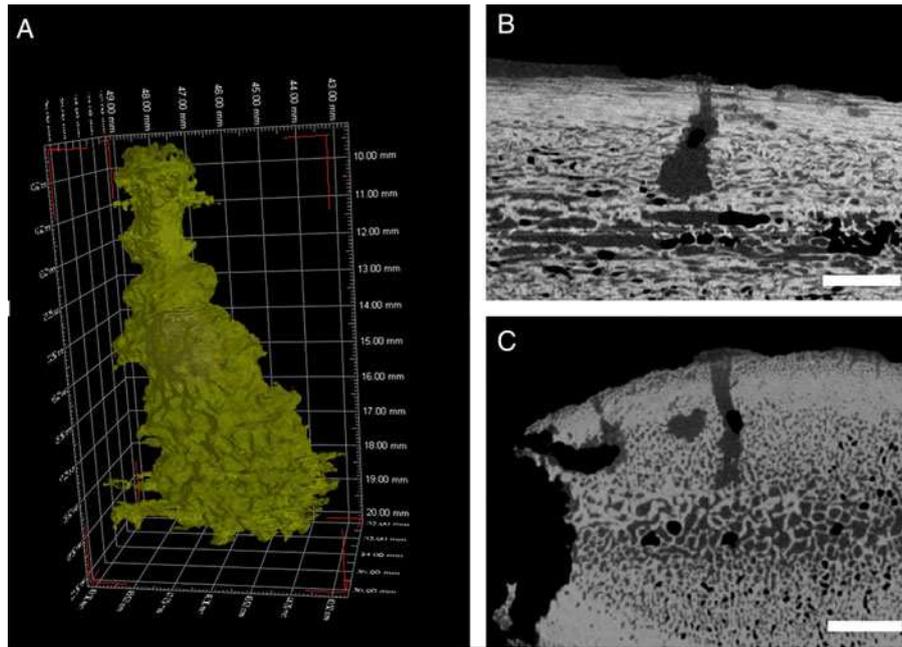
### Additional *Osedax* borings from Monterey Canyon

Another large boring found in the same bone as *Osedax frankpressi* & *O.* 'nude palp #20' (Table 1), is assumed to belong to a separate *Osedax* species, having a distinct hemispherical morphology (Figure 6D). Unlike the *O. mucifloris* or *O. japonicus* borings however, these had many fine micro-tunnels emanating from the main chamber, which densely penetrated the surrounding bone. It can be assumed that expansion of the boring by this species of *Osedax* occurs when these many micro-tunnels grow together and merge.

A similar pattern of small micro-tunnels extending from the main chamber was also identified in *Osedax* borings in a whale ear-bone (tympanic bulla) from Monterey Canyon (Figure 6A). Other *Osedax* borings in this bone showed similar structure, but also have defined tube sections leading from the surficial aperture (Figure 6B, C). The chamber of the borings found in the ear bone tend to be globular, but

Table 2. Measurements for the largest boring produced by each *Osedax* species in this study. SA:V is the surface area to volume ratio of the boring. (N) number of individual borings of each species examined; (-) measurements not possible because no individual borings could be isolated from others.

Species (N)	Aperture diameter (mm)	Length (mm)	Width (mm)	Depth (mm)	Depth of tube part (mm)	Volume (mm <sup>3</sup> )	Surface area (mm <sup>2</sup> )	SA:V
<i>O. rubiplumus</i> (5)	1.66	4.36	3.93	3.52	0.50	35.89	122.54	3.41
<i>O. frankpressi</i> (1)	1.33	9.37	4.43	5.63	3.38	89.93	246.44	2.70
<i>O. japonicas</i> (>12)	0.72	5.90	5.50	2.51	0.26	34.89	68.93	1.98
<i>O.</i> 'yellow-palp' (3)	1.30	13.46	5.54	4.53	0.35	98.52	473.69	4.81
<i>Osedax</i> sp., San Diego Trough (4)	1.90	6.63	3.38	6.52	3.64	50.11	185.37	3.70
<i>O.</i> 'nude palp #20' (>6)	0.38	1.82	0.39	3.62	0.97	2.18	19.85	9.11
<i>O. antarcticus</i> (4)	1.12	4.74	-	4.03	0.89	-	-	-
<i>O.</i> 'nude palp M' (2)	0.39	12.13	-	0.90	0.21	-	-	-
<i>O.</i> 'nude palp C' (4)	0.31	7.96	-	1.24	0.20	-	-	-



