

This is a repository copy of Macroevolutionary consequences of profound climate change on niche evolution in marine molluscs over the past three million years.

White Rose Research Online URL for this paper: http://eprints.whiterose.ac.uk/82681/

Version: Accepted Version

Article:

Saupe, EE, Hendricks, JR, Portell, RW et al. (4 more authors) (2014) Macroevolutionary consequences of profound climate change on niche evolution in marine molluscs over the past three million years. Proceedings of the Royal Society B: Biological Sciences, 281 (1795). ARTN 20141995. ISSN 0962-8452

https://doi.org/10.1098/rspb.2014.1995

Reuse

Items deposited in White Rose Research Online are protected by copyright, with all rights reserved unless indicated otherwise. They may be downloaded and/or printed for private study, or other acts as permitted by national copyright laws. The publisher or other rights holders may allow further reproduction and re-use of the full text version. This is indicated by the licence information on the White Rose Research Online record for the item.

Takedown

If you consider content in White Rose Research Online to be in breach of UK law, please notify us by emailing eprints@whiterose.ac.uk including the URL of the record and the reason for the withdrawal request.



eprints@whiterose.ac.uk https://eprints.whiterose.ac.uk/

1 2 3	Macroevolutionary consequences of profound climate change on niche evolution in marine molluscs over the past three million years
4 5 6	Saupe, E.E. ^{1*} ; Hendricks, J.R. ² ; Portell, R.W. ³ ; Dowsett, H.J. ⁴ ; Haywood, A. ⁵ ; Hunter, S.J. ⁶ ; Lieberman, B.S. ⁷
7 8 9	¹ Biodiversity Institute and Department of Geology, University of Kansas, 1475 Jayhawk Boulevard, Room 120 Lindley Hall, Lawrence, KS 66045
10 11 12 13	² Department of Geology, San José State University, Duncan Hall 321, San José, CA 95192 and Paleontological Research Institution, 1259 Trumansburg Road, Ithaca, NY 14850
14 15 16	³ Division of Invertebrate Paleontology, Florida Museum of Natural History, University of Florida, 1659 Museum Road, PO Box 117800, Gainesville, FL 32611
10 17 18	⁴ U.S. Geological Survey, 926A National Center, Reston, VA 20192
10 19 20 21	⁵ School of Earth and Environment, University of Leeds, Leeds, LS2 9JT, United Kingdom
22 23 24 25	⁶ Sellwood Group for Palaeo-Climatology, University of Leeds, Room 9.127 Earth and Environment Building, School of Earth and Environment, West Yorkshire, LS2 9JT, United Kingdom
23 26 27 28	⁷ Biodiversity Institute and Department of Ecology & Evolutionary Biology, University of Kansas, 1345 Jayhawk Boulevard, Dyche Hall, Lawrence, KS 66045
29 30	*Corresponding author: Erin E. Saupe; <u>eesaupe@gmail.com</u>
31	Summary
32 33 34 25	In order to predict the fate of biodiversity in a rapidly changing world, we must first understand how species adapt to new environmental conditions. The long-term evolutionary dynamics of species' physiological tolerances to differing climatic regimes remains observe. Here, we units palacentalogical and poentalogical data to apply
35 36 37 38	whether species' environmental tolerances remain stable across three million years of profound climatic changes using ten phylogenetically, ecologically, and developmentally diverse mollusc species from the Atlantic and Gulf Coastal Plains, USA. We additionally
39 40 41 42	investigate whether these species' upper and lower thermal tolerances are constrained across this interval. We find that these species' environmental preferences are stable across the duration of their lifetimes, even when faced with significant environmental perturbations. The results suggest that species will respond to current and future warming
43	by altering distributions to track suitable habitat, or, if the pace of change is too rapid, by

- going extinct. Our findings also support methods that project species' present-day environmental requirements to future climatic landscapes to assess conservation risks.

47

Keywords Atlantic Coastal Plain | conservation palaeobiology | fundamental niche |
 macroevolution | mid-Pliocene Warm Period | Mollusca

50

51 **1. Introduction**

52 Earth's climate is rapidly changing, altering all facets of our planet at an 53 unprecedented rate, from the biosphere, to the hydrosphere, to the atmosphere. Given 54 these changes, debate exists as to whether species can adapt their physiological tolerances, 55 or niches, to altered environmental conditions [1-4]. Determining whether species' niches 56 evolve or remain stable in the face of environmental change is important for 57 implementing proper conservation measures, mitigating threats posed to biodiversity [5-58 7], and for shedding light on macroevolutionary dynamics [8-11]. 59 Here, we unite palaeontological and neontological data [12] to test niche stability 60 across three million years of environmental changes using ten phylogenetically, 61 ecologically, and developmentally diverse bivalve and gastropod species from the 62 Atlantic and Gulf Coastal Plains, USA, and surrounding region (electronic supplementary 63 material, table S1). Species' niches were quantified using ecological niche modelling [13] 64 for three time periods from the Pliocene-recent: the mid-Pliocene Warm Period 65 (mPWP; ~3.264–3.025 Ma); the Eemian Last Interglacial (LIG; ~130–123 Ka); and the 66 present-day interval (PI). We test whether these species' niches changed across both long 67 (Pliocene to Eemian; millions of years) and short (Eemian to present-day; thousands of 68 years) time scales. We additionally investigate whether these species' upper and lower 69 thermal tolerances changed across millions of years. Recent research suggests that 70 tolerances to heat are largely conserved within terrestrial species, but that tolerances to 71 cold are more variable [14]. This asymmetry is thought to diminish in the marine realm,

72 where ectotherms are limited by both cold and warm conditions due to decreased aerobic 73 capacity [15]. This study is the first to incorporate both modern and fossil data across 74 millions of years to understand ecological and evolutionary responses of species to 75 changes in their environment, though see [16-18] for analyses in deep time. 76 Theoretical [19, 20] and empirical studies have both supported [21, 22] and 77 questioned [16, 23, 24] niche stability. The debate has even continued at the genetic level, 78 where recent research indicates that genetic reshuffling in Drosophila species can occur 79 in response to climate change [25, 26]. Whether these genetic changes translate into 80 evolution of actual physiological tolerances, however, remains unclear. The context in 81 which niche evolution is considered is important with respect to whether change occurred 82 in actual physiological tolerances (i.e., the fundamental niche; FN), or whether change 83 occurred because of differences in resource utilization or underlying environmental 84 structure (i.e., changes in the realized niche; RN). Studies may incorrectly indicate niche 85 evolution if the environmental conditions that are available to a species are not taken into 86 account [4, 27, 28]. 87 The aforementioned studies have contributed much to our understanding of how

species' environmental tolerances evolve, but questions about the relative dominance of
niche evolution *versus* stability remain, particularly since most studies lack a temporal
component that would allow for analysis of change across the entire duration of a
species' lifetime, which may span millions of years [8].

The region encompassing and surrounding the Atlantic and Gulf Coastal Plains is ideal for elucidating the coevolution of species' niches and the environment. Not only has it experienced profound environmental changes associated with the closure of the Central American Seaway beginning in the Pliocene [29], but these environmental changes have

96 been linked to patterns of extinction, species turnover, and ecological change [30, 31]. 97 The mPWP is considered a climatic analogue for conditions expected at the end of this 98

99 climate scenarios [32]. Results such as those presented here are vital for proper mitigation

century and can contribute information on how target species may fare under future

100 of the risks posed by current and future climate changes to Earth's biodiversity [7, 33].