**Fig. 5.** Undetermined *Osedax* sp. boring from San Diego Trough: (A) three-dimensional reconstruction; (B) longitudinal section; (C) Transverse cross section. Scale mesh spacing: A, 1 mm. Scale bars: B, C, 5 mm.

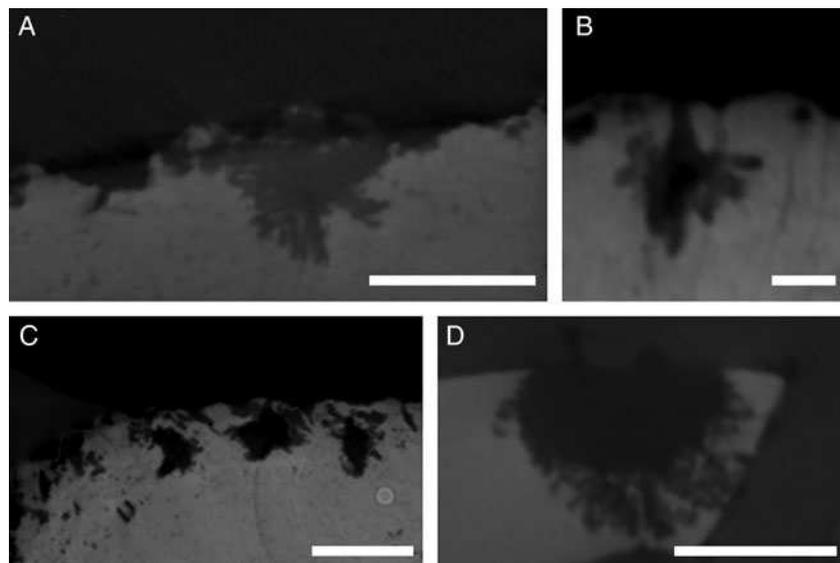
are more irregular than those from other bone. There is no information on the species of *Osedax* in this bone.

### Additional borings from San Nicolas slope

Inspection of the bones recovered from a natural whale-fall discovered on the San Nicolas slope revealed small holes in surface of an epiphyseal vertebral disc similar to those produced by *Osedax*. Thin sections of bone that bisected these holes revealed areas of bone trabeculae that had been hollowed out (Figure 7), as observed in CT scans of bone inhabited by *O. antarcticus* specimens described above (Figure 3D).

### DISCUSSION

*Osedax* worms are not physiologically restricted to exploiting whale bones, and experimental studies have shown them living on the bones of fish (Rouse *et al.*, 2011), cows (e.g. Jones *et al.*, 2008), pigs (Vrijenhoek *et al.*, 2008a) and even birds (R. Vrijenhoek, personal observation). Borings found in a similar array of fossil vertebrate bones have been attributed to *Osedax* (Kiel *et al.*, 2010, 2012). Previous investigations of *Osedax* borings have established computed tomography as an ideal method of studying these features (Higgs *et al.*, 2011a, 2012). The results presented here are the first systematic examination of the borings of multiple *Osedax* species using



**Fig. 6.** Undetermined *Osedax* spp. borings from Monterey Canyon: (A–C) cetacean tympanic bulla (ear bone); (D) cow femur. Scale bars: A, C and D, 5 mm; B, 1 mm.