101

102 2. Materials and methods

103 In order to test for within-lineage niche stability, we used ecological niche 104 modelling (ENM), a correlative process whereby known occurrences of species are 105 associated with environmental parameters to characterize a species' environmental 106 requirements [13]. Models of species' abiotic niche parameters were constructed for each 107 of three temporal intervals—the mPWP, LIG, and PI—using taxon occurrence data and 108 environmental parameters unique to each time slice. The resulting niche estimates were 109 compared through time to statistically assess similarity using both environmental and 110 geographic approaches [28, 34, 35]. In both approaches, an observed similarity metric is 111 computed and compared to a simulated null distribution. Details of our methodology are 112 outlined below.

113

114 (a) Taxa

115 We selected ten species that occur in both the modern and fossil (from ~3.1 Ma to 116 recent) records of the Atlantic and Gulf Coastal Plains, USA, and surrounding region 117 (Table 1). Species were chosen because they have diverse phylogenetic positions, varied 118 ecological habits and larval developmental modes, and abundant distributional data 119 available from fossil and modern localities (electronic supplementary material, table S1).

We used morphological criteria to identify target species, as each taxon is readily diagnosable. All evidence suggests that these lineages represent species that have distinct evolutionary trajectories, a supposition supported by the fact that many invertebrate species have durations greater than three million years [8]. Consequently, we studied within-lineage rather than across-lineage niche evolution, although see the Discussion section for potential caveats.

126

127 (b) Distributional data

128 **Present day.** Presence-only distributional data were derived from [36] (electronic 129 supplementary material, table S1 and figures S1-3). Only records with spatial uncertainty 130 <15 km were retained, ensuring that they were matched correctly with corresponding 131 environmental data of a coarser spatial resolution (i.e., 1.25 x 1.25°) [37]. We 132 subsampled distributional data to leave one record per environmental pixel to account for 133 sampling biases in R.15.2 (R Core Team, 2012), which resulted in 20-58 unique 134 occurrences per species (electronic supplementary material, table S1). This process did 135 not affect the resultant overall distribution of the species, but rather prevented certain 136 localities with multiple records from being unduly weighted in the niche modelling 137 analyses [38, 39]. 138 *Fossil.* We considered fossil distributional data from mPWP (~3.264–3.025 Ma) 139 and LIG (~130–123 Ka) strata of the Atlantic and Gulf Coastal Plains, USA, and 140 surrounding region. To ensure distributional data were derived from geologic units of 141 similar ages to our periods of interest, we generated a stratigraphic database for all 142 Pliocene-recent geologic units of the Atlantic Coastal Plain (electronic supplementary 143 material, dataset S1). Correlations and unit ages were determined by extensive literature

survey and use of various stratigraphic databases, resulting in ten viable formations for
the Pliocene and 16 for the LIG. The formations from which occurrence data were
derived are documented in dataset S1.

147 Distributional records were obtained from onsite investigations of collections to 148 ensure proper species identification, including the Florida Museum of Natural History, 149 Paleontological Research Institution, Virginia Museum of Natural History, Academy of 150 Natural Sciences of Drexel University, and Yale Peabody Museum. As with present-day 151 distributional data, we subsampled fossil distributional data to leave one record per 152 environmental pixel, resulting in six to 16 unique occurrences per species (electronic 153 supplementary material, table S1). At least six spatially-explicit distributional records 154 were used for model calibration for any given species/time period; studies have shown 155 this number to be statistically robust for extant species [40, 41].

156

157 (c) Environmental data

158 Environmental data were derived from the coupled atmosphere-ocean HadCM3 159 global climate model (GCM) [42, 43] for three time slices: mPWP (~3.264–3.025 Ma), 160 LIG (~130–123 Ka), and PI (considering the pre-industrial interval from ~1850–1890). 161 Ideally, we would use an ensemble-modelling approach that considered multiple GCMs 162 [44]; however, model output for the LIG was available to us only from HadCM3 and 163 consisted of variations of temperature and salinity parameters. This GCM has been 164 successfully used in a variety of Quaternary and pre-Quaternary modelling studies [45-165 47]. Boundary conditions for the LIG were from [46] and [48]. Here, atmospheric gas 166 concentrations were derived from ice core records [49-51], and orbital parameters were from Berger and Loutre [52]. The mPWP GCM used the alternate PRISM3D PlioMIP 167

168 dataset [53], and the pre-industrial experiment was equivalent to [54]. All GCM

169 experiments were run for 500 model years, and environmental parameters were averaged 170 from the final 30 years of each experiment at 1.25 x 1.25° resolution (~140 x 140 km at 171 the equator). Where ocean data were unavailable (i.e., sites presenting macrofossil data, 172 but where the GCM indicated land), we used an inverse-distance weighted algorithm to 173 extrapolate model data.

174 Modelled monthly salinity and temperature outputs were converted to maximum, 175 minimum, and average yearly coverages for both surface and bottom conditions using 176 ArcGIS. From these 12 coverages, we eliminated variables that significantly co-varied 177 (assessed using the 'cor' function in R.15.2; R Core Team, 2012). Ultimately, two bottom 178 variables (yearly average salinity and temperature) and four surface variables (maximum 179 and minimum salinity, and maximum and minimum temperature) were retained. These 180 six variables were preserved because they did not significantly co-vary and are deemed 181 biologically important for marine ectotherms [55-57].

182

183 (d) Modelling algorithm

184 To approximate niche parameters for these species, we generated ENMs using 185 Maxent v.3.3.3 [58] (figure 1 and electronic supplementary material, figures S4-5). 186 Maxent finds suitable environmental combinations for species under a null expectation 187 that suitability is proportional to availability. Thus, Maxent minimized the relative 188 entropy of observed environments relative to those in the background [59]. We enabled 189 only quadratic features to simulate realistic bell-shaped response curves that are known 190 from physiological experiments of plants and animals [60-62]. We calibrated models 191 within a region bounded by the Americas and 34°W longitude, and 48°N and 44°S

latitude (figure 1). We sought the union of the area sampled by researchers and most
likely accessible to the species across spatial and temporal dimensions [13, 63, 64]. We
used all spatially-explicit data points for each species/time slice, running 100 bootstrap
replicates with a ten per cent random test percentage. The mean value of the suitability
grids was used to threshold to binary predictions [65, 66].

To correct for potential biases in fossil distributional data, we implemented what is called a 'bias file' within Maxent for past modelling [67]. The bias file describes the probability that an area was sampled; thus, regions with rock outcrop (i.e., areas where species may actually be sampled) were weighted twice as heavily as regions without rock outcrop. Maxent will then factor out this bias during the modelling process (see [67] for details). This method essentially accounts for incomplete knowledge of species'

distributions sensu [68].

204 Although characterizing the entirety of a species' FN is often difficult without 205 mechanistic studies [14], we study close approximations here, given that recent 206 biophysical approaches have determined that FNs can be represented by limited sets of 207 parameters such as temperature [69, 70]. This is particularly true for marine ectotherms, 208 which have been shown to closely match range limits within their thermal tolerances [15]. 209 That being said, our estimates may reflect some quantity between the RN and the FN, 210 since our niche parameters are ultimately derived from the areas occupied by a species 211 [13, 14, 27].