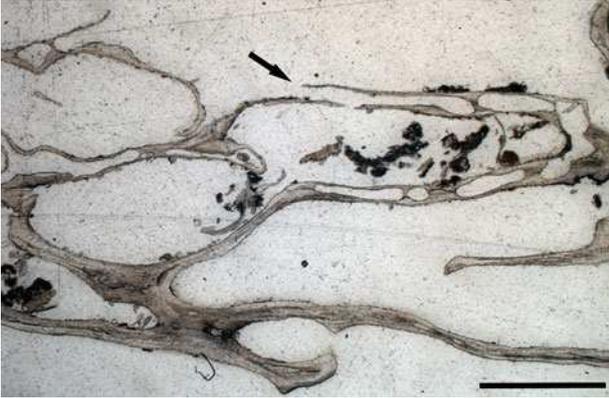


Fig. 7. Petrographic thin section through a boring in whale bone from San Nicolas slope. Hollowed out trabeculae (arrow) near external surface of the bone.

CT scanning. Below, we discuss the results with reference to the autecology, synecology and palaeontology of this diverse genus of polychaete worms.

### Factors driving morphological diversity in *Osedax* borings

The range of boring morphologies illustrated above shows that *Osedax* borings are highly variable between species and previous studies have shown that the boring morphology is also variable within a species (Higgs *et al.*, 2011a). Disentangling this intraspecific variation from interspecific variation is dependant on identifying the factors that govern the growth of root tissues in each species. A detailed study of borings created by *Osedax mucofloris* in whale bones showed that boring morphology was primarily determined by bone structure: borings were consistent on the same bone but highly variable when compared between different bone types (Higgs *et al.*, 2011a). This phenomenon is also observed here for *Osedax japonicus*, *O. rubiplumus*, *O.* 'nude palp C', *O.* 'nude palp E', *O.* 'nude palp #20' and the *Osedax* specimens on the San Diego Trough whale bone. In all of these cases the morphology of borings within a particular species is consistent when examined from the same bone, generally confirming the role of bone structure in determining the morphology of *Osedax* borings.

Where multiple species co-exist on the same bone, each species may display a different boring morphology, showing that there are also species-specific factors that influence

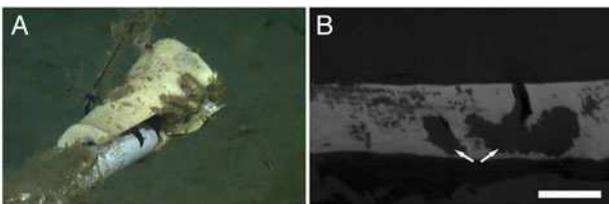


Fig. 8. Cow femur from Monterey Canyon colonized by multiple *Osedax* species; (A) experimentally implanted bone *in situ*, with the gelatinous tubes of *Osedax* 'nude palp #20' visible on the bone; (B) computed tomography scan showing relative position of a single large *O. frankpressi* boring (arrows) and multiple borings of *Osedax* 'nude palp #20' (other voids in the bone matrix).

boring morphology. For example, the cow bone collected from Monterey Canyon housed one specimen of *Osedax frankpressi* and multiple individuals of two undescribed species. All three species displayed contrasting boring morphologies (Figures 6D & 8B). One of the undescribed species (*O.* 'nude palp #20') was more prevalent on this bone and all individuals of this species showed the same boring morphology. The consistent differences in boring morphology between species may lie in the detailed mechanisms of how they erode the bone.

In addition to phylogeny and bone structure, ontogenetic development of root tissues can lead to changes in the morphology of *Osedax* borings such that complex boring morphologies develop from simpler ones. For example, the multi-annexed borings produced by large individuals of *Osedax* 'yellow-palp' are more complex than the simple hemispherical borings created by smaller individuals. Similarly, the simple bulb shaped chamber created by the small individuals of *O. rubiplumus* shown above lacks the discrete root projections figured in the original description of the species, which develop as individuals mature (Greg Rouse, personal communication). The ontogenetic development of complexity contrasts with the simple hemi-ellipsoidal shape of *O. mucofloris* borings, which remains consistent in shape over a range of sizes, indicating that there is little ontogenetic variation in this species (Higgs *et al.*, 2011a).