212

213 (e) Model verification

Two model validation methods were used, depending on the prevalence ofdistributional records (electronic supplementary material, table S2). For species/time

slices with <25 points, we assessed statistical significance using a jackknife procedure

217 under a least training presence threshold [41]. This method, however, may produce over-

218 optimistic estimates of predictive power for sample sizes >25, and these species/time

slices were tested using partial Receiver Operating Characteristic analyses [71].

220

221 (f) Niche comparisons

Characterizations of species' niches were compared through time using two statistical approaches: a kernel smoothing script [28] and ENMTools [35]. Both frameworks use randomization tests to compare observed similarity to that expected under a null hypothesis. The null is rejected if models are more or less similar than expected by chance, based on the environment within the geographical regions of interest. Similarity is quantified using Schoener's *D* [72], with values ranging from 0 to 1, or more to less similar, respectively.

229 For each of the ten species, we compared observed niches across the three 230 different time periods (mPWP, LIG, and PI). Comparisons were made in two directions 231 [28, 35]; e.g., comparing the mPWP to the LIG, and the LIG to the mPWP, since it is 232 possible for two niches to be more similar than expected based on the environment 233 available for one time slice, but less similar than expected based on the environment 234 available for the other. If the observed value fell outside the null distribution to the high 235 end, niches were more similar than expected by chance, whereas if the observed value 236 fell outside the null distribution to the lower end, niches were more different than 237 expected by chance. Observed values that fell within the null distribution did not allow 238 for discrimination of similarity or differences based on the environment available to the 239 entities in question. We performed similarity tests using (1) a PCA applied to all six

240 environmental variables; (2) a PCA applied to the three most important environmental 241 variables; (3) raw average bottom temperature and maximum surface temperature in two-242 dimensional environmental space; (4) maximum surface temperature only; (5) minimum 243 surface temperature only; and (6) ENMTools on projections of ecological niche models. 244 The first five sets of tests compare niches in environmental space, with the first three 245 multi-dimensional in nature, whereas the sixth compares niches in geographic space. 246 Each of these tests resulted in 60 comparisons (i.e., 10 species x three time slices x two 247 directions), for a total of 360. Details of the comparisons are provided below.

248 *Environmental comparisons.* We calculated metrics of niche overlap in gridded 249 environmental space using the methodology of [28]. Here, ordination techniques [73] 250 allow for direct comparison of species-environment relationships in environmental space 251 [27]. Observed densities for each region are corrected in light of the availability of 252 environmental space using kernel density functions (table 1 and electronic supplementary 253 material, table S3 and dataset S2). Niche overlap is measured along gradients of a 254 multivariate analysis, and statistical significance is assessed using the framework 255 described above.

256 We tested for similarity using a principal component analysis (PCA) (1) applied 257 to all six environmental parameters, and (2) when niche dimensionality was reduced to 258 three variables, including surface coverages for maximum salinity, maximum 259 temperature, and minimum temperature. These variables were retained because they 260 explained the most variance in the dataset [57, 74, 75]. Analyses performed with this 261 reduced set of variables are potentially more informative, as over-parameterization can 262 constrict niche estimates and lead to approximations closer to the RN [13]. PCA analyses 263 were calibrated on environments of both time slices (setting project equal to false). We

264 used both the PCA-occ and PCA-env functions; the former calibrates the PCA based only 265 on the distributional data, whereas the latter uses data from the entire environmental 266 space of the two study systems. The results were equivalent, and thus we present only 267 those from PCA-env. A bin size of 100 was used to characterize the environment, 268 running 1,000 replicates for similarity tests. Since prevalence of distributional data varies 269 through time, we generated input data from ENMs outside of the framework of [28], 270 subsampling one point per pixel in binary predictions such that comparisons were 271 unbiased with regard to the quantity of input data. Doing so ensures that we capture all of 272 the environments that a species finds suitable, rather than the portion that happened to be 273 occupied most frequently.

We also tested similarity in raw variables (table 1 and electronic supplementary material, table S3 and dataset S2). We used the script of [28] to analyse each of the six variables individually, and we modified the script to compare raw variables in two dimensions, while still accounting for differences in availability of environments in a given time period. We were interested in testing for evolution in overall temperature parameters, and thus we assessed similarity using average bottom temperature and maximum surface temperature.

Geographic projections. In addition to the comparisons made entirely in
environmental space, we used ENMTools [35] to compare the geographic projections of
niches. Null distributions consisted of 100 random models generated within Maxent, with
model parameters drawn from and constrained by the study system. To ensure accurate
response curves when projecting, we disabled clamping and enabled extrapolation within
Maxent [76].

287

288 **3. Results**

Model verification exercises suggest that models of species' niches are statistically significant for each time slice (P<0.05; see electronic supplementary material, table S2). The niche model depictions are shown in figure 1 and electronic supplementary material, figures S4-5.

293 Together, the suite of niche comparisons (360 in total) indicates these species' 294 environmental preferences are stable across millions of years. In 359 of 360 cases, we 295 found no evidence of niche dissimilarity across all comparisons. Indeed, of the ten 296 ecologically diverse species studied, nine show the opposite pattern: statistically similar 297 niches for the majority of the comparisons. Probabilistically, this result would be 298 obtained < 1% of the time, assuming equal likelihood for evolution *versus* stability of 299 niche attributes. We obtain evidence of niche similarity for tests on both principle 300 component analyses (PCAs) and raw variables. Moreover, minimum and maximum 301 temperature tolerances are generally conserved through time.

302

303 (a) Environmental comparisons

304 Comparisons on multi-dimensional niches indicate overwhelming signals of niche 305 stability across the time slices. Of these 180 comparisons, 149 indicate statistically 306 similar niches through time, and no comparison found evidence of niche dissimilarity. 307 Comparisons considering all six environmental variables indicate niches are 308 statistically similar for most species and time slices (46 of 60 comparisons) (electronic 309 supplementary material, table S3). When niche dimensionality was reduced to the most 310 important variables, nine species show statistically similar niches for all comparisons, 311 with the exception of one or two inconclusive tests for Crepidula fornicata, Dinocardium

312 robustum, Lucina pensylvanica, and Neverita duplicata (49 of 60 comparisons; figure 2 313 and table 1). Bulla occidentalis is the only species with non-significant tests across 314 multiple time slices. This species does not have any readily identifiable traits—such as 315 larval strategy or feeding ecology-that would predispose it to occupying new 316 environments relative to the other species that we studied. Niches also show stability 317 when raw variables are considered. Seven of the ten species have statistically similar 318 niches across all time comparisons (42 of 60 comparisons; electronic supplementary 319 material, table S3). Two other species, Oliva sayana and Crassostrea virginica, have 320 statistically similar niches with the exception of one and two inconclusive tests, 321 respectively. Quantifying niche similarity for *B. occidentalis* proved more difficult, as 322 three of six niche comparisons are non-significant (but not statistically different). 323 Species seem to conserve their upper thermal tolerance limits, but results are less 324 conclusive for minimum temperature tolerances (table 1 and electronic supplementary 325 material, dataset S2). Across the suite of species, the majority of comparisons are 326 statistically similar with regard to maximum surface temperature, although five species 327 have one or two comparisons that are inconclusive (B. occidentalis, D. robustum, L. 328 pensylvanica, N. duplicata, O. sayana, and Terebra dislocata). Comparisons also indicate 329 statistical similarity with regard to minimum temperature tolerances. However, the 330 structure of this variable changes through time, making it difficult to quantify similarities 331 or differences. For example, all mPWP–LIG comparisons are inconclusive with the 332 exception of N. duplicata, as are at least half of the comparisons for B. occidentalis and L. 333 pensylvanica.