### Resource partitioning and coexistence of *Osedax* species

Vertebrate skeletons are spatially and temporally patchy resources in the deep-sea, which are degraded by *Osedax* worms over time, eventually leading to the demise of the *Osedax* populations living on them. The bones are often densely populated and it is common for multiple *Osedax* species to occupy the same skeleton, or even the same bone in close proximity (see Table 1). For example, one whale skeleton monitored in Monterey Canyon had seven distinct species of *Osedax* living on it concurrently, with six of them coexisting for several years (Lundsten *et al.*, 2010). Assuming limiting resources, classic niche theory suggests that these species must differ in some ecological trait in order to coexist, i.e. niche differentiation (Chesson, 2000). Just as plants may segregate their root systems in soil to avoid competition for resources (Schenk *et al.*, 1999; Schenk, 2006), it may be hypothesized that *Osedax* root tissues will be segregated spatially within a single bone when resources are limiting so as to access different parts of the resource spectrum. Root tissues could be segregated by depth within the bone or by type of bone tissue. At least three of the bones used in this study contained multiple species, allowing us to test this hypothesis.

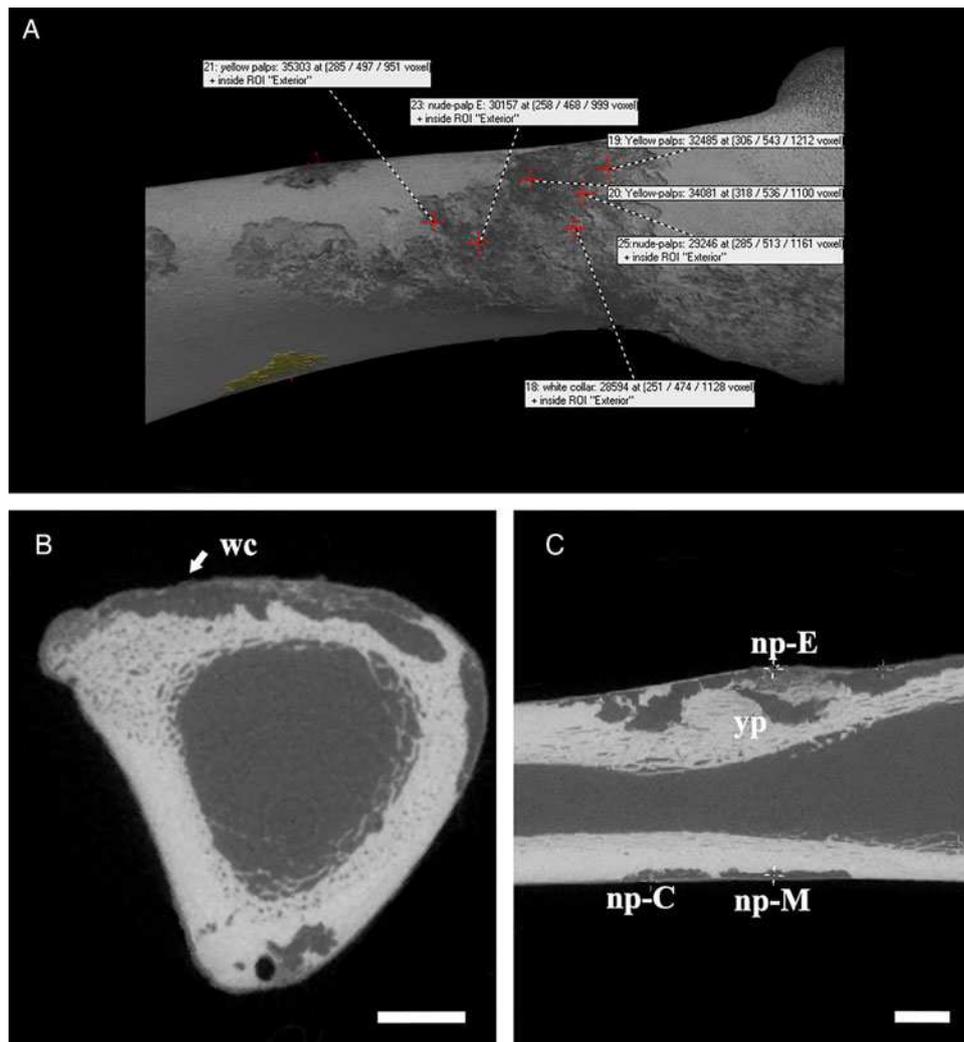
The cow bone from Monterey Canyon contained three species of *Osedax*, each with a distinct boring morphotype. The root tissue of *Osedax* 'nude palp sp. #20' and that of the unidentified species were both located just below the surface of the bone, whereas that of *O. frankpressi* sits deeper into the bone, at the base of a relatively long tube (Figure 8). Although only one individual of *O. frankpressi* was available for study with CT, deep-penetration of their roots into the bone is commonly observed (Greg Rouse, personal communication). Similarly, the roots and ovisac of the *Osedax* species

from the San Diego Trough sit at the base of a long tube section that penetrates deep into the spongy (and lipid-rich) part of the bone (Figure 5; Table 2). This is in stark contrast to the root growth of *O. mucofloris*, which closely follows the cortical (collagen-dense) part of the bone near the surface (Higgs *et al.*, 2010, 2011a). These two contrasting boring types show that there may be different spatial niches within a bone, occupied by different *Osedax* species, although differences in size and tissue distribution between cow and whale bones of various sizes could also conceivably influence burrow morphologies. An even more extreme example of spatial differentiation has apparently evolved in another undescribed OTU, *Osedax* 'spiral', which exploits bones that are buried in the sediment (Braby *et al.*, 2007). In this species, the root tissues are highly filamentous and bear more of a resemblance to their botanical analogues.

The distinction between *Osedax* species in a bone is not always clear-cut. At least four species co-occurred on the pig bones from Sagami Bay, and no distinction could be made between the borings of the closely related *Osedax* 'nude palp' species (Figure 9A–C). Borings of *Osedax* 'yellow palp' were noticeably deeper than those of the other species and

their roots seemed to undercut those of the 'nude palp' species (Figure 9C). Additionally, the *Osedax* 'white collar' boring is not at all distinguishable from those of the *Osedax* 'nude palp' species. Caution must be exercised in interpreting niche differentiation in cow bones, however, because cow bones are unlikely to be natural habitat for *Osedax*. Nonetheless, taken together, these examples suggest that some degree of spatial niche differentiation exists among different *Osedax* species occupying a particular bone, but the diversity of spatial niches appears to be far less than the diversity of *Osedax* species. In other words many species still display spatial niche overlap. However, we cannot say from the present data whether this niche overlap persists when resources are limiting (e.g. at very high population densities), and in natural habitats such as adult whale bones, where the full potential niche dimensionality is likely to be present.

The three bones analysed here only provide a snapshot in time and cannot provide information on temporal dynamics of recruitment, population growth and resource utilization that may facilitate co-existence of species (e.g. Leibold *et al.*, 2004). At the regional scale, bathymetric segregation plays a role for many of the *Osedax* species living in Monterey Bay,



**Fig. 9.** Pig limb bone colonized by multiple *Osedax* species: (A) computed tomography reconstruction showing species marked with crosses; (B) transverse cross-section through bone; (C) longitudinal cross-section through bone. np-C, *O.* 'nude palp C'; np-E, *O.* 'nude palp E'; np-M, *O.* 'nude palp M'; wc, *O.* 'white collar'; yp, *O.* 'yellow palp'. Scale bars: B and C, 5 mm.