334

335 (b) Geographic comparisons

336 Results from comparisons of the geographic projections of niches mirror those 337 from the environmental comparisons. Niches are statistically similar for seven of the ten 338 species across all comparisons (42 of 60 comparisons; electronic supplementary material, 339 table S3 and dataset S2). Crassostrea virginica and L. pensylvanica have one comparison 340 that is inconclusive (LIG-mPWP and PI-mPWP, respectively), while the niche of B. 341 occidentalis is significantly dissimilar for the LIG-mPWP comparison and non-342 significant for the PI-mPWP comparison. 343 344 4. Discussion 345 Our analyses find no support for niche evolution. Instead, we observe statistically

346 significant niche stability across three million years of considerable environmental 347 changes, from extreme warmth during the mPWP to glacial cycles during the Pleistocene 348 [29]. This is true for all of ten of the species analysed. Importantly, niche stability will 349 not be recovered within analyses for reasons other than similarity, whereas niche 350 differences can be obtained as a function of changing parameters of the RN [14]. 351 Therefore, the lack of any net change suggests that species were either shifting their niche 352 preferences in response to oscillating climatic conditions at scales too rapid to be detected 353 by our analyses, or their preferences remained stable across this temporal interval. In 354 either case, overall niche stability has profound implications for understanding 355 conservation priorities and for elucidating macroevolutionary dynamics. 356 357 (a) Implications for survival of taxa during times of change 358 These results aid our understanding of how species may respond to climate 359 change on both long and short time scales. As climate continues to change, species that

360 are unable to adapt to new conditions face two futures: extinction or shifting distributions 361 to follow suitable areas. Already, both responses have been documented or predicted as a 362 result of current climate change. Marine and terrestrial species are forecast to experience 363 climate-driven extinctions into the 22nd century [77, 78]. Indeed, the niche stability we 364 have documented may doom many marine species to extinction over the next 100+ years, 365 particularly if they live at their thermal tolerance limits and are unable to alter their upper 366 thresholds [57]. The target species considered here are predicted to experience severe 367 distributional reductions by the end of this century when variables other than temperature 368 and salinity are considered, but wholesale extinction is unlikely [36]. This prediction is 369 supported by their survival in the Pliocene, albeit in geographically-reduced areas, when 370 conditions were purportedly similar to those expected at the end of this century [32]. 371 These small areas of suitability—or refugia—are thought to have played an important 372 role in species' survival during past episodes of climate change [79]. 373 If species are able to keep pace with the changing environment, distributional 374 shifts, rather than extinctions, are expected [33]. Under this scenario, dispersal ability 375 becomes an important parameter predicting species' responses to climate change [80]. 376 Present-day elevational, latitudinal, and bathymetric shifts [81] have already been 377 observed in response to current warming patterns, and, indeed, the fossil record provides 378 abundant evidence for habitat tracking during rapid Pleistocene climate cycles [82], often 379 creating non-analogous community assemblages [83]. The rate at which climate changes 380 also dictates whether species can track preferred environments, and future rates are 381 anticipated to exceed those experienced during the geologic intervals analysed within this 382 study [57, 84, 85]. In a rapidly changing world, species will most likely be forced to 383 move to suitable areas or face extinction, since it seems unlikely they will alter their

abiotic preferences on extremely short time scales if they are unable to do so on longertime scales, as we demonstrated here.

386	Methodologically, niche stability provides support for ENM and species
387	distribution modelling (SDM) analyses that attempt to predict how species will respond
388	to altered climatic conditions [13]. In particular, our results may somewhat alleviate
389	concerns over inaccurate forecasts due to changing niches [1, 3]. Problems still remain,
390	however, in that ENM and SDM methods typically do not account for dispersal
391	limitations or altered biotic interactions [86], though see [84], nor do they consider that
392	species can alter their behaviour or microhabitat preferences to buffer against
393	environmental changes [2, 87].
394	
395	(b) Macroevolutionary implications of stable niches
396	We show that large-scale parameters of species' niches, in this case temperature
397	and salinity, do not change for a phylogenetically and ecologically diverse set of marine
398	molluscs. Although species may modify their behaviour or resource utilization, the FN
399	places constraints on species' interactions with the environment, which potentially
400	governs speciation and extinction processes over long time scales [10, 88]. Some
401	researchers have suggested that niche stability may promote allopatric speciation [89, 90].
402	That is, environmental perturbations may separate two populations, with those
403	populations prevented from merging back together because of constraints imposed by the
404	FN, which will then eventually lead to diversification.
405	Niche stability also provides a potential mechanism for the morphological stasis
406	observed within species over millions of years [8]. More specifically, niche stability
407	requires species to track preferred habitats as the environment changes, thereby

408 continuously joining and separating populations on scales less than 10,000 years or so. In
409 this framework, any localized phenotypic adaptation is unlikely to be fixed across an
410 entire species, such that no overall net changes are observed for the species as a whole [8].
411

412 (c) Potential caveats

413 Although our analyses are quantitatively robust, our study is not without 414 limitations. First, our models may approximate the existing or realized niche, rather than 415 the FN [91], because FNs are difficult to characterize without detailed physiological 416 studies [13, 14]. With that said, niche estimates were calculated from environmental 417 preferences that were averaged over a period of time, which may broaden estimates such 418 that real physiological limits are captured [57]. The recovered pattern of niche stability is 419 even more robust if we studied RNs, since change is expected to occur over time in RN 420 parameters owing to differences in resource utilization or underlying environmental 421 structure [4, 13, 27]. Second, estimates of present-day and past niches may not be 422 equivalent and thus incomparable. This, however, is of less concern here since we 423 documented niche stability rather than niche evolution. Third, we acknowledge that 424 recognition of 'species'-especially in the fossil record-is sometimes contentious, and 425 while these species are diagnosably distinct throughout their duration, they may not 426 constitute single evolutionary lineages. Our results, however, are even more robust if we 427 studied aggregated collections of closely-related lineages, since we would expect more 428 change in niche parameters at speciation. We support conservatism of niches across 429 speciation events if the entities in question represent closely-related species complexes. 430 Fourth, we analysed data from warm time periods, as distributional data do not exist for 431 glacial periods (e.g., the last glacial maximum; ~ 21 Ka). Therefore, our analyses may

432	have missed rapid (but reversible) niche evolution that occurred in response to these
433	colder conditions. Although possible, the scenario is unlikely because of the rate at which
434	niche evolution would have had to occur, and because of the paucity of evidence for
435	niche adaptation both in the fossil record [82] and in experimental studies [14]. Moreover,
436	environmental conditions at the mPWP, LIG, and PI differ to a significant degree, such
437	that we are still able to discern whether species adapted to new conditions or tracked
438	stable climate envelopes. Finally, and related to this issue, because palaeoclimate models
439	were only available for certain key temporal intervals, we could not capture the entire
440	temporal history of these species in the context of an ENM framework. We did, however,
441	examine changes across both long (mPWP to LIG) and short (LIG to PI) time scales.

443 Acknowledgements

444 We are grateful to Alycia Stigall, Geerat Vermeij, Gary Carvalho, Norman MacLeod, and 445 an anonymous reviewer for improving the quality of this contribution. We thank the following individuals for museum collection help: Lauck Ward and Alton Dooley 446 (Virginia Museum of Natural History), Judith Nagel-Myers, Greg Dietl, David Campbell 447 and Warren Allmon (Paleontological Research Institution), Paul Callomon (The 448 Academy of Natural Sciences of Drexel University), Jessica Utrup (Yale Peabody 449 450 Museum), and Alex Kittle and Sean Roberts (Florida Museum of Natural History). We also thank numerous geologists for discussions on stratigraphy, including: Lauck Ward 451 452 (Virginia Museum of Natural History), Christopher Williams and Guy Means (Florida 453 Geological Survey), Paul Huddlestun (Georgia Geological Survey), Helaine Markewich 454 (U.S. Geological Survey), Kathleen Farrell (North Carolina Geological Survey), John 455 Wehmiller (University of Delaware), Ervin Otvos (University of Southern Mississippi), 456 and Lyell Campbell (University of South Carolina Upstate). We benefited from 457 discussion of ENM with Naravani Barve, Andres Lira, A. Townsend Peterson, Jorge Soberón, and the KU ENM Working Group (University of Kansas). Olivier Brönnimann 458 459 is thanked for assistance with his niche overlap script. An NSF GK-12 Fellowship, KU 460 Madison and Lila Self Graduate Fellowship, KU Biodiversity Institute Panorama Grant, 461 and John W. Wells Grant-in-Aids of Research provided funding for this work to EES. 462 BSL was supported by NSF grant EF-1206757. A.M.H. and S.H. acknowledge that the research leading to these results received funding from the European Research Council 463 464 under the European Union's Seventh Framework Programme (FP7/2007-2013)/ERC 465 grant agreement no. 278636. A.M.H and S.H thank Paul Valdes and Jov Singaraver for 466 making available LIG model boundary conditions.

468 Data accessibility

469 The stratigraphic database (dataset S1) and output from niche comparison tests (dataset

470 S2) are available in the electronic supplementary material. Climate and distributional data

471 are available on Dryad, doi: XXX.

472

473 References

Pearman P.B., Guisan A., Broennimann O., Randin C.F. 2008 Niche dynamics
 in space and time. *Trends Ecol Evol* 23, 149–158.
 Lavergne S., Mouquet N., Thuiller W., Ronce O. 2010 Biodiversity and climate
 change: integrating evolutionary and ecological responses of species and

- 478 communities. *Annu Rev Ecol Evol Syst* **41**, 321–350.
- 479 3. Hoffmann A.A., Sgrò C.M. 2011 Climate change and evolutionary adaptation.
 480 *Nature* 470, 479–485.
- 481 4. Guisan A., Petitpierre B., Broennimann O., Daehler C., Kueffer C. 2014
 482 Unifying niche shift studies: insights from biological invasions. *Trends Ecol Evol* 29, 260–269.
- 484 5. Pereira H.M., Leadley P.W., Proença V., Alkemade R., Scharlemann J.P.W.,
 485 Fernandez-Manjarrés J.F., Araújo M.B., Balvanera P., Biggs R., Cheung W.W.L., et al.
 486 2010 Scenarios for global biodiversity in the 21st Century. *Science* 330, 1496–1501.
- 487 6. Dawson T.P., Jackson S.T., House J.I., Prentice I.C., Mace G.M. 2011 Beyond
 488 predictions: biodiversity conservation in a changing climate. *Science* 332, 53–58.
 489 7. Moritz C., Agudo R. 2013 The future of species under climate change:
- 490 resilience or decline? *Science* **341**, 504–508.
- 8. Eldredge N., Thompson J.N., Brakefield P.M., Gavrilets S., Jablonski D., Jackson
 J.B.C., Lenski R.E., Lieberman B.S., McPeek M.A., Miller W.I. 2005 The dynamics of
 evolutionary stasis. *Paleobiology* **31**, 133–145.
- 494 9. Romdal T., Araújo M.B., Rahbek C. 2013 Life on a tropical planet: niche
 495 conservatism explains the global diversity gradient. *Glob Ecol Biogeogr* 22, 344–350.
 496 10. Jablonski D., Belanger C.L., Berke S.K., Huang S., Krug A.Z., Roy K.,
- Tomasovych A., Valentine J.W. 2013 Out of the tropics, but how? Fossils, bridge
 species, and thermal ranges in the dynamics of the marine latitudinal diversity
 gradient. *Proc Natl Acad Sci* **110**, 10487–10494.
- 500 11. Vermeij G.J. 1991 When biotas meet: understanding biotic interchange.
 501 *Science* 253, 1099–1104.
- 502 12. Fritz S.A., Schnitzler J., Eronen J.T., Hof C., Böhning-Gaese K., Graham C.H.
 503 2013 Diversity in time and space: wanted dead and alive. *TREE* 28, 509–516.

504 13. Peterson A.T., Soberón J., Pearson R.G., Anderson R.P., Martínez-Meyer E., 505 Nakamura M., Araújo M.B. 2011 Ecological Niches and Geographic Distributions. 506 Princeton. Princeton University Press. 507 Araújo M.B., Ferri-Yanez F., Bozinovic F., Marguet P.A., Valladares F., Chown 14. 508 S.L. 2013 Heat freezes niche evolution. *Ecol Lett* **16**, 1206–1219. 509 (doi:10.1111/ele.12155). 510 15. Sunday J.M., Bates A.E., Dulvy N.K. 2011 Global analysis of thermal tolerance 511 and latitude in ectotherms. *Proc R Soc B* **278**, 1823–1830. 512 16. Stigall A.L. 2012 Using ecological niche modelling to evaluate niche stability 513 in deep time. *J Biogeogr* **39**, 772–781. 514 Stigall A.L. 2014 When and how do species achieve niche stability over long 17. 515 time scales? *Ecography*. (doi:10.1111/ecog.00719). 516 Malizia R.W., Stigall A.L. 2011 Niche stability in Late Ordovician articulated 18. 517 brachiopod species before, during, and after the Richmondian invasion. *Palaeogeogr* 518 Palaeoclimatol Palaeoecol **311**. 154–170. 519 19. Holt R.D. 1996 Adaptive evolution in source-sink environments: direct and 520 indirect effects of density-dependence on niche evolution. *Oikos* 75, 182–192. 521 20. Kawecki T.J. 1995 Demography of source-sink populations and the evolution 522 of ecological niches. Evol Ecol 9, 38-44. 523 Martínez-Meyer E., Peterson A.T. 2006 Conservatism of ecological niche 21. 524 characteristics in North American plant species over the Pleistocene-to-Recent 525 transition. *J Biogeogr* 33, 1779–1789. 526 Strubble D., Broennimann O., Chiron F., Matthysen E. 2013 Niche 22. 527 conservatism in non-native birds in Europe: niche unfilling rather than niche 528 expansion. *Glob Ecol Biogeogr* **22**, 962–970. 529 Broennimann O., Treier U.A., Müller-Schärer H., Thuiller W., Peterson A.T., 23. 530 Guisan A. 2007 Evidence of climatic niche shift during biological invasion. Ecol Lett 531 **10**. 701–709. 532 24. Rödder D., Lötters S. 2009 Niche shift versus niche conservatism? Climatic 533 characteristics of the native and invasive ranges of the Mediterranean house gecko 534 (Hemidactylus turcicus). Glob Ecol Biogeogr 18, 674–687. 535 Umina P.A., Weeks A.R., Kearney M.R., McKechnie S.W., Hoffmann A.A. 2005 A 25. 536 rapid shift in a classic clinal pattern in Drosophila reflecting climate change. Science 537 **308**, 691–693. 538 26. Balanyá J., Oller J.M., Huey R.B., Gilchrist G.W., Serra L. 2006 Global genetic 539 change tracks global warming in *Drosophila subobscura*. Science **313**, 1773–1775. 540 Araújo M.B., Peterson A.T. 2012 Uses and misuses of bioclimatic envelope 27. 541 modeling. *Ecology* **93**, 1527–1539. 542 28. Broennimann O., Fitzpatrick M.C., Pearman P.B., Petitpierre B., Pellissier L., Yoccoz N.G., Thuiller W., Fortin M.-J., Randin C., Zimmermann N.E., et al. 2012 543 544 Measuring ecological niche overlap from occurrence and spatial environmental data. 545 *Glob Ecol Biogeogr* **21**, 481–497. 546 29. Cronin T.M. 1988 Evolution of marine climates of the U.S. Atlantic Coast 547 during the past four million years. *Phil Trans R Soc B* **318**, 661–678.