California (Lundsten *et al.*, 2010). At the local scale, temporal segregation also plays a role, as *Osedax* species exhibit changing dominance as bones decompose, with some species disappearing altogether (Braby *et al.*, 2007; Pradillon *et al.*, 2009; Lundsten *et al.*, 2010). The composition of nutritional symbionts associated with various *Osedax* species also change over time, but the symbionts do not appear to be linked to particular hosts (Salathé & Vrijenhoek, 2012). Heterogeneity in the skeletal resource (see Higgs *et al.*, 2011b) may provide another mechanism for niche differentiation, but given the strong role of dispersal and recruitment in shaping the composition of *Osedax* communities (Rouse *et al.*, 2008; Vrijenhoek *et al.*, 2008b) it is also likely that neutral processes are important in maintaining *Osedax* species coexistence. Neutral theory assumes that all species are functionally equivalent and that community structure is primarily shaped by stochastic demographic mechanisms (Hubbell, 2005). It was initially assumed that neutral and niche theories were mutually exclusive, but current opinion posits a continuum where 'niche and neutral processes combine to generate coexistence' (Adler *et al.*, 2007). We suggest that such a combination of mechanisms may explain why spatial niche differentiation is not more evident here, given the high global and local diversity of what appear to be functionally similar *Osedax* species coexisting on vertebrate carcasses in space and time.

### *Osedax* borings as trace fossils

Detailed information on the morphology of borings is essential for documenting the fossil record of *Osedax* (Kiel *et al.*, 2010; Higgs *et al.*, 2012) and elucidating their evolutionary history (Vrijenhoek *et al.*, 2009), as the soft bodied animals themselves are unlikely to be preserved in the geological record. The diverse array of borings illustrated here greatly expands the range of borings known to be created by *Osedax*, allowing more detailed information to be gleaned from the fossil record. For example, fossil borings in fish bones shown by Kiel *et al.* (2012) are almost identical in size and shape to those of *O. japonicus* shown here. Additionally, the consistency of boring morphology by different *Osedax* species in the same bone (see above) allows palaeontologists to estimate the minimum number of species that were living on a fossil bone where different boring morphotypes are present; i.e. multiple ichnospecies of trace fossil found on the same bone probably represent multiple biological species.

*Osedax* traces in fossil bones may take several forms depending on how much the borings have been degraded before burial (Higgs *et al.*, 2012). In cancellous bone with only a thin layer of cortical bone, borings are likely to collapse and appear as shallow chambers (*sensu* Pirrone *et al.*, 2014) in the bone surface (Figures 4D and 9A; see also Higgs *et al.* (2011a), figure 6A). In dense cortical bone borings are more likely to be preserved intact and simply appear as small round holes in the bone surface. Borings created by species that lack a substantial tube section (such as *O. japonicus*) may also collapse, leaving deep, hemispherical chambers in the dense bone. Such features may be caused by a number of organisms, so more detailed information is needed to identify *Osedax* in fossil bones (Higgs *et al.*, 2012). The peculiar way in which *Osedax* hollow out individual bone trabeculae (Figures 3D & 7) as part of the boring process was first shown by Higgs *et al.* (2011a) for *O. mucofloris*. This

pattern of bioerosion is also shown here for *O. antarcticus* and has been associated with other *Osedax* borings (Higgs, 2012). The hollowed out trabeculae may be taken as indicators of *Osedax* activity and offer additional diagnostic evidence when assignment of fossil borings to *Osedax* is not straightforward (see discussion in Higgs *et al.*, 2012). Furthermore, it may provide a relatively cheap and quick way of identifying *Osedax* activity in fossil bones, since micro-CT technology is not widely available at low cost. A similar pattern of bone erosion was also observed on fossil whale bone from the Miocene (Amano *et al.*, 2007; Figure 2E), although no other signs of *Osedax* traces were described in this instance.

The presence of hollowed out trabeculae on bones from the San Nicolas slope indicates that *Osedax* was once present on this carcass, but had died out by the time that it was first sampled over fifty years after the carcass arrived on the seafloor (Schuller *et al.*, 2004). Exactly why the *Osedax* population died out is not clear, but may be related to natural succession following the diminishing collagen resources in the surficial portions of the bone as the dense parts of bones were eaten away. Resource exhaustion by *Osedax* may explain the apparent disparity in rates of decomposition between the large San Nicolas skeleton and those of juvenile whale skeletons of Monterey Canyon that were estimated to degrade or disappear in under 10 yr (Lundsten *et al.*, 2010). Factors such as bone size and degree of bone calcification are also likely to be important for bone resistance to *Osedax* boring and the time scales of whale-skeleton persistence at the deep seafloor (Smith & Baco, 2003).

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