- 548 Todd J.A., Jackson J.B.C., Johnson K.G., Fortunato H.M., Heitz A., Alvarez M., 30. 549 Jung P. 2002 The ecology of extinction: molluscan feeding and faunal turnover in the 550 Caribbean Neogene. *Proc R Soc B* **269**, 571–577. 551 Vermeij G.J. 2009 One-way traffic in the western Atlantic: causes and 31. 552 consequences of Miocene to early Pleistocene molluscan invasions in Florida and 553 the Caribbean. *Paleobiology* **31**, 624–642. 32. 554 Robinson M.M., Dowsett H.J. 2008 Pliocene role in assessing future climate 555 impacts. EOS, Trans Am Geo Un 89, 501-502. 556 33. Warren R., VanDerWal J., Price J., Welbergen J.A., Atkinson I., Ramirez-557 Villegas J., Osborn T.J., Jarvis A., Shoo L.P., Williams S.E., et al. 2013 Quantifying the 558 benefit of early climate change mitigation in avoiding biodiversity loss. *Nature Clim* 559 *Change* **3**, 678–682. 560 Warren D.L., Glor R.E., Turelli M. 2008 Environmental niche equivalency 34. 561 versus conservatism: Quantitative approaches to niche evolution. Evolution 62, 562 2868-2883. 563 35. Warren D.L., Glor R.E., Turelli M. 2010 ENMTools: a toolbox for comparative 564 studies of environmental niche models. *Ecography* **33**, 607–611. 565 36. Saupe E.E., Hendricks J.R., Townsend A.T., Lieberman B.S. 2014 Climate 566 change and marine molluscs of the western North Atlantic: future prospects and 567 perils. J Biogeogr 41, 1352–1366. 568 Graham C.H., Elith J., Hijmans R.J., Guisan A., Peterson A.T., Loiselle B.A., 37. 569 Group T.N.P.S.D.W. 2008 The influence of spatial errors in species occurrence data 570 used in distribution models. *J Appl Ecol* **45**, 239–247. 571 Royle J.A., Chandler R.B., Yackulic C., Nichols J.D. 2012 Likelihood analysis of 38. 572 species occurrence probability from presence-only data for modelling species 573 distributions. *Methods Ecol Evol* **3**, 545–554. 574 Yackulic C.B., Chandler R., Zipkin E.F., Royle J.A., Nichols J.D., Campbell Grant 39. 575 E.H., Veran S. 2013 Presence-only modelling using MAXENT: when can we trust the 576 inferences? Methods Ecol Evol 4, 236-243. 577 40. Hernandez P.A., Graham C.H., Master L.L., Albert D.L. 2006 The effect of 578 sample size and species characteristics on performance of different distribution 579 modeling methods. *Ecography* **29**, 773–785. 580 Pearson R.G., Raxworthy C.I., Nakamura M., Peterson A.T. 2007 Predicting 41. 581 species distributions from small numbers of occurrence records: a test case using 582 cryptic geckos in Madagascar. *J Biogeogr* **34**, 102–117. 583 Gordon C., Cooper C., Senior C.A., Banks H., Gregory J.M., Johns T.C., Mitchell 42. 584 J.F.B., Wood R.A. 2000 The simulation of SST, sea ice extents and ocean heat 585 transports in a version of the Hadley Centre coupled model without flux 586 adjustments. ClDy 16, 147–168. 587 Pope V.D., Gallani M.L., Rowntree P.R., Stratton R.A. 2000 The impact of new 43. 588 physical parameterizations in the Hadley Centre Climate model: HadAM3. ClDy 16, 589 123-146. 590 44. Fordham D.A., Wigley T.M.L., Watts M.J., Brook B.W. 2012 Strengthening 591 forecasts of climate change impacts with multi-ensemble averaged projections using
- 592 MAGICC/SCENGEN 5.3. *Ecography* **45**, 4–8.

593 Eriksson A., Betti L., Friend A.D., Lycett S.J., Singarayer J.S., von Cramon-45. 594 Taubadel N., Valdes P.J., Balloux F., Manica A. 2012 Late Pleistocene climate change 595 and the global expansion of anatomically modern humans. *Proc NatlAcad Sci* **109**, 596 16089-16094. 597 46. Singarayer J.S., Valdes P.J. 2010 High-latitude climate sensitivity to ice-sheet 598 forcing over the last 120 kyr. *Quat Sci Rev* 29, 43–55. 599 Haywood A.M., Ridgwell A., Lunt D.J., Hill D.J., Pound M.J., Dowsett H.J., Dolan 47. 600 A.M., Francis J.E., Williams M. 2011 Are there pre-Quaternary geological analogues 601 for a future greenhouse gas-induced global warming? Phil Trans R Soc A 369, 933-602 956. 603 48. Singarayer J.S., Valdes P.J., Friedlingstein P., Nelson S., Beerling D.J. 2011 Late 604 Holocene methane rise caused by orbitally controlled increase in tropical sources. 605 *Nature* **470**, 82–85. 606 49. Petit J.R., Jouzel J., Raynaud D., Barkov N.I., Barnola J.-M., Basile I., Bender M., 607 Chappellaz J., Davis J., Delaygue G., et al. 1999 Climate and atmospheric history of 608 the past 420,000 years from the Vostok Ice Core, Antarctica. *Nature* **399**, 429–436. 609 50. Loulergue L., Schilt A., Spahni R., Masson-Delmotte V., Blunier T., Lemieux B., 610 Barnola J.-M., Raynaud D., Stocker T.F., Chappellaz J. 2008 Orbital and millennial-611 scale features of atmospheric CH₄ over the past 800,000 years. *Nature* **453**, 383–386. 612 Spahni R., Chappellaz J., Stocker T.F., Loulergue L., Hausammann G., 51. 613 Kawamura K., Fluckiger J., Schwander J., Raynaud D., Masson-Delmotte V., et al. 2005 Atmospheric methane and nitrous oxide of the Late Pleistocene from Antarctic ice 614 615 cores. Science 310, 1317-1321. 616 52. Berger A., Loutre M.F. 1991 Insolation values for the climate of the last 10 617 million years. Quat Sci Rev 10, 297–317. 618 53. Haywood A.M., Dowsett H.J., Robinson M.M., Stoll D.K., Dolan A.M., Lunt D.J., 619 Otto-Bliesner B., Chandler M.A. 2011 Pliocene Model Intercomparison Project 620 (PlioMIP): experimental design and boundary conditions (Experiment 2). 621 *Geoscientific Model Development* **4**, 571–577. 622 54. Braconnot P., Otto-Bliesner B., Harrison S., Joussaume S., Peterchmitt J.-Y., Abe-Ouchi A., Crucifix M., Driesschaert E., Fichefet T., Hewitt C.D., et al. 2007 Results 623 624 of PMIP2 coupled simulations of the Mid-Holocene and Last Glacial Maximum – Part 625 1: experiments and large-scale features. *CliPa* **3**, 261–277. 626 55. Tomašových A., Kidwell S.M. 2009 Preservation of spatial and environmental 627 gradients by death assemblages. *Paleobiology* **35**, 119–145. 628 Buckley L.B., Hurlbert A.H., Jetz W. 2012 Broad-scale ecological implications 56. 629 of ectothermy and endothermy in changing environments. *Glob Ecol Biogeogr* 9, 630 873-885. 631 57. Sunday J.M., Bates A.E., Dulvy N.K. 2012 Thermal tolerance and the global 632 redistribution of animals. *Nature Clim Change* **2**, 686–690. 633 58. Phillips S.I., Anderson R.P., Schapire R.E. 2006 Maximum entropy modeling of 634 species geographic distributions. *Ecol Modell* **190**, 231–259. Elith J., Phillips S.J., Hastie T., Dudik M., Chee Y.E., Yates C.J. 2011 A statistical 635 59. 636 explanation of MaxEnt for ecologists. *Divers Distrib* **17**, 43–57.

637 Austin M.P., Nicholls A.O., Doherty M.D., Meyers J.A. 1994 Determining 60. 638 species response functions to an environmental gradient by means of a beta-639 function. *J Veg Sci* **5**, 215-228. 640 Angilletta M. 2009 Thermal Adaptation: A Theoretical and Empirical Synthesis. 61. 641 Oxford, Oxford University Press. 642 Hooper H.L., Connon R., Callaghan A., Fryer G., Yarwood-Buchanan S., Biggs J., 62. 643 Maund S.J., Hutchinson T.H., Sibly R.M. 2008 The ecological niche of *Daphnia magna* 644 characterized using population growth rate. *Ecology* **89**, 1015–1022. 645 63. Phillips S.J., Dudík M., Elith J., Graham C.H., Lehmann A., Leathwick J., Ferrier S. 646 2009 Sample selection bias and presence-only distribution models: implications for 647 background and pseudo-absence data. *Ecol Appl* **19**, 181–197. 648 VanDerWal J., Shoo L.P., Graham C., Williams S.E. 2009 Selecting pseudo-64. 649 absence data for presence-only distribution modeling: how far should you stray 650 from what you know? Ecol Modell 220, 589–594. 651 Liu C., Berry P.M., Dawson T.P., Pearson R.G. 2005 Selecting thresholds of 65. 652 occurrence in the prediction of species distributions. *Ecography* **28**, 385–393. 653 Freeman E.A., Moisen G.G. 2008 A comparison of the performance of 66. 654 threshold criteria for binary classification in terms of predicted prevalence and 655 kappa. *Ecol Modell* **217**, 48–58. 656 Dudík M., Schapire R.E., Phillips S.J. 2005 Correcting sample selection bias in 67. 657 maximum entropy density estimation. In Adv Neural Inf Process Syst (pp. 323–330, 658 The MIT Press. 659 68. Svenning J.-C., Fløjgaard C., Marske K.A., Nógues-Bravo D., Normand S. 2011 660 Applications of species distribution modeling to paleobiology. Quat Sci Rev 30, 661 2930-2947. 662 69. Kearney M.R., Simpson S.J., Raubenheimer D., Kooijman S.A.L.M. 2013 663 Balancing heat, water and nutrients under environmental change: a thermodynamic framework. Funct Ecol 27, 950–966. 664 665 70. Kearney M.R., Wintle B.A., Porter W.P. 2010 Correlative and mechanistic 666 models of species distribution provide congruent forecasts under climate change. 667 Conserv Lett 3, 203–213. 668 Peterson A.T., Papes M., Soberón J. 2008 Rethinking receiver operating 71. 669 characteristic analysis. *Ecol Modell* **213**, 63–72. 670 72. Schoener T.W. 1968 Anolis lizards of Bimini: resource partitioning in a 671 complex fauna. *Ecology* **49**, 704–726. Hof C., Rahbek C., Araújo M.B. 2010 Phylogenetic signals in the climatic 672 73. 673 niches of the world's amphibians. *Ecography* **33**, 242–250. 674 74. Tewksbury J.J., Huey R.B., Deutsch C.A. 2008 Putting the heat on tropical 675 animals. *Science* **320**, 1296–1297. 676 75. Tunnell J.W., Andrews J., Barrera N.C., Moretzsohn F. 2010 Encyclopedia of Texas Seashells: Identification, Ecology, Distribution & History. College Station, Texas 677 678 A&M University Press. 679 Owens H.L., Campbell L.P., Dornak L.L., Saupe E.E., Barve N., Soberón J., 76. Ingenloff K., Lira-Noriega A., Hensz C.M., Myers C.E., et al. 2013 Constraints on 680 681 interpretation of ecological niche models by limited environmental ranges on 682 calibration areas. Ecol Modell 263, 10–18.

683	77. Bijma J., Pörtner HO., Yesson C., Rogers A.D. 2013 Climate change and the						
684	oceans – what does the future hold? <i>Mar Pollut Bull</i> 74 , 495–505.						
685	78. Zu Ermgassen P.S.E., Spalding M.D., Blake B., Coen L.D., Dumbauld B., Geiger						
686	S., Grabowski J.H., Grizzle R., Luckenbach M., McGraw K., et al. 2012 Historical						
687	ecology with real numbers: past and present extent and biomass of an imperilled						
688	estuarine habitat. <i>Proc R Soc B</i> 279 , 3393–3400.						
689	79. Willis K.J., MacDonald G.M. 2011 Long-term ecological records and their						
690	relevance to climate change predictions for a warmer world. Annu Rev Ecol Evol Syst						
691	42 , 267–287.						
692	80. Trakhtenbrot A., Nathan R., Perry G., Richardson D.M. 2005 The importance						
693	of long-distance dispersal in biodiversity conservation. <i>Divers Distrib</i> 11 , 173–181.						
694	81. Chen IC., Hill J.K., Ohlemüller R., Roy D.B., Thomas C.D. 2011 Rapid range						
695	shifts of species associated with high levels of climate warming. Science 333, 1024-						
696	1026.						
697	82. Hof C., Levinsky I., Araújo M.B., Rahbek C. 2011 Rethinking species' ability to						
698	cope with rapid climate change. <i>Glob Change Biol</i> 17 , 2987–2990.						
699	83. Williams J.W., Jackson S.T. 2007 Novel climates, no-analog communities, and						
700	ecological surprises. <i>Front Ecol Environ</i> 5 , 475–482.						
701	84. Fordham D.A., Mellin C., Russell B.D., Akçakaya R.H., Bradshaw C.J.A., Aiello-						
702	Lammens M.E., Caley J.M., Connell S.D., Mayfield S., Shepherd S.A., et al. 2013						
703	Population dynamics can be more important than physiological limits for						
704	determining range shifts under climate change. <i>Glob Change Biol</i> 19 , 3224–3237.						
705	85. IPCC. 2013 Climate Change 2013: The Physical Science Basis. Geneva,						
706	Intergovernmental Panel on Climate Change Secretariat.						
707	86. Davis A.J., Jenkinson L.S., Lawton J.H., Shorrocks B., Wood S. 1998 Making						
708	mistakes when predicting shifts in species range in response to global warming.						
709	<i>Science</i> 391 , 783–786.						
710	87. Kearney M., Shine R., Porter W.P. 2009 The potential for behavioral						
711	thermoregulation to buffer "cold-blooded" animals against climate warming. Proc						
712	Natl Acad Sci 106 , 3835–3840.						
713	88. van Dam J.A., Aziz H.A., Álvarez Sierra M.Á., Hilgen F.J., van den Hoek Ostende						
714	L.W., Lourens L.J., Mein P., van der Meulen A.J., Palaez-Campomanes P. 2006 Long-						
715	period astronomical forcing of mammal turnover. <i>Nature</i> 443 , 687–691.						
716	89. Peterson A.T., Soberón J., Sanchez-Cordero V. 1999 Conservatism of						
717	ecological niches in evolutionary time. <i>Science</i> 285 , 1265–1267.						
718	90. Kozak K.H., Wiens J.J. 2006 Does niche conservatism promote speciation? A						
719	case study in North American salamanders. <i>Evolution</i> 60 , 2604–2621.						
720	91. Soberón J., Nakamura M. 2009 Niches and distributional areas: concepts,						
721	methods, and assumptions. <i>Proc Natl Acad Sci</i> 106 , 19644–19650.						
722							
722							
123							
724							
725							

727 Figure and Table Legends

Figure 1 | Representative ecological niche models. Model results for the present, Last Interglacial, and mid-Pliocene Warm Period for two species: Anomia simplex and Oliva savana. Binary and continuous predictions are presented, with binary predictions thresholded using the mean suitability value from the continuous output. For the binary predictions, yellow=suitable and dark blue=unsuitable, whereas for the continuous predictions, darker greys indicate higher suitability. All analyses were conducted within the geographic extent shown. Note that the modelled shorelines do not match the continental shorelines because of the nature of our GCM data and the need to capture the higher sea levels characteristic of the mid-Pliocene Warm Period. See electronic supplementary material, figures S4-5, for remaining species analysed.



- / 10

- 748 Figure 2 | Representative results from niche comparison analyses. Comparisons for
- 749 Anomia simplex and Oliva sayana using a PCA on the three most important
- environmental variables: maximum and minimum surface temperature, and maximum
- surface salinity. Comparisons are shown for the Last Interglacial (LIG, ~130 Ka), mid-
- Pliocene Warm Period (mPWP, ~3.1 Ma), and present-day (PI). The histograms show the
- null distribution of similarity values (D) drawn from the study area, with the observed
- similarity value in red. All comparisons indicate that niches are statistically more similar
- than expected given the environmental backgrounds. For other comparisons, see table 1
- and electronic supplementary material, table S3 and dataset S2.



. . .

772 Table 1 | Comparisons of multi-dimensional niches and thermal tolerance limits.

773 Bold values indicate non-significant results. All other comparisons are statistically more

similar than expected given the environmental background of the variable in question.

Note that it is possible for two niches to be more similar than expected based on the environment available for one time slice, but less similar than expected based on the

environment available for the other. See electronic supplementary material, dataset S2,

for graphical depictions of similarity tests

Spec	cies/Comparison	mPWP-LIG	LIG-mPWP	PI-LIG	LIG-PI	mPWP-PI	PI-mPWP
Envi	ronmental comparison: p-valu	les for tests u	sing PCA on 3	most imp	ortant va	riables	
	Anomia simplex	0.01	0.00	0.00	0.00	0.01	0.00
Bivalvia	Crassostrea virginica	0.04	0.00	0.03	0.00	0.00	0.00
	Dinocardium robustum	0.01	0.00	0.00	0.02	0.09	0.00
	Lucina pensylvanica	0.25	0.01	0.00	0.00	0.03	0.03
	Mercenaria campechiensis	0.01	0.00	0.00	0.02	0.00	0.00
	Bulla occidentalis	0.34	0.54	0.10	0.29	0.31	0.20
da	Crepidula fornicata	0.03	0.00	0.02	0.00	0.29	0.58
obc	Neverita duplicata	0.08	0.00	0.01	0.44	0.01	0.00
Basti	Oliva sayana	0.02	0.00	0.00	0.00	0.00	0.00
9	Terebra dislocata	0.02	0.00	0.00	0.01	0.00	0.00
Envi	ironmental comparison: ma	aximum surf	ace temperat	ure			
	Anomia simplex	0.04	0.00	0.00	0.00	0.00	0.00
ia.	Crassostrea virginica	0.00	0.00	0.00	0.02	0.01	0.00
alv	Dinocardium robustum	0.03	0.00	0.00	0.00	0.09	0.00
Bi	Lucina pensylvanica	0.00	0.00	0.00	0.00	0.81	0.42
	Mercenaria campechiensis	0.01	0.00	0.00	0.03	0.00	0.00
_	Bulla occidentalis	0.00	0.00	0.02	0.60	0.96	0.02
oda	Crepidula fornicata	0.00	0.00	0.00	0.00	0.01	0.00
rop	Neverita duplicata	0.01	0.00	0.00	0.06	0.00	0.00
Gast	Oliva sayana	0.00	0.00	0.01	0.15	0.54	0.00
Ŭ	Terebra dislocata	0.03	0.00	0.00	0.02	0.43	0.02
Envi	ironmental comparison: mi	nimum surfa	ace temperat	ure			
	Anomia simplex	0.21	0.00	0.00	0.00	0.00	0.00
via	Crassostrea virginica	0.91	0.05	0.00	0.03	0.01	0.00
Gastropoda Bivalv	Dinocardium robustum	0.11	0.00	0.00	0.00	0.00	0.00
	Lucina pensylvanica	0.68	0.00	0.00	0.00	0.77	0.22
	Mercenaria campechiensis	0.82	0.00	0.00	0.00	0.01	0.00
	Bulla occidentalis	0.66	0.00	0.09	0.10	0.85	0.10
	Crepidula fornicata	0.17	0.00	0.01	0.00	0.44	0.00
	Neverita duplicata	0.50	0.00	0.05	0.00	0.00	0.00
	Oliva sayana	0.07	0.00	0.03	0.00	0.29	0.00
0	Terebra dislocata	0.18	0.00	0.00	0.00	0.05	0.